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Anxiogenic CO₂ Stimulus Elicits Exacerbated Hot Flash-like Responses in a Rat Menopause Model and Hot Flashes in Menopausal Women

Lauren M. Federici, B.S.^{1,2}, Sarah Dorsey Roth, B.S.N.⁵, Connie Krier, B.S.⁵, Stephanie D. Fitz³, Todd Skaar, Ph.D.⁴, Anantha Shekhar, M.D., Ph.D.^{2,3,6}, Janet S. Carpenter, Ph.D., RN, FAAN⁵, and Philip L. Johnson, Ph.D.^{1,2}

¹Department of Anatomy and Cell Biology, Indiana University School of Medicine, Indianapolis, IN USA

²Program in Medical Neurosciences, Paul and Carole Stark Neurosciences Research Institute, Indiana University School of Medicine, Indianapolis, IN USA

³Department of Psychiatry, Indiana University School of Medicine, Indianapolis, IN USA

⁴Division of Clinical Pharmacology, Indiana University School of Medicine, Indianapolis, IN USA

⁵Indiana University School of Nursing, Indianapolis, IN USA

⁶Indiana Clinical and Translational Sciences Institute; Indiana University School of Medicine, Indianapolis, IN USA

Abstract

Objective—Since longitudinal studies determined that anxiety is a strong risk factor for hot flashes, we hypothesized that an anxiogenic stimulus that signals air hunger (hypercapnic, normoxic gas) would trigger an exacerbated hot flash-associated increase in tail skin temperature (TST) in a rat ovariectomy (OVEX) model of surgical menopause and hot flashes in symptomatic menopausal women. We also assessed TST responses in OVEX serotonin transporter (SERT)^{+/-} rats that models a common polymorphism that is associated with increased climacteric symptoms in menopausal women and increases in anxiety traits.

Methods—OVEX and sham-OVEX rats (initial experiment) and wildtype and SERT^{+/-} OVEX rats (subsequent experiment) were exposed to a 5 min infusion of 20% CO₂ normoxic gas while measuring TST. Menopausal women were given brief 20% and 35% CO₂ challenges, and hot flashes were self-reported and objectively verified.

Results—Compared to controls, OVEX rats had exacerbated increases in TST, and SERT^{+/-} OVEX rats had prolonged TST increases following CO₂. Most women reported mild/moderate hot flashes after CO₂ challenges, and the hot flash severity to CO₂ was positively correlated with daily hot flash frequency.

Corresponding Author, Philip L. Johnson, Ph.D., Neuroscience Research Building, 320 W. 15th St., room 308, Indianapolis, IN USA
46202, philjohn@iupui.edu.

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Conclusions—The studies demonstrate that this anxiogenic stimulus is capable of inducing cutaneous vasomotor responses in OVEX rats, and eliciting hot flashes in menopausal women. In rats, the severity of the response was mediated by loss of ovarian function and increased anxiety traits (SERT^{+/-}), and, in women, by daily hot flash frequency. These findings may provide insights into anxiety related triggers and genetic risk factors for hot flashes in thermoneutral environments.

Keywords

hypercapnia; stress; anxiety; serotonin; thermoregulation; acidosis

Introduction

Hot flashes are intense heat sensations that occur primarily in the chest and face and affect the vast majority of women at midlife or following estrogen inhibition therapies or oophorectomy (1, 2). In the laboratory setting, the subjective hot flash sensation (as indicated with button press or self-report) correlates well with objectively measured, sympathetically-mediated increases in cutaneous vasodilation which pools 37°C blood to the skin. This in turn interacts with peripheral thermal sensors in skin to signal heat. Sweating sometimes also occurs during a hot flash, but this induces an evaporative cooling sensation. Under normal circumstances, both cutaneous vasodilation and sweating are effective means of dissipating heat to maintain core body temperature (T_c) in warmer environments. While environmental temperature is often thought to affect hot flash parameters (i.e., frequency, severity, duration), many studies have found weak or inconsequential relationships to ambient temperature (3-5), and in a self-reported survey of hot flash precipitants, temperature was identified less frequently than other stimuli (6). While exact figures are unavailable, these data suggest that many women may experience hot flashes in thermoneutral environments with no clear identifiable trigger. In these circumstances, the collective heat dissipation mechanisms that occur during a hot flash could lead to decreases in T_c which does occur and could explain chills that also occur in menopausal women [(7) and see review (8)].

While there are many variables that influence hot flashes, recent findings from large longitudinal studies have identified the presence of anxiety, early life stress, and lower socioeconomic status as key risk factors for more severe and problematic hot flashes (9, 10). Notably, anxiety has been identified as the best predictor of hot flashes, even when controlling for typical confounds, and women with greater anxiety levels experience more severe hot flashes (10, 11). Additionally, many women report that stressful stimuli in everyday life precipitate hot flashes (6), and stressful stimuli in a laboratory setting have been shown to increase objectively measured hot flashes (12). Taken together, there is evidence that stressful events may provoke hot flashes, and that pre-existing anxiety states or traits may contribute to increased symptoms. However, the mechanisms by which anxiety and stress-related factors contribute to hot flashes remain poorly understood.

One contributing factor to the poor understanding of the mechanisms mediating hot flashes is a scarcity of appropriate, validated animal modeling (13). Previous models such as the morphine withdrawal model (14) elicit robust cutaneous vasomotor responses in the tail, but

lack an obvious relevance to causes and triggers of hot flashes (15, 16). Consequently, the use of models for mechanistic investigation into the central and peripheral neural circuitry and associated pathways mediating hot flashes has suffered. Recently, we found profound decreases in T_c in rats following an acute 20% carbon dioxide (CO_2) exposure, which produces anxiety-associated behavior (social and open field avoidance, fear-associated freezing, and avoidance of a 15-20% CO_2 filled chamber in a two chamber place preference test) and sympathetic associated cardioexcitation (17-21). In human studies, 20% CO_2 inhalation induces anxiety symptoms (22, 23), and in another study the participants reported anxiety that was also associated with strong heat sensations following a single inhalation of 35% CO_2 (24). However, it was unknown if this would have relevance to menopause-associated symptoms.

Therefore, we designed a series of studies to determine if CO_2 exposure represented a novel method of provoking hot flashes. Thus, in a rodent model, we first hypothesized that 20% CO_2 -induced decreases in T_c were due to cutaneous vasomotor responses in the tail which would increase the tail skin temperature (TST) to dissipate heat in a thermoneutral environment. We further posited that the TST response would be exacerbated in ovariectomized (OVEX) rats, modeling surgical oophorectomy-induced menopause, which is linked to more severe hot flashes (2, 25). Second, in a small, proof of concept study, we hypothesized that 20% CO_2 and 35% CO_2 inhalation tests would elicit a subjective hot flash in symptomatic menopausal women. Finally, recent work has begun to elucidate some of the genetic contributions to increased or problematic hot flashes. The short ('s') version of the serotonin transporter (SERT) gene, whose protein product mediates the termination of serotonin signaling, leads to reduced transcriptional efficiency and less protein, and is linked to anxiety-associated traits in humans (26-29) as well as increased climacteric symptoms in menopausal women (30). Therefore, our last hypothesis was that OVEX rats with a heterozygous null mutation of the SERT gene ($SERT^{+/-}$) which leads to similar reduced transcriptional efficiency, would have exacerbated and/or prolonged hot flashes (increased TST) in response to CO_2 compared to wildtype (WT) OVEX controls.

Materials and Methods

Animal Experiments

Animals and Housing Conditions—Adult female (225-250 g) Sprague-Dawley rats were purchased from Harlan Laboratories (Indianapolis, IN USA). Rats with heterozygous expression of the serotonin transporter (SERT) were purchased from Charles River Laboratories (Wilmington, MA USA) and bred in-house on a Wistar background from Wistar rats purchased from Harlan Laboratories (Indianapolis, IN). These rats possess a null mutation of the SERT generated by *N*-methyl-*N*-nitrosurea-mutation which results in an approximately 50% reduction in expression of SERT mRNA within the brain as described previously (31). New Wistars from Harlan (Indianapolis) were bred into the colony every three generations to prevent genetic drift. Female Wistar rats (225-250 g) were purchased (Harlan, Indianapolis) as wildtype controls and were housed in the same conditions as the other experimental rats (maintained at 22°C on a 12:12 light/dark cycle with lights on at 0700h) for at least 5 days prior to experimental procedures and had *ad libitum* access to food

and water. We have also confirmed that, compared to Wistar male wildtype controls, the male *SERT*^{-/+} rats have an approximately 50% reduction in *SERT* mRNA within a brain region densely populated with serotonergic neurons (i.e., 300µM coronal brain punches of dorsal and median raphe nuclei (unpublished observations). All procedures were approved by the Institutional Animal Care and Use Committee at Indiana University-Purdue University, Indianapolis, and in accordance with the NIH Guidelines for the Care and Use of Laboratory Animals (NIH publication no. 80-23) revised 1996.

Ovariectomy—Female rats were anesthetized under isoflurane (2-3% by volume in atmospheric air; MGX Research Machine, Vetamac, Rossville, IN USA; Praxair Inc., Indianapolis, IN USA) delivered through a nosecone. The skin was shaved and sterilized prior to incision; the ovaries were visualized and excised bilaterally. For sham-ovariectomized rats, the ovaries were visualized but not excised. Rats were sutured, given pain medication, and allowed 12 days of recovery prior to experimental manipulations.

CO₂ provocation of hot flash—Flow cages (12 in. width × 12 in. height × 24 in. length) were custom-built using Plexiglas®. When the lid of the cage was latched, gases could only enter the cage through an inlet connector (for the gas infusion) and could only exit the cage through an outlet connector. The gas flow into the cages was controlled using a 2-stage regulator (Praxair, Inc., Danbury, CT USA) at a pressure of 0.6 Bar. We previously validated the consistency of the rate of CO₂ delivery using state-of-the-art infrared CO₂ (ProCO₂) and electrochemical O₂ (ProO₂) sensors (32). Specifically, concentrations of O₂ remain at 21% throughout the gas infusion in the control and experimental cages [see (Johnson et al., 2005)]. The CO₂ concentration remains constant at < 1 % in the control cage during exposure of rats to atmospheric air (< 1% CO₂ / 21% O₂ / 79% N₂). Infusion of the premixed normoxic, hypercapnic gas (20% CO₂ / 21% O₂ / 59% N₂) results in a rapid increase in CO₂ concentration from < 1% CO₂ up to 20% CO₂ at the 5 min time point. After terminating gas infusion and opening the cages, the concentration of CO₂ rapidly decreases from 20% CO₂ to < 2.5% CO₂ during the following 5 min. Using a portable iSTAT gas analyzer (HESKA, Des Moines, IA USA) we have also determined that this hypercapnic, normoxic challenge leads to arterial pCO₂/pH levels of ~ 130mmHg/7.01 during the challenge that are back to normal physiological range (~50mmHg/7.37) within 2 min post challenge (unpublished data). Prior to placement in the apparatus, a thermistor (Omega Precision Fine Wire Thermocouples, Part #5SRTC-TT-K-30-36; Omega Engineering, Stamford, CT USA) was secured to the ventral side of the tail approximately 1 cm from the base using 3M Durapore® tape to measure tail skin temperature. The thermistor was attached to a T-type pod and connected to a Powerlab system running LabChart software (ADInstruments, Colorado Springs, CO USA) for continuous monitoring of temperature. After obtaining a stable baseline temperature in atmospheric air, rats were exposed to hypercapnic gas for 5 min, and then atmospheric air was infused again for 10 min following the challenge.

Atmospheric Air Control Experiment—To determine if the experimental environment alone caused a change in tail skin temperature, the above experiment was performed, but only atmospheric air was infused for the duration of the experiment. After obtaining a stable

baseline, the air was turned off and immediately turned back on to mimic the gas change during a CO₂ infusion. Rats recovered in atmospheric air for 10 min following the challenge.

Infrared Thermography—As an additional verification of temperature change, thermal images were acquired following hot flash provocations using a FLIR T440 thermal imaging camera (FLIR Systems, Boston, MA USA) with standard settings taken at a height of 1 m. Analysis of images was performed using FLIR Tools to standardize scaling between thermal images.

Human procedures

Sample—Six peri- or post-menopausal English-speaking women with at least 4 hot flashes per 24 hour day were recruited. Each participant gave written informed consent after one of the investigators explained the nature, purpose, and risks of the study. All participants were screened for medical and psychiatric history. To be eligible, all participants reported daily hot flashes (verified on 7-day diary), were afebrile, had a resting heart rate < 110 beats per minute, a respiratory rate of 8-18 breaths per minute, oxygen saturation above 94%, and were normotensive (BP < 160/100). Women were excluded if they had respiratory or cardiac disease (COPD, asthma, emphysema, hyperventilation syndrome, heart disease), hypertension, insulin-dependent diabetes, cancer, Axis I diagnosis (depression, panic disorder, generalized anxiety disorder), personal/familial history of cerebral aneurysm, were taking any medications for hot flashes, were substance abusers (including excessive smoking, i.e. >10 per day), were trained athletes, or were pregnant or breastfeeding. Participants were compensated \$50 at the end of the study.

Measures—Demographic information was elicited by the investigator using a standard questionnaire. Participants completed a checklist type form to provide age, race, ethnicity, education, menopausal status, and other information.

Hot flashes were measured as follows. Using a hot flash diary, women were asked to record the number and severity of their hot flashes for seven consecutive days. This information was used to verify eligibility. To evaluate hot flash severity during the study, participants completed a numeric rating scale (NRS). During the study visit, participants rated the severity of any reported hot flashes from 0 (not at all) to 10 (extremely) during the study visit, and all self-reported hot flashes were verified with objective sternal skin conductance monitoring (Biolog, UFI Serving Science, Morro Bay, CA USA).

Anxiety was rated using two self-report scales. Using a numeric rating scale (NRS) of 0 (not at all) to 10 (extremely), participants rated how anxious they felt at baseline, after the control room air inhalation, after the CO₂ challenge conditions, and at discharge. Higher scores on the NRS indicated greater anxiety. In addition, participants also completed the State version of the short form of the State-Trait Anxiety Inventory, a 10-item inventory, to assess anxiety “in the moment”. All items on the scale are positively worded (e.g., I feel calm) and each item is rated from 1 (not at all) to 4 (very much so) so that higher total scores indicate less state anxiety. The STAI was completed at baseline and immediately prior to discharge.

Procedures—Procedures were HIPAA compliant and approved by the Indiana University-Purdue University, Indianapolis Institutional Review Board prior to study implementation. Women were recruited through self-referral and emails to potentially eligible research registry members. For registry participants, once emails had been sent by the registry, the research team attempted to contact each woman by telephone. Self-referrals phoned or emailed the research team on their own. Once contact was made with a potential participant, trained personnel used a script to screen them for eligibility. If eligible, women were emailed the hot flash diary and instructed on how to complete it. Once diary responses were sent via phone, email, mail or secure fax, eligibility was confirmed (i.e., daily hot flashes met threshold) and study visits were scheduled at a mutually convenient time. Email or phone call reminders were done prior to study visits.

Each participant arrived at the university Clinical Research Center at a pre-determined time. Written informed consent and authorization to use health information was obtained. Trained research staff assessed baseline vital signs (heart rate, respiratory rate, blood pressure, and temperature), peripheral oxygen saturation levels, and height and weight, and women were connected to the hot flash monitor. Next, demographic information and anxiety scales (STAI, NRS) were completed. A non-rebreather mask was fitted for each participant. Through this mask, the participant inhaled normal room air for 10 minutes to become acclimated to the mask. The anxiety NRS was completed by the participant, followed by a 3-minute rest period. Next, the participant inhaled 20% CO₂ for 40 seconds via the mask. The hot flash NRS and the anxiety NRS were completed, followed by a 15-minute rest (the rest period was extended by 5 minutes if the participant had a hot flash), and then a vital-capacity, double-breath inhalation of 35% CO₂. A final anxiety NRS and STAI anxiety were completed by the participant; see Fig. 2a for a schematic of study procedures. As mentioned in the introduction, 20% and 35% CO₂ inhalation procedures have been utilized previously with healthy humans to induce and study anxiety symptoms (23) and reliably produce stable autonomic responses, including increases in skin conductance and heart rate, across multiple sessions (33). Here we utilized the NRS to measure anxiety symptoms at baseline, following mask and control air and immediately post CO₂ inhalation, but the NRS and STAI were also utilized to verify that there were no lingering anxiety effects 15-20 min post CO₂ inhalation. Previous work has demonstrated that there are no increases in severe anxiety at 6 and 12 months following 20% CO₂ challenges [compared to participants exposed to control conditions; (34)]. Vital signs were taken and the participant was discharged to home.

Statistical Analyses—For animal experiments, dependent variables for analyses of tail skin temperature (TST) were analyzed using a one-way ANOVA with repeated measures, using gas infusion as the between-subjects factor and time as a within-subjects factor. In the presence of significant main effects or main effect × time interactions, Fisher's Least Significant Difference (LSD) test was used for post hoc testing between-subjects comparisons. For the clinical component, an ANOVA over time was used to determine the effect of 20% CO₂ on NRS anxiety scores; Dunnett's post hoc test was used to determine specific time point differences. Pearson's correlation was used to determine relationships between hot flash frequency (daytime, nighttime, and average daily) and NRS anxiety scores or hot flash intensity following 20% CO₂ or 35% CO₂ challenges. The alpha level was set at

0.05 in all cases. All statistical analyses were carried out using SPSS 21.0 (SPSS Inc., Chicago, IL, USA), and all graphs were generated using SigmaPlot 12.0 (SPSS Inc.) and an illustration program (CorelDraw X5 for Windows, Viglen Ltd., Alperton, UK).

Results

Experiments 1-2: Ovariectomized rats exhibit exacerbated hot flash-associated increases in tail skin temperature in response to hypercapnic gas infusion while atmospheric air infusion does not change tail skin temperature

An ANOVA with ovex surgery and hypercapnic gas/CO₂ as main factors and time as a repeated measure revealed that hypercapnic gas infusion produced an exacerbated increase in tail skin temperature in ovex rats while atmospheric air caused no changes in temperature [Fig 1a-Representative thermal image; Fig. 1b- CO₂ tail skin temperature (n=15/group) overall CO₂ effect $F_{(1,38)}=45.9$, $p<0.0001$, overall ovex effect $F_{(1,38)}=5.3$, $p=0.027$, and overall CO₂ by time interaction $F_{(114,532)}=20.1$, $p<0.0001$ but no ovex by time interaction $F_{(14,532)}=0.23$, $p=0.999$ or ovex by CO₂ by time interaction $F_{(14,532)}=0.34$, $p=0.989$]. Fisher's Least Significant Difference Test was used for post hoc testing to determine specific time point differences between groups, and Dunnett's test was used to determine significance of specific time points against t-1.

Experiment 3: CO₂ challenges elicit hot flashes in symptomatic women

A total of 113 women were contacted (109 research registry and 4 self-referrals). Of these 113, 56 were successfully reached and completely screened for eligibility. Of the 56, 37 were not eligible and 7 were not interested, leaving 12 eligible and interested women. These 12 women were sent diaries but 5 were not eligible on diary return and 1 did not return her diary. Of the remaining 6 women, all completed the study visit.

Sample demographics were as follows for the five participants that successfully completed at least one CO₂ inhalation. The participants had a mean age of 49.8 (SD=2.77) and 17.00 years of education (SD=3.32). Four of five participants were non-Hispanic White and one was non-Hispanic Black, 4/5 were married (1 was other), 4/5 reported no difficulty paying for basics (1 reported some difficulty), and all worked full-time outside the home. Comorbidities included hypothyroidism (n=1), chronic interstitial cystitis (n=1), and non-insulin dependent diabetes (n=1). Total number of prescription medications taken per participant was 1.00 (SD=1.0, range 0 to 3).

The screening week hot flash diaries showed an average of 7.00 hot flashes during the daytime (SD=4.15, range 3.42 to 13.57), and 3.29 hot flashes during the nighttime (SD=0.96, range 2.29 to 4.86), for an average of 10.29 total hot flashes per 24-hours per participant (SD=4.24, range 5.86 to 16.43).

Five of six participants reported hot flashes after CO₂ challenge (see Fig. 2b). One participant had no confirmed inhalations during either CO₂ challenge and was removed from all analyses. Three of the four participants with confirmed inhalations of 20% CO₂ reported hot flashes within 5 min of the challenge that ranged from mild to moderate, and two of the three participants with confirmed inhalations of 35% CO₂ reported a hot flash within 5 min

of the challenge. Numbers below “NRS hot flashes post CO₂” respectively represent hot flash severity rating following 20% CO₂ (Fig. 2b column 9), and 35% CO₂ with the parenthetical number representing the mean NRS of confirmed inhalations; a “-” indicates no confirmed inhalation which was evidenced by removal of the mask and coughing, or confirmation that inhalations were not completed. Numbers below “NRS anxiety post CO₂” respectively represent 20% CO₂ and 35% CO₂ (Fig. 2b, column 10, a “-” indicates no confirmed inhalation). Average severity of CO₂-induced hot flashes was a 4 (range 3-5) after 20% CO₂, and 2.5 (range 1-4) after 35% CO₂. NRS ratings of anxiety approached significance following inhalation of 20% CO₂, when compared to NRS anxiety ratings post inhalation of air ($p=0.062$ with paired Wilcoxon post hoc test). However, there was a positive correlation between average total number of hot flashes per day and the mean hot flash severity following confirmed CO₂ inhalations, $r=0.921$, $p=0.026$, $n=5$ (Fig. 2c). There was a positive correlation between average total number of hot flashes per day and NRS scores for anxiety following verified CO₂ inhalation, $r=0.942$, $p=0.016$, $n=5$. (Fig. 2d). There was a positive correlation between NRS hot flash severity following confirmed CO₂ inhalation and NRS anxiety following confirmed inhalation, $r=0.877$, $p=0.049$, $n=5$ (Fig. 2e). Vital signs were in the normal range at the end of the study visit for all participants, and there were no lingering effects on anxiety or vital signs. STAI scores of 39 to 40 out of 40 possible (no anxiety) were measured in all participants at both baseline and end of the study visit.

Experiment 4: Ovariectomized SERT^{+/-} rats exhibit prolonged hot flash-associated increases in tail skin temperature in response to hypercapnic gas infusion

An ANOVA with genotype as the main factor and time as a repeated measure revealed that hypercapnic gas infusion produced a prolonged increase in tail skin temperature in ovex rats [Fig 3a- Representative thermal image; Fig. 3b- CO₂ tail skin temperature ($n=7,8$) overall *time effect* $F_{(13,392)}=47.8$, $p<0.0001$ and overall *genotype \times time interaction* $F_{(15,180)}=2.1$, $p=0.013$]. Fisher’s Least Significant Difference Test was used for post hoc testing to determine specific time point differences, and Dunnett’s test was used to determine significance of specific time points against t-1.

Discussion

These studies have demonstrated that an anxiogenic stimulus can precipitate an exacerbated hot flash-associated increase in TST responses in OVEX rats compared to sham-OVEX controls, which, to the best of our knowledge, is the first demonstration of threshold differences between OVEX and intact rats in response to CO₂. Following similar CO₂ inhalations, symptomatic menopausal women reported mild to moderate objectively verified hot flashes which approached significance for an increase in anxiety ratings following 20% CO₂ inhalation. The mean number of daily hot flashes was positively correlated with the severity of hot flashes and anxiety ratings post confirmed inhalations of CO₂. Our studies provide potential insights into triggers of hot flashes in thermoneutral environments and evidence that anxiety or anxiety-generating stimuli may have a causal relationship to hot flashes, not merely an associative one. Earlier clinical provocations used the drug yohimbine to facilitate hot flashes (35), which, at high doses, elicits strong anxiety (36, 37), and also

produces robust TST responses in OVEX rats (38), and thus yohimbine may represent a pharmacological model preclinically and clinically. A weakness of the present rodent studies was that only CO₂ was used as a stressor. However, we recently demonstrated that the GABAergic compound FG-7142 (a partial inverse benzodiazepine receptor agonist) which causes anxiety and flushing clinically (39) elicited rapid hot flash-associated increases in TST in OVEX rats but not sham control rats (40). Other studies have previously documented that stress-related stimuli, including mental arithmetic or emotionally salient films, can increase the rate of objective and subjective hot flashes (12) and self-reports implicate stressful stimuli (e.g. interpersonal family conflict or social stress) precipitate hot flashes in breast cancer patients (6). These data complement the emerging literature relating to the role of anxiety and stress-related factors in hot flashes and may help to start addressing a critical gap in knowledge of hot flash triggers.

In our preclinical rodent experiments, the results exhibit validity in three domains: 1) face validity, as the increase in TST is a rapid heat loss mechanism (cutaneous vasodilation) analogous to and what would be expected during a hot flash; 2) construct validity, as the TST change is exaggerated in ovariectomized rats, which is a model of surgical/oophorectomy-induced menopause in women (a condition with more severe hot flashes (2, 25)); and 3) predictive validity, as the same stimulus causes subjective hot flashes in menopausal women, and is exacerbated in ovariectomized SERT^{+/-} rats, which mimic a common human polymorphism that was recently linked to increased hot flash pathology (30).

This modeling approach is unique in that the provocation does not use pharmacological manipulations; rather, it triggers an endogenous challenge to homeostasis by provoking air hunger through CO₂ exposure. It is important to note that increasing concentrations of CO₂ in the blood combine with water to form carbonic acid to induce acidosis, and CO₂ also easily crosses the blood-brain barrier to interact with central chemoreceptive systems (41, 42). The 20%CO₂ exposure used here produces a brief drop in blood pH to ~7.1-7.2 (19), and also alters pH within anxiety brain circuits (21). Importantly, these studies suggest that disease states or conditions associated with respiratory or metabolic acidosis may be risk factors for hot flashes and may reduce the threshold for a hot flash to occur. Such conditions include chronic obstructive pulmonary disease [COPD; pH has been reported to be decreased to 7.35 during sleep (43), diabetic ketoacidosis [pH 7.2 with a range of 6.78-7.39 (44), and other factors that make breathing (and expiring adequate CO₂) difficult, such as in smokers, asthmatic persons, or obese persons. Indeed, obesity and smoking are well-established risk factors for hot flashes (45, 46). Additionally, stress induced hypoventilation or environments with poor ventilation and higher than normal levels of CO₂ could contribute to lowering the hot flash threshold by inducing dyspnea. Finally, the anxiety response to acidosis or other stressors may also be relevant triggers and not just a response to the hot flash. Changes in pH are revealing in light of other work that has demonstrated a key role for pH in regard to thermoregulation and anxiety. The amygdala, a key brain structure for fear processing and anxiety, and 20%CO₂ exposure used here produces acidosis in the amygdala, and local induction of acidosis induces robust fear associated freezing in mice (21). In persons with limbic encephalitis, which reversibly lesions the amygdala, emotional sweating is absent, yet thermoregulatory sweating was intact (47). Thus, emotional stimuli and

emotional circuits also appear to be relevant to hot flashes, and it is possible that under conditions of reduced pH or elevated CO₂ (which causes some peripheral vasodilation), anxiety is elicited and emotional sweating occurs. This anxiety, in combination with environmental conditions (i.e., slightly increased ambient temperature) may further exacerbate the heat dissipation response and trigger a hot flash.

Currently, the role of blood pH in hot flashes is largely unknown. Paced respiration (which could moderate blood pH) such as controlled breathing of 6-10 breaths per minute, is often recommended as a non-pharmacological treatment for reducing hot flashes, yet clinical trials have not demonstrated significantly greater efficacy than usual treatment or active comparators (48-50).

Here our challenges utilized systemic administration through either a mask or environmental exposure (the latter necessitating inhalation leading to systemic exposure). Dissection of the central and peripheral components of the CO₂-induced hot flashes and TST responses in rats is beyond the scope of this investigation, but the paradigms we used most likely triggered central mechanisms, as previous applications were used to elicit and study anxiety in humans and in rodents that are accompanied by sympathetic associated cardiovascular responses. However, CO₂ has also been shown to have local cutaneous vasodilatory action in both humans and rodents (51, 52). Thus, there are likely central and peripheral contributions to hot flashes (53).

In this case, evidence for central mechanisms comes from *ex vivo* assessments of cellular responses in rodents where 20%CO₂ exposure increases activity in anxiety and thermoregulatory associated brain circuitry such as the hypothalamus and brainstem (18, 32). Within the hypothalamus, there is substantial overlap between nuclei that respond to 20% CO₂ and expression patterns of estrogen receptors α and β (ER α , ER β). Specifically, the paraventricular nucleus responds strongly to hypercapnic gas, and also has very dense expression of both ER β (54) and an excitatory transmembrane estrogen receptor (e.g., GRP30) (55), whereas the more posterior hypothalamic regions are similarly responsive to CO₂ yet expresses ER α (54) and GRP30 (55). Noradrenergic and serotonergic neurons in the midbrain pons and medulla also exhibit hypercapnic responsivity (28). Hypothalamic and brainstem sites including serotonergic nuclei (dorsal raphe and raphe pallidus) and noradrenergic nuclei (locus ceruleus) were found to significantly increase in response to FG-7142 hot flash provocation as well (40). These latter sites may be especially relevant to hot flashes in women because both the noradrenergic and serotonergic systems are treatments for hot flashes with selective serotonin and/or norepinephrine reuptake inhibitors (56, 57). Taken together, these results provide additional validity that the neural mechanisms mediating cutaneous vasomotor responses induced by CO₂ may be relevant to spontaneous hot flashes.

Our rodent studies also contribute to an emerging literature concerning the *5-HTTLPR* on climacteric symptoms and anxiety trait vulnerability. Short form 's' carriers (*s/s* or *s/l* genotype) of the *5-HTTLPR* experience higher rates of psychiatric disorders later in life, including anxiety and depression (27-29, 58), which are two conditions that have been linked to increased hot flashes (10, 59). Analyses from the multiethnic Study of Women's

Heath Across the Nation Mental Health Study found that early life stress (childhood abuse or neglect) also predicted later vasomotor symptoms (9). Menopausal women with the short form ('s' carrier) of the *5-HTTLPR* also experience increased climacteric symptoms (30), which could be related to increased anxiety traits associated with this polymorphism. This animal model could contribute mechanistic insight into the interactions between early life events, genetic contributions, and later anxiety and physiological phenotypes, including hot flash-associated responses, which may ultimately lead to a better understanding of the role of serotonin and its complement of proteins to menopausal symptoms.

Indeed, this is an important research area in women's health at midlife, as drugs that target the serotonin transporter are among the most frequently used pharmacotherapies for vasomotor symptom relief and currently represent the only FDA-approved non-hormonal treatment for hot flashes (60, 61). Despite this indication, SSRIs do not fully alleviate hot flashes and treatment switching is common due to tolerability issues (62). Some of this variability in response and side effects may be due to differences in SERT genotype, as persons with the *s/s* genotype have been shown to have a poor response to SSRIs (for treating anxiety disorders) (63). Specifically, *s/s* carriers are at greater risk for treatment-emergent side effects from these medications (64). Investigations into *5-HTTLPR* genotype by hot flash treatment efficacy could prove useful in advancing personalized medicine by informing treatment strategies, though to our knowledge, no such studies have been performed. This is likely to be a substantial challenge, as there is evidence that the effect of the polymorphism can be complex and modulated by ethnic background; for example the 'l' allele, as opposed to the 's' allele, confers increased risk for anxiety in a Chinese population (65).

Limitations and Future Directions

The sample size for our clinical component was small, but reflects a pilot investigation design. A follow-up study with a larger and more clinically diverse sample, including *s/s* or *s/l* carriers, women taking estrogen or other medications for hot flashes and those with additional risk factors such as asthma, is warranted to further validate this method and preclinical results. We would like to further delineate the prevalence and intensity of hot flashes following CO₂ inhalation at different concentrations and accompanying physiological events (i.e. skin conductance and temperature, core body temperature, cardiovascular profile, and blood chemistry, including pH throughout the laboratory session) and time course of evoked hot flashes. These are important considerations, because current procedures to elicit hot flashes in a clinical study typically use warming methods, such as water-perfused suits and/or heating pads(66) which are relevant to hot flashes occurring in warm environments but have questionable validity for hot flashes occurring in thermoneutral environments. In addition, these methods can require up to 20 minutes for a hot flash to occur. Importantly, while the ratings of severity of the hot flashes in this preliminary investigation were mild to moderate, we did note that the subjective hot flash intensity appeared to be worse in menopausal women with more frequent hot flashes, though with the small sample size, these correlations should be considered cautiously. Future studies will also need to determine how pre-existing anxiety or stress contributes to the presentation of

objective and subjective hot flashes post stress-induced hot flashes (e.g., mental arithmetic, emotionally salient films, or hypercapnic gas inhalation).

Potential Clinical Value

These studies demonstrated a novel method of provoking a hot flash using CO₂ inhalation and suggests that decreased breathing during stress, or conditions associated with acidosis [e.g. respiratory (e.g., COPD, bronchitis, or asthma) or metabolic (e.g., diabetes or lactic acidosis)] may increase the risk for hot flashes by reducing the threshold for hot flashes to occur, especially in combination with anxiety or thermal associated stimuli. Further study is warranted to fully validate this method, as our sample size was small and limited to otherwise healthy symptomatic women. A fine-grained delineation of the response to CO₂ will allow reverse translational studies (in highly controlled environments) that have the potential to help elucidate mechanisms underlying hot flashes.

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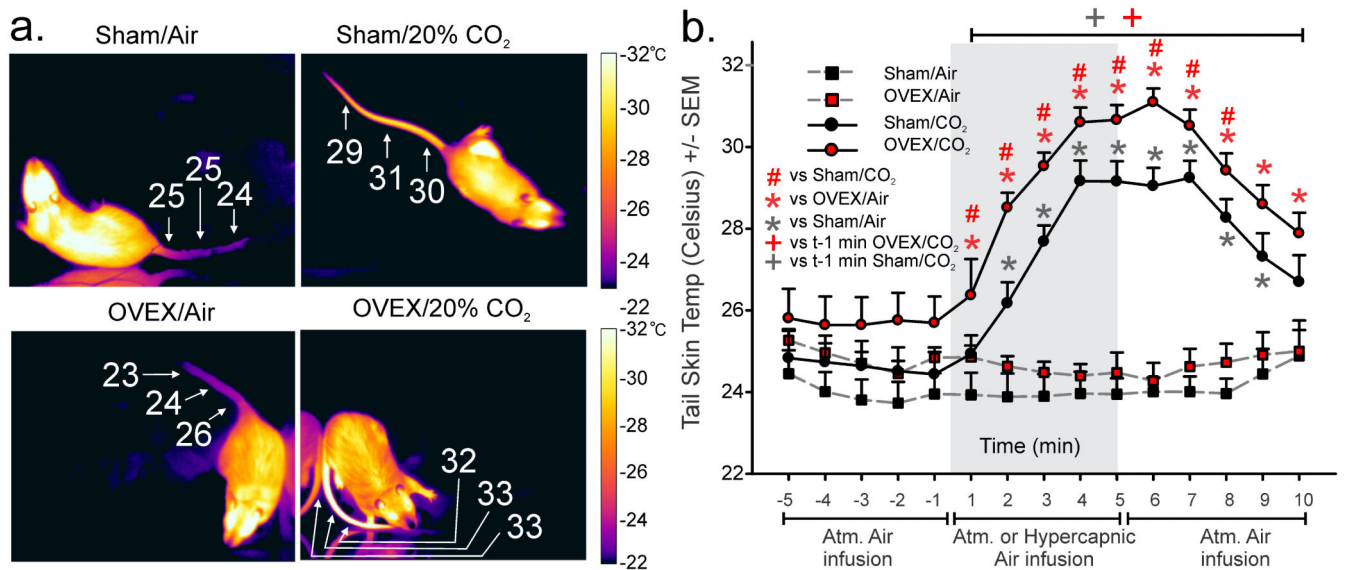


Figure 1. Rats modeling surgical menopause exhibit exacerbated hot flash-associated tail skin temperature responses

Effects of ovariectomy or sham surgery on baseline tail skin temperature in response to hypercapnic or atmospheric air infusion. a) Representative thermal images with scale (to the right) of a sham-OVEX (top) or OVEX rat (bottom) after 5 min exposure to atmospheric air (left) or 20% CO₂ (right). b) Line graph with error bars (SEM) represents mean tail skin temperature prior to and following atmospheric air or hypercapnic gas challenge in OVEX or sham-OVEX groups assessed with a tail thermistor at the base of the tail (n=15/group for CO₂ challenge and n=6/group for atmospheric air challenge). *denotes significance of surgical treatment at specific time points in (b) with Fisher's Least Significant Difference test protected by an ANOVA, and + denotes significant differences over time from t-1 as measured by Dunnett's test.

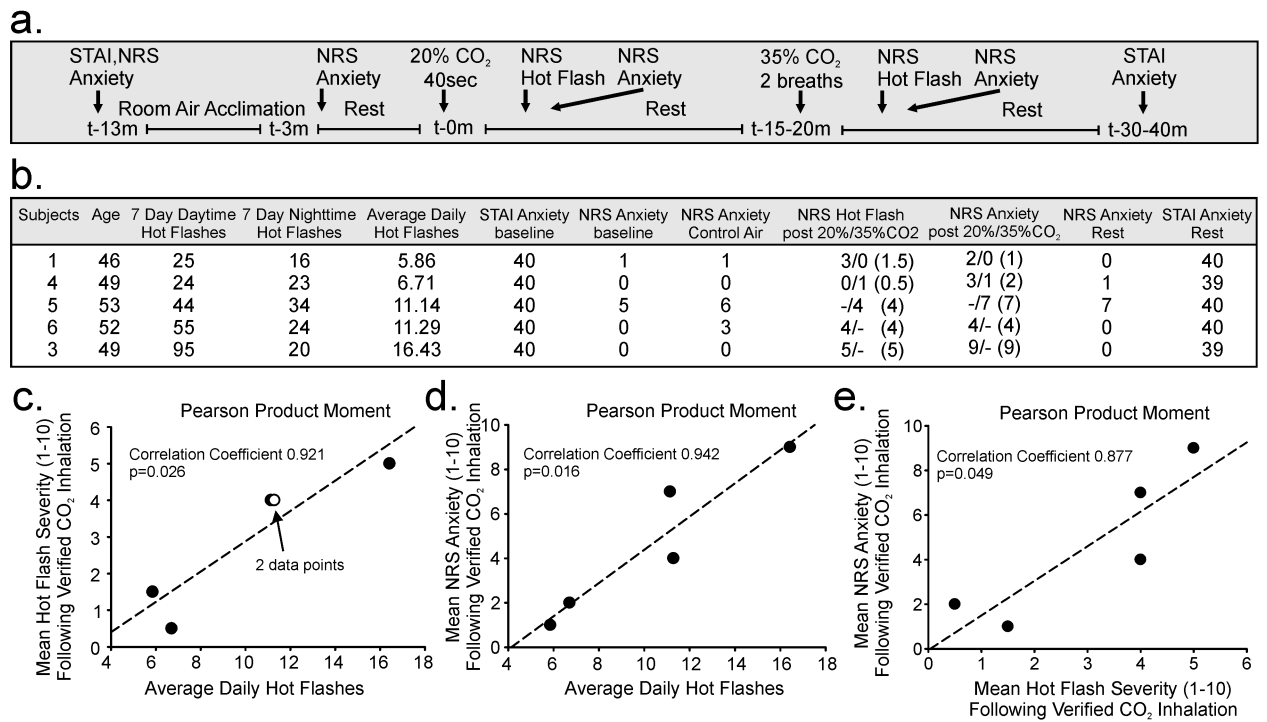


Figure 2. Small clinical study of hot flash provocation with CO₂ challenges in highly symptomatic women

a) Timeline for the study procedures; a room air acclimation (through a mask) was given for 10 min prior to a rest period before a 40s, 20% CO₂ (normoxic) challenge. Participants rested for 15-20 min prior to the 35% CO₂ administration. **b)** Table illustrates participants assigned number, age, total hot flashes during the day and night for a 7 day period prior to the study, average hot flashes per day, baseline STAI and NRS anxiety, NRS anxiety post control air, NRS hot flash and anxiety (respectively) severity post 20% and 35% CO₂ inhalation, followed by the NRS Anxiety and STAI anxiety at rest. Participants are ranked in order of increasing daily hot flashes. 3 of the 4 participants, and 2 of the 3 participants with confirmed inhalations of 20% CO₂ and 35%CO₂ reported hot flashes within 5 min, respectively; hot flashes were rated from mild to moderate. Numbers below NRS hot flashes or anxiety (columns 9 and 10, respectively) post CO₂ respectively represent 20% CO₂ and 35% CO₂ with the parenthetical number representing the mean of the confirmed inhalations (“-” indicates no confirmed inhalation for respective challenge). Correlation analyses revealed that the frequency of daily hot flashes (total per 24h day) was positively correlated with the **c)** mean severity of the hot flash post confirmed CO₂ inhalations and **d)** mean severity of the anxiety response post confirmed CO₂ inhalations. Correlation analysis also demonstrated that the **e)** mean hot flash severity following verified CO₂ inhalation was positively correlated with the mean anxiety rating following verified CO₂ inhalation.

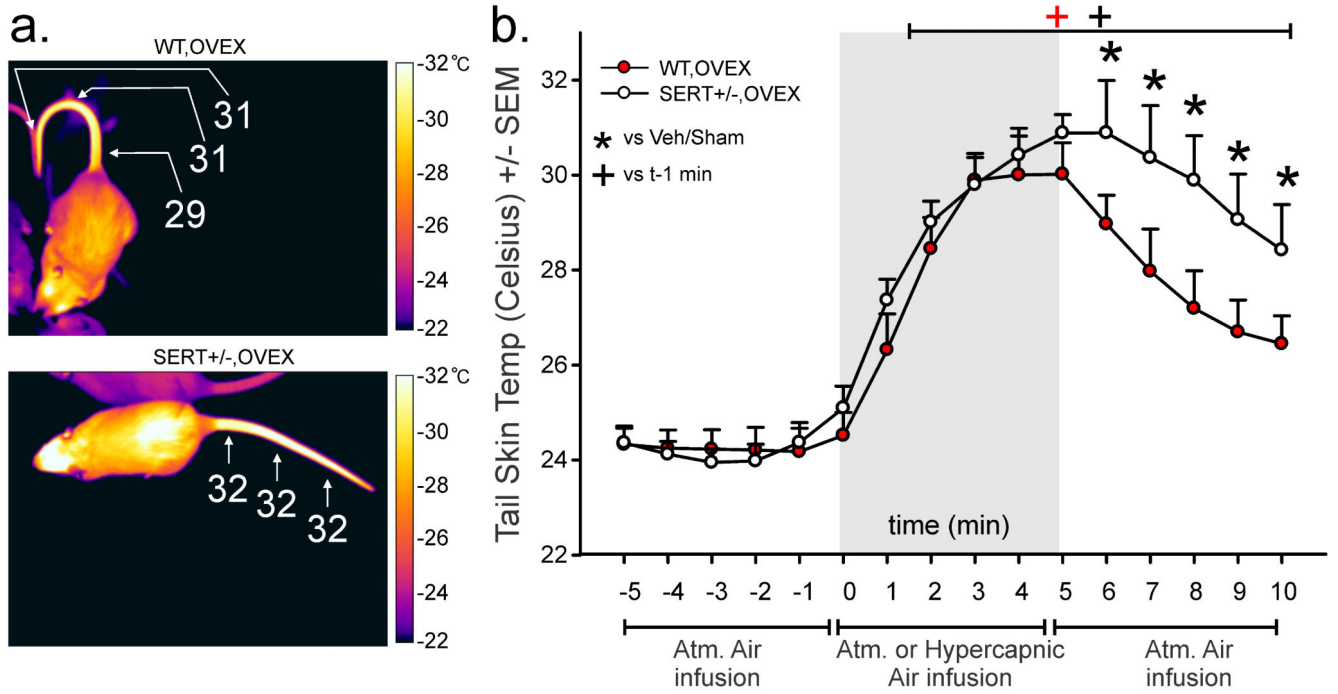


Figure 3. Rats with a heterozygous null mutation of the serotonin transporter ($SERT^{+/-}$) have prolonged hot flash-associated tail responses to hypercapnic gas infusion
 Effects of ovariectomy in $SERT^{+/-}$ and WT rats tail skin temperature in response to hypercapnic gas infusion. a) Representative thermal image with scale (to the right) of a WT-ovex (top) or $SERT^{+/-}$ rat (bottom) after 5 min exposure to 20% CO_2 . b) Line graph with error bars (SEM) represents mean tail skin temperature prior to and following hypercapnic gas challenge in ovex $SERT^{+/-}$ or WT rats assessed with a tail thermistor at the base of the tail (n=8,7). *denotes significant effect of estrogen treatment with a two-way ANOVA p=0.001 in (a). *denotes significance of genotype at specific time points in (c) with Fisher's Least Significant Difference test protected by an ANOVA, and + denotes significant differences over time from t-1 as measured by Dunnett's test.