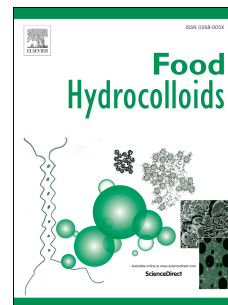


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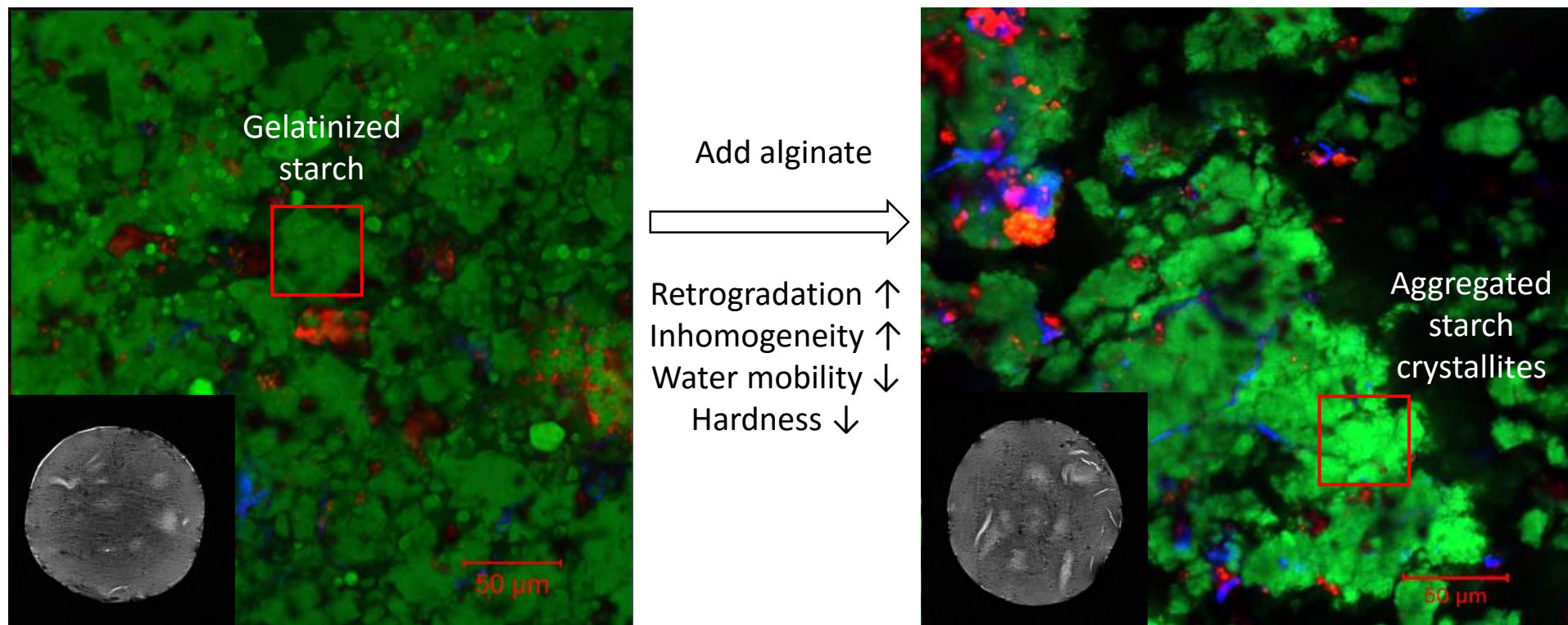
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Anti-staling action of alginate in steamed rice cakes



1 Anti-staling of high-moisture starchy food: effect of hydrocolloids,
2 emulsifiers and enzymes on mechanics of steamed-rice cakes

3

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16

17 Abstract

18 High-moisture rice snacks, such as steamed rice cakes, develop firmness on storage which
19 decreases shelf life significantly. By analogy with lower moisture bread systems, this staling
20 was hypothesised to be due to a combination of starch retrogradation and moisture re-
21 distribution. Therefore, food additives which are commonly used to retard starch
22 retrogradation during bread staling, including enzymes, hydrocolloids and emulsifiers (alpha-
23 amylase, alginate, xanthan, guar gum, carrageenan, carboxymethyl cellulose, distilled
24 monoglyceride, and sodium stearoyl lactylate) were investigated for their anti-hardening
25 effects in high-moisture rice snacks. The results showed that only alginate significantly
26 reduced the firming rate of rice cakes. However, differential scanning calorimetry
27 measurements surprisingly indicated that rice cakes with alginate had higher levels of starch
28 retrogradation than the control after storage for 7 days. Magnetic resonance imaging results
29 were characterised by a redistribution of signal intensity from the edge to the centre of rice
30 cakes and the formation of high intensity regions. These features were stronger with the
31 addition of alginate. We propose that the alginate forms a continuous phase with water that
32 has high mobility, whereas the partially gelatinized starch granules are an included phase
33 distributed within the continuous phase. The reduced hardness of aged rice cakes with
34 alginate is more dependent on the soft continuous phase than the hard starch granules,
35 therefore leading to a softer texture. This mechanism is different to that proposed to operate
36 for lower water content baked systems, therefore hydrocolloid and other anti-staling agents
37 which are effective in bread systems may not be applicable in higher moisture starchy foods.

38 Key words: firming; high-moisture; water mobility; additives

39

40 1 Introduction

41 High-moisture starchy foods that contain 35% or greater moisture content or a water activity
42 above 0.9 experience significant staling processes which result in hardened texture and off
43 flavours that limit shelf life. The staling mechanisms that are widely studied in intermediate-
44 moisture starchy foods are also thought to be responsible for the hardening of high-moisture
45 foods, namely moisture loss and redistribution as well as starch retrogradation (Ji, Zhu, Zhou,
46 & Qian, 2010; Lee, *et al.*, 2014).

47 Various anti-staling agents such as enzymes, hydrocolloids and emulsifiers have been
48 applied successfully in intermediate-moisture foods, particularly breads, to control staling.
49 The enzymes α -amylase and β -amylase have an anti-staling effect on bread by partially
50 degrading starch and generating low-molecular-weight dextrans (De Stefanis, Ponte Jr,
51 Chung, & Ruzza, 1977; Hebeda, Bowles, & Teague, 1991; Katina, Salmenkallio-Marttila,
52 Partanen, Forsell, & Autio, 2006; Nguyen, *et al.*, 2015; Outtrup & Norman, 1984). The
53 dextrans are thought to hinder gluten-starch interactions and interrupt the starch network
54 (Goesaert, Leman, Bijttebier, & Delcour, 2009; Goesaert, Slade, Levine, & Delcour, 2009).

55 Hydrocolloids enhance water retention and limit its redistribution within starchy baked food
56 structures (Sozer, Bruins, Dietzel, Franke, & Kokini, 2011), which provides an anti-staling
57 effect. The type, source and molecular weight of hydrocolloids are all important factors that
58 influence the anti-staling ability in different food products (Guarda, Rosell, Benedito, &
59 Galotto, 2004; Gujral, Haros, & Rosell, 2004). Guarda, *et al.* (2004) reported that alginate
60 and hydroxypropyl methyl cellulose (0.1% - 0.5%) showed wheat bread crumb hardening
61 retardation and reduction of moisture loss while xanthan and carrageenan only reduced
62 moisture loss.

63 Effective emulsifiers retard staling of intermediate-moisture bread by influencing the firming
64 rate instead of simply reducing the initial hardness of bread (Knightly, 1977) which
65 shortening is normally capable of. The interactions between emulsifiers and starch and/or
66 protein are thought to be the reason for anti-staling effects (Gray & Bemiller, 2003).
67 Emulsifiers form single helical V-type complexes with amylose, which may reduce starch
68 swelling and amylose leaching from starch granules or prevent migration of water from
69 gluten to facilitate starch crystallisation, further reducing hardness of bread (Dragsdorf, 1980;
70 Zobel, 1973, 1988). This complex-forming ability relies on amylose structure, the type, pH,
71 ion concentration of emulsifiers (Krog, 1973), and their physical state before adding to the
72 dough, with aqueous, liquid-crystalline gels showing best results (Krog & Jensen, 1970).
73 However, this may not be the only reason for the anti-staling effects of emulsifiers, as
74 emulsifiers can complex with amylopectin as well, although at a lower level, to reduce

75 amylose-amylopectin network formation (Gudmundsson & Eliasson, 1990). Mikus, Hixon,
76 and Rundle (1946) and Tang and Copeland (2007) suggested that monoglycerides form
77 helical complexes of aggregated structure with amylose to decrease hardness caused by
78 amylose retrogradation and amylose complexes with lipids and other components. In
79 contrast to the many reported studies of anti-staling effects in breads and other intermediate
80 moisture foods, there have been few comprehensive studies of anti-staling effects in high
81 moisture foods. We hypothesised that anti-staling agents with proven effectiveness in breads
82 would also have anti-staling properties in high moisture foods.

83 The research example of high-moisture starchy foods in this study, steamed rice cakes, are
84 a traditional snack food in some Asian countries. Steamed rice cakes have a moisture
85 content of 40%-65% and a water activity of around 0.92 (Eunhye Choi & Ko, 2014; Ji, Zhu,
86 Qian, & Zhou, 2007; Sang, Shao, & Jin, 2015). A few studies have shown that additives such
87 as tea polyphenols, oligosaccharides and polysaccharides (β -glucan, β -cyclodextrin,
88 xanthan gum, carrageenan etc.) can reduce the rate and extent of retrogradation in rice
89 starch (Banchathanakij & Suphantharika, 2009; Tang, Hong, Gu, Zhang, & Cai, 2013; Tian,
90 *et al.*, 2009; Wu, Chen, Li, & Li, 2009). However, the mechanisms behind the retardation of
91 starch retrogradation in these cases are not well characterised. The oligo- and poly-
92 saccharides might function as hydrocolloids by holding water to control the amount of water
93 available for inclusion in starch crystallites. Tea polyphenols may act as plasticizers to
94 interact with starch by hydrogen bonding, and reduce self-entanglement of side chains of
95 starch polymers (Wu, *et al.*, 2009). The hydrogen bonding depends on size, number and
96 reactivity of OH groups in the polyphenol additives (Smits, Kruiskamp, Van Soest, &
97 Vliegenthart, 2003). No work has been reported for ready-to-eat high-moisture rice products
98 to investigate starch retrogradation retardation and anti-hardening effect of additives, or to
99 reveal the potential mechanisms.

100 The aim of this study is to evaluate the anti-staling effect of several enzymes, hydrocolloids
101 and emulsifiers on high-moisture steamed rice cakes. Uniaxial compression testing was
102 used to determine the mechanical properties of rice cakes, particularly hardness, as a
103 function of storage time. Factors thought to be closely related to hardness development were
104 investigated with the goal of defining the anti-staling mechanisms: distribution of water
105 (magnetic resonance imaging) and major components (confocal laser scanning microscopy);
106 starch recrystallization (differential scanning calorimetry); and molecular mobility (magnetic
107 resonance imaging).

108 2 Materials and Methods

109 2.1 Materials

110 Sticky rice flour and rice flour were obtained from Erawan marketing Co., Bangkok, Thailand.
111 The amylose contents of the sticky rice flour and rice flour were 1.87 and 20.84%,
112 respectively, measured using the size exclusion chromatography method of Fitzgerald, *et al.*
113 (2009). The protein content of sticky rice flour and rice flour are 6.61% and 6.98%,
114 respectively (from the manufacturer's nutrition information table). The total fat content of both
115 flours is less than 1%. Sugar was obtained from Sugar Australia Pty Limited, Australia.
116 Enzyme (fungal α -amylase with enzyme activity of min. 1000 units/g), hydrocolloids (guar,
117 carboxymethyl cellulose (CMC), carrageenan, alginate (extracted from brown algae
118 *Laminaria digitate* with M/G ratio of 1.5:1), and xanthan) and emulsifiers (sodium stearoyl
119 lactylate (SSL), distilled monoglyceride (DM)) were provided by Danisco Australia Pty Ltd
120 (Banksmeadow, Australia).

121 2.2 Preparation of steamed rice cake

122 Control steamed rice cakes were produced by mixing sticky rice flour, rice flour, sugar and
123 water together in a ratio of 3:2:1:3.6 (w/w) using a bench mixer (GS-6118, Homemaker,
124 Kmart Australia) at low speed for 1 min following a traditional recipe for steamed rice cakes
125 in Asian countries. The dough was then shaped into 50 g balls and cooked in a steamer
126 (RC-4700-A, Homemaker, Kmart Australia) for 15 min. After cooling down, each rice cake
127 was packaged into a 12 μ m polyethylene terephthalate/120 μ m linear low density
128 polyethylene stand up pouch (10010001, West's Packaging Services P/L, Australia) and
129 sealed by a food packaging machine (Sous Vide 260, Australia Vacuum Packaging
130 Machines Pty Ltd, Australia). Anti-staling additives including enzymes, hydrocolloids and
131 emulsifiers were mixed with water first in concentrations as shown in Table 1, then the
132 mixture of sticky rice flour, rice flour and sugar was added to the anti-staling additive-water
133 system using the same ratio as control steamed rice cakes. The same mixing and shaping
134 processes were performed. The two sets of doughs with amylase were covered with cling
135 wraps (Cling Wrap, Berry Plastics Pty Ltd, Australia) to prevent moisture loss and left at
136 room temperature for 0.5h and 1h, respectively, allowing time for amylase action on starch.
137 All the samples were then steamed and packaged in the same way as control steamed rice
138 cakes.

139 2.3 Properties of steamed rice cakes during storage

140 The packaged steamed rice cakes were stored at room temperature ($22\pm 1^\circ\text{C}$, tested using
141 wireless thermocouples (Hitemp 140-FP-36, MadgeTech Inc., USA)) for 2, 5 and 7 days.

142 The hardness of all samples was measured on days 0, 2, 5 and 7 to screen for effective anti-
143 staling additives in steamed rice cakes. The rice cakes with effective anti-staling additives
144 and control rice cakes were further used to measure starch retrogradation, microstructure,
145 and moisture content on days 0, 2, 5 and 7. Moisture distributions of rice cakes were
146 measured on days 0 and 7.

147 2.3.1 Hardness

148 An Instron universal testing machine (5800, Illinois Tool Works Inc., USA) was used to
149 measure the compression response of rice cakes. Bluehill2 software (Instron Corp., USA)
150 was used to operate the experiment and record data. The centre of the rice cake samples
151 was cut into a cylinder of 20 mm (diameter) × 20 mm (height) for instrumental texture
152 measurement. The height of each cylindrical sample was measured by a 150mm digital
153 Vernier Caliper (Work Zone, Australia) and recorded in the software. Then the sample was
154 placed on an Instron compression cylindrical plate (T1223-1021) with a diameter of 50 mm.
155 Another cylindrical compression plate (T1223-1021 with diameter of 50 mm) was placed
156 directly above the sample and the gauge length was reset as 0mm. A 2000 N load cell was
157 used to compress the sample until reaching the maximum load of 1500 N at a crosshead
158 speed of 2 mm/s. The compressive stress and strain were recorded and graphed. The
159 hardness was reported as the modulus in a linear region of compressive strain between 10%
160 and 20%.

161 2.3.2 Starch retrogradation by differential scanning calorimetry (DSC)

162 Analyses were performed in a DSC Q2000 (TA Instruments, United States), using Tzero
163 aluminium hermetic sample pans and lids (901683.901). 10 mg of rice cake was weighed by
164 an ABT 120-5DM analytical balance (Kern & Sohn GmbH, Balingen, Germany) into Tzero
165 aluminium sample pans. All the sample pans were hermetically sealed using a Tzero sample
166 press and hermetic die kit (TA Instruments, United States). An empty sealed pan was used
167 as a reference. Each sample was heated from 30 to 130 °C at a speed of 10 °C/min with
168 nitrogen flow of 50 mL/min. The heat flow curves were recorded and TA Universal Analysis
169 software was used to analyse the data. The temperature values obtained were for the onset
170 (T_o), peak (T_p), and completion (T_c) of the endothermic transition. The enthalpy of transition
171 was estimated from the integrated heat flow over the temperature range of the transition,
172 and is expressed as joule per gram sample (J/g).

173 2.3.3 Microstructural properties of rice cake by confocal laser scanning microscopy (CLSM)

174 The micro-structural properties of rice cakes were investigated using confocal microscopy
175 (LSM 700, Carls Zeiss, Germany). A triple-staining technique including the combination of
176 Calcofluor White, FITC (1% w/v, ethanol) and Rhodamine B (0.1% v/w, ethanol) was used.

177 A very thin slice of rice cake (obtained by hand using a razor blade ~ 10 mg) was incubated
178 with 1 ml of FITC for 48 hours under gentle shaking conditions followed by washing with
179 water until the washed water was observed to be clear. The FITC stained cake was further
180 incubated with 1 ml of Rhodamine B for 4 hours under shaking conditions followed by
181 washing until the supernatant was clear. A small section of double stained cake was placed
182 on a glass slide and infiltrated with Calcofluor white for 5 min. The excess of Calcofluor white
183 was absorbed by a filter paper tip. The sample was observed after excitation at 405, 488
184 and 555 nm for Calcofluor White, FITC and Rhodamine B respectively. Normally, double-
185 staining of mixed starch and protein systems with a mixture of Rhodamine B and FITC leads
186 to staining of proteins by Rhodamine B and starch by FITC. However, both Rhodamine B
187 and FITC can bind non-covalently with each of starch and protein, depending upon the
188 concentration of the components in the system. In a low protein system, such as rice cake,
189 and a high concentration of FITC (FITC 1% vs Rhodamine B 0.1%) the FITC binds with both
190 starch and protein. In contrast, Rhodamine B binds more specifically with protein allowing
191 the visualisation of both starch and protein simultaneously (Zheng, Stanley, Gidley, & Dhital,
192 2016). The use of Calcofluor White allows visualisation of the non-starch polymers, mainly
193 the distribution of added hydrocolloid and rice cell wall material.

194 2.3.4 Moisture content

195 Moisture was determined by drying the sample to a constant weight in an oven at 105 °C for
196 24h. The difference in the weight of the sample before and after drying was used to calculate
197 the moisture content (AOAC, 1990).

198 2.3.5 Water distribution by magnetic resonance imaging (MRI)

199 Rice cakes were imaged in a preclinical MRI system, comprising a 300mm bore 7T ClinScan
200 (Bruker, Germany), running Siemens software version VB17. A 72 mm ID MRI rf coil was
201 used to acquire the images. The rice cakes were placed on a ruler taped to the bottom of a
202 rat-sized bed. The following images were acquired.

203 1. Localiser images: Three sets of orthogonal gradient echo images were acquired with
204 the following parameters: repetition time (TR) = 41 msec, echo time (TE) = 4.43 msec, field
205 of view 130 X 130 mm, slice thickness = 1 mm, number of slices in each direction = 5, matrix
206 = 128 X 128, flip angle = 25°, bandwidth = 400 Hz/Px, averages = 1, concatenations = 3,
207 total scan time = 16 sec.

208 2. T2 TSE images – Coronal T2 weighted spin echo images were obtained with the
209 following parameters: TR = 2800 msec, TE = 49 msec, field of view 60 X 54 mm, matrix =
210 256 X 256, slice thickness 0.6 mm, number of slices = 19, averages = 1, bandwidth = 130
211 Hz/Px, turbo factor = 7, total scan time = 3 min 12 sec.

212 3. T1 parameter images were generated using the 3D_VIBE_t1mapit sequence using
213 the following parameters: TR = 12msec, TE = 1.02 msec, field of view 60 X 55 X 60 mm,
214 image matrix = 160 X 160 X 120, flip angle = 6°, and 33°, bandwidth = 400 Hz/Px, averages
215 = 2, total scan time = 8 min 31 sec. A 3D T1 map was automatically generated following
216 acquisition of the images.

217 4. T2 parameter images were generated using multi-echo T2 images acquired with the
218 following parameters: TR = 2500 msec, TE = 10-120 ms in 10 msec increments, field of view
219 60 X 58 mm, matrix = 192 X 192, slice thickness 1.0 mm, number of slices = 20, averages =
220 2, bandwidth = 196 Hz/Px, total scan time = 15 min 34 sec. T2 parameter images were
221 automatically generated following acquisition of the images.

222 2.4 Statistical analysis

223 All measurements were performed with at least 3 replicates. The mean values and standard
224 deviations were reported. Microsoft Excel 2013 and R software (3.2.3) were used for
225 statistical analysis.

226 3 Results and Discussion

227 3.1 Hardness

228 Staling of starchy foods is typically associated with increases in compressive hardness due
229 to starch retrogradation and redistribution of moisture. Thus the hardness of the rice cakes
230 was monitored to represent staling over time and to differentiate between successful and
231 unsuccessful anti-staling interventions. Examples of the full range of compressive stress-
232 strain curves of rice cakes are shown in Figure 1a. The stress-strain curves are divided into
233 4 regions: a region of partial contact because of the uneven surfaces of rice cakes; a linear
234 region before breaking (Figure 1b) which was used to calculate the modulus to represent the
235 hardness of the rice cakes; the breaking region; and a linear compressive region after
236 breaking.

237 The compression modulus of all rice cakes, shown in Figure 2, increases significantly after 7
238 days of storage. All the concentrations of amylases added to rice cakes with both incubation
239 times showed significantly higher hardness than the control (Figure 2a). In contrast to bread
240 (Goesaert, Slade, *et al.*, 2009), amylase appears to be ineffective at preventing staling of
241 high-moisture rice cakes. The mechanism of anti-staling effect of amylase in intermediate-
242 moisture bread is complex and incompletely understood but is likely to involve effects on
243 both starch retrogradation and consequent firming processes (Akers & Hosney, 1994; León,
244 Durán, & de Barber, 1997). Low-molecular-weight amylase products, such as branched
245 dextrins, maltotriose or maltotetraose, are likely to reduce starch retrogradation as indicated

246 by a lower endotherm enthalpy on re-heating (Lin & Lineback, 1990; Schults, Schoonover,
247 Fisher, & Jackel, 1952). Conversely, amylase products with higher molecular weight might
248 show the opposite effect because the starch recrystallization processes underlying
249 retrogradation is promoted by the more mobile polymers produced by enzyme action (Akers,
250 *et al.*, 1994). In this study, the increased hardness of steamed rice cakes following amylase
251 treatment suggests that the effects on the starch network may have been caused by the
252 presence of staling-accelerating hydrolysis products in rice cakes.

253 Most concentrations of DM and SSL show significantly higher hardness than control across
254 the whole storage time (Figure 2a). Similar to amylase, staling retardation due to emulsifiers
255 is not observed in high-moisture rice cakes, which is opposite to the effect normally reported
256 in intermediate-moisture bread (Gomes-Ruffi, Cunha, Almeida, Chang, & Steel, 2012).

257 Most of the rice cakes with suitable amounts of hydrocolloid have a lower hardness
258 compared to the control after storage for 5 and 7 days (Figure 2b), presumably due to the
259 general water holding capacity of hydrocolloids. This is in line with Davidou, Le Meste,
260 Debever, and Bekaert (1996) where addition of 0.6% w/w of locust bean gum, 0.3% alginate
261 or xanthan in wheat bread, and Sim, Noor Aziah, and Cheng (2011) and Sim, Noor Aziah,
262 and Cheng (2015) where addition of 0.2% w/w flour of alginate or 0.8% w/w flour of konjac
263 glucomannan in Chinese steamed bread reduced hardness by reducing the dehydration rate
264 of samples, or influencing protein-starch interaction or macromolecular entanglement. The
265 statistical analysis shows that only the rice cakes with an addition of 0.3% w/w alginate show
266 significantly lower hardening rate ($p < 0.05$), while the rest are not significantly different from
267 the control rice cakes. This suggests that alginate is a potential anti-staling agent for
268 application in high-moisture starchy foods such as steamed rice cakes, although the
269 mechanism needs to be uncovered for predictable application. One potential mechanism is
270 that, in high-moisture starchy foods, hydrocolloids might absorb more water which reduces
271 the amount of water available to be included in starch crystallites. Xanthan is reported to
272 have a greater water holding ability (Sánchez, Bartholomai, & Pilosof, 1995), but its anti-
273 staling effect on steamed rice cakes is less than alginate. However, water holding ability as
274 an isolated hydrocolloid may not be a good indicator of water holding in a high starch and
275 sugar environment as found in rice cakes. One possibility is that xanthan binds more to
276 (neutral) starch than the more highly charged alginate does, and therefore contributes less to
277 water-holding. Further work (e.g. using labelled xanthan to identify its location within rice
278 cakes) would be needed to test this possibility. In order to further investigate the
279 mechanism(s) by which alginate exerts its anti-staling effect, the enthalpy required for
280 melting retrograded amylopectin, and the rice cake moisture content were measured.

281 3.2 Starch retrogradation

282 Starch retrogradation has been found to be an important factor influencing the hardness of
283 starchy food systems such as bread and cake (Katz, 1928; Maga & Ponte, 1975; Zhu, 2016).
284 As the main component on a dry basis of steamed rice cakes, starch experiences structural
285 changes both in the steaming process (gelatinization) and on storage (retrogradation) which
286 greatly influence the texture. Therefore, the retrograded starch may play a major role in the
287 staling mechanism. DSC tests were applied to control and 0.3% alginate-added rice cake
288 samples to capture the heat flow curves and compare the levels of starch retrogradation.

289 A typical peak representing the melting of retrograded amylopectin was detected and the
290 onset temperature, peak temperature, conclusion temperature, and enthalpy were recorded,
291 as shown in Table 2. With increasing storage time, the width of the transition ($T_c - T_o$)
292 increased and the peak shifted to a lower temperature in the control group, while the alginate
293 group did not show this shift. This suggests that after long term storage, amylopectin in rice
294 starch forms a broad range of imperfect and (for control) slightly more labile crystalline
295 structures, consistent with the ageing process involving more of the starch in crystallisation
296 rather than an annealing of the same crystallites into more perfect crystals. Rice cakes with
297 0.3% alginate show lower onset and peak temperatures than the control, suggesting that
298 alginate influences the formation of crystalline structures by interacting with starch molecules
299 and/or interacting with water to control the availability of water within starch networks.

300 The enthalpy values increase with storage time, consistent with increasing amounts of
301 retrograded amylopectin. Control rice cakes have less retrograded amylopectin (smaller
302 enthalpies) compared to rice cakes with 0.3% alginate at the same storage time (Table 2).
303 Alginate is a macromolecule and is unlikely to be able to penetrate either intact starch
304 granules as only molecules smaller than 1000 g/mol are able to penetrate starch granules
305 (Lathe & Ruthven, 1956) or swollen granules (Appelqvist & Debet, 1997). In addition,
306 alginate absorbs water first as it was pre-dissolved prior to mixing with dry flour. Due to
307 competition with the alginate, less water would be expected to be available in this
308 concentrated system for the starch granule swelling (Gonera & Cornillon, 2002; Ramírez, *et*
309 *al.*, 2015). The more intact granules and ordered structure of starch in alginate-added rice
310 cakes are proposed to result in larger DSC enthalpies than the control. However, from the
311 compression test results (Figure 2), the rice cakes with addition of 0.3% alginate have a
312 lower hardness than controls. This suggests that amylopectin retrogradation is not
313 responsible for the firming of high-moisture rice cakes and that the anti-firming action of
314 alginate has another origin. From phase separation theory (Appelqvist, *et al.*, 1997), alginate
315 might form a continuous soft phase with water and leached amylose, which separates from a
316 second phase made of stiffened starch granules. The balance of these two phases may

317 affect the hardness of steamed rice cakes. Possibly the soft alginate-water phase is
318 dominant and yields the softer rice cake texture found in this study. To test this hypothesis,
319 direct observation of the microstructure of rice cakes using microscopy can be used to
320 examine if there are two phases. In addition, the softness of the continuous phase is likely
321 related to the water behaviour for which MRI can be used to derive information on molecular
322 mobility.

323 3.3 Microstructure

324 Since amylopectin retrogradation was not correlated with hardness development of high-
325 moisture steamed rice cakes, the microstructure is of interest to reveal any difference
326 between samples with and without 0.3% alginate. Figure 3 shows CLSM images of stained
327 samples from control and alginate-containing rice cakes. Both groups showed similarities in
328 changes during storage for 7 days. In fresh rice cake samples (day 0, Figure 3 (a1), (b1)),
329 most of the starch molecules, stained green, formed a continuous phase as they were
330 gelatinised. Bright green areas show that some starch granules are kept intact and
331 aggregate with limited swelling (Figure 3c). With increasing storage time, starch granule
332 aggregates are clearly observed and their size increases by day 7. This suggests that some
333 phase separation is occurring, perhaps driven by starch molecules crystallizing and forming
334 a more ordered structure compared to the fresh samples (as evidenced by DSC – Table 2).
335 Proteins (stained red) are dispersed between starch granules apparently uniformly. Even
336 though the two groups of samples show a similar overall behaviour, the control group
337 appears to have less and smaller aggregates of residual partially-swollen granule structures
338 compared to the alginate group. In addition, they tend to be more continuous and have a
339 looser structure. This observation is consistent with the DSC results that the control group
340 has a smaller enthalpy for retrograded amylopectin than the alginate group.

341 Calcofluor stained non-starch polymers blue, including cell walls and alginate in this study.
342 The blue parts observed in Figure 3 (a1-3) and (b1-3) are believed to be the cell walls, as
343 cell wall polysaccharides account for 2% of the dry weight of polished rice grains (Palmer, *et*
344 *al.*, 2015), while the alginate content is only 0.3% of rice flour basis, which is less than cell
345 wall polysaccharides. Alginate appears to be distributed uniformly in the inter-granular
346 spaces of the rice cakes (in the background of Figure 3 (a1-3) and (b1-3)). This reduces the
347 inter-particle contact, interrupting the continuity of the network of starch granules and
348 weakening the composite network structure (Biliaderis, Arvanitoyannis, Izydorczyk, &
349 Prokopowich, 1997). From these results, it is hypothesised that aged rice cakes with addition
350 of alginate have a lower overall hardness because of a softer continuous alginate/water
351 continuous phase despite containing harder swollen starch granules as an included phase.

352 3.4 Moisture content

353 The results of the present study (Figure 2) show that only 0.3% alginate addition in the rice
354 cakes exhibited less firming than the control. As starch retrogradation is not positively
355 correlated with the firming of aged rice cakes, it is possible that water content and/or
356 distribution play an important role in the firming process. Therefore, the moisture contents of
357 the control group and the alginate 0.3% group were tested, but the results (Figure 4) show
358 no significant difference of moisture content between the two groups before or after different
359 storage periods. Moisture loss can be an important factor influencing staling of starchy food
360 products, however, in this study, moisture loss was prevented by sealed polyethylene
361 packaging and was not responsible for rice cake staling. Even though the total moisture
362 content of the rice cakes did not change significantly during storage, there may be a change
363 of water mobility and moisture distribution. In studies related to bread staling, the moisture
364 redistribution between components, particularly starch and protein, changes the physical
365 structure of the molecules and their properties, and influences the staling process (Schiraldi
366 & Fessas, 2000).

367 3.5 Moisture distribution

368 MRI was used to capture the moisture distribution and water mobility of rice cakes on day 0
369 (fresh) and day 7 (stale) to better understand the effect of water on staling. Three central
370 images of the proton-density, T1 (longitudinal relaxation time) and T2 (spin-spin relaxation
371 time)-weighted images of fresh (day 0) and stale (day 7) control and 0.3% alginate-added
372 steamed rice cakes are shown in Figure 5. The MRI images (Figure 5) consist of dark areas
373 representing air bubbles in the rice cake samples and an intermediate-intensity background
374 representing the rice cake base. In the stale samples, high image intensity (bright) regions
375 develop, consistent with areas of higher water density and/or mobility. Steamed rice cakes
376 have a foam-like structure with an extensive number of fine pores, while the void areas might
377 be filled with air and/or liquid. The porous structure of steamed rice cakes makes it hard to
378 convert proton density to water content in local areas and calculate the precise T1 and T2
379 values for analysing the difference in water mobility between samples quantitatively.
380 However, a difference between fresh and aged samples is observed in the proton density,
381 T1 and T2-weighted MRI images with the appearance of bright areas in aged samples. The
382 pixel intensity distribution in the proton density, T1 and T2 images are shown in Figure 6.

383 The proton density images of fresh control and fresh alginate added samples are very similar
384 in that they both have higher signal intensity near the edges than in the centre, which
385 suggests that the water distribution in fresh samples is not uniform. The porous structure of
386 rice cakes leads to two possibilities. Firstly, the pores may not be evenly distributed, with an

387 outer layer containing a lower density of pores and/or smaller pore sizes below the image
388 resolution compared to the central region. Secondly, the outer layer of pores may be filled
389 with water to a greater extent than the inner pores. Water vapour transfers to the pores near
390 the edges during the steaming process, but the short time (15min) and the hydrophilic nature
391 of starch may limit diffusion of water from the surface into the centre. With increasing storage
392 time, water tends to move to the centre as the whole system moves towards an equilibrium
393 state.

394 This is different from water movement in intermediate-moisture bread which has a higher
395 initial moisture content in the internal crumb (25%-35%) than the external crust (15%-20%)
396 (Czuchajowska & Pomeranz, 1989; Mandala, Karabela, & Kostaropoulos, 2007) due to
397 water evaporation during baking. During equilibration, water moves from the crumb to the
398 crust and evaporates from the crust to the surrounding air (Maga, *et al.*, 1975), i.e. the
399 opposite direction to steamed rice cakes. The cling wrap covering of the rice cakes in this
400 study would minimize evaporation, and enhance water re-distribution in the rice cakes. This
401 is likely to be one of the main reasons for the apparently different mechanisms of staling in
402 intermediate- and high-moisture starchy foods.

403 Both fresh and aged alginate-added samples have brighter centres than the control and
404 water movement is more pronounced in control samples, which may be due to the water
405 holding ability of alginate. Fresh and aged samples can be differentiated as a few areas with
406 high proton density are observed in both aged samples (Figure 5a). These areas might be
407 composed of gel structures with limited porosity. Generally, the pixel intensity in the centre of
408 the stale rice cakes displayed higher maximum intensity and greater variation in the pixel
409 intensity, than the fresh samples after 7 days' storage (Figure 6a). These bright areas are
410 consistent with regions of higher water content and reduced air bubbles. This may occur as
411 air bubbles are filled with water expelled from starch structure in the rice cakes. However,
412 the alginate added samples have more bright areas than the control, consistent with
413 comparatively more water binding with the alginate. Morris (1990) suggested that water can
414 be lost from a starch-rich phase due to the molecular associations between starch chains as
415 occurs during retrogradation. The DSC results show that rice cakes with alginate added
416 have more retrograded starch than the control, which may expel more water and result in
417 brighter areas (more inhomogeneity) in the alginate group. There is also experimental
418 evidence that some types of hydrocolloids work as a coating agent to reduce interaction of
419 starch molecules and retard syneresis (Charoenrein, Tatirat, Rengsutthi, & Thongngam,
420 2011; Ferrero, Martino, & Zaritzky, 1994; Hahm & Kuei, 2015; Lee, Baek, Cha, Park, & Lim,
421 2002). Shi and BeMiller (2002) suggested that this effect was likely due to interactions
422 between certain leached molecules, primarily between amylose and certain gums. In this

423 study, alginate neither reduced starch retrogradation, nor inhibited syneresis, so this
424 mechanism does not seem to apply.

425 T1-weighted images (Figure 5b) of both fresh control and fresh alginate-added samples
426 have greater intensity towards the surface than in the centre, with water distribution moving
427 towards a more homogeneous state after 1 week's storage, similar to that observed in
428 proton density images. The T1 images also showed higher pixel intensity and greater
429 variation in the pixel intensity following 7 days of storage. The reduced variation in the
430 profiles in the stale samples is consistent with the high intensity regions not being as
431 prominent in the T1 images. This suggests similar T1 times for the water distributed
432 throughout the rice cake and in the regions of high intensity. Figure 6b shows that the
433 intensity of fresh alginate added samples are slightly higher than that of the fresh control,
434 which may indicate a general shortening of T1 or different porosity caused by the alginate. In
435 this study, alginate was first added to water, followed by the mixture of rice flours and sugar,
436 thus less water is available for binding with starch in alginate-added samples. This is
437 consistent with a reduced overall water mobility / porosity after addition of alginate. The
438 intensity difference between the surface and the centre is more pronounced in the control,
439 suggesting that an alginate-water phase facilitates moisture diffusion into the rice cake. This
440 is consistent with the presence of alginate increasing the phase volume of the extra-granular
441 phase by reducing the amount of water associated with swollen starch granules. The slower
442 hardness development in alginate-added samples might therefore result from a continuous
443 phase with alginate which remains soft with less reduction of water mobility.

444 Bread staling has been studied extensively, however, the movement of water between
445 components at the molecular level is still not clear. In bread staling research, water
446 transportation from starch to gluten and from gluten to starch have both been hypothesized
447 based on instrumental measurements (Bachrach & Briggs, 1947; Cluskey, Taylor, & Senti,
448 1959; Eliasson, 1983; Ribotta & Le Bail, 2007; Senti & Dimler, 1960). However, steamed rice
449 cake has a very different composition to wheat bread, particularly more water, less protein
450 and fat, and no gluten, which makes the mechanisms of staling behaviour different. With no
451 gluten and less protein content, water might be more mobilized and involved in the starch
452 network to reduce crystal formation and starch retrogradation, as a smaller DSC enthalpy
453 (1.35 J/g) was observed in the control samples after 7 days' storage than was reported for
454 bread (more than 3 J/g) (Baik & Chinachoti, 2000; Rogers, Zeleznak, Lai, & Hosney, 1988).

455 T2-weighted images (Figure 5c) of both control and alginate-added samples have a bright
456 layer on the surface, indicating that during steaming free water diffuses into the pores in this
457 layer or there is a thin layer of water between the packaging and the rice cake which

458 provides the high image intensity. The contrast between the background rice cake and the
459 regions of high intensity in the stale cakes was greatest in the T2 images, in particular the
460 alginate stale sample (blue dashed line, Figure 6c). After 1 week's storage, there was a
461 general increase in the image intensity at the centre of both samples (Figure 6c). This is
462 consistent with water diffusing from the surface to the centre resulting in a more
463 homogeneous water distribution. In addition, the aged samples have distinct bright areas,
464 more obvious in alginate-added samples (Figure 6c), similar to what was found in the proton
465 density images. The bright areas are consistent with longer T2 values and/or lower porosity.
466 The higher intensity regions may result from areas with lower porosity, pores filled with water
467 or free water filling cracks in the rice cakes. The aggregated bright areas possibly represent
468 a liquid phase with high water mobility, similar to the hypothesis of a continuous aqueous
469 phase leading to soft texture of fresh rice cakes. For wheat bread samples, T2
470 measurements suggest three distinct ranges: 8-14 μ s, 280-360 μ s, and 2000-3000 μ s (Chen,
471 Long, Ruan, & Labuza, 1997). These ranges are proposed to represent water (8-14 μ s)
472 strongly associated with other molecules by hydrogen bonding, particularly macromolecules
473 such as starch and gluten in bread; water (280-360 μ s) associated with macromolecules to a
474 certain degree; and mobile water (2000-3000 μ s) which was closely correlated with the
475 firming process. Some studies related two ranges of water (microsecond and millisecond
476 range) to two mobility fractions of water (Ruan, *et al.*, 1996). Two fractions are observed in
477 T2 images of rice cakes as well. One is the portion of water with low mobility shown as the
478 intermediate intensity background of rice cakes, while the other is the portion with high
479 mobility shown as high intensity separated areas in Figure 5c. The changes of water mobility
480 might be caused by macromolecules such as starch including water to form crystals during
481 retrogradation or expelling water as the physical structure of the macromolecules change
482 due to e.g. re-crystallisation. Overall, the MR imaging results are characterised by movement
483 of signal intensity from the edge to centre of the rice cakes and formation of high intensity
484 regions in aged cakes.

485 4 Conclusions

486 Among three categories of additives tested (hydrocolloids, enzymes and emulsifiers),
487 alginate was the only one that showed a significant reduction of hardening rate for steamed
488 rice cakes during storage. The additives were chosen based on their anti-staling effect on
489 low-moisture foods such as breads, but the results suggest most are not effective in higher
490 moisture systems. Amylase treatment and emulsifier addition to rice cakes led to higher
491 hardness compared to the control, while other hydrocolloids had no significant effect on
492 hardness. The staling of rice cakes is accompanied by starch retrogradation, but addition of

493 alginate increased the DSC enthalpy for melting retrograded amylopectin, whilst decreasing
494 hardness. The microstructure of rice cakes with and without alginate is similar, however after
495 staling, the alginate samples had a smaller reduction in water mobility and more
496 inhomogeneity than in the control, which might be related to the reduced hardness. We
497 hypothesise and provide evidence to suggest that the properties of high-moisture steamed
498 rice cakes are due to a phase-separated system containing a low water mobility included
499 phase of stiffened starch granules and a continuous soft aqueous phase containing added
500 alginate with high water mobility. The continuous phase apparently plays a key role in
501 determining hardness and storage ability of steamed rice cakes. More generally, for high
502 moisture starchy foods, anti-staling agents function by different mechanisms in high moisture
503 starchy foods than for the well-studied but lower moisture bread systems.

504

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641

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Figures

643 Figure 1 Compressive stress-strain curves: (a) full strain range, regions from left to right are region
644 for partial contact, linear region before breaking, breaking region, and linear region after breaking;
645 (b) linear region before breaking for modulus calculation

646 Figure 2 Firmness of rice cakes with added anti-staling agents: (a) enzyme and emulsifiers; (b)
647 hydrocolloids. DM: distilled monoglyceride; SSL: sodium stearyl lactylate; CMC: carboxymethyl
648 cellulose.

649 Figure 3 Confocal laser scanning microscopy images of control (a1-3), 0.3% alginate (b1-3) added rice
650 cake samples on storage time of day 0, 2 and 7. c: polarised light microscopy image of alginate
651 added sample on day 0. Starch were stained by FITC to green, proteins were stained by Rhodamine B
652 to red, and the non-starch polymers, mainly alginate and rice cell wall material were stained by
653 Calcofluor White to blue..

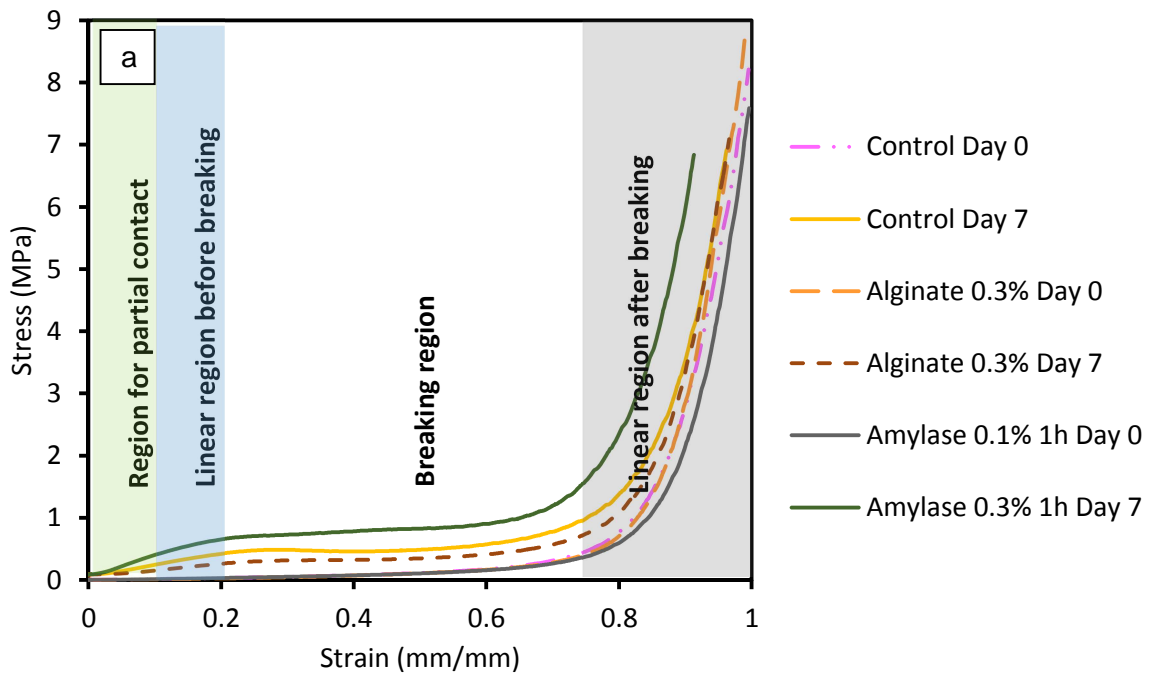
654 Figure 4 Moisture contents of rice cakes of control and alginate 0.3% addition groups

655 Figure 5 The proton-density (a), T1 (b) and T2-weighted (c) magnetic resonance images of control
656 and 0.3% alginate added steamed rice cakes.

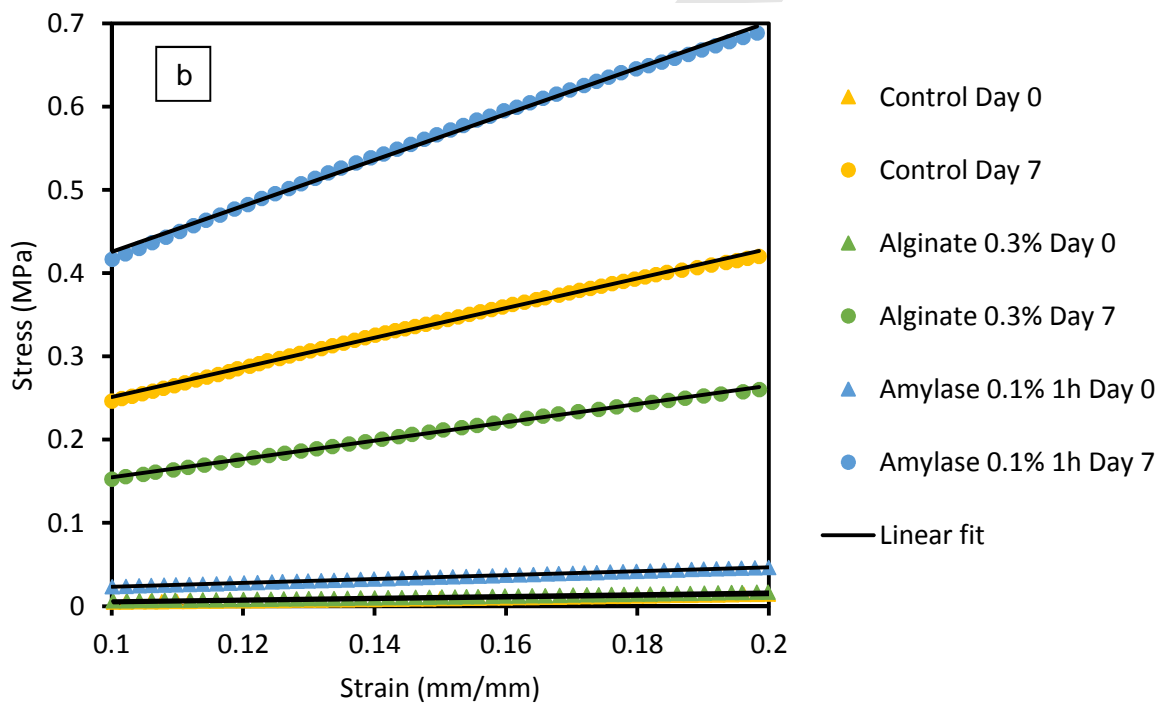
657 Figure 6 Pixel intensity distribution of proton density (a), T1 (b) and T2 (c) images as a function of
658 distance from rice cake centre.

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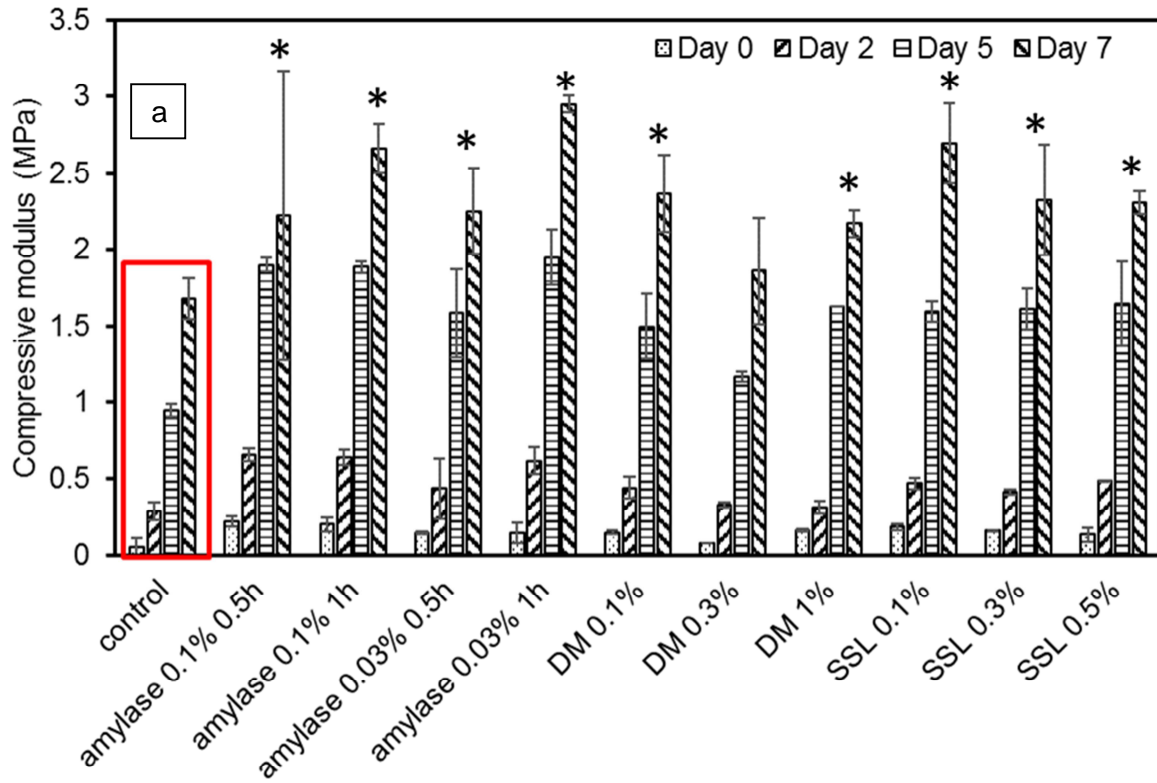
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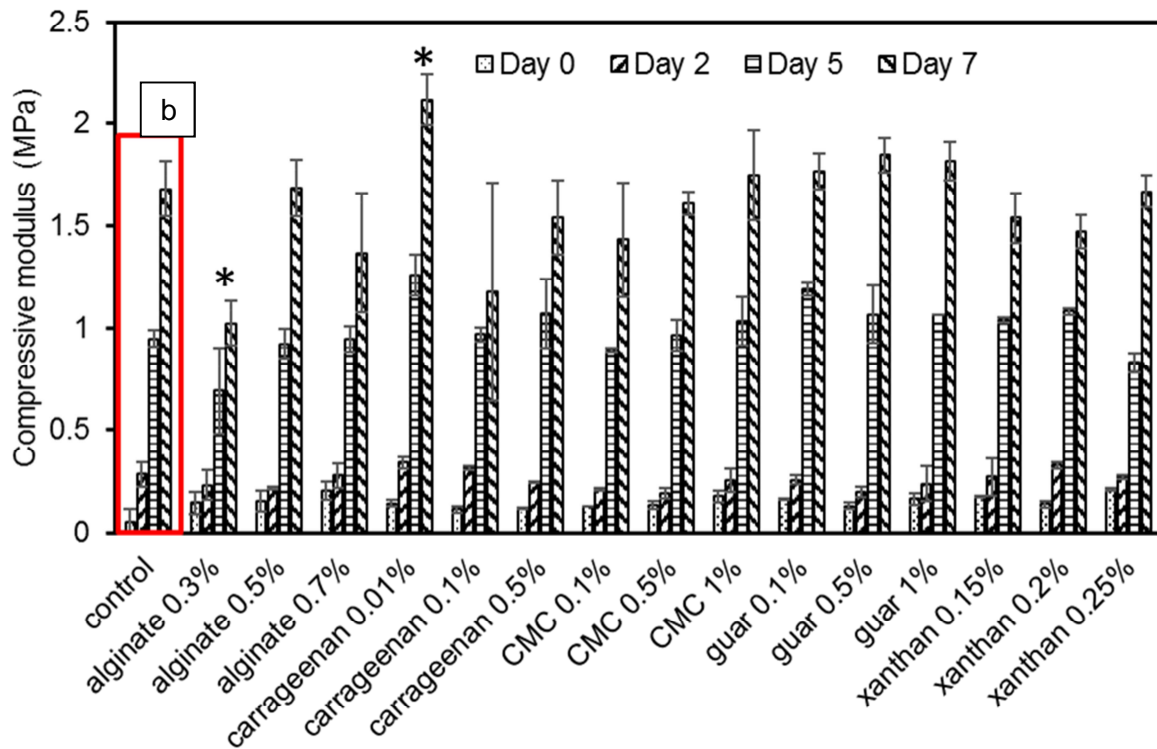
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Figure 1 Compressive stress-strain curves: (a) full strain range, regions from left to right are region for partial contact, linear region before breaking, breaking region, and linear region after breaking; (b) linear region before breaking for modulus calculation



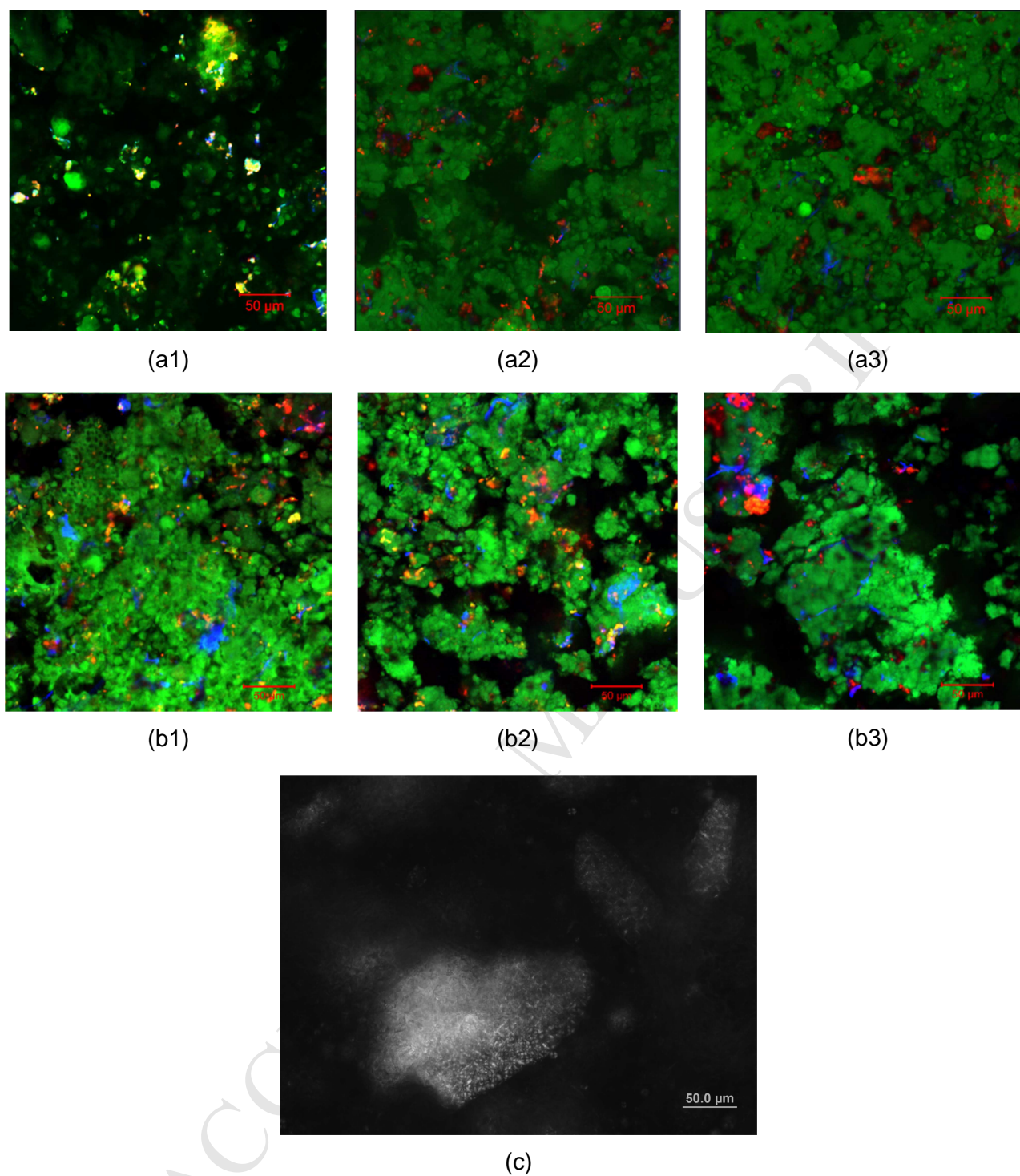
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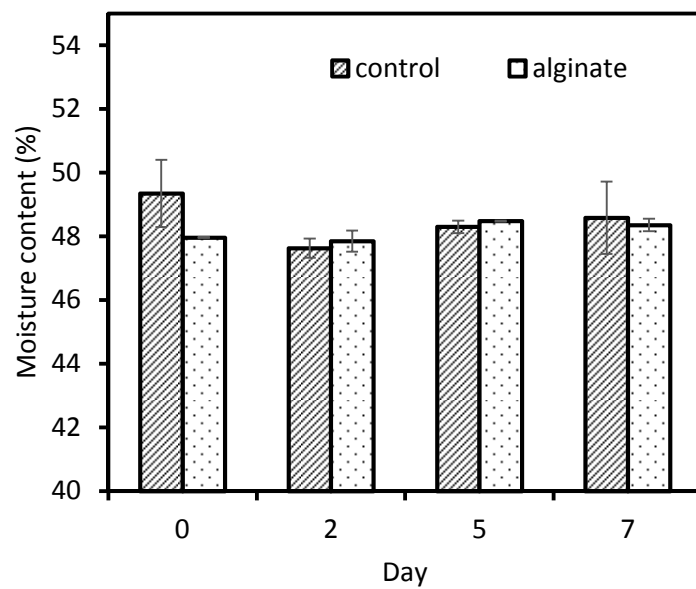
668 Figure 2 Firmness of rice cakes with added anti-staling agents: (a) enzyme and emulsifiers; (b) hydrocolloids. DM:
 669 distilled monoglyceride; SSL: sodium stearyl lactylate; CMC: carboxymethyl cellulose.

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671 *Figure 3 Confocal laser scanning microscopy images of control (a1-3), 0.3% alginate (b1-3) added rice cake*
 672 *samples on storage time of day 0, 2 and 7. c: polarised light microscopy image of alginate added sample on day*
 673 *0. Starch were stained by FITC to green, proteins were stained by Rhodamine B to red, and the non-starch*
 674 *polymers, mainly alginate and rice cell wall material were stained by Calcofluor White to blue..*

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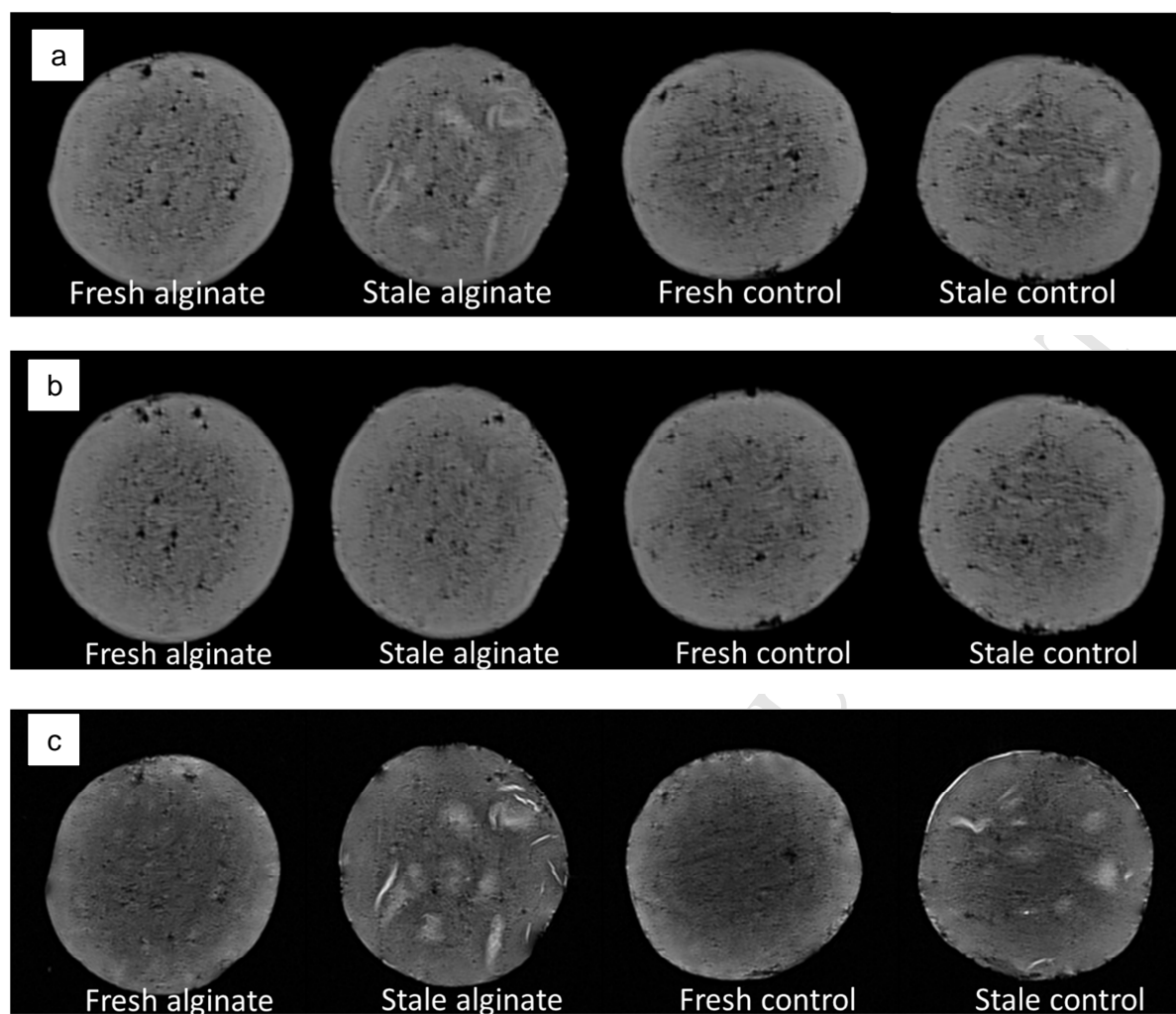


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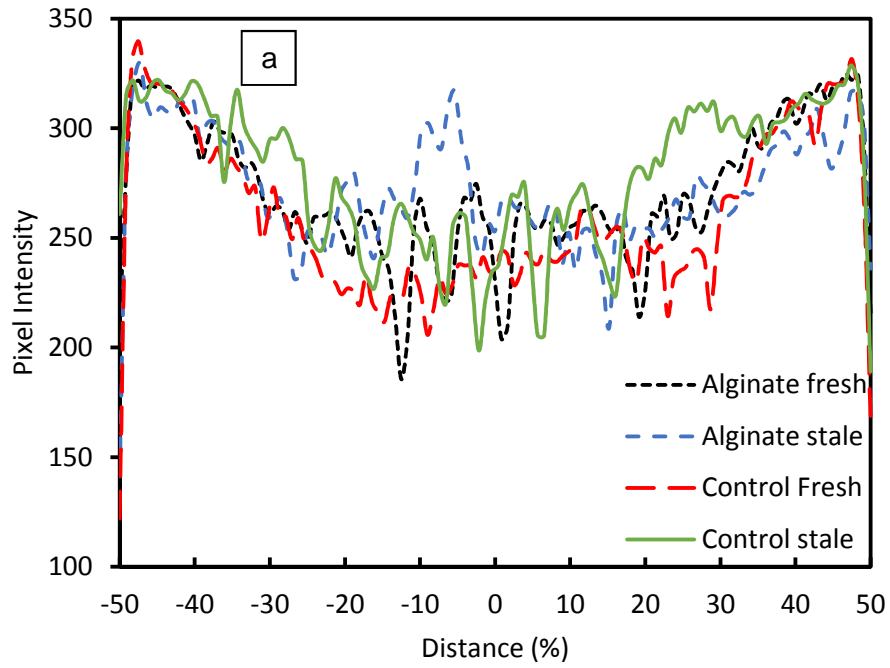
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Figure 4 Moisture contents of rice cakes of control and alginate 0.3% addition groups

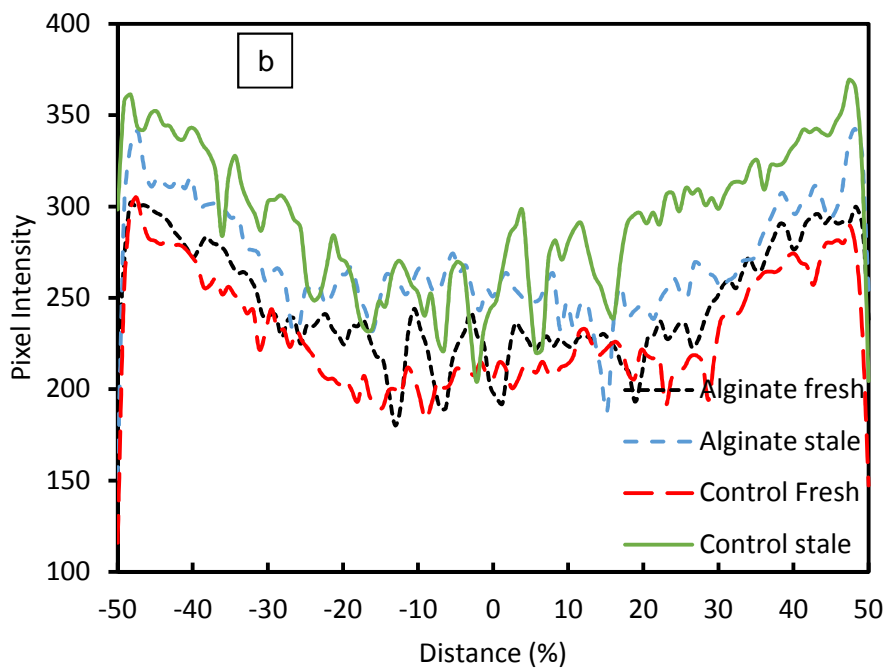


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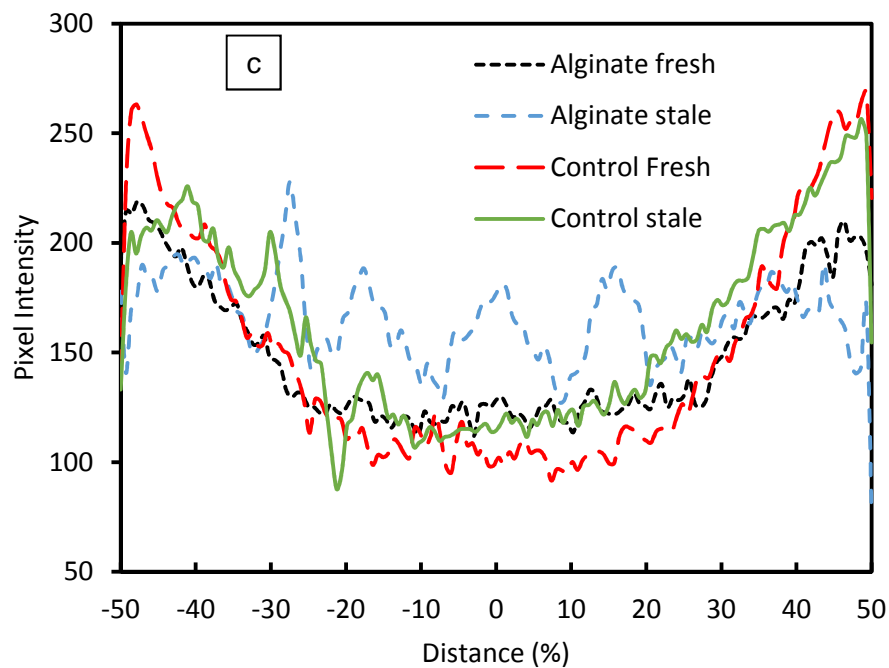
Figure 5 The proton-density (a), T1 (b) and T2-weighted (c) magnetic resonance images of control and 0.3% alginate added steamed rice cakes.



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685 Figure 6 Pixel intensity distribution of proton density (a), T1 (b) and T2 (c) images as a function of distance from
686 rice cake centre.

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Tables

688 Table 1 Additives and their concentrations used in this study

689 Table 2 DSC characteristics of rice cakes during storage for a week

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Table 1 Additives and their concentrations used in this study

Additives	Concentrations tested (w/w rice flour)		
Xanthan	0.15%	0.2%	0.25%
Guar gum	0.1%	0.5%	1%
Carrageenan	0.01%	0.1%	0.5%
CMC	0.1%	0.5%	1%
Alginate	0.3%	0.5%	0.7%
Fungal α -amylase	0.03%	0.1%	Incubation time 0.5h, 1h
SSL	0.1%	0.3%	0.5%
DM	0.1%	0.3%	1%

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Table 2 DSC characteristics of rice cakes during storage for a week

Storage time (day)	Retrograded amylopectin				
	T _o (°C)	T _p (°C)	T _c (°C)	ΔH (J/g)	
0	-	-	-	-	
Alginate 0.3%	2	52.5±1.1	62.2±1.7	75.1±0.6	0.37±0.09
	5	46.9±0.1	58.5±1.1	73.4±0.4	2.00±0.27
	7	48.2±0.7	61.5±0.2	78.3±1.1	2.94±0.51
0	-	-	-	-	
Control	2	58.7±5.0	68.3±1.7	77.5±0.8	0.19±0.17
	5	56.0±2.3	66.2±1.2	76.7±1.0	0.47±0.39
	7	52.9±1.2	64.2±0.8	77.2±0.5	1.35±0.23

694 “-” indicates no peak detected.

Highlights

Effects of anti-staling treatments used in baking evaluated for steamed rice cakes

Alginate showed the most effective anti-firming effect but promoted starch retrogradation

Anti-firming mechanism proposed to be due to structuring of the soft continuous phase

Hydrocolloid anti-staling mechanisms are different for high and low moisture foods