

Aphidicidal potential of *Azadirachta indica*, *Adhatoda vasica*, *Vitex negundo*, *Parthenium hysterophorus* and *Lantana camara* against mustard aphid, *Lipaphis erysimi* Kaltenbach (Hemiptera:Aphididae)

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Received: December 17, 2010; Revised received: March 16, 2012; Accepted: August 10, 2012

Abstract: Use of eco-friendly naturally occurring plant products in the management of the mustard aphid, *Lipaphis erysimi* Kaltenbach (Hemiptera:Aphididae) under laboratory conditions was carried out. The results revealed that the nymphs and adults mortality of mustard aphid varied significantly with three different concentrations (0.5, 1.0 and 2.0%) of five plant products. The mortality was high with seed kernel extracts of neem, *Azadirachta indica* killed nymphs and adults of *L. erysimi* (70.82 %) followed by leaves extracts of lagundi, *Vitex negundo* Linn. (69.40 %), leaves extract of *Parthenium hysterophorus* Linn. (65.38 %), leaves extract of arusa, *Adhatoda vasica* Nees. (55.81%), aerial parts of aripple, *Lantana camara* Linn. (51.70%) and untreated control (1.5 %), respectively. The seed kernel extracts of *A. indica* differed significantly from the remaining ones except *V. negundo* from which it does not differ significantly to one another. The effects of carrot grass, *P. hysterophorus* was recorded moderate aphidicidal action while *L. camara* caused the lowest mortality of nymphs and adults of *L. erysimi* among all treated natural products. Of the five plant products tested *A. indica* performed better under all the experimental conditions where as untreated control gave only 01.5% aphid mortality. The concentrations 2.0% are superior to concentration 1.0% and 0.5% concentrations. Similarly maximum aphid mortality was observed after 24 hours of exposure and minimum after 6 hours. The exposure periods of 24 hours was significantly superior to 12 hours and 6 hours in both control and plant products.

Keywords: *Adhatoda vasica*, Aphid mortality, *Azadirachta indica*, *Lipaphis erysimi*, *Parthenium hysterophorus*, *Vitex negundo*

INTRODUCTION

Plant - derived extracts and photochemical have been intensively investigated for the past thirty years in an effort to develop alternative to conventional insecticides but with reduced health and environmental impacts. Synthetic insecticides can leave potentially toxic residues in food products and can be deleterious to non- target organisms in the environment (Isman, 2006). Among the plant families studied, the Meliaceae, Asteraceae, Annonaceae, Acanthaceae, Verbenaceae, Rutaceae, Pipraceae, are among the most promising : Roomi and Ariquiddin (1977), Joshi *et al.* (1978), Graingage *et al.* (1985), Devakumar *et al.* (1986), Zehnder and Warthen (1988), Jacobson (1989), Bathal *et al.* (1991), Nichol and Schmutterer (1991), Gujar (1997), Muthukrishna *et al.* (1997), Ranganath *et al.* (1997). Breuer and De (1999), Das *et al.* (2001), Mandal and Bhattacharya (2003), Tripathi and Singh (2003), Pinto *et al.* (2005), Mumtaz *et al.* (2006), Isman (2006), Singh and Yadav (2007), Nazrussalam *et al.* (2008), Pande *et al.* (2008), Raghuraman *et al.* (2008), Yadav and Ali (2008), Lalnuntluanga *et al.* (2008), Bhagwati *et al.* (2009).

Family crucifereae provides some of the most important oil and vegetable yielding crops in the country. Mustard was a common spice in ancient Greek and Egyptian civilizations, where it was often eaten raw, chewed with meat to mask off flavors, to aid digestion, and for its antimicrobial properties. Romans recorded mustard application as a preservative and its use in sauces with meat, fish and vegetables. There are more than twenty-four insect-pest damaged the mustard at its various stages of crop growth (Kundu and Pant, 1967). Among them mustard aphid, *Lipaphis erysimi* is polyphagous insect- pests and causes severe damage to most of plant of economy importance which may cause yield losses of 66.00% to 99.00% in *Brassica campestris* Linn. and 27.00 to 68.00 % in *B. juncea* Linn. (Gupta and Rawat, 2004). This insect-pest of mustard and rapeseed cultivated during the rabi season October to March has been reported to serious damage on vegetables like cabbage, cauliflower, turnip, radish, broccoli, kale and pulses, bean, spinach, cucumber, jute, sweet potato and many other important crops (Tripathi, 1982). Keeping this in view, use of some naturally occurring plant products under laboratory conditions was carried out

in the management of the mustard aphid, *L. erysimi* Kaltenbach (Hemiptera: Aphididae).

MATERIALS AND METHODS

Experiment was conducted to test the efficacy of five plant products with three replications in the research laboratory, Department of Zoology, D.B.S.College, Kanpur, during winter seasons of 2009-2010 against mustard aphid, *L. erysimi* Kalt. infesting mustard crop. Seed kernel extracts of neem, *Azadirachta indica*, leaves extracts of lagundi, *Vitex negundo* Linn., leaves extracts of carrot grass, *Parthenium hysterophorus* L., leaves extract of arusa, *Adhatoda vasica* Nees. and aerial parts of ariipple, *Lantana camara* Linn. were prepared in the laboratory by using the standard method suggested by Chandel *et al.* (2009).

The fresh leaves of mustard were taken from unsprayed field and washed thoroughly with tap water. The different concentrations of five botanicals were tested by dry film technique. The spraying of the insecticides was done in glass petridishes (10cm diameter) by potters spray tower, using 1.0 ml. of insecticidal preparation per petridish. Three concentrations (0.5, 1.0, 2.0 %) were tested in three replications, along with over control (emulsified water). To record the mortality, the spray petridishes were gently shaken under an electric fan until the liquid phase evaporated leaving behind a uniform coat (dry) film of herbal extract on the glass surface. The spray tower was thoroughly rinsed with the insecticide solution. The stock of mustard aphid, *L. erysimi* Kaltenbach was maintained under laboratory conditions. Ten known aged nymphs and adults of *L. erysimi* were released inside each pair of petridishes and allow remaining their upto two hours. After which, they were transferred to the fresh petridishes containing fresh food for feeding. These petridishes were kept as such under control conditions (16±2°C temp. 80±10% relative humidity) and mortality count was taken after 6, 12, 24 hours of exposure. The observed mortality %age of nymphs and adults were converted into angular transformed values, which were statistically analyzed to test the significance and compared the respective three concentrations of selected plant extracts and period with the control (Abbott, 1925). Based on the %age reduction of the nymphs and adults, the detail of each laboratory experiment against *L. erysimi* Kalt. are summarized in Table 1.

RESULTS

The data recorded on the mustard crop and vegetable damage by cabbage aphid, *L. erysimi* Kalt. as effected by different treatments revealed that all the treatments were found superior over the control in reducing the aphid population. The maximum aphid mortality (70.82 %) was observed in the treatment petridish consisting of *A. indica* seed kernel extracts (0.5, 1.0 and 2.0% concentration) while the minimum aphid mortality was recorded in untreated control (07.04%) during 2008-2009 and their average, respectively.

Table 2 indicated the effect of different concentration of plant extract on mortality %age of nymphs and adults of *L. erysimi* after 6, 12, 24 hrs of treatment in the laboratory condition. It was observed that 6 hrs of treatment, maximum mortality (67.86 %) was with 2.0 % of *V. negundo*, followed by *A. indica* (66.14) and *P. hysterophorus* (61.92). Twelve hrs after maximum mortality (90.00%) was noted with 2.0 % *A. indica*. After 24 hrs the value was highest (83.85 %) with 2.0% of *A. indica* and *V. negundo* followed by 2.0% of *P. hysterophorus* (83.85 %).

The analysis of variance indicated that control vs. treated, insecticides used, concentrations used, periods, Periods x control were highly significant. However, insecticides x control and period x insecticides x control was not significant.

The Table 3 revealed that insecticide *A. indica* A. Juss. gave the best results when compared to other phyto-extracts. Seed kernel extracts of neem, *A. indica* killed (70.82%) of nymphs and adults of *L. erysimi* followed by leaves extracts of lagundi, *V. negundo* (69.40 %), leaves extracts of carrot grass, *P. hysterophorus* (65.38 %), leaves extract of arusa, *A. vasica* Nees (55.81 %), aerial parts of ariipple, *L. camara* Linn. (51.70%) and control (1.5 %) respectively. The seed kernel extracts of *A. indica* are differs significantly from the remaining ones except *V. negundo* and, *P. hysterophorus* from which it does not differ significantly to one another. Among all the phyto-extracts, *L. camara* prove the least toxic giving moderate mortality (51.70 %) of nymphs and adults of *L. erysimi* where, as untreated control gave only (01.5%) aphid mortality.

The mortality %age of nymphs and adults of *L. erysimi* with plant extracts (Table 3; Fig. 2) are arranged in

Table 1. List of selected herbal plant materials and their respective parts.

S. No.	Botanical name	Common name	Family	Part used
1.	<i>Adhatoda vasica</i> Nees.	Arusa	Acanthaceae	Leaves Extracts
2.	<i>Azadirachta indica</i> A.Juss	Neem	Meliaceae	Seeds kernel
3.	<i>Lantana camara</i> Linn.	Ariipple	Verbenaceae	Aerial parts
4.	<i>Parthenium hysterophorus</i> L.	carrot grass	Asteraceae	Leaves
5.	<i>Vitex negundo</i> Linn.	Lagundi	Verbenaceae	leaves

Table 2. Effect of phyto-chemicals on mean mortality % age of *L. erysimi* Kalt. in the laboratory.

Treatments Herbal products	Con. %	Mean mortality % after						Mean (%)	
		6 hrs.		12 hrs.		24 hrs.		MT	TBV _M
		T ₁	TBV ₁	T ₂	TBV ₂	T ₃	TBV ₃		
<i>A. vasica</i>	0.5	37.22	36.6	48.84	56.7	59.00	73.5	44.45	49.0
<i>A. vasica</i>	1.0	41.14	43.3	50.85	60.1	70.07	88.4	54.37	66.1
<i>A. vasica</i>	2.0	54.09	67.1	63.44	80.0	77.70	95.4	68.92	87.1
<i>A. indica</i>	0.5	43.07	46.6	59.00	73.5	77.70	95.5	52.69	63.2
<i>A. indica</i>	1.0	48.84	56.7	66.14	83.6	83.85	98.8	77.71	90.1
<i>A. indica</i>	2.0	66.14	83.6	90.00	100.0	90.00	100.0	83.85	98.8
<i>L. camara</i>	0.5	35.00	32.9	46.92	53.4	50.85	60.1	44.89	49.8
<i>L. camara</i>	1.0	46.92	53.4	52.77	63.4	59.21	73.8	52.23	62.5
<i>L. camara</i>	2.0	52.77	63.4	57.00	70.3	63.93	80.7	57.99	71.9
<i>P. hysterophorus</i>	0.5	48.93	56.8	57.00	70.3	63.93	80.7	55.21	67.5
<i>P. hysterophorus</i>	1.0	54.78	65.7	61.92	77.8	75.00	93.3	66.68	84.3
<i>P. hysterophorus</i>	2.0	61.92	77.8	81.14	97.6	83.85	98.9	74.06	92.5
<i>V. negundo</i>	0.5	48.93	56.8	61.92	77.8	63.93	80.7	56.58	69.7
<i>V. negundo</i>	1.0	61.92	77.8	67.86	85.8	81.14	97.6	78.35	95.9
<i>V. negundo</i>	2.0	67.86	85.8	81.14	97.6	90.00	100.0	69.33	87.5
Control		00.00	00.0	00.00	00.0	21.14	13.04	07.04	01.5

(i) C.D. for control vs. treated = 5.0499, (ii) C.D. for insecticide means= 4.3879; (iii) C.D. for concentration means= 2.0977,(iv) C.D. for period means = 2.0813; (v) C.D. for insecticide and concentration means at the same period= 8.3183

descending order as: *A. indica* (70.82) > *V. negundo* (69.40) > *P. hysterophorus* (65.38) > *A. vasica* (55.81) > *L. camara* (51.70) > control (01.5), respectively.

Table 4 revealed that all the three concentrations differ from each other. Concentration 2.0 % was superior to concentration 1.0, and 0.5 %, respectively. The mean mortality %age was observed to be 53.48 %, 61.49% and 72.06% with 0.5%, 1.0% and 2.0% concentrations respectively. The observation of concentration means taken over all of extracts, which indicated that all the three concentrations (0.5 %, 1.0 % and 2.0 %) differed significantly from one another.

The Table 4 revealed that the concentration 2.0 % gave the maximum (60.55 %) mean mortality to the nymphs and adults of *L. erysimi* Kalt., followed by 1.0 %

concentration which gave the mean mortality %age (50.72 %) of aphids, whereas 0.5 % concentration gave only (42.63 %) mean mortality %age of *L. erysimi* after 6 hours exposure periods. After 12 hour exposure period aphid mortality increased in 2.0 % concentration (74.54 %), 1.0 % concentration (59.90 %) and 0.5 % concentration it was (54.73 %). Whereas in 24 hour exposure period 2.0. 1.0 and 0.5% concentration showed 81.09,73.85 and 63.08% aphid mortality. It can be concluded that concentrations 2.0 % are superior to concentration 1.0 % and 0.5 % concentrations.

Table 5 indicated that the maximum mean nymphs and adults mortality %age (38.60 %) of *L. erysimi* noticed was after 24 hrs and minimum after 6 hr. (29.56 %). In 6hrs. 59.13 % larvae were killed and 18 hrs later the %age

Table 3. Effect of phyto-chemicals on mean mortality % of *L. erysimi* Kalt. in the laboratory.

Treatments Herbal products	Mean mortality % after						Mean (%)	
	6 hrs.		12 hrs.		24 hrs.		MT	TBV _M
	T ₁	TBV ₁	T ₂	TBV ₂	T ₃	TBV ₃		
<i>A. vasica</i>	44.15	48.5	54.37	66.1	68.92	87.1	55.81	68.4
<i>A. indica</i>	52.67	63.2	71.71	90.2	87.18	99.7	70.82	89.2
<i>L. camara</i>	44.89	49.8	52.23	62.5	57.99	71.9	51.70	61.6
<i>P.hysterophorus</i>	55.21	62.5	66.68	84.3	74.26	92.6	65.38	82.6
<i>V. negundo</i>	59.57	74.4	70.30	88.6	78.35	95.9	69.40	87.6
Control	00.00	00.0	00.00	00.0	21.14	13.04	07.04	01.5

(I) C.D. for periods means at the same insecticide = 5.4097,(II) C.D. for insecticide means at the same periods = 4.4142

Table 4. Effect of phyto-chemicals on mean mortality %age of *L. erysimi* Kalt. in the laboratory.

Concentration	Mean mortality % after						Mean (%)	
	6 hrs.		12 hrs.		24 hrs.		MT	TBV _M
	T ₁	TBV ₁	T ₂	TBV ₂	T ₃	TBV ₃		
0.5	42.63	45.9	54.73	66.7	63.08	79.5	53.48	64.4
1.0	50.72	59.9	59.90	74.8	73.85	92.3	61.49	77.2
2.0	60.55	75.8	74.54	92.9	81.09	97.6	72.06	90.5

(i) C.D. for concentration means = 5.7633,(ii) C.D. for period concentration x period means for = 3.2367

increase was just by 11.36 %.

DISCUSSION

The finding of the study is in conformity with the earlier findings of various workers who conducted experiments in varied conditions. Ranganath *et al.* (1997) have reported Neem oil (1.2%) and Neem cake (4.0%) to be as effective as dichlorvos (0.2%) the application of molasses + Malathion and water (in ratio of 1:0.1:100) provided good control of melon fruit fly (Akhraruzzaman *et al.* 2000).

Several reports have shown toxic effect of plants on insects (Grainage *et al.*, 1985) This has been possible because the plant kingdom is a store house of diversified chemicals, which has evolved in the course of evolution (Maxwell and Jennings, 1980 and Singh, 2003). Seenivasan and Sundra Basu (2000) tested the efficacy of new insecticides against *Phyllocnistis citrella* Stainton and compared with commonly used insecticides along with neem- based formulations. Among different insecticides evaluated neem formulations viz., neem seed kernel and azadirachtin were found in causing high mortality of the larvae. Singh and Kanaujia, (2003) evaluated certain biopesticides against third instar larvae of *Spilosoma obliqua* Walk. on castor. Out of which NSKE (5.0 %) exhibited 1.44 % residual toxicity.

Similarly several reports have appeared on the toxic effect of neem on various insect pests of crop and stored grains as Dhingra *et al.* (2007) found that *Caesalpinia crista* and neem oil extract and other derivatives act as antifeedant and insecticides. Hexane and methanol extracts of *C. crista* were 2.0 and 2.6 times less effective than neem oil, and 1.75 and 1.3 times more effective than NSKE, correspondingly. Chandel *et al.* (2009) tested some plant insecticides against *Spilarctia obliqua* and

reported that 2.00 % *A. vasica* leaf extract gave 93.33 % larval mortality followed by *A. indica* seed kernel extracts (89.99 %) and *Curcuma longa* (87.77 %). Viswakarma *et al.* (2009) tested five plant products and two synthetic insecticides against larvae of *Spodoptera litura*. Among them *Beauveria bassiana*, Vanguard, neemarin and Multineem gave 18.8, 17.7, 13.3 and 10.0 % mean mortality, respectively. There fore a successful and environmentally acceptable pest management system can be developed of these bio-chemicals are identified and exploited as insect control agent. The experiment gives higher yield and posed less hazards to natural enemies and there fore indicate their suitability for inclusion in the integrated pest management.

Thus, considering the various aspects as discussed above, it is obvious that so many plant products have been investigated for their toxic effect, but information about selected phyto-products for their insecticidal properties against the nymph and adults of *L. erysimi* Kalt., which are the serious pest of cruciferous vegetable crops, is not available. It can be concluded that the mortality was high with seed kernel extracts of neem, *A. indica* killed nymphs and adults of *L. erysimi* (70.82 %) followed by leaves extracts of lagundi, *V. negundo* Linn. (69.40 %), leaves extract of *P. hysterophorus* Linn. (65.38 %), leaves extract of arusa, *A. vasica* Nees. (55.81%), aerial parts of *L. camara* (51.70%) and control (01.5%), respectively.

It can be concluded that the mortality was high with seed kernel extracts of neem, *A. indica* killed nymphs and adults of *L. erysimi* (70.82 %) followed by leaves extracts of lagundi, *V. negundo* Linn. (69.40 %), leaves extract of *P. hysterophorus* Linn. (65.38 %), leaves extract of arusa, *A. vasica* Nees. (55.81%), aerial parts of ariple, *L. camara* Linn. (51.70%) and untreated control (1.5 %),

Table 5. Effect of phyto-chemicals on mean mortality %age of *L. erysimi* Kalt. in the laboratory.

Treatments	Mean mortality % after						Mean (%)	
	6 hrs.		12 hrs.		24 hrs.		MT	TBV _M
	T ₁	TBV ₁	T ₂	TBV ₂	T ₃	TBV ₃		
Plant Extracts	49.24	57.4	57.95	71.8	69.09	87.3	58.89	73.3
Control	55.31	67.6	64.05	80.9	76.56	94.6	65.70	83.1

(Figures in parenthesis represent the transformed back values) i) C. D. for period means at control=5.2769,(ii) C.D. for period means at treated = 2.1273 (iii) C.D. for control vs. treated at the same period = 6.3128,(iv) C.D. for period means= 2.0913.

respectively, *L. camara* Linn. (51.70%) and untreated control (1.5 %), respectively. No physical injury was noticed during experiments.

ACKNOWLEDGEMENT

The authors are grateful to Dr. N. D. Pandey, Ex- Professor and Head, Division of Entomology, C. S. Azad University of Agriculture and Technology, Kanpur, for help during experimental work. The work reported here has been financially supported by U.G.C., New Delhi by way of Major Research Project sanctioned to principal author for which the authors are thankful.

REFERENCES

- Abbott, M.S. (1925). A method of computing the effectiveness of an insecticide. *Journal of economic Entomology*, 18 :265-270.
- Aktharruzzaman, M., Alam, M., Ali, Z. and Sardar, M. M. (2000). Efficiency of different bait sprayer for suppressing fruit fly on cucumber. *Bulletin of the Institute of Tropical Agriculture, Kyushu University*, 23 : 15-26.
- Bathal, S.S., Singh, D., Bhillon, R.S., Nayyar Singh, K. and Singh, D. (1991). Ovicidal effects of neem oil and plant extract of *Ageratum conyzoides* Linn. on *Dysdecus koenigii* Fab. *J. Insect. Sci.*, 4 (2) : 1852
- Bhagwati, B., Deka, M.K. and Patgira, P. (2009). Bio-efficacy of botanicals against banana pseudostem borer, *Odoiporus longicollis*. *Ann. Pl. protec. Sci.*, 17 (2) 366-369.
- Breuer, M. and De, L.A. (1999). Field studies on the efficacy of meliaceous plant preparations against the oak processionary, *Thaumetopea processionea* Linn. (Lepidoptera: Thaumetopoeidae). *Mededelingen Facultiet Landbouwkundigeen Toegepaste Bilogische Watenschapper Universiteiet Gent*. 64 (3a): 311-317.
- Chandel, B.S, Vajpai, R., Vajpai, Shail and Rajni (2009). Bioefficacy of botanicals against *Spilarctia obliqua*. *Ann. Pl. protec. Sci.*, 17 (2): 465-466.
- Das, A. N. Mukherjee, S.K. and Sontakke, B.K. (2001). Efficacy of some commercial neem formulation against major pests of rice and their safely to natural enemies. *Pest management & Economic Zoology*, 9 (1) : 59 – 64.
- Devakumar, C., Saxena, B.S. and Mukharjee, S.K. (1986). Evaluation of neem, *Azadirachta indica* A. Juss. liminoids and azadirachtin against sawflower aphid, *Dactynotus carthami*. *Indian J. Ent.*, 48 (4): 467-470.
- Dhingra, S., Srivastava, C., Haider, J., Bhandari, J.K.S. and Jha, A.N. (2007). Efficacy of different extracts of *Caesalpinia crista* as grain protectant against *Callosobruchus chinensis* Linn. and their effects on seed germination. *Indian J. Ent.*, 69 (2): 131-134.
- Gujar, G.T. (1997). Biological effects of azadirachtin and plumbagia on *Helicoverpa armigera*. *Indian J. Ent.*, 59 (4): 415-422
- Gupta, M.P. and Rawat, G.S. (2004). Evaluation of neem products and their admixture with insecticides against budfly incidence in linseed. *Ann. Pl. Protec. Sci.*, 12 : 1-4.
- Graingage M., Ahmed, S., Mitchell, W.C. and Hylin, J. (1985). Plant species repeatedly possessing pest control properties. *An EWVI/VH Data base resource system Institute East – West Centre, Honolulu, Hawaii, U.S.A.*
- Isman, M.B. (2006). Botanical insecticides, deterrents and repellents in modern agriculture and an increasingly regulated world. *Annu. Rev. Entomol.*, 51:45-56.
- Jacobson, M. (1989). Botanical insecticides. Past, present and future. In: Arnason, J.T., Philene.B.J.R., Morand, P. (Eds.). *Insecticides of Plant Origin*. American Chemical Society Symposium, Series No.387, Wshington, D.C. pp.1-10.
- Joshi, B.G., Prasad R. and Satyanarayana, S.V.V. (1978). Relative efficacy of neem kernel fentin acetate and fentin hydroxide as antifeedants against tobacco caterpillar, *Spodoptera litura* Fabr. in the nursery. *Indian Jour. Agric. Sci.* 48 (1) : 19-22.
- Lalnuntluanga, Joseph and Singh., H.K. (2008). Performance of certain chemicals and neem formulations against ginger shoot bore (*Dichocrocis punctiferalis* Guen.). *Indian Journal of Entomology*, 70(2) : 183.
- Kundu, S.S. and Pant, N.C. (1967). Studies on *Lipaphis erysimi* (Kalt.) with special reference to insect plant relationship. I. susceptibility of different varieties of *Brassica* and *Eruca sativa* to mustard aphid infestation. *Indian Journal of Entomology*, 29: 24-251.
- Mandal, P. and Bhattacharya, A.K. (2003). Azadirachtin, the most potent neem derivative: effect on *Spilosoma obliqua*. *Indian J. Ent.*, 65 (2): 170-176.
- Maxwell, G.F. and Jennings, R.P. (1980). Breeding plant resistance to insect. *New York, Chcheter, Bribane, Toronto, Jonn Wiley & Sons P.* 683.
- Mumtaz, R., Sunil Kumar and Khan, M.A. (2006). Evaluation of *Azadirachta indica* and *Mentha piperata* extracts against *Chilo auriculus*. *Annals of Plant Protection Science*, 14 (2) : 287-293.
- Muthukrishna, J., Pushpalatha, E. and Kasthuribhai, A. (1997). Biological effect of four plant extracts on *Culex quinquefasciatus* Say. larval stages. *Insect. Sci. Applic.*, 17 (3/4): 389-394.
- Nazrussalam, Ansari, M.S., Ali, H. and Ahmad, T. (2008). Efficacy of multineem and NSKE with insecticides for management of *Amrasca biguttula biguttula* and *Earies villevella* on okra. *Ann. Pl. Protec. Sci.*, 16 : 17-20.
- Nichol, C.M.Y. and Schmutterer (1991). Contact effect of seed oil from the neem tree *Azadirachta indica* (A. Juss.) on nymphs of the gregarious phase of the desert locust, *Schistocerca gregaria* (Forsk.). *Journal of Applied Entomology*, 111:197-205.
- Pande., R., Dnyaneshwar., Firke, M., and Yadav, S. (2008). Effect of NSKE with additives, synthetic and bio-pesticides against *Spilarctia obliqua*, defoliators of soyabean. *Ann. Pl. Protec. Sci.*, 16 (2): 488-490.
- Pinto, A.C., Cordeiro, M.C.R., Anndrade, S.R.M., Ferreira, F.R., Filgueiras, H.A., Alves, R.E. and Kinpara, D. J. (2005). *Annona* species. International center for under utilized crops., University of Southampton, Southampton, U.K.
- Raghuraman, M., Birah Ajanta and Gupta, G. P. (2008). Management of *Helicoverpa armigera* in chickpea with botanical formulations. *Indian J. Ent.*, 70 (2) :118-122.
- Ranganath, H.R., Suryanarayanarayana, M.A. and Vennakumari, K.(1997). Management of melon fly (*Bactrocera Zeugodacus*) cucurbitae in cucurbits in south Andaman. *Insect Enviroment*, 3 : 32-33.
- Roomi, M.W. and Ariquiddin, M. (1977). Observation on the repellency of neem plant *Azadirachta indica* A. Juss. on

- some stored again pests from Pakistan. *Z. Angew Ent.*, 84 (2): 124-129.
- Singh, S.C. (2003). Effect of neem leaf powder on infestation of the pulse beetle, *C. chinensis* in stored khesari. *Indian J. Ent.*, 65 (2): 188-192.
- Singh, S.S. and Yadav, S.K.(2007). Comparative efficacy of insecticides, biopesticides and neem formulations, against *Helicoverpa armigera* on chick pea. *Ann. Pl. Protec. Sci.*, 15 : 299-302.
- Singh, A.N. and Kanaujia, K.R.(2003). Residual toxicity of some biopesticides against *Spilarctia obliqua* Walker on castor." *Indian J. Ent.*, 65 (2): 297-298.
- Sreenivasan, G and Sundara Babu, P.C. (2000). Comparative efficacy of neem products against brinjal leaf hopper, *Amrasca biguttula biguttula*. *Indian J. Ent.*, 62 (1): 18-23.
- Tripathi, N.L.M. (1982). Developmental behaviour of *Lipaphis erysimi* (Kaltenbach) on *Brassica campestris* var. *toria* and evaluation of some insecticides for its control. M.Sc. (Agric.) Thesis, G.B. Pant University of Agriculture and Technology, Pantnagar pp. 107.
- Tripathi, M.K. and Singh, H.N. (2003). Synergistic potential of neem seed powder extract to susceptibility of several insecticides in *H. armigera* larvae in India. *Indian J. Ent.*, 65 (3): 373-378.
- Viswkarma,R., Deepa, M.and Kumari, K. (2009). Mortality response of *Spodoptera litura* against biopesticides and insecticides. *Ann. Pl. protec. Sci.*, 17(2) 478-479.
- Yadav, A. and Ali, S. (2008). Bio-efficacy and economics of neem based formulations against *Dasyneura lini* Barnes. *Ann. Pl. protec. Sci.*, 16(2) : 350-352.
- Zehnder, Geoffrey and Warthen, J. D. (1988). Feeding inhibition and mortality effects of neem seed extract on the Colorado potato beetle (Coleoptera: Chrysomelidae). *Jour. Econ. Ent.*, 81 (4): 1040-1044.