

Draft Genome Sequence of *Oenococcus oeni* Strain X₂L (CRL1947), Isolated from Red Wine of Northwest Argentina

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We report the draft genome sequence of *Oenococcus oeni* strain X₂L, a potential starter culture of malolactic fermentation, isolated from Malbec wine of Argentina. Genes encoding for enzymes involved in the metabolism of malate, citrate, and nitrogen compounds, as well as aroma compounds, were found in this genome, showing its ability to improve the sensorial characteristics of wines.

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Oenococcus oeni is the best-adapted wine lactic acid bacterium and is almost exclusively used for the induction of malolactic fermentation (MLF), the conversion of malic acid into lactic acid during the wine-making process (1). MLF is a critical step to improve the quality of wine because acidity decreases, microbiological stability increases, and sensory characteristics are enhanced (2, 3). However, the harsh wine conditions represent a challenge to the survival and MLF of *O. oeni*. Therefore, a better understanding of the molecular mechanisms related to the stress tolerance of *O. oeni* is important for the selection of strains as culture starters (4, 5). Genome sequences of several *O. oeni* strains are available in the GenBank database, with the *O. oeni* PSU-1 genome being the only complete sequence reported (6–8). Comparative studies show genetic variation among strains with differences in sugar utilization, amino acid and exopolysaccharide biosynthesis, bacteriophages, and plasmids presence. *O. oeni* X₂L (CRL1947; CERELA culture collection, Tucumán, Argentina) was isolated from Malbec red wine of northwest Argentina (9) and selected for its great malolactic activity in the fermentation of Malbec grape must (10). To our knowledge, *O. oeni* X₂L is the first published genome of a strain from Argentinian wines.

Total DNA of *O. oeni* X₂L was extracted according to the protocol described by Pospiech and Neumann (11). The genome sequence was obtained using a whole-genome shotgun strategy (40-fold genome coverage) with an Ion Torrent personal genome machine (Life Technologies). Quality filtered reads were *in silico* assembled using the DNASTAR NGEN assembler, giving 114 large contigs. The draft genome sequence consists of 1,812,711 bp with an average GC content of 37.90%. Genome analysis was performed using the RAST server (12), and tRNA and rRNA genes were annotated by tRNAscan-SE (13) and RNAmmer (14), respectively. Results of RAST analysis showed that there are 247 subsystems represented in the chromosome, which represent only 45% of the sequences assigned. The genome sequence was annotated by the NCBI Prokaryotic Genomes Annotation Pipeline. A total of 1,458 coding DNA sequences, 46 tRNAs, and 6 rRNAs were predicted.

Among genes of biotechnological importance in the wine-making process, the complete cluster of genes related to metabolism of malic acid (*mleA*, *mleP*, *mleR*) and citrate (*citR*, *maeP*, *citC*, *citD*, *citE*, *citF*, *citX*, *citG*) were found in the *O. oeni* X₂L genome sequence. Genes encoding for esterases/lipases and glucosidases were also identified. In addition, genes encoding for enzymes involved in the metabolism of nitrogen compounds such as proteases, carboxypeptidases, aminopeptidases, and dipeptidases were detected, whereas genes related to biogenic amines synthesis (histamine, tyramine) were not found, although genes of putrescine transport were identified. The genes predicted through genome analysis showed the genetic potential of *O. oeni* X₂L as a starter culture for MLF and its ability to improve sensorial characteristics of red wines.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number [JROK00000000](https://www.ncbi.nlm.nih.gov/nuclink/JROK00000000). The version described in this paper is version [JROK01000000](https://www.ncbi.nlm.nih.gov/nuclink/JROK01000000).

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REFERENCES

- Liu SQ. 2002. Malolactic fermentation in wine—beyond deacidification. *J Appl Microbiol* 92:589–601. <http://dx.doi.org/10.1046/j.1365-2672.2002.01589.x>.
- Bartowsky EJ, Borneman AR. 2011. Genomic variations of *Oenococcus oeni* strains and the potential to impact on malolactic fermentation and aroma compounds in wine. *Appl Microbiol Biotechnol* 92:441–447. <http://dx.doi.org/10.1007/s00253-011-3546-2>.
- Lonvaud-Funel A. 1999. Lactic acid bacteria in the quality improvement and depreciation of wine. *Antonie van Leeuwenhoek* 76:317–331. <http://dx.doi.org/10.1023/A:1002088931106>.
- Mohedano ML, Russo P, de los Rios V, Capozzi V, Spano G, López P. 2014. A partial proteome reference map of the wine lactic acid bacteria

- Oenococcus oeni* ATCC BAA-1163. *Open Biol* 4:130154. <http://dx.doi.org/10.1098/rsob.130154>.
5. Cappello MS, De Domenico S, Logrieco A, Zapparoli G. 2014. Bio-molecular characterisation of indigenous *Oenococcus oeni* strains from Negroamaro wine. *Food Microbiol* 42:142–148. <http://dx.doi.org/10.1016/j.fm.2014.02.004>.
 6. Mills DA, Rawsthorne H, Parker C, Tamir D, Makarova K. 2005. Genomic analysis of *Oenococcus oeni* PSU-1 and its relevance to winemaking. *FEMS Microbiol Rev* 29:465–475. <http://dx.doi.org/10.1016/j.fm.2005.04.011>.
 7. Borneman AR, McCarthy JM, Chambers PJ, Bartowsky EJ. 2012. Comparative analysis of the *Oenococcus oeni* pan genome reveals genetic diversity in industrially-relevant pathways. *BMC Genomics* 13:373. <http://dx.doi.org/10.1186/1471-2164-13-373>.
 8. Lamontanara A, Orrù L, Cattivelli L, Russo P, Spano G, Capozzi V. 2014. Genome sequence of *Oenococcus oeni* OM27, the first fully assembled genome of a strain isolated from an Italian wine. *Genome Announc* 2(4):e00658-14. <http://dx.doi.org/10.1128/genomeA.00658-14>.
 9. Strasser de Saad AM, Manca de Nadra MC. 1987. Isolation and identification of the lactic acid bacteria from Cafayate (Argentina) wines. *Microbiol Alim Nutr* 5:45–49.
 10. Mendoza LM, Merin MG, Morata VI, Fariás ME. 2011. Characterization of wines produced by mixed culture of autochthonous yeasts and *Oenococcus oeni* from the Northwest region of Argentina. *J Ind Microbiol Biotechnol* 38:1777–1785. <http://dx.doi.org/10.1007/s10295-011-0964-1>.
 11. Pospiech A, Neumann B. 1995. A versatile quick-prep of genomic DNA from Gram-positive bacteria. *Trends Genet* 11:217–218. [http://dx.doi.org/10.1016/S0168-9525\(00\)89052-6](http://dx.doi.org/10.1016/S0168-9525(00)89052-6).
 12. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. *BMC Genomics* 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.
 13. Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res* 25:955–964. <http://dx.doi.org/10.1093/nar/25.5.0955>.
 14. Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, Ussery DW. 2007. RNAMmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res* 35:3100–3108. <http://dx.doi.org/10.1093/nar/gkm160>.