



Neurotoxicity of Ammonia

Simo S. Oja¹ · Pirjo Saransaari¹ · Esa R. Korpi²Received: 16 March 2016 / Revised: 15 July 2016 / Accepted: 21 July 2016 / Published online: 28 July 2016
© Springer Science+Business Media New York 2016

Abstract Abnormal liver function has dramatic effects on brain functions. Hyperammonemia interferes profoundly with brain metabolism, astrocyte volume regulation, and in particular mitochondrial functions. Gene expression in the brain and excitatory and inhibitory neurotransmission circuits are also affected. Experiments with a number of pertinent animal models have revealed several potential mechanisms which could underlie the pathological phenomena occurring in hepatic encephalopathy.

Keywords Hepatic encephalopathy · Astrocytes · Mitochondria · *N*-methyl-D-aspartate receptors · Reactive oxygen species · Glutamine-glutamate cycle

Background

Ammonia is produced in the body by intermediary amino acid metabolism or arises from the actions of intestinal bacteria. In human adults approximately 1000 mmol (17 g) of ammonia is produced daily [1]. A part of this is reutilized in biosynthesis, while the remainder is waste and neurotoxic. Its normal concentration in the portal blood varies from 300 to 600 μM , but in the blood leaving the liver the concentration is reduced to 20–60 μM . The liver thus occupies a central position in the regulation of ammonia levels in the

organism. The failure of liver functions in hepatic cirrhosis or for other reasons may thus result in an uncontrolled increase in levels of ammonia in the circulating blood. Ammonia at high concentrations penetrates from the blood into practically all organs. Although the brain is partially protected by the blood–brain barrier from toxic agents such as ammonia, excessive amounts of ammonia can pass into the brain, constituting a principal factor in the syndrome of hepatic encephalopathy. In patients, hepatic encephalopathy may result from acute liver failure or portal-systemic bypass with no intrinsic hepatocellular disease, or may be associated with cirrhosis and portal hypertension [2], all involving enormous costs to society. Ammonia concentrations can be experimentally elevated in a variety of ways, e.g., by administration of hepatotoxins as Professor Jan Albrecht has done in his many studies on ammonia toxicity, by peripheral administration of massive doses of ammonium chloride, or by directing the blood flow from the alimentary canal by end-to-side portacaval anastomosis, thus by-passing the liver. In addition to this, inborn errors in the urea cycle may lead to congenital hyperammonemia.

The brain is much more susceptible to the deleterious effects of ammonium during development than in adulthood. The concentration of ammonia in the blood is higher in newborns than in adults [3]. In the brain the normal ammonia content also diminishes during maturation [4]. Depending on the extent of hyperammonemia and its duration, more or less serious irreversible damage is caused to the brain, leading to mental retardation [5]. Hyperammonemia can provoke irreversible damage to the developing central nervous system, which leads to cortical atrophy, ventricular enlargement and demyelination, responsible for cognitive impairment, seizures and cerebral palsy [6]. An increased exposure to ammonia during the prenatal and lactation periods has been shown to cause long-lasting impairment of

✉ Simo S. Oja
simo.oja@uta.fi

¹ Medical School, 33014 University of Tampere, Finland

² Department of Pharmacology, Faculty of Medicine, University of Helsinki, Helsinki, Finland

N-methyl-*D*-aspartate (NMDA) receptor functions [7]. The disruption of energy metabolism by ammonia and disturbances in axonal growth during development could also be factors contributing to mental retardation [8].

Increased accumulation of ammonia in the brain due to liver dysfunction is a major factor in the pathogenesis of hepatic encephalopathy [9]. Hyperammonemia apparently affects brain functions by several mechanisms. It blocks chloride efflux from postsynaptic neurons [10], causes depression of synaptic transmission [11], inhibits neuron-astrocyte trafficking of glutamate and affects postsynaptic glutamate receptors [12]. The firing of glutamatergic neurons in the CA1 region of the hippocampus evoked by applied glutamate is then abolished by ammonia. On the other hand, glutamate exocytosis is evoked by ammonia in cultured rat astrocytes [13]. Ammonia also affects other neurotransmitter systems in addition to the glutamatergic. The synthesis of histamine, serotonin, dopamine and noradrenaline in the brain is altered by hyperammonemia [14]. For example, ammonium chloride directly administered to the rat striatum via reversed microdialysis evokes a prompt accumulation of dopamine in the microdialysates [15]. Furthermore, ammonia influences the passage of different molecules across the blood–brain barrier. It modulates the transcellular passage of low- to medium-size molecules by affecting their carriers located at this barrier [16]. Ammonia also inhibits GABA uptake and enhances its release [17]. On the other hand, ammonia has been shown to stimulate glutamine uptake into non-synaptic mitochondria isolated from rat cerebral hemispheres [18]. We here briefly review these proposed mechanisms.

Astrocyte Swelling and Shrinking

Ammonia induces astrocytic swelling [19]. Astrocyte swelling is believed to be a key component in the cytotoxic brain edema [20] associated with acute liver failure and the increase in intracranial pressure and eventually brain herniation which is often the cause of death in patients with hepatic encephalopathy [21]. Elevated ammonia has also been shown to produce astrocytic swelling, tissue swelling and neuronal toxicity in organotypic slice cultures of cerebral tissue [22]. Astrocyte swelling has thus been generally assumed to be the key factor in the generation of ammonia toxicity and the increase in intracranial pressure leading to brain herniation and death [19, 23].

Glutamine has been thought to be the principal factor in ammonia detoxification. More recently, however, glutamine has been considered to mediate ammonia toxicity when in excess [24, 25]. In keeping with this assumption the cerebral glutamine content has been shown to correlate positively with the grade of hepatic encephalopathy [26]. Glutamine

may impair the mitochondrial function in astrocytes secondarily to its excessive accumulation in them. Glutamine has been shown to increase mitochondrial permeability [27]. Ammonia itself does not induce mitochondrial swelling, even though it increases glutamate uptake in mitochondria [18].

It has been demonstrated that activation of the neuronal NKCC1 ($\text{Na}^+ \text{K}^+ 2\text{Cl}^-$ -cotransporter, encoded by the *SLC12A2* gene) is involved in the astrocyte swelling induced by ammonia and in brain edema [28]. Preclinical results in non-anesthetized mouse model of childhood epilepsy (ornithine transcarbamylase-deficient mice), which are contradictory to earlier conceptions as to the critical role of astrocyte swelling, have recently been published. According to them an acute increase in extracellular ammonia does not lead primarily to astrocyte swelling but rather to unbalanced astrocyte buffering of potassium ions, with increases extracellular potassium overactivating the NKCC1, which in turn compromises inhibitory neurotransmission in the cerebral cortex and depolarizes the neuronal GABA reversal potential (E_{GABA}) [29]. Consequently, intracellular chloride is increased in neurons and the main fast-acting inhibitory system, GABA_A receptor-mediated inhibition, is blunted, leading to a myoclonic seizure phenotype in this mouse model. This abnormal GABAergic excitation might be exacerbated by the known increase in extracellular GABA due to ammonia-induced inhibition of GABA uptake and enhancement of GABA release [17], and by increased synthesis of GABA by glutamate decarboxylase due to increased glutamate levels [30]. The potassium uptake mechanisms by Na^+/K^+ -ATPase are then saturated by increased ammonium ion levels. According to these studies increased ammonia would thus seem not to lead to astrocyte swelling but rather to transient astrocyte shrinking. Astrocyte swelling or brain edema only occurs in the terminal stages of ammonia toxicity [31]. Importantly, the clinically used diuretic inhibitor of NKCC1, bumetanide, has been seen to block the effects of acute ammonia on GABAergic neurotransmission and prevent seizures in the mouse model [29]. It remains to be established whether these acute mechanisms show adaptation during more modest and prolonged hyperammonemia.

Molecular Mechanisms Involved

At present, several factors and pathways have been surmised to be associated in ammonia toxicity [32]. To date, these include oxidative and nitrosative stress [9, 33], adverse alterations in the glutamate-glutamine cycle, changes in mitochondrial permeability transition (MPT) [34], effects on neural transmission, activation of mitogen-activated protein kinases (MAPKs) and effects on the transcription factor nuclear factor-kappa B (NF- κ B) [35]. Such effects

strongly suggest the involvement of phosphorylation of MAPKs in the mechanism of ammonia-induced astrocyte dysfunction associated with ammonia neurotoxicity [36]. Blockade of the MPT, MAPKs and NF- κ B has been shown to reduce the extent of astrocyte swelling [35]. The cytoplasmic level of tumor suppressor phosphoprotein p53 is also increased in acute toxicity with large ammonium doses [37, 38]. These above effectors generate additional reactive oxygen–nitrogen species, phosphorylate various proteins and transcription factors, and cause mitochondrial dysfunction. Astrocytes exposed to ammonia also exhibit a reduction in intra- and extracellular levels of thrombospondin-1, an astrocytic factor involved in the maintenance of synaptic integrity. A defective release of thrombospondin-1 may impair synaptic integrity in chronic hepatic encephalopathy [39].

Ammonia markedly enhances the generation of reactive oxygen and nitrogen species (ROS and RNS), including the highly toxic peroxyxynitrite, in astrocytes [40, 41]. The production of hydroxyl radicals is also increased *in vivo* in the rat striatum upon microdialysis of ammonium chloride [42]. The generation of reactive oxygen and nitrogen species, in turn, induces protein tyrosine nitration [43], lipid peroxidation [44], S-nitrosylation of cysteine residues in proteins, and nucleic acid oxidation [45]. Glutathione is the major antioxidant in the brain and could counteract the harmful effects of reactive oxygen and nitrogen species. The synthesis of glutathione in cultured astrocytes [46] and the uptake of its precursor cysteine [47] are fomented by ammonia. In line with these observations, the glutathione content is increased in the brain extracellular spaces after administration of ammonium chloride [48].

Elevated concentrations of ammonia induce the formation of free radicals in astrocytes, which is associated with the synthesis of glutamine [40]. When the glutamine transport into cultured astrocytes is prevented, the generation of ammonia-induced reactive oxygen species production, cell swelling, MPT, and loss of ATP are completely blocked or significantly attenuated [49]. These findings clearly implicate mitochondrial glutamine transport in the mechanism of ammonia neurotoxicity. It has been proposed that the glutamine-derived ammonia within mitochondria interferes with mitochondrial functions, giving rise to excessive production of free radicals and induction of MPT, two phenomena which bring about astrocyte dysfunction, including cell swelling [50]. Glutamine thus induces oxidative stress and MPT, being critical in the development of astrocyte swelling in hyperammonemia [23].

There is strong evidence to indicate that oxidative stress is involved in the induction of MPT by ammonia, and that oxidative stress and the subsequent induction of MPT contribute to the pathogenesis of hepatic encephalopathy and other hyperammonemic disorders [51, 52]. Altered

bioenergetics and oxidative stress appear to be critical factors in this pathogenesis [52]. MPT seems thus to represent an important component in the pathogenesis of hepatic encephalopathy and other hyperammonemic states [53]. In line with these speculations, direct application of glutamine to cultured astrocytes increases free radical production and induces MPT [54]. Only astrocytes, but not neurons, generate free radicals following glutamine exposure [55].

Brain glucose consumption is diminished in portacaval shunt-induced [56] and thioacetamide-induced [57] hyperammonemic states. Hyperammonemia can induce cerebral energy failure by several mechanisms [58, 59]. The high concentration of ammonia interferes with oxidative metabolism in the brain through an inhibitory effect on the tricarboxylic acid cycle [60]. Ammonia also induces ATP depletion due to activation of Na⁺/K⁺-ATPase, which, in turn, is a consequence of decreased phosphorylation by protein kinase C (PKC) [61]. Ammonium chloride also affects energy metabolism by increasing the neuronal tricarboxylic acid cycle activity and switching it in astrocytes towards glutamine synthesis [62]. At variance with this assumption the excess ammonia has been reported to interfere with brain energy metabolism by inhibiting the tricarboxylic acid cycle and this inhibition may result in depletion of ATP in the brain cells [63]. Moreover, there are also other controversial findings on this topic. It has been consistently reported that hepatic encephalopathy and concomitant hyperammonemia lead to reduced cerebral oxygen consumption. However, this may not be directly linked to an effect of ammonia but related to the fact that hepatic encephalopathy is always associated with reduced brain activity [64]. The whole-brain oxidative metabolism in patients with hepatic encephalopathy may not be due to malfunction of oxidative metabolism in astrocytes. The observed decline of brain oxidative metabolism may result from changes in neurons and their energy turnover [65].

One of the primary roles of astrocytes is to protect neurons against excitotoxicity by taking up excess ammonia and glutamate and converting them into glutamine via glutamine synthetase, which is located almost exclusively in astrocytes [66, 67]. Changes in the expression of this enzyme reflect changes in astroglial functions, hence also affecting neuronal functions [68]. Newly synthesized glutamine is transferred to neurons and hydrolyzed by glutaminase to glutamate [30]. In hepatic encephalopathy the expression of glutamate transporter (EEAT-2) is decreased, which impairs the cycling of glutamate–glutamine between astrocytes and neurons. Consequently, extracellular level of the main fast-acting excitatory neurotransmitter glutamate is increased, the NMDA receptor-mediated signaling activated, including RNS production, and tyrosine residues are nitrated. This sequence of events has been considered a cornerstone in the pathogenesis of hepatic encephalopathy [69].

Involvement of *N*-methyl-D-aspartate Receptors

Ionotropic NMDA receptors are involved in many functions in the central nervous system. The severity of the symptoms caused by hyperammonemia is positively correlated with the activation of NMDA receptors [70]. The acute neurotoxic effects of ammonia may thus be due mainly to overactivation of NMDA receptors, possibly potentiated by impaired control of their function by metabotropic glutamate receptors [71]. The sequence of events consists of increased extracellular glutamate stimulating NMDA receptors, which leads to increased intracellular Ca^{2+} and subsequent activation of NADPH oxidase (superoxide production, ROS) and NO synthase (NO production, RNS). Superoxide and NO can then promote the formation of peroxynitrite and protein tyrosine nitration. On the other hand, long-term exposure to ammonia of cultured cerebellar neurons impairs the glutamate-NO pathway in a dose- and time-dependent manner. The glutamate-induced formation of cGMP is reduced without effects on NO synthase [72].

The ammonia-induced swelling of rat cerebral cortical slices is significantly attenuated by NMDA receptor antagonists, inhibitors of the NO synthase, and taurine [73]. Ammonia treatment in vivo reduces synthesis of kynurenic acid, which is an endogenous, broad-spectrum antagonist of ionotropic glutamate receptors [74]. Inhibition of excitatory synaptic transmission by elevated brain ammonia has been assumed to underlie the central nervous system depression in hepatic encephalopathy [75]. Ammonia may stimulate the expression of inducible NO synthase in astrocytes, leading to excessive formation of NO, which in turn could trigger the formation of peroxynitrite in adjacent neurons, inducing their death [76]. The NMDA receptor antagonist dizocilpine (MK-801) blocks the ammonia-induced generation of reactive oxygen and nitrogen species in astrocytes [77]. On the other hand, administration of ammonium chloride has been reported to reduce the expression of two NMDA receptor subunits (GluN2A and GluN2B) in the rat hippocampus [78]. Ammonium chloride infusion into the rat striatum in vivo via a microdialysis probe increases glutamine, NO oxidation products and cGMP in the microdialysate [79]. Likewise, it activates NMDA receptors and foments the generation of hydroxyl radicals [42]. Ammonia also induces apoptosis as a result of a complex interplay of at least three signalling molecules: NO, PKC and NF- κ B. The NF- κ B is possibly involved in the induction of iNOS and the generation of toxic levels of NO in C6 glioma cells [80]. Figure 1 summarizes the main sequelae of ammonia neurotoxicity.

More recently, Cauli et al. [81, see for refs] have used cerebellar in vivo microdialysis to assess the mechanisms of ammonium-induced impairment of the glutamate-NMDAR-NO-cGMP pathway. NMDA-triggered citrulline and cGMP production was monitored in dialysates. The cGMP and

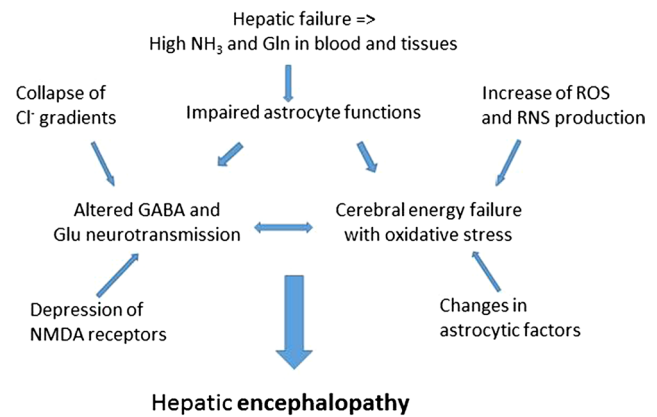


Fig. 1 Main factors responsible for hepatic encephalopathy, the most severe form of neurotoxicity induced by hyperammonemia

citrulline levels could be regulated by both NMDAR and GABA_AR activities, and importantly, the neurosteroid sensitivities of both receptor systems were altered depending on the increased ammonium levels. Several neurosteroids, for example allopregnanolone, tetrahydrodeoxycorticosterone and dehydroepiandrosterone sulphate, reduced the pathway, whereas pregnenolone sulphate enhanced it. The results were taken to indicate that certain neurosteroids might contribute to cognitive symptoms of hyperammonemia while others alleviate them [82, 83]. In line, Zorumski et al. [84, 85] have reported that 100 μM ammonium in vitro inhibits the induction of NMDAR-dependent long-term potentiation (LTP) via increased neurosteroid synthesis in hippocampal slices. The effect of ammonium could be blocked by finasteride, a selective inhibitor of 5 α -reductase needed for neurosteroid synthesis, and by GABA_AR blocker picrotoxin, as well as by L-carnitine. The key step in the neurosteroid synthesis is mediated by the mitochondrial 18 kDa translocator protein (TSPO, formerly known as peripheral benzodiazepine receptor), which enhances the uptake of the precursor cholesterol for the synthesis [86]. The TSPO is strongly upregulated in animal models of hyperammonemia and in patients with hepatic encephalopathy [87–91], which then increases neurosteroid levels [92, 93]. These interesting observations await for translation to clinical trials, particularly as finasteride has alleviated various symptoms in a model of thioacetamide-induced hepatic encephalopathy in rats [94].

How to Alleviate Ammonia Toxicity? Preclinical vs. Clinical?

The severity of symptoms in ammonia toxicology has been the impetus in the search for methods and compounds which could alleviate them, in addition to possible neurosteroid-related mechanisms (see above). Some

neuroprotective strategies such as the potential use of NMDA receptor antagonists, NO inhibitors, creatine and acetyl-L-carnitine have been suggested to counteract the toxic effects of ammonia [6]. L-Carnitine has been found to suppress ammonia-induced seizures and biochemical alterations in the brain in mice [95, 96]. L-Carnitine and its analogues thus have a potential to suppress ammonia neurotoxicity [97]. For example, treatment with acetyl-L-carnitine has preserved ATP in the brain, while lowering ammonia in the blood and brain less markedly [98]. By assuming that acute ammonia toxicity is mediated by activation of the NMDA receptors, the protective effect of L-carnitine against glutamate toxicity may result from its ability to increase the affinity of glutamate for the metabotropic receptors [96, 99]. Especially the mGluR5 receptors have been implicated [100]. In human patients in whom valproate has caused hepatic encephalopathy as a serious adverse effect, treatment with L-carnitine speeds up the decrease in ammonemia [101]. The inhibitor of NO synthase nitroarginine also attenuates acute ammonia toxicity and ammonia-induced alterations in brain energy metabolites by a mechanism which does not involve the activation of NMDA receptors [102].

Increased accumulation of cGMP in the rat striatum by intrastriatal infusions of ammonium chloride or NMDA has been virtually abolished by co-infusion of taurine [42]. This inhibitory compound has also attenuated the simultaneous accumulation of hydroxyl radicals. Similar co-infusion of the potent glycine site-specific NMDA receptor antagonist CGP 78608 ((1S)-1-[[[7-bromo-1,2,3,4-tetrahydro-2,3-dioxo-5-quinoxaliny]methyl]amino]ethylphosphonate) abolished ammonia-induced cGMP synthesis [103]. Melatonin and dimethylsulfoxide have also been found to lower the thioacetamide-induced increase in brain ammonia [104]. Dimethylsulfoxide has also significantly reduced the basal glutamine concentration in the rat striatum and attenuated the basal concentration of cGMP in microdialysates [80]. Kynurenic acid, an endogenous NMDA receptor antagonist with a high affinity towards its glycine site, may counter the over-activation or depression of glutamergic transmission observed at the different stages of hyperammonemia [105]. Glutathione can also counteract the generation of oxygen radicals provoked by ammonia. Upregulation of cystine uptake may contribute to this response [48]. Resveratrol, a polyphenol found in grapes and red wines, has also prevented ammonia toxicity by modulating oxidative stress and glial and inflammatory responses in astrocytes [106]. Inhibition of NKCC1 with bumetadine or more specific drugs, which also affect potassium regulation by astrocytes, may be a new and promising approach in the treatment of ammonia neurotoxicity [31].

Concluding Remark

Rapid advances in knowledge of the mechanisms involved in hyperammonemia-induced alterations in the brain are the basis for an understanding of the neurochemical, cellular, functional and structural effects caused by ammonia, hopefully leading to the invention of novel strategies in the treatment of hepatic encephalopathy.

References

- Walker V (2014) Ammonia metabolism and hyperammonemic disorders. *Adv Clin Chem* 67:73–150
- Ferenci P, Lockwood A, Mullen K, Tarter R, Weissenborn K, Blei AT (2002) Hepatic encephalopathy definition, nomenclature, diagnosis, and quantification: final report of the working party at the 11th World Congresses of Gastroenterology, Vienna, 1998. *Hepatology* 35:716–721
- Donn SM, Banagale RC (1984) Neonatal hyperammonemia. *Ped Rev* 5:203–208
- Oja SS, von Bonsdorff HA, Lindroos OF (1966) Ammonia content of developing brain. *Nature* 212:937–938
- Msall M, Batshaw ML, Suss R, Brusilow SW, Mellits ED (1984) Neurologic outcome in children with inborn errors of urea synthesis. Outcome of urea cycle enzymopathies. *N Engl J Med* 310:1500–1505
- Cagnon L, Braissant O (2007) Hyperammonemia-induced toxicity for the developing central nervous system. *Brain Res Rev* 56:183–197
- Miñana MD, Marcaida G, Grisolia S, Felipe V (1995) Prenatal exposure of rats to ammonia impairs NMDA receptor function and affords delayed protection against ammonia toxicity and glutamate neurotoxicity. *J Neuropathol Exp Neurol* 54:644–650
- Bachmann C (2002) Mechanisms of hyperammonemia. *Clin Chem Lab Med* 40:653–662
- Skowrońska M, Albrecht J (2013) Oxidative and nitrosative stress in ammonia neurotoxicity. *Neurochem Int* 82:731–737
- Raabe WA (1989) Neurophysiology of ammonia intoxication. In Butterworth RF, Layrargues RG (eds) *Hepatic encephalopathy: pathophysiology and treatment*, Humana Press, Clifton, pp 49–77
- Fan P, Lavoie J, Le NL, Szerb JC, Butterworth RF (1990) Neurochemical and electrophysiological studies on the inhibitory effect of ammonium ions on synaptic transmission in slices of rat hippocampus: evidence for a postsynaptic action. *Neuroscience* 37:327–334
- Butterworth RF (1993) Portal-systemic encephalopathy: a disorder of neuron-astrocytic metabolic trafficking. *Dev Neurosci* 15:313–319
- Görg B, Morwinsky A, Keitel V, Qvartskhava N, Schror K, Häussinger D (2010) Ammonia triggers exocytotic release of L-glutamate from cultured rat astrocytes. *Glia* 58:691–705
- Hawkins RA, Mans AM (1994) Brain metabolism in encephalopathy caused by hyperammonemia. *Adv Exp Med Biol* 368:11–21
- Anderzhanova E, Oja SS, Saransaari P, Albrecht J (2003) Changes in the striatal extracellular levels of dopamine and dihydroxyphenylacetic acid evoked by ammonia and N-methyl-D-aspartate: modulation by taurine. *Brain Res* 977:290–293
- Skowrońska M, Albrecht J (2012) Alterations of blood brain barrier function in hyperammonemia: an overview. *Neurotox Res* 21:236–244

17. Bender AS, Norenberg MD (2000) Effect of ammonia on GABA uptake and release in cultured astrocytes. *Neurochem Int* 36:389–395
18. Dolińska M, Hilgier W, Albrecht J (1996) Ammonia stimulates glutamine uptake to the cerebral non-synaptic mitochondria of the rat. *Neurosci Lett* 213:45–48
19. Norenberg MD, Baker I, Norenberg LO, Blicharska J, Bruce-Gregorios JH, Neary JT (1991) Ammonia-induced astrocyte swelling in primary culture. *Neurochem Res* 16:833–836
20. Blei AT (2005) The pathophysiology of brain edema in acute liver failure. *Neurochem Int* 47:71–77
21. Blei AT (2007) Brain edema in acute liver failure: can it be prevented? Can it be treated? *J Hepatol* 46:564–569
22. Back A, Tupper KY, Bai T, Chirananand P, Goldenberg FD, Frank JI, Brorson JR (2011) Ammonia-induced brain swelling and neurotoxicity in an organotypic slice model. *Neurol Res* 33:1100–1108
23. Norenberg MD, Rama Rao KV, Jayakumar AR (2005) Mechanisms of ammonia-induced astrocyte swelling. *Metab Brain Dis* 20:303–318
24. Cooper AJ (2001) Role of glutamine in cerebral nitrogen metabolism and ammonia neurotoxicity. *Ment Retard Dev Disabil Res Rev* 7:280–286
25. Albrecht J, Zielińska M, Norenberg MD (2010) Glutamine as a mediator of ammonia neurotoxicity: a critical reappraisal. *Biochem Pharmacol* 80:1303–1308
26. Laubenberger J, Häussing D, Bayer S, Gufler H, Hennig J, Langer M (1997) Proton magnetic resonance spectroscopy of the brain in symptomatic and asymptomatic patients with liver cirrhosis. *Gastroenterology* 112:1610–1616
27. Ziemińska E, Dolińska M, Lazarewicz JW, Albrecht J (2000) Induction of permeability transition and swelling of rat brain mitochondria by glutamine. *Neurotoxicology* 21:295–300
28. Jayakumar AR, Norenberg MD (2010) The Na-K-Cl co-transporter in astrocyte swelling. *Metab Brain Dis* 25:31–38
29. Rangroo Thrane V, Thrane AS, Wang F, Cotrina ML, Smith NA, Chen M, Xu Q, Kang N, Fujita T, Nagelhus EA, Nedergaard M (2013) Ammonia triggers neuronal disinhibition and seizures by impairing astrocyte potassium buffering. *Nat Med* 19:1643–1648
30. Albrecht J, Sidoryk-Węgrzynowicz M, Zielińska M, Aschner M (2010) Roles of glutamine in neurotransmission. *Neuron Glia Biol* 6:263–276
31. Eid T, Lee T-SW (2013) Reassessing the role of astrocytes in ammonia neurotoxicity. *Nat Med* 19:1572–1574
32. Norenberg MD, Rama Rao KV, Jayakumar AR (2009) Signaling factors in the mechanism of ammonia neurotoxicity. *Metab Brain Dis* 24:103–117
33. Norenberg MD (2003) Oxidative and nitrosative stress in ammonia neurotoxicity. *Hepatology* 37:245–248
34. Alvarez VM, Rama Rao KV, Brahmabhatt M, Norenberg MD (2011) Interaction between cytokines and ammonia in the mitochondrial permeability transition in cultured astrocytes. *J Neurosci Res* 89:2028–2040
35. Sinke AP, Jayakumar AR, Panickar KS, Moriyama MR, Pichili VBR, Norenberg MD (2008) NFκB in the mechanism of ammonia-induced astrocyte swelling in culture. *J Neurochem* 106:2302–2311
36. Jayakumar AR, Panickar KS, Murthy CK, Norenberg MD (2006) Oxidative stress and mitogen-activated protein kinase phosphorylation mediate ammonia-induced cell swelling and glutamate uptake inhibition in cultured astrocytes. *J Neurosci* 26:4774–4784
37. Kosenko E, Kaminsky Y, Solomadin I, Marov N, Venediktova N, Felipo V, Montoliu C (2007) Acute ammonia neurotoxicity in vivo involves increase in cytoplasmic protein P53 without alterations in other markers of apoptosis. *J Neurosci Res* 85:2491–2499
38. Panickar KS, Jayakumar AR, Rama Rao KV, Norenberg MD (2009) Ammonia-induced activation of p53 in cultured astrocytes: role in cell swelling and glutamate uptake. *Neurochem Int* 55:98–105
39. Jayakumar AR, Tong XY, Curtis KM, Ruiz-Cordero R, Sharmaladevi N, Abuzamel M, Johnstone J, Gaidosh G, Rama Rao KV, Norenberg MD (2014) Decreased astrocytic thrombospondin-1 secretion after chronic ammonia treatment reduces the level of synaptic proteins: in vitro and in vivo studies. *J Neurochem* 131:333–347
40. Murthy CR, Rama Rao KV, Bai G, Norenberg MD (2001) Ammonia-induced production of free radicals in primary cultures of rat astrocytes. *J Neurosci Res* 66:282–288
41. Skowrońska M, Zielińska M, Albrecht (2010) Stimulation of natriuretic peptide receptor C attenuates accumulation of reactive oxygen species and nitric oxide synthesis in ammonia-treated astrocytes. *J Neurochem* 115:1068–1076
42. Hilgier W, Anderzhanova E, Oja SS, Saransaari P, Albrecht J (2003) Taurine reduces ammonia- and N-methyl-D-aspartate-induced accumulation of cyclic GMP and hydroxyl radicals in microdialysates of the rat striatum. *Eur J Pharmacol* 468:21–25
43. Häussinger D, Görg B, Reinehr R, Schliess F (2005) Protein tyrosine nitration in hyperammonemia and hepatic encephalopathy. *Metab Brain Dis* 20:285–294
44. Swapna I, Sathya Sai Kumar KV, Murthy CR, Senthilkumar B (2006) Membrane alterations and fluidity changes in cerebral cortex during acute ammonia intoxication. *Neurotoxicology* 27:402–408
45. Görg B, Qvartskhava N, Keitel V, Bidmon HJ, Selbach O, Schliess F, Häussinger D (2008) Ammonia induces RNA oxidation in cultured astrocytes and brain in vivo. *Hepatology* 48:567–579
46. Murthy CR, Bender AS, Dombro RS, Bai G, Norenberg MD (2000) Elevation of glutathione levels by ammonium ions in primary cultures of rat astrocytes. *Neurochem Int* 37:255–268
47. Węgrzynowicz M, Hilgier W, Dybel A, Oja SS, Saransaari P, Albrecht J (2007) Upregulation of cerebral cortical glutathione synthesis by ammonia in vivo and in cultured glial cells: the role of cystine uptake. *Neurochem Int* 50:883–889
48. Hilgier W, Węgrzynowicz M, Ruszkiewicz J, Oja SS, Saransaari P, Albrecht J (2010) Direct exposure to ammonia and hyperammonemia increase the extracellular accumulation and degradation of astroglia-derived glutathione in the rat prefrontal cortex. *Toxicol Sci* 117:163–168
49. Pichili VBR, Rama Rao KV, Jayakumar AR, Norenberg MD (2007) Inhibition of glutamine transport into mitochondria protects astrocytes from ammonia toxicity. *Glia* 55:801–809
50. Albrecht J, Norenberg MD (2006) Glutamine: a Trojan horse in ammonia neurotoxicity. *Hepatology* 44:788–794
51. Norenberg MD, Jayakumar AR, Rama Rao KV (2004) Oxidative stress in the pathogenesis of hepatic encephalopathy. *Metab Brain Dis* 19:313–329
52. Rama Rao KV, Jayakumar AR, Norenberg MD (2005) Role of oxidative stress in the ammonia-induced mitochondrial permeability transition in cultured astrocytes. *Neurochem Int* 47:31–38
53. Rama Rao KV, Jayakumar AR, Norenberg MD (2003) Ammonia neurotoxicity: role of the mitochondrial permeability transition. *Metab Brain Dis* 18:113–127
54. Norenberg MD, Rama Rao KV, Jayakumar AR (2004) Ammonia neurotoxicity and the mitochondrial permeability transition. *J Bioenerg Biomembr* 36:303–307
55. Jayakumar AR, Rama Rao KV, Schousboe A, Norenberg MD (2004) Glutamine-induced free radical production in cultured astrocytes. *Glia* 46:296–301
56. DeJoseph MR, Hawkins RA (1991) Glucose consumption decreases throughout the brain only hours after portacaval shunting. *Am J Physiol* 260:E613–E619
57. Hilgier W, Beveniste H, Diemer NH, Albrecht J (1991) Decreased glucose utilization in discrete brain regions of rat in

- thioacetamide-induced hepatic encephalopathy as measured with [³H]-deoxyglucose. *Acta Neurol Scand* 83:353–355
58. Hindfelt B, Plum F, Duffy TE (1977) Effect of acute ammonia intoxication on cerebral metabolism in rats with portacaval shunts. *J Clin Invest* 59:386–396
 59. Therrien G, Giguère JF, Butterworth RF (1991) Increased cerebrospinal fluid lactate reflects deterioration of neurological status in experimental porta-systemic encephalopathy. *Metab Brain Dis* 6:225–231
 60. Haghighat N, McCandless DW (1997) Effect of ammonium chloride on energy metabolism of astrocytes and C6-glioma cells in vitro. *Metab Brain Dis* 12:287–298
 61. Felipo V, Kosenko E, Miñana MD, Marcaida G, Grisolia S (1994) Molecular mechanism of acute ammonia toxicity and of its prevention by L-carnitine. *Adv Exp Med Biol* 368:65–77
 62. Leke R, Bak LK, Anker M, Melo TM, Sørensen M, Keiding S, Vilstrup H, Ott P, Portela LV, Sonnewald U, Schousboe A, Waagepetersen HS (2011) Detoxification of ammonia in mouse cortical GABAergic cell cultures increases neuronal oxidative metabolism and reveals an emerging role for release of glucose-derived alanine. *Neurotox Res* 19:496–510
 63. Haghighat N, McCandless DW, Geraminegad P (2000) The effect of ammonium chloride on metabolism of primary neurons and neuroblastoma cells in vitro. *Metab Brain Dis* 15:151–162
 64. Schousboe A, Waagepetersen HS, Leke R, Bak LK (2014) Effects of hyperammonemia on brain energy metabolism: controversial findings in vivo and in vitro. *Metab Brain Dis* 29:913–917
 65. Iversen P, Mouridsen K, Hansen MB, Jensen SB, Sørensen M, Bak LK, Waagepetersen HS, Schousboe A, Ott P, Vilstrup H, Keiding S, Gjedde A (2014) Oxidative metabolism of astrocytes is not reduced in hepatic encephalopathy: a PET study with [¹¹C]acetate in humans. *Front Neurosci* 8:353
 66. Martinez-Hernandez A, Bell KP, Norenberg MD (1977) Glutamine synthetase: glial localization in brain. *Science* 195:1356–1358
 67. Rose CF, Verkhratsky A, Parpura V (2013) Astrocyte glutamine synthetase: pivotal in health and disease. *Biochem Soc Trans* 41:1518–1524
 68. Suárez I, Bodega G, Fernández B (2002) Glutamine synthetase in brain: effect of ammonia. *Neurochem Int* 41:123–143
 69. Butterworth RF (2010) Altered glial-neuronal crosstalk: cornerstone in the pathogenesis of hepatic encephalopathy. *Neurochem Int* 57:383–388
 70. Hermenegildo C, Monfort P, Felipo V (2000) Activation of N-methyl-D-aspartate receptors in rat brain in vivo following acute ammonia intoxication: characterization by in vivo brain microdialysis. *Hepatology* 31:709–715
 71. Albrecht J (1998) Roles of neuroactive amino acids in ammonia neurotoxicity. *J Neurosci Res* 51:133–138
 72. Hermenegildo C, Montoliu C, Llansola M, Muñoz M-D, Gaztelu J-M, Miñana M-D, Felipo V (1998) Chronic hyperammonemia impairs the glutamate-nitric oxide-cyclic GMP pathway in cerebellar neurons in culture and in the rat in vivo. *Eur J Neurosci* 10:3201–3209
 73. Zielińska M, Law RO, Albrecht J (2003) Excitotoxic mechanism of cell swelling in rat cerebral cortical slices treated with ammonia. *Neurochem Int* 43:299–303
 74. Saran T, Hilgier W, Kocki T, Urbańska EM, Turski WA, Albrecht J (1998) Acute ammonia treatment in vitro and in vivo inhibits the synthesis of a neuroprotectant kynurenic acid in rat cerebral cortical slices. *Brain Res* 787:348–350
 75. Szerb JC, Retondo IM (1993) Astrocytes and the entry of circulating ammonia into the brain: effect of fluoroacetate. *Metab Brain Dis* 8:217–234
 76. Schliess F, Görg B, Fischer R, Desjardins P, Bidmon HJ, Herrmann A, Butterworth RF, Zilles K, Häussinger D (2002) Ammonia induces MK-801-sensitive nitration and phosphorylation of protein tyrosine residues in rat astrocytes. *FASEB J* 16:739–741
 77. Kruczek C, Görg B, Keitel V, Bidmon HJ, Schliess F, Häussinger D (2011) Ammonia increases nitric oxide, free Zn²⁺, and metallothionein mRNA expression in cultured rat astrocytes. *Biol Chem* 392:1155–1165
 78. Yonden Z, Aydin M, Kilbas A, Demirin H, Sutcu R, Delibas N (2010) Effects of ammonia and allopurinol on rat hippocampal NMDA receptors. *Cell Biochem Funct* 28:159–163
 79. Hilgier W, Węgrzynowicz M, Mączewski M, Beręsewicz A, Oja SS, Saransaari P, Albrecht J (2008) Effect of glutamine synthesis inhibition with methionine sulfoximine on the nitric oxide-cyclic GMP pathway in the rat striatum treated acutely with ammonia: a microdialysis study. *Neurochem Res* 33:267–272
 80. Bużańska L, Zabłocka B, Dybel A, Domańska-Janik K, Albrecht J (2000) Delayed induction of apoptosis by ammonia in C6 glioma cells. *Neurochem Int* 37:287–297
 81. Cauli O, Gonzalez-Usano A, Agusti A, Felipo V (2011) Differential modulation of the glutamate-nitric oxide-cyclic GMP pathway by distinct neurosteroids in cerebellum in vivo. *Neuroscience* 190:27–36
 82. Gonzalez-Usano A, Cauli O, Agustí A, Felipo V (2013) Hyperammonemia alters the modulation by different neurosteroids of the glutamate-nitric oxide-cyclic GMP pathway through NMDA-, GABA_A- or sigma receptors in cerebellum in vivo. *J Neurochem* 125:133–143
 83. Gonzalez-Usano A, Cauli O, Agusti A, Felipo V (2014) Pregnenolone sulfate restores the glutamate-nitric-oxide-cGMP pathway and extracellular GABA in cerebellum and learning and motor coordination in hyperammonemic rats. *ACS Chem Neurosci* 5:100–105
 84. Izumi Y, Izumi M, Matsukawa M, Funatsu M, Zorumski CF (2005) Ammonia-mediated LTP inhibition: effects of NMDA receptor antagonists and L-carnitine. *Neurobiol Dis* 20:615–624
 85. Izumi Y, Svrakic N, O'Dell K, Zorumski CF (2013) Ammonia inhibits long-term potentiation via neurosteroid synthesis in hippocampal pyramidal neurons. *Neuroscience* 233:166–173
 86. Zorumski CF, Paul SM, Izumi Y, Covey DF, Mennerick S (2013) Neurosteroids, stress and depression: potential therapeutic opportunities. *Neurosci Biobehav Rev* 37:109–122
 87. Itzhak Y, Roig-Cantisano A, Dombro RS, Norenberg MD (1995) Acute liver failure and hyperammonemia increase peripheral-type benzodiazepine receptor binding and pregnenolone synthesis in mouse brain. *Brain Res* 705:345–348
 88. Desjardins P, Bandeira P, Raghavendra Rao VL, Ledoux S, Butterworth RF (1997) Increased expression of the peripheral-type benzodiazepine receptor-isoquinoline carboxamide binding protein mRNA in brain following portacaval anastomosis. *Brain Res* 758:255–258
 89. Desjardins P, Butterworth RF (2002) The “peripheral-type” benzodiazepine (omega 3) receptor in hyperammonemic disorders. *Neurochem Int* 41:109–114
 90. Cagnin A, Taylor-Robinson SD, Forton DM, Banati RB (2006) In vivo imaging of cerebral “peripheral benzodiazepine binding sites” in patients with hepatic encephalopathy. *Gut* 55:547–553
 91. Agusti A, Dziedzic JL, Hernandez-Rabaza V, Guilarte TR, Felipo V (2014) Rats with minimal hepatic encephalopathy due to portacaval shunt show differential increase of translocator protein (18 kDa) binding in different brain areas, which is not affected by chronic MAP-kinase p38 inhibition. *Metab Brain Dis* 29:955–963
 92. Ahboucha S, Coyne L, Hirakawa R, Butterworth RF, Halliwell RF (2006) An interaction between benzodiazepines and neuroactive steroids at GABA_A receptors in cultured hippocampal neurons. *Neurochem Int* 48:703–707

93. Ahboucha S, Gamrani H, Baker G (2012) GABAergic neurosteroids: the “endogenous benzodiazepines” of acute liver failure. *Neurochem Int* 60:707–714
94. Mladenovic D, Hrcic D, Petronijevic N, Jevtic G, Radosavljevic T, Rasic-Markovic A, Puskas N, Maksic N, Stanojlovic O (2014) Finasteride improves motor, EEG, and cellular changes in rat brain in thioacetamide-induced hepatic encephalopathy. *Am J Physiol Gastrointest Liver Physiol* 307:G931–G940
95. Matsuoka M, Igisu H, Kohriyama K, Inoue N (1991) Suppression of neurotoxicity of ammonia by L-carnitine. *Brain Res* 567:328–331
96. Felipo V, Miñana M-D, Cabedo H, Grisolia S (1994) L-Carnitine increases the affinity of glutamate for quisqualate receptors and prevents glutamate neurotoxicity. *Neurochem Res* 19:373–377
97. Miñana M-D, Hermenegildo C, Llansola M, Montoliu C, Grisolia S, Felipo V (1996) Carnitine and choline derivatives containing a trimethylamine group prevent ammonia toxicity in mice and glutamate toxicity in primary cultures of neurons. *J Pharmacol Exp Ther* 279:194–199
98. Matsuoka M, Igisu H (1993) Comparison of the effects of L-carnitine, D-carnitine and acetyl-L-carnitine on the neurotoxicity of ammonia. *Biochem Pharmacol* 46:159–164
99. Llansola M, Erceg S, Hernandez-Viadel M, Felipo V (2002) Prevention of ammonia and glutamate neurotoxicity by carnitine: molecular mechanisms. *Metab Brain Dis* 17:389–397
100. Felipo V, Hermenegildo C, Montoliu C, Llansola M, Miñana MD (1998) Neurotoxicity of ammonia and glutamate: molecular mechanisms and prevention. *Neurotoxicology* 19:675–681
101. Lheureux PER, Hantson P (2009). Carnitine in the treatment of valproic acid-induced toxicity. *Clin Toxicol (Phila)* 47:101–111
102. Kosenko E, Kaminsky Y, Grau E, Miñana M-D, Grisolia S, Felipo V (1995) Nitroarginine, an inhibitor of nitric oxide synthetase, attenuates ammonia toxicity and ammonia-induced alterations in brain metabolism. *Neurochem Res* 20:451–456
103. Hilgier W, Oja SS, Saransaari P, Albrecht J (2004) A novel glycine site-specific N-methyl-D-aspartate receptor antagonist prevents activation of the NMDA/NO/cGMP pathway by ammonia. *Brain Res* 1015:186–188
104. Túnez I, Muñoz MC, Villavicencio MA, Medina FJ, de Prado EP, Espejo I, Barcos M, Salcedo M, Feijóo M, Montilla P (2005) Hepato- and neurotoxicity induced by thioacetamide: protective effects of melatonin and dimethylsulfoxide. *Pharmacol Res* 52:223–228
105. Albrecht J, Węgrzynowicz M (2005) Endogenous neuro-protectants in ammonia toxicity in the central nervous system: facts and hypotheses. *Metab Brain Dis* 20:253–263
106. Bobermin LD, Quincozes-Santos A, Guerra MC, Leite MC, Souza DO, Goncalves C-A, Gottfried C (2012) Resveratrol prevents ammonia toxicity in astroglial cells. *PLoS One* 7(12):e52164. doi:[10.1371/journal.pone.0052164](https://doi.org/10.1371/journal.pone.0052164)