

The relative compliance of energy-storing tendons may
be due to the helical fibril arrangement of their fascicles

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Abstract

A non-linear elastic microstructural model is used to investigate the relationship between structure and function in energy-storing and positional tendons. The model is used to fit mechanical tension test data from the equine common digital extensor tendon (CDET) and superficial digital flexor tendon (SDFT), which are used as archetypes of positional and energy-storing tendons, respectively. The fibril crimp and fascicle helix angles of the two tendon types were used as fitting parameters in the mathematical model in order to predict their values. The fibril Young's modulus and collagen volume fraction were taken from the literature and the matrix shear modulus was estimated from previously collected mechanical test data. The fibril crimp angles were predicted to be $15.9^\circ \pm 2.9^\circ$ in the CDET and $17.9^\circ \pm 5.7^\circ$ in the SDFT and were not found to be statistically significantly different between the two tendon types ($p = 0.420$). The fascicle helix angles were predicted to be $8.8^\circ \pm 9.4^\circ$ in the CDET and $24.9^\circ \pm 14.2^\circ$ in the SDFT and were found to be highly statistically significantly different between the two tendon types ($p = 0.001$). This supports previous qualitative observations that helical sub-structures are more likely to be found in energy-storing tendons than in positional tendons and explains that the relative compliance of energy-storing tendons may be directly caused by these helical sub-structures.

Key words: Collagen, Mathematical modelling, Micromechanics, Non-linear elasticity, Structure-function

1 Nomenclature

E	fibril Young's modulus
μ	ground state shear modulus of tendon interfascicular matrix
ϕ	collagen volume fraction
θ_o	outer fibril crimp angle
α	fascicle helix angle
\mathbf{M}	fascicle alignment vector
λ, e	longitudinal stretch/strain
\mathbf{F}	deformation gradient
$\mathbf{e}_i, \mathbf{E}_J$	basis vectors in deformed/undeformed configuration
r, θ, z	circular cylindrical coordinates in deformed configuration
R, Θ, Z	circular cylindrical coordinates in undeformed configuration
W	strain energy function
I_1, I_4	isotropic/anisotropic strain invariant
λ^*	critical stretch at which toe-region ends
β	$2(1 - \cos^3 \theta_o)/(3 \sin^2 \theta_o)$
γ, η	constants defined in equations (5) and (6), respectively
\mathbf{S}	nominal stress
p	Lagrange multiplier
S_{zz}	longitudinal nominal stress
F	force in interfascicular matrix at 10% of failure load
l, A	interfascicular matrix thickness/contact area
Δx	extension of interfascicular matrix at 10% of failure load

3 **1. Introduction**

4 Tendons have varying mechanical requirements depending on their function.
5 Positional tendons need to be stiff in order to keep joints in place, whereas
6 energy-storing tendons play a role in locomotion (Alexander, 1991) and are
7 necessarily more compliant (Lichtwark and Wilson, 2007 and Lichtwark and
8 Wilson, 2008). This specialisation of mechanical properties between tendon
9 types occurs despite them being composed of the same elementary materials -
10 primarily collagen type I, which is organised into a hierarchical structure con-
11 sisting of fibrous sub-units of varying diameters, each of which is interspersed
12 with a small amount of primarily non-collagenous matrix (Kastelic et al., 1978).
13 The fundamental building block of the collagen hierarchy is the fibril, which
14 has a diameter of 10-20 nm and aggregates to build fibres (diameter: 10-50
15 μm) and fascicles (50-400 μm). It is thought that structural and compositional
16 differences in this hierarchy give rise to the differing mechanical properties of
17 different tendons (Thorpe et al., 2013a, 2013b).

18 One approach to determine how the geometrical arrangement of tendon
19 sub-units affects gross mechanical properties is to use mathematical modelling.
20 Many models have been proposed over the last several decades to describe the
21 mechanical behaviour of tendons, and soft tissues in general; however, many
22 of these are either phenomenological, e.g. (Gou, 1970), or contain a very large
23 number of parameters, e.g. (Limbirt, 2011), some of which may be extremely
24 challenging to measure experimentally. An overview of the approaches to tendon
25 modelling is given in the introduction of (Shearer et al., 2016).

26 To avoid the problems associated with earlier approaches, two models have
27 previously been developed (Shearer, 2015a, 2015b) with the specific aim of en-
28 suring a microstructural basis, whilst keeping the number of necessary parame-
29 ters to a minimum. The latter of these requires only two constitutive parameters
30 and four structural quantities, namely: the collagen fibril Young's modulus E ,
31 the matrix shear modulus μ , the collagen volume fraction ϕ , the outer fibril
32 crimp angle θ_o , the fascicle helix angle α (this term was referred to as the *fibril*
33 helix angle in (Shearer, 2015b)) and the fascicle alignment vector \mathbf{M} (see Figure
34 1). All of these quantities can potentially be measured via either mechanical
35 testing (Wenger et al., 2007), histology (Screen et al., 2005) or X-ray micro-
36 computed tomography (Shearer et al., 2014; Balint et al., 2016; Shearer et al.,
37 2016). In the current paper, this model (Shearer, 2015b) is used to investigate
38 the stress-strain behaviour of two types of equine tendon: one positional - the
39 common digital extensor tendon (CDET), and one energy-storing - the super-
40 ficial digital flexor tendon (SDFT). It is demonstrated that the differences in
41 mechanical properties between the two tendon types can be explained as being
42 entirely the result of differences in the geometical arrangement of collagen with-
43 ing the fascicles as opposed to differences in their constitutive parameters. The
44 model predicts that the SDFT is likely to have a considerably larger fascicle
45 helix angle than the CDET - a prediction that supports previous experimental
46 observations (Thorpe et al., 2013b).

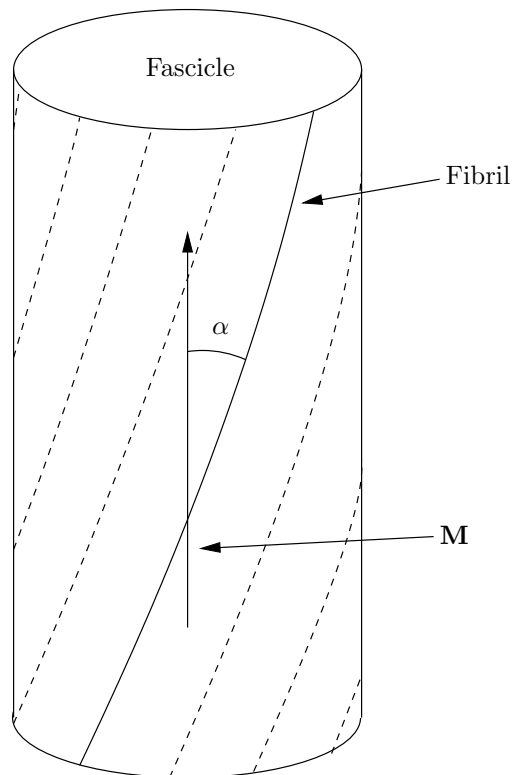


Figure 1: Diagram illustrating the fascicle helix angle α and the fascicle alignment vector \mathbf{M} .

Note that this diagram omits fibril crimp and instead shows the *average* fibril direction.

47 **2. Methods**

48 *2.1. Mechanical testing*

49 The mechanical test data were collected for a previous study (Thorpe et
50 al., 2012) and the testing protocol is described in detail therein. Briefly, the
51 CDET and SDFT were dissected from the left forelimbs of 18 horses aged 3-
52 20 years and frozen until the day of testing. On the day of testing, tendons
53 were thawed and their cross-sectional areas were measured, as described by
54 (Goodship and Birch, 2005). The tendons were mounted vertically in a servo-
55 hydraulic materials testing machine (Dartec Ltd., Stroubridge, UK) with a 50
56 kN load cell and were gripped with cryoclamps cooled by liquid carbon dioxide
57 (Riemersa and Schamhardt, 1982). They were pre-loaded to 25 N (CDET) or
58 100 N (SDFT) and were subjected to 20 preconditioning cycles between 0% and
59 5.25% strain at a frequency of 0.5 Hz, using a protocol adapted from (Batson et
60 al., 2003). The load was then removed so that slack was visible in the tendons,
61 which were then tested to failure at a rate of 5%/s. The stresses in the tendons
62 were recorded as forces per unit undeformed areas, so that the reported values
63 are nominal stresses, and the displacements at which the initial pre-loads were
64 reached were taken as the start points for the tests in all specimens.

65 *2.2. Mathematical modelling*

66 Each tendon is modelled as an incompressible, transversely isotropic, non-
67 linear elastic cylinder, subjected to a longitudinal stretch λ (≥ 1), so that the

68 deformation gradient is given by (Shearer, 2015b)

$$\mathbf{F} = F_{iJ} \mathbf{e}_i \otimes \mathbf{E}_J, \quad F_{iJ} = \begin{pmatrix} \lambda^{-\frac{1}{2}} & 0 & 0 \\ 0 & \lambda^{-\frac{1}{2}} & 0 \\ 0 & 0 & \lambda \end{pmatrix}, \quad (1)$$

69 where \mathbf{e}_i , $i = (r, \theta, z)$, and \mathbf{E}_J , $J = (R, \Theta, Z)$, are deformed and undeformed

70 unit vectors in the radial, azimuthal and longitudinal directions, respectively.

71 The longitudinal stretch is related to the longitudinal strain e via $\lambda = 1 + e$.

72 To calculate the theoretical nominal stresses, the strain energy function from

73 (Shearer, 2015b) is utilised:

$$W = (1 - \phi) \frac{\mu}{2} (I_1 - 3), \quad I_4 < 1, \quad (2)$$

$$\begin{aligned} W = (1 - \phi) \frac{\mu}{2} (I_1 - 3) + \phi \frac{E}{3 \sin^2 \theta_o} & \left(2 \cos \alpha \sqrt{I_4} - \right. \\ & \left. 3 \log \left(2 \left(\cos^2 \alpha \sqrt{I_4} + \cos \alpha \sqrt{\sin^2 \alpha + I_4 \cos^2 \alpha} \right) \right) + \right. \\ & \left. \frac{\cos \alpha \sqrt{I_4}}{\sin^2 \alpha \sqrt{\sin^2 \alpha + I_4 \cos^2 \alpha}} \right) + \gamma, \quad 1 \leq I_4 \leq \lambda^{*2}, \quad (3) \end{aligned}$$

$$\begin{aligned} W = (1 - \phi) \frac{\mu}{2} (I_1 - 3) + \phi E & \left(\beta \cos \alpha \sqrt{I_4} - \right. \\ & \left. \log \left(\cos^2 \alpha \sqrt{I_4} + \cos \alpha \sqrt{\sin^2 \alpha + I_4 \cos^2 \alpha} \right) \right) + \eta, \quad I_4 > \lambda^{*2}, \quad (4) \end{aligned}$$

74 where I_1 and I_4 are strain invariants as defined in (Holzapfel and Ogden, 2010),

75 for example, $\lambda^* = 1 / \cos \alpha \sqrt{1 / \cos^2 \theta_o - \sin^2 \alpha}$ is the critical stretch at which

76 the toe-region ends (Shearer, 2015b), $\beta = 2(1 - \cos^3 \theta_o) / (3 \sin^2 \theta_o)$,

$$\gamma = -\frac{E}{3 \sin^2 \theta_o} \left(2 \cos \alpha - 3 \log (\cos^2 \alpha + \cos \alpha) + \frac{\cos \alpha}{\sin^2 \alpha} \right), \quad (5)$$

and

$$\eta = \gamma + E \left(\left(\frac{\cos \theta_o}{\sin^2 \theta_o} \frac{1}{\sin^2 \alpha} + \frac{2}{\sin^2 \theta_o} - 3\beta \right) \sqrt{\frac{1}{\cos^2 \theta_o} - \sin^2 \alpha} - 3 \frac{\cos^2 \theta_o}{\sin^2 \theta_o} \log \left(\cos \alpha \left(\frac{1}{\cos \theta_o} + \sqrt{\frac{1}{\cos^2 \theta_o} - \sin^2 \alpha} \right) \right) \right). \quad (6)$$

77 Equations (1)–(4) can be substituted into the general equation for the nominal
 78 stress in a transversely isotropic non-linear elastic material, which, for a strain
 79 energy function that is only dependent on I_1 and I_4 is given by:

$$\mathbf{S} = -p\mathbf{F}^{-1} + 2W_1\mathbf{F}^T + 2W_4\mathbf{M} \otimes \mathbf{F}\mathbf{M}, \quad (7)$$

80 where p is a Lagrange multiplier associated with the incompressibility constraint,
 81 $W_i = \partial W / \partial I_i$ and, \mathbf{M} is a unit vector oriented in the direction of the fascicles
 82 in the undeformed configuration. It is assumed that the fascicles are coaligned
 83 with the longitudinal axis of the tendon in both the CDET and SDFT, so that
 84 $\mathbf{M} = \mathbf{E}_Z$. In reality, this is not the case; however, it is assumed that the
 85 deviation from longitudinal alignment is small enough to be negligible.

86 Upon applying stress-free boundary conditions on the curved surface of the
 87 cylinder, thus determining the value of p , the following expression is obtained
 88 for the longitudinal nominal stress:

$$S_{zz} = \begin{cases} (1 - \phi)\mu(\lambda - \lambda^{-2}) + \phi \frac{E \cos \alpha}{3 \sin^2 \theta_o} \times \\ \left(2 - \frac{3}{\sqrt{\sin^2 \alpha + \lambda^2 \cos^2 \alpha}} + \frac{1}{(\sin^2 \alpha + \lambda^2 \cos^2 \alpha)^{\frac{3}{2}}} \right), & 1 \leq \lambda \leq \lambda^*, \\ (1 - \phi)\mu(\lambda - \lambda^{-2}) + \\ \phi E \cos \alpha \left(\beta - \frac{1}{\sqrt{\sin^2 \alpha + \lambda^2 \cos^2 \alpha}} \right), & \lambda > \lambda^*. \end{cases} \quad (8)$$

89 This expression is used to model the mechanical test data obtained for the
90 CDET and SDFT.

91 *2.3. Parameter selection*

92 In order to fit equation (8) to the experimental data, it is necessary to select
93 values for the constitutive parameters in the model.

94 *2.3.1. Constitutive parameters*

95 For the collagen Young's modulus, there is a wide range of reported values
96 in the literature, ranging from 32 MPa (Graham et al., 2004) to 12 GPa (Eppell
97 et al., 2006). To the authors' knowledge there is no data available for equine
98 collagen fibrils; therefore, bovine data was used as a substitute - the value
99 selected here was 1.9 GPa, which is the value reported by Grant et al. (2008)
100 for bovine collagen fibrils under ambient conditions.

101 A lack of data is available in the literature for the matrix shear modulus due
102 to the difficulties involved in measuring it experimentally; therefore, a custom
103 method was developed to estimate the values of this parameter in the CDET
104 and SDFT based on mechanical test data from a previous study (Thorpe et al.,
105 2012). The testing protocol is described in detail within that paper; however,
106 briefly, groups of two fascicles bound together by the interfascicular matrix
107 were dissected from the CDET and SDFT (n=17, 12 samples per tendon). The
108 fascicles were secured into a custom-made dissection rig and the opposing end of
109 each fascicle was cut transversely, leaving 10mm of intact interfascicular matrix.
110 The intact end of each fascicle was then secured in a materials testing machine

111 and pulled apart to failure at a speed of 1 mm/s. Force and extension data
112 were recorded and the point at which a load of 0.02N was reached was defined
113 as the test start point. The matrix shear modulus was then estimated using the
114 following equation:

$$\mu = \frac{Fl}{A\Delta x}, \quad (9)$$

115 where F is the force and Δx is the extension in the matrix at 10% of the failure
116 load, l is its thickness and A is its contact area. The contact area was estimated
117 by multiplying the average fascicle diameter by the test length (10mm). The
118 thickness was estimated based on values calculated by Ali et al. (2015). Using
119 this method, it was estimated that the matrix shear modulus of the CDET is
120 1.01 kPa and of the SDFt is 1.62 kPa.

121 2.3.2. Structural parameters

122 For the collagen volume fraction, an estimate was made based on the collagen
123 area fractions reported in (Screen et al., 2005) for nonincubated rat tail tendon -
124 the selected value was 0.8. The fibril crimp and helix angles were used as fitting
125 parameters in order to predict their values. The function (8) was used to fit each
126 of the experimental data sets up to 10% strain, beyond which it was assumed
127 that the deformation was no longer elastic. The experimental data was fitted
128 using the *NonlinearModelFit* command in *Mathematica 9.0* (Wolfram Research,
129 Inc., Champaign, Illinois, 2008) subject to the constraints $0 \leq \theta_o \leq 90^\circ$ and
130 $0 \leq \alpha \leq 90^\circ$.

Tendon	Crimp angle (θ_o)	Helix angle (α)
CDET	$15.9^\circ \pm 2.9^\circ$	$8.8^\circ \pm 9.4^\circ$
SDFT	$17.9^\circ \pm 5.7^\circ$	$24.9^\circ \pm 14.2^\circ$

Table 1: Predicted fibril crimp and helix angles.

131 3. Results

132 The predicted fibril crimp and fascicle helix angles according to the model
133 fit are listed in table 1 (given as mean \pm standard deviation) and an example fit
134 to the experimental data is plotted in figure 2 (plots of all 18 fits are provided in
135 supplementary material). The minimum coefficient of determination (R^2 value)
136 across all 36 data sets was 0.979. There was no statistically significant difference
137 between the crimp angles of the CDET and SDFT ($p = 0.420$); however, there
138 was a highly statistically significant difference between the helix angles of the
139 CDET and SDFT ($p = 0.001$) according to the Mann-Whitney test.

140 4. Discussion

141 The fitting process predicted a much larger fascicle helix angle in the SDFT
142 than in the CDET. These predictions agree with the qualitative observations
143 in (Thorpe et al., 2013b) and support the hypothesis that helical substructures
144 are more likely to be found in energy-storing tendons than in positional
145 tendons. The model also provides a link between the microstructures and me-
146 chanical functions of these tendons, explaining that the relative compliance of
147 energy-storing tendons is caused directly by the helical fibril arrangement of

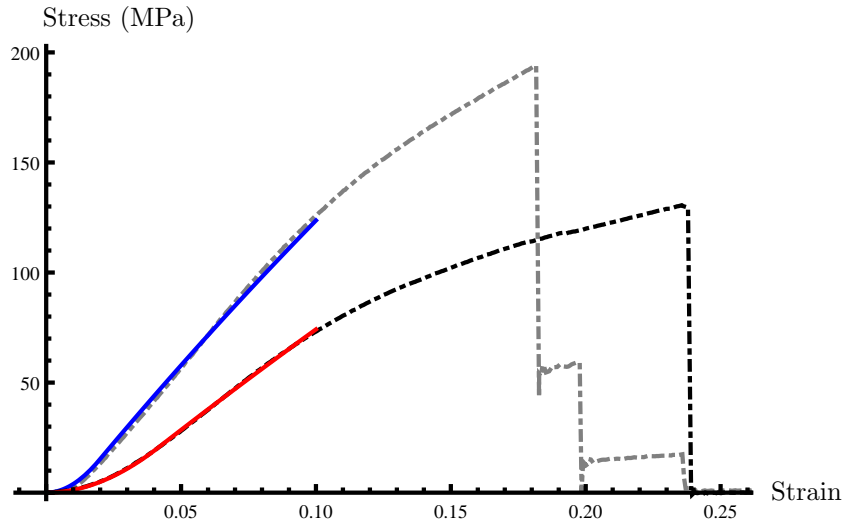


Figure 2: Example experimental (dashed) and theoretical (solid) stress-strain curves for the CDET (grey/blue) and SDFT (black/red).

148 their fascicles, and *not* by differences in their fibril Young's modulus or crimp
 149 angles.

150 Whilst different values of the matrix shear modulus were used to model each
 151 tendon, this does not affect the conclusions above as the stress in the matrix
 152 at 10% strain was 0.0001% of the total stress in the tendon on average and
 153 therefore the contribution of this phase was not important compared to that of
 154 the fibrils. The modelling was repeated three times with the same matrix shear
 155 modulus being used for both the CDET and SDFT using the values: $\mu = 1.01$
 156 kPa, $\mu = 1.62$ kPa and $\mu = 10$ kPa. In all three cases, the predicted helix and
 157 crimp angles differed from those reported in table 1 by less than 0.01° .

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160 **Data statement**

161 TBC.

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