

DR. GINA RHIANNA WEBSTER (Orcid ID : 0000-0001-9899-0855)

DR. RAJKO RELJIC (Orcid ID : 0000-0003-4351-8355)

Article type : Research Article

A polymeric immunoglobulin - antigen fusion protein strategy for enhancing vaccine immunogenicity

Gina R. Webster¹, Craig van Dolleweerd², Thais Guerra¹, Szymon Stelter¹, Sven Hofmann¹, Mi-Young Kim¹, Audrey Y-H. Teh¹, Gil Reynolds Diogo¹, Alastair Copland¹, Mathew Paul¹, Peter Hart¹, Rajko Reljic¹ & Julian K-C Ma^{1*}.

¹ Institute for Infection and Immunity, St. George's University of London, UK

² University of Canterbury, Christchurch, New Zealand

*Corresponding author - Professor Julian K-C. Ma

Institute for Infection and Immunity

St. George's Hospital Medical School

Cranmer Terrace

London SW17 0RE, UK

Tel: +44 208 725 5818

jma@sgul.ac.uk

This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the Version of Record. Please cite this article as doi: 10.1111/pbi.12932

This article is protected by copyright. All rights reserved.

Key Words: Plant Biotechnology, Fusion Protein, Tuberculosis, Ag85B, Immune Complex, FcγR

Non-standard abbreviations

ADCC	Antibody-dependent cell-mediated cytotoxicity
APCs	Antigen presenting cells
BCG	Bacille Calmette-Guerin
CHO	Chinese Hamster Ovary
Cys	cysteine
DAPI	4,6-diamidino-2-phenylindole
DLS	Dynamic Light Scattering
ELISA	Enzyme-linked immunosorbent assay
HIV	Human Immunodeficiency Virus
IFN-γ	Interferon gamma
IL	Interleukin
MFI	Mean fluorescent intensity
MPL	Monophosphoryl lipid A
N. benthamiana	Nicotiana benthamiana
PBS	Phosphate buffered saline
PIGS	Polymeric Immunoglobulin G scaffold
poly(I:C)	Polyriboinosinic-polyribocytidylic acid
Pro	proline
RICs	Recombinant immune complexes
RNAi	RNA interference

RU _s	Response Units
SDS-PAGE	Sodium dodecylsulphate-polyacrylamide gel electrophoresis
SEC-UV	Size Exclusion Chromatography-Ultra-Violet spectrophotometry
Ser	serine
SPR	Surface plasmon resonance
TB	Tuberculosis
TB-PIGS	Tuberculosis antigen 85B fused to PIGS
Thr	threonine
TNF- α	Tumour necrosis factor alpha
TTFC	Tetanus toxin fragment C
μ tp	Mu immunoglobulin chain tail piece

Abstract

In this study, a strategy based on polymeric immunoglobulin G scaffolds (PIGS), was used to produce a vaccine candidate for *Mycobacterium tuberculosis*. A genetic fusion construct comprising genes encoding the mycobacterial Ag85B antigen, an immunoglobulin γ -chain fragment and the tailpiece from immunoglobulin μ chain was engineered. Expression was attempted in Chinese Hamster Ovary (CHO) cells and in *Nicotiana benthamiana*. The recombinant protein assembled into polymeric structures (TB-PIGS) in *N. benthamiana*, similar in size to polymeric IgM. These complexes were subsequently shown to bind to the complement protein C1q and Fc γ Rs with increased affinity. Modification of the N-glycans linked to TB-PIGS by removal of xylose and fucose residues that are normally found in plant glycosylated proteins, also resulted in increased affinity for low affinity Fc γ Rs. Immunisation studies in mice indicated that TB-PIGS are highly immunogenic with and without adjuvant. However, they did not improve protective efficacy in mice against challenge with *M.*

tuberculosis compared to conventional vaccination with BCG, suggesting that additional or alternative antigens may be needed to protect against this disease. Nevertheless, these results establish a novel platform for producing polymeric antigen-IgG γ -chain molecules with inherent functional characteristics that are desirable in vaccines.

Introduction

Vaccination against infectious disease has been one of the great medical success stories of the last century. However, there remain many diseases for which vaccines remain undeveloped, as well as diseases where better vaccines are urgently needed. Modern vaccine development also prioritises strategies to improve accessibility, availability and acceptability, which include cost, mode of administration and administration regime.

Vaccines based on recombinant protein antigens are attractive for many reasons, but administration of antigen alone is rarely sufficient to stimulate a potent, long-lasting immune response. In almost all cases, an adjuvant is required to boost immunogenicity or to favour a specific kind of immune response. Adjuvant development is a major challenge for the vaccine industry (Reed et al. 2013). For this reason, antigens that have “self-adjuvanting” properties are of interest, as they would greatly simplify vaccine manufacture and development.

In the early stages of a normal immune response, the formation of natural immune complexes is an important step for targeting antigen presenting cells (APCs), activation of the classical complement cascade and other immunoregulatory functions (Wen et al. 2016). An approach to produce recombinant monoclonal immune complexes has been described using plant

biotechnology, which results in the desired Fc effector functionality and proof of concept for enhanced immunogenicity was described for tetanus toxin (Chargelegue et al. 2005), Ebola virus glycoprotein (Phoolcharoen et al. 2011), *Mycobacterium tuberculosis* Ag85B (Pepponi et al. 2014) and a dengue vaccine candidate (Kim et al. 2015).

An alternative approach to assembling and expressing recombinant immune complexes has recently been described, using a strategy involving a polymeric immunoglobulin G scaffold (Mekhaïel et al. 2011). Hexa-Fc fusion proteins were engineered using a variety of proteins of interest, fused to the Fc region of Human IgG1. The addition of the 18 amino acid tail piece from human IgM resulted in well-defined hexameric structures, which bound differently to complement, Fc Receptors and B-cells compared to their monomeric counterparts (Mekhaïel et al. 2011).

In this study, we investigate the use of the polymeric immunoglobulin G scaffold to create a multivalent vaccine candidate (TB-PIGS) based on the Ag85B antigen of *M. tuberculosis*.

Ag85B is the major component of the Ag85 complex, which in turn, is the major secreted component in mycobacterial culture fluids (Wiker & Harboe 1992). Early promising results with Ag85B led to its inclusion in the vaccine candidate rBCG30 as well as being fused to a plethora of other proteins such as ESAT-6 in subunit vaccine candidates (Horwitz et al. 1995; Horwitz et al. 2000) and it is still a priority candidate for current vaccine developers (Babaki et al. 2017).

We compare the expression of TB-PIGS incorporating mouse IgG2a and human IgG1 sequences, and we also assess their expression in CHO cells and transient expression in *Nicotiana benthamiana*. We report characterisation of the assembly of the TB-PIGS, their size and *in vitro* functional properties. Finally, we describe the immunogenicity of TB-PIGS in a mouse model, as well as the response to challenge with *M. tuberculosis*.

Results

Design and expression of TB-PIGS in wild-type and transgenic ΔXF plants

DNA sequences for murine (Mse γ_{2a}) and human (Hu γ_1) heavy chain scaffolds were synthesised using commercial codon optimisation algorithms for plant and mammalian expression. The scaffolds contained the nucleotide sequence encoding the C_{H2} and C_{H3} domains as well as the last 9-10 amino acid residues of the C_{H1} domain for either Mse γ_{2a} or Hu γ_1 (Figure 1). For Hu γ_1 constructs, Cys²³⁰ was substituted to Ser in the hinge region, to prevent disulphide bond formation occurring, in the absence of light chain. The Ag85B sequence was incorporated at the 5' end of each scaffold sequence.

The μ tp from IgM was added to the C-terminus of the C_{H3} domain of the γ -chains and in both mouse and human versions, Pro⁴⁷⁶ was substituted to Thr to ensure that the terminal residues of the C_{H3} domain of the γ -chain are identical to those of the C_{H4} domain of the μ -chain. This had previously been suggested to help integrate the two immunoglobulin domains together and prevent disruption of IgM polymeric structural abilities (Smith et al. 1995). IgM predominantly forms hexameric molecules in the absence of J-chain and pentameric molecules in the presence of J-chain (Randall et al. 1992). Assembly of TB-PIGS into

monomeric (IgG-like) structures would result in molecular structures of approximate M_r 120K. Subsequent hexameric assembly (as no J-chain will be present) would result in molecular structures of approximate M_r 720K (Figure 1).

Initially, expression was attempted in mammalian CHO cells which were transfected with pEF-DEST51::TB-PIGS vectors, using both murine and human-based scaffolds. The transfected CHO cells were selected using blasticidin. Although CHO cell transfections were attempted repeatedly, no successful transfectants were recovered for TB-PIGS. Successful transfections were however recovered on each occasion for a control PIGS protein, comprising the immunoglobulin scaffold without antigen (Supplementary Figure S1).

Two *N. benthamiana* plant lines were vacuum infiltrated with *Agrobacteria* containing pTRAk.2::TB-PIGS vectors, using both murine and human-based scaffolds. Wild-type *N. benthamiana* results in recombinant protein production with typical plant glycosylation, whereas a glyco-engineered *N. benthamiana* line in which xylosyl- and fucosyl-transferases were deleted, results in glycoproteins with glycans that are more similar to those commonly found in mammalian systems (Strasser et al. 2008). Five days post-infiltration, the leaves were harvested and soluble protein was extracted. Recombinant antibody molecules were purified by protein A affinity chromatography.

Expression and assembly of TB-PIGS

Plant samples were analysed by SDS-PAGE and Western blotting using anti-serum specific for Ag85B, human γ 1 or mouse γ 2a immunoglobulin chains (Figure 2). Under non-reducing conditions, coomassie blue stained SDS-PAGE demonstrates three major protein groups for both Human TB-PIGS and Murine TB-PIGS (Figure 2A panels i-ii). There was a major band at approx. $M_r \sim 120K$ (labelled M), which is the predicted size for the PIGS monomer (Figure 1). Protein bands of higher M_r are also present ($\sim 250K$ and greater, labelled P), which are likely to represent polymeric structures. There are also major bands at $M_r \sim 60K$ (labelled S) which may be single chain, unassembled fusion proteins, or degradation products. For comparison, commercial purified human IgG1, human IgM and murine IgG2a were also analysed on the same gels. Under reducing conditions, the human TB-PIGS sample resolved into a predominant band of approximate $M_r \sim 60K$, which is likely to represent unassembled single fusion proteins (Figure 2A panels iii-iv). A similar size band was observed for the murine TB-PIGS, but the predominant band was $M_r \sim 100K$, which may be a dimer. Some lower molecular bands were also evident, particularly in the case of the Human TB-PIGS, which are likely to represent truncated degradation products. For comparison, reduced samples of commercial purified human IgG1, human IgM and murine IgG2a were also run, demonstrating bands at the expected M_r for immunoglobulin heavy and light chains.

The identity of the bands was supported by Western blotting using antigen and antibody-specific antisera. Under non-reducing conditions, all the major protein bands in the human and murine TB-PIGS samples were detected using both anti-Ag85B and anti-immunoglobulin heavy chain (anti-Human γ 1 or anti-Mouse γ 2a) antisera (Figure 2B panels i-iii). Under reducing conditions, the major $M_r \sim 60K$ was detected with both anti-Ag85B and

anti-immunoglobulin heavy chain antisera, but the lower M_r bands <37K were only detected by the anti-immunoglobulin heavy chain antisera (Figure 2B panels iv-vi).

Similar results were obtained when the human and murine TB-PIGS were expressed in glyco-engineered (Δ XF) *N. benthamiana* (data not shown). Successful glyco-engineering was confirmed by Western blotting using specific antisera to confirm the absence of xylose and fucose residues on recombinant proteins expressed in Δ XF plants (results not shown).

Fully assembled hexameric TB-PIGS are very large proteins that would be difficult to visualise by SDS-PAGE. Two different systems were therefore utilised to demonstrate polymeric assembly of TB-PIGS more accurately; native PAGE (Figure 2C), and size exclusion chromatography-ultraviolet spectrophotometry (SEC-UV; Figure 2D).

Native PAGE (Figure 2C) indicated that human TB-PIGS assembled into four major protein bands, that were consistent with polymers (800-1000K), dimers (two bands ~240-300K) and probably monomers (~140K). The murine TB-PIGS however appeared predominantly as a single band (~250K). Commercial human and murine IgM standards were run in parallel. Under, SEC-UV (Figure 2D), the murine TB-PIGS (expressed in Δ XF plants) resolved into two peaks (retention time 19.2 and 20.5 mins), which are consistent with results expected for dimer/monomer and a single chain. Four species were observed in the human TB-PIGS: Δ XF sample; with retention times consistent with those expected for polymers, dimers, monomers and single chain. The molecular weights of TB-PIGS species were determined by comparison with retention times for proteins of known molecular weight within the BioRad standards

sample (from left to right - Vitamin B12 (1.35K), myoglobin (17K), ovalbumin (44K), γ -globulin (158K) and thyroglobulin (670K)). Yield estimations are provided in supplementary table S1.

Functional characterisation of TB-PIGS

To assess functionality of TB-PIGS, their binding to C1q (the first component of the classical complement pathway) was determined by ELISA (Figure 3A). Both the human and murine TB-PIGS, produced in wild-type and glyco-engineered Δ XF plants, bound to C1q in a concentration dependent manner. WT crude plant extract, Ag85B, MseIgG2a and HuIgG1 (all used as negative controls) only resulted in minimal or no binding to C1q. In contrast, ICMs (immune complex mimetics positive control; consisting of murine IgG1 monoclonal antibody bound to polymeric *M. tb* antigen Acr in an immune complex), bound to C1q.

Surface plasmon resonance (SPR) was used to determine affinity and kinetics of binding between human TB-PIGS and the high affinity receptor CD64 (Fc γ RI) or low affinity receptors CD16a and CD32a (Fc γ RIIIa and Fc γ RIIIa) (Supplementary Figure S2 and Table S2). This experiment was to determine the individual interaction of Fc γ R with a single ‘arm’ of the polymeric TB-PIGS, to determine that the Fc γ R-binding site on the engineered IgG-like structure retains its proper activity. Human TB-PIGS produced in Δ XF plants had high affinity binding to CD64 (0.41nM) which was similar to the affinity of human IgG1 with the same receptor (0.27nM) as expected. Human TB-PIGS expressed with normal plant glycosylation had approximately 10-fold lower affinity binding to CD64. The affinity of human TB-PIGS to CD16a was superior to human IgG1 as expected. In particular, the K_D for

glyco-engineered human TB-PIGS was 19.7 nM compared with the K_D for human IgG1 of 673 nM. The binding affinities of human IgG1 and human TB-PIGS expressed in WT or Δ XF plants were similar, and in the micromolar range.

SPR was also used to demonstrate differences in avidity between glycoengineered human TB-PIGS: Δ XF with low affinity Fc γ Rs, compared to monomeric (commercial) HuIgG1. The experimental set-up involved tethering multiple Fc γ Rs to the chip surface allowing polymeric TB-PIGS to bind to multiple receptors simultaneously, allowing avidity measurements. Although avidity cannot be quantified accurately in this assay, the differences between TB-PIGS: Δ XF and HuIgG1 are evident from the dissociation curves (Figure 3B and C). For both CD32a and CD16a, the dissociation of HuIgG1 is rapid and almost complete within seconds. In contrast, the TB-PIGS: Δ XF dissociate more slowly, particularly from CD32a.

Binding of TB-PIGS to monocyte cell lines

SPR demonstrated binding of TB-PIGS to soluble Fc γ Rs. This activity was next confirmed *ex vivo* by demonstrating binding to human TB-PIGS by THP-1 cells (a human monocyte cell line expressing Fc γ Rs on their cellular surface) by both flow cytometry (Supplementary Figure S3) and confocal microscopy (Figure 4).

THP-1 cells were pre-treated with commercial “Fc block”, to block binding to CD64, then incubated with either medium, a rabbit IgG antiserum, or HuIgG1 as controls, and human TB-PIGS produced in Δ XF plants as indicated. THP-1 nuclear staining was performed with DAPI (blue- first column). Detection of antibody or TB-PIGS binding to the cells was with

either rabbit α -Ag85B antiserum, followed by a goat α -Rabbit Ig-Alexa Fluor488 antiserum (green– second column), or sheep α -HuIgG1 followed by donkey α -sheep IgG-Alexa Fluor633 (magenta– third column). Superimposed images are shown in the fourth column.

After blocking CD64, HuIgG1 did not bind to the low affinity Fc γ Rs as expected (Figure 4 row 3). Human TB-PIGS: Δ XF binding to THP-1 was detected using both α -Ag85B and α -HuIgG1 antisera (Figure 4 row 4). This result was supported by the flow cytometry analysis, which demonstrated very low level of fluorescence for HuIgG1 (Supplementary Figure S3), increased fluorescent signal for human TB-PIGS: Δ XF, consistent with the signal for heat aggregated IgG1 which was used as the positive control.

Similar results were observed for binding of murine TB-PIGS to Fc γ Rs on the surface of J774 cells (a mouse monocyte/macrophage cell line), although in this case, high affinity receptors were not blocked so binding to these receptors was observed using MseIgG2a control (Supplementary Figure S5). Use of phalloidin and DAPI staining for cytoplasm and nuclei, respectively, indicated that TB-PIGS bind to receptors on the surface of the cell and may be taken-up by the cell (Supplementary Figures S4).

***In vivo* immunogenicity and protective efficacy elicited by TB-PIGS**

BALB/c mice were immunised with murine TB-PIGS, produced in both wild-type and Δ XF *N. benthamiana*, with and without adjuvant. TB-PIGS were highly immunogenic, but plant-produced Ag85B alone was not (Figure 5 A-C). An Ag85B specific antibody response was observed in all animals immunised with Murine TB-PIGS (Figure 5A). Murine TB-PIGS

alone in particular induced a strong IgG1 (Th2-type) response but this shifted to a mixed Th1/Th2 response with the addition of MPL adjuvant, as indicated by production of α -Ag85B-specific IgG2a antibodies.

Ag85B specific T-cell proliferative responses were also observed following *in vitro* stimulation of splenocyte cultures with Ag85B. Figures 5B and 5C show the analysis of specific poly-functional CD4⁺ and CD8⁺ cells respectively. Surprisingly, the PBS control group was found to have some specific CD4⁺ T-cell response. However, TB-PIGS from Δ XF *N. benthamiana* but not WT plants induced a higher proportion of triple positive cells compared to either PBS or BCG groups. Addition of MPL adjuvant increased the proportion of double positive cells but abolished the triple positive cells, similar to antigen stimulation alone. In the CD8⁺ T-cell compartment, the striking finding was that TB-PIGS from Δ XF *N. benthamiana* induced exclusively triple positive cells and was superior to all other groups, including when combined with the MPL adjuvant. Overall, the data showed that TB-PIGS from Δ XF *N. benthamiana* are highly potent in inducing triple positive CD4⁺ and CD8⁺ T-cells and that this was largely reversed by the addition of MPL adjuvant.

Finally, the potential protective efficacy of TB-PIGS was assessed following intranasal aerosol challenge with *M. tuberculosis*. By assessment of recoverable bacterial counts from lungs and spleens, no improvement in protective efficacy compared to PBS immunised mice was observed (Figure 5 D).

To test the human TB-PIGS, a similar study was performed using transgenic CD64 (human Fc γ RI) mice. Immunisation was with human TB-PIGS, produced in Δ XF *N. benthamiana*, with and without BCG priming. Two adjuvants were tested, MPL and poly(I:C).

As with the Murine TB-PIGS, an Ag85B specific antibody response was elicited in all animals immunised with Human TB-PIGS (Figure 6A). Similar mixed IgG1/IgG2a responses were seen in the three groups, all of which received an adjuvant. BCG priming did not have a significant effect.

Analysis of splenocyte cytokine responses (Figure 6B) also suggests a mixed Th1/Th2 response. All groups except PBS + MPL control group, resulted in high levels of IFN- γ . If mice did not receive BCG, statistically lower levels of IL-2 were produced. Poly(I:C), with or without BCG priming, resulted in increased levels of TNF- α , compared to control mice or those receiving MPL adjuvant. However, the most balanced cytokine response appeared in the group that was primed with BCG and received Human TB-PIGS: Δ XF + MPL adjuvant. Furthermore, analysis of polyfunctional T-cells (Figure 7A) indicated that this was the only group in which CD4⁺ quadruple cytokine producing T-cells were elicited. A high proportion of triple (IFN- γ , IL-2 and TNF- α) and double (IFN- γ and IL-17) cytokine producing CD4⁺ cells was also observed in this group, as well as in the BCG primed group that received human TB-PIGS: Δ XF with poly(I:C) adjuvant. CD8⁺ cells were largely restricted to single cytokine producers in all groups, except for this latter group that was primed with BCG and boosted with human TB-PIGS: Δ XF + poly(I:C) adjuvant. Here there was a significant proportion of double and triple cytokine producing T-cells (Figure 7B). In both cases, mice

primed with BCG and boosted with human TB-PIGS resulted in greater numbers of α -Ag85B-specific T-cells compared to control groups.

Finally, none of the vaccine candidate groups resulted in a statistical reduction in bacterial load in the lungs, compared to mice immunised with BCG (Figure 7C left). However, mice primed with BCG and boosted with human TB-PIGS: Δ XF + poly(I:C) adjuvant resulted in a statistically significant reduction in bacterial load in the spleens, compared to mice immunised with BCG (Figure 7C right).

Discussion

Tuberculosis is a major global health problem, due to limitations of the BCG vaccine, the rise of multidrug resistant and extensively drug resistant TB cases, as well as the increased prevalence of immunocompromised people (Cayabyab et al. 2012). A considerable effort is therefore being made to develop a new TB vaccine to either replace or boost BCG-immunisation. Ag85B is an established vaccine candidate and was used in this study as a model antigen to assess a novel molecular platform for vaccine design. PIGS is an attractive platform because it is designed to present multiple copies of antigen whilst also directly targeting APCs through Fc receptors to enhance antigen uptake and presentation.

Two expression systems were tested for production of the TB-PIGS. CHO cells were selected because they are currently the industry standard for recombinant immunoglobulin pharmaceuticals, and transient expression in *Nicotiana benthamiana*, an emerging plant expression system with significant potential advantages in relation to pharmaceuticals that are

urgently needed in low and middle income countries (Chen & Davis 2016). *Nicotiana* plants are well documented to express antibodies and antibody complexes extremely efficiently and accurately (Chargelegue et al. 2005; Hiatt et al. 1989; Ma et al. 1995; Teh et al. 2014). Unexpectedly, we were unable to express any TB-PIGS in CHO cells. The PIG scaffold alone, lacking Ag85B, was expressed successfully in CHO cells, suggesting a protein specific issue, such as toxicity of the antigen or the overall size of TB-PIGS.

We generated a murine $\gamma 2a$ scaffold first because we planned to perform pilot *in vivo* testing in mice, but we also designed and expressed a human $\gamma 1$ in anticipation of developing a vaccine product. Both could be expressed in *Nicotiana benthamiana* although differences in polymerisation states were observed. Native PAGE indicated that the human scaffold resulted in a greater proportion of polymers (~35-60%) compared to monomers (~1-10%), whereas Mse $\gamma 2a$ scaffolds resulted in predominantly monomers (~45-60%) compared to polymers (~5-20%). This finding was partly corroborated by SEC-UV spectrophotometry. Mekhaïel *et al.* also found that Human IgG1-based hexa-Fc constructs could form hexameric structures, yet, their Mouse IgG2a-based constructs did not form anything larger than a dimer (Mekhaïel et al. 2011). The reason for these differences is unknown. Further improvement in polymer yields may require additional mutations, such as the L309C mutation in human IgG1 heavy chain, which has been indicated to increase yields of polymers, as it is homologous to Cys414 in IgM, which has been shown to have a role in inter-heavy chain disulphide bonds (Mekhaïel et al. 2011, Sorensen et al. 1999, Yoo et al. 1999). However, this mutation does not always improve polymer yields (Teye et al. 2017).

The PIG scaffold is designed to mimic polymeric IgM and the Fc mediated properties of that molecule. Functional assessment of the TB-PIGS included assays to demonstrate binding to complement and Fc γ receptors. The classical complement pathway is normally activated by polymeric antigen-bound IgG or IgM binding to C1q, leading to activation of the enzyme cascade. IgG normally exists in a monomeric state, in which the C1q binding site is accessible but the affinity is too low to bind and activate C1q (Diebolder et al. 2014). IgG needs to polymerise to bind C1q with high avidity (Diebolder et al. 2014). The demonstration that TB-PIGS could bind to C1q was therefore an indication of their polymeric status. These results support similar findings by Smith *et al.* but contrast with Mekhaïel *et al.*, who showed that hexa-Fc fusion molecules bound C1q poorly compared to dimers and monomers (Mekhaïel et al. 2011; Smith & Morrison 1994; Smith et al. 1995). Mekhaïel et al., also showed that the interaction of their hexa-Fc molecules with Fc γ Rs was dependent on the fusion partner. They hypothesised that these results may be due to steric hindrance, caused by the antigen blocking the C1q or Fc γ R binding site located in or near the hinge region. Therefore, in our constructs, we included the last 9-10 amino acid residues of the C_H1 domain, corresponding to a β -strand, to add molecular flexibility and to avoid any potential steric interference.

The only known difference between antibodies expressed in plants and those expressed by mammalian cells is in N-glycosylation. Although the initial steps of N-glycosylation and N-glycan processing are highly conserved between plants and mammals, subsequent N-glycan maturation differs in plants resulting in complex N-glycans with β 1,2-linked xylose, core α 1,3-linked fucose and Lewis A-type structures that are not found in mammalian antibody glycosylation (Cabanes-Macheteau et al. 1999; Strasser 2016). Glycosylation is known to impact molecular interactions of IgG (Hayes et al. 2014; Hodoniczky et al. 2005; Shields et

al. 2002; Zhou et al. 2008). In particular, *N*-linked glycan structures present at Asn 297 in IgG, can modulate Fc γ R binding, which affects the immune response induced (Mimura et al. 2001). For example, removal of fucose has been shown to improve binding of antibody to Fc γ RIIIa and results in enhanced antibody dependent cell-mediated cytotoxicity (ADCC) (Ferrara et al. 2011; Shields et al. 2001), so here we made use of a glyco-engineered line of *N. benthamiana* (Strasser et al. 2008) in which the endogenous α 1,3-fucosyltransferase was silenced. Our results showed that the use of the glyco-engineered *Nicotiana benthamiana* host had no impact on expression or assembly of TB-PIGS. There was no functional loss associated with TB-PIGS expressed in glyco-engineered plants, for example they bound equally well to C1q, but there was an important enhancement of binding affinity to CD64 and CD16a as predicted from previous reports (Hiatt et al. 2014).

Physiologically, IgG binds with high affinity to Fc γ RI/CD64, whereas immune complexes bind preferentially to low affinity receptors (Fc γ RII/CD32 and Fc γ RIII/CD16) (Klaassen et al. 1988; Lux et al. 2013; White et al. 2001). In our studies, the human TB-PIGS bound with high affinity to Fc γ RI/CD64 and the glyco-engineered human TB-PIGS bound with enhanced affinity to Fc γ RII/CD32a and Fc γ RIII/CD16a. Potentially, this indicates that like naturally formed immune complexes, human TB-PIGS could interact efficiently with immature dendritic cells (DCs) (reviewed in Platzer et al. 2014), resulting in endocytosis and leading to maturation and migration of the DCs to secondary lymphoid organs, where cytokines are produced that activate specific naïve T-cells. *M. tuberculosis* has evolved to evade the human immune system, for example, by ‘hiding’ inside macrophages. This indicates that a vaccine candidate should elicit T-cell responses (reviewed in Scriba et al. 2017). Recently, B cells and antibodies have also been indicated to be important in controlling *M. tuberculosis* disease

(reviewed in du Plessis et al. 2016). Moreover, phagocytosis and ADCC may be impaired during active tuberculosis disease, possibly due to a decrease in antibody avidity (Arias-bouda et al. 2003; Perley et al. 2014) and/or downregulation of Fc receptor expression (Jacobsen et al. 2007; Laux da Costa et al. 2015; J. Maertzdorf et al. 2011; Jeroen Maertzdorf et al. 2011). Therefore, improvement of antibody avidity and Fc receptor engagement may be a crucial area of research for protection against *M. tuberculosis*.

Mouse TB-PIGS affinities for mouse Fc γ Rs were inconsistent, but generally lower than observed for human TB-PIGS against human Fc γ Rs (data not shown). These differences may be due to less efficient polymerisation of the PIG scaffold. In addition, murine Fc γ R have been less well studied than their human counterpart, so there may be unrecognised biological differences. The classification of murine Fc γ R homologs is based on amino acid similarity, rather than ligand binding or cellular expression (Bruhns 2015), for example, mouse Fc γ RIV has been suggested to be the functional equivalent of both human Fc γ RI and human Fc γ RIIIa (Mechetina et al. 2002; Nimmerjahn et al. 2005). Investigation of half-lives for murine TB-PIGS with mouse Fc γ RI were inconclusive, but suggested that the dissociation is much faster than for human reagents (data not shown), which is a consideration in the analysis of *in vivo* results.

In vivo murine and human TB-PIGS were tested in conventional and human CD64 transgenic BALB/c mice respectively. Murine TB-PIGS (produced in wild-type and Δ XF plants) generated humoral and cellular immunity with and without adjuvant, but failed to reduce the bacterial load in challenged mice, compared to BCG immunised mice. Murine TB-PIGS: Δ XF did however, result in a greater percentage of triple-cytokine producing CD4⁺ and CD8⁺ cells,

which have been suggested to be important in protecting against *M. tuberculosis* infection. In the second *in vivo* study, human TB-PIGS were used to immunise transgenic CD64 mice with and without BCG priming. The best IgG2a-specific response with quadruple-cytokine producing CD4⁺ cells was generated by mice primed with BCG and boosted with human TB-PIGS:ΔXF + MPL adjuvant. In contrast, the greatest proportion of triple-cytokine producing CD8⁺ cells was found for mice immunised with BCG and boosted with human TB-PIGS:ΔXF + poly(I:C). CFU counts indicated this was the most promising vaccine candidate as there was a statistically significant reduction in bacterial burden in spleens of challenged mice compared to BCG immunised mice, suggesting that this vaccine candidate could impair MTB dissemination *in vivo*. However, only a marginal reduction was observed in lungs of challenged mice, which is important as the lungs are the primary site of entry for the pathogen.

Tuberculosis poses some fundamental challenges in the evaluation of vaccine candidates. There are questions about the predictive value of animal models for human disease, and there is a lack of correlates of vaccine-induced protection (Karp et al. 2015). In addition to testing humoral and cellular immunity, we tested protection against mycobacterial challenge, but this is a demanding end-point for *in vivo* assessment of TB-PIGS as a vaccine candidate for a number of reasons. Importantly, Ag85B, although extensively studied, has not yet been demonstrated to provide consistent protection against infection. So its role in this study was as a model immunogen, rather than as a confirmed vaccine protein. Secondly, one of the main attractions of the PIG scaffold is its potential for targeting Fcγ receptors on APCs. As noted above, the biology of murine Fcγ receptors in relation to their human counterparts still needs clarification, and as our results suggested less polymerisation for murine TB-PIGS and

diminished binding affinity of murine TB-PIGS to murine FcRs compared with human TB-PIGS to human FcRs, the murine model may not be optimal for protection studies. Furthermore, although we were able to test the human TB-PIGS in human CD64 transgenic mice, the animals were not transgenic for low affinity FcγRs such as CD16a, which are known to play an important role in ADCC (Gupta et al. 2005).

In this study, we have demonstrated the feasibility of polymeric IgG scaffolds as a vaccine platform. We illustrated expression, assembly and purification from *Nicotiana benthamiana*, and demonstrated *in vitro* and *in vivo* functional characteristics based on our hypothesis that PIGS would promote enhanced antigen presentation to the immune system. It builds on a parallel project using a dengue vaccine candidate (Kim et al. 2017 and Kim et al. 2018) and demonstrates the generic potential of this platform to be used for vaccine candidates against a range of infectious diseases. Further work is now needed to optimise the formation of polymers, to enhance their stability and to maximise yields in the plant expression system in order to advance to safety and immunisation studies in humans.

Experimental Procedures

Construction of TB-PIGS vectors

GeneArt synthesised four codon optimised constructs in pMA-T vector. Two genes were codon optimised for *N. tabacum* (human PIGS and murine PIGS) and the other two for *H. sapiens* (human PIGS^H and murine PIGS^H). The PIGS sequence included leader sequence, truncated γ-chain (derived from either human IgGγ₁ or mouse IgGγ_{2a}) and μtp from IgM, all flanked by NcoI and XbaI restriction sites as well as attB gateway cloning sites.

Primer pairs containing BsaI restriction sites: 5'-cagagcggctcgcgatgcgaggtgttctctcgtcctggttgct-3' and 5'-

tggtcaggtctcaaagcacctgctcccaaagaagattgaag-3' were designed to amplify *N*-terminally truncated Ag85B (signal peptide removed) DNA fragment from pGEM::Ag85B-ESAT6 construct (Dedieu et al. 2010) into pMA-T plasmid containing PIGS sequence by PCR in order to create pMA-T::human TB-PIGS and pMA-T::murine TB-PIGS.

AttB1 and attB2 gateway cloning sites were used to clone Ag85B-human PIGS and Ag85B-murine PIGS into pDONR-Zeo, using the BP Clonase II enzyme protocol (Invitrogen). The flanking NcoI and XbaI sites were used to clone the desired genes into pTRAk.2 (plant expression destination vector; van Dolleweerd et al. 2014; Sack et al. 2007) using T4 DNA Ligase (NEB).

Expression of TB-PIGS and Ag85B in plants

E. coli DH5 α cells (Invitrogen) were transformed to amplify the plasmid, prior to electroporation of *A. tumefaciens* (strain GV3101::pMP90RK). A 400 ml *A. tumefaciens* culture was grown, centrifuged at 4,000 rpm for 20 minutes, and the pellet re-suspended in plant infiltration buffer (10 mM MES, 10 mM MgCl₂ and 100 μ g/ml acetosyringone at pH 5.6) to an OD₆₀₀ of 0.25. 6-8 week old *N. benthamiana* plants were immersed in *A. tumefaciens* solution inside a vacuum chamber with a vacuum pressure of <100 mbar for ~2 minutes. After infiltration, the plants were maintained at 26°C for ~5-6 days prior to leaf harvest for recombinant protein analysis.

Purification and characterisation of TB-PIGS from plants

Whole leaves were frozen in liquid nitrogen and PBS (pH 7.0) was used as extraction buffer. A blender was used to homogenise the leaf material and then passed through miracloth (Merck) prior to centrifugation at 17,000g for 30 minutes. The supernatant was syringed through a 0.22 μm filter before being purified on protein-A-sepharose (SIGMA), except Ag85B which was purified on a nickel (II) affinity column.

Eluted fractions were pooled, concentrated using Amicon Ultra-4 centrifugal filter devices and dialysed to exchange buffer for PBS (pH 7.0). The integrity of the proteins was determined by SDS-PAGE on NuPAGE 4-12% Bis-Tris gels (Novex), both non-reducing and reducing (10% β -mercaptoethanol), and the gels were stained with Coomassie Instant Blue stain (Expedeon). Additionally, NativePAGE 3-12% Bis-Tris gels (Novex) were utilised with NativeMark Unstained Protein Standard as per manufacturer's protocol. ImageJ software was used to analyse the intensity of bands within each lane of the gel. A chromatogram was plotted and integration of the area under the curve used to determine a value for the percent of each species present.

Proteins were also separated on a high performance SEC Yarra 300x7.8 column utilised on a HPLC LC2010A machine with PBS (pH 7.0) as buffer. Eluted fractions were compared against known high molecular weight standards (BioRad).

C1q binding ELISA

Microtiter wells were coated with 5 µg/ml of human C1q (Merck Calbiochem) in PBS (pH 7.0) and incubated at room temperature for two hours prior to blocking with 5% skimmed milk for 2 hours. The purified proteins, in PBS, were added to wells at a starting concentration of 100 µg/ml and in 5-fold dilutions in blocking buffer. Plates were incubated at room temperature for 2 hours before being washed with dH₂O + 0.1% Tween20. Plates were incubated with detection antiserum (α -Hu IgG γ ₁-HRP, α -Mse IgG γ ₁-HRP, or α -Mse IgG γ _{2a}-HRP), in blocking buffer, for 2 hours. Finally, plates were washed with dH₂O + 0.1% Tween20 before being developed with 50 µl of HRP substrate in buffer (Sigmafast OPD tablet and Sigmafast buffer with urea H₂O₂ tablet in 20 ml dH₂O). Absorbance was measured at 450nm (Tecan Infinite F200 PRO).

Surface plasmon resonance

SPR experiments were performed using the Biacore X100 instrument (GE Healthcare). Filter-sterilised HBS-EP⁺ buffer was used as running buffer and dilution buffer. For determining affinity values; flow cells of CM5 sensor chips were coupled with recombinant protein A (SIGMA) using amine-coupled chemistry as described by the manufacturer. TB-PIGS were captured onto the surface of the chip. For analyte HuCD64 or MseCD64 (R&D systems) a flow rate of 40 µl/min, a contact time of 135 seconds, dissociation time of 600 seconds and regeneration buffer of 100 mM glycine (pH 1.5) was used. For analyte HuCD32a and HuCD16a (R&D systems) a flow rate of 40 µl/min, a contact time of 90 seconds, dissociation time of 300 seconds and regeneration buffer of 100 mM glycine (pH 1.5) was used. In all experiments, data were zero adjusted and evaluated using Biacore x100 software. For determining avidity; a His capture kit (GE Healthcare) was used to bind anti-histidine

antibody to the surface of a CM5 chip using amine-coupled chemistry. HuCD32a or HuCD16a were captured onto the surface of the chip. For analyte TB-PIGS a flow rate of 35 $\mu\text{l}/\text{min}$, a contact time of 90 seconds, a dissociation time of 300 seconds and His capture kit regeneration buffer was used.

Confocal Microscopy

If Human FcR block (BD Pharmingen) was utilised, cells were incubated with 2.5 μg per 1×10^6 cells for 20 minutes at room temperature before addition of 25 μg of PIGS or control protein (HuIgG1 or MseIgG2a). Cells were incubated at room temperature for 2 hours before supernatant was discarded. Cells were washed with PBS before addition of 150 μl blocking buffer (PBS + 2% FBS + 1% BSA + 0.2% Triton X-100 + 0.05% Tween20) per well. The plate was incubated at room temperature for 1 hour before washing with PBS and fixing cells with 4% para-formaldehyde (SIGMA) for 20 minutes. Cells were washed with PBS + 7% glycine and permeabilised with 0.1% Triton X-100 for 5 minutes. Again, cells were washed with PBS before addition of primary antibodies (sheep α -HuIgG (TBS), sheep α -MseIgG (TBS), rabbit α -Ag85B (Abcam) or phalloidin-Alexa488 (LifeTechnologies)) in blocking buffer. Plates were incubated for 1 hour at room temperature, in the dark. Cells were washed with PBS and then secondary antibodies (donkey α -sheep IgG-AlexaFluor633 (Life Technologies) or goat α -rabbit IgG-AlexaFluor488 (Abcam)), in blocking buffer, was added for 1 hour, in the dark. Finally, cells were washed with IF wash (PBS + 0.2% Triton X-100 + 0.05% Tween20), before DAPI was added for 15 seconds, washed twice more with IF wash and then addition of 200 μl PBS and stored at 4°C in the dark until imaging using a Zeiss Confocal LSM 510 and analysis using ImageJ software.

***In vivo* immunisation of mice**

All animals were housed in the Biological Research Facilities (BRF) at St. George's, University of London, under Home Office regulations. Six to eight-week old female BALB/c mice purchased from Charles River/Harlan were used in all experiments with murine TB-PIGS and were maintained under specific pathogen free conditions. Previously determined positive human CD64 mice were used to breed off-spring and tail-bleeds were screened by PCR to identify positive human CD64 transgenic mice. Infected animals were housed in cages contained within purpose-built plastic isolators under negative pressure, in a containment level 3 facility.

Mice were immunised with TB-PIGS at 25 µg/dose administered subcutaneously (s.c.) or intranasally (i.n.) at 3 week intervals on three separate occasions, either alone or suspended with 20% v/v MPL (SIGMA) or poly(I:C) adjuvant (Invivo Gen). BCG priming was performed 9 weeks before first boost, by immunising mice with 2×10^6 CFU/dose (BCG-Pasteur).

3 mice per vaccine group were culled 3-4 weeks after final immunisation. Mice were anaesthetised under isoflurane and cardiac puncture performed to collect blood. Spleens were removed into 5 ml R10 media. Blood was placed into a 37°C incubator for 30 minutes, then at 4°C for 2 hours, centrifuged at 13,000 rpm for 15 minutes and serum collected and stored at -20°C. Spleens for each group were combined and macerated through a 70 µm cell strainer using a sterile syringe plunger and washed into a petri-dish with 25ml of R10 media. Cells were centrifuged at 400g for 10 minutes and re-suspended in 5 ml/spleen ACK red blood cell

lysis buffer (ThermoFisher Scientific) for 7 minutes. 40 ml of R10 media was added to halt the reaction, cells were centrifuged for 10 minutes at 400g, washed with 20 ml R10 buffer, centrifuged 10 minutes at 400g and pellet re-suspended in 10 ml R10 buffer. Viable cells were counted using 0.4% trypan blue dye, counting slides and BioRad TC20 automated cell counter.

Humoral Responses

Microtiter wells were coated for two hours at 37°C with 10 µg/ml of *N. benthamiana* produced Ag85B protein in PBS (pH 7.0). Plates were washed with dH₂O before being blocked for 1 hour, at 37°C, with PBS + 5% BSA. Plates were washed with dH₂O. The mouse serum was added to wells (1:100) and a 3-fold serial dilution in blocking buffer. Plates were incubated at 37°C for 2 hours before being washed again with dH₂O. Plates were incubated with secondary antibody (α -Mse IgG-HRP (TBS), α -Mse γ_1 -HRP (Abcam) or α -Mse γ_{2a} -HRP (STAR)), in blocking buffer, for 1 hour at 37°C. Finally, plates were washed with dH₂O before being developed with 50 µl of HRP substrate in buffer (Sigmafast OPD tablet and Sigmafast buffer with urea H₂O₂ tablet in 20 ml dH₂O) and the reaction stopped with 25 µl 2M H₂SO₄. Absorbance was measured at 450 nm (Tecan infinite F200 PRO).

Cytokine responses

Splenocytes were seeded at a concentration of 5 x 10⁵/ml per well in a 96-well U-bottomed plate (Corning). Cells were stimulated with 10 µg/ml of *N. benthamiana* produced Ag85B or 5 µg/ml concanavalin A (ConA; SIGMA) as a positive control or R10 media as a negative control. Plates were incubated at 37°C, 5% CO₂ and 90% humidity for 72 hours and

supernatants were harvested for cytokine ELISAs. Cytokine ELISAs were performed as per manufacturer's instructions (eBiosciences). eBiosciences Mouse Th1/Th2 ELISA Ready-SET-Go! kit detected IFN- γ , IL-2 and IL10. Mouse TNF α ELISA Ready-SET-Go! kit detected TNF α .

Polyfunctional T-cell analysis

Splenocytes were seeded at a concentration of 1.5×10^6 /ml per well in a 96-well V-bottomed plate. Cells were stimulated with R10 media containing 100 μ g/ml of Brefeldin A (SIGMA) and 5 μ g/ml of *N. benthamiana* produced Ag85B or 100 μ g/ml Brefeldin A, 4 μ g/ml Phorbol Myristate Acetate (PMA; SIGMA) and 20 μ g/ml ionomycin calcium salt (SIGMA) as a positive control or 100 μ g/ml Brefeldin A as a negative control. Plates were incubated at 37°C, 5% CO₂ and 90% humidity for 4 hours. Plates were centrifuged at 400g for 5 minutes at 4°C, and supernatant discarded. All cells (except two wells which were left as negative controls) were stained with viability dye eFluor 780 (1:2000; eBioscience) in PBS for 30 minutes at 4°C, in the dark. Plates were washed with 200 μ l PBS, centrifuged at 400g for 5 minutes, and supernatant discarded. 100 μ l Cytfix (BD) was added to each cell pellet and incubated at 4°C for 30 minutes, in the dark. The plates were washed with 200 μ l FACS buffer, centrifuged at 400g for 5 minutes, and supernatants discarded. Pellets were re-suspended in 200 μ l FACS buffer and stored at 4°C, in the dark, overnight.

The plates were centrifuged at 400g for 5 minutes and supernatants were discarded. 200 μ l Permeabilisation Buffer (PBS + 0.5% BSA + 0.1% sodium azide + 0.5% saponin) was added to each well, centrifuged at 400g for 5 minutes and supernatant discarded. 25 μ l of staining

cocktail (eFluor 780, α -TNF- α -APC, α -IL2-PE, α -IL17-Cy7, α -IFN- γ -PE-Dazzle, α -CD3-FITC, α -CD4-PerCP-Cy5.5, and α -CD8-AlexaFluor700) was added to each pellet and incubated for 45 minutes at 4°C, in the dark. Fluorescence Minus One (FMO) stains were prepared as necessary and added to appropriate wells for 45 minutes at 4°C, in the dark. Similarly, Ultra compensation beads (eBiosciences) were stained for 45 minutes at 4°C in the dark and used for compensation. Cells were washed in 200 μ l Permeabilisation buffer, centrifuged at 400g for 5 minutes, and supernatants discarded. Cells were washed in 200 μ l FACS buffer, centrifuged at 400g for 5 minutes, and supernatants discarded. Finally, cells were re-suspended in 200 μ l FACS buffer and stored at 4°C until acquisition on the flow cytometer (LSRII; BD). Analysis performed using FlowJo version 10.

***M. tuberculosis* challenge**

Mice were transferred to Category III facilities and infected with aerosolised 200-500 CFU *M. tuberculosis* H37Rv for 10 minutes using a Biaera machine with Aero MP software (Management Platform) with nose-only chambers. They were culled 3 weeks *post*-challenge.

Lungs and spleens were removed and homogenised in 5 ml of dH₂O + 0.1% Triton X-100 using a Seward 80 stomacher and serially diluted in dH₂O + 0.1% Triton X-100 (1:5, 1:25 and 1:125 for lungs and 1:5, 1:25 and 1:75 for spleens), 30 μ l of each dilution was plated in duplicate onto 7H11 agar plates. CFUs were counted 3 weeks later. Due to limits of detection a log 2.2 value was attributed to spleen plates where no colonies were observed. Similarly, log 1.9 was attributed to lung plates where no colonies were observed.

Additional Methods

Cloning of Ag85B as control protein for expression in *N. benthamiana*, Gateway cloning of TB-PIGS into mammalian expression vector, Western blotting to determine structure of PIGS, expression and detection of PIGS in CHO cells, flow cytometry methodology and maintenance of cell lines are described in supplementary material.

Acknowledgments

The authors gratefully acknowledge the European Research Council Future-Pharma Advanced grant, the EC H2020 EMI-TB grant and the Hotung Foundation for funding this research project.

We would like to thank staff in the BRF at St. George's for help with the *in vivo* studies, Prof. Herta Steinkellner, BOKU Vienna for provision of glyco-engineered *N. benthamiana*, Dr. Anne Rawkins, PHE Porton Down for providing BCG-Pasteur and Prof. Rainer Fischer, Fraunhofer Institute Aachen, for provision of pTRAk.2 vector.

Author Contributions

Gina R. Webster designed and performed research, analyzed data and wrote the paper. Craig van Dolleweerd and Audrey Teh designed research and analyzed data. Thais Guerra & Miyoung Kim aided with research. Szymon Stelter designed and aided SPR experiments. Sven Hofmann designed and aided SEC experiments. Gil Reynolds Diogo performed cardiac punctures, category III laboratory work and aided with animal experiments. Mathew Paul

performed category III laboratory work and aided with animal experiments. Alastair Copland designed the polyfunctional assay and performed category III laboratory work. Peter Hart performed category III laboratory work. Rajko Reljic aided in design and analysis of animal work and performed category III laboratory work. Julian K-C Ma designed research, analyzed data and wrote the paper.

Competing financial interests: the authors declare no competing financial interests.

Figure Legends

Figure 1: Design of TB-PIGS. (A) Linearized map of gene sequence for TB-PIGS. The main structural elements are indicated for both human and murine immunoglobulin-based scaffolds with the amino acid substitutions highlighted. **(B) Diagrammatic representation of TB-PIGS ‘monomer’.** Expected size of ~120K, not including any glycosylation. Diagram is not to scale; Ag85B portion is ~30K and truncated γ -chain is ~30K. **(C) Diagrammatic representation of TB-PIGS ‘polymer’.** Expected size of ~720K for hexamer, not including any glycosylation. There are potentially 6 glycosylation sites in one single-chain; 4 within Ag85B nucleotide sequence and 2 within truncated γ -chain. However, only two sites are predicted to be accessible for glycosylation to occur when protein is folded into 3D structure.

Figure 2: Characterisation of assembly and size of TB-PIGS. In all cases 4-12% Bis-Tris gels with MOPS buffer were used. ~5-10 μ g of protein was loaded per well.

(A) SDS-PAGE of purified human and murine TB-PIGS detected by Coomassie blue staining. Human IgG1, IgM and murine IgG2a are shown as controls. Panels i-ii are under non-reducing conditions, panels iii-iv are under reducing conditions.

(B) Western blotting –Panels i and iv show detection using α -Ag85B antiserum under non-reducing and reducing conditions respectively. Positive control is plant expressed Ag85B, negative control is an extract from a wild type *N. benthamiana* plant (W.T.). Samples are purified human (H) and murine (M) TB-PIGS; Panels ii and v show detection using α -human IgG1 antiserum under non-reducing and reducing conditions respectively. Positive control is commercial purified human IgG1; Panels iii and vi show detection using α -murine IgG2a antiserum under non-reducing and reducing conditions respectively. Positive control is commercial purified murine IgG2a; **P** indicates polymer; **M** indicates monomer and **S** indicates single-chain size of TB-PIGS. Molecular weight markers in K are indicated to the left of each gel.

(C) Native PAGE of TB-PIGS. A 3-12% bis-tris gel in dark blue cathode and anode buffer was used. Molecular size markers are indicated on right-hand side of gel. **P** ~ polymer size; **D** ~ dimer size and **M** ~ monomer size.

(D) Size exclusion chromatography-ultraviolet spectrophotometry (SEC-UV). Bio Rad Standards include (from right to left) 1.35 kDa Vitamin B12 (0.5 mg), 17 kDa Horse Myoglobin (2.5 mg), 44 kDa Chicken Ovalbumin (5 mg), 158 kDa Bovine γ -globulin (5 mg) and 670 kDa Bovine Thyroglobulin (5 mg). A total volume of 20 μ l was run for each sample. Human TB-PIGS: Δ XF was run at a sample concentration of 100 μ g/ml and yielded four peaks. Murine TB-PIGS: Δ XF was run at a concentration of 100 μ g/ml and yielded 2 peaks. Black numbers indicate the retention time of each peak.

Figure 3: Functional characterisation of TB-PIGS *in vitro*. (A) **Complement-binding ELISA.** ELISA plates were coated with 5 µg/ml human C1q. Positive control was ICMS (TBG65 and ACR). α -Mse γ 1-HRP was used as detection antibody. Human TB-PIGS and HuIgG1 control were detected using α -Hu γ 1 antiserum. Murine TB-PIGS and MseIgG2a control were detected using α -Mse γ 2a antiserum. (B) **SPR analysis of avidity interaction between glyco-engineered human TB-PIGS and human CD32a.** 1 µM of human TB-PIGS: Δ XF or HuIgG1 were passed over the His-captured human CD32a. (C) **Avidity analysis between glyco-engineered human TB-PIGS and human CD16a.** 540 nM of human TB-PIGS: Δ XF or HuIgG1 were passed over the His-captured human CD16a.

Figure 4: Confocal microscopy of THP1 cells with human TB-PIGS: Δ XF. Nuclei were stained with DAPI (blue), Ag85B was detected with rabbit α -Ag85B antiserum and goat α -rabbit Ig-Alexa Fluor 488 (green), HuIgG γ -chain was detected with sheep α -HuIgG1 antiserum and donkey α -sheep IgG-Alexa Fluor 633 (magenta). Superimposed images are shown (far right). In descending order, samples incubated with cells were; media only, isotype control (rabbit IgG-Alexa Fluor 488), HuIgG1 and human TB-PIGS: Δ XF.

Figure 5: *In vivo* study of murine TB-PIGS in BALB/c mice. (A) **Ag85B-specific-IgG responses.** Three weeks after last immunisation, three mice were culled per group and serum was obtained. ELISA plates were coated with Ag85B and mean (\pm SEM) are depicted for total Ag85b specific IgG responses (top), IgG1 specific response (middle) and IgG2a specific response (bottom). (B) **Polyfunctional CD4⁺ T-cell responses.** Splenocytes were stimulated with Ag85B and stained for four different cytokines (IFN- γ , IL-2, IL-17 and TNF- α). Analysis of the percentage of positive CD4⁺ T-cells for each cytokine alone and in

combination was determined and represented in pie-charts (top). The total percentage of cells out of the events counted (before gating) are listed under the pie charts. The frequency of the antigen-specific cytokine producing cells are indicated in the graph (below). (C) **Polyfunctional CD8⁺ T-cell responses.** As for (B) but for CD8⁺ cells. (D) ***M. tuberculosis* challenge and tissue colonisation.** Extracts of lungs (left) and spleens (right) were plated out at three different dilutions per mouse, incubated for three weeks and the number of mycobacterial colonies counted. Mean (\pm SEM) are plotted per group. A one-way ANOVA with Tukey's multiple comparisons test was performed to determine statistical significance.

Figure 6: *In vivo* study of human TB-PIGS in transgenic CD64 mice. (A) α -Ag85B-specific-IgG responses. Three weeks after last immunisation, three mice were culled per group and serum was obtained. ELISA plates were coated with Ag85B and mean (\pm SEM) are depicted for total IgG responses (top), IgG1 specific response (middle) and IgG2a specific response (bottom). (B) **Cytokine response.** Splenocytes were stimulated with Ag85B and levels of four different cytokines, in the supernatant, were determined by ELISA. A one-way ANOVA with Tukey's multiple comparisons test was performed to determine statistical significance; * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ and **** $p < 0.0001$.

Figure 7: *In vivo* study of human TB-PIGS in transgenic CD64 mice. (A) Polyfunctional CD4⁺ T-cell responses. Splenocytes were stimulated with Ag85B and stained for four different cytokines (IFN- γ , IL-2, IL-17 and TNF- α). Analysis of the percentage of positive CD4⁺ T-cells for each cytokine alone and in combination was determined and represented in pie-charts (top). The total percentage of cells out of the events counted (before gating) are listed under the pie charts. The frequency of the antigen-specific cytokine producing cells are

indicated in the graph (below). **(B) Polyfunctional CD8⁺ T-cell responses.** As for (A) but for CD8⁺ cells. **(C) Protective efficacy of human TB-PIGS against challenge with *M. tuberculosis*.** Extracts of lungs (left) and spleens (right) were plated out at three different dilutions per mouse, incubated for three weeks and then the number of mycobacterial colonies counted. Mean (\pm SEM) are plotted per group. A one-way ANOVA with Tukey's multiple comparisons test was performed to determine statistical significance. For simplification purposes, not all statistical significance has been indicated; * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ and **** $p < 0.0001$.

Supplementary Figure & Table Legends

S1: SDS-PAGE of purified murine and human PIGS (without antigen) produced from CHO cells, under non-reducing and reducing conditions. A 4-12% Bis-Tris gel in MOPS buffer was used and ~ 20 μg of protein was loaded in all cases. Lane 1 was *N. benthamiana* produced human PIGS, lane 2 was *N. benthamiana* produced murine PIGS, lane 3 was CHO cell produced human PIGS under non-reducing conditions, lane 4 was CHO cell produced murine PIGS under non-reducing conditions, lane 5 was CHO cell produced human PIGS under reducing conditions and lane 6 was CHO cell produced murine PIGS under reducing conditions.

S2: SPR kinetics and affinity analysis of TB-PIGS binding to Fc γ Rs. (A) Human TB-PIGS with human CD64. Eight different concentrations (240 nM down to 1.875 nM, 1:2 dilutions) of CD64 were passed over the protein A-captured antibodies. 1:1 binding interaction model (black lines) was used for HuIgG1 and human TB-PIGS: Δ XF, whereas 'Heterogeneous antibody with aglycosylation' model^{*1} (black lines) was used for wild-type

N. benthamiana produced human TB-PIGS and the curves were fitted to the experimental data (coloured lines). **(B) Murine TB-PIGS with mouse CD64.** Eight different concentrations (240 nM down to 1.875 nM, 1:2 dilutions) of CD64 were passed over the protein A-captured antibodies/PIGS. 1:1 binding interaction model (black lines) was used for all samples and the curves were fitted to the experimental data (coloured lines). **(C) Steady-state analysis of human TB-PIGS with human CD32a.** Six different concentrations (4000 nM to 125 nM in 2-fold dilutions) of CD32a were passed over the protein A-captured antibodies/PIGS. Affinity values were derived from steady states (using steady-state affinity analysis available in the evaluation software). **(D) Human TB-PIGS with human CD16a.** Five-six different concentrations of CD16a were passed over the protein A-captured antibodies/PIGS. Two-state binding interaction model (black lines) was used for all samples and the curves were fitted to the experimental data (coloured lines).

*¹This model was created by Stelter *et al.* (manuscript in preparation) and was created based on heterogeneous ligand model, which calculates two separate sets of kinetic and affinity values. One set is defined and corresponds to kinetic/affinity values for aglycosylated heavy chains (as determined for a plant-produced aglycosylated human IgG1) and the second set calculates kinetic/affinity values for the glycosylated portion.

S3: Flow cytometric analysis of human TB-PIGS:ΔXF with THP1 cells. This data is representative of two repeats. Pseudocolour plots for each sample are shown with percentage values for 'negative' population (left of each plot) and percentage values for 'positive' FITC population (right of each plot). Half-offset histogram for FITC fluorescence for each sample with their corresponding mean fluorescence intensity (MFI) values given on the right. THP1 cells were incubated with **(A)** isotype FITC labelled MseIgG2a; **(B)** FITC labelled α-

HuIgG γ_1 ; (C) HuIgG1 and then FITC labelled α -HuIgG γ_1 ; (D) heat aggregated HuIgG1 and then FITC labelled α -HuIgG γ_1 ; (E) human TB-PIGS: Δ XF and then FITC labelled α -HuIgG γ_1 .

S4: Confocal microscopy of J774 cells with murine TB-PIGS: Δ XF. Nuclei were stained with DAPI (blue), actin filaments/cytoplasm stained with phalloidin (green; left), MseIgG γ_{2a} stained with AlexaFluor633 (pink; right) and merged images (far right). In descending order, samples incubated with cells were; media only, secondary antibody only, MseIgG2a and murine TB-PIGS: Δ XF.

S5: Flow cytometric analysis of murine TB-PIGS: Δ XF with J774 cells. This data is representative of two repeats. Pseudocolour plots for each sample are shown with percentage values for 'negative' population (left of each plot) and percentage values for 'positive' FITC population (right of each plot). Half-offset histogram for FITC fluorescence for each sample with their corresponding mean fluorescence intensity (MFI) values given on the right. J774 cells were incubated with (A) isotype FITC labelled rat IgG1; (B) FITC labelled α -MseIgG γ_{2a} ; (C) MseIgG2a and then FITC labelled α -MseIgG γ_{2a} ; (D) heat aggregated MseIgG2a and then FITC labelled α -MseIgG γ_{2a} ; (E) murine TB-PIGS: Δ XF and then FITC labelled α -MseIgG γ_{2a} .

Table S1: Yields for TB-PIGS produced in plants in mg of protein per kg of leaf mass.

The yield of polymers, dimers and monomers was calculated based on the total yield of protein (mg/kg) and the percentages of each species calculated by native PAGE (NP) or size exclusion chromatography (SEC).

Table S2: Kinetic and Affinity values of TB-PIGS with FcγRs. Two sets of values were generated for wild-type plant produced TB-PIGS with CD64 – one for aglycosylated TB-PIGS and one for glycosylated TB-PIGS present within the sample.

References

- Arias-bouda LMP, Kuijper S, Werf A Van Der, Nguyen LN, Jansen HM, Kolk AHJ. 2003. Changes in Avidity and Level of Immunoglobulin G Antibodies to Mycobacterium tuberculosis in Sera of Patients Undergoing Treatment for Pulmonary Tuberculosis. *Clinical and Diagnostic Laboratory Immunology* 10: 702–709.
- Babaki KZ, Soleimanpour MS, Rezaee SA. 2017. Antigen 85 complex as a powerful Mycobacterium tuberculosis immunogene: Biology, immune-pathogenicity, applications in diagnosis, and vaccine design. *Microb Pathog* 112: 20–29.
- Bruhns P. 2015. Properties of mouse and human IgG receptors and their contribution to disease models. *Blood* 119: 5640–5650.
- Cabanes-Macheteau M, Fitchette-Lainé AC, Loutelier-Bourhis C, Lange C, Vine ND, Ma JK, Lerouge P, Faye L. 1999. N-Glycosylation of a mouse IgG expressed in transgenic tobacco plants. *Glycobiology* 9: 365–372.
- Cayabyab MJ, Macovei L, Campos-Neto A, Izzo A, State C. 2012. Current and novel approaches to vaccine development against tuberculosis. *Frontiers in cellular and infection microbiology* 2: 154.
- Chargelegue D, Drake PMW, Obregon P, Prada A, Fairweather N, Ma JK. 2005. Highly

Immunogenic and Protective Recombinant Vaccine Candidate Expressed in Transgenic Plants. 73: 5915–5922.

Chen Q, Davis KR. 2016. The potential of plants as a system for the development and production of human biologics. *F1000Research* 5: 912.

Dedieu L, Floss DM, Mockey M, Zanello G, Brosson D, Diogon M, Frutos R, Bruel T, Rodrigues V, Garzon E, Chevaleyre C, Berri M, Salmon H, Conrad U. 2010. Expression and immunogenicity of the mycobacterial Ag85B/ESAT-6 antigens produced in transgenic plants by elastin-like peptide fusion strategy. *Journal of Biomedicine and Biotechnology* 2010:

Diebolder CA, Beurskens FJ, de Jong RN, Koning RI, Strumane K, Lindorfer MA, Voorhorst M, Ugurlar D, Rosati S, Heck AJ, van de Winkel JG, Wilson IA, Koster AJ, Taylor RP, Sapphire EO, Burton DR, Schuurman J, Gros P, Parren PW. 2014. Complement is activated by IgG hexamers assembled at the cell surface. *Science* 343: 1260–3.

van Dolleweerd CJ, Teh AY-H, Banyard AC, Both L, Lotter-Stark HCT, Tsekoa T, Phahladira B, Shumba W, Chakauya E, Sabeta CT, Gruber C, Fooks AR, Chikwamba RK, Ma JK-C. 2014. Engineering, Expression in Transgenic Plants and Characterisation of E559, a Rabies Virus-Neutralising Monoclonal Antibody. *The Journal of Infectious Diseases* 210: 200-208.

Ferrara C, Grau S, Jäger C, Sondermann P, Brünker P, Waldhauer I, Hennig M, Ruf A, Rufer AC, Stihle M, Umaña P, Benz J. 2011. Unique carbohydrate–carbohydrate interactions are required for high affinity binding between FcγRIII and antibodies lacking core fucose. *Proceedings of the National Academy of Sciences* 108: 12669–12674.

Gupta N, Arthos J, Khazanie P, Steenbeke TD, Censoplano NM, Chung EA, Cruz CC,

Chaikin MA, Daucher M, Kottlilil S, Mavilio D, Schuck P, Sun PD, Rabin RL, Radaev S, Van Ryk D, Cicala C, Fauci AS. 2005. Targeted lysis of HIV-infected cells by natural killer cells armed and triggered by a recombinant immunoglobulin fusion protein: implications for immunotherapy. *Virology* 332: 491–7.

Hayes JM, Frostell A, Cosgrave EFJ, Struwe WB, Potter O, Davey GP, Karlsson R, Anneren C, Rudd PM. 2014. Fc gamma receptor glycosylation modulates the binding of IgG glycoforms: A requirement for stable antibody interactions. *Journal of Proteome Research* 13: 5471–5485.

Hiatt A, Bohorova N, Bohorov O, Goodman C, Kim D, Pauly MH, Velasco J, Whaley KJ, Piedra PA, Gilbert BE, Zeitlin L. 2014. Glycan variants of a respiratory syncytial virus antibody with enhanced effector function and in vivo efficacy. *P.N.A.S.* 111: 5992–5997.

Hiatt A, Cafferkey R, Bowdish K. 1989. Production of antibodies in transgenic plants. *Nature* 342: 76–78.

Hodoniczky J, Yuan ZZ, James DC. 2005. Control of recombinant monoclonal antibody effector functions by Fc N-glycan remodeling in vitro. *Biotechnology Progress* 21: 1644–1652.

Horwitz MA, Harth G, Dillon BJ, Maslesa-Galic' S. 2000. Recombinant bacillus calmette-guerin (BCG) vaccines expressing the Mycobacterium tuberculosis 30-kDa major secretory protein induce greater protective immunity against tuberculosis than conventional BCG vaccines in a highly susceptible animal model. *Proceedings of the National Academy of Sciences of the United States of America* 97: 13853–8.

Horwitz MA, Lee BW, Dillon BJ, Harth G. 1995. Protective immunity against tuberculosis

induced by vaccination with major extracellular proteins of *Mycobacterium tuberculosis*. Proceedings of the National Academy of Sciences of the United States of America 92: 1530–4.

Jacobsen M, Repsilber D, Gutschmidt A, Neher A, Feldmann K, Mollenkopf HJ, Ziegler A, Kaufmann SHE. 2007. Candidate biomarkers for discrimination between infection and disease caused by *Mycobacterium tuberculosis*. *Journal of Molecular Medicine* 85: 613–621.

Karp CL, Wilson CB, Stuart LM. 2015. Tuberculosis vaccines: Barriers and prospects on the quest for a transformative tool. *Immunological Reviews* 264: 363–381.

Kim M, van Dolleweerd C, Copland A, Paul MJ, Hofmann S, Webster GR, Julik E, Ceballos-Olvera I, Reyes-del Valle J, Yang M-S, Jang Y-S, Reljic R, Ma JK-C. 2017. Molecular engineering and plant expression of an immunoglobulin heavy chain scaffold for delivery of a dengue vaccine candidate. *Plant biotechnology journal* 15: 1590–1601.

Kim M, Copland A, Nayak K, Chandele A, Ahmed M, Zhang Q, Diogo G, Paul M, Hofmann S, Yang M, Jang Y, Ma JK-C, Reljic R. 2018. Plant-expressed Fc-fusion protein tetravalent dengue vaccine with inherent adjuvant properties. *Plant Biotechnol Journal*.

Kim M, Reljic R, Kilbourne J, Ceballos-olvera I, Yang M, Valle JR, Mason HS. 2015. Novel vaccination approach for dengue infection based on recombinant immune complex universal platform. *Vaccine* 33: 1830–1838.

Klaassen RJ, Goldschmeding R, Tetteroo PA, Von dem Borne AE. 1988. The Fc valency of an immune complex is the decisive factor for binding to low-affinity Fc gamma receptors. *European journal of immunology* 18: 1373–7.

Laux da Costa L, Delcroix M, Dalla Costa ER, Prestes I V, Milano M, Francis SS, Unis G,

Silva DR, Riley LW, Rossetti MLR. 2015. A real-time PCR signature to discriminate between tuberculosis and other pulmonary diseases. *Tuberculosis* 95: 421–425.

Lux A, Yu X, Scanlan CN, Nimmerjahn F. 2013. Impact of immune complex size and glycosylation on IgG binding to human FcγRs. *Journal of immunology (Baltimore, Md. : 1950)* 190: 4315–23.

Ma J, Hiatt A, Hein M, Vine N, Wang F, Stabila P, van Dolleweerd C, Mostov K, Lehner T. 1995. Generation and assembly of secretory antibodies in plants. *Science* 268: 716–9.

Maertzdorf J, Ota M, Repsilber D, Mollenkopf HJ, Weiner J, Hill PC, Kaufmann SHE. 2011. Functional correlations of pathogenesis-driven gene expression signatures in tuberculosis. *PLoS ONE* 6: 1–8.

Maertzdorf J, Repsilber D, Parida SK, Stanley K, Roberts T, Black G, Walzl G, Kaufmann SHE. 2011. Human gene expression profiles of susceptibility and resistance in tuberculosis. *Genes and Immunity* 12: 15–22.

Mechetina L V., Najakshin AM, Alabyev BY, Chikaev NA, Taranin A V. 2002. Identification of CD16-2, a novel mouse receptor homologous to CD16/FcγRIII. *Immunogenetics* 54: 463–468.

Mekhaieel DNA, Czajkowsky DM, Andersen JT, Shi J, El-Faham M, Doenhoff M, McIntosh RS, Sandlie I, He J, Hu J, Shao Z, Pleass RJ. 2011. Polymeric human Fc-fusion proteins with modified effector functions. *Scientific reports* 1: 124.

Mimura Y, Sondermann P, Ghirlando R, Lund J, Young SP, Goodall M, Jefferis R. 2001. Role of Oligosaccharide Residues of IgG1-Fc in FcγRIIb Binding. *Journal of Biological Chemistry* 276: 45539–45547.

Nimmerjahn F, Bruhns P, Horiuchi K, Ravetch J V. 2005. FcγRIV: A novel FcR with distinct IgG subclass specificity. *Immunity* 23: 41–51.

Pepponi I, Diogo GR, Stylianou E, van Dolleweerd CJ, Drake PMW, Paul MJ, Sibley L, Ma JK-C, Reljic R. 2014. Plant-derived recombinant immune complexes as self-adjuvanting TB immunogens for mucosal boosting of BCG. *Plant biotechnology journal* 1–11.

Perley CC, Frahm M, Click EM, Dobos KM, Ferrari G, Stout JE, Frothingham R. 2014. The human antibody response to the surface of *Mycobacterium tuberculosis*. *PloS one* 9: e98938.

Phoolcharoen W, Bhoo SH, Lai H, Ma J, Arntzen CJ, Chen Q, Mason HS. 2011. Expression of an immunogenic Ebola immune complex in *Nicotiana benthamiana*. *Plant biotechnology journal* 9: 807–16.

Platzer B, Stout M, Fiebiger E. 2014. Antigen cross-presentation of immune complexes. *5: 1–10*.

du Plessis WJ, Walzl G, Loxton AG. 2016. B cells as multi-functional players during *Mycobacterium tuberculosis* infection and disease. *Tuberculosis* 97: 118–125.

Randall TD, Brewer JW, Corley RB. 1992. Direct evidence that J-chain regulates the polymeric structure of IgM in antibody-secreting B-cells. *The Journal of Biological Chemistry* 267: 18002-18007.

Reed SG, Orr MT, Fox CB. 2013. Key roles of adjuvants in modern vaccines. *Nat Med* 19: 1597–608.

Sack M, Paetz A, Kunert R, Bomble M, Hesse F, Stiegler G, Fischer R, Katinger H, Stoeger E, Rademacher T. 2007. Functional analysis of the broadly neutralizing human anti-

HIV-1 antibody 2F5 produced in transgenic BY-2 suspension cultures. *FASEB J* 21: 1655-64.

Scriba TJ, Coussens AK, Fletcher HA. 2017. Human Immunology of Tuberculosis. *Microbiology spectrum* 5: 1–23.

Shields RL, Lai J, Keck R, O'Connell LY, Hong K, Gloria Meng Y, Weikert SHA, Presta LG. 2002. Lack of fucose on human IgG1 N-linked oligosaccharide improves binding to human FcγRIII and antibody-dependent cellular toxicity. *Journal of Biological Chemistry* 277: 26733–26740.

Shields RL, Namenuk AK, Hong K, Meng YG, Rae J, Briggs J, Xie D, Lai J, Stadlen A, Li B, Fox JA, Presta LG. 2001. High Resolution Mapping of the Binding Site on Human IgG1 for FcγRI, FcγRII, FcγRIII, and FcRn and Design of IgG1 Variants with Improved Binding to the FcγR. *Journal of Biological Chemistry* 276: 6591–6604.

Smith RI, Morrison SL. 1994. Recombinant polymeric IgG: an approach to engineering more potent antibodies. *Biotechnology (NY)* 12: 683–8.

Smith RI, Coloma MJ, Morrison S. 1995. Addition of a mu-tailpiece to IgG results in polymeric antibodies with enhanced effector functions including complement-mediated cytotoxicity by IgG4. *The Journal of Immunology* 2226–2236.

Sorensen V, Sundvold V, Michaelsen TE, Sandlie I. 1999. Polymerization of IgA and IgM: Roles of Cys³⁰⁹/Cys⁴¹⁴ and the Secretory Tailpiece. *The Journal of Immunology* 162: 3448-3455.

Strasser R. 2016. Plant protein glycosylation. *Glycobiology* 26: 926–939.

Strasser R, Stadlmann J, Schähs M, Stiegler G, Quendler H, Mach L, Glössl J, Weterings K,

Pabst M, Steinkellner H. 2008. Generation of glyco-engineered *Nicotiana benthamiana* for the production of monoclonal antibodies with a homogeneous human-like N-glycan structure. *Plant biotechnology journal* 6: 392–402.

Teh AYH, Maresch D, Klein K, Ma JKC. 2014. Characterization of VRC01, a potent and broadly neutralizing anti-HIV mAb, produced in transiently and stably transformed tobacco. *Plant Biotechnology Journal* 12: 300–311.

Teye K, Hashimoto K, Numata S, Ohta K, Haftek M, Hashimoto T. 2017. Multimerization is required for antigen binding activity of an engineered IgM/IgG chimeric antibody recognizing a skin-related antigen. *Scientific Reports* 7:8212-8223.

Wen YM, Mu L, Shi Y. 2016. Immunoregulatory functions of immune complexes in vaccine and therapy. *EMBO Molecular Medicine* 8: 1120–1133.

White DM, Jensen MA, Shi X, Qu Zx, Arnason BG. 2001. Design and expression of polymeric immunoglobulin fusion proteins: a strategy for targeting low-affinity Fcγ receptors. *Protein expression and purification* 21: 446–55.

Wiker HG, Harboe M. 1992. The antigen 85 complex: a major secretion product of *Mycobacterium tuberculosis*. *Microbiological reviews* 56: 648–61.

Yoo EM, Coloma MJ, Trinh KR, Nguyen TQ, Vuong L-UC, Morrison SL, Chintalacheruvu KR. 1999. Structural Requirements for Polymeric Immunoglobulin Assembly and Association with J Chain. *The Journal of Biological Chemistry* 274: 33771-33777.

Zhou Q, Shankara S, Roy A, Qiu H, Estes S, McVie-Wylie A, Culum-Merdek K, Park A, Pan C, Edmunds T. 2008. Development of A Simple and Rapid Method for Producing Non-Fucosylated Oligomannose Containing Antibodies With Increased Effector Function. *Biotechnology and Bioengineering* 99: 652–665.









