Thrifty Phenotype vs Cold Adaptation: Trade-offs in Upper Limb Proportions of Himalayan Populations of Nepal

Stephanie Payne¹, Rajendra Kumar BC², Emma Pomeroy³, Alison Macintosh¹, Jay Stock^{1,4}

¹Department of Archaeology, University of Cambridge, UK

²Pokhara University Research Centre, Nepal

³School of Natural Sciences and Psychology, Liverpool John Moores University, UK

⁴Department of Anthropology, University of Western Ontario, London, Ontario, Canada

Keywords: human adaptation, altitude, energetic stress

SUMMARY

The multi-stress environment of high altitude has been associated with growth deficits in humans, particularly in zeugopod elements (forearm, lower leg). This is consistent with the thrifty phenotype hypothesis, which has been observed in Andeans, but has yet to be tested in other high altitude populations. In Himalayan populations, other factors, such as cold stress, may shape limb proportions. The current study investigated whether relative upper limb proportions of Himalayan adults (n=254) differ between highland and lowland populations, and whether cold adaptation or a thrifty phenotype mechanism may be acting here. Height, weight, humerus length, ulna length, hand length, and hand width were measured using standard methods. Relative to height, total upper limb and ulna lengths were significantly shorter in highlanders compared to lowlanders in both sexes, whilst hand and humerus length were not. Hand width did not significantly differ between populations. These results support the thrifty phenotype hypothesis, as hand and humerus proportions are conserved at the expense of the ulna. The reduction in relative ulna length could be attributed to cold adaptation, but the lack of difference between populations in both hand length and width indicate that cold adaptation is not shaping hands proportions in this case.

INTRODUCTION

Life at high altitude is associated with extreme environmental stresses [1-8]. Hypoxia, low temperatures, a physically demanding lifestyle, and nutritional constraints create a multi-stress environment which is inhospitable to longer term occupation by many human populations [8]. Populations who reside permanently in high altitude regions have adapted to deal with the extreme stresses. Quantitatively different phenotypes have developed across the globe in high altitude regions, demonstrating that multiple adaptive pathways have evolved to deal with high altitude stresses [4-9] (Table 1). Hypoxia is one of the few environmental stresses that cannot be effectively buffered by cultural adaptation [10], and so adaptive responses to hypoxia must occur through biological pathways to enable long term survival of populations at high altitude [4,6,8]. High altitude populations have evolved efficient mechanisms for dealing with hypoxia (Table 1), but energetic deficits associated with life at high altitude often result in trade-offs during growth, creating a different phenotype from lowland populations [11–20].

Plastic Growth

Linear growth during infancy and childhood appears to be moderately reduced with increasing altitude in Andean and Himalayan populations relative to their lowland counterparts [15,21–23], likely due to developmental plasticity. This height deficit has been commonly attributed to hypoxic stress, whereby limited oxygen compromises growth [15,16,18,19,24–28]. However recent evidence suggests that oxygen saturation does not correlate with height in high altitude Andean populations, indicating that nutrition and socioeconomic factors may play a more important role in stunted growth patterns [13,18]. Indeed, it is likely to be multiple high altitude-related stresses contributing to reduced growth in high altitude populations.

Clarifying where in the body the reduction in growth occurs is a strong indicator of the reason behind reduced height. The most significant decrement in height relative to lowland populations occurs in tibial

growth, whilst sitting height remains the same [14,29]. The reduction in tibia length is mirrored by a reduction in radius length in some Andean populations [17], although this currently remains untested in Himalayan populations. This relative reduction in zeugopod length with altitude has been attributed to a thrifty phenotype mechanism [30], whereby exposure to environmental stress during early life can lead to growth trade-offs between different body elements. In an Andean population, autopod lengths (hands and feet) were seen to be conserved at the expense of other limb segments (forearm and lower leg) [17]. The authors argued that this pattern preserved function in the hands and feet, and that this pattern was inconsistent with the alternative distal blood flow hypothesis [31], which would predict a gradient of decreasing relative distal segment length with increased distance from the body as a result of progressively reduced nutrient availability. It remains untested whether the same pattern of relative size in different segments of the extremities is observed in high altitude Himalayans. Greater cold stress in the Himalayas may result in different limb proportions from those of Andeans.

Potential Cold Adaptation

Whilst both the Himalayas and the Andes have considerable local variation in temperature and humidity, high altitude populations in the Himalayas are exposed to lower temperatures on average compared to Andeans due to differences in latitude, topography, rainfall and ecology [32]. The highland populations of Peru, Ecuador, and Bolivia, residing up to as high as 4500m above sea level, are likely to experience limited seasonality, but a significant range in diurnal temperature [33]. During winter, highlanders in cold, arid regions such as Oruro, Bolivia, will tend to experience daily temperatures such as 5-10°C, with minimum temperatures dropping to around -10°C. Minimum temperatures are significantly lower in Himalayan settlements, reaching below -40°C in winter [34,35]. These lower temperatures may be greater selection pressures for good thermoregulation and minimising risk of cold injury, and thus thermal selection pressures may have shaped the limb morphology of Himalayan populations unlike other high altitude populations. Himalayan limb morphology may resemble the cold adapted patterns found in other populations exposed to low temperatures [36], such as shorter and broader first metacarpals in individuals residing in cold climates than individuals from hot climates. This supports Allen's rule [37], where appendage length is reduced and appendage breadth increased to reduce heat loss in a cold climate.

Thus, applying Allen's rule to predict limb proportions in Himalayan populations, we would expect them to have shorter and broader limbs to minimise heat loss. Minimising heat loss would reduce energetic demands on the body from maintaining body temperature, which may well be selected for as energetic stress is already strong in these populations as a result of multiple altitude-related stresses. Furthermore, low temperatures would also put individuals at greater risk of cold injury in the extremities [38,39]. Although there are individually reported cases of Sherpas with frostbite [38,40], they tend to have a lower incidence than recreational mountaineers [41,42]. These findings suggest that Sherpa hands may be better adapted to life in cold conditions, but whether hand dimensions play a role remains untested. By measuring hand dimensions of a sample of Sherpas, it may be possible to infer whether both their absolute and relative hand dimensions are suited to heat preservation or not.

As the extremity proportions of permanent Himalayan populations remain poorly documented [23,43,44], it is currently not possible to infer the key environmental stresses in Himalayan high altitude upper limb morphology and how the trade-off is balanced between dexterity and thermoregulation. Thus, the current study investigates the limb proportions of highland and lowland groups from the Himalayas to determine how the multi-stress environment of high altitude influences limb morphology.

METHODS

Study Sample

The lowland population (n=71) was sampled from a migrant Tibetan community in Jawalakhel, Kathmandu, Nepal (1400 m above sea level, 27.6744° N, 85.3123° E; average minimum winter temperature= 3.1°C [34]). This community was selected as they share common genetic ancestry with the highland population

[45], and have similar diets and activity levels. The highland population (n=183) was sampled from several Sherpa communities in Namche Bazaar and surrounding villages, Nepal (3500 m+ above sea level, 27.8069° N, 86.7140° E; average minimum winter temperature= -7.9°C [34]). Each participant self-identified as Tibetan or Sherpa in the lowland and highland populations respectively, and evidence of birthplace was confirmed when possible through birth certificates or school records. A convenience sample of 254 participants between the age of 18 and 59 was measured.

Methods

Height was measured to the nearest mm using a Seca Leicester Height Measure following standard protocols with participants dressed in light clothing and unshod [3,4]. Body mass was measured to the nearest 0.05 kg using SECA-807 weighing scales (Seca, Birmingham, United Kingdom). Upper limb segment measurements were taken using Trystom anthropometer a-226 (Trystom, spol s.r. o, Czech Republic). Both humerus and ulna length were measured following standard definitions [48]. Humerus length was measured from the lateral border of the acromion to the inferior extent of the olecranon (elbow flexed at 90 degrees), while ulna length was taken from the olecranon to the head of the styloid process. Hand dimensions were measured following definitions by Davies *et al.* [49], with palm facing upwards, fingers and palm fully extended and hand flat, with dorsum of the hand resting on a horizontal surface. Hand length was measured from the level of the ulna styloid to the greatest extension of the middle finger perpendicular to the long axis of the hand. Hand width was measured as the linear distance between the radial side of the second metacarpophalangeal joint and the ulnar side of the fifth metacarpophalangeal joint. Humerus, ulna, and hand lengths were summed to give total upper limb length.

Statistical Analysis

To take account of differences in body size, upper limb segments relative to height were compared between populations. Relative segment lengths were calculated as follows:

$$Relative segment length = \frac{Absolute segment length (cm)}{Height (cm)}$$

Both absolute and relative segment lengths were analysed using independent t-tests between the highland and lowland populations. To remove any sex differences, male and female data were analysed separately. Normality was tested using the Shapiro-Wilk test on all data. All statistical analysis was carried out using SPSS 25.0 for Windows.

RESULTS

Absolute ulna length was significantly longer in lowlanders than in highlanders in both sexes (Table 2). In males, highlanders were significantly shorter in height, total upper limb length, humerus length, ulna length and hand length. Absolute hand width did not significantly differ between populations in either sex.

Relative to height, total upper limb and ulna lengths were significantly shorter in highlanders compared to lowlanders in both sexes, whilst relative hand length and width and relative humerus length were not significantly different between the two populations (Figure 1: p>0.05 for both sexes).

DISCUSSION

These results are consistent with previous findings from Andean populations [17], as relative hand and humerus proportions are conserved at the expense of the ulna. This provides further support for a thrifty phenotype mechanism in shaping limb segment proportions in the presence of high altitude stresses, and demonstrates that limb growth responds to environmental stress in Himalayan populations in a similar way to that seen in Andean populations. Whilst the current study only investigated adults (aged 18-59), it indicates that the adult phenotype reflects the pattern which develops during childhood [17].

The current study aligns with prior evidence of selective growth under environmental stress [17,50]. No difference was found in relative hand length or width between the populations, indicating that no

compromise in growth was made in hand dimensions. Relative ulna length was significantly shorter in highlanders relative to lowlanders, indicating reduced growth of this limb segment. Differences in altitude may result in this limb segment difference as limited oxygen availability may reduce growth in the highland population, as previously seen in other high altitude populations [14,51]. However, this explanation is based on hypothetical assumptions relating to prioritisation of functional elements, and thus requires further investigation to fully understand the underlying mechanisms behind the limb segment pattern found here and elsewhere [11,21,29,52]

The reduction in relative ulna length could be attributed to cold adaptation [53][54], but the lack of difference between populations in both hand length and width indicate that cold adaptation is not shaping hand proportions in this case. It is possible that the forearms, but not hand proportions, are shaped by climate; Steegman (2007) suggested that extreme vasoconstriction in the hands as a response to cold may negate any effect of hand proportions, as hand temperature may reach close to the surrounding temperature, and thus little heat is transferred to the surroundings from the hand. This is supported by cold immersion tests, whereby heat flux from the hand is consistently lower than heat flux from the forearm, even when a temporary cold-induced vasodilation response occurs in the fingers [55]. The forearm does not have such vasoregulatory responses, and thus maybe more susceptible to heat loss, and thus shortening of the zeugopod segment may have a significant effect on reducing energy expenditure via reduction in heat loss [56]. The mechanism for this adaptive limb segment shortening is unknown, but plasticity may play a role. It is well documented that temperature influences long bone elongation during postnatal development in several species, including mice [57–61], rats [62–64], rabbits [65], and pigs [66]. This plastic growth response to temperature may influence high altitude long bone proportions; however, this plasticity in response to temperature has yet to be investigated in humans.

The hand proportions measured in the current study do not appear to align with cold adaptation theory. This may be for several reasons. Firstly, cold stress may not be the dominant factor influencing limb proportions; maintenance of hand dimensions for dexterity may be acting here [67]. Evidence in the skeletal record suggests that cold adaptation theory may explain patterns in hand proportions of high latitude-dwelling populations [36], but may not be applicable to high altitude populations. The highland population in the current study may not show cold adaptation patterns in the hands as they may not be exposed to extreme low temperatures as regularly or for such prolonged periods as populations at very high latitudes and the high insolation of the Himalayas during the day may alleviate cold stress [34,35]. Alternatively, the results here may indicate that in Himalayan populations, temperature does not act on hand proportions through plastic mechanisms. As the lowland population had a shared genetic ancestry with the highland population [45], both populations may have the same genetic-based long term adaptations which shape the hands, which may or may not relate to cold adaptation. Finally, there could be other modifying factors here, such as the use of gloves or insulative clothing in highlanders to alleviate any cold stress effects, but this was not measured in our study.

The results here do not support the distal blood flow hypothesis [31], as the hand was not significantly reduced in length or width relative to the rest of the body in highlanders compared to lowlanders. This again aligns with findings from Andean populations [17]. However, this limb proportion pattern may indirectly be linked to differential blood supply to hand and forearm segments. When blood vessels are fully perfused, blood supply is greater in autopod segments than zeugopod segments, due to dense capillary networks in the hands and feet [68], where blood moves slowly and thus nutrient delivery is highly efficient. Even if there is significant vasoconstriction in the highland populations during cold exposure, there may still be sufficient nutrient delivery to the deep tissue and bones of the hands, ensuring essential bone development and regeneration [69,70]. Whether vasoconstriction negates any effect of differential blood supply requires further investigation.

Although overall the diet and activity of the two populations was similar, there may have been some differences which were difficult to quantify. Lowland individuals self-reported a traditionally Tibetan diet, but may also have had access to westernised food as globalisation has increased the diversity of food

products available in Kathmandu. Differences in activity may also have occurred; the women in both populations were homemakers and living relatively sedentary lifestyles; the men in the lowland population were factory workers, whilst the men in the highland population were porters. Whilst the men in both populations were manual labourers, energy expenditure of activity was not directly measured in this case, so any differences in activity were unknown. Previous work indicates a very high daily energy expenditure of highland porters [71]; further investigation would be required to determine the daily energy expenditure of Jawalakhel factory workers.

The significant differences between males in all absolute variables other than hand width may be due to greater sensitivity to environmental stresses in males [72]. As five different variables show the same pattern between the male populations (height, total upper limb length, humerus length, ulna length, hand length), this is unlikely to be a chance outcome. Alternatively, confounding factors such as unknown differences in diet or activity, as discussed above, may result in differences in body form between highland and lowland males. Although there is a discrepancy in sample size between males, there are no assumptions relating to sample size when applying the independent samples t-test, and thus differences in sample size should not have an effect. However, it is possible that the lack of differences identified in the female samples, other than the significant difference in relative ulna length, may result from a lack of power due to the relatively small sample sizes.

Although the absolute differences were greater in males, the differences in relative ulna length and total upper limb length were greater in females. This may indicate differential investment in segment lengths between the sexes during energetic stress, or alternatively, that the greater deficit in height in highland males reduces the relative differences in upper limb segment lengths. This outcome needs further investigation to determine why absolute differences between highland and lowland upper limb segment lengths are greater in males, but relative differences are greater in females.

Conclusion

In conclusion, the current study showed heterogeneous reductions in different upper limb segments in association with altitude-related stresses in Himalayan populations. Relative to height, total upper limb length was significantly shorter in highlanders than lowlanders, a difference driven largely by reduced ulna length. These results provide further support for the thrifty phenotype hypothesis, as hand dimensions are prioritised over other upper limb segments for their manipulative function. Cold adaptation patterns in the hand were not found in this study, indicating that other selection pressures dictate limb proportions in the Himalayan high altitude environment.

Ethics

Participation was voluntary and the study was conducted according to accepted international ethical standards for research involving human subjects (Declaration of Helsinki) [73]. The study was approved by the Human Biology Research Ethics Committee at the University of Cambridge (HBREC.2016.22), and the Nepal Health Research Council (Reference Number: 1571). Written informed consent was obtained from all participants by signature, or fingerprint if not literate.

Data Accessibility

The datasets supporting this article can be found at Dryad: doi:10.5061/dryad.25p96

Authors' Contributions

SP designed the study, acquired the data, analysed the data and drafted the article. RK assisted in data acquisition. EP and AM made significant contributions to drafting and revising the article. JS assisted in formulating study design and revising the article.

Competing Interests

The authors declare no competing interests.

Funding

This study was funded by the following institutions: National Geographic Society; British Association for Biological Anthropology and Osteoarchaeology; Sigma Xi Scientific Research Society; Trinity Hall, University of Cambridge; Department of Archaeology, University of Cambridge

Acknowledgements

We would like to thank all the participants who took part in this study, and Oliver Melvill, who helped SP collect data. The authors would like to thank the anonymous reviewers who helped to improve this paper.

Correspondence: Stephanie Payne, sp627@cam.ac.uk
Department of Archaeology, University of Cambridge, Cambridge UK, CB2 3QG, UK

- Morpurgo G, Arese P, Bosia A, Pescarmona G, Luzzana M, Modiano G. 1976 Sherpas living permanently at high altitude: a new pattern of adaptation. Proceedings of the National Academy of Sciences 73, 747–751.
- 2. Moore L, Niermeyer S, Zamudio S. 1998 Human adaptation to high altitude: regional and life-cycle perspectives. American Journal of Physical Anthropology Suppl 27, 25–64.
- 3. Moore L. 2001 Human genetic adaptation to high altitude. High Altitude Medicine & Biology 2, 257–279.
- 4. Beall. 2006 Andean, Tibetan, and Ethiopian patterns of adaptation to high-altitude hypoxia. *Human Biology* **46**, 18–24. (doi:10.1093/icb/ici004)
- 5. West JB, Milledge JS, Schoene RB, Luks A. 2013 High altitude medicine and physiology. Boca Raton: Francis Group.
- 6. Beall CM. 2014 Adaptation to High Altitude: Phenotypes and Genotypes. *Annual Review of Anthropology* **43**, 251–272. (doi:10.1146/annurev-anthro-102313-030000)
- 7. Bigham AW, Lee FS. 2014 Human high-altitude adaptation: Forward genetics meets the HIF pathway. *Genes* and Development 28, 2189– 2204.
- (doi:10.1101/gad.250167.114)
 8. Gilbert-Kawai ET, Milledge JS, Grocott MPW, Martin DS. 2014 King of the Mountains: Tibetan and Sherpa Physiological Adaptations for Life at High Altitude. Physiology 29, 388–402. (doi:10.1152/physiol.00018.20
- 9. Pawson I. 1976 Growth and development in high altitude populations: a review of Ethiopian, Peruvian and Nepalese Studies. *Proceedings of the Royal Society B: Biological Sciences* **194.** 83–98.
- 10. Beall CM, Jablonski N, Steegman A. 2012 Human Adaptation to Climate: Temperature, Ultraviolet Radiation, and Altitude. In Human Biology: An Evolutionary and Biocultural Perspective (eds S Stinson, B Bogin, D O'Rourke), pp. 177–250. John Wiley & Sons, Inc.

- 11. Pawson I. 1977 Growth characteristics of populations of Tibetan origin in Nepal.

 American Journal of Physical Anthropology 47, 473–482.
- 12. Smith C. 1997 The Effect of Maternal Nutritional Variables on Birthweight Outcomes of Infants Born to Sherpa Women at Low and High Altitudes in Nepal. American Journal of Human Biology 763, 751–763.
- 13. Weitz C a, Garruto RM, Chin C-TC, Liu JJ-C. 2004
 Morphological growth and thorax dimensions among Tibetan compared to Han children, adolescents and young adults born and raised at high altitude. Annals of Human Biology 31, 292–310. (doi:10.1080/03014460420001 96316)
- 14. Bailey S, Xu J, Feng J, Hu X, Zhang C, Qui S. 2007
 Tradeoffs Between Oxygen and Energy in Tibial Growth.

 American Journal of Human Biology 19, 662–668.

 (doi:10.1002/ajhb)
- 15. Argnani L, Cogo A, Gualdi-Russo E. 2008 Growth and nutritional status of Tibetan children at high altitude. Collegium Antropologicum 32, 807–812.
- Moore L, Charles S, Julian C.
 2011 Humans and high altitude: hypoxia and fetal growth.
 Physiology and Neurobiology 178, 181–190.
- 17. Pomeroy E, Stock JT, Stanojevic S, Miranda JJ, Cole TJ, Wells JCK. 2012 Trade-Offs in Relative Limb Length among Peruvian Children: Extending the Thrifty Phenotype Hypothesis to Limb Proportions. PLoS ONE 7, e51795. (doi:10.1371/journal.pone.005 1795)
- 18. Pomeroy E, Stock JT, Stanojevic S, Miranda JJ, Cole TJ, Wells JCK. 2013 Associations between arterial oxygen saturation, body size and limb measurements among high-altitude andean children. American Journal of Human Biology 25, 629–636. (doi:10.1002/ajhb.22422)

- 19. Pomeroy E, Stock JT, Stanojevic S, Miranda JJ, Cole TJ, Wells JCK. 2014 Stunting, adiposity, and the individual-level 'dual burden' among urban lowland and rural highland peruvian children. American Journal of Human Biology 26, 481–490. (doi:10.1002/ajhb.22551)
- Weitz CA, Garruto RM. 2015
 Stunting and the Prediction of Lung Volumes Among Tibetan Children and Adolescents at High Altitude. High altitude medicine & biology 16, 306–317.

(doi:10.1089/ham.2015.0036)

- 21. Beall C. 1981 Growth in a population of Tibetan origin at high altitude. *Annals of Human Biology* **8**, 31–38.
- 22. Beall C. 1984 Aging and growth at high altitudes in the Himalayas. In *The People of South Asia* (ed C Beall), pp. 365–385. New York: Plenum Press.
- 23. Gupta R, Basu A. 1981
 Variations in body dimensions in relation to altitude among the Sherpas of the eastern Himalayas. *Annals of Human Biology* **8**, 145–52.
- 24. Bateson P et al. 2004
 Developmental plasticity and human health. Nature 430, 419–421.

(doi:10.1038/nature02725)

- 25. Weinstein KJ. 2005 Body Proportions in Ancient Andeans From High and Low Altitudes. **585**, 569–585. (doi:10.1002/ajpa.20137)
- Julian C, Vargas E, Armaza J, Wilson M, Niermeyer S, Moore L. 2007 High-altitude ancestry protects against hypoxiaassociated reduction in fetal growth. Archives of Disease in Childhood Fetal and Neonatal Edition 92, F372-377.
- Eichstaedt C, Antao T, Cardona A, Pagani L, Kivisild T, Mormina M. 2015 Genetic and phenotypic differentiation of an Andean intermediate altitude population. *Physiological Reports* 3, e12376–e12376. (doi:10.14814/phy2.12376)
- 28. Ge R-L, Simonson TS, Gordeuk V, Prchal JT, McClain D a. 2015 Metabolic aspects of high-altitude adaptation in Tibetans. *Experimental Physiology* **100**, 1247–1255. (doi:10.1113/EP085292)

- Bailey S, Hu X. 2002 Highaltitude growth differences among Chinese and Tibetan children. In *Human Growth* from Conception to Maturity (eds G Gilli, L Schell, L Benso), pp. 237–247. London: Smith-Gordon.
- 30. Hales CN, Barker DJP. 1992 Type 2 (non-insulin-dependent) diabetes mellitus: the thrifty phenotype hypothesis. International Journal of Epidemiology 42, 1215–1222. (doi:10.1093/ije/dyt133)
- 31. Lampl M, Kuzawa C, Jeanty P.
 2003 Prenatal smoke exposure
 alters growth in limb
 proportions and head shape in
 the midgestation human fetus.

 *American Journal of Human
 Biology 15, 533–546.
 (doi:10.1002/ajhb.10140)
- 32. Barry R. 1992 *Mountain* weather and climate. 2nd edn. London: Routledge.
- 33. Thomas R, Winterhalder B.
 1976 Physical and biotic
 environment of southern
 highland Peru. In Man in the
 Andes: A Multidisciplinary
 Study of High-Altitude
 Quechua. (eds P Baker, M
 Little), Stroudsburg: Dowden,
 Hutchinson and Ross Inc.
- 34. Merkel A. 2016 Nepal Climate Data. Climate-Data.org Accessed 2.
- Vuillermoz E. 2016 Nepal Climate Observatory. EVK2-CNR Everest Pyramid GAW Station , Accessed 06.05.2016.
- 36. Betti L, Lycett SJ, Von Cramon-Taubadel N, Pearson OM. 2015 Are human hands and feet affected by climate? A test of Allen's rule. American Journal of Physical Anthropology 158, 132–140. (doi:10.1002/ajpa.22774)
- 37. Allen J. 1877 The influence of physical conditions on the genesis of species. *Radical Review* 1, 108–140.
- 38. Subedi BH, Pokharel J, Thapa R, Banskota N, Basnyat B. 2010 Frostbite in a Sherpa. Wilderness and Environmental Medicine 21, 127–129. (doi:10.1016/j.wem.2009.12.0 31)
- 39. Moore GWK, Semple JL. 2011
 Freezing and Frostbite on
 Mount Everest: New Insights
 into Wind Chill and Freezing
 Times at Extreme Altitude.
 High Altitude Medicine &
 Biology 12, 271–275.
 (doi:10.1089/ham.2011.0008)

- 40. Macdonald EB, Shrestha S, Chhetri MK, Sherpa R, Sherpa DG, Murray K, Sanati KA. 2015 Work-health needs of high-altitude mountain guides (Sherpas) in Nepal a pilot study. International Journal of Occupational Safety and Ergonomics 21, 9–14. (doi:10.1080/10803548.2015.1 017945)
- 41. Takeoka M, Yanagidaira Y, Sakai A, Asano K, Fujiwara T, Yanagisawa K. 1993 Effects of high altitudes on finger cooling test in Japanese and Tibetans at Qinghai Plateau. *International Journal of Biometeorology* 37, 27–31.
- 42. Maley MJ, Eglin CM, House JR, Tipton MJ. 2014 The effect of ethnicity on the vascular responses to cold exposure of the extremities. *European Journal of Applied Physiology* 114, 2369–2379. (doi:10.1007/s00421-014-2962-2)
- 43. Sloan A, Masali M. 1978 Anthropometry of Sherpa men. Annals of Human Biology 5, 453–458.
- 44. Tripathy V, Gupta R. 2007 Growth among Tibetans at high and low altitudes in India. American Journal of Human Biology 19, 789–800. (doi:10.1002/ajhb)
- 45. Bhandari S et al. 2015 Genetic evidence of a recent Tibetan ancestry to Sherpas in the Himalayan region. Scientific Reports 5, 16249. (doi:10.1038/srep16249)
- 46. Norton K, Olds T. 1996 The Anthropometric Profile. In Anthropometrica, pp. 33–186. Sydney, Australia: UNSW Press.
- 47. Cameron N. 2013 Essential anthropometry: Baseline anthropometric methods for human biologists in laboratory and field situations. American journal of human biology: the official journal of the Human Biology Council 25, 291–9. (doi:10.1002/ajhb.22388)
- 48. Lohman T, Roche A, Martorell R. 1988 Anthropometric standardization reference manual. Human Kinetics Books.

- 49. Davies BT, Benson a K, Courtney a, Minto I. 1980 A comparison of hand anthropometry of females in three ethnic groups.

 Ergonomics 23, 179–82. (doi:10.1080/00140138008924
- Bogin B, Smith P, Orden AB, Varela Silva MI, Loucky J. 2002 Rapid change in height and body proportions of Maya American children. American Journal of Human Biology 14, 753-761.
- 51. Greksa L. 1990 Developmental responses to high-altitude hypoxia in Bolivian children of European ancestry. *American Journal of Human Biology* **2**, 603–612.
- Singh ASP, Sidhu LS, Malhotra P, Zeitschrift S, Juni H, Singh BSP, Sidhu LS, Malhotra P. 1986 Body morphology of high altitude Spitians of North West Himalayas. Zeitschrift fur Morphologie und Anthropologie 2, 189–195.
- 53. Ruff CB. 1991 Climate and body shape in hominid evolution. *Journal of Human Evolution* 21, 81–105. (doi:10.1016/0047-2484(91)90001-C)
- 54. Wilberfoss P. 2012 Cold case: cold induced vasodilation response, and the origins of Polynesian body morphology as an adaptation to a cold environment. 1994.
- 55. Wang D, Zhang H, Arens E, Huizenga C. 2007
 Observations of upper-extremity skin temperature and corresponding overall-body thermal sensations and comfort.

 Building and Environment 42, 3933–3943.
 (doi:10.1016/j.buildenv.2006.0 6.035)
- Steegmann AT. 2007 Human Cold Adaptation: An Unfinished Agenda. American Journal of Human Biology 19, 165–180. (doi:10.1002/ajhb)
- 57. Sumner F. 1909 Some effects of external conditions upon the white mouse. *Journal of Experimental Zoology* **7**, 97–154.
- 58. Sundstroem ES. 1922 The adaptation of albino mice to an artificially produced tropical climate. I. Effect of the various factors composing a tropical climate on growth and fertility in mice. *American Journal of Physiology* **60**, 397–415.

- 59. Ogle C. 1934 Climatic influence on the growth of the male albino mouse. *The American Journal of Physiology* **107**, 635–640.
- 60. Serrat MA, King D, Lovejoy CO. 2008 Temperature regulates limb length in homeotherms by directly modulating cartilage growth. Proceedings of the National Academy of Sciences of the United States of America 105, 19348–19353.
- (doi:10.1073/pnas.0803319105)
 Serrat MA, Williams RM,
 Farnum CE. 2009 Temperature
 alters solute transport in growth
 plate cartilage measured by in
 vivo multiphoton microscopy.

 Journal of Applied Physiology
 106, 2016–2025.
 (doi:10.1152/japplphysiol.002
 95.2009)
- Chevillard L, Portet R, Cadot M. 1963 Growth rate of rats born and reared at 5 and 30 C. Fed. Proc 22, 699–703.
- Lee MM, Chu PC, Chan HC.
 1969 Magnitude and pattern of compensatory growth in rats after cold exposure. *Journal of* embryology and experimental morphology 21, 407–416.
- 64. Riesenfeld A. 1973 The effect of extreme temperatures and starvation on the body proportions of the rat. *American Journal of Physical Anthropology* **39**, 427–459. (doi:10.1002/ajpa.1330390311)
- 65. Ogle C. 1933 Animal adaptation to environmental temperature conditions. *The American Journal of Physiology* **103**, 606–628.
- 66. Weaver ME, Ingram D. 1969
 Morphological Changes in
 Swine Associated with
 Environmental Temperature.
 Ecology 50, 710–713.
- 67. Marzke MW, Marzke R. 2000 Evolution of the human hand: approaches to acquiring, analysing and interpreting the anatomical evidence. *Journal* of Anatomy 197, 121–140.
- 68. Standring S. 2008 *Upper Limb*. 40th edn. Edinburgh: Churchill Livingstone, Elvesier.
- 69. Ramasamy SK et al. 2016
 Blood flow controls bone
 vascular function and
 osteogenesis. Nature
 Communications 7, 1–13.
 (doi:10.1038/ncomms13601)

- 70. Tomlinson RE, Silva MJ. 2013 Skeletal Blood Flow in Bone Repair and Maintenance. *Bone* Research 1, 311–322. (doi:10.4248/BR201304002)
- 71. Malville NJ, Byrnes WC, Lim HA, Basnyat R. 2001 Commercial Porters of Eastern Nepal: Health Status, Physical Work Capacity, and Energy Expenditure. 56, 44–56.
- 72. Stinson S. 1985 Sex
 Differences in Environmental
 Sensitivity During Growth and
 Development. Yearbook of
 Physical Anthropology 28,
 123–147.
 (doi:10.1002/ajpa.1330280507)
- World Medical Association.
 World Medical Association Declaration of Helsinki: ethical principles for medical research involving human subjects. Journal of the American Medical Association 310, 2191–2194.
- 74. Harrison GA et al. 1969 The Effects of Altitudinal Variation in Ethiopian Populations. Philosophical Transactions of the Royal Society B: Biological Sciences 256, 147–182. (doi:10.1098/rsta.1892.0001)
- 75. Trowbridge F, Marks J, Lopez de Romana G, Madrid S, Boutton T, Klein P. 1987 Body composition of Peruvian children with short stature and high weight-for-height. II. Implications for the interpretation for weight-for-height as an indicator of nutritional status. American Journal of Clinical Nutrition 46, 411–418.
- Clegg E, Pawson I, Ashton E, Flinn R. 1972 The Growth of Children at Different Altitudes in Ethiopia. *Philosophical Transactions of the Royal Society B: Biological Sciences* 264, 403–437. (doi:10.1098/rsta.1892.0001)
- Boyer SJ, Blume FD. 1984
 Weight loss and changes in body composition at high altitude. *Journal of Applied Physiology* 57, 1580–1585.
- Haas J, Baker P, Hunt E. 1977
 The effects of high altitude on body size and composition of the newborn infant in Southern Peru. Human Biology 49, 611–628.
- 79. Greksa L. 1986 Chest morphology of young Bolivian high-altitude residents of European ancestry. *Human Biology* **58**, 427–443.

- 80. Erzurum SC et al. 2007 Higher blood flow and circulating NO products offset high-altitude hypoxia among Tibetans. Proceedings of the National Academy of Sciences of the United States of America 104, 17593–17598.
- (doi:10.1073/pnas.0707462104)
 81. Beall C, Laskowski D, Strohl K,
 Soria R, Villena M, Vargas E.
 2001 Pulmonary nitric oxide in
 mountain dwellers. *Nature* **414**,
 411–412.
- 82. Beall CM, Laskowski D, Erzurum SC. 2012 Nitric oxide in adaptation to altitude. Free Radical Biology and Medicine 52, 1123–1134. (doi:10.1016/j.freeradbiomed.2 011.12.028)
- 83. Xing G et al. 2008 Adaptation and mal-adaptation to ambient hypoxia; Andean, Ethiopian and Himalayan patterns. PLoS ONE

 (doi:10.1371/journal.pone.000 2342)
- 84. Beall CMCCM et al. 1997
 Ventilation and hypoxic
 ventilatory response of Tibetan
 and Aymara high altitude
 natives. American Journal of
 Physical Anthropology 104,
 427–447.
- 85. Beall C, Strohl K, Gothe B, Britenham G, Barragan M, Vargas E. 1992 Respiratory and hematological adaptations of young and older Aymara men native to 3600M. American Journal of Human Biology 4, 17–26.

Table 1 List of traits found in high altitude populations (>3000m) compared to local lowland native groups

	High Altitude Region				
Trait	Himalayas/ Tibet	Andes	Ethiopia		
Height	↓ [9,11]	↓ [4]	↑ [74]		
Sitting Height	↑ [44]	↑ [75]	↑ [76 <u>]</u>		
Relative zeugopod length	↓ [29]	↓ [17]	↓ [76]		
Fat mass	↓ [77]	↓ [78]	↓ [74]		
Chest volume	↑ [44]	↑ [79]	↑ [74]		
Exhaled nitric oxide	↑ [80]	↑ [81]	↑ [82]		
Erythrocytosis	↔ [6]	↑ [6]	↔ [83]		
Arterial oxygen concentration	↓ [84]	↑ [84]	↔ [4]		
Altitude sickness with age	↑ [83]	↑ [85]	↔ [83]		

 $[\]uparrow$ - Increase; \downarrow - Decrease; \leftrightarrow - No difference

Table 2 Descriptive statistics of highland and lowland populations

	Female			Male		
	Lowland (n=42) Mean (SD) (cm)	Highland (n=48) Mean (SD) (cm)	Sig.	Lowland (n=29) Mean (SD) (cm)	Highland (n=135) Mean (SD) (cm)	Sig.
Height	154.1 (±5.7)	155.5 (±6.3)	p>0.05	168.2 (±7.0)	165.1 (±7.0)	p<0.01
Total Upper Limb Length	71.3 (±3.7)	70.0 (±3.3)	p>0.05	77.6 (±3.9)	74.6 (±3.8)	p<0.01
Humerus Length	29.1 (±1.8)	$29.0 (\pm 2.0)$	p>0.05	31.4 (±1.8)	30.3 (±2.1)	p<0.01
Ulna Length	24.3 (±1.5)	23.2 (±1.4)	p<0.01	26.8 (±1.7)	25.4 (±1.8)	p<0.01
Hand Length	17.8 (±0.9)	17.7 (±0.9)	p>0.05	19.4 (±1.3)	18.9 (±0.1)	p<0.05
Hand Width	9.2 (±0.5)	9.1 (±0.6)	p>0.05	10.2 (±0.7)	9.9 (±0.6)	p>0.05

Sig. = significance. **Bold** indicates statistically significant differences (p < 0.05)

Figure and Table Captions

Table 1 List of traits found in high altitude populations (>3000m) compared to local lowland native groups

Table 2 Descriptive statistics of highland and lowland populations

Figure 1 Bar chart of mean difference in upper limb segment length relative to height between lowland and highland populations (Mean relative difference calculated as lowland relative mean – highland relative mean) *** = p < 0.01

