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# Appetite Suppression and Altered Food Preferences Coincide with Changes in Appetite-Mediating Hormones During Energy Deficit at High Altitude, But Are Not Affected by Protein Intake

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#### **Abstract**

Karl, J. Philip, Renee E. Cole, Claire E. Berryman, Graham Finlayson, Patrick N. Radcliffe, Matthew T. Kominsky, Nancy E. Murphy, John W. Carbone, Jennifer C. Rood, Andrew J. Young, and Stefan M. Pasiakos. Appetite suppression and altered food preferences coincide with changes in appetite-mediating hormones during energy deficit at high altitude, but are not affected by protein intake. High Alt Med Biol. 19:156-169, 2018.—Anorexia and unintentional body weight loss are common during high altitude (HA) sojourn, but underlying mechanisms are not fully characterized, and the impact of dietary macronutrient composition on appetite regulation at HA is unknown. This study aimed to determine the effects of a hypocaloric higher protein diet on perceived appetite and food preferences during HA sojourn and to examine longitudinal changes in perceived appetite, appetite mediating hormones, and food preferences during acclimatization and weight loss at HA. Following a 21-day level (SL) period, 17 unacclimatized males ascended to and resided at HA (4300 m) for 22 days. At HA, participants were randomized to consume measured standard-protein (1.0 g protein/kg/d) or higher protein (2.0 g/kg/d) hypocaloric diets (45% carbohydrate, 30% energy restriction) and engaged in prescribed physical activity to induce an estimated 40% energy deficit. Appetite, food preferences, and appetitemediating hormones were measured at SL and at the beginning and end of HA. Diet composition had no effect on any outcome. Relative to SL, appetite was lower during acute HA (days 0 and 1), but not different after acclimatization and weight loss (HA day 18), and food preferences indicated an increased preference for sweetand low-protein foods during acute HA, but for high-fat foods after acclimatization and weight loss. Insulin, leptin, and cholecystokinin concentrations were elevated during acute HA, but not after acclimatization and weight loss, whereas acylated ghrelin concentrations were suppressed throughout HA. Findings suggest that appetite suppression and altered food preferences coincide with changes in appetite-mediating hormones during energy deficit at HA. Although dietary protein intake did not impact appetite, the possible incongruence with food preferences at HA warrants consideration when developing nutritional strategies for HA sojourn.

**Keywords:** ingestive behavior, hypoxia, energy restriction, hunger, satiety, craving

### Introduction

OWLANDERS SOJOURNING at high altitude (HA; 2500–5000 m) commonly experience unintentional weight loss (Westerterp and Kayser, 2006; Pasiakos et al., 2017). The

etiology is multifactorial, but reduced energy intake is a contributing factor (Hamad and Travis, 2006; Westerterp and Kayser, 2006), and coincides with a loss of appetite putatively characterized by a rapid reduction in hunger during meals and early satiation (Westerterp-Plantenga et al., 1999).

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Some evidence suggests that this HA anorexia may be related to hypoxia-induced increases in the anorexigenic hormones, leptin (Tschop et al., 1998; Shukla et al., 2005; Barnholt et al., 2006; Sierra-Johnson et al., 2008; Snyder et al., 2008) and cholecystokinin (CCK) (Bailey et al., 2000), and suppression of the orexigenic hormone ghrelin (Shukla et al., 2005; Riepl et al., 2012; Wasse et al., 2012; Matu et al., 2017a, 2017b). However, findings are not consistent across all studies (Zaccaria et al., 2004; Benso et al., 2007; Vats et al., 2007; Riepl et al., 2012; Aeberli et al., 2013; Debevec et al., 2014, 2016; Mekjavic et al., 2016; Morishima and Goto, 2016). These inconsistencies and heterogeneous study designs have impeded reaching consensus on the role of appetitemediating hormones in the development and persistence of HA anorexia (Hamad and Travis, 2006; Westerterp and Kayser, 2006; Raff et al., 2008; Sierra-Johnson et al., 2008; Debevec, 2017). An improved understanding of the contribution of appetite-mediating hormones to HA anorexia, and how that impact evolves with acclimatization, could help develop optimized strategies for mitigating HA anorexia.

Higher protein (>1.2 g protein/kg body weight/d) diets preserve fat-free mass during weight loss (Wycherley et al., 2012; Leidy et al., 2015; Pasiakos et al., 2015a, 2015b) which may be beneficial for maintaining physical function and health. These benefits underpin recommendations supporting higher protein diets for athletes (Thomas et al., 2016) and military personnel (Pasiakos et al., 2013a, 2013b, 2015a, 2015b) and have stimulated recent interest in studying the effectiveness of higher protein diets for mitigating fat-free mass loss during HA sojourn (Pasiakos et al., 2017). However, protein increases fullness (Dhillon et al., 2016) and is more satiating than carbohydrate or fat (Leidy et al., 2015). These effects may be potentiated at HA due to the higher thermogenic effect of protein relative to fat (Veldhorst et al., 2008) which could worsen hypoxia by increasing postprandial oxygen consumption (Westerterp and Kayser, 2006; Veldhorst et al., 2008). Higher protein diets may also be incompatible with food preferences at HA, which have favored carbohydrate in some (Boyer and Blume, 1984; Kayser et al., 1993; Westerterp-Plantenga et al., 1999; Bailey et al., 2004; Hamad and Travis, 2006; Matu et al., 2017a, 2017b), but not all (Rose et al., 1988; Reynolds et al., 1998; Aeberli et al., 2013), studies. Collectively, these effects could prove counterproductive to retaining fat-free mass during HA-induced weight loss by exacerbating HA anorexia and further reducing energy intake. Although these possibilities have been acknowledged (Westerterp and Kayser, 2006; Pasiakos et al., 2017), to date, the effects of higher protein diets on appetite and food preferences during HA sojourn are undetermined.

We recently reported that consuming a controlled higher protein hypocaloric diet relative to a controlled standard-protein hypocaloric diet did not protect fat-free mass during weight loss over 22 days at HA (Berryman et al., 2017). This report details changes in perceived appetite, appetite-mediating hormones, and food preferences measured as secondary outcomes during that study. The objectives associated with these outcomes were: (1) to determine the effects of a higher protein hypocaloric diet relative to a standard-protein hypocaloric diet on perceived appetite and food preferences during HA sojourn and (2) to examine longitudinal changes in perceived appetite, appetite-mediating hormones, and food preferences during acclimatization and weight loss at HA.

#### **Materials and Methods**

#### **Participants**

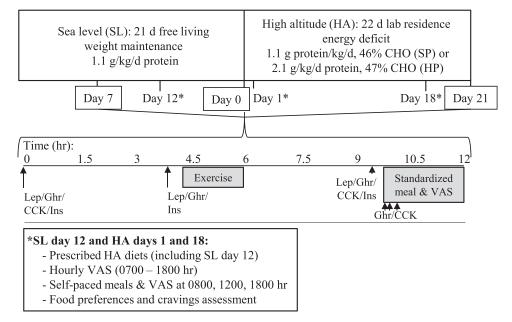
Twenty-one men gave their free and informed voluntary consent to participate in this randomized controlled trial. Participants were 18–42 years, exercised ≥2 days/week, were born at altitudes <2100 m, had not traveled to altitudes >1200 m for ≥5 days within 2 month of participation, and were not taking any medications. Exclusion criteria included musculoskeletal injury, metabolic, cardiovascular, and gastrointestinal abnormalities or disorders, diseases affecting macronutrient metabolism or ability to participate in strenuous exercise, sleeping disorders or apnea, history of HA pulmonary or cerebral edema, alcoholism, substance abuse, anabolic steroid use, anemia, sickle cell trait, and elevated prothrombin time. Participants were asked to abstain from alcohol, caffeine, dietary supplements, and nicotine products throughout the study. The study was approved by the Institutional Review Board at the U.S. Army Research Institute of Environmental Medicine (Natick, MA) and conducted May-Aug 2016. Investigators adhered to the policies regarding the protection of human subjects as prescribed in Army Regulation 70-25, and the research was conducted in adherence with the provisions of 32 CFR Part 219. The trial was registered on www.clinicaltrials.gov as NCT02731066.

#### Study design

The 43-day study included a 21-day sea level (SL) phase (50 m; Natick, MA), which was immediately followed by 22 days at HA (4300 m; Pikes Peak, CO) (Berryman et al., 2017) (Fig. 1). Upon enrollment, investigators randomized participants using computer-generated randomization to consume a standard-protein or higher protein diet at HA. During SL, participants were free living but visited the laboratory daily and maintained habitual exercise. Compliance with diet instructions and body weight maintenance was verified by diet records and daily body weight monitoring.

On SL day 21 participants were flown from Boston, MA to Denver, CO where they were placed on supplemental oxygen until ascent. In the early morning after arrival in Colorado (HA day 0), participants ascended to 4300 m by car where they resided for 22 days in a temperature-controlled indoor environment at the U.S. Army Research Institute of Environmental Medicine Maher Memorial Laboratory.

During HA, participants consumed a measured higher protein or standard-protein diet designed to provide 70% of SL weight maintenance energy requirements. To induce a 40% total energy deficit, supervised low-to-moderate intensity exercise was conducted to increase physical activity expenditure by 10% of weight-maintenance energy needs. The duration and magnitude of the energy deficit were designed to replicate those used in a recent study from our group investigating the effects of varying protein intake on body composition during weight loss at SL (Pasiakos et al., 2013a, 2013b) and was in accord with the primary study objective of determining the efficacy of a higher protein diet for fat-free mass retention during weight loss at HA (Berryman et al., 2017). This magnitude of energy deficit was within ranges reported during HA sojourns in which dietary intake was ad libitum (Hoyt et al., 1994; Westerterp et al., 1994) and was consistent with previous studies investigating endocrine responses at HA (Barnholt et al., 2006).



**FIG. 1.** Study design. Volunteers were followed for 21 days at SL, then for 22 days while living at HA (4300 m) and consuming standard-protein or higher protein hypocaloric diets. CCK, cholecystokinin; CHO, carbohydrate; Ghr, acylated ghrelin; Ins, insulin; Lep, leptin; VAS, visual analog scale measuring appetite; SL, sea level; HA, high altitude.

The shortened version of the Environmental Symptoms Questionnaire was administered at SL and daily during HA to assess the incidence of acute mountain sickness, which was defined as a Lake Louise score of >0.7 (Beidleman et al., 2007). Oxygen saturation was measured at SL and daily during HA by finger pulse oximetry (Nonin Model 7500 or 8600; Plymouth, MN).

## Study diets

During SL, all participants consumed a self-selected weight maintaining diet. Registered dietitians provided instruction on consuming a diet containing 1.0 g protein/kg/d, 45%–65% energy from carbohydrate, and 20%–35% energy from fat. Twenty-four hour food records were completed and reviewed every 2–3 days, and additional education was provided as needed to promote adherence. During SL, most participants consumed meals from the local cafeteria or local restaurants.

During HA, all food and caloric beverages consumed were strictly controlled, weighed and measured, and provided by study staff. Meals and snacks were supervised, and participants were instructed to consume all provided food and beverages. Any uneaten foods were weighed and documented. Diets were individualized, followed a five-day rotating menu, and were designed to provide a standard (SP;  $1.0\pm02$  g/kg/d) or higher (HP;  $2.0\pm0.2$  g/kg/d) amount of protein, and 45% of total energy as carbohydrate each day. Fat intake was reduced in HP to accommodate the higher protein intake (Table 1). Most of the foods were entrées, sides, and snack items included in U.S. military Meals Ready-to-Eat rations. These items were supplemented with fresh fruits and vegetables, fruit snacks, olive oil, and ranchflavored salad dressing. Protein intake was also manipulated using a whey-protein beverage (Isopure<sup>®</sup> Zero Carb; Isopure Co., Hauppauge, NY). Although the diets differed in appearance, participants were not told to which dietary group they were assigned. Whether participants could correctly identify their group assignment was not assessed. Water and noncaffeinated calorie-free sodas were allowed *ad libitum*. Dietary intake was assessed using daily food records, the Combat Rations Database (U.S. Army Natick Soldier Research Development and Engineering Center and USAR-IEM), and Food Processor SQL v.10.14 (ESHA Research, Salem, OR).

Effects of protein intake on appetite and food preferences during energy deficit at HA (objective 1)

Appetite testing. To determine the effects of protein intake on appetite at HA, perceived appetite was measured using visual analog scales administered hourly, and immediately before and after meals on SL day 12 and HA days 1 and 18. Participants consumed the same individualized menu

Table 1. Baseline Participant Characteristics and Actual Dietary Intakes Over 22 Days at High Altitude (4300 m)

	Standard protein (n=8)	Higher protein (n=9)	
Age, year	23±3	24±7	
BMI, $kg/m^2$	$27.0 \pm 4.0$	$25.5 \pm 3.1$	
Body fat,%	$22.8 \pm 7.0$	$22.6 \pm 5.5$	
TDEE, kcal/d	$2744 \pm 264$	$2635 \pm 344$	
Energy intake, kcal/d	$1950 \pm 186$	$1885 \pm 269$	
Protein intake, g/kg/d	$1.1 \pm 0.0$	$2.1 \pm 0.0 *$	
Protein intake,%	$18 \pm 2$	$33 \pm 2*$	
Carbohydrate intake, %	$46 \pm 1$	$47 \pm 1$	
Fat intake,%	$39\pm2$	$23 \pm 3*$	

Values are mean ± SD. TDEE, total daily energy expenditure during weight maintenance at sea level. Adapted from Berryman et al. (2017).

<sup>\*</sup>Independent samples *t*-test, p < 0.05.

on all three test days and ate all meals and snacks at the same times each day. On SL day 12 the prescribed meals matched the HA energy prescription and diet assignment to eliminate any confounding resulting from differences in energy intake between SL and HA. Diets were constant within participants across test days, but because the diets were individualized, the types of foods and the volume of food and beverages consumed differed between participants. Water was allowed ad libitum between meals.

Food preferences and cravings. Questionnaires were administered on SL day 12 and HA days 1 and 18 to determine the effects of protein intake on food preferences and cravings at HA. Food preferences were measured using the Leeds Food Preference Questionnaire (LFPQ), which was administered before and after lunch. The LFPQ is a computerized platform that uses pictures of individual food items selected from a validated database of common foods to measure different components of food preferences and hedonics (Finlayson et al., 2007). Pictures for each test varied in two separate dimensions; fat (low and high) and taste (sweet and savory) or protein (low and high) and taste (sweet and savory). Four pictures from each food type were selected resulting in 16 separate pictures being used within each test (Supplementary Table S1; Supplementary materials are available online at http://www.liebertpub.com/ham).

The LFPQ measures three constructs as follows: explicit liking (i.e., perceived hedonic impact of the food), explicit wanting (i.e., conscious desire to consume a food), and implicit wanting (i.e., subconscious motivation for a food). To measure explicit liking, the participant was shown each picture in random order and asked to respond to the question "how pleasant would you find the taste of this food right now" using a visual analog scale. To measure explicit wanting, the participant was shown each picture in random order and asked to respond to the question "how much do you want to eat this food right now" using a visual analog scale. Implicit wanting was measured using forced choice methodology in which pairs of food images from separate food categories were presented in randomized order on the computer screen. Participants were asked to as quickly as possible select the food that they most wanted to eat at that moment.

Both frequencies of selections within each food category and response time (i.e., relative preference) were recorded. The preference for different food types was computed for each metric by subtracting the mean scores (visual analog scale scores or frequency of selection and reaction time) from a comparator group (low fat/protein and savory) relative to the matched reference group (high fat/protein and sweet, respectively). Food cravings were assessed before lunch using the Food Cravings Inventory (White and Grilo, 2005) and Control of Eating Questionnaire (Dalton et al., 2015).

Longitudinal changes in appetite and appetitemediating hormones during acclimatization and weight loss at HA (objective 2)

Longitudinal changes in appetite and related hormones were examined on SL day 7 and HA days 0 and 21 by measuring circulating hormone concentrations throughout the day and by measuring appetite, CCK, and acylated ghrelin responses to a standardized meal. On these days participants did not consume their prescribed diets to conduct testing for

unrelated study objectives which have been (Berryman et al., 2017) or will be reported elsewhere. Hormone concentrations were not measured on SL day 12 and HA days 1 and 18 (i.e., full-day appetite testing) for logistical reasons. Therefore, because prescribed SP and HP diets were not consumed until after HA day 0, and because diet habituation has been reported not to influence fasting hormone concentrations independent of body weight changes (Ellis et al., 2012), we did not expect hormone concentrations to differ by diet group unless differential changes in body composition were observed. As such, determining the effects of protein intake on appetite-mediating hormones was not a study objective. However, these study days provided an opportunity to examine appetite-mediating hormone responses to acute HA exposure and whether those responses persisted after acclimatization and weight loss. In addition, these days allowed us to examine whether the rates of change of postprandial appetite responses, as was previously reported (Westerterp-Plantenga et al., 1999), and related hormones differed between HA and SL.

Fasting blood samples were collected shortly after waking following a >10-hour fast on SL day 7 and HA days 0 and 21. Additional blood samples were collected from an indwelling venous catheter 4 hours and 9.5 hours later and during a standardized meal test beginning 10 hours after the fasting blood draw (Fig. 1). No food was consumed after dinner the evening prior until the standardized meal test. However, participants consumed beverages containing 145 g (n=9) or 0 g (n=8) of carbohydrate during an 80-minute submaximal aerobic exercise bout 3.5–5 hours before the meal test, and all participants drank 25 g of whey-protein (Isopure<sup>®</sup> Zero Carb) 3 hours before the meal test. The rationale for providing these beverages was to conduct testing for unrelated study objectives (Berryman et al., 2017). Potential confounding due to carbohydrate intake was addressed by including this variable in statistical models.

Standardized meal tests occurred between 1530 and 1630 hours. For each meal test, participants were provided a fixed-portion meal to consume at a controlled eating rate and 240 g water to drink. The meal was macaroni and cheese with added butter and provided 20% of individual weight maintenance energy needs determined during SL (41%, 49%, and 10% energy from carbohydrate, fat, and protein, respectively). Participants were instructed to consume all of the food and water provided in 12 minutes while eating at a constant rate (Karl et al., 2011). Over the subsequent 105 minutes, water intake was prescribed, and no additional food or beverage was allowed. Blood samples were collected approximately 30 minutes before the meal and at 7.5, 15, and 30 minutes after starting the meal. Perceived appetite was measured before, during, and periodically for 105 minutes after completing the meal using visual analog scales.

Serum insulin and leptin and plasma acylated ghrelin and CCK were measured throughout SL day 7 and HA days 0 and 21. However, only acylated ghrelin and CCK responses were measured during the standardized meal test. For serum measures, blood was collected into Monovettes containing clotting activator (Sarstedt, Newton, NC) and stored at -20°C to -80°C until analysis. Serum insulin was determined using the IMMULITE<sup>®</sup> 2000 (Siemens Healthcare Diagnostics, Deerfield, IL). Serum leptin was determined by RIA (EMD Millipore, St. Charles, MO) according to manufacturer's instructions. For plasma measures, blood was collected into

chilled Monovettes containing EDTA-K<sub>3</sub> and 4-(2-aminoethyl)benzenesulfonyl fluoride hydrochloride (1 mg/ mL whole blood) for acylated ghrelin determination or protease inhibitor cocktail (40  $\mu$ L/mL whole blood; cOmplete, EDTA-free) for CCK determination. Acylated ghrelin aliquots were combined with 50  $\mu$ L 1N HCl/mL plasma, and all samples were stored at –20°C to –80°C until analysis. Plasma acylated ghrelin was determined by RIA (EMD Millipore) and plasma CCK by ELISA (Cusabio, College Park, MD) according to manufacturer's instructions.

### Visual analog scales

Four separate visual analog scales were used to measure self-perceived appetite (Blundell et al., 2010). Participants rated their levels of hunger and fullness, their desire to eat, and the amount of food they thought they could eat "right now" (i.e., prospective consumption) by marking anywhere on a 10-cm scale anchored by phrases representing opposite extremes of a spectrum (e.g., "not at all hungry" and "extremely hungry"). A total appetite score was computed as follows:

Overall appetite = (hunger + desire to eat + prospective consumption + (10 - fullness))/4

## Statistical analyses

Sample size estimates based on the primary study outcome, fat-free mass, indicated n=8/grp were required to detect a 1 kg between-group difference in fat-free mass loss (Berryman et al., 2017). This sample size was estimated to allow detection of a 25% difference in mean between-group appetite ratings at  $\alpha = 0.05$  and power = 0.80 based on variability measured in our previous work (Karl et al., 2016). Although a 25% difference in appetite ratings is larger than the 8%–10% difference deemed to be of practical relevance (Blundell et al., 2010), >25% differences have been reported in previous studies comparing the impact of higher and lower protein meals on appetite (Dhillon et al., 2016). Statistical analyses were completed using SPSS v.21 (IBM Analytics; Armonk, NY). All data were checked to verify adherence to model assumptions and log<sub>10</sub>-transformed when necessary. Data are presented as mean  $\pm$  SD unless otherwise noted, and statistical significance was set at  $p \le 0.05$ .

The effects of study day, diet (SP and HP), and their interaction on outcomes measured on SL day 12 and HA days 1 and 18 were tested using linear mixed models, generalized linear mixed models, and repeated measures ANOVA. Appetite ratings measured hourly on SL day 12 and HA days 1 and 18 were analyzed using area under the curve calculated from ground (AUC; where ground is y=0).

The effects of study day, diet, carbohydrate ingestion during exercise (145 g and 0 g carbohydrate), time, and their interactions on appetite ratings and hormone concentrations measured on SL day 7 and HA days 0 and 21 were tested using linear mixed models. All possible interactions except any between dietary protein group and carbohydrate supplement group were included in these models. Appetite ratings measured during and after the standardized meal test were analyzed using the metrics AUC, rate of change in appetite ratings during the meal, and rate of change in appetite

ratings after the meal, and therefore, models did not include time as a variable. To calculate rates of change, slopes were extracted from linear regression models run separately for each individual using the appetite ratings measured from 0 to 15 minutes during the meal and from 15 to 120 minutes after the meal. To aid in interpretation of hormone analyses, models were reduced by removing effects that did not significantly contribute to the overall model fit until the most parsimonious model was reached. As expected, dietary protein group did not impact outcomes measured on SL day 7 and HA days 0 and 21, and therefore, results are presented with groups combined for outcomes measured on those days. In addition, hormones were analyzed both as measured and after adjusting for plasma volume changes (Dill and Costill, 1974). Plasma volume adjustment did not affect interpretation of the results, and only unadjusted values are reported. For all models, when a significant interaction or main effect was observed, t-tests with Bonferroni corrections were used to identify pairwise differences.

#### Results

During SL testing, one individual from each diet group withdrew, and one individual from each diet group was withdrawn for nonadherence with study procedures. As such, 17 of 21 enrolled participants completed both the SL and HA phases (Table 1). Primary study outcomes are reported elsewhere (Berryman et al., 2017). Briefly, participants were weight stable during SL (mean weight change  $0.7 \, \text{kg}$  [95% CI:  $-0.3 \, \text{to} \, 1.7$ ], p = 0.21). Total weight loss during HA (SP:  $7.2 \pm 2.4 \, \text{kg}$  vs. HP:  $7.0 \pm 0.9 \, \text{kg}$ ; diet-by-day interaction, p = 0.88) and fat-free mass loss during HA as measured by dual energy X-ray absorptiometry (SP:  $4.0 \pm 3.3 \, \text{kg}$  vs HP:  $3.2 \pm 1.5 \, \text{kg}$ ; diet-by-day interaction, p = 0.53) did not differ by dietary protein group.

Lake Louise scores were elevated on HA days 0–3 relative to SL, but not thereafter (main effect of day, p<0.01), and did not differ by diet group (data not shown). On HA days 1 and 18 (i.e., full-day appetite testing) the incidence of acute mountain sickness was higher relative to SL (main effect of day, p<0.001), but did not differ by diet group (Table 2). Oxygen saturation decreased upon arrival at HA and remained below SL values throughout HA (main effect of time, p<0.01) and did not differ by diet group (data not shown). However, when only SL day 12 and HA days 1 and 18 were included in the model, oxygen saturation was lower in HP relative to SP on HA day 1 (diet-by-day interaction, p=0.02; Table 2).

Effects of protein intake on appetite and food preferences during energy deficit at HA (SL day 12 and HA days 1 and 18)

Appetite ratings measured on SL day 12 and HA days 1 and 18 indicated changes in appetite over time that were independent of dietary protein group (Fig. 2). Specifically, hunger, prospective consumption, desire to eat, and overall appetite AUC were lower, whereas fullness AUC was higher on HA day 1 relative to SL (main effect of day, p < 0.01). On HA day 18 none of these measures differed from SL. Of note, overall appetite AUC was 19% higher in SP relative to HP on HA day 18, but this difference was not statistically significant. Self-selected meal duration ranged from  $6.1\pm1.4$  minutes at SL to  $7.8\pm5.3$  minutes on HA day 18 and did not differ by day (p = 0.36) or by dietary protein group (p = 0.80).

TABLE 2. ACUTE MOUNTAIN SICKNESS, OXYGEN SATURATION, ACTUAL DIETARY INTAKES, AND MEAL ACCEPTABILITY MEASURED DURING APPETITE TESTING DAYS CONDUCTED AT SEA LEVEL AND WHILE LIVING AT HIGH ALTITUDE (4300 M)

	Standard protein (n=8)			Higher protein (n = 9)		
	SL day 12	HA day 1	HA day 18	SL day 12	HA day 1	HA day 18
$AMS, \%n^{1,2}$	0	50	12	0	78	44
Nausea, $\%n^{1,3}$	0	37	25	0	44	44
O <sub>2</sub> saturation,% <sup>4</sup>	$96.6 \pm 1.1^{a}$	$83.0 \pm 1.4^{b}$	$87.0 \pm 1.9^{c}$	$97.1 \pm 1.1^{a}$	$79.0 \pm 3.6^{b*}$	$87.2 \pm 1.9^{c}$
Energy intake, kcal/d <sup>4</sup>	$1981 \pm 185$	$2019 \pm 193$	$1966 \pm 243$	$1966 \pm 274^{a}$	$1775 \pm 387^{b}$	$1968 \pm 267^{a}$
Protein intake, $g/d^5$	$93 \pm 15$	$93 \pm 14$	$91 \pm 16$	$166 \pm 26$	$148 \pm 37$	$163 \pm 26$
Carbohydrate intake, $g/d^4$	$214 \pm 23$	$219 \pm 23$	$211 \pm 30$	$215 \pm 32^{a}$	$199 \pm 34^{b}$	$216 \pm 32^{a}$
Fat intake, $g/d^5$	$90 \pm 7$	$92 \pm 7$	$90 \pm 9$	$55 \pm 9$	$48 \pm 15$	$56 \pm 9$
Meal acceptability <sup>6,7</sup>	$7\pm2$	$6\pm2$	$5\pm3$	$7\pm2$	$6\pm2$	$5\pm3$

Values are %n or mean  $\pm$  SD.

Incidence of nausea defined as slight to extreme nausea reported on the Environmental Symptoms Questionnaire.

Overall meal acceptability was lower on HA days 1 and 18 relative to SL (main effect of day, p < 0.001) independent of dietary protein group (Table 2).

Responses to the LFPQ indicated changes in food preferences over time independent of dietary protein group (Fig. 3). Specifically, an increased preference for sweet over savory foods across all domains of the LFPQ and a conscious preference for low-protein over high-protein foods was observed on HA day 1 (main effect of day, p < 0.05), but not HA day 18, relative to SL. An increased preference for high-fat over low-fat food across all domains of the LFPQ was observed on HA day 18 relative to SL day 12 and HA day 1 (main effect of day, p < 0.05).

Scores on the Control of Eating Questionnaire indicated that the overall strength and frequency of food cravings were higher during the final week of HA relative to the final week at SL (main effect of day, p = 0.03) (Supplementary Table S2). Similarly, scores on the Food Craving Inventory indicated higher total food cravings during HA relative to the final week at SL (main effect of day, p=0.03) which was attributable to an increase in cravings for high fat and fast foods (main effect of day,  $p \le 0.02$ ) (Supplementary Table S2). No food craving metric differed between dietary protein groups.

Longitudinal changes in appetite and appetitemediating hormones during acclimatization and weight loss at HA (SL day 7 and HA days 0 and 21)

Appetite responses to a standardized test meal. Appetite responses to the standardized meal test administered on SL day 7 and HA days 0 and 21 indicated changes in appetite responses over time that were independent of both dietary protein group and carbohydrate supplementation during exercise. Specifically, hunger AUC was lower on HA day 0, but not HA day 21, relative to SL, whereas fullness AUC was higher on HA day 21, but not HA day 0, relative to SL (main effect of day,  $p \le 0.03$ ; Fig. 4A, B). Overall appetite, pro-

spective consumption, and desire to eat AUC were lower on HA days 0 and 21 relative to SL (main effect of day,  $p \le 0.01$ ; Fig. 4C–E). These differences were attributable to a combination of differences in premeal appetite ratings (main effect of day, p < 0.05) and differences in the rate of change in appetite ratings during the postprandial period (main effect of day,  $p \le 0.01$ ) across study days (Fig. 4). No differences between days in the rate of change in any appetite rating measured during the meal (0–12 minutes) were observed (main effect of day, p > 0.05). The overall acceptability of the meal was rated as "moderately liked" (mean score =  $7.1 \pm 1.6$ ), which did not change over time (main effect of day, p = 0.10) or differ between dietary protein groups (main effect of diet, p = 1.00).

Appetite-mediating hormones. Insulin, leptin, acylated ghrelin, and CCK concentrations measured before the standardized meal test indicated changes over time that were independent of dietary protein group. Specifically, fasting insulin concentrations were higher on SL day 7 and HA day 0 relative to HA day 21 and decreased during the day on SL day 7 and HA day 21, but not on HA day 0 (Fig. 5A). Leptin concentrations decreased during the day at SL but not during HA days 0 and 21 and were higher at all time points on SL day 7 and HA day 0 relative to HA day 21 (Fig. 5B).

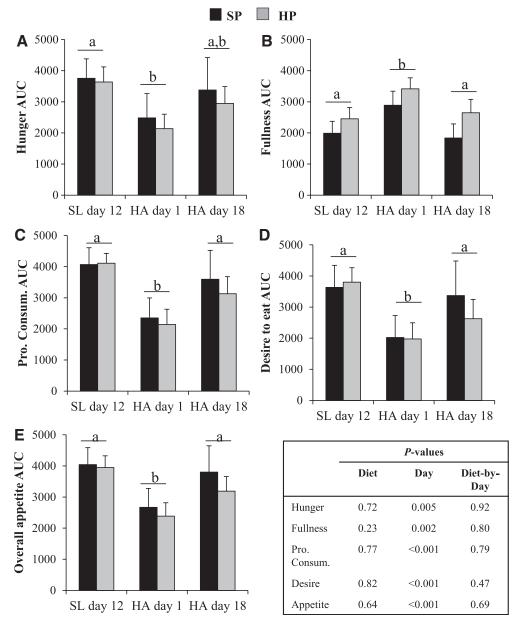
Acylated ghrelin concentrations measured during the day were affected by carbohydrate supplementation during exercise (Fig. 5C). In the carbohydrate group, mean acylated ghrelin concentrations were lower during HA days 0 and 21 relative to SL. In the placebo group, ghrelin concentrations did not differ across study days. A diet-by-day interaction was observed for aclyated ghrelin concentrations measured before the fixed meal (p = 0.01). However, post hoc testing indicated that the interaction was due to higher acylated ghrelin concentrations at HA day 0 in HP relative to SP (p=0.03). As consumption of the intervention diets did not start until HA day 1, this result was considered unrelated to

<sup>&</sup>lt;sup>1</sup>Generalized linear mixed model with Bonferroni corrections; subject included as a random factor, and diet, day, and their interaction as fixed factors. Main effect of day (p < 0.001); incidence higher on HA days 1 and 18 relative to SL.

Acute mountain sickness (AMS) was defined as a Lake Louise score >0.7 on the shortened version of the Environmental Symptoms Questionnaire (Beidleman et al., 2007).

<sup>&</sup>lt;sup>4-6</sup>Linear mixed model with Bonferroni corrections; subject included as a random factor, and diet, day, and their interaction as fixed factors. <sup>4</sup>Diet-by-day interaction, p < 0.05. <sup>a-c</sup>Values within a dietary protein group sharing a superscript letter are not significantly different within that group. <sup>5</sup>Main effect of group, p < 0.001. <sup>6</sup>Main effect of day (p < 0.001); lower acceptability on HA days 1 and 18 relative to SL. Overall meal acceptability rated from 1 (dislike extremely) to 9 (like extremely). Values are the mean of ratings at breakfast, lunch, and

<sup>\*</sup>Significantly different from the same day in the standard-protein group, p < 0.05.



**FIG. 2.** Appetite at SL and HA (4300 m) while consuming hypocaloric standard-protein (SP, 1.1 g/kg/d, n = 8) or higher protein (HP; 2.1 g/kg/d, n = 9) diets. AUC for perceived hunger (**A**), fullness (**B**), prospective consumption (Pro. Consum.,) (**C**), and desire to eat (**D**) measured at SL and after 1 and 18 days of living at HA. Metrics measured by visual analog scales administered hourly (0700–1800) and after meals. Overall appetite (**E**) was computed from hunger, fullness, prospective consumption, and desire to eat ratings. (**A**–**E**) Bars are mean  $\pm$  SEM. Effects of diet, day, and their interaction tested using linear mixed models with subject included as a random factor and Bonferroni corrections. Superscripts indicate main effect of day, p < 0.05. Bars not sharing a superscript letter are significantly different, p < 0.05. AUC, area under the curve calculated from ground.

the study diets. CCK concentrations decreased during the day on SL day 7 and HA day 21, but not HA day 0 (Fig. 5D).

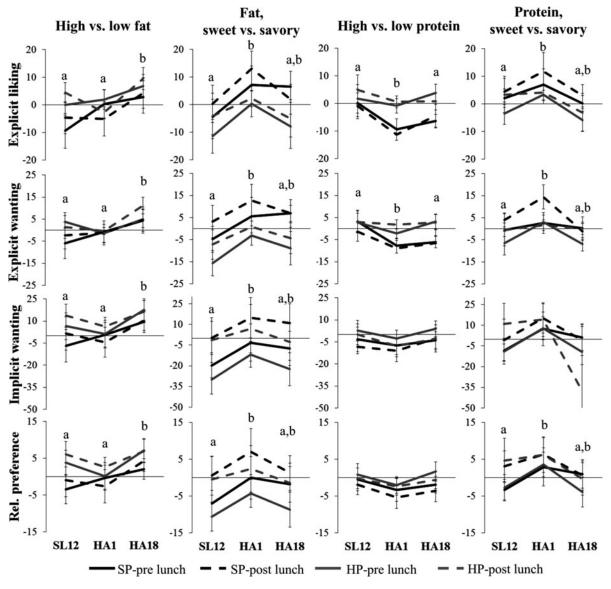
During the standardized meal test, CCK concentrations increased and were higher throughout the test during HA day 0 relative to SL day 7 and HA day 21 (Fig. 6A). Acylated ghrelin concentrations decreased following the standardized meal test and were higher at SL relative to HA (Fig. 6B).

#### **Discussion**

The primary finding of this study was that dietary protein intake did not impact appetite during energy deficit at HA.

However, acute HA exposure was associated with appetite suppression concomitant to elevated concentrations of the anorexigenic hormones insulin, leptin, and CCK, depressed concentrations of the orexigenic hormone acylated ghrelin, and preferences for sweet and low-protein foods. Although anorexigenic hormone concentrations did not remain elevated following acclimatization and weight loss, appetite and acylated ghrelin concentrations remained depressed while food preferences shifted toward high fat foods.

To our knowledge, this study is the first to examine differential effects of controlled standard- and higher protein hypocaloric diets on appetite regulation at HA. Contrary to



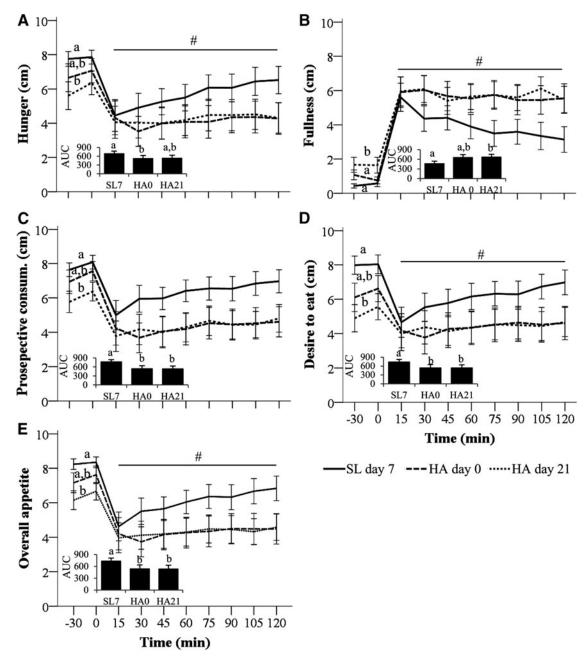
**FIG. 3.** Food preferences at SL and HA (4300 m) while consuming hypocaloric standard-protein (SP, 1.1 g/kg/d, n=8) or higher protein (HP; 2.1 g/kg/d, n=9) diets. Food preferences were measured before and after lunch on SL day 12 and after 1 and 18 days of living at HA using the Leeds Food Preference Questionnaire. Positive values indicate preference for high fat/ protein and sweet foods, negative values indicate preference for low fat/protein and savory foods. Values are mean  $\pm$  SEM. Effects of diet, day, and their interaction tested using repeated measures ANOVA. Superscripts indicate main effect of day, p < 0.05. Values not sharing a superscript letter are significantly different, p < 0.05.

our hypothesis, protein intake did not significantly impact appetite at SL or during HA exposure (Fig. 2). Of note, between-group differences in overall appetite following acclimatization and weight loss were sufficiently large to be considered of practical relevance if statistically significant (Blundell et al., 2010). Unfortunately, logistical constraints prevented obtaining a larger sample size, and one limitation of the study is limited statistical power due to the small sample sizes within dietary protein groups.

Additional factors may also have decreased the ability to detect significant effects of protein intake on appetite. First, between-group differences in protein intake ranged from 8 to 19 g/meal, which is less than the 30 g/meal difference others have suggested as a threshold for observing satiety-enhancing effects of higher protein meals (Paddon-Jones and Leidy, 2014). Second, the proportion of total energy intake

derived from liquids was higher in HP relative to SP as a result of the beverage used to increase protein intake, and liquids have been shown to elicit weaker satiety effects than solid foods (Leidy et al., 2011). Given these limitations, the hypothesis that higher protein diets could be counterproductive for maintaining fat-free mass at HA by exacerbating HA anorexia cannot be conclusively refuted and warrants further investigation.

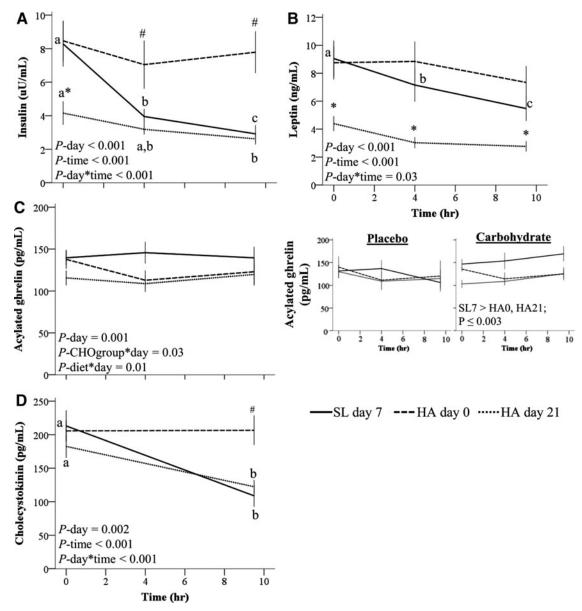
A second aim of this study was to explore mechanisms underlying acute HA anorexia and their persistence. Several recent studies have likewise aimed to characterize appetitemediating hormone responses to acute and chronic hypoxia (Debevec et al., 2014, 2016; Mekjavic et al., 2016), but were conducted at lower simulated altitudes in normobaric hypoxia. That environment may elicit different physiological responses than the hypobaric hypoxia at 4300 m studied in



**FIG. 4.** Appetite response to a standardized meal at SL and HA (4300 m). Perceived hunger (**A**), fullness (**B**), prospective consumption (**C**), and desire to eat (**D**) measured during a standardized meal test at sea level (SL7), on the first day of HA exposure (HA0) and after 21 days of living at HA (HA21). Overall appetite (**E**) was computed from hunger, fullness, prospective consumption, and desire to eat ratings. Values and bars (AUC) are mean  $\pm$  SEM. Effects of diet (standard- or higher protein), carbohydrate group (carbohydrate beverage or placebo), day, and their interactions tested using linear mixed models with subject included as a random factor and Bonferroni corrections. Diet and carbohydrate groups had no effects on any outcome. Premeal values (0 minutes) and bars not sharing a superscript are significantly different (main effect of day, p < 0.05). #Slopes from 15 to 120 minutes on SL day 7 are significantly different from those on HA days 0 and 21 (main effect of day,  $p \le 0.01$ ).

this investigation (Millet et al., 2012). Furthermore, in the present study, we tightly controlled physical activity, climate, and diet, factors which may have confounded several previous field studies and contributed to inconsistencies in observations (Sierra-Johnson et al., 2008; Debevec, 2017). However, the absence of a SL control group is a limitation of the present study that prevents establishing causal effects of HA on study outcomes. Although this limitation is less likely

to impact study outcomes measured during acute HA, it does preclude definitively separating the impact of acclimatization to HA on study outcomes from factors such as weight loss, diet monotony, and living in a confined environment. Potential confounding due to weight loss is particularly important to consider because weight loss independently influences appetite, food preferences, and appetite-mediating hormones (Maclean et al., 2011; Sumithran and Proietto,



**FIG. 5.** Appetite-mediating hormones at HA (4300 m). (**A**) Insulin (n=17;  $\log_{10}$ -transformed for analysis), (**B**) leptin (n=16), (**C**) acylated ghrelin (n=17), and (**D**) cholecystokinin (n=17) measured at SL, on the first day of HA exposure and after 21 days of living at HA. Volunteers consumed a hypocaloric standard-protein (1.1 g/kg/d, n=8) or higher protein (2.1 g/kg/d, n=9) diet while living at HA. Values are mean  $\pm$  SEM. Effects of diet group (standard- or higher protein), carbohydrate group (carbohydrate beverage or placebo), day, time, and all interactions not including those between diet group and carbohydrate group were tested using linear mixed models with subject included as a random factor and Bonferroni corrections. Only significant effects (p < 0.05) from the parsimonious model are listed. Within a study day, values not sharing a superscript letter are significantly different, p < 0.05. \*Different from SL day 7 and HA day 0,  $p \le 0.03$ . \*Different from SL day 7 and HA day 21,  $p \le 0.03$ . CHOgroup, carbohydrate supplementation during exercise at 4.5 hours (145 g or 0 g).

2013). As such, observed changes following acclimatization must be interpreted cautiously and within the context of the known effects of weight loss on study outcomes.

Appetite suppression during acute HA exposure is a well-established response (Hamad and Travis, 2006; Westerterp and Kayser, 2006) and was supported in the present study (Figs. 2 and 4). While previous studies report that appetite remains suppressed at very high altitudes (>5000 m) for several weeks (Rose et al., 1988; Westerterp-Plantenga et al., 1999), others suggest that appetite suppression may be alleviated during chronic (>7 days) exposure to altitudes below

5000 m, at least under normobaric hypoxic conditions [3000–4000 m (Debevec et al., 2014, 2016; Mekjavic et al., 2016)]. In the present study, appetite was lower during the meal test on HA day 21 relative to SL and did not differ from SL on HA day 18 despite substantial weight loss. It is well established that weight loss stimulates compensatory mechanisms which act to stimulate appetite and restore body weight (Maclean et al., 2011; Sumithran and Proietto, 2013). Therefore, as we are not aware of any reports demonstrating weight regain in people sojourning at >4000 m for ≥3 weeks, these findings suggest that normal physiologic adaptations to weight loss

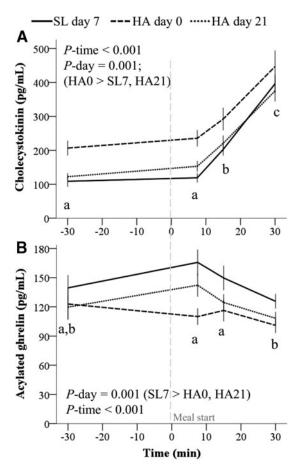


FIG. 6. Appetite-mediating hormone responses to a standardized meal at HA (4300 m). (A) Cholecystokinin (n = 17) and (B) acylated ghrelin (n = 17) responses to a fixed portion meal measured at SL, on the first day of HA exposure and after 21 days of living at HA. Volunteers consumed a hypocaloric standard-protein (1.1 g/kg/d, n=8) or higher protein (2.1 g/kg/d, n=9) diet while living at HA. Values are mean ± SEM. Effects of diet group (standard- or higher protein), carbohydrate group (carbohydrate beverage or placebo), day, time, and all interactions not including those between diet group and carbohydrate group were tested using linear mixed models with subject included as a random factor and Bonferroni corrections. Only significant effects (p < 0.05) from the parsimonious model are listed. Within a study day, values not sharing a superscript letter are significantly different, p < 0.05.

that stimulate weight regain may be blunted and persistent appetite suppression may be a relevant concern during sojourn at  $\geq$ 4300 m for several weeks. Studies with appropriate control groups are needed to verify this hypothesis.

Our findings suggest, but do not prove, that the orexigenic hormone ghrelin may be one factor in the persistence of HA anorexia. Previous studies have reported that total ghrelin concentrations are transiently suppressed in response to hypoxia (Shukla et al., 2005; Riepl et al., 2012; Debevec et al., 2014, 2016; Mekjavic et al., 2016). However, the relevance of these findings to appetite regulation is unclear because the appetite-stimulating effects of ghrelin require posttranslational acylation (Lim et al., 2010), and acylated ghrelin comprises only a small variable fraction of total ghrelin concentrations (Liu et al., 2008). Previous studies measuring acylated ghrelin responses to HA have reported

suppressed concentrations during 5–7 hours of exposure to normobaric hypoxia simulating altitudes of 3500–4000 m (Wasse et al., 2012; Matu et al., 2017a, 2017b), but not <3000 m (Morishima and Goto, 2016), and suppressed concentrations during a 12-day trek to 5140 m (Matu et al., 2017a, 2017b). Our findings are consistent with and extend those results by demonstrating that the suppression of acylated ghrelin concentrations upon acute HA exposure may be sustained for up to 21 days (Figs. 5C and 6B). Importantly, this observation is not likely confounded by weight loss as weight loss at SL is associated with increased acylated ghrelin concentrations (Martins et al., 2010).

Whether the 12%–22% lower mean acylated ghrelin concentrations observed at HA relative to SL in our study would be sufficient to cause changes in energy intake had volunteers been given free access to food was not assessed. However, Wasse et al. (2012) reported an 18% reduction in acylated ghrelin concentrations and 31% reduction in voluntary energy intake in response to 7-hour exposure to hypoxia simulating 4000 m altitude, while Matu et al. (2017a, 2017b) recently reported a  $\sim 25\%$  reduction in both fasting acylated ghrelin concentrations and in voluntary energy intake during a 12-day trek to 5140 m. Although appetite regulation at HA is undoubtedly influenced by multiple factors, collectively these studies suggest, but do not prove, a role for acylated ghrelin in HA anorexia. Additional studies with appropriate controls are warranted to confirm the hypothesis that HA suppresses acylated ghrelin concentrations and that this effect meaningfully impacts energy intake.

Our findings suggest that elevations in insulin, leptin, and CCK occur concomitant to appetite suppression during acute HA exposure. Elevated insulin concentrations during initial HA exposure are consistent with previous reports of transient hypoxia-induced impairments in insulin sensitivity and glucose homeostasis (Barnholt et al., 2006; Woolcott et al., 2015). Leptin concentrations also reportedly increase during the initial 24 hours of hypoxia (Tschop et al., 1998; Snyder et al., 2008). However, whether this effect persists is controversial because of inconsistent findings across multiple studies (Zaccaria et al., 2004; Shukla et al., 2005; Barnholt et al., 2006; Vats et al., 2007; Debevec et al., 2014, 2016; Mekjavic et al., 2016) attributable to differences in the control of confounding factors (Raff et al., 2008; Sierra-Johnson et al., 2008; Debevec, 2017). In particular, differences in the timing of measurements may be critical as reductions in food intake due to HA anorexia would provide a strong stimulus to suppress leptin concentrations within 48–72 hours (Karl et al., 2016).

Although results from the present study support a transient elevation in insulin and leptin concentrations during HA sojourn (Fig. 5), both hormones decrease with fat mass loss. As such, similar to most previous studies (Zaccaria et al., 2004; Shukla et al., 2005; Vats et al., 2007; Riepl et al., 2012; Debevec et al., 2014, 2016; Mekjavic et al., 2016), the lack of a weight stable control group precludes isolating the independent effect of acclimatization to HA on insulin and leptin.

CCK is secreted during meals and limits meal size by stimulating satiation (Woods, 2009). We hypothesized that hypoxia may induce a faster rise in CCK concentrations during meals thereby accounting for the more rapid onset of satiation previously observed at HA (Westerterp-Plantenga et al., 1999). Although the data did not support a faster rise in CCK at HA (Fig. 6A), mean CCK concentrations were higher

relative to SL after 10 hours of exposure to HA (Fig. 6A, D). However, this difference did not occur concomitant to increased fullness immediately following the standard test meal where the effects of CCK on satiation would be expected to occur. That observation, in combination with previous studies reporting decreases [3454 m, <72 hours; (Riepl et al., 2012)], no changes [4559 m, two and four days; (Aeberli et al., 2013)], and increases [20-day trek to 5100 m (Bailey et al., 2000)] in CCK concentrations at HA, indicates that additional research is needed to clarify the role of CCK, if any, in HA anorexia.

A potential strategy for overriding physiologic signals contributing to HA anorexia may be to provide highly palatable foods that exploit hedonic (i.e., pleasure and reward) pathways impacting eating behavior (Berthoud, 2006). Presently, the effects of hypoxia on food hedonics and subsequent food intake are unclear as previous reports on changes in taste perception and food hedonics (Premavalli et al., 2009; Aeberli et al., 2013) and self-selected macronutrient intake (Boyer and Blume, 1984; Rose et al., 1988; Reynolds et al., 1998; Westerterp-Plantenga et al., 1999; Bailey et al., 2004; Aeberli et al., 2013) at HA are inconsistent.

In the present study, HA exposure was acutely associated with a preference for sweet over savory foods and liking of low-protein over higher protein foods suggesting that sweet, low-protein foods may encourage eating during acute HA. The subsequent transition to a preference for high-fat foods suggests that a high-fat diet may have greater utility for promoting food intake following acclimatization. However, these hypotheses require confirmation as the design of this study precludes determining whether observed changes would have influenced ad libitum food intake, and factors such as diet monotony, limited food variety, and weight loss may have also impacted results. Nonetheless, our findings extend those of Aeberli et al. (2013) who used the same food preference assessment method to demonstrate that perceived liking of high-fat foods increased and was positively associated with ad libitum energy intake during a four day sojourn at HA (4559 m). One implication of these findings is that diet recommendations for HA sojourn may differ according to the duration of sojourn if the aim is to leverage hedonic mechanisms to promote energy intake and minimize weight loss.

Finally, while the present findings suggest that high protein and savory foods may not be preferred upon initial HA exposure, dietary protein intake within the levels studied did not appear to differentially influence food preferences. Using the same food preference assessment methodology, Griffioen-Roose et al. (2012) reported that a 14-day low-protein (0.5 g/kg/d) diet stimulated a compensatory response characterized by an increased drive to consume protein and protein intake. That food preferences were not influenced by dietary protein intake in the present study suggests that the protein content of the standard-protein diet was sufficient to prevent this response. It is also acknowledged that an increased drive to consume protein when protein intake is low, known as the protein leverage hypothesis, has not been observed in all studies (Martens et al., 2013).

# Conclusion

Taken together, these findings suggest that maximizing the effectiveness of dietary strategies for mitigating weight loss at HA may require overriding physiologic signals that sup-

press appetite, accommodating food preferences, and adjusting diets to both the duration and elevation of the sojourn. These observations in combination with the absence of a SL control group highlight the need for additional research with appropriate controls to establish causal effects of prolonged HA exposure on appetite, appetite-mediating hormones, and food preferences. Contrary to our hypothesis, a higher protein diet did not exacerbate HA anorexia in the present study. However, study limitations preclude definitive conclusions, and the potential for the satiating properties of protein to reduce *ad libitum* food intake at HA and the possible incongruence of higher protein diets with food preferences at HA warrant further investigation.

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#### **Disclaimers**

The views, opinions, and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy, or decision, unless so designated by other official documentation. Citation of trade names in this report does not constitute an official Department of the Army endorsement or approval of the use of such commercial items.

# **Author Disclosure Statement**

No authors report a conflict of interest.

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