

planar interactions stabilize the GQ. Adapted from Phan AT et al. (2007). (d) Transcriptionactivated GQ formation in gene promoter. Adapted from Zhang C et al. (2013).

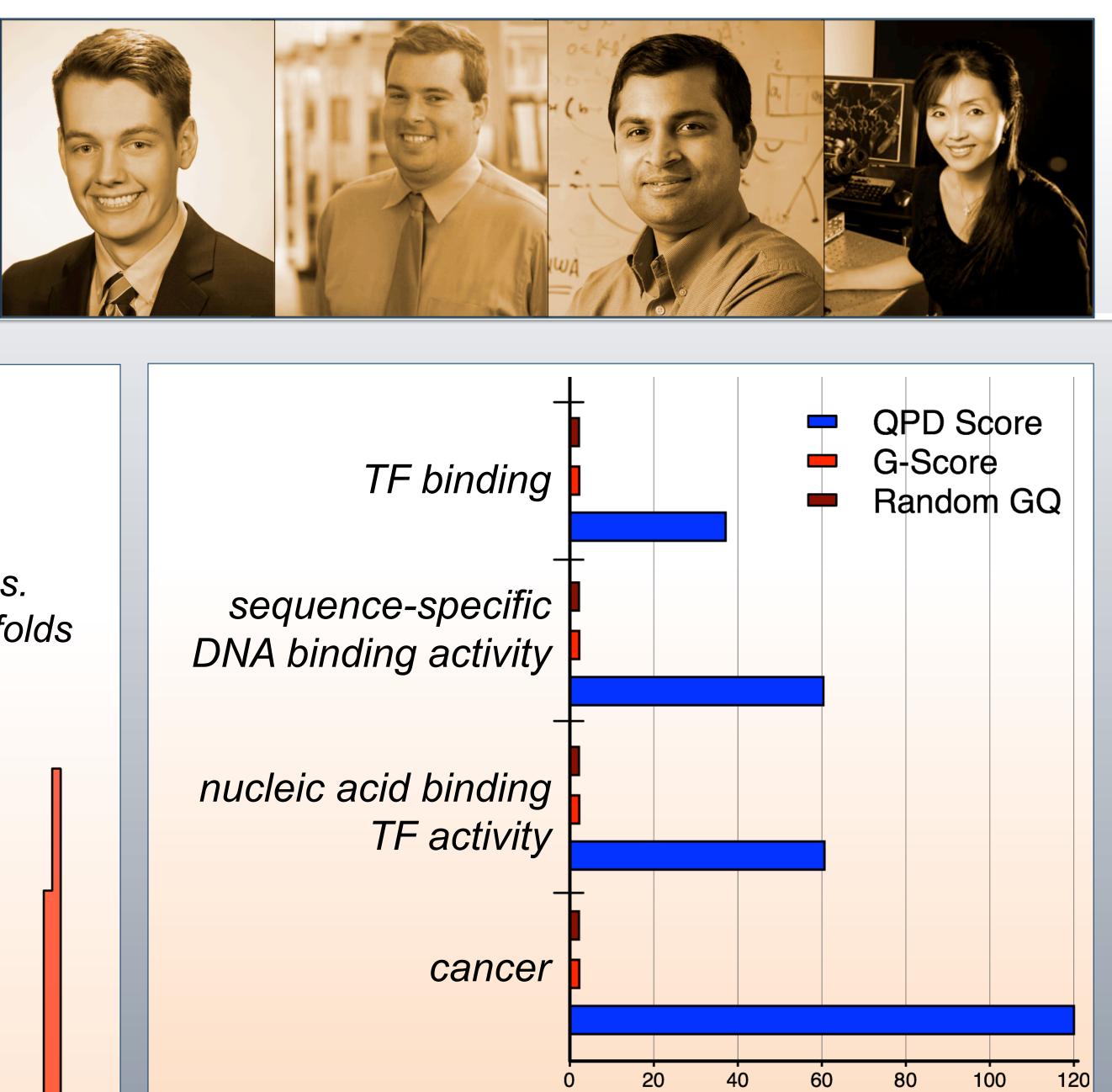
Predicting G-quadruplex Formation

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Figure 2. (a) GQ sequence motif. N represents any base, N_{1-9} are called intervening sequences. (b) Comparison of base composition and (c) the total intervening length (sum of three N_{1-9} loops).

tion **b** 0.03 b Rate Φ



Results

Q: How are the pull-down data used?

A: We use a probabilistic model to detect the unique features of the pulled-down sequences. We translate the probability that a sequence folds into a score, which we call the "QPD Score."

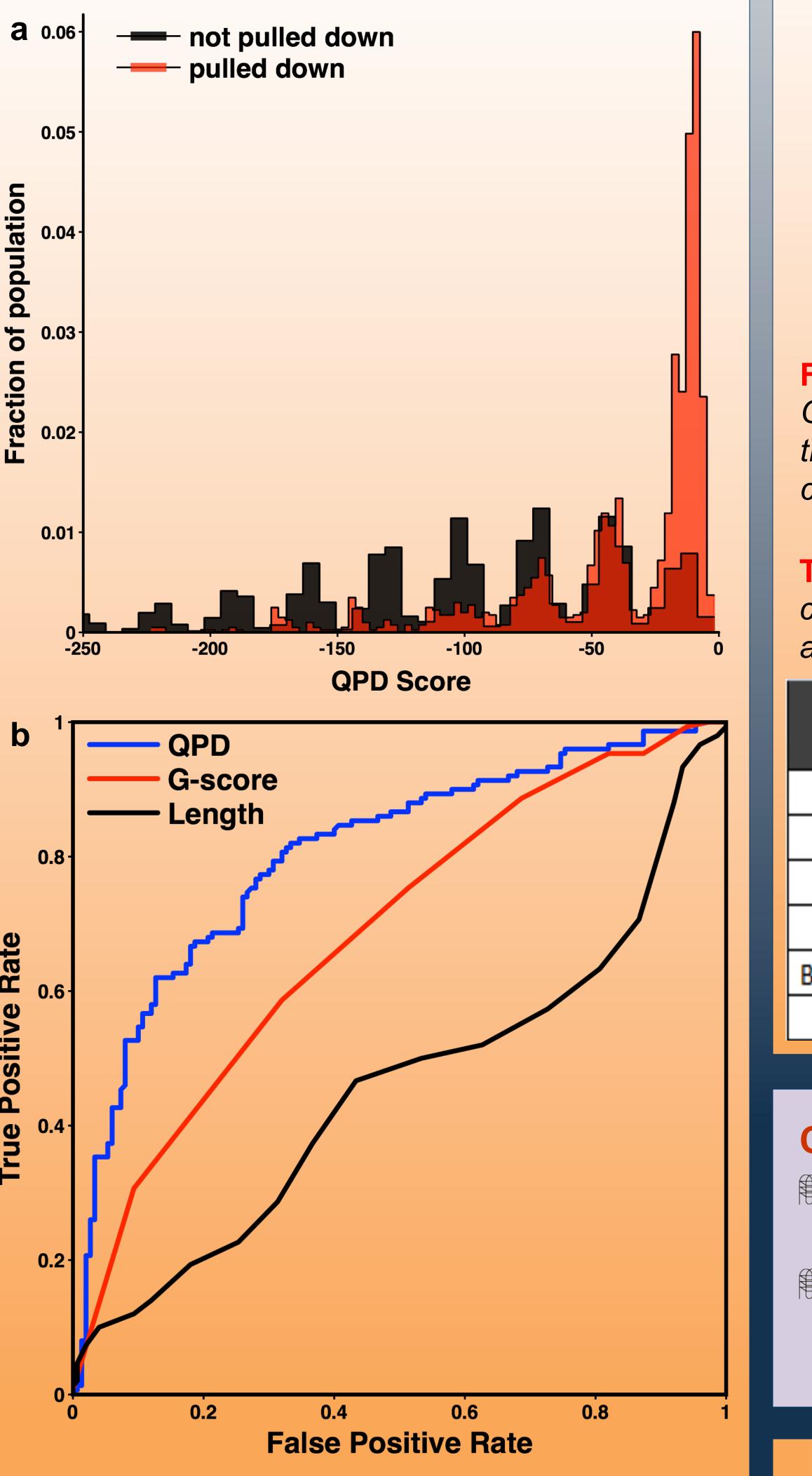
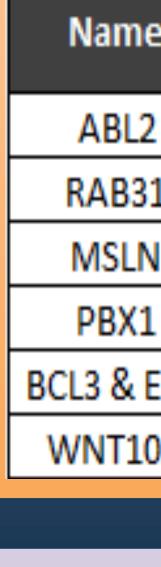


Figure 3. (a) Distribution of QPD scores for all genomic GQs. (b) ROC curve comparison of QPD score against two existing methods for predicting GQ folding. Plot was generated with sequences not included in the training set.

Figure 4. Selected ontologies. High QPD-scoring GQs tend to localize near genes of importance to transcription factor (TF) activity, regulation, and cancer. Sequences chosen in other ways do not.



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p-value (-log10)

 Table 1. A selection of genes whose promoters
contain GQs predicted to fold by QPD. Sequences are listed with G_3 omitted.

e	Sequence	G-Score Percentile	QPD Percentile
2	AAGGAAA	54	98
1	TAGTAGA	69	99
N	TTGAAGT	69	99
L	AATAGTAGT	82	97
ERG	AA	95	99
DA	TTG	95	99

Conclusions

Our model outperforms existing methods of GQ folding prediction.

Highly-scoring sequences localize near genes important in regulation and disease