

Original citation:

Zheng, Bo, Wang, Hongwei, Shang, Wenting, Xie, Fengwei, Li, Xiaoxi, Chen, Ling and Zhou, Zhongkai (2018)*Understanding the digestibility and nutritional functions of rice starch subjected to heat-moisture treatment*. Journal of Functional Foods, 45 . pp. 165-172. doi:10.1016/j.jff.2018.03.041

Permanent WRAP URL:

http://wrap.warwick.ac.uk/103033

Copyright and reuse:

The Warwick Research Archive Portal (WRAP) makes this work by researchers of the University of Warwick available open access under the following conditions. Copyright © and all moral rights to the version of the paper presented here belong to the individual author(s) and/or other copyright owners. To the extent reasonable and practicable the material made available in WRAP has been checked for eligibility before being made available.

Copies of full items can be used for personal research or study, educational, or not-for-profit purposes without prior permission or charge. Provided that the authors, title and full bibliographic details are credited, a hyperlink and/or URL is given for the original metadata page and the content is not changed in any way.

Publisher's statement:

© 2018, Elsevier. Licensed under the Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International http://creativecommons.org/licenses/by-nc-nd/4.0/

A note on versions:

The version presented here may differ from the published version or, version of record, if you wish to cite this item you are advised to consult the publisher's version. Please see the 'permanent WRAP URL' above for details on accessing the published version and note that access may require a subscription.

For more information, please contact the WRAP Team at: wrap@warwick.ac.uk

1	Understanding the digestibility and nutritional functions of rice starch
2	subjected to heat-moisture treatment
3	Bo Zheng ^a , Hongwei Wang ^a , Wenting Shang ^b , Fengwei Xie ^c , Xiaoxi Li ^a , Ling Chen ^{a*} ,
4	Zhongkai Zhou ^{b*}
5	
6	^a Ministry of Education Engineering Research Center of Starch & Protein Processing, Guangdong
7	Province Key Laboratory for Green Processing of Natural Products and Product Safety, School of
8	Food Science and Engineering, South China University of Technology, Guangzhou 510640, China
9	^b Key Laboratory of Food Nutrition and Safety, Ministry of Education, Tianjin University of Science
10	and Technology, Tianjin, 300457, China
11	^c Institute of Advanced Study, University of Warwick, Coventry CV4 7HS, United Kingdom
12	
13	*Corresponding author.
14	Email addresses: felchen@scut.edu.cn, zkzhou@tust.edu.cn; Tel: +86 20 8711 3252

17 Abstract: In this study, rice starch with well-controlled digestion resistibility achieved by heatmoisture treatment (HMT) was chosen as a supplementary diet for high-fat-diet-fed mice. Then, the 18 19 nutritional functions of HMT-modified rice starch were evaluated by the physiological and 20 biochemical indices, proliferation and distribution of intestinal microflora, and functional diversity 21 by putative metagenomes analysis. Compared with the native-rice-starch mice (DM) group, the 22 blood glucose, serum lipid, oxidative stress, and liver function metabolic levels/indices of the HMT-23 rice-starch mice (HMT-DM) group were worse due to the declined level of slowly digestible starch (SDS) in HMT-modified rice starch. Meanwhile, the species diversity index was observed to be 24 25 higher in the DM group and Bifidobacteria was identified as a type of bacteria related to the relatively higher content of RS in HMT-modified rice starch. Overall, our results provide important 26 27 information for the rational design of rice starch-based health-promoting foods with nutritional 28 functions.

29

30 Keywords: Rice starch; heat-moisture treatment; digestion; nutritional functions; intestinal
 31 microflora

33 Nomenclature

34	SDS	slowly digestible starch
35	НМТ	heat-moisture treatment
36	RDS	rapidly-digestible starch
37	SDS	slowly-digestible starch
38	RS	resistant starch
39	TG	triglyceride
40	ТСН	total cholesterol
41	HDL-c	high-density lipoprotein cholesterol
42	LDL-c	low-density lipoprotein cholesterol
43	MDA	Malondialdehyde
44	SOD	Superoxide dismutase
45	GSH-PX	Glutathione peroxidase
46	T-AOC	Total antioxidant capacity
47	ALT	alanine aminotransferase
48	ALP	alkaline phosphatase
49	AST	aspartate aminotransferase
50	OTUs	Operational Taxonomic Units
51	SEM	standard error of the mean
50		

53 **1. Introduction**

With the improvement of living standard and the increased diversity of diet and disease spectra, 54 55 foods with nutritional functions such as physiological accommodation and disease prevention have 56 attracted increasing attention (Barratt, Lebrilla, Shapiro, & Gordon, 2017; Caballero, 2013; Link & 57 Reue, 2017). Therefore, designing personalized healthy foods with particular nutritional functions has been one of the hotspots in food science. Rice, one of the most widely consumed staple food, is 58 59 mainly composed of starch. Modulating the digestibility and nutritional functions of rice starch have 60 a direct impact on human health. For positive physiological effects and nutritional benefits, starch, depending on the rate and extent 61 of digestion, can be classified into three categories, namely rapidly-digestible starch (RDS), slowly 62 digestible starch (SDS), and resistant starch (RS) (Dhital, Warren, Butterworth, Ellis, & Gidley, 63 2017; Englyst, Kingman, & Cummings, 1992). Previous research has shown that RDS, after rapidly 64 digested in the digestive tract, releases glucose, which increases the level of serum glucose, promotes 65 66 insulin secretion, stimulates the liver and lipocytes to create fat, and increases the serum lipid level in 67 our body (Hung, Chau, & Phi, 2016; Lee et al., 2014). Meanwhile, SDS and RS can retard or restrain 68 the release of glucose, lower the glycemic index, improve the insulin sensitivity, promote the secretion or expression of insulin and adipocyte-secreted factors, and prevent the fat from deposition 69 70 (Dhital, Bhattarai, Gorham, & Gidley, 2016; Wei, Sissons, Warren, Gidley, & Gilbert, 2016). Interestingly, RS will produce a series of short-chain fatty acid metabolites at a lower pH. Many 71 72 health benefits and functional properties of RS have been reported including the prebiotic effect on 73 colon microbes (Shang et al., 2017), the adjustment of lipid metabolism (Nathalie et al., 2016), the

74	improvement in cholesterol metabolism (Newman et al., 2017), and the reduction in the risk of
75	ulcerative colitis (Bindels et al., 2017) and colon cancer (Si, Strappe, Blanchard, & Zhou, 2016; Si,
76	Zhou, Strappe, & Blanchard, 2016). However, gelatinized rice starch consists of over 95% RDS,
77	which can be easily digested and absorbed, leading to a high glycemic response. This negative
78	physiological effect of rich starch does not correspond to the modern concept of nutrition and health.
79	Therefore, recent research has focused on changing the multi-scale structure of rice starch under
80	different ways of processing or physical modification to regulate its digestion, absorption, and
81	metabolism in the human gastrointestinal tract (GIT) and to improve its nutritional functions.
82	Different ways of changing the starch multi-scale structure and regulating the starch digestibility
83	have been widely reported (Klein et al., 2013; Puncha-Arnon & Uttapap, 2013; Silva et al., 2017;
84	Tan et al., 2017). As a new physical modification method, heat-moisture treatment (HMT) only
85	involves the use of thermal energy and moisture, which has advantages such as environmental
86	protection, high efficiency, and safety (Wang, Zhang, Chen, & Li, 2016; Zavareze & Dias, 2011).
87	HMT has already played an important role in the green processing of starch (Arns et al., 2015; Hung,
88	Vien, & Phi, 2016; Pratiwi, Faridah, & Lioe, 2017; Silva et al., 2017). However, limited work has
89	been undertaken for evaluating the effect of HMT on the nutritional functions of starch in terms of
90	blood lipid metabolism, blood sugar metabolism, and intestinal microflora. Thus, it is necessary to
91	establish a systemic connection between digestibility and nutritional functions, which would
92	contribute to the development of healthy starch food by HMT.
93	Therefore, in this study, we investigated the physiological and biochemical indexes and the

94 proliferation and distribution of intestinal microbiota for 10 high-fat-diet-fed mice supplemented

with rice starch (the DM group) and another 10 with HMT-modified rice starch (the HMT-DM
group) to understand the alteration of nutritional functions by HMT. To the best of our knowledge, it
is the first study comprehensively comparing these detailed nutritional function parameters between
native rice starch and HMT-modified rice starch.

99

100 **2. Material and methods**

101 2.1 sample preparation

102 Native rice starch was purchased from the South China Agricultural University (Guangzhou,

103 China). The RDS, SDS and RS contents of this starch were 32.6%, 52.4%, 9.0%, respectively, which

104 was analyzed with a modified Englyst procedure according to a previous study in our lab (Wang et

al., 2018). The HMT of rice starch was conducted under a moisture content of 25%. The sample was

106 equilibrated at 4 °C for 24 h. Then, they were placed in a 500 mL screwed stainless steel reactor with

107 continuous rotation and heated with oil at 110 °C for 4 h, followed by cooling to room temperature.

- 108 During HMT, the starch granules were stirred by the paddle when the reactor was continuously
- 109 rotated. Subsequently, the treated samples were dried at 40 °C and then ground. The RDS, SDS and
- 110 RS contents of HMT-modified rice starch were measured to be 60.4%, 20.7%, 18.9%, respectively.

111 2.2 Animals

112 Twenty healthy male C57BL-6 mice (non-obese) of 20±5 g weight were purchased from the

animal house, the Academy of Military Medical Sciences (China). These mice were fed in a clean-

114 grade facility in the Laboratory Animal Center at Tianjin University of Science and Technology.

115	After one week's adaptive feeding with a basal diet, no difference in the body weight was observed.
116	These mice were divided into two groups randomly. One group of mice were fed a high-fat diet
117	supplemented with rice starch (the DM group) and the other group supplemented with HMT-
118	modified rice starch (the HMT-DM group). For both groups, the intervention was implemented for 8
119	weeks. Their body weights and food intakes were monitored weekly. Feces were collected on the day
120	of necropsy.
121	All animal procedures were approved by the Ethical Committee for the Experimental Use of
122	Animals in the Center for Drug Safety Evaluation, Tianjin University of Science & Technology
123	(Approval No: 13/051/MIS).
124	2.3 Experimental diets
125	The experimental high-fat diet was prepared by Research Diets (Choi, Gwon, Ahn, Jung, & Ha,

126 2013), which consisted of 54.5% carbohydrate source, 10% lard oil, 20% casein, 5% corn oil, 0.5%

127 cholesterol, 5% cellulose, 3.5% mineral mix (based on AIN76), 1% vitamins (based on AIN76),

128 0.3% methionine, and 0.2% choline bitartrate. The two groups of mice were fed with the

129 experimental high-fat diet where carbohydrate source was replaced by rice starch and HMT-modified

130 rice starch, respectively.

131 2.4 Blood and tissue analysis

132 At the end of the intervention period, the mice were fasted overnight, before the blood glucose was

133 recorded. Blood samples were taken from the arteria femoralis for the determination of physiological

134 and biochemical indexes. The liver tissue, epididymal white adipose tissue, and perirenal white

135	adipose tissue were quickly removed after sacrifice and then weighed. The liver tissue was
136	immediately stored at -80 °C for further pathological slides.

137 2.4.1 Analysis of blood lipids composition

138 Blood lipid indexes evaluated in this study included triglyceride (TG), total cholesterol (TCH),

139 high-density lipoprotein cholesterol (HDL-c), and low-density lipoprotein cholesterol (LDL-c). The

140 specific measurement procedures and final calculations were performed according to the

141 specification of the TG, TCH, HDL-c, and LDL-c assay kits (Jian Cheng Biotechnology Co., Ltd.,

142 Nanjing, China).

151

143 **2.4.2** Analysis of oxidative stress index

China) according to the manufacturer's instructions.

There are four main indexes for characterizing oxidative stress. Malondialdehyde (MDA) is a final product of the lipid oxidation process and was measured according to an MDA assay kit (Jian Cheng Biotechnology Co., Ltd., Nanjing, China). Superoxide dismutase (SOD) was determined using an SOD assay kit (Jian Cheng Biotechnology Co., Ltd., Nanjing, China). Glutathione peroxidase (GSH-PX) was measured using an assay kit (Jian Cheng Biotechnology Co., Ltd., Nanjing, China) and the activity was calculated according to the manufacturer's instructions. The Total antioxidant capacity (T-AOC) was determined using a T-AOC assay kit (Jian Cheng Biotechnology Co., Ltd., Nanjing,

152	2.4.3	Determination	of hepatic	lipid	metabolism	enzymes
-----	-------	---------------	------------	-------	------------	---------

153	The detection of the liver functional metabolism mainly includes three indices reflecting the
154	aspartate transaminase (Nathalie et al.), alanine aminotransferase (ALT), and alkaline phosphatase
155	(ALP) activities. The aspartate aminotransferase (AST) activity was measured by colorimetric
156	analysis using an AST assay kit (Jian Cheng Biotechnology Co., Ltd, Nanjing, China). The ALT
157	activity was measured by colorimetric analysis using an ALT assay kit (Jian Cheng Biotechnology
158	Co., Ltd, Nanjing, China). The ALP activity was measured by standard oxidation using potassium
159	ferricyanide.
160	2.4.4 Liver histology and analysis
161	The liver tissues were placed in 10% neutral formalin liquid for 12 h, which were then dehydrated
162	using 30%, 50%, 70%, 80%, 90%, 95% and 100% ethanol solutions, respectively, and washed in
163	xylol. Then, the liver tissues were embedded in paraffin (BMJ-III embedding machine, Jiangsu,
164	China). Finally, they were stained with hematoxylin and eosin (H & E) and observed by light
165	microscopy at 200× magnification.
166	2.5 Microbial community analysis

167 2.5.1 Feces samples collection, DNA extraction, and 168 rRNA gene sequencing

Feces samples were collected in a sterile container and immediately stored at -80 °C until further
processing. Total genome DNA from samples was extracted using the hexadecyltrimethylammonium
bromide/sodium dodecyl sulfate (CTAB/SDS) method (Caporaso et al., 2010). The DNA

171	concentration and purity were monitored on 1% agarose gels. The 16S rRNA genes of distinct
172	regions in V4 were amplified using the specific primer of 515F-806R. All polymerase chain reaction
173	(PCRs) were carried out with a Phusion® High-Fidelity PCR Master Mix (New England Biolabs).
174	The same volumes of 1× loading buffer (containing SYBR Green I) were mixed with PCR products
175	and electrophoresis was operated on 2% agarose gels. Sequencing libraries were generated using a
176	TruSeq® DNA PCR-Free Sample Preparation Kit (Illumina, USA) following manufacturer's
177	recommendations, and index codes were added. Finally, the library was sequenced on an
178	IlluminaHiSeq2500 platform and 250 bp paired-end reads were generated.
179	2.5.2 Intestinal microbiota analysis
180	After the removal of the low-quality sequences, pyrosequencing errors, and chimera, all the
181	sequencing reads were denoised. Sequences of >97% similarity levels were assigned to the same
182	Operational Taxonomic Units (OTUs) (Edgar, 2013). Representative sequences for each OTU were
183	screened for further annotation. The rare fraction curves and alpha diversity indices (Chao1, ACE,
184	Shannon, and Simpson) were performed using MOTHUR software, and the beta diversity (among
185	samples) was analyzed using principal component analysis (PCA). The function of intestinal
186	microbiota was performed by online phylogenetic investigation of communities by reconstruction of
187	an unobserved states program (PICRUSt, http://picrust.github.io/picrust/).
188	2.6 Statistical analysis

189 All analyses were conducted in triplicate and statistical analyses were performed using the
190 Statistical Package for Social Science (SPSS) software version 22. Body weight and food intake

191	were analyzed using General Linear Model repeated measures (Mixed Design ANOVA). P
192	interaction indicates the effect of time and group interactions. Once a significant difference ($P \le$
193	0.05) was detected, post hoc multi-comparisons were performed by LSD adjustment. Other
194	physiological and biochemical indexes (blood glucose and insulin) were analyzed by one-way
195	analysis of variance followed by Tukey's multiple comparison analysis. Different lowercase letters
196	above the same column indicate a significant difference ($P \le 0.05$). Results were expressed as mean
197	\pm standard error of the mean (SEM).

3. Results and discussions

200 3.1 Body weight, and liver and adipose tissue weights

201 Fig. 1 shows the changes in body weight of the two groups of mice after the 8-week intervention. 202 The initial body weights between the two groups were not significantly different (P > 0.05), showing the rationality of the grouping method. From the start to Week 4, there were apparent decreasing 203 204 trends of the body weight and food intake for both of the groups. Afterwards, both the body weight 205 and food intake gradually increased. At the end of Week 8, the body weights of mice in these two groups were 30% higher than their initial values, and General Linear Model (GLM) analysis showed 206 207 no significant difference (week \times group interaction) between the two groups (P > 0.05). 208 After the 8-week intervention, the perirenal fat, epididymal fat, and total fat-to-body ratio of the

209 HMT-DM group mice were significantly higher than those of the DM group mice (Table 1). These

210 results indicate that native rice starch had better performance in controlling the fat tissue deposit in 211 contrast to HMT-modified rice starch.

212 3.2 Blood glucose, insulin and lipid levels

Table 2 shows the blood glucose, insulin, and blood lipid levels of the two groups of mice. 213 214 Previous research has shown that the blood glucose level of C57BL-6 mice was normally 6–10 mmol/L (Rodríguez, Limón-Pacheco, Del Razo, & Giordano, 2016), which was slightly lower than 215 216 those of two experimental groups after the 8-week intervention. This observation indicated that both 217 native rice starch and HMT-modified rice starch could stabilize the blood glucose level and reduce 218 the speed of change into a hyperglycemia condition. In addition, the blood glucose level of the HMT-219 DM group was significantly higher than that of the DM group (P < 0.05), which might be related to 220 the increased RDS content and greatly decreased SDS content after HMT. The result confirmed that the SDS content has a strong impact on the regulation of the blood glucose level as also shown in 221 222 recent studies (Kittisuban, Lee, Suphantharika, & Hamaker, 2014; Wolever et al., 2016). 223 After the intervention, the serum insulin levels of these two groups were all within the normal

range (Seghers, Nakazaki, Aguilar, & Bryan, 2000). Nonetheless, compared with the DM group, the

HMT-DM group displayed a significantly higher serum insulin level (P < 0.05), indicating a higher

226 insulin resistance of the HMT-DM group than that of the DM group. Thus, the HMT-modified rice

starch intervention reduced the biological effect of insulin as well as the sensitivity of the insulin

228 receptor tissue to insulin.

According to a previous study (Roza, Possignolo, Palanch, & Gontijo, 2016), the degree of
dyslipidemia and the levels of TG, TC and LDL-c were all positively proportional to obesity, while

231 the HDL-c level was negatively related to obesity. It has been proved that the abnormal deposition of 232 TG in the liver and skeletal muscles can impair the activity of oxidase and affect the metabolism of 233 glucose and lipids (Zhang, 2016). In addition, it is widely believed that the abnormal changes in 234 individual blood lipid levels, such as increased TC, TG and LDL-c levels and a reduced HDL-c level, 235 could be the causes of coronary heart disease (Sugawara et al., 2000). In other words, the blood 236 lipids have a strong relationship with health. Table 2 summarizes the blood lipid levels of the two 237 groups at the end of the intervention period. It can be seen that the TC and TG levels of blood lipids 238 in the HMT-DM group were significantly higher than those of the DM group (P < 0.05), while the 239 HDL-c level was significantly lower (P < 0.05). These results suggest that after the high-fat-diet intervention for 8 weeks, the blood lipid indices of mice in the DM group were better than those of 240 HMT-DM group. In other words, the ability of rice starch in regulating the blood lipid levels was 241 242 actually reduced by HMT.

243 3.3 Oxidative stress indices

Oxidative stress causes cytotoxicity and the accumulation of reactive oxygen species in the 244 245 body (Reeves, Nielsen, & Fahey, 1993). In this study, the changes in MDA, SOD, GSH-PX and T-246 AOC levels in the serum of mice were examined to understand the influence of native and HMT-247 modified rice starches on these oxidative stress indices. After a high-fat diet intake, a large amount of 248 calorie led to exuberant energy metabolism and hyperglycemia status in mice, which then intensified 249 glucose oxidation to produce excessive oxidative products (Cristani et al., 2016). Meanwhile, the 250 stronger oxidative ability of excessive oxidative products than the antioxidant capacity could lead to 251 oxidative stress in the body. The reason for the reduced amounts of antioxidant enzymes could be the

252 rapid consumption and exhaustion of their storage in the body when fighting free radicals generated during development of obesity (Sathiavelu, Senapathy, Devaraj, & Namasivayam, 2009). The results 253 254 (Table 3) show a statistically significant increase in serum levels of MDA and GSH-PX, as well as a 255 decrease in T-AOC level for the HMT-DM group compared with for the DM group (P < 0.05). 256 Overall, the serum oxidative stress status of mice in the DM group was more effectively improved 257 than that of mice in the HMT-DM group. Thus, it could be suggested that native rice starch, 258 compared with HMT-modified rice starch, could better suppress oxidant stress induced by the high-259 fat diet.

260 3.4 hepatic lipid metabolism enzymes

261 The detection of hepatic metabolism enzymes mainly includes ALT, AST, and ALP. In the 262 present study, the activities of AST, ALT, and ALP are increased in the serum, which can be 263 considered an indication of liver damage. Previous studies have shown that cell membrane damage 264 leads to a more prominent increase in ALT and AST activities (Borlak, Chougule, & Singh, 2014). 265 The data in Table 3 also show that the ALT and AST levels of mice in the HMT-DM group were significantly higher than those in the DM group (P < 0.05). Thus, the increased levels of these 266 267 enzymes indicated that HMT-modified rice starch may have a significant impact on cellular 268 homeostasis. Overall, these results suggest that compared to the intervention with the HMT-modified 269 rice starch, native rice starch could alleviate liver dysfunction to a certain degree, and could prevent or delay the abnormal liver function caused by fat accumulation. 270

Fig. 2 shows that the liver tissue of mice in the DM group had a large amount of fatty degeneration, with different sizes of lipid droplet vacuoles (marked by black circles) in the cytoplasm

and some degree of perivascular infiltration by inflammatory cells. While the liver tissue of the
HMT-DM group mice had visible steatosis as well, it contained fewer lipid vacuoles, higher vascular
permeability, and more perivascular infiltration than that of DM group did. In other words, the
resistance to abnormal liver function caused by the high-fat diet was reduced by the intervention with
HMT-modified rice starch.

278 3.5 Intestinal microflora

279 The gut is a complex, active, relatively balanced system, of which the type and flora amounts can 280 be changed when the body acquires different kinds of flora from the environment and food (Clemente, Ursell, Parfrey, & Knight, 2012). Recent research has revealed the function of intestinal 281 282 microflora for the normal homeostasis of the human body (Hartstra, Nieuwdorp, & Herrema, 2016). 283 Fig. 3A shows the gut microbiota composition in mice after the 8-week intervention for the two 284 groups. The observed species indices in the DM group was significantly higher than those in the 285 HMT-DM group (P < 0.05), which indicates a higher intestinal flora abundance of the DM group. 286 This result confirmed that when RS reached the colon and degraded by microbes, the degradation 287 products would change the distribution structure of intestinal flora and reduce its species abundance. 288 In addition, the PCA analysis diagram based on OTU abundance showed two relatively separated 289 macroscopic distribution profiles, accounting for the apparent difference in the microbial structure 290 between the DM group and the HMT-DM group. To further analyze the difference in microbial 291 structure between the two groups, a weighted UniFrac distance matrix was constructed (shown in 292 Fig. 3B). Interestingly, the difference within the HMT-DM group was larger than that within the DM 293 group based on the level of a single sample of OTUs.

294	Then, we analyzed the statistical results of species percentage among different species
295	classification levels. At the phylum level (shown in Fig. 3C), bacteria between the two groups were
296	mainly composed of Bacteroidete, Firmicutes, and Proteobacteria. These three bacteria took up
297	about 45.8%, 31.6% and 17.7% in the DM group mice whereas those in the HMT-DM group mice
298	were 45.2%, 24.3.0%, and 24.4%, respectively. Thus, there were significant differences in the
299	percentage of <i>Firmicutes</i> and <i>Proteobacteria</i> between the two groups ($P < 0.05$).
300	At the family level, the microbial structure and relative abundance in the two groups of mice were
301	also shown in Fig. 3C. The proportions of Ruminococcaceae, Lachnospiraceae,
302	Porphyromonadaceae, and other bacteria in the DM group were slightly higher than that in the
303	HMT-DM group. The ratios were 7.1% vs. 4.2%, 7.6% vs. 4.1%, and 3.6% vs. 1.9% (DM vs. HMT-
304	DM), respectively ($P < 0.05$). At the same time, the proportions of some species of the intestinal
305	microflora in the HMT-DM group, namely Helicobacteraceae, S24-7, Erysipelotrichacea,
306	Desulfovibrionaceae, and Bifidobacteriaceae, were significantly higher than that in the DM group,
307	and the ratios were 17.5% vs. 6.6%, 23.0 vs. 18.6%, 10.6% vs. 6.4%, 4.5% vs. 1.3% (HMT-DM vs.
308	DM), respectively ($P < 0.05$). Studies have shown that the digestive tract adenoma or cancer of the
309	organism are correlated positively with Ruminococcaceae, Porphyromonadaceae, and
310	Lachnospiraceae (Frank et al., 2007; Meehan & Beiko, 2014), and negatively with Bacteroides,
311	Bifidobacteriaceae and Desulfovibrionaceae (Panasevich et al.). Therefore, it could be speculated
312	that when RS entered the colon and degraded by microbes, the degradation products could
313	effectively inhibit the growth of some species of intestinal bacteria such as Ruminococcaceae,
314	Porphyromonadaceae and Lachnospiraceae, but meanwhile stimulate the growth of Bacteroides,

Bifidobacteriaceae and Desulfovibrionaceae, which could suppress the occurrence of digestive tract
 adenoma and cancer.

317 Metagenomics analysis using the linear discriminant analysis effect size (LEfSe) method was 318 performed to compare the microbial community composition and its abundance diversity in feces 319 between the two groups of mice from the phylum, class, order, family and genus levels, and to further select the dominant flora of microbial communities in the two groups. It can be seen in Fig. 4 320 321 that there were large differences in each level of microbial communities between the two groups. 322 According to the LDA score, the dominant bacteria in the HMT-DM group were Bifidobacterium, 323 while S24-7, β -proteobacteria, and Proteobacteria occupied important places in the DM group. Some previous opinions considered that bifidobacteria were one of the probiotics associated with the 324 325 glucose and lipid metabolism (Martorell et al., 2016; Patterson et al., 2017). When the disorder of 326 glucose and lipid metabolism occurred, there was a decrease in the bifidobacteria content in the gut. 327 In contrast, a higher amount of bifidobacteria contributed to normalizing the blood glucose and 328 blood lipid levels. However, here, the content of bifidobacteria in the HMT-DM group mice was 329 higher than that in the DM group, though the indices on blood glucose and blood lipids in the HMT-330 DM group mice were still worse than those in the DM group (see Table 1 and Table 2). These differences were mainly due to the high sugar environment produced by the high-fat diet, which led 331 332 to lower activity of functional enzymes in the gut. Therefore, SDS showed more powerful effects on 333 regulating blood glucose and lipids.

334 Besides, Parabacteroides and S24-7 were pathogenic bacteria, which were closely related to the incidence of cancer in the digestive tract. Our results showed that HMT-modified rice starch could 335 336 effectively inhibit the pathogenic bacteria, and reduce the occurrence of cancer in the digestive tract. 337 The results of animal experiments in our study indicate that physiological and biochemical indices 338 were mainly depended on the content of enzyme-resistant components (SDS + RS) in rice starch, 339 whilst the intestinal microflora was determined by the RS content. 340 We also assessed the functional diversity of the different putative metagenomes using PICRUSt software (Langille et al., 2013), which allows the prediction of metabolic pathways from the 16S 341 342 rRNA reads. Most of the genes were detected to annotate in the pathways of environmental information processing, genetic information processing, and metabolism. Moreover, the pathways 343 344 displayed a difference in the mean proportion between the DM and HMT-DM groups (Fig. 5). Some 345 pathways including carbohydrate metabolism, lipid metabolism, and membrane transport were over-346 represented in the DM group, whereas the metabolisms of cofactors, vitamins and amino acids and 347 the cell motility were over-represented in the HMT-DM group. These results indicate that rice starch 348 associated with HMT may also influence the functional diversity, especially predicted by putative 349 metagenomes.

350 **4. Conclusions**

This research is focused on the nutritional function of rice starch after HMT, and the relationship between digestibility and nutritional function was established. The results indicate that the physiological and biochemical indices of the HMT-DM group were slightly worse than those of the DM group due to the relatively lower content of SDS in HMT-modified rice starch. *Bifidobacteria*

355	were identified as a type of bacteria related to HMT-modified rice starch, which might be ascribed to
356	the relatively higher content of RS in HMT-modified rice starch, although the specific functions of
357	this type of bacteria require further studies. Moreover, some pathways influenced by these two
358	starches were annotated to reveal the functional metabolisms in the body. Thus, this work is of great
359	significance in modulating the nutritional functions of rice starch and is instrumental to the
360	development of starch-based healthy food.
361	
362	Potential conflict of interest statement
363	The authors declare no competing financial interest.
364	
365	Acknowledgements
366	This research has been financially supported by the National Natural Science Foundation of China
367	(NSFC)-Guangdong Joint Foundation Key Project (U1501214), YangFan Innovative and
368	Entrepreneurial Research Team Project (2014YT02S029), the Science and Technology Program of
369	Guangzhou (201607010109). F. Xie acknowledges the European Union's Marie Skłodowska-Curie
370	Actions (MSCA) and the Institute of Advanced Study (IAS), University of Warwick for the Warwick
371	Interdisciplinary Research Leadership Programme (WIRL-COFUND).
372	
373	References

Arns, B., Bartz, J., Radunz, M., Evangelho, J. A. D., Pinto, V. Z., Zavareze, E. D. R., & Dias, A. R.

375	G. (2015). Impact of heat-moisture treatment on rice starch, applied directly in grain paddy
376	rice or in isolated starch. LWT - Food Science and Technology, 60(2), 708-713.
377	Barratt, M. J., Lebrilla, C., Shapiro, H. Y., & Gordon, J. I. (2017). The Gut Microbiota, Food
378	Science, and Human Nutrition: A Timely Marriage. Cell Host & Microbe, 22(2), 134.
379	Bindels, L. B., Munoz, R. R. S., Gomesneto, J. C., Mutemberezi, V., Martínez, I., Salazar, N.,
380	Reyesgavilán, C. G. D. L. (2017). Resistant starch can improve insulin sensitivity
381	independently of the gut microbiota. Microbiome, 5(1), 12.
382	Borlak, J., Chougule, A., & Singh, P. K. (2014). How useful are clinical liver function tests in in vitro
383	human hepatotoxicity assays? Toxicology in Vitro, 28(5), 784-795.
384	Caballero, B. (2013). Sucrose: Dietary Sucrose and Disease - Encyclopedia of Human Nutrition
385	(Third Edition). Encyclopedia of Human Nutrition, 231–233.
386	Caporaso, J. G., Kuczynski, J., Stombaugh, J., Bittinger, K., Bushman, F. D., Costello, E. K.,
387	Gordon, J. I. (2010). QIIME allows analysis of high-throughput community sequencing data.
388	Nature Methods, 7(5), 335.
389	Choi, W. H., Gwon, S. Y., Ahn, J., Jung, C. H., & Ha, T. Y. (2013). Cooked rice prevents
390	hyperlipidemia in hamsters fed a high-fat/cholesterol diet by the regulation of the expression
391	of hepatic genes involved in lipid metabolism. Nutrition Research, 33(7), 572-579.
392	Clemente, J. C., Ursell, L. K., Parfrey, L. W., & Knight, R. (2012). The impact of the gut microbiota
393	on human health: an integrative view. Cell, 148(6), 1258.
394	Cristani, M., Speciale, A., Saija, A., Gangemi, S., Minciullo, P. L., & Cimino, F. (2016). Circulating
395	Advanced Oxidation Protein Products as Oxidative Stress Biomarkers and Progression

396	Mediators in Pathological Conditions Related to Inflammation and Immune Dysregulation.
397	Current Medicinal Chemistry, 23(34), 3862-3882.
398	Dhital, S., Bhattarai, R. R., Gorham, J., & Gidley, M. J. (2016). Intactness of cell wall structure
399	controls the in vitro digestion of starch in legumes. Food & Function, 7(3), 1367.
400	Dhital, S., Warren, F. J., Butterworth, P. J., Ellis, P. R., & Gidley, M. J. (2017). Mechanisms of Starch
401	Digestion by α -amylase-structural Basis for Kinetic Properties. Critical Reviews in Food
402	Science & Nutrition, 57(5), 875.
403	Edgar, R. C. (2013). UPARSE: highly accurate OTU sequences from microbial amplicon reads.
404	<i>Nature Methods, 10</i> (10), 996.
405	Englyst, H. N., Kingman, S. M., & Cummings, J. H. (1992). Classification and measurement of
406	nutritionally important starch fractions. European Journal of Clinical Nutrition, 46 Suppl
407	2(Suppl 2), S33.
408	Frank, D. N., Amand, A. L. S., Feldman, R. A., Boedeker, E. C., Harpaz, N., & Pace, N. R. (2007).
409	Molecular-phylogenetic characterization of microbial community imbalances in human
410	inflammatory bowel diseases. Proceedings of the National Academy of Sciences of the United
411	States of America, 104(34), 13780.
412	Hartstra, A. V., Nieuwdorp, M., & Herrema, H. (2016). Interplay between gut microbiota, its
413	metabolites and human metabolism: dissecting cause from consequence. Trends in Food
414	Science & Technology, 57.
415	Hung, P. V., Chau, H. T., & Phi, N. T. L. (2016). In vitro digestibility and in vivo glucose response of

416 native and physically modified rice starches varying amylose contents. *Food Chemistry*, 191,

417 74.

418	Hung, P. V., Vien, N. L., & Phi, N. T. L. (2016). Resistant starch improvement of rice starches under
419	a combination of acid and heat-moisture treatments. Food Chemistry, 191, 67.
420	Kittisuban, P., Lee, B. H., Suphantharika, M., & Hamaker, B. R. (2014). Slow glucose release
421	property of enzyme-synthesized highly branched maltodextrins differs among starch sources.
422	Carbohydrate Polymers, 107(1), 182-191.
423	Klein, B., Pinto, V. Z., Vanier, N. L., Zavareze, E. D. R., Colussi, R., Evangelho, J. A. D., Dias,
424	A. R. G. (2013). Effect of single and dual heat-moisture treatments on properties of rice,
425	cassava, and pinhao starches. Carbohydrate Polymers, 98(2), 1578-1584.
426	Langille, M. G., Zaneveld, J., Caporaso, J. G., Mcdonald, D., Knights, D., Reyes, J. A., Knight,
427	R. (2013). Predictive functional profiling of microbial communities using 16S rRNA marker
428	gene sequences. Nature Biotechnology, 31(9), 814.
429	Lee, B. H., Lin, A. H., Nichols, B. L., Jones, K., Rose, D. R., Quezadacalvillo, R., & Hamaker, B. R.
430	(2014). Mucosal C-terminal maltase-glucoamylase hydrolyzes large size starch digestion
431	products that may contribute to rapid postprandial glucose generation. Molecular Nutrition &
432	<i>Food Research</i> , <i>58</i> (5), 1111–1121.
433	Link, J. C., & Reue, K. (2017). The Genetic Basis for Sex Differences in Obesity and Lipid
434	Metabolism. Annual Review of Nutrition.
435	Martorell, P., Llopis, S., González, N., Chenoll, E., Lópezcarreras, N., Aleixandre, A., Genovés,
436	S. (2016). Probiotic Strain Bifidobacterium animalis subsp. lactis CECT 8145 Reduces Fat
437	Content and Modulates Lipid Metabolism and Antioxidant Response in Caenorhabditis

438 elegans. J Agric Food Chem, 64(17), 3462-3472. Meehan, C. J., & Beiko, R. G. (2014). A phylogenomic view of ecological specialization in the 439 440 Lachnospiraceae, a family of digestive tract-associated bacteria. Genome Biology & 441 *Evolution*, *6*(3), 703-713. Nathalie, B., Williams, P. T., Regina, L., Nastaran, F., Alyssa, G., Li, X., . . . Hazen, S. L. (2016). 442 443 Diets high in resistant starch increase plasma levels of trimethylamine-N-oxide, a gut 444 microbiome metabolite associated with CVD risk. British Journal of Nutrition, 116(12), 2020-2029. 445 446 Newman, M. A., Zebeli, Q., Eberspächer, E., Grüll, D., Molnar, T., & Metzler-Zebeli, B. U. (2017). Transglycosylated Starch Improves Insulin Response and Alters Lipid and Amino Acid 447 Metabolome in a Growing Pig Model. Nutrients, 9(3), 291. 448 449 Panasevich, M. R., Morris, E. M., Chintapalli, S. V., Wankhade, U., Shankar, K., Britton, S. L., ... 450 Rector, R. S. Reduced Short-chain Fatty Acid Producing Microbiota are Linked to Increased 451 Energy Intake and Susceptibility to High Fat Diet Induced Hepatic Steatosis in Low Aerobic 452 Capacity Rats. 453 Patterson, E., Wall, R., Lisai, S., Ross, R. P., Dinan, T. G., Cryan, J. F., ... Shanahan, F. (2017). 454 Bifidobacterium breve with α -linolenic acid alters the composition, distribution and 455 transcription factor activity associated with metabolism and absorption of fat. Sci Rep, 7, 456 43300. 457 Pratiwi, M., Faridah, D. N., & Lioe, H. N. (2017). Structural changes to starch after acid hydrolysis, debranching, autoclaving-cooling cycles, and heat moisture treatment (HMT): A review. 458

Starch - Stärke.

460	Puncha-Arnon, S., & Uttapap, D. (2013). Rice starch vs. rice flour: differences in their properties
461	when modified by heat-moisture treatment. Carbohydrate Polymers, 91(1), 85.
462	Reeves, P. G., Nielsen, F. H., & Fahey, G. C. (1993). AIN-93 purified diets for laboratory rodents:
463	final report of the American Institute of Nutrition ad hoc writing committee on the
464	reformulation of the AIN-76A rodent diet. Journal of Nutrition, 123(11), 1939.
465	Rodríguez, V. M., Limón-Pacheco, J. H., Del Razo, L. M., & Giordano, M. (2016). Effects of
466	inorganic arsenic exposure on glucose transporters and insulin receptor in the hippocampus of
467	C57BL/6 male mice. Neurotoxicology & Teratology, 54, 68.
468	Roza, N. A. V., Possignolo, L. F., Palanch, A. C., & Gontijo, J. A. R. (2016). Effect of long-term
469	high-fat diet intake on peripheral insulin sensibility, blood pressure, and renal function in
470	female rats. Food & Nutrition Research, 60.
471	Sathiavelu, J., Senapathy, G. J., Devaraj, R., & Namasivayam, N. (2009). Hepatoprotective effect of
472	chrysin on prooxidant-antioxidant status during ethanol-induced toxicity in female albino
473	rats. Journal of Pharmacy & Pharmacology, 61(6), 809.
474	Seghers, V., Nakazaki, M., F, Aguilar, B. L., & Bryan, J. (2000). Sur1 knockout mice. A model for
475	K(ATP) channel-independent regulation of insulin secretion. Journal of Biological Chemistry,
476	275(13), 9270.
477	Shang, W., Si, X., Zhou, Z., Wang, J., Strappe, P., & Blanchard, C. (2017). Studies on the unique
478	properties of resistant starch and chito-oligosaccharide complexes for reducing high-fat diet-
479	induced obesity and dyslipidemia in rats. Journal of Functional Foods, 38, 20-27.

480	Si, X., Strappe, P., Blanchard, C., & Zhou, Z. (2016). Enhanced anti-obesity effects of complex of
481	resistant starch and chitosan in high fat diet fed rats. Carbohydrate Polymers, 157, 834-841.
482	Si, X., Zhou, Z., Strappe, P., & Blanchard, C. (2016). A comparison of RS4-type resistant starch to
483	RS2-type resistant starch in suppressing oxidative stress in high-fat-diet-induced obese rats.
484	Food & Function, 8.
485	Silva, W. M., Biduski, B., Lima, K. O., Pinto, V. Z., Hoffmann, J. F., Vanier, N. L., & Dias, A. R.
486	(2017). Starch digestibility and molecular weight distribution of proteins in rice grains
487	subjected to heat-moisture treatment. Food Chemistry, 219(15), 260-267.
488	Sugawara, J., Tazuke, S. I., Suen, L. F., Powell, D. R., Kaper, F., Giaccia, A. J., & Giudice, L. C.
489	(2000). Regulation of insulin-like growth factor-binding protein 1 by hypoxia and 3',5'-cyclic
490	adenosine monophosphate is additive in HepG2 cells. Journal of Clinical Endocrinology &
491	Metabolism, 85(10), 3821-3827.
492	Tan, X., Li, X., Chen, L., Xie, F., Li, L., & Huang, J. (2017). Effect of heat-moisture treatment on
493	multi-scale structures and physicochemical properties of breadfruit starch. Carbohydrate
494	Polymers, 161, 286.
495	Wang, H., Liu, Y., Chen, L., Li, X., Wang, J., & Xie, F. (2018). Insights into the multi-scale structure
496	and digestibility of heat-moisture treated rice starch. Food Chemistry, 242, 323-329.
497	Wang, H., Zhang, B., Chen, L., & Li, X. (2016). Understanding the structure and digestibility of
498	heat-moisture treated starch. International Journal of Biological Macromolecules, 88, 1.
499	Wei, Z., Sissons, M., Warren, F. J., Gidley, M. J., & Gilbert, R. G. (2016). Compact structure and
500	proteins of pasta retard in vitro digestive evolution of branched starch molecular structure.

- 501 *Carbohydrate Polymers, 152,* 441.
- 502 Wolever, T. M., van Klinken, B. J., Bordenave, N., Kaczmarczyk, M., Jenkins, A. L., Chu, Y., &
- 503 Harkness, L. (2016). Reformulating cereal bars: high resistant starch reduces in vitro
- 504 digestibility but not in vivo glucose or insulin response; whey protein reduces glucose but
- 505 disproportionately increases insulin. *American Journal of Clinical Nutrition*, 104(4).
- Zavareze, E. D. R., & Dias, A. R. G. (2011). Impact of heat-moisture treatment and annealing in
 starches: A review. *Carbohydrate Polymers*, *83*(2), 317-328.
- 508 Zhang, X. Y. (2016). The Effects of Sitagliptin Combined with High Dose of Insulin on the Blood
- 509 Glucose and Blood Fat Levels in Patients with Type 2 Diabetes. *Diabetes New World*.

Figure captions

512	Figure 1. Changes in body weight and food intake of mice during the 8-week intervention for the
513	two groups. Statistical analysis was performed by General Linear Model repeated measures (Mixed
514	Design ANOVA). P interaction indicates the effect of time and group interactions.
515	Figure 2. Light microscope images of liver tissue of mice after the 8-week intervention for the two
516	groups (A: DM group; B: HMT-DM group)
517	Figure 3. Gut microbiota composition in mice after the 8-week intervention for the two groups. A:
518	Alpha diversity index; B: β diversity of intestinal microflora (left: PCA analysis; right: analysis of
519	weighted UniFrac distance matrix) (adjusted P value < 0.05); C: The relative abundance of bacterial
520	community at the taxa level (left: Phylum; right: Family) for the two groups (C1: DM group; C2
521	HMT-DM group)
522	Figure 4. Significant analysis of intestinal microflora in mice for the two groups (C1: DM group;
523	C2: HMT-DM group)
524	Figure 5. Function in KEGG module prediction using 16S data with PICRUSt. (A: KEGG genes
525	annotation; B: different notability functions between the two groups; C1: DM group; C2: HMT-DM

526 group).



Figure 1



Figure 2







Figure 3



Figure 4



Figure 5

 Group
 Organ coefficient
 Perirenal fat (g)
 Epididymal fat (g)
 Fat ratio (100%)

 DM
 0.054±0.0063^a
 0.094±0.0074^a
 0.418±0.0139^a
 2.084±0.10^a

 HMT-DM
 0.058±0.0065^a
 0.120±0.0102^b
 0.464±0.0105^b
 2.119±0.13^b

Table 1 Changes in perirenal fat, epididymal fat, fat-to-body ratio, and organ coefficient of miceafter the 8-week intervention for the two groups (mean \pm SEM).

Different lowercase letters above the same column indicate a significant difference ($P \le 0.05$).

Groups	Blood glucose (mmol/L)	Insulin (mU/L)	TC (nmol/L)	TG (nmol/L)	HDL-c (µmol/L)	LDL-c (µmol/L)
DM	11.12±0.28 ^b	0.115±0.002 ^b	1.922±0.059 ^b	12.629±0.789 ^b	104.676±5.310 ^a	88.551±2.384ª
HMT-DM	12.62±0.37 ^a	0.123±0.002ª	1.990±0.065ª	15.644±0.200ª	97.523±1.753 ^b	88.855±1.690ª

Table 2 Changes in blood glucose, insulin levels, and serum lipid of mice after the 8-weekintervention for two groups (mean \pm SEM).

Different lowercase letters above the same column indicate a significant difference ($P \le 0.05$).

Crosse	MDA	SOD	GSH-PX	T-AOC	ALT	AST	ALP
Groups	(nmol/mL)	(U/mL)	(U/mL)	(U/mL)	(U/L)	(U/L)	(IU/L)
DM	$7.26{\pm}0.42^{b}$	139.89±9.35ª	600.00 ± 46.82^{b}	10.38±0.68ª	12.47±1.98 ^b	124.07±11.78 ^b	0.11±0.01ª
HMT-DM	8.41±0.99ª	141.50±5.68ª	715.86±35.47ª	9.52±0.23 ^b	15.15±0.88ª	137.13±11.15 ^a	0.12±0.01ª

Table 3 Changes in oxidative stress and liver function metabolic levels of mice after the 8-weekintervention for two groups (mean \pm SEM).

Different lowercase letters above the same column indicate a significant difference ($P \le 0.05$).