

AMERICAN SOCIETY FOR MICROBIOLOGY

# brought to you by

## EUKARYOTES



# Transcriptome Sequence of the Bloodstream Form of *Trypanoplasma borreli*, a Hematozoic Parasite of Fish Transmitted by Leeches

# Mark Carrington,<sup>a</sup> Eva Dóró,<sup>b</sup> <sup>(D)</sup>Maria Forlenza,<sup>b</sup> Geert F. Wiegertjes,<sup>b</sup> <sup>(D)</sup>Steven Kelly<sup>c</sup>

Department of Biochemistry, University of Cambridge, Cambridge, United Kingdom<sup>a</sup>; Cell Biology and Immunology Group, Department of Animal Sciences, Wageningen University, Wageningen, The Netherlands<sup>b</sup>; Department of Plant Sciences, University of Oxford, Oxford, United Kingdom<sup>c</sup>

**ABSTRACT** Here, we report a transcriptome sequence of *Trypanoplasma borreli* isolated from its natural host, the common carp, *Cyprinus carpio*. The transcriptome allows an analysis of abundant cell surface proteins and acts as a comparator for understanding the evolution and pathogenicity of other *Kinetoplastida*, including several that infect humans.

The *Kinetoplastida* comprise a class of protozoa that separated early in the evolutionary diversification of eukaryotes (1). Within the *Kinetoplastida*, the *Trypanosomatidae* comprise a monophyletic group of parasites and pathogens that have evolved to colonize a diverse range of eukaryotic hosts (2). Most *Trypanosomatidae* spp. are transmitted between hosts by biting insects, and several species cause economic and health losses in developing countries (3). The *Bodonida* comprise a second group within the *Kinetoplastida* and are more diverse with free-living, commensal, and pathogenic species (4, 5). Here, we report a transcriptome assembly from a pathogenic *Bodonida* species, *Trypanoplasma borreli* (taxon ID 5710), isolated from experimentally infected carp. *T. borreli* infection can cause severe anemia and splenomegaly in both wild and farmed fish (6).

The isolate used here originated from a common carp, Cyprinus carpio, in a hatchery near Celle in Germany and was cloned in 1984 at the Hannover Medical School by the Fish Disease Research Unit at Tierärztliche Hochschule in Hannover, Germany (6, 7), and has been maintained in laboratories through serial infections of carp and stored as stabilates in liquid nitrogen. T. borreli was grown in carp to a parasitemia of ~1  $\times$  10<sup>8</sup>/ml and isolated from heparinized blood (8). Total RNA was prepared using the QIAgen RNeasy protocol. A library was made from polyA-selected RNA using random hexamer priming for reverse transcriptase. Sequencing was performed on an Illumina HiSeq using a single library of 90-nucleotide paired-end reads with a 170-bp inset size. Quality-filtered reads were assembled into contigs using the string graph assembler (SGA) (9). Contigs were then subject to scaffolding using SSPACE (10) and the full set of reads using the settings -k, 10; -a, 0.7; -n, 50; and -o, 20. Scaffolds were subject to gap filling using the SGA gap-filling function. Gene models were predicted using GeneMark-ST (11) and clustered into orthogroups using OrthoFinder (12). The assembled transcriptome contained 15,713 contigs encoding 13,640 putative proteins greater than 100 amino acids in length.

Accession number(s). This transcriptome shotgun assembly project has been deposited at GenBank under the accession number GFCF00000000. The version de-

Received 19 December 2016 Accepted 23 December 2016 Published 2 March 2017

**Citation** Carrington M, Dóró E, Forlenza M, Wiegertjes GF, Kelly S. 2017. Transcriptome sequence of the bloodstream form of *Trypanoplasma borreli*, a hematozoic parasite of fish transmitted by leeches. Genome Announc 5:e01712-16. https://doi.org/10.1128/ genomeA.01712-16.

**Copyright** © 2017 Carrington et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Steven Kelly, steven.kelly@plants.ox.ac.uk.

scribed in this paper is the first version, GFCF01000000. The raw reads are available from NCBI SRA under the accession number SRX2395293.

### ACKNOWLEDGMENTS

M.C. was funded by the Wellcome Trust (085956/Z/08/Z). E.D. was supported by the European Marie-Curie Initial Training Network FishForPharma (contract no. PITN-GA-2011-289209). S.K. is a Royal Society University Research Fellow.

### REFERENCES

- Adl SM, Simpson AG, Farmer MA, Andersen RA, Anderson OR, Barta JR, Bowser SS, Brugerolle G, Fensome RA, Fredericq S, James TY, Karpov S, Kugrens P, Krug J, Lane CE, Lewis LA, Lodge J, Lynn DH, Mann DG, McCourt RM, Mendoza L, Moestrup O, Mozley-Standridge SE, Nerad TA, Shearer CA, Smirnov AV, Spiegel FW, Taylor MF. 2005. The new higher level classification of eukaryotes with emphasis on the taxonomy of protists. J Eukaryot Microbiol 52:399–451. https://doi.org/10.1111/j.1550 -7408.2005.00053.x.
- Leonard G, Soanes DM, Stevens JR. 2011. Resolving the question of trypanosome monophyly: a comparative genomics approach using whole genome data sets with low taxon sampling. Infect Genet Evol 11:955–959. https://doi.org/10.1016/j.meegid.2011.03.005.
- Sutherland CS, Yukich J, Goeree R, Tediosi F. 2015. A literature review of economic evaluations for a neglected tropical disease: human African trypanosomiasis ("sleeping sickness"). PLoS Negl Trop Dis 9:e0003397. https://doi.org/10.1371/journal.pntd.0003397.
- Simpson AG, Stevens JR, Lukes J. 2006. The evolution and diversity of kinetoplastid flagellates. Trends Parasitol 22:168–174. https://doi.org/ 10.1016/j.pt.2006.02.006.
- Poynton SL, Whitaker BR, Heinrich AB. 2001. A novel trypanoplasm-like flagellate Jarrellia atramenti n. g., n. sp. (Kinetoplastida: Bodonidae) and ciliates from the blowhole of a stranded pygmy sperm whale Kogia breviceps (Physeteridae): morphology, life cycle and potential pathogenicity. Dis Aquat Organ 44:191–201. https://doi.org/10.3354/dao044191.
- 6. Steinhagen D, Kruse P, Körting W. 1989. The parasitemia of cloned

*Trypanoplasma borreli* Laveran and Mesnil, 1901, in laboratory-infected common carp (*Cyprinus carpio* L.). J Parasitol 75:685–689. https://doi.org/10.2307/3283050.

- Kruse P, Steinhagen D, Körting W, Friedhoff KT. 1989. Morphometries and redescription of *Trypanoplasma borreli* Laveran & Mesnil, 1901 (Mastigophora, Kinetoplastida) from experimentally infected common carp (*Cyprinus carpio* L.). J Protozool 36:408–411. https://doi.org/ 10.1111/j.1550-7408.1989.tb05536.x.
- Saeij JP, de Vries BJ, Wiegertjes GF. 2003. The immune response of carp to *Trypanoplasma borreli*: kinetics of immune gene expression and polyclonal lymphocyte activation. Dev Comp Immunol 27:859–874. https:// doi.org/10.1016/S0145-305X(03)00083-1.
- Simpson JT, Durbin R. 2012. Efficient de novo assembly of large genomes using compressed data structures. Genome Res 22:549–556. https://doi.org/10.1101/gr.126953.111.
- Boetzer M, Pirovano W. 2014. SSPACE-LongRead: scaffolding bacterial draft genomes using long read sequence information. BMC Bioinformatics 15:211. https://doi.org/10.1186/1471-2105-15-211.
- Tang S, Lomsadze A, Borodovsky M. 2015. Identification of protein coding regions in RNA transcripts. Nucleic Acids Res 43:e78. https:// doi.org/10.1093/nar/gkv227.
- Emms DM, Kelly S. 2015. OrthoFinder: solving fundamental biases in whole genome comparisons dramatically improves orthogroup inference accuracy. Genome Biol 16:157. https://doi.org/10.1186/s13059-015 -0721-2.