

Technical University of Denmark



A critical review of producers of small lactone mycotoxins: patulin, penicillic acid and moniliformin

Frisvad, Jens Christian

Published in:
World Mycotoxin Journal

Link to article, DOI:
[10.3920/WMJ2017.2294](https://doi.org/10.3920/WMJ2017.2294)

Publication date:
2018

Document Version
Publisher's PDF, also known as Version of record

[Link back to DTU Orbit](#)

Citation (APA):
Frisvad, J. C. (2018). A critical review of producers of small lactone mycotoxins: patulin, penicillic acid and moniliformin. World Mycotoxin Journal, 11(1), 73-100. DOI: 10.3920/WMJ2017.2294

DTU Library
Technical Information Center of Denmark

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.



A critical review of producers of small lactone mycotoxins: patulin, penicillic acid and moniliformin

J.C. Frisvad

Department of Biotechnology and Biomedicine, Technical University of Denmark, Soltofts Plads Building 221, 2800 Kgs. Lyngby, Denmark; jcf@bio.dtu.dk

Received: 19 December 2017 / Accepted: 26 January 2018

© 2018 Wageningen Academic Publishers

OPEN ACCESS



REVIEW ARTICLE

Abstract

A very large number of filamentous fungi has been reported to produce the small lactone mycotoxins patulin, penicillic acid and moniliformin. Among the 167 reported fungal producers of patulin, only production by 29 species could be confirmed. Patulin is produced by 3 *Aspergillus* species, 3 *Paecilomyces* species, 22 *Penicillium* species from 7 sections of *Penicillium*, and one *Xylaria* species. Among 101 reported producers of penicillic acid, 48 species could produce this mycotoxin. Penicillic acid is produced by 23 species in section *Aspergillus* subgenus *Circumdati* section *Circumdati*, by *Malbranchea aurantiaca* and by 24 *Penicillium* species from 9 sections in *Penicillium* and one species that does not actually belong to *Penicillium* (*P. megasporum*). Among 40 reported producers of moniliformin, five species have been regarded as doubtful producers of this mycotoxin or are now regarded as taxonomic synonyms. Moniliformin is produced by 34 *Fusarium* species and one *Penicillium* species. All the accepted producers of patulin, penicillic acid and moniliformin were revised according to the new one fungus – one name nomenclatural system, and the most recently accepted taxonomy of the species.

Keywords: patulin, penicillic acid, moniliformin, terrestriac acid, *Penicillium*, *Aspergillus*, *Fusarium*

1. Introduction

Small toxic lactones, especially patulin, penicillic acid and moniliformin are recognised mycotoxins (Bemi, 2004; Ciegler *et al.*, 1971; Gaucher, 1979; McKinley and Carlton, 1991; Moake *et al.*, 2005; Puel *et al.*, 2010; Schütt *et al.*, 1998; Scott, 1974; Wilson, 1976), even though only patulin is under regulation (EC, 2003). They share some properties apart from being toxic and having some structural similarities. They are all polar, acidic, secondary metabolites often produced in large amounts in foods and feeds (Ciegler *et al.*, 1971), even though they differ in the niches their producers occupy: patulin is often associated with fruits and penicillic acid and moniliformin with cereals. Patulin and penicillic acid were early on recognised as effective antibiotics (Chain *et al.*, 1942; Florey *et al.*, 1949; Korzybski *et al.*, 1967; Oxford *et al.*, 1942; Singh, 1967), and have later been shown also to inhibit quorum sensing in bacteria (Rasmussen *et al.*, 2005). Patulin and penicillic acid also share a specific reaction with the sulphydryl-

containing amino acids cysteine and glutathione (Larsen and Olson, 1992; Morgavi *et al.*, 2003). While this may help inactivating these mycotoxins in foods containing cysteine and glutathione, those data indicate that part of the toxic effect of the small lactones is the binding of S-containing amino acids in proteins.

Apart from being an effective antibiotic, patulin can also produce petit mutants in *Saccharomyces cerevisiae* (Mayer and Legator, 1969). Patulin can act as a phytotoxin (Ellis *et al.*, 1977; Florey *et al.*, 1949; Ismail and Papenbrock, 2015; Korzybski *et al.*, 1967; Scott, 1974) and is active against pathogenic fungi (Gilliver, 1946; Herrick, 1945; Sanders, 1946). Patulin was initially suggested for treatment of common cold (Birkinshaw *et al.*, 1943; Gye, 1943), as it is anti-viral (Detry and Still, 1975) in addition to being antibacterial, but the trials on humans were quickly dropped because of the toxicity of patulin.

Patulin has been reported to be acutely toxic (Ciegler, 1977; Devaraj *et al.*, 1986; Drusch and Ragab, 2003; Engel and Teuber, 1984; Escoula *et al.*, 1977; Fung and Clark, 2004; Hayes *et al.*, 1979; Hopkins, 1993; McKinley and Carlton, 1980a,b; McKinley *et al.*, 1982; Wouters and Speijers, 1996), teratogenic (Ciegler *et al.*, 1976; Vesely and Vesela, 1991), cardiotoxic (Vesely and Vesela, 1991), it influences the colonic epithelial permeability (Maidana *et al.*, 2016; Mohan *et al.*, 2012), and it is immunotoxic (Bourdiol *et al.*, 1990; Escoula *et al.*, 1988; Paucod *et al.*, 1990; Sharma, 1993). Patulin is first of all found in pomaceous foods like apples and pears, but also in other fruits (Beretta *et al.*, 2000; Jackson and Dombrinck-Kurtzman, 2006; Yang *et al.*, 2014).

Penicillic acid was first reported by Alsberg and Black (1913) in an examination of fungal growth in maize and potential fungal involvement in pellagra. Its structure was elucidated by Birkinshaw *et al.* (1936) and its antibacterial effect against Gram positive and Gram negative bacteria was first reported by Oxford in 1942, Oxford *et al.* in 1942 and Geiger and Conn in 1945, as reviewed by Korzybski *et al.* (1967). Later the antibiotic effect of penicillic acid has also been reported by Ezzat *et al.* (2007). Penicillic acid is also very inhibitory towards *Phytophthora* species (Kang and Kim, 2004). Furthermore, penicillic acid is phytotoxic (Ismail and Papenbrock, 2015).

Penicillic acid has been shown to be acutely toxic to poultry, mice, rats and rabbits (Cole and Cox, 1981). Furthermore, it dilates blood vessels and possesses antidiuretic properties (Murnaghan, 1946). Penicillic acid has been involved in broiler chicken mycotoxicosis (Ghadevaru, 2013; Samadha and Balachandran, 2008; Sarmadha *et al.*, 2008) but gingerol has been shown to alleviate the toxic effects of penicillic acid (Pazhavinel *et al.*, 2015). Penicillic acid is cytotoxic (Grabsch *et al.*, 2006), hepatotoxic (Chan and Hayes, 1981), and has been claimed to be carcinogenic and cardiotoxic (Dickens and Jones, 1961, 1963, 1965; Vesely and Vesela, 1991), and a sodium, potassium and calcium channel blocker in frog hearts (Pandiyan *et al.*, 1990). It is also toxic to rat alveolar macrophages (Sorenson and Simpson, 1986).

Ochratoxin A and penicillic acid show synergistic toxicity on bovine macrophage cell lines (Oh *et al.*, 2015), in chickens (Kubena *et al.*, 1984; Micco *et al.*, 1989), in the corn ear worm and the fall armyworm (Dowd, 1989). The two toxins together also increase degeneration in kidney proximal tubules (Klarić *et al.*, 2013; Stoev, 2013; Stoev and Denev, 2013). The synergistic worm toxicity of penicilllic acid and ochratoxin A is interesting as the fungi producing these combinations of toxins are often growing in corn and other cereals (Lund and Frisvad, 1994; Visagie *et al.*, 2014a). Other studies have only shown an additive toxic effect on porcine lymphocytes of ochratoxin A and penicillic acid (Bernhoft *et al.*, 2004; Stoev *et al.*, 1999). These two mycotoxins are often co-occurring as *Aspergillus* from section *Circumdati*

produce penicillic acid and ochratoxin simultaneously in most species from that section (Visagie *et al.*, 2014a). Furthermore, species from *Penicillium* section *Fasciculata* series *Viridicata* and *Verrucosa* co-occur in cereals (Lund and Frisvad, 1995), and thus penicillic acid and ochratoxin A are also both found often in contaminated cereals.

It has been suggested that ochratoxin A and penicillic acid may both be involved in Balkan Endemic Nephropathy (Stoev, 2017; Stoev *et al.*, 2001). *Penicillium polonicum* and *Penicillium aurantiogriseum*, both producing penicillic acid, verrucosidin and nephrotoxic glycopeptides, are often found in the Balkan countries, but *Penicillium verrucosum*, the main ochratoxin A producing *Penicillium* species in cereals, is found less often (MacGeorge and Mantle, 1990; Mantle, 1994; Mantle *et al.*, 2010; Miljkovic *et al.*, 2003; Stoev *et al.*, 2010; Yeulet *et al.*, 1988). Penicillic acid is mostly found in cereals (Bokhari and Flannigan, 1996; Chelkowski and Golinski, 1983; Chelkowski *et al.*, 1983, 1987; Kurtzman and Ciegler, 1970; Szeklak *et al.*, 1981; Tangni and Pussemier, 2007) and some other products like cassava (Wareing *et al.*, 2001).

Terrestric acid is rather closely related to penicillic acid, and is produced by *P. aurantiogriseum*, *Penicillium crustosum* (= *Penicillium terrestre*), *Penicillium hirsutum*, *Penicillium hordei*, *Penicillium radicum*, *Penicillium tricolor*, *Penicillium tulipae*, *Penicillium venetum* and *Magnaporthe grisea* = *Pyricularia oryzae* (Birkinshaw and Raistrick, 1936; Frisvad and Samson, 2004; Frisvad *et al.*, 1994, 2004; Lund and Frisvad, 1994; Overy and Frisvad, 2003; Overy *et al.*, 2005; Sonjak *et al.*, 2005; Yu *et al.*, 2010). Production of terrestric acid by *Penicillium griseoroseum* (Da Silva *et al.*, 2013), and *Penicillium viride* (Birkinshaw and Samant, 1960) could not be confirmed (Frisvad and Samson, 2004), other metabolites produced by these organisms indicate they were in fact *P. crustosum* (Frisvad *et al.*, 2014). Terrestric acid has also been reported from *P. hirsutum* var. *albocoremium* (Frisvad and Filtenborg, 1989), but this taxon was later subdivided in *Penicillium albocoremium*, *P. radicum* and *P. tulipae*. Only the last two species produce terrestric acid (Overy and Frisvad, 2003; Overy *et al.*, 2005). This small acidic lactone is also phytotoxic (Nukina, 1988) and cardiotoxic (Giarman, 1948, 1949), thus resembling penicillic acid in some of its bioactivities. *P. aurantiogriseum*, *P. radicum* and *P. tulipae* can produce both penicillic acid and terrestric acid (Frisvad *et al.*, 2004; Overy and Frisvad, 2003; Overy *et al.*, 2005).

Like penicillic acid, moniliformin has also been found in cereals and other plant products (Herrare *et al.*, 2017; Kovalsky *et al.*, 2016; Neme and Mohammed, 2017; Uhlig *et al.*, 2004). An important fact is that while moniliformin is mostly produced by important field-fungi, such as *Fusarium* spp., penicillic acid is produced by storage fungi only. Moniliformin has not been reported as an antibiotic or

quorum sensing inhibitor in bacteria (Bacon *et al.*, 2017), but moniliformin is acutely toxic to poultry (Burmeister *et al.*, 1979; Cole *et al.*, 1973; Sharma *et al.*, 2008) and rats (Chen *et al.*, 1990; Jonsson *et al.*, 2013, 2015; Thiel, 1978), in the latter study indicating a severe impact on the immune system. It inhibits several enzymes important in metabolism (Burka *et al.*, 1982). It is cardiotoxic and effects mammalian smooth muscle (Kamyar *et al.*, 2006; Peltonen *et al.*, 2010). Moniliformin is also phytotoxic and has plant growth-regulating properties (Cole *et al.*, 1973).

There is one example of co-production of penicillic acid and patulin by *Penicillium roqueforti* (Olivigni and Bullerman, 1977, 1978), but the producing organism was reidentified to *Penicillium carneum* later (Boysen *et al.*, 1996; Frisvad and Samson, 2004; Frisvad *et al.*, 2004). *Penicillium melanoconidium* has been reported to produce moniliformin and penicillic acid (Hallas-Møller *et al.*, 2016), but no species has been reported to produce patulin and moniliformin concomitantly.

It was the aim of this review to revise the lists of producers of patulin, penicillic acid and moniliformin according to the most recent taxonomic treatments of the genera *Aspergillus* (Chen *et al.*, 2016a,b, 2017; Frisvad, 2015; Frisvad and Larsen, 2015; Hubka *et al.*, 2015, 2016; Jurjevics *et al.*, 2015; Kocsubé *et al.*, 2016; Samson *et al.*, 2011a,b, 2014, 2017; Sklenář *et al.*, 2017; Varga *et al.*, 2007a,b, 2011a,b; Visagie *et al.*, 2014a, 2017), *Penicillium* (Frisvad and Samson, 2004; Houbraken *et al.*, 2016; Visagie *et al.*, 2014b) and *Fusarium* (Aoki *et al.*, 2014; Burgess, 2014; Gerlach and Nirenberg, 1982; Lesley and Summerell, 2016; Nelson *et al.*, 1983), as correct species-mycotoxin relations are important for prevention of mycotoxin formation in foods, feeds and biotechnological products.

2. Criteria for accepting a species as a producer of one or more small toxic lactones

Confirmation of mycotoxin production by a species of a filamentous fungus requires that both the fungus and the mycotoxin are correctly identified. Furthermore, it should be verified that the fungus examined is a pure culture. Also the fungal strain producing the mycotoxins should be available to the scientific community. In several cases the producing fungi were never accessioned in a culture collection, so the experiments could not be repeated and the identity of the fungal strain could not be verified or validated. If the mycotoxinogenic isolate was identified using a polyphasic approach, using a combination of morphological, physiological, chemical, and molecular features, the identifications are often correct. Molecular identification should be based on sequences of several genes, because ITS sequences are not always sufficient for correct identification at the species level (Raja *et al.*, 2017; Seifert *et al.*, 2007). Several independent reports of production of a

particular mycotoxin by a particular fungal species adds to the credibility of a species-mycotoxin connection (Frisvad, 1989; Frisvad and Samson, 2004). Often production of a particular secondary metabolite is taxonomically restricted to rather few, often phylogenetically related species, so unexpected mycotoxin formation from phylogenetically unrelated species to known producers should be confirmed with special care. It should be noted that some groups of species have been revised several times, so a name that was valid earlier now may just be a synonym or a rejected name. Furthermore, the one fungus – one name system introduced in 2011 has caused a lot of changes. For example species earlier called *Eurotium*, *Neosartorya*, *Emericella* are now all called *Aspergillus* (Samson *et al.*, 2014) and *Fusarium* is the preferred name for species formerly allocated to *Gibberella* (Geiser *et al.*, 2013). It also adds to the credibility of the reported specific mycotoxin production, if the toxin has been detected in a large number of isolates in the species examined.

It should also be documented that the growth medium used for mycotoxin production does not contain the mycotoxin already. Often secondary metabolites are accumulated, so repeated reporting of trace amounts of mycotoxins on media otherwise suited for production may indicate that the mycotoxin was already present in the growth medium. This is most important for media based on cereals, as these can contain trace amounts of mycotoxins if field or storage fungi have grown in such substrates. It is in general recommended to analyse control samples for mycotoxins and use chemically defined media in a confirmation of mycotoxin production, if unexpected results have been obtained using plant based media.

Chemical analysis, for example using HPLC methods for separation and combined with sensitive detection methods will also inherently carry the risk of carry over of mycotoxins, so here proper controls are also needed. Chromatographic and spectrometric verification of the identity of the mycotoxin is needed to guard against false positives. If the retention times in several chromatographic systems is equal to the retention time of an authentic chemical standard, this is valuable confirmation of identity, but on top of that high resolution mass spectra (MS) and/or UV spectra, as compared to an authentic standard are necessary to confirm identity. It is recommended to have at least 3 different independent confirmations of identity, and one of them should preferably be high resolution mass spectrometry (identical mass ions and relevant adducts as compared to a standard) and even the same mass fragmentation pattern as an authentic standard. Occasionally chemical identity is confirmed by nuclear magnetic resonance (NMR) characterisation, circular dichroism (CD) spectra, fourier transform infrared (FTIR) spectra, etc., especially the first time a secondary metabolite is structure elucidated and isolated from a

particular species. NMR, CD and FTIR can also be used for a very solid confirmation of identity. Other types of data, for example detection of biosynthetic precursors, and discovery of the gene cluster or part of it for the particular mycotoxin under scrutiny adds further data to support a mycotoxin-fungal species connection. The criteria for accepting a fungus as a producer of patulin, penicillic acid or moniliformin include the following: production has been reported several times and in several different strains (more than one time and from different research groups), fungal identification has been documented, the fungal producing strains are available for the scientific community, and at least three chromatographic and spectrometric kinds of confirmatory data have been provided. If, in addition, parts of the gene cluster for the mycotoxins have been found, NMR data for the mycotoxin have been made available, or the biosynthesis has been confirmed, for example by detecting biosynthetic precursors, the documentation for such relationships is of course even stronger.

3. Patulin and producing organisms

The biosynthesis of patulin, starting from 6-methyl salicylic acid is well known (Barad *et al.*, 2016c; Birch *et al.*, 1955; Bu'Lock and Ryan 1958; Dimroth *et al.*, 1976; Forrester and Gaucher, 1972a,b; Priest, 1989; Puel *et al.*, 2010; Scott *et al.*, 1971, 1972; Sekiguchi, 1983; Sekiguchi and Gaucher, 1978; Tanenbaum and Bassett, 1959), and the gene cluster for patulin has been found in different producing filamentous fungi (Ballester *et al.*, 2015; Barad *et al.*, 2016c; Dombrink-Kurtzman, 2007; Li *et al.*, 2015; Nielsen *et al.*, 2017; Puel *et al.*, 2010; Snini *et al.*, 2014; Tannous *et al.*, 2014). The role of patulin itself in the fruit infection process has not been fully elucidated (Barad *et al.*, 2016a,b), but it has been suggested that accumulation of both gluconic acid and patulin (Barad *et al.*, 2012, 2014, 2016a; Snini *et al.*, 2015) will make *Penicillium expansum* a more aggressive post-harvest pathogen. However, patulin may also be important for *P. expansum* itself in the competition with other microorganisms thriving in the fruit. It should be noted that the two other apple pathogenic Penicillia, *Penicillium solitum* and *P. crustosum*, do not produce patulin (Frisvad and Samson, 2004; Sanderson and Spotts, 1995), questioning the importance of patulin in the infection process.

The main end-product is patulin itself, but other related toxins are known, such as neopatulin, isopatulin, desoxypatulinic acid, ascladiol, longianone and pintulin (Appell *et al.*, 2009; Bennett *et al.*, 1990, 1991; Ghisalberti *et al.*, 2000; Lykakis *et al.*, 2009; Mikami *et al.*, 1996; Scott *et al.*, 1972; Sekiguchi *et al.*, 1979).

Many strains producing patulin have been genome sequenced, including *P. expansum* (Ballester *et al.*, 2015; Barad *et al.*, 2016c; Julca *et al.*, 2015; Yin *et al.*, 2017; Yu *et*

al., 2014), *P. griseofulvum* (Banani *et al.*, 2016), *Penicillium antarcticum*, *P. carneum*, *Penicillium paneum*, and *Penicillium vulpinum* (Nielsen *et al.*, 2017). A comparison of the gene clusters for patulin production in the different species shows they are quite similar, and that *P. roqueforti* has part of the gene cluster needed to produce patulin, but lacks essential genes, making it unable to produce patulin (Nielsen *et al.*, 2017).

Patulin has been reported from 166 different species, varieties and chemotypes (Table 1), but the documentation for some of all these producers is occasionally doubtful. In many of the publications, the original producers reported are not available to the scientific community. For example, in the papers by Steiman *et al.* (1989), Okeke *et al.* (1993), and Oh *et al.* (1998), accession numbers of the fungi examined are not available, and thus it is not possible to verify the identity of the producers. In the paper by Okeke *et al.* (1993) some very efficient producers of patulin are reported, often known producers of patulin, such as *P. expansum*, while most of the trace producers of patulin are fungi not expected to produce patulin, such as *Trichothecium roseum*. It has been a general experience that when a fungus can produce patulin, it will always accumulate large amounts of the toxin, never a small trace of a toxin (Frisvad, personal observations). It is tempting to speculate that the detection of patulin in unexpected species was caused by carry over in the HPLC chromatographic system or because of contamination with efficient patulin producers. Patulin has been found in sections *Canescens*, *Formosana*, *Gladioli*, *Osmophila*, *Penicillium*, *Ramosa* and *Robsamsonia* in *Penicillium*, but only in section *Clavati* in *Aspergillus*.

Patulin has the formula $C_7H_6O_4$, and a molecular mass of 154, and a monoisotopic mass of 154.026. Searching in the Microbial secondary metabolite database AntiBase with approximately 40,000 natural products, 95 secondary metabolites have the molecular mass of 154, while 13 fungal secondary metabolites had a monoisotopic mass of 154.026, including members of the patulin biosynthetic pathway patulin, isopatulin, neopatulin, gentisic acid, longianone and (-)-phylllostine. However, other secondary metabolites, such as terreic acid and terremutin also have the monoisotopic mass of 154.026. This may be the reason *Aspergillus terreus* was reported to produce patulin (Draughon and Ayers, 1980; El-Shanawany *et al.*, 2005; Escoula, 1974; Girisham and Reddy, 1986a,b; Giridhar and Reddy, 1998; Kent and Heatley, 1945; Reddy and Reddy, 1984, 1988), while it was probably terreic acid that was detected. Given several references on patulin production by *A. terreus*, it cannot be excluded that this species, or one of the closely related species, can produce it. Obviously for such small molecules, mass spectrometric (MS) verification is of value, but has to be combined with other data, such as an UV spectrum identical to that of patulin and retention time verification,

Table 1. Filamentous fungi claimed to produce patulin (also called clavacin, clavatin, claviformin, expansin and penicidin).

Fungal species	References	Synonymised species
<i>Acremonium implicatum</i>	Ismail <i>et al.</i> , 2016	
<i>Acremonium persicinum</i>	Okeke <i>et al.</i> , 1993	
<i>Acremonium sclerotigenum</i>	Okeke <i>et al.</i> , 1993	
<i>Acremonium zaeae</i>	Steiman <i>et al.</i> , 1989	
<i>Alternaria alternata</i>	Drusch and Ragab, 2003; Leidou <i>et al.</i> , 2001; Steiman <i>et al.</i> , 1989	
<i>Alternaria papaveris</i>	Steiman <i>et al.</i> , 1989	
<i>Alternaria tenuissima</i>	Okeke <i>et al.</i> , 1993	
<i>Ascochyta imperfecta</i>	Steiman <i>et al.</i> , 1989	
<i>Aspergillus amstelodami</i>	Steiman <i>et al.</i> , 1989	
<i>Aspergillus candidus</i>	Frank, 1977	
<i>Aspergillus clavatus</i>	Bergel <i>et al.</i> , 1943, 1944; Diaz and Flannigan, 1997; Escoula, 1974; Lopez-Varga <i>et al.</i> , 2007; Paterson, 2004 (IDH gene); Umezawa <i>et al.</i> , 1947; Wiesner, 1942	
<i>Aspergillus echinulatus</i>	Steiman <i>et al.</i> , 1989	
<i>Aspergillus flavus</i>	Luque <i>et al.</i> , 2011	
<i>Aspergillus fumigatus</i>	Steiman <i>et al.</i> , 1989	
<i>Aspergillus giganteus</i>	Florey <i>et al.</i> , 1944; Okeke <i>et al.</i> , 1993; Paterson, 2004 (IDH gene); Varga <i>et al.</i> , 2007a	
<i>Aspergillus longivesica</i>	Varga <i>et al.</i> , 2007a	
<i>Aspergillus manginii</i>	Steiman <i>et al.</i> , 1989	
<i>Aspergillus ochraceus</i>	Abu-Seidah, 2003	
<i>Aspergillus oryzae</i>	Chunmei <i>et al.</i> , 2013; Luque <i>et al.</i> , 2011	
<i>Aspergillus parasiticus</i>	Steiman <i>et al.</i> , 1989	
<i>Aspergillus petrakii</i>	Okeke <i>et al.</i> , 1993	
<i>Aspergillus repens</i>	Steiman <i>et al.</i> , 1989	
<i>Aspergillus sydowii</i>	Chunmei <i>et al.</i> , 2013	
<i>Aspergillus tamarii</i>	Luque <i>et al.</i> , 2011	
<i>Aspergillus terreus</i>	Couch and Gaucher, 2004 (MSA gene); El-Shanawany <i>et al.</i> , 2005; Escoula, 1974; Draughon and Ayers, 1980; Giridhar and Reddy, 1998; Girisham and Reddy, 1986a,b; Kent and Heatley, 1945; Paterson, 2004 (IDH gene); Reddy and Reddy, 1984, 1988	
<i>Aspergillus variecolor</i>	Steiman <i>et al.</i> , 1989	
<i>Aspergillus versicolor</i>	Steiman <i>et al.</i> , 1989	
<i>Aureobasidium pullulans</i> var. <i>pullulans</i>	Steiman <i>et al.</i> , 1989	
'Basidio 2'	Steiman <i>et al.</i> , 1989	
<i>Botrytis allii</i>	Steiman <i>et al.</i> , 1989	
<i>Byssochlamys fulva</i>	Escoula, 1974; Paterson, 2004 (IDH gene); Sant'Ana <i>et al.</i> , 2010; rejected by Puel <i>et al.</i> , 2007	see <i>Paecilomyces fulvus</i>
<i>Byssochlamys nivea</i>	Escoula, 1974; Kis <i>et al.</i> , 1969; Paterson, 2004 (IDH gene); Sant'Ana <i>et al.</i> , 2010; Scurti <i>et al.</i> , 1973	see <i>Paecilomyces niveus</i>
<i>Calcarosporium arbuscula</i>	Okeke <i>et al.</i> , 1993; Steiman <i>et al.</i> , 1989	
<i>Chaetomium atrobrunneum</i>	Okeke <i>et al.</i> , 1993	
<i>Chondrostereum purpureum</i>	Okeke <i>et al.</i> , 1993	
<i>Chrysosporium pannorum</i>	Steiman <i>et al.</i> , 1989	
<i>Cladobotryum varium</i>	Steiman <i>et al.</i> , 1989	
<i>Cladobotryum verticillatum</i>	Steiman <i>et al.</i> , 1989	
<i>Cladorrhinium</i> sp.	Okeke <i>et al.</i> , 1993; Steiman <i>et al.</i> , 1989	
<i>Colletotrichum musae</i>	Steiman <i>et al.</i> , 1989	
<i>Coniothyrium</i> sp.	Okeke <i>et al.</i> , 1993	
<i>Cunninghamella bainieri</i>	Steiman <i>et al.</i> , 1989	
<i>Curvularia lunata</i>	Steiman <i>et al.</i> , 1989	
<i>Cylindrocarpon cylindroides</i>	Okeke <i>et al.</i> , 1993	
<i>Cylindrocarpon ianthotele</i>	Okeke <i>et al.</i> , 1993	
<i>Cylindrocarpon olidum</i>	Okeke <i>et al.</i> , 1993	

Table 1. Continued.

Fungal species	References	Synonymised species
<i>Dichotomomyces cepii</i>	Okeke et al., 1993	
<i>Emericella quadrilineata</i>	Luque et al., 2011	
<i>Emericella rugulosa</i>	Luque et al., 2011	
<i>Emericella variecolor</i>	Luque et al., 2011	
<i>Eupenicillium brefeldianum</i>	Okeke et al., 1993; Steiman et al., 1989	
<i>Eupenicillium javanicum</i>	Okeke et al., 1993	
<i>Eupenicillium</i> sp. 1	Okeke et al., 1993	
<i>Eupenicillium</i> sp. 2	Okeke et al., 1993	
<i>Fusarium culmorum</i>	Steiman et al., 1989	
<i>Fusarium proliferatum</i> var. <i>proliferatum</i>	Steiman et al., 1989	
<i>Fusarium</i> sp.	Xie et al., 2011	
<i>Gymnoascus</i> sp.	Karow and Forster, 1944; Kuehn, 1958	
<i>Gymnoascus reesii</i>	Okeke et al., 1993; Steiman et al., 1989	
<i>Mortierella bainieri</i>	Steiman et al., 1989	
<i>Mucor hiemalis</i>	Steiman et al., 1989	
<i>Mucor racemosus</i> var. <i>globosus</i>	Steiman et al., 1989	
<i>Oidiodendron echinulatum</i>	Steiman et al., 1989	
<i>Oidiodendron tenuissimum</i>	Steiman et al., 1989	
<i>Paecilomyces lilacinus</i>	Okeke et al., 1993; Steiman et al., 1989	
<i>Paecilomyces variotii</i>	Escoula, 1975; Paterson, 2004 (IDH gene)	
<i>Paecilomyces fulvus</i>	renamed here from <i>Byssochlamys fulva</i>	
<i>Paecilomyces niveus</i>	renamed here from <i>Byssochlamys nivea</i>	
<i>Paecilomyces saturatus</i>	Samson et al., 2009	
<i>Penicillium antarcticum</i>	Vansteelant et al., 2012	
<i>Penicillium asperosporum</i>	Moslem et al., 2011	
<i>Penicillium aurantiogriseum</i>	Luque et al., 2011; Moslem et al., 2010; Oh et al., 1998; Okeke et al., 1993; Paterson, 2004 (IDH gene); Paterson et al., 2003; Steiman et al., 1989	
<i>Penicillium brevicompactum</i>	Paterson, 2004 (IDH gene); Paterson et al., 2003	
<i>Penicillium camemberti</i>	Luque et al., 2011	
<i>Penicillium canescens</i>	Steiman et al., 1989	
<i>Penicillium carneum</i>	Boysen et al., 1996; Dombrinck-Kurtzman, 2007; Nielsen et al., 2006	
<i>Penicillium chrysogenum</i>	Chunmei et al., 2013; Leistner and Pitt, 1977; Oh et al., 1998; Steiman et al., 1989	
<i>Penicillium citreonigrum</i>	Steiman et al., 1989	
<i>Penicillium citrinum</i>	El-Samawaty et al., 2013; Frank, 1977; Oh et al., 1998	
<i>Penicillium claviforme</i>	Afiyatullov et al., 2015; Bergel et al., 1943; Borkowska Opacka and Escoula, 1977; Chain et al., 1942; Frisvad and Filtenborg, 1983	see <i>P. vulpinum</i>
<i>Penicillium clavigerum</i>	Dombrinck-Kurtzman, 2007; Dombrinck-Kurtzman and Blackburn, 2005; Svendsen and Frisvad, 1994	
<i>Penicillium commune</i>	Luque et al., 2011; Oh et al., 1998	
<i>Penicillium compactum</i>	Houbraaten et al., 2016	
<i>Penicillium concentricum</i>	Dombrinck-Kurtzman, 2007; Frisvad and Samson, 2004; Frisvad et al., 2004; Leistner and Eckardt, 1979	
<i>Penicillium concentricum</i> II	Frisvad and Filtenborg, 1983; Houbraaten et al., 2016	see <i>P. coprobum</i>
<i>Penicillium coprobum</i>	Frisvad and Filtenborg, 1989; Frisvad and Samson, 2004; Frisvad et al., 2004; Dombrinck-Kurtzman and Blackburn, 2005; Houbraaten et al., 2016	
<i>Penicillium corylophilum</i>	Paterson, 2004 (IDH gene); Vismer et al., 1996	
<i>Penicillium crustosum</i>	Northolt et al., 1978; Yun et al., 2006	
<i>Penicillium cyaneofulvum</i>	Berestetskii et al., 1974	
<i>Penicillium cyaneum</i>	Okeke et al., 1993	

Table 1. Continued.

Fungal species	References	Synonymised species
<i>Penicillium cyclopium</i>	Efimenko and Yakimov, 1960; Frank, 1972; 1977; Leistner and Pitt, 1977	
<i>Penicillium dipodomyicola</i>	Dombrinck-Kurtzman, 2007; Frisvad and Samson, 2004; Frisvad et al., 1987, 2004; Houbraken et al., 2016; Luque et al., 2011	
<i>Penicillium dipodomys</i>	Koteswara Rao et al., 2011	
<i>Penicillium divergens</i>	Barta and Mecir, 1948;	see <i>P. glandicola</i>
<i>Penicillium diversum</i>	Okeke et al., 1993	
<i>Penicillium duclauxii</i>	Okeke et al., 1993	
<i>Penicillium echinulatum</i>	Okeke et al., 1993	
<i>Penicillium equinum</i>	Burton, 1949; Burton and Pausacker, 1947	see <i>P. expansum</i>
<i>Penicillium expansum</i>	Luijk, 1938; Andersen et al., 2004; Anslow et al., 1943; Borkowska Opacka and Escoula, 1977; Casquete et al., 2017; Chunmei et al., 2013; Dombrinck-Kurtzman, 2007; Dombrinck-Kurtzman and Blackburn, 2005; El-Samawaty et al., 2013; Frisvad and Filtenborg, 1983; Harwig et al., 1973; Larsen et al., 1998; Okeke et al., 1993; Paster et al., 1995; Paterson, 2004 (IDH gene); Paterson et al., 2003; Sommer et al., 1974; Steiman et al., 1989; Welke et al., 2011	
<i>Penicillium fellutanum</i>	Vismer et al., 1996	
<i>Penicillium funiculosum</i>	Steiman et al., 1989; Vismer et al., 1996; Yassin et al., 2010	
<i>Penicillium glabrum</i>	Okeke et al., 1993	
<i>Penicillium gladioli</i>	Dombrinck-Kurtzman, 2007; Frisvad and Samson, 2004; Frisvad et al., 2004	
<i>Penicillium glandicola</i>	Frisvad and Filtenborg, 1989; Houbraken et al., 2016; Paterson, 2004 (IDH gene)	see <i>P. concentricum</i>
<i>Penicillium glandicola</i> var. <i>glaucovenetum</i>	Dombrinck-Kurtzman, 2007; Frisvad and Filtenborg, 1989; Paterson, 2004 (IDH gene)	
<i>Penicillium granulatum</i>	Borkowska Opacka and Escoula, 1977; Frisvad et al., 1983	see <i>P. glandicola</i>
<i>Penicillium griseofulvum</i>	Chunmei et al., 2013; Dombrinck-Kurtzman, 2007; Dombrinck-Kurtzman and Blackburn, 2005; Frisvad and Filtenborg, 1983, 1989; Frisvad et al., 1987; Houbraken et al., 2016; Kent and Heatley, 1945; Moslem et al., 2010; Okeke et al., 1993; Oh et al., 1998; Paterson, 2004 (IDH gene); Simonart and Lathouwer, 1956; Steiman et al., 1989; Welke et al., 2011	
<i>Penicillium griseofulvum</i> var. <i>dipodomycola</i>	Frisvad and Filtenborg, 1989; Frisvad et al., 1987; Paterson, 2004 (IDH gene)	see <i>P. dipodomycola</i>
<i>Penicillium hirsutum</i>	Okeke et al., 1993; Paterson et al., 2004	
<i>Penicillium italicum</i>	Okeke et al., 1993	
<i>Penicillium janczewskii</i>	only IDH gene found, Paterson, 2004	
<i>Penicillium lanosum</i>	Kharchenko, 1970	
<i>Penicillium lapidosum</i>	Myrchink, 1967	
<i>Penicillium leucopus</i>	Anslow et al., 1943; Brian et al., 1956; Umezawa et al. 1947	see <i>P. expansum</i>
<i>Penicillium lignorum</i>	Okeke et al., 1993	see <i>P. griseofulvum</i>
<i>Penicillium maltum</i>	Ukai et al., 1954	
<i>Penicillium marinum</i>	Frisvad and Samson, 2004; Frisvad et al., 2004	
<i>Penicillium melanoconidium</i>	Luque et al., 2011	
<i>Penicillium melinii</i>	Frisvad and Filtenborg, 1990; Karow and Forster, 1944; Okeke et al., 1993; Paterson, 2004 (IDH gene)	
<i>Penicillium miczynskii</i>	Chunmei et al., 2013	
<i>Penicillium novae-zeelandiae</i>	Burton, 1949; Burton and Pausacker, 1947; Frisvad and Filtenborg, 1990	
<i>Penicillium paneum</i>	Boysen et al., 1996; Dombrinck-Kurtzman, 2007; Frisvad and Samson, 2004; Nielsen et al., 2006	
<i>Penicillium patulum</i>	Anslow et al., 1943; Birkinshaw et al., 1943; Chain et al., 1942; Frisvad, 1981	see <i>P. griseofulvum</i>
<i>Penicillium polonicum</i>	Luque et al., 2011	
<i>Penicillium psychrosexualis</i>	Houbraken et al., 2010	
<i>Penicillium puberulum</i>	El-Samawaty et al., 2013; Moslem et al., 2011	
<i>Penicillium purpurogenum</i>	Xie et al., 2011	

Table 1. Continued.

Fungal species	References	Synonymised species
<i>Penicillium raistrickii</i>	Veselá and Veselý, 1995	
<i>Penicillium rivolii</i>	Berestetskii <i>et al.</i> , 1975	
<i>Penicillium roqueforti</i>	Bullerman, 1978; Cakmakci <i>et al.</i> , 2015; Chunmei <i>et al.</i> , 2013; Erdogan <i>et al.</i> , 2003; Leistner and Pitt, 1977; Malekinejad <i>et al.</i> , 2015; Olivigni and Steiman <i>et al.</i> , 1989; Müller and Amend, 1997; Paterson <i>et al.</i> , 2003; Vismér <i>et al.</i> , 1996	
<i>Penicillium roqueforti</i> chemotype II	Frisvad and Filtenborg, 1983	see <i>P. paneum</i>
<i>Penicillium roqueforti</i> var. <i>carneum</i>	Frisvad and Filtenborg, 1989; Vismér <i>et al.</i> , 1996	see <i>P. carneum</i>
<i>Penicillium rugulosum</i>	Leistner and Pitt, 1977; Vismér <i>et al.</i> , 1996	
<i>Penicillium samsonianum</i>	Houbraken <i>et al.</i> , 2016	
<i>Penicillium sclerotigenum</i>	Dombrinck-Kurtzman, 2007; Frisvad <i>et al.</i> , 2004	
<i>Penicillium selandiae</i>	<i>nomen nudum</i> , in IMI Culture Collection Catalogue, 2010; Frisvad, unpublished	see <i>P. antarcticum</i>
<i>Penicillium simplicissimum</i>	Okeke <i>et al.</i> , 1993; Paterson, 2004 (IDH gene)	
<i>Penicillium</i> sp.	Atkinson, 1942	
<i>Penicillium spinulosum</i>	only IDH gene, Paterson, 2004	
<i>Penicillium terrestre</i>	Atkinson <i>et al.</i> , 1944	
<i>Penicillium thomii</i>	Borkowska Opacka and Escoula, 1977	
<i>Penicillium urticae</i>	Kent and Heatley, 1945; Yamamoto, 1954	see <i>P. griseofulvum</i>
<i>Penicillium variabile</i>	Leistner and Pitt, 1977; Steiman <i>et al.</i> , 1989	
<i>Penicillium verrucosum</i>	Lüke <i>et al.</i> , 2011; Moslem <i>et al.</i> , 2013; Oh <i>et al.</i> , 1998	
<i>Penicillium viridicatum</i>	Frank, 1972	
<i>Penicillium vulpinum</i>	Dombrinck-Kurtzman, 2007; Frisvad <i>et al.</i> , 2004; Houbraken <i>et al.</i> , 2016; Ismaiel and Papenbroek, 2015; Ismaiel <i>et al.</i> , 2016	
<i>Pestalotiopsis</i> sp.	Okeke <i>et al.</i> , 1993	
<i>Phialophora hoffmannii</i>	Okeke <i>et al.</i> , 1993	
<i>Pseudodiplodia</i> sp.	Steiman <i>et al.</i> , 1989	
<i>Rhinocladiella atrovirens</i>	Steiman <i>et al.</i> , 1989	
<i>Scopulariopsis</i> sp.	Steiman <i>et al.</i> , 1989	
<i>Scopulariopsis flava</i>	Okeke <i>et al.</i> , 1993	
<i>Scytalidium lignicola</i>	Okeke <i>et al.</i> , 1993	
<i>Spicelleum roseum</i>	Okeke <i>et al.</i> , 1993	
<i>Sporormiella minimoides</i>	Steiman <i>et al.</i> , 1989	
<i>Sporothrix schenckii</i>	Steiman <i>et al.</i> , 1989	
<i>Stemphylium</i> sp.	Okeke <i>et al.</i> , 1993	
<i>Stemphylium vesicarium</i>	Laidou <i>et al.</i> , 2001	
<i>Talaromyces purpureogenus</i>	Ismaiel <i>et al.</i> , 2016	
<i>Talaromyces trachyspermus</i>	Okeke <i>et al.</i> , 1993	
<i>Trametes squalens</i>	Steiman <i>et al.</i> , 1989	
<i>Trichoderma pseudokoningii</i>	Steiman <i>et al.</i> , 1989	
<i>Trichoderma polysporum</i>	Steiman <i>et al.</i> , 1989	
<i>Trichophyton mentagrophytes</i>	Steiman <i>et al.</i> , 1989	
<i>Trichophyton persicolor</i>	Okeke <i>et al.</i> , 1993	
<i>Trichothecium roseum</i>	Okeke <i>et al.</i> , 1993	
<i>Xylaria longiana</i>	Edwards <i>et al.</i> , 1999; Goss <i>et al.</i> , 1999; Lykakis <i>et al.</i> , 2009	

¹ Confirmed producers are in bold.

and comparison to an authentic standard (Kildgaard *et al.*, 2014). A MS/MS analysis will only yield few fragments for patulin, and thus several other verification tests have to be done to ensure that the metabolite detected is indeed patulin. In the paper by Luque *et al.* (2011) patulin was reported for several known producers and in addition species, such as *Aspergillus flavus*, *Penicillium camemberti*, and *P. verrucosum*. In tests of those fungi for patulin this has never been verified in those species (Frisvad *et al.*, 2004; Varga *et al.*, 2011b). Luque *et al.* (2011) used micellar capillary electrophoresis (retention time) and/or HPLC-MS (retention time and MS) in addition to reporting on a PCR amplicon (496 basepairs) for the FC2/IDH gene involved in the patulin biosynthesis, and despite this, they detected patulin in species that do obviously not produce patulin. Other records of patulin producers are correct, but sometimes the fungi are just synonyms of another species. An example of this is *Penicillium griseofulvum*, a very efficient and consistent producer of patulin, and three of its synonyms *P. maltum*, *P. patulum* and *P. urticae* (Pitt, 1979) have been repeatedly reported to produce patulin (Table 1).

Given the dubious accounts of patulin production by many species, a revision of confirmed and efficient producers of patulin is given in Table 1 (species in bold). Only 29 species can be reliably said to produce patulin. Among these *P. expansum* is the most important, producing patulin in apples, pears, plums and other fruits (Dombrink-Kurtzman and McGovern, 2007; Filtenborg *et al.*, 1996; Frisvad and Samson, 2004). *Penicillium sclerotigenum*, specifically associated to sweet potatoes, is also an important patulin-producing species (Yamamoto *et al.*, 1955), as is *P. griseofulvum* that may produce patulin in wheat and malted barley (Jiménez *et al.*, 1991; Ukai *et al.*, 1954; Yamamoto *et al.*, 1954). Similarly *Aspergillus clavatus* can produce patulin in malted barley (Lopez-Diaz and Flannigan, 1997). *Penicillium dipodomyicola*, another efficient patulin producer is occasionally found in foods (Dombrink-Kurtzman and McGovern, 2007; Frisvad and Samson, 2004). Members of *Penicillium* section *Roquefortorum* all produce patulin, except *P. roqueforti*. Since *P. roqueforti* is used for blue mould cheese production, it is particularly important to know whether this fungus produces this mycotoxins. The data of Nielsen *et al.* (2017) show that *P. roqueforti* has most of the gene cluster needed for production of patulin, but some genes are missing, explaining the inability of *P. roqueforti* to produce patulin. The reports of patulin production by *P. roqueforti* (Cakmakci *et al.*, 2015; Chunmei *et al.*, 2013; Erdogan *et al.*, 2003; Leistner and Pitt, 1977; Malekinejad *et al.*, 2015; Müller and Amend, 1997; Olivigni and Bullerman, 1978; Paterson *et al.*, 2003; Steiman *et al.*, 1989; Vismer *et al.*, 1996) appear to be caused by misidentifications, because the closely related species *P. carneum* and *P. paneum* both produce patulin (Boysen *et al.*, 1996). *Paecilomyces fulvus* (formerly *Byssochlamys fulva*),

Paecilomyces niveus (formerly *Byssochlamys nivea*) and *Paecilomyces saturatus* have all been reported to produce patulin. There has been some doubt whether *Pae. fulvus* can produce patulin (reported by Escoula, 1974; Paterson, 2004 and Sant'Ana *et al.*, 2010, but rejected by Puel *et al.*, 2007), but these species all have heat resistant ascospores (Samson *et al.*, 2009), and thus may survive in pasteurised fruit juices and produce patulin (Rice *et al.*, 1977; Sant'Ana *et al.*, 2010). *Paecilomyces niveus* and *Penicillium paneum* can grow in substrates with a high concentration of acetic acid and thus can also be found in silage, and may produce patulin there (Gallo *et al.*, 2015; O'Brien *et al.*, 2006; Scurti *et al.*, 1973). *P. antarcticum* is a marine-derived *Penicillium* species and may produce patulin on seaweed or even in shellfish (Geiger *et al.*, 2013; Vansteelandt *et al.*, 2012). *Penicillium marinum* is also marine-derived and may produce patulin in the same seaborne organisms (Frisvad and Samson, 2004).

The remaining species producing patulin are of less consequence for mycotoxin production in foods. *Penicillium gladioli* may potentially produce patulin in gladiolus bulbs, and *Penicillium glandicola* can potentially produce patulin on acorns, but flower bulbs and acorns are not used for food or feed. *Penicillium compactum*, *Penicillium novaezealandiae* and *Penicillium samsonianum* are soil or grass-borne fungi (Houbreken *et al.*, 2016) and *Aspergillus longivesica*, *Aspergillus giganteus*, *Penicillium clavigerum*, *Penicillium concentricum*, *Penicillium coprobiuum*, *Penicillium formosanum*, and *P. vulpinum* are all dung fungi and of no consequence for food safety (Frisvad and Samson, 2004).

For all of the species listed above, patulin has been detected in a large number of isolates of the producing species, for example patulin was detected in 83 out of 85 isolates of *P. expansum* by Andersen *et al.* (2004), and produced by 350/357 isolates out of *P. expansum*, by 40 out of 40 isolates in *P. griseofulvum*, by 13/13 isolates in *P. coprobiuum*, by 14/14 isolates in *P. glandicola*, by 14/15 strains of *P. concentricum* (= *P. glandicola* var. *glaucovenetum*), by 10/12 isolates of *P. carneum* (= *P. roqueforti* var. *carneum*), by 4/4 isolates of *P. dipodomyicola* (= *P. griseofulvum* var. *dipodomyicola*) and by 6/6 isolates of *P. vulpinum* (Frisvad and Filtenborg, 1989), showing a high consistency in patulin production by these species.

4. Penicillic acid and producing organisms

Penicillic acid is the main product of the biosynthetic pathway that starts with orsellinic acid and has penicillic acid as end product (Axberg and Gatenbeck, 1975a,b; Eldridge *et al.*, 1977). However there are other members of the penicillic acid pathway, including 5,6-dihydropenicillic acid, 5,6-dihydro-6-hydroxypenicillic acid, 6-methoxy-5,6-dihydropenicillic acid, coclunol, and (4R,5R)-4,5-

dihydroxy-3-methoxy-5-methylcyclohex-2-en-1-one (He *et al.*, 2004; Kimura *et al.*, 1996; Nonaka *et al.*, 2015; Obana *et al.*, 1995a,b; Phainuphong *et al.*, 2017; Qi *et al.*, 2015; Raphael, 1947a,b; Shiono *et al.*, 2005).

Penicillic acid has been reported in 101 species, varieties and chemotypes of fungi (Table 2), of which 48 are regarded as well documented. Some of the data may be incorrect connections between fungal species and penicillic acid because of carry over in the chromatographic system, or contamination by penicillic acid producing species. In the paper by Gutarowska *et al.* (2010), *Aspergillus niger* and *Aspergillus flavus* are reported to produce penicillic acid, and this has never been reported from these Aspergilli in any other papers. Again the isolates have not been accessioned in any fungal collection, but furthermore, the concomitant detection of penicillic acid and viridicatin indicates that the cultures of *A. flavus* and *A. niger* were contaminated with *Penicillium cyclopium*, which is a known producer of these two secondary metabolites. In the control sample on malt extract agar (MEA) with no fungi inoculated, Gutarowska *et al.* (2010) found kojic acid, indicating that a kojic acid producer had been growing in the barley malt before it was made into malt extract.

Producers of penicillic acid are especially common in *Penicillium* section *Viridicata* and in *Aspergillus* section *Circumdati* (Frisvad and Samson, 2000; Frisvad *et al.*, 2004a; Visagie *et al.*, 2014a). One can therefore expect a mycotoxin cocktail in mouldy cereals consisting of citrinin, ochratoxin A, penicillic acid, verrucosidin, nephrotoxic glycopeptides, xanthomegnin, viomellein, vioxanthin, moniliformin, and penitrem A when associated storage fungal species, such as *P. verrucosum*, *P. aurantiogriseum*, *P. polonicum*, *P. viridicatum*, *P. cyclopium* and *Penicillium freii* have grown on those cereals in a worst case scenario (Frisvad and Samson, 2004; Frisvad *et al.*, 2004; Hallas-Møller *et al.*, 2016; Lund and Frisvad, 1994). Most species in *Aspergillus* section *Circumdati*, such as *Aspergillus westerdijkiae*, *Aspergillus steynii* and *Aspergillus ochraceus* also produce one or more of the mycotoxins ochratoxin A, penicillic acid, xanthomegnin, viomellein and vioxanthin (Visagie *et al.*, 2014a). Penicillic acid has been found in many sections of *Penicillium*, including *Brevicompacta*, *Canescens*, *Chrysogena*, *Exilicaulis*, *Fasciculata*, *Lanata-Divaricata*, *Ramosa*, *Roquefortorum* and *Turbatum*, so this is found in many phylogenetic groups of *Penicillium*. Penicillic acid has only been found in one section of *Aspergillus*: *Circumdati*.

Other species listed in Table 2 producing penicillic acid include soil borne species, such as *Penicillium baarnense*, *Penicillium egyptiacum*, and *Penicillium jamesonlandense*, grass-associated fungi, such as *Penicillium brasiliense* and dung fungi, such as *Penicillium bovifimosum* also produce

penicillic acid, but such fungi will probably not grow in foods or feeds, and are less relevant for food safety.

Among the common species producing penicillic acid, there is a high consistency in producing this mycotoxin. In *P. aurantiogriseum*, more than 278 isolates out of 385 examined produced penicillic acid, and it was produced by 17/17 isolates of *P. melanoconidium*, by 35/53 isolates of *Penicillium neoechinulatum*, by 234/276 isolates of *P. polonicum*, and by 324/350 isolates of *P. viridicatum* (Frisvad and Filtenborg, 1989).

Terrestric acid, often co-occurring with penicillic acid, is produced by cereal borne fungi such as *P. aurantiogriseum*, *P. tricolor* and *P. hordei*, by onion and bulb associated fungi, such as *P. radicicola*, *P. tulipae*, and *P. venetum* in addition to being produced by the apple, cheese and nut associated *P. crustosum* and the rice associated fungus *M. grisea* (Frisvad and Samson, 2004; Yu *et al.*, 2010) (Table 3). This toxin may thus co-occur in cereals with penicillic acid, moniliformin, ochratoxin A, verrucosidins, and xanthomegnins (Frisvad and Samson, 2004).

5. Moniliformin and producing microorganisms

Moniliformin is a mycotoxin that was first reported from *Fusarium moniliforme* (Cole *et al.*, 1973), but has since been reported to be produced by many species in that genus (Table 4). It is biosynthesised from two acetyl units (Franck and Breipohl, 1984; Gathercole *et al.*, 1986). Most of the producers of moniliformin have been confirmed by data from later papers (Table 4), and rejected producers mostly are only synonyms of names later accepted. Most of these 40 species reported to produce moniliformin do indeed produce it, except for 6 species, where Schütt *et al.* (1998) could not confirm production by these species. It should be noted that moniliformin production is difficult to verify because of the low molecular weight and simple MS fragmentation pattern, so often TLC and the characteristic UV spectrum has been used to confirm identity. In the database on natural products called AntiBase, among more than 40,000 compounds, only one has a formula of $C_4H_2O_3$, so simple HR-MS data are also confirmative of production. In addition to its production by *Fusarium* species, *Penicillium melanoconidium* has been reported to produce moniliformin (Hallas-Møller *et al.*, 2016). A large number of these species are cereal-borne, including *P. melanoconidium* which has, however, only been found on stored cereals. However, other species of *Fusarium* are pathogens on potatoes, tomatoes, onions, etc. (Lesley and Summerell, 2016), and thus moniliformin can occur in many cereals, fruits and vegetables. Furthermore, moniliformin is produced by isolates of *Fusarium fujaroides* from millet, sorghum, dried fish and peanuts (Rabie *et al.*, 1978). In addition *Fusarium* species produce several other mycotoxins concomitantly, such as trichothecenes, zearalenone,

Table 2. Filamentous fungi claimed as producers of penicillic acid.¹

Fungal species	References	Synonymised species
<i>Aspergillus affinis</i>	Visagie et al., 2014a	
<i>Aspergillus auricomus</i>	Ciegler 1972; Frisvad et al., 2004a; Visagie et al., 2014a	
<i>Aspergillus bridgeri</i>	Frisvad et al., 2004a; Kumar et al., 2011; Visagie et al., 2014a	
<i>Aspergillus cervinus</i>	He et al., 2004	
<i>Aspergillus cretensis</i>	Frisvad et al., 2004a; Visagie et al., 2014a	
<i>Aspergillus elegans</i>	Obana et al., 1995b	
<i>Aspergillus flavus</i>	Gutarowska et al., 2010	
<i>Aspergillus flocculosus</i>	Frisvad et al., 2004a; Montenegro et al., 2012; Visagie et al., 2014a	
<i>Aspergillus fumigatus</i>	Şenyuva et al., 2008	
<i>Aspergillus insulicola</i>	Frisvad et al., 2004a; Visagie et al., 2014a	
<i>Aspergillus melleus</i>	Ciegler, 1972; Frisvad et al., 2004a; Gill-Carey 1949; Obana et al., 1995b; Visagie et al., 2014a	
<i>Aspergillus muricatus</i>	Frisvad and Samson, 2000; Frisvad et al., 2004a; Visagie et al., 2014a	
<i>Aspergillus neobridgeri</i>	Frisvad et al., 2004a; Visagie et al., 2014a	
<i>Aspergillus nidulans</i>	Abu-Seidah, 2003	
<i>Aspergillus niger</i>	Gutarowska et al., 2010	
<i>Aspergillus occultus</i>	Visagie et al., 2014a	
<i>Aspergillus ochraceopetaliformis</i>	Visagie et al., 2014a	
<i>Aspergillus ochraceus</i>	Ciegler, 1972; El-Shanawany et al., 2005; Frisvad et al., 2004a; Garza et al., 1993; Karow et al., 1944; Northolt et al., 1979; Obana et al., 1995b; Visagie et al., 2014a	
<i>Aspergillus ostianus</i>	Ciegler, 1972; Frisvad et al., 2004a; Namikoshi et al., 2003; Obana et al., 1995b; Visagie et al., 2014a	
<i>Aspergillus pallidofulvus</i>	Visagie et al., 2014a	
<i>Aspergillus persii</i>	Frisvad et al., 2004a; Visagie et al., 2014a	
<i>Aspergillus petrakii</i>	Frisvad et al., 2004a	
<i>Aspergillus pseudoelegans</i>	Frisvad et al., 2004a	
<i>Aspergillus pulvericola</i>	Visagie et al., 2014a	
<i>Aspergillus quercinus</i>	Gill-Carey, 1949	
<i>Aspergillus salwaensis</i>	Visagie et al., 2014a	
<i>Aspergillus roseoglobulosus</i>	Frisvad et al., 2004a; Visagie et al., 2014a	
<i>Aspergillus sclerotiorum</i>	Ciegler, 1972; Frisvad et al., 2004a; Kang and Kim, 2004; Kang et al., 2007; Obana et al., 1995b; Visagie et al., 2014a; Zheng et al., 2010	
<i>Aspergillus</i> sp.	Li et al., 2010	
<i>Aspergillus subramanianii</i>	Visagie et al., 2014a	
<i>Aspergillus sulphureus</i>	Ciegler, 1972; Frisvad et al., 2004a; Gill-Carey, 1949	
<i>Aspergillus violaceus</i>	Abu-Seidah, 2003	
<i>Aspergillus wentii</i>	He et al., 2004	
<i>Aspergillus westerdijkiae</i>	Frisvad et al., 2004a; Visagie et al., 2014a	
<i>Aspergillus westlandensis</i>	Visagie et al., 2014a	
<i>Eupenicillium bovinosum</i>	Tuthill and Frisvad, 2002	see <i>P. bovinosum</i>
<i>Exophiala</i> sp.	Zhang et al., 2008	
<i>Malbranchea aurantiaca</i>	Martínez-Luis et al., 2005	
<i>Paecilomyces ehrlichii</i>	Gorbach and Friederick, 1949	
<i>Penicillium atramentosum</i>	Bridge et al., 1989	
<i>Penicillium aurantiogriseum</i>	Bridge et al. 1989; Bokhari and Flannigan, 1996; El-Banna et al., 1987; Frisvad and Filtenborg 1983; Frisvad et al., 2004b; Garza et al., 1993; Khaddor et al., 2007; Lund and Frisvad 1994; Mills et al., 1995a,b; Oh et al., 1998	
<i>Penicillium aurantiogriseum</i> var. <i>polonicum</i>	Frisvad and Filtenborg, 1989	see <i>P. polonicum</i>
<i>Penicillium aurantiogriseum</i> var. <i>neoechinulatum</i>	Frisvad et al., 1987	see <i>P. neoechinulatum</i>

Table 2. Continued.

Fungal species	References	Synonymised species
<i>Penicillium aurantiogriseum</i>	Frisvad and Filtenborg, 1989	see <i>P. melanoconidium</i>
var. <i>melanoconidium</i>		
<i>Penicillium aurantiovirens</i>	Lund and Frisvad, 1994; Wirth and Klosek, 1972	presently regarded as a synonym of <i>P. cyclopium</i>
<i>Penicillium baarnense</i>	Burton, 1949; Mosbach, 1960	
<i>Penicillium bovifinosum</i>	Tuthill and Frisvad, 2002; Visagie et al., 2014b	
<i>Penicillium brasiliatum</i>	Frisvad, 1989; Frisvad and Filtenborg, 1990	
<i>Penicillium brevicompactum</i>	Schurman et al., 2010	
<i>Penicillium canescens</i>	Paterson et al., 1987	
<i>Penicillium carneum</i>	Keromnes and Thouvenot, 1985; Kharchenko, 1970	
<i>Penicillium castellae</i>	Boysen et al., 1996; Frisvad et al., 2004	
<i>Penicillium castellae</i>	Quintanilla, 1982	see <i>P. raistrickii</i>
<i>Penicillium chrysogenum</i>	Jiménez et al., 1991; Leistner and Pitt, 1977; Oyero and Oyefulo, 2009	
<i>Penicillium citrinum</i>	Delgado et al., 2011; El-Samawaty et al., 2013	
<i>Penicillium claviforme</i>	Ueno, 1994	
<i>Penicillium commune</i>	Ciegler et al., 1972; Mintzlaff et al., 1972	
<i>Penicillium cordubense</i>	Skóra et al., 2017	see <i>P. polonicum</i>
<i>Penicillium cremeogriseum</i>	Frisvad and Filtenborg, 1990	
<i>Penicillium cyclopium</i>	Bentley and Keil, 1962; Birkinshaw et al., 1936; Ciegler et al., 1972; Frisvad and Samson, 2004; Jiménez et al., 1991; Keromnes and Thouvenot, 1985; Lei et al., 2010; Lindenfelser and Ciegler, 1977; Lund and Frisvad, 1994; Mintzlaff et al., 1972; Northolt et al., 1979	
<i>Penicillium expansum</i>	Ciegler et al., 1972; Mintzlaff et al., 1972; Patterson and Damoglou, 1985	
<i>Penicillium fennelliae</i>	Van Eijk, 1969	
<i>Penicillium freii</i>	Frisvad et al., 2004; Lund and Frisvad, 1994	
<i>Penicillium frequentans</i>	Yamaji et al., 2005	
<i>Penicillium funiculosum</i>	Nasser, 2008	
<i>Penicillium granulatum</i>	Bridge et al., 1989	
<i>Penicillium griseofulvum</i>	Moslem et al., 2010; Reio, 1958	
<i>Penicillium griseum</i>	Gorbach and Friederick, 1949	
<i>Penicillium hirsutum</i>	Bridge et al., 1989; Ezzat et al., 2007	
<i>Penicillium hirsutum</i> var. <i>albocoremium</i>	Frisvad and Filtenborg, 1989	
<i>Penicillium islandicum</i>	Nasser, 2008	
<i>Penicillium jamesonlandense</i>	Frisvad et al., 2006	
<i>Penicillium janczewskii</i>	Frisvad and Filtenborg, 1990	
<i>Penicillium janthinellum</i>	Ciegler et al., 1972; Mintzlaff et al., 1972	
<i>Penicillium lilacinum</i>	Kharchenko, 1970	
<i>Penicillium lividum</i>	Gorbach and Friederick, 1949	
<i>Penicillium madriti</i>	Birkinshaw and Gowlland, 1962	also named <i>P. matriti</i>
<i>Penicillium martensii</i>	Kurtzman and Ciegler, 1970; Lillehoj et al., 1972; Mintzlaff et al., 1972; Northolt et al., 1979; Wirth et al., 1956	
<i>Penicillium megasporum</i>	Nozawa et al., 1989; not a <i>Penicillium</i> (Frisvad and Filtenborg, 1990; Visagie et al., 2014b)	
<i>Penicillium melanoconidium</i>	Lund and Frisvad 1994; Frisvad and Samson, 2004	
<i>Penicillium neoechinulatum</i>	Lund and Frisvad 1994; Frisvad and Samson, 2004	see <i>P. viride</i>
<i>Penicillium ochraceum</i>	Tsunoda et al., 1978	see <i>P. viride</i>
<i>Penicillium olivinoviride</i>	Kobayashi et al., 1971	see <i>P. viride</i>
<i>Penicillium oxalicum</i>	Oh et al., 1998	

Table 2. Continued.

Fungal species	References	Synonymised species
<i>Penicillium palitans</i>	Ciegler and Kurtzman, 1970	
<i>Penicillium paraherquei</i>	Leistner and Eckardt, 1979	
<i>Penicillium piscarium</i>	Leistner and Pitt, 1977	
<i>Penicillium polonicum</i>	Frisvad and Samson, 2004; Lund and Frisvad, 1994; Skóra et al., 2017	
<i>Penicillium puberulum</i>	Alsberg and Black 1913; El-Samawaty et al., 2013; Moslem et al., 2011	
<i>Penicillium pulvillorum</i>	Frisvad and Filtenborg, 1990	see <i>P. cyclopium</i>
<i>Penicillium radicicola</i>	Frisvad and Samson, 2004; Overy and Frisvad, 2003, 2005	
<i>Penicillium raistrickii</i>	Bridge et al., 1989; Frisvad, 1988; Frisvad and Filtenborg, 1990	
<i>Penicillium rolfssii</i>	Frisvad, 1989	
<i>Penicillium roqueforti</i>	Bridge et al., 1989; Cakmakci et al., 2012, 2015; Erdogan et al., 2003; Moubasher et al., 1978; Malekinejad et al., 2015; Mioso et al., 2015; Müller and Amend, 1997; Olivigni and Bullerman, 1978	
<i>Penicillium simplicissimum</i>	Betina et al., 1969; Ciegler et al., 1972; El-Banna et al., 1987; Mintzlaff et al., 1972; Takahashi et al., 2008	
<i>Penicillium solitum</i>	Bridge et al., 1989	
<i>Penicillium</i> sp.	Komagata et al., 1996; Tachibana et al., 2008	
<i>Penicillium stoloniferum</i>	Alsberg and Black, 1913; Clutterbuck et al., 1932	
<i>Penicillium suavolens</i>	Ciegler et al., 1972; Lindenfelser and Ciegler, 1977	
<i>Penicillium subrubescens</i>	Karow et al., 1944	see <i>P. carneum</i>
<i>Penicillium thomi/P. thomii</i>	Karow et al. 1944; Casquete et al., 2017	
<i>Penicillium tulipae</i>	Frisvad and Samson, 2004; Overy and Frisvad, 2003; Overy et al., 2005;	
<i>Penicillium vanderhammenii</i>	Houbraken et al., 2011	
<i>Penicillium verrucosum</i>	Gedek et al., 1981; Moslem, 2013; Oh et al., 1998	
<i>Penicillium viridicatum</i>	Bresler et al., 1995; Ciegler et al., 1972, 1973; Frisvad and Samson, 2004; Jiménez et al., 1991; Lund and Frisvad, 1994; Mintzlaff et al., 1972; Northolt et al., 1979; Oh et al., 1998	
<i>Trichoderma</i> spp. ²	Lebed et al., 1978	

¹ Confirmed producers are in bold.

² In this case penicillic acid is referring to a member of the penicilin biosynthetic pathway.

enniatins and several other mycotoxins or emerging mycotoxins (Bräse et al., 2009; Gruber-Dorninger et al., 2017; Neme and Mohammed, 2017). Since moniliformin is produced by so many species one can maybe expect that apart from its mammal and plant toxicity, it could have antibiotic or quorum sensing inhibition abilities.

6. Conclusions

The small acidic lactone mycotoxins, such as patulin, penicillic acid and moniliformin can occur in many foods and feeds, and are toxic by themselves, but often there may be a potential synergistic or additive effect with other co-occurring mycotoxins. Based on the species producing patulin, fruit products are most prone to contamination with this mycotoxin. Penicillic acid is more common on stored cereals, as the best producers are mostly cereal-borne.

Moniliformin is produced by a large number of *Fusarium* species, and many of those are common on cereals, but also on fruits and vegetables. Among a large number of species reported to produce patulin and penicillic acid, only 29 and 48 species, respectively, have been reliably reported to produce them. The species producing them are mostly belonging to *Penicillium* and *Aspergillus*. Moniliformin is produced by 34 *Fusarium* species and by one species of *Penicillium*. The revised lists of producers of these mycotoxins will aid in preventive mycotoxin work, but also help in de-selecting known producers of these antibiotic toxins, when screening for new antibiotics.

Table 3. Filamentous fungi reported as producers of terrestriac acid.¹

Fungal species	References	Synonymised species
<i>Magnaporthe oryzae</i>	Couch and Kohn, 2002 (taxonomy); Yu et al., 2010	
<i>Pyricularia oryzae</i>	Yu et al., 2010	see <i>M. oryzae</i>
<i>Penicillium aurantiogriseum</i>	Frisvad and Samson, 2004; Lund and Frisvad, 1994	
<i>Penicillium aurantiogriseum</i> var. <i>aurantiogriseum</i>	Frisvad and Filtenborg, 1989	see <i>P. aurantiogriseum</i>
<i>Penicillium crustosum</i>	Frisvad and Filtenborg, 1989; Frisvad and Samson, 2004; Sonjak et al., 2005	
<i>Penicillium griseoroseum</i>	Da Silva et al., 2013	
<i>Penicillium hirsutum</i>	Frisvad and Samson, 2004; Overy and Frisvad, 2003; Overy et al., 2005	
<i>Penicillium hirsutum</i> var. <i>albocoremium</i>	Frisvad and Filtenborg, 1989	
<i>Penicillium hirsutum</i> var. <i>hirsutum</i>	Frisvad and Filtenborg, 1989	see <i>P. hirsutum</i>
<i>Penicillium hirsutum</i> var. <i>hordei</i>	Frisvad and Filtenborg, 1989	see <i>P. hordei</i>
<i>Penicillium hirsutum</i> var. <i>venetum</i>	Frisvad and Filtenborg, 1989	see <i>P. venetum</i>
<i>Penicillium hordei</i>	Frisvad and Samson, 2004; Overy et al., 2005	
<i>Penicillium terreste</i>	Birkinshaw and Samant, 1960	
<i>Penicillium tricolor</i>	Frisvad and Samson, 2004; Frisvad et al., 1994	
<i>Penicillium radicicola</i>	Overy and Frisvad, 2003; Overy et al., 2005	
<i>Penicillium tulipae</i>	Overy and Frisvad, 2003; Overy et al., 2005	
<i>Penicillium venetum</i>	Frisvad and Samson, 2004; Overy and Frisvad, 2003; Overy et al., 2005	
<i>Penicillium viridicatum</i>	Birkinshaw and Samant, 1960	

¹ Confirmed producers are in bold.

Acknowledgements

This work was supported by The Novo Nordic Foundation (NNF13OC0005201) and the Danish National Research Foundation (DNRF137) for the Center for Microbial Secondary Metabolites, and by an Agilent Thought Leader Award (#2871).

References

- Abbas, H.K., Boyette, C.D. and Hoagland, R.E., 1995. Phytotoxicity of *Fusarium*, other fungal isolates, and of the phytotoxins fumonisin, fusaric acid, and moniliformin to jimsonweed. *Phytoprotection* 76: 17-25.
- Abu-Seidah, A.A., 2003. Secondary metabolites as co-markers in the taxonomy of aspergilli. *Acta Microbiologica Polonica* 52: 15-23.
- Alsberg, C.I. and Black, O.F., 1913. Contribution to the study of maize deterioration; biochemical and toxicological investigations of *Penicillium puberulum* and *Penicillium stoloniferum*. USDA – Bureau Plant Industry Bulletin 270: 7-48.

Andersen, B., Smedsgaard, J. and Frisvad, J.C., 2004. *Penicillium expansum*: consistent production of patulin, chaetoglobosins, and other secondary metabolites in culture and their natural occurrence in fruit products. *Journal of Agricultural and Food Chemistry* 52: 2421-2428.

Anslow, W.K., Raistrick, H. and Smith, G., 1943. Antifungal substances from molds; patulin (anhydro-3-hydroxymethylene-tetrahydro-1:4-pyrone-2-carboxylic acid), a metabolic product of *Penicillium patulum* Bainier and *Penicillium expansum* (Link) Thom. *Journal of the Indian Chemical Society* 62: 236-238.

Aoki, T., O'Donnell, K. and Geiser, D.M., 2014. Systematics of key phytopathogenic *Fusarium* species: current status and future challenges. *Journal of General Microbiology* 80: 189-201.

Appell, M., Dombrinnk-Kurtzman, M.A. and Kendra, D.F., 2009. Comparative study of patulin, ascladiol, and neopatulin by density function theory. *Journal of Molecular Structure: THEOCHEM* 894: 23-31.

Atkinson, N., 1942. Antibacterial substances produced by moulds. I. Penicidin, a product of the growth of a *Penicillium*. *Australian Journal of Experimental Biology and Medical Science* 20: 287.

Table 4. Filamentous fungi reported as producers of moniliformin.¹

Fungal species	References
<i>Fusarium acuminatum</i>	Bosch and Mirocha, 1992; Hussein et al., 1991; Logrieco et al., 1992; Rabie et al., 1982; Schütt et al., 1998
<i>Fusarium acutatum</i>	Schütt et al., 1998
<i>Fusarium anthophilum</i>	Marasas et al., 1986; Schütt et al., 1998
<i>Fusarium arthrosporoides</i>	Schütt et al., 1998
<i>Fusarium avenaceum</i>	Bosch and Mirocha, 1992; Rabie et al., 1982; Schütt et al., 1998
<i>Fusarium begoniae</i>	Fotso et al., 2002; Schütt et al., 1998
<i>Fusarium beomiforme</i>	Marasas et al., 1991; Schütt et al., 1998
<i>Fusarium bulbicola</i>	Schütt et al., 1998
<i>Fusarium chlamydosporum</i>	Schütt et al., 1998
<i>Fusarium concentricum</i>	Schütt et al., 1998
<i>Fusarium concolor</i> ²	Rabie et al., 1982
<i>Fusarium culmorum</i> ²	Scott et al., 1987
<i>Fusarium denticulatum</i>	Fotso et al., 2002; Schütt et al., 1998
<i>Fusarium denticulatum</i>	Schütt et al., 1998
<i>Fusarium dlamini</i>	Schütt et al., 1998
<i>Fusarium equiseti</i>²	Bosch and Mirocha, 1992; Hussein et al., 1991; Rabie et al., 1982
<i>Fusarium fujikuroi</i> (Bakane strains)	Marasas et al., 1986, 1991; Schütt et al., 1998
<i>Fusarium fusaroides</i>	Rabie et al., 1978, 1982
<i>Fusarium lactis</i>	Fotso et al., 2002; Schütt et al., 1998
(<i>Fusarium moniliforme</i>) ²	Cole et al., 1973; Burmeister et al., 1979; Hussein et al., 1991; Marasas et al., 1986; Rabie et al., 1982; Tseng, 1993
<i>Fusarium moniliforme</i> var. <i>subglutinans</i> (<i>Fusarium sacchari</i> var. <i>subglutinans</i>) ² = <i>Fusarium subglutinans</i> (see this)	Bosch and Mirocha, 1992; Bosch et al., 1992; Hussein et al., 1991; Kriek et al., 1977; Lew et al., 1996; Logrieco et al., 1993; Marasas et al., 1986; Rabie et al., 1982
<i>Fusarium napiforme</i>	Marasas et al., 1991; Schütt et al., 1998
<i>Fusarium nisikadoi</i>	Fotso et al., 2002; Schütt et al., 1998
<i>Fusarium nygamai</i>	Marasas et al., 1988; 1991; Schütt et al., 1998
<i>Fusarium oxysporum</i>	Abbas et al., 1995; Hussein et al., 1991; Rabie et al., 1982; Schütt et al., 1998
<i>Fusarium phyllophilum</i>	Fotso et al., 2002; Schütt et al., 1998
<i>Fusarium proliferatum</i>	Logrieco and Bottalico, 1988; Logrieco et al., 1995; Marasas et al., 1986; Miller et al., 1995; Scarpino et al., 2015; Schütt et al., 1998
<i>Fusarium pseudoanthophilum</i>	Schütt et al., 1998
<i>Fusarium pseudocircinatum</i>	Fotso et al., 2002; Schütt et al., 1998
<i>Fusarium pseudonygamai</i>	Fotso et al., 2002; Schütt et al., 1998
<i>Fusarium ramigenum</i>	Fotso et al., 2002; Schütt et al., 1998
<i>Fusarium redolens</i>	Schütt et al., 1998
<i>Fusarium sacchari</i>	Schütt et al., 1998
<i>Fusarium semitectum</i>	Rabie et al., 1982;
<i>Fusarium sporotrichioides</i> ²	Scott et al.; 1987
<i>Fusarium subglutinans</i>	Bosch and Mirocha, 1992; Bosch et al., 1992; Hussein et al., 1991; Kriek et al., 1977; Marasas et al., 1986; Lew et al., 1996; Logrieco et al., 1993; Rabie et al., 1982; Schütt et al., 1998
<i>Fusarium temperatum</i>	Scauflaire et al., 2011; Sewram et al., 1999
<i>Fusarium thapsinum</i>	Schütt et al., 1998
<i>Fusarium tricinctum</i>	Schütt et al., 1998
<i>Penicillium melanoconidium</i>	Hallas-Møller et al., 2016

¹ Confirmed producers are in bold.² Not confirmed by Schütt et al. (1998), regarded here as the authoritative statement regarding taxonomy of the Fusaria.

- Atkinson, N., Sheppard, R.A.W., Stanley, N.F. and Rainsford, K.M., 1944. Antibacterial substances produced by moulds. 7. The activity of a further group of Australian strains of *Penicillium* and *Aspergillus*. Australian Journal of Experimental Biology and Medical Science 22: 227-230.
- Axberg, K. and Gatenbeck, S., 1975a. Intermediates in the penicillic acid biosynthesis in *Penicillium cyclopium*. Acta Chemica Scandinavica B29: 749-751.
- Axberg, K. and Gatenbeck, S., 1975b. The enzymatic formation of penicillic acid. FEBS Letters 54: 18-20.
- Bacon, C.W., Hinton, D.M. and Motchell, T.R., 2017. Is quorum signalling by mycotoxins a new risk-mitigating strategy for bacterial control of *Fusarium verticillioides* and other endophytic fungal species? Journal of Agricultural and Food Chemistry 65: 7071-7080.
- Ballester, A.R., Marcet-Houben, M., Levin, E., Sela, N., Selma-Lazaro, C., Carmona, L., Wisniewski, M., Droby, S., Gonzales-Candelas, L. and Galbadon, T., 2015. Genome, transcriptome, and functional analyses of *Penicillium expansum* provide new insights into secondary metabolism and pathogenicity. Molecular Plant-Microbe Interactions Journal 28: 232-248.
- Banani, H., Marcet-Houben, M., Ballester, A.-R., Abbruscato, P., González-Candelas, L., Gabaldón, T. and Spadero, D., 2016. Genome sequencing and secondary metabolism of the postharvest pathogen *Penicillium griseofulvum*. BMC Genomics 17: 19.
- Barad, S., Espeso, E.A., Sherman, A. and Prusky, D., 2016a. Ammonia activates *pacC* and patulin accumulation in an acidic environment during apple colonization by *Penicillium expansum*. Molecular Plant Pathology 17: 727-740.
- Barad, S., Horowitz, S.B., Kobiler, I., Sherman, A. and Prusky, D., 2014. Accumulation of the mycotoxin patulin in presence of gluconic acid contributes to pathogenicity of *Penicillium expansum*. Molecular Plant-Microbe Interactions Journal 27: 66-77.
- Barad, S., Horowitz, S.B., Moskovitch, O., Lichter A., Sherman, A. and Prusky, D., 2012. A *Penicillium expansum* glucose oxidase-encoding gene, *GOX2*, is essential for gluconic acid production and acidification during colonization of deciduous fruit. Molecular Plant-Microbe Interactions Journal 25: 779-788.
- Barad, S., Sela, N., Kumar, D., Kumar-Dubey, A., Glam-Matana, N., Sherman, A. and Prusky, D., 2016c. Fungal and host transcriptome analysis of pH-regulated genes during colonization of apple fruits by *Penicillium expansum*. BMC Genomics 17: 330.
- Barad, S., Sionov, E. and Prusky, D., 2016b. Role of patulin in post-harvest diseases. Fungal Biology Reviews 30: 24-32.
- Barta, J. and Mecir, R., 1948. Antibacterial activity of *Penicillium divergens* Bainier. Experientia 4: 277-278.
- Bemi, E., 2004. Patulin: properties, occurrence in food and regulation. A review. Industria Conserve 79: 425-440.
- Bennett, M., Gill, G.B., Pattenden, G. and Shuker, A.J., 1990. A total synthesis of neopatulin. Synlett 1990: 455-456.
- Bennett, M., Gill, G.B., Pattenden, G., Shuker, A.J. and Stapleton, A., 1991. Ylidenebutenolide mycotoxins – concise synthesis of patulin and neopatulin from carbohydrate precursors. Journal of the Chemical Society, Perkin Transactions 1: 929-937.
- Bentley, R. and Keil, J.G., 1962. Tetronic acid biosynthesis in molds. II. Formation of penicillic acid in *Penicillium cyclopium*. Journal of Biological Chemistry 237: 867-873.
- Berestetskii, O.A., Patyka, V.F. and Kalmykova, N.A. 1975. Isolation and identification of a phytotoxic substance from *Penicillium rivotii* strain 1066. Microbiologichny Zhurnal (Kiev) 37: 11-14.
- Berestetskii, O.A., Nadkernichnyi, S.P. and Patyka, V.F., 1974. Formation of patulin from *Penicillium cyaneofulvum*. Khimiya Prirodnykh Soedinenii 1974: 420-421. (Chemical Abstracts 13062r, 1975).
- Beretta, B., Gaiaschi, A., Galli, C.L. and Restani, P., 2000. Patulin in apple-based foods: occurrence and safety evaluation. Food Additives and Contaminants 17: 399-406.
- Bergel, F., Morrison, A.L., Klein, R., Moss, A.R., Rinderknecht, H. and Ward, J.L., 1943. An antibiotic substance from *Aspergillus clavatus* and *Penicillium claviforme* and its probable identity with patulin. Nature 152: 750.
- Bergel, F., Morrison, A.L., Moss, A.R. and Rinderknecht, H., 1944. An antibiotic substance from *Aspergillus clavatus*. Journal of the Chemical Society: 415-421.
- Bernhoft, A., Keblys, M., Morison, E., Larsen, H.J.S. and Flaoyen, A., 2004. Combined effects of selected *Penicillium* mycotoxins on *in vitro* proliferation of porcine lymphocytes. Mycopathologia 158: 441-450.
- Betina, V., E. Gasperíková and Nemec, P., 1969. Isolation of penicillic acid from *Penicillium simplicissimum*. Biologia (Bratislava) 24: 482-485.
- Birch, A.J., Massey-Westropp, R.A. and Moye, C.J., 1955. Studies in relation to biosynthesis. VII. 2-Hydroxy-6-methyl-benzoic acid in *Penicillium griseofulvum*. Australian Journal of Chemistry 8: 539-544.
- Birkinshaw, H., Bracken, A., Michael, S.E. and Raistrick, H., 1943. Patulin in the common cold. II. Biochemistry and chemistry. Lancet 1943: 625-631.
- Birkinshaw, J.H. and Gowlland, A., 1962. Studies in the biochemistry of microorganisms 110. Production and biosynthesis of orsellinic acid by *Penicillium madriti* G. Smith. Biochemical Journal 84: 342-347.
- Birkinshaw, J.H. and Raistrick, H., 1936. Studies in the biochemistry of microorganisms. LII. Isolation, properties and constitution of terrestriac acid (ethylcarolic acid), a metabolic product of *Penicillium terrestris* Jensen. Biochemical Journal 30: 2194-2200.
- Birkinshaw, J.H. and Samant, M.S., 1960. Studies in the biochemistry of microorganisms. 107. Metabolites of *Penicillium viridicatum* Westling: Viridicatic acid (ethylcarlosic acid). Biochemical Journal 74: 369-373.
- Birkinshaw, J.H., Oxford, A.E. and Raistrick, H., 1936. Studies in the biochemistry of microorganisms. XLVIII. Penicillic acid, a metabolic product of *Penicillium puberulum* Bainier and *Penicillium cyclopium* Westling. Biochemical Journal 30: 394-411.
- Bokhari, F.M. and Flannigan, B., 1996. Synthesis of penicillic acid and other metabolites by *Penicillium aurantiogriseum* in stored cereals and lentils. In: Sand, W. (ed.) Biodeterioration and biodegradation, Dachema Monographs 133, Dachema, Frankfurt, pp. 237-244.
- Borkowska Opacka, B. and Escoula, L., 1977. Production de la patuline en milieu liquide par des moisissures appartenant aux genres: *Aspergillus* et *Penicillium*. Annales de Recherches Veterinaires 8: 129-133.

- Bosch, U. and Mirocha, C.J., 1992. Toxin production by *Fusarium* species from sugar beets and natural occurrence of zearalenone in beets and beet fibers. Applied and Environmental Microbiology 58: 3233-3239.
- Bosch, U., Mirocha, C.J. and Wen, Y., 1992. Production of zearalenone, moniliformin and trichothecenes in intact sugar beets under laboratory conditions. Mycopathologia 119: 167-173.
- Bourdiol, D., Escoula, L. and Salvayre, R., 1990. Effect of patulin on microbial activity of mouse peritoneal macrophages. Food and Chemical Toxicology 11: 617-624.
- Boysen, M., Skouboe, P., Frisvad, J.C. and Rossen, L., 1996. Reclassification of the *Penicillium roqueforti* group into three species on the basis of molecular genetic and biochemical profiles. Microbiology-SGM 142: 541-549.
- Brase, S., Encinas, A., Keck, J. and Nising, C.F., 2009. Chemistry and biology of mycotoxins and related fungal metabolites. Chemical Reviews 109: 3903-3990.
- Bresler, G., Brizzio, S.B. and Vaamonde, G., 1995. Mycotoxin-producing potential of fungi isolated from amaranth seeds in Argentina. International Journal of Food Microbiology 25: 101-108.
- Brian, P.W., Elson, G.W. and Lowe, D., 1956. Production of patulin in apple fruits by *Penicillium expansum*. Nature 178: 263.
- Bridge, P.D., Hawksworth, D.L., Kozakiewicz, Z., Onions, A.H.S., Paterson, R.R.M., Sackin, M.J. and Sneath, P.H.A., 1989. A reappraisal of the terverticillate penicillia using biochemical, physiological and morphological features. I. Numerical taxonomy. Journal of General Microbiology 135: 2941-2966.
- Bu'Lock, J.D. and Ryan, A.J., 1958. The biogenesis of patulin. Proceedings of the Chemical Society 1958: 222-223.
- Burgess, L.W., 2014. McAlpine memorial lecture – a love affair with *Fusarium*. Australasian Plant Pathology 43: 359-368.
- Burka, L.T., Doran, J. and Wilson, B.J., 1982. Enzyme inhibition and the toxic action of moniliformin and other vinylous α -ketoacids. Biochemical Pharmacology 31: 79-84.
- Burmeister, H.R., Ciegler, A. and Vesonder, R.F., 1979. Moniliformin, a metabolite of *Fusarium moniliforme* NRRL 6322: purification and toxicity. Applied and Environmental Microbiology 37: 11-13.
- Burton, H.S. and Pausacker, B.F., 1947. Unpublished. Cited by: Abraham, E.P. and Florey, H.W. 1949. Substances produced by fungi imperfecti and ascomycetes. In: Florey, H.W., Chain, E., Heatley, N.G., Jennings, M.A., Sanders, A.G., Abraham, E.P. and Florey, M.E. (eds.) Antibiotics. Vol. 1. Oxford University Press, London, UK, pp. 273-283.
- Burton, H.S., 1949. Antibiotics from *Penicillia*. British Journal of Experimental Pathology 30: 151-158.
- Cakmakci, S., Cetin, B., Gurses, M., Dagdemir, E. and Hayaloglu, A.A., 2012. Morphological, molecular, and mycotoxicogenic identification of dominant filamentous fungi from moldy civil cheese. J Food Prot. 75: 2045-2049.
- Cakmakci, S., Gurses, M., Hayaloglu, A.A., Cetin, B., Sekerci, P. and Dagdemir, E., 2015. Mycotoxin production capability of *Penicillium roqueforti* in strains isolated from mould-ripens traditional Turkish civil cheese. Food Additives & Contaminants: Part A: Chemistry, Analysis, Control, Exposure & Risk Assessment 32: 245-249.
- Casquete, R., Rodríguez, A., Hernández, A., Martín, A., Bartolomé, T., Córdoba, J.J. and Córdoba, M.G., 2017. Occurrence of toxigenic fungi and mycotoxins during smoked paprika production. Journal of Food Protection 80: 2068-2077.
- Chain, E.H., Florey, W. and Jennings, M.A., 1942. An antibacterial substance produced by *Penicillium claviforme*. British Journal of Experimental Pathology 23: 202-205.
- Chan, P.K. and Hayes, A.W., 1981. Hepatotoxicity of the mycotoxin penicillic acid – a pharmacokinetics consideration. Journal of the American Oil Chemists' Society 58: 1017-1022.
- Chełkowski, J. and Golinski, P., 1983. Mycotoxins in cereal grain. 7. A simple method to assay mycotoxin potential of cereal grain and cereal products. Die Nahrung 27: 305-310.
- Chełkowski, J., Golinski, P. and Wiewiorowska, M., 1987. Mycotoxins in cereal grain. 12. Contamination with ochratoxin A and penicillic acid as indicator of improper storage of cereal grain. Die Nahrung 31: 81-84.
- Chełkowski, J., Trojanowska, K. and Wiewiorowska, M., 1983. Mycotoxins in cereal grain. 8. Microbiological evaluation of cereal grain quality, connected with mycotoxins occurrence. Die Nahrung 27: 311-318.
- Chen, A.J., Frisvad, J.C., Sun, B.D., Varga, J., Kocsimbé, S., Dijksterhuis, J., Kim, D.H., Hong, S.B., Houbraken, J. and Samson, R.A., 2016b. *Aspergillus* section *Nidulantes* (formerly *Emericella*). Polyphasic taxonomy, chemistry and biology. Studies in Mycology 84: 1-118.
- Chen, A.J., Hubka, V., Frisvad, J.C., Visagie, C.M., Houbraken, J., Meijer, M., Varga, J., Rasine, D., Jurjević, Ž., Kubátová, A., Sklenář, F. and Samson, R.A., 2017. Polyphasic taxonomy of *Aspergillus* section *Aspergillus* (formerly *Eurotium*) and its occurrence in indoor environment and food. Studies in Mycology 88: 37-135.
- Chen, A.J., Varga, J., Frisvad, J.C., Jiang, X.Z. and Samson, R.A., 2016a. Polyphasic taxonomy of *Aspergillus* section *Cervini*. Studies in Mycology 85: 65-89.
- Chen, L.Y., Tain, X.L. and Yang, B., 1990. A study on the inhibition of rat myocardium glutathione peroxidase and glutathione reductase by moniliformin. Mycopathologia 110: 119-124.
- Chunmei, J., Junling, S., Qi'an, H. and Yanlin, L., 2013. Occurrence of toxin-producing fungi in intact and rotten table and wine grapes and related influencing factors. Food Control 31: 5-13.
- Ciegler, A. and Kurtzman, C.P., 1970. Penicillic acid production by blue-eye fungi on various agricultural commodities. Journal of Applied Microbiology 20: 761-764.
- Ciegler, A., 1972. Bioproduction of ochratoxin A and penicillic acid by members of the *Aspergillus ochraceus* group. Canadian Journal of Microbiology 18: 631-636.
- Ciegler, A., 1977. Patulin. In: Rodricks, J.V., Hesseltine, C.W. and Mehlman, M.A. (eds.) Mycotoxins in human and animal health. Pathotox Publishers, Park Forest South, IL, USA, pp. 609-624.
- Ciegler, A., Beckwith, A.C. and Jackson, L.K., 1976. Teratogenicity of patulin and patulin adducts formed with cysteine. Applied and Environmental Microbiology 31: 644-667.
- Ciegler, A., Detroy, R.W. and Lillehoj, E.B., 1971. Patulin, penicillic acid and other carcinogenic lactones. In: Ciegler, A., Kadis, S. and Ajl, S.J. (eds.) Microbial toxins. Vol. VI. Fungal toxins. Academic Press, New York, NY, USA, pp. 404-434.

- Ciegler, A., Fennell, D.I., Sansing, G.A., Detroy, R.W. and Bennett, G.A., 1973. Mycotoxin producing strains of *Penicillium viridicatum*: Classification into subgroups. *Journal of Applied Microbiology* 26: 271-278.
- Ciegler, A., Mintzlaff, H.J., Machnick, W. and Leistner, L., 1972. Untersuchungen über das Toxinbildungsvermögen von Rohwürsten isolierter Schimmelpilze der Gattung *Penicillium*. *Fleischwirtschaft* 52: 1311-1314.
- Ciegler, A., Mintzlaff, H.-J., Weisleder, D. and Leistner, L., 1972. Potential production and detoxification of penicillic acid in mild-fermented sausage (salami). *Journal of Applied Microbiology* 24: 114-119.
- Clutterbuck, P.W., Oxford, A.E., Raistrick, H. and Smith, G., 1932. Studies in the biochemistry of microorganisms XXIV. The metabolic products of the *Penicillium brevicompactum* series. *Biochemistry Journal* 26: 1441-1458.
- Cole, R.J. and Cox, R.H., 1981. *Handbook of toxic fungal metabolites*. Academic Press, New York, NY, USA.
- Cole, R.J., Kirksey, J.W., Cutler, H.G., Doubnik, B.L. and Peckham, J.C., 1973. Toxin from *Fusarium moniliforme*: effect on plants and animals. *Science* 179: 1324-1326.
- Couch, B.C. and Kohn, L.M., 2002. A multilocus gene genealogy concordant with host preference indicates segregation of a new species, *Magnaporthe oryzae*, from *M. grisea*. *Mycologia* 94: 683-693.
- Couch, R.D. and Gaucher, G.M., 2004. Rational elimination of *Aspergillus terreus* sulochrin production. *Journal of Biotechnology* 108: 171-177.
- Da Silva, J.V., Fill, T.P., Da Silva, B.F. and Rodriguez-Fo, E., 2013. Diclavatol and tetric acids from *Penicillium griseoroseum*. *Natural Product Research* 27: 9-16.
- Delgado, S., Núñez, F., Sánchez, B., Bermúdez, E. and Rodríguez, J.M., 2011. Toxinogenic microorganisms in medicinal plants used for ritual protection of infants. *Food Research International* 44: 304-309.
- Detroy, R.W. and Still, P.E., 1975. Patulin inhibition of mycovirus replication in *Penicillium stoloniferum*. *Journal of General Microbiology* 92: 167-174.
- Devaraj, H., Suseela, R.E. and Devaraj, N., 1986. Patulin toxicosis in chicks. *Current Science* 55: 998-999.
- Dickens, F. and Jones, H.E.H., 1961. Carcinogenic activity of a series of reactive lactones and related substances. *British Journal of Cancer* 15: 85-100.
- Dickens, F. and Jones, H.E.H., 1963. Further studies on the carcinogenic and growth-inhibitory activity of a series of reactive lactones and related substances. *British Journal of Cancer* 17: 100-108.
- Dickens, F. and Jones, H.E.H., 1965. Further studies on the carcinogenic action of certain lactones and related substances in the rat and mouse. *British Journal of Cancer* 19: 392-403.
- Dimroth, P., Ringelmann, E. and Lynen, F., 1976. 6-Methylsalicylic acid synthetase from *Penicillium patulum*. *European Journal of Biochemistry* 68: 591-596.
- Dombrink-Kurtzman, M.A., 2007. The sequence of the isoepoxydon dehydrogenase genes of the patulin biosynthesis pathway in *Penicillium* species. *Antonie van Leeuwenhoek* 91: 179-189.
- Dombrink-Kurtzman, M.A. and Blackburn, J.A., 2005. Evaluation of several culture media for production of patulin by *Penicillium* species. *International Journal of Food Microbiology* 98: 241-248.
- Dombrink-Kurtzman, M.A. and McGovern, A.E., 2007. Species-specific identification of *Penicillium* linked to patulin contamination. *Journal of Food Protection* 70: 2646-2650.
- Dowd, P.F., 1989. Toxicity of naturally occurring levels of the *Penicillium* mycotoxins citrinin, ochratoxin A, and penicillic acid to the corn earworm *Heliothis zea*, and the fall armyworm, *Spodoptera frugiperda* (*Lepidoptera: Noctuidae*). *Environmental Entomology* 18: 24-29.
- Draughon, F.A. and Ayers, J.C., 1980. Insecticide inhibition of growth and patulin production in *Penicillium expansum*, *Penicillium urticae*, *Aspergillus clavatus*, *Aspergillus terreus*, and *Byssochlamys nivea*. *Journal of Agricultural and Food Chemistry* 28: 1115-1117.
- Drusch, S. and Ragab, W., 2003. Mycotoxins in fruits, fruit juices, and dried fruits. *Journal of Food Protection* 66: 1514-1527.
- Edwards, R.L., Maitland, D.J., Oliver, C.L., Pacey, M.S., Shileds, L. and Whalley, A.J.S., 1999. Metabolites of the higher fungi. Part 31. Longianone, a $C_7H_6O_4$ spiro bicyclic lactone from the fungus *Xylaria longiana*. *Journal of the Chemical Society, Perkin Transactions 1*: 715-719.
- Efimenko, O.M. and Yakimov, P.A., 1960. Antibiotic from *Penicillium cyclopium*. Trudy Leningradskii Khimiko-Farmatsevticheskii Institut, pp. 88-92; *Chemical Abstracts* 55: 21470f, 1961.
- El-Banna, A.A., Pitt, J.I. and Leistner, L., 1987. Production of mycotoxins by *Penicillium* species. System. *Journal of Applied Microbiology* 10: 42-46.
- Eldridge, J.A., Jaiswal, D.K., Jones, J.R. and Thomas, R., 1977. Tritium nuclear magnetic resonance spectroscopy. Part 7. New information of the tritium distribution in biosynthetically labelled penicillic acid. *Journal of the Chemical Society Perkin Transactions I*: 1080-1083.
- Ellis, J.R., Norstadt, F.A. and McCalla, T.M., 1977. Patulin effects wheat plants in field treatments. *Plant Soil* 47: 679-686.
- El-Samawaty, A.M.A., Abd El-Rahim, M.A., Moslem, M.A., Yassin, M.A., Sayed, S.R.M., El-Shikh, M.S., 2013. Control of grape blue molding penicillia by *Allium sativum*. *Journal of Pure and Applied Microbiology* 7: 1047-1053.
- El-Shanawany, A.A., Moustafa, M.E. and Barakat, A., 2005. Fungal populations and mycotoxins in silage in Assiut and Sohag governates in Egypt, with a special reference to characteristic *Aspergillus* toxins. *Mycopathologia* 159: 281-289.
- Engel, G. and Teuber, M., 1984. Patulin and other small lactones. In: Betina, V. (ed.) *Mycotoxins – production, isolation, separation, and purification*. Elsevier, Amsterdam, the Netherlands, pp. 291-298.
- Erdogan, A., Gurses, M. and Sert, S., 2003. Isolation of moulds capable of producing mycotoxins from blue mouldy Tulum cheeses produced in Turkey. *International Journal of Food Microbiology* 85: 83-85.
- Escoula, L., 1974. Moisissures toxinogènes des fourrages ensilés. I. Présence de patuline dans les fronts de coupe d'ensilages. *Annales de Recherches Veterinaires* 5: 423-432.
- Escoula, L., 1975. Toxinogenic moulds in silage IV. Patulin production in liquid medium using fungus species isolated from silage. *Annales de Recherches Veterinaires* 6: 303-310.

- Escoula, L., Moore, J. and Baradat, C., 1977. The toxins of *Byssochlamys nivea*. Part I. Acute toxicity of patulin in adult rats and mice. *Annales de Recherches Veterinaires* 8: 41-49.
- Escoula, L., Thomsen, M., Bourdoliol, M., Pipy, B., Peuriere, S. and Roubinet, S., 1988. Patulin immunotoxicology: effect on phagocyte activation and cellular and humoral immune system of mice and rabbits. *International Journal of Immunopharmacology* 10: 983-989.
- European Union, 2003. Commission regulation (EC) No. 1425/2003 of 11 August 2003 amending Regulation (EC) № 466/2001 as regards patulin. *Official Journal of the European Union* L 203/13.
- Ezzat, S.M., El-Sayed, A.E., Abou El-Hawa, M.I., Ismaiel, A.S., 2007. Morphological and ultrastructural studies for the biological action of penicillic acid on some bacterial species. *Research Journal of Microbiology* 2: 303-314.
- Filtenborg, O., Frisvad, J.C. and Thrane, U., 1996. Moulds in food spoilage. *International Journal of Food Microbiology* 33: 85-102.
- Florey, H.W., Chain, E., Heatley, N.G., Jennings, M.A., Sanders, A.G., Abraham, E.P. and Florey, M.E., 1949. Antibiotics. Vol. 1. Oxford University Press, Oxford, UK, pp. 273-283.
- Florey, H.W., Jennings, M.A. and Philpot, F.J., 1944. Claviformin from *Aspergillus giganteus* Wehm. *Nature* 153: 139.
- Forrester, P.I. and Gaucher, G.M., 1972a. Conversion of 6-methylsalicylic acid into patulin by *Penicillium urticae*. *Biochemistry* 11: 1102-1107.
- Forrester, P.I. and Gaucher, G.M., 1972b. m-Hydroxybenzyl alcohol dehydrogenase from *Penicillium urticae*. *Biochemistry* 11: 1108-1114.
- Fotso, J., Leslie, J.F. and Smith, J.S., 2002. Production of beauvericin, moniliformin, fusaproliferin, and fumonisin B₁, B₂, and B₃ by fifteen ex-type strains of *Fusarium* species. *Applied and Environmental Microbiology* 68: 5195-5197.
- Franck, B. and Breipohl, G., 1984. Biosynthesis of moniliformin, a fungal toxin with cyclobutanedione structure. *Angewandte Chemie International Edition* 23: 996-998.
- Frank, H.K., 1972. Das Mykotoxinprodukt bei Lebensmitteln und Getränken. *Zentralblatt für Bakteriologie und Hygiene*. 1. Abt. Originale B 159: 324-334.
- Frank, H.K., 1977. Occurrence of patulin in fruits and vegetables. *Annales de la Nutrition et de l'Alimentation* 31: 459-465.
- Frisvad, J.C., 1981. Physiological criteria and mycotoxin production as aids in identification of common asymmetric Penicillia. *Applied and Environmental Microbiology* 41: 568-579.
- Frisvad, J.C., 1989. The connection between the penicillia and aspergilli and mycotoxins with special emphasis on misidentified isolates. *Archives of Environmental Contamination and Toxicology* 18: 452-467.
- Frisvad, J.C., 2015. Taxonomy, chemodiversity, and chemoconsistency of *Aspergillus*, *Penicillium*, and *Talaromyces* species. *Frontiers in Microbiology* 5: 773.
- Frisvad, J.C. and Filtenborg, O., 1983. Classification of terverticillate penicillia based on profiles of mycotoxins and other secondary metabolites. *Applied and Environmental Microbiology* 46: 1301-1310.
- Frisvad, J.C. and Filtenborg, O., 1989. Terverticillate Penicillia: chemotaxonomy and mycotoxin production. *Mycologia* 81: 837-861.
- Frisvad, J.C. and Filtenborg, O., 1990. Revision of *Penicillium* subgenus *Furcatum* based on secondary metabolites and conventional characters. In: Samson, R.A. and Pitt, J.I. (eds.) *Modern concepts in Penicillium and Aspergillus classification*. Plenum Press, New York, NY, USA, pp. 159-170.
- Frisvad, J.C. and Larsen, T.O., 2015. Chemodiversity in the genus *Aspergillus*. *Journal of Applied Microbiology Biotechnology* 99: 7859-7877.
- Frisvad, J.C. and Samson, R.A., 2000. *Neopetromyces* gen nov. and an overview of teleomorphs of *Aspergillus* subg. *Circumdati*. *Studies in Mycology* 45: 201-207.
- Frisvad, J.C. and Samson, R.A., 2004. Polyphasic taxonomy of *Penicillium* subgenus *Penicillium*. A guide to identification of the food and air-borne terverticillate Penicillia and their mycotoxins. *Studies in Mycology* 49: 1-173.
- Frisvad, J.C., Filtenborg, O. and Wicklow, D.T., 1987. Terverticillate Penicillia isolated from underground seed caches and cheek pouches of banner-tailed kangaroo rats (*Dipodomys spectabilis*). *Canadian Journal of Botany* 65: 765-773.
- Frisvad, J.C., Frank, J.M., Houbraken, J.A.M.P., Kuijpers, A.F.A. and Samson, R.A., 2004a. New ochratoxin A producing species in *Aspergillus* section *Circumdati*. *Studies in Mycology* 50: 23-43.
- Frisvad, J.C., Seifert, K.A., Samson, R.A. and Mills, J.T., 1994. *Penicillium tricolor*, a new mold species from Canadian wheat. *Canadian Journal of Botany* 72: 933-939.
- Frisvad, J.C., Smedsgaard, J., Larsen, T.O. and Samson, R.A., 2004. Mycotoxins, drugs and other extrolites produced by species in *Penicillium* subgenus *Penicillium*. *Studies in Mycology* 49: 201-241.
- Frisvad, J.C., Smedsgaard, J., Larsen, T.O. and Samson, R.A., 2004b. Mycotoxins, drugs and other extrolites produced by species in *Penicillium* subgenus *Penicillium*. *Studies in Mycology* 49: 201-242.
- Fung, F. and Clark, R.F., 2004. Health effects of mycotoxins: a toxicological overview. *Journal of Toxicology – Clinical Toxicology* 42: 217-234.
- Gallo, A., Giuberti, G., Frisvad, J.C., Bertuzzi, T. and Nielsen, K.F., 2015. Review on mycotoxin issues in ruminants: occurrence in forages, effects of mycotoxin ingestion on health status and animal performance and practical strategies to counteract their negative effect. *Toxins* 7: 3057-3111.
- Garza, S., Canela, R., Vinas, I. and Sanchis, V., 1993. Effects of potassium sorbate on growth and penicillic acid production by *Aspergillus ochraceus* and *Penicillium aurantiogriseum*. *Zentralblatt für Mikrobiologie* 148: 343-350.
- Gathercole, P.S., Thiel, P.G. and Hofmeyr, J.H.S., 1986. Inhibition of pyruvate dehydrogenase complex by moniliformin. *Biochemistry Journal* 233: 719-723.
- Gaucher, G.M., 1979. Mycotoxins – their biosynthesis in fungi: patulin and related carcinogenic lactones. *Journal of Food Protection* 42: 810-814.
- Gedek, B., Bauer, J. and Schreiber, H., 1981. Mycotoxin production of silage deteriorating molds. *Wiener Tierärztliche Monatsschrift* 68: 299-301.

- Geiser, D.M., Aoki, T., Bacon, C.W., Baker, S.E., Bhattacharyya, M.K., Brandt, M.E., Brown, D.W., Burgess, L.W., Chulze, S., Coleman, J.J., Correll, J.C., Covert, S.F., Crous, P.W., Cuomo, C.A., De Hoog, G.S., Di Pietro, A., Elmer, W.H., Epstein, L., Frandsen, R.J.N., Freeman, S., Gagkaeva, T., Glenn, A.E., Gordon, T.R., Gregory, N.E., Hammond-Kosack, K.E., Hanson, L.E., Del Mar Jimenez-Gasco, M., Kang, S., Kistler, H.C., Kulda, G.A., Leslie, J.F., Logrieco, A., Lu, G., Lysøe, E., Ma, L.-J., McCormick, S.P., Michel, Q., Moretti, A., Munaut, F., O'Donnell, K., Pfennig, L., Ploetz, R.C., Proctor, R.H., Rehner, S.A., Robert, V.A.R.G., Rooney, A.P., Bin Salleh, B., Scandiani, M.M., Scauflaire, J., Short, D.P.G., Steenkamp, E., Suga, H., Summerell, B.A., Sutton, D.A., Thrane, U., Trail, F., Van Diepeningen, A., VanEtten, H.D., Viljoen, A., Waalwijk, C., Ward, T.J., Wingfield, M.J., Xu, J.-R., Yang, X.-B., Yli-Mattila, T. and Zhang, N., 2013. One fungus, one name: defining the genus *Fusarium* in a scientifically robust way that preserves longstanding use. *Phytopathology* 103: 400-408.
- Geiger, M., Guitton, Y., Vansteelandt, M., Kerzaon, I., Blanchet, E., Robiou du Pont, T., Frisvad, J., Hess, P., Pouchus, Y.F. and Grovel, O., 2013. Cytotoxicity and mycotoxin production of shellfish-derived *Penicillium* spp., a risk for shellfish consumers. *Letters in Applied Microbiology* 57: 385-392.
- Geiger, W.B. and Conn, J.E., 1945. The mechanism of the antibiotic action of clavacin and penicillic acid. *Journal of the American Chemical Society* 67: 112-116.
- Gerlach, W. and Nirenberg, H.I., 1982. The genus *Fusarium* – a pictorial atlas. Biologische Bundesanstalt für Land- und Forstwirtschaft. Paul Parey, Berlin-Dahlem, Germany.
- Ghadevaru, G., 2013. *In vitro* evaluation of toxin binding efficacy for aflatoxin B1, ochratoxin A, citrinin, zearalenone, T2 toxin, penicillic acid and fumonisin in broiler chickens. *Journal of Veterinary Pharmacology and Therapeutics* 35, Suppl. 3: 50.
- Ghisalberti, E.L., Hargraves, J.R., Skelton, B.W. and White, A.H., 2000. New patulin derivatives. *Australian Journal of Chemistry* 53: 995-997.
- Giarman, N.J., 1948. Antibiotic lactones and synthetic analogs. 1. Cardiotoxin effects on the isolated frog heart. *J. Pharmacol. Exp. Therap.* 94: 232-243.
- Gill-Carey, D.E., 1949. The nature of some antibiotics from aspergilli. *British Journal of Experimental Pathology* 30: 119-123.
- Gilliver, K., 1946. The inhibitory action of antibiotics on plant pathogenic bacteria and fungi. *Annals of Botany* 10: 271-282.
- Giridhar, P. and Reddy, S.M., 1998. The effect of polyamine biosynthesis inhibitors on mycelial growth and mycotoxin production by *Aspergillus terreus*. *National Academy Science Letters (India)* 21: 5-8.
- Girisham, S. and Reddy, S.M., 1986a. Interaction of different seed-borne fungi of pearl millet (*Pennisetum americanum*) and its effect on patulin production by *Aspergillus terreus*. *Current Science* 55: 730-732.
- Girisham, S. and Reddy, S.M., 1986b. Efficacy of volatile compounds and food preservatives in the control of growth and patulin production by *Aspergillus terreus*, *National Academy Science Letters (India)* 9: 373-374.
- Gorbatch, G. and Friederick, W., 1949. Beiträge zur Kenntnis der Penicillinsäure. *Oesterreichische Chemiker Zeitung* 50: 93-97.
- Goss, R.J.M., Fuchser, J. and O'Hagan, D., 1999. Biosynthesis of longianone from *Xylaria longiana*: a metabolite with a biosynthetic relationship to patulin. *Chemical Communications* 1999: 2255-2256.
- Grabsch, C., Wichmann, G., Loffhagen, N., Herbarth, O. and Müller, A., 2006. Cytotoxicity assessment of gliotoxin and penicillic acid in *Tetrahymena pyriformis*. *Environmental Toxicology* 21: 111-117.
- Gruber-Dorninger, C., Novak, B., Nagl, V. and Berthiller, F., 2017. Emerging mycotoxins: beyond traditionally determined food contaminants. *Journal of Agricultural and Food Chemistry* 65: 7052-7070.
- Gutarowska, B., Sulyok, M. and Kraska, R., 2010. A study of the toxicity of moulds isolated from dwellings. *Indoor and Built Environment* 19: 668-675.
- Gye, W.E., 1943. Patulin in the common cold. III. Preliminary trial in the common cold. *Lancet* 1943: 630-631.
- Hallas-Møller, M., Nielsen, K.F. and Frisvad, J.C., 2016. Production of the *Fusarium* mycotoxin moniliformin by *Penicillium melanoconidium*. *Journal of Agricultural and Food Chemistry* 64: 4505-4510.
- Harwig, J., Chen, Y.-K., Kennedy, B.P.C. and Scott, P.M., 1973. Occurrence of patulin and patulin-producing strains of *Penicillium expansum* in natural rots of apple in Canada. *Canadian Institute of Food Science and Technology Journal* 6: 22-25.
- Hayes, A.W., Phillips, T.D., Williams, W.L. and Ciegler, A., 1979. Acute toxicity of patulin in mice and rats. *Toxicology* 13: 91-100.
- He, J., Wijeratne, E.M.K., Bashyal, B.P., Zhan, J., Seliga, C.J., Liu, M.X., Pierson, E.E., Pierson III, L.S., VenEtten, H.D. and Gunatilaka, A.A.L., 2004. Cytotoxic and other metabolites of *Aspergillus* inhabiting the rhizosphere of Sonoran desert plants. *Journal of Natural Products* 67: 1985-1991.
- Herrare, M., Van Dam, R., Spanjer, M., De Stoppelaar, J., Mol, H., De Nijs, M. and López, P., 2017. Survey of moniliformin in wheat- and corn-based products using a straightforward analytical method. *Mycotoxin Research* 33: 333-341.
- Herrick, J.A., 1945. Antifungal properties of clavacin. *Proceedings of the Society for Experimental Biology and Medicine* 59: 41-42.
- Hopkins, J., 1993. The toxicological hazards of patulin. *Food and Chemical Toxicology* 31: 455-459.
- Houbraek, J., Frisvad, J.C. and Samson, R.A., 2010. Sex in *Penicillium* series *Roqueforti*. *IMA Fungus* 1: 171-180.
- Houbraek, J., López, C., Frisvad, J.C., Boekhout, T., Theelen, B., Molano, A.E.F. and Samson, R.A., 2011. Five new *Penicillium* species, *P. aracuaraense*, *P. elleniae*, *P. penarojaense*, *P. vanderhammenii* and *P. wotroi*, from Colombian leaf litter. *International Journal of Systematic and Evolutionary Microbiology* 61: 1462-1475.
- Houbraek, J., Wang, L., Lee, H.B. and Frisvad, J.C., 2016. New sections in *Penicillium* containing novel species producing patulin, pyripyropes or other bioactive compounds. *Persoonia* 36: 299-314.
- Hubka, V., Nováková, A., Kolarik, M., Jurjevics, Z. and Peterson, S.W., 2015. Revision of *Aspergillus* section *Flavipedes*: seven new species and proposal of section *Jani* sect. nov. *Mycologia* 107: 169-208.
- Hubka, V., Nováková, A., Peterson, S.W., Frisvad, J.C., Sklenář, F., Matsusawa, T., Kubátová, A. and Kolařík, M., 2016. A reappraisal of *Aspergillus* section *Nidulantes* with descriptions of two new sterigmatocystin-producing species. *Plant Systematics and Evolution* 302: 1267-1299.

- Hussein, H.M., Baxter, M., Andrew, I.G. and Franich, R.A., 1991. Mycotoxin production by *Fusarium* species isolated from New Zealand maize fields. *Mycopathologia* 113: 35-40.
- Ismaiel, A.A. and Papenbrock, J., 2015. Mycotoxins: producing fungi and mechanisms of phytotoxicity. *Agriculture* 5: 492-537.
- Ismaiel, A.A., Bassyouni, R.H., Makel, Z. and Gabr, S.M., 2016. Detoxification of patulin by Kombucha tea culture. *CyTA – Journal of Food* 14: 271-279.
- Jackson, L. and Dombrinck-Kurtzman, M.A., 2006. Patulin. In: Sapers, G.M., Gorny, J.R. and Yousef, A.E. (eds.) *Microbiology of fruits and vegetables*. CRC Press, Boca Raton, FL, USA, pp. 281-311.
- Jímenez, M., Mateo, R., Querol, A., Huerta, T. and Hernández, E., 1991. Mycotoxins and mycotoxicogenic moulds in nuts and sunflower seeds for human consumption. *Mycopathologia* 115: 121-127.
- Jonsson, M., Atosuo, J., Jestoi, M., Nathanail, A.V., Kokkonen, U.-M., Anttila, M., Koivisto, P., Lilius, E.-M. and Peltonern, K., 2015. Repeated doses 28-day oral toxicity study of moniliformin in rats. *Toxicology Letters* 233: 38-44.
- Jonsson, M., Jestoi, M., Nathanail, A.V., Kokkonen, U.-M., Anttila, M., Koivisto, P., Karhunen, P. and Peltonen, K., 2013. Application of OECD guideline 423 in assessing the acute oral toxicity of moniliformin. *Food and Chemical Toxicology* 53: 27-32.
- Julca, I., Drobys, S., Sela, N., Marcat-Houben, M. and Gabaldón, T., 2015. Contrasting genomic diversity in two closely related postharvest pathogens: *Penicillium digitatum* and *Penicillium expansum*. *Genome Biology and Evolution* 8: 218-227.
- Jurjevic, Z., Kubátová, A., Lolarik, M. and Hubka, V., 2015. Taxonomy of *Aspergillus* section *Petersonii* sect. nov. encompassing indoor and soil-borne species with predominant tropical distribution. *Plant Systematics and Evolution* 301: 2441-2462.
- Kamyar, M.R., Kouri, K., Rawnduzi, P., Studenic, C. and Lemmens-Gruber, R., 2006. Effects of moniliformin in presence of cyclohexadepsipeptides on isolated mammalian tissue and cells. *Toxicology in Vitro* 20: 1284-1291.
- Kang, S.W. and Kim, S.W., 2004. New antifungal activity of penicillic acid against *Phytophthora* species. *Biotechnology Letters* 26: 695-698.
- Kang, S.W., Park, C.H., Hong, S.I. and Kim, S.W., 2007. Production of penicillic acid by *Aspergillus sclerotiorum* CGF. *Bioresource Technology* 98: 191-197.
- Karow, E.O. and Forster, J.W., 1944. An antibiotic substance from species of *Gymnoascus* and *Penicillium*. *Science* 99: 265-266.
- Karow, E.O., Woodruff, H.B. and Forster, J.W., 1944. Penicillic acid from *Aspergillus ochraceus*, *Penicillium thomi* and *Penicillium suavolens*. *Archives of Biochemistry* 5: 279-282.
- Kent, J. and Heatley, N.G., 1945. Antibiotics from moulds. *Nature* 156: 295-296.
- Kerromnes, J. and Thouvenot, D., 1985. Role of penicillic acid in the phytotoxicity of *Penicillium cyclopium* and *Penicillium canescens* to germination of corn seeds. *Applied and Environmental Microbiology* 49: 660-663.
- Khaddor, M., Saidi, R., Aidoun, A., Lamarti, A., Tantaoui-Eleraki, A., Ezziyyani, M., Castillo, M.-E.C. and Badoc, A., 2007. Antibacterial effects and toxigenesis of *Penicillium aurantiogriseum* and *P. viridicatum*. *African Journal of Biotechnology* 6: 2314-2318.
- Kharchenko, S.M., 1970. Identification of antibiotics from *Penicillia* active against agents of mottled leaves and bunt fungi. *Microbiologichny Zhurnal (Kiev)* 32: 115-119.
- Kildgaard, S., Mansson, M., Dosen, I., Klitgaard, A., Frisvad, J.C., Larsen, T.O. and Nielsen, K.F., 2014. Accurate dereplication of bioactive secondary metabolites from marine-derived fungi by UHPLC-DAD-QTOFMS and MS/HRMS library. *Marine Drugs* 12: 3681-3705.
- Kimura, Y., Nakahara, S. and Fujioka, S., 1996. Aspyrone, a nematicidal compound isolated from the fungus *Aspergillus melleus*. *Bioscience, Biotechnology, and Biochemistry* 60: 1375-1376.
- Kis, Z., Furger, P. and Sigg, H.P., 1969. Über die Isolierung von Pyrenophenol. *Experientia* 25: 123.
- Klarić, M.S., Rašić, D. and Peraica, M., 2013. deleterious effects of mycotoxin combinations involving ochratoxin A. *Toxins* 5: 1965-1987.
- Kobayashi, H., Tsunoda, H. and Tatsuno, T., 1971. Recherches toxicologique sur les mycotoxines qui polluent de fourrages artificiel du porc. *Chemical and Pharmaceutical Bulletin* 19: 829-842.
- Kocsimbé, S., Perrone, G., Magistà, D., Houbraken, J., Varga, J., Szigeti, G., Hubka, V., Hong, S.-B., Frisvad, J.C. and Samson, R.A., 2016. *Aspergillus* is monophyletic: evidence from multiple gene phylogenies and extrolite profiles. *Studies in Mycology* 85: 199-213.
- Komagata, D., Fujita, S., Yamashita, N., Saito, S. and Morino, T., 1996. Novel neuritogenic activities of pseurotin A and penicillic acid. *Journal of Antibiotics* 49: 958-959.
- Korzybski, T., Kowszyk-Gindifer, Z. and Kuryłowicz, W., 1967. *Antibiotics. Origin, nature and properties*. Pergamon Press, Oxford, UK.
- Koteswara Rao, V., Shilpa, P., Girisham, S. and Reddy, S.M., 2011. Incidence of mycotoxicogenic penicillia in feeds of Andhra Pradesh, India. *International Journal of Biotechnology and Molecular Biology Research* 2: 46-50.
- Kovalsky, P., Kos, G., Nahrer, K., Schwab, C., Jenkins, T., Schatzmayer, G., Solyok, M. and Krská, R., 2016. Co-occurrence of regulated, masked and emerging mycotoxins and secondary metabolites in finished feed and maize – an extensive survey. *Toxins* 8: 12.
- Kriek, N.P.J., Marasas, W.F.O., Steyn, P.S., Van Rensburg, S.J. and Steyn, M., 1977. Toxicity of moniliformin-producing strain of *Fusarium moniliforme* var. *subglutinans* isolated from maize. *Food and Cosmetics Toxicology* 15: 579-587.
- Kubena, L.F., Phillips, T.D., Witzel, D.A. and Heidelbaugh, N.D., 1984. Toxicity of ochratoxin A and penicillic acid to chicks. *Bulletin of Environmental Contamination and Toxicology* 32: 711-716.
- Kuehn, H.H., 1958. A preliminary survey of the *Gymnoascaceae*. I. *Mycologia* 50: 417-439.
- Kumar, C.G., Mongolla, P., Pombala, S., Kamle, A. and Joseph, J., 2011. Physicochemical characterization and antioxidant activity of melanin from a novel strain of *Aspergillus bridgeri* ICTF-201. *Journal of Applied Microbiology* 53: 350-358.
- Kurtzman, C.P. and Ciegler, A., 1970. Mycotoxin from a blue-eyed mold of corn. *Journal of Applied Microbiology* 20: 204-207.
- Laidou, I.A., Thanassoulopoulos, C.C. and Liakopoulou-Kyriakidis, M., 2001. Diffusion of patulin in the flesh of pears inoculated with four post-harvest pathogens. *Journal of Phytopathology* 149: 457-461.

- Larsen, J. and Olson, L.W., 1992. Mode of action of α,β unsaturated carbonyl-compounds. *Journal of Phytopathology* 135: 1-5.
- Larsen, T.O., Frisvad, J.C., Ravn, G. and Skaanning, T., 1998. Mycotoxin production by *Penicillium expansum* on black currant and cherry juice. *Food Additives and Contaminants* 15: 671-675.
- Lebed, E.S., Novikova, S.D. and Kuznetsov, V.D., 1978. Production of penicillic and 6-amino penicillanic acid by some *Trichoderma* species. *Mikol Fitopatol* 12: 103-106.
- Lei, H.Y., He, Z.P., Yuan, H., Wu, J., Wen, L.X., Li, R.F., Zhang, M., Yuan, L.Y. and Yuan, Z.H., 2010. Generation and characterization of a monoclonal antibody to penicillic acid from *Penicillium cyclopium*. *African Journal of Biotechnology* 9: 3026-3031.
- Leistner, L. and Eckardt, C., 1979. Vorkommen toxinogener Penicillien bei Fleischerzeugnisse. *Fleischwirtschaft* 59: 1892-1896.
- Leistner, L. and Pitt, J.I., 1977. Miscellaneous *Penicillium* toxins. In: Rodricks, J.V., Hesseltine, C.W. and Mehlman, M.A. (eds.) *Mycotoxins in human and animal health*. Chem-Orbital, pp. 639-653.
- Lesley J. and Summerell, B.A., 2016. The *Fusarium* laboratory manual. Blackwell Publishing, Ames, IA, USA.
- Lew, H., Chelkowski, J., Pronczuk, P. and Edinger, W., 1996. Occurrence of the mycotoxin moniliformin in maize (*Zea mays* L) ears infected by *Fusarium subglutinans* (Wollenw. and Reinking) Nelson et al. *Food Additives & Contaminants* 13: 321-324.
- Li, B., Zong, Y., Du, Z., Zhang, Z., Qin, G., Zhao, W. and Tian, S., 2015. Genomic characterization reveals insights into patulin biosynthesis and pathogenicity in *Penicillium* species. *Molecular Plant-Microbe Interactions* 28: 635-647.
- Li, H.J., Cai, Y.T., Chen, Y.Y., Lam, C.K. and Lan, W.J., 2010. Metabolites of marine fungus *Aspergillus* sp. collected from soft coral *Sarcophyton tortuosum*. *Chemical Research in Chinese Universities* 26: 415-419.
- Lillehoj, E.B., Milburn, M.S. and Ciegler, A., 1972. Control of *Penicillium martensii* development and penicillic acid production by atmospheric gases and temperatures. *Applied Microbiology* 24: 198-201.
- Lindenfelser, L.A. and Ciegler, A., 1977. Penicillic acid production in submerged culture. *Applied and Environmental Microbiology* 34: 553-556.
- Logrieco, A. and Bottalico, A., 1988. *Fusarium* species of the *Liseola* section associated with stalk and ear rot of maize in southern Italy, and their ability to produce moniliformin. *Transactions of the British Mycological Society* 90: 215-219.
- Logrieco, A., Altomare, C., Moretti, A. and Bottalico, A., 1992. Cultural and toxicogenic variability in *Fusarium acuminatum*. *Mycological Research* 96: 518-523.
- Logrieco, A., Moretti, A., Altomare, C., Bottalico, A. and Carbonell Torres, E., 1993. Occurrence and toxicity of *Fusarium subglutinans* from peruvian maize. *Mycopathologia* 122: 185-190.
- Logrieco, A., Moretti, A., Ritieni, A., Bottalico, A. and Corda, P., 1995. Occurrence and toxicogenicity of *Fusarium proliferatum* from preharvest maize ear rot, and associated mycotoxins, in Italy. *Plant Disease* 79: 727-731.
- Lopez-Diaz, T.M. and Flannigan, B., 1997. Production of patulin and cytochalasin E by 4 strains of *Aspergillus clavatus* during malting of barley and wheat. *International Journal of Food Microbiology* 35: 129-136.
- Lund, F. and Frisvad, J.C., 1994. Chemotaxonomy of *Penicillium aurantiogriseum* and related species. *Mycological Research* 98: 481-492.
- Luque, M.I., Rodríguez, A., Casado, E., Asensio, M.A. and Córdoba, J.J., 2011. Development of a PCR protocol to detect patulin producing moulds in food products. *Food Control* 22: 1831-1838.
- Lykakis, I.N., Zaravinos, I.-P., Raptis, C. and Stratakis, M., 2009. Divergent synthesis of the co-isolated mycotoxins longianone, isopatulin, and (Z)-ascladiol via furan oxidation. *Journal of Organic Chemistry* 74: 6339-6342.
- MacGeorge, K.M. and Mantle, P.G., 1990. Nephrotoxicity of *Penicillium aurantiogriseum* and *P. commune* from an endemic nephropathy area of Yugoslavia. *Mycopathologia* 112: 139-145.
- Maidana, L., Gerez, J.R., El Khoury, R., Pinho, F., Puel, O., Oswald, I.P., Bracarense, A.P.F.R.L., 2016. Effects of patulin and ascladiol on porcine intestinal mucosa: an *ex vivo* approach. *Food and Chemical Toxicology* 98: 189-194.
- Malekinejad, H., Aghazadeh-Attari, J., Rezabakhsh, A., Sattari, M. and Ghasemoltani, B., 2015. Neurotoxicity of mycotoxins produced *in vitro* by *Penicillium roqueforti* isolated from maize and grass silage. *Human & Experimental Toxicology* 34: 997-1005.
- Mansouri, S., Houbraken, J., Samson, R.A., Frisvad, J.C., Christensen, M., Tuthill, D., Koutaniemi, S., Hatakka, A. and Lankinen, P., 2013. A new *Penicillium* species efficiently producing inulinase. *Antonie van Leeuwenhoek* 103: 1343-1357.
- Mantle, P.G., 1994. Renal histopathological responses to nephrotoxic *Penicillium aurantiogriseum* in the rat during pregnancy, lactation and after weaning. *Nephron* 66: 903-998.
- Mantle, P.G., McHugh, K.M. and Fincham, J.E., 2010. Contrasting nephropathic responses to oral administration of extract of cultured *Penicillium polonicum* in rat and primate. *Toxins* 2: 2083-2097.
- Marasas, W.F.O., Nelson, P.E. and Toussoun, T.A., 1988. Reclassification of two important moniliformin-producing strains of *Fusarium*, NRRL 6022 and NRRL 6322. *Mycologia* 80: 407-410.
- Marasas, W.F.O., Thiel, P.G., Rabie, C.J., Nelson, P.E. and Toussoun, T.A., 1986. Moniliformin production in *Fusarium* section *Liseola*. *Mycologia* 78: 242-247.
- Marasas, W.F.O., Thiel, P.G., Sydenham, E.W., Rabie, C.J., Lübben, A. and Nelson, P.E., 1991. Toxicity and moniliformin production by four recently described species of *Fusarium* and two uncertain taxa. *Mycopathologia* 113: 191-197.
- Martínez-Luis, S., González, M.C., Ulloa, M. and Mata, R., 2005. Phytotoxins from the fungus *Malbranchea aurantiaca*. *Phytochemistry* 66: 1012-1016.
- Mayer, V.W. and Legator, M.S., 1969. Production of petite mutants of *Saccharomyces cerevisiae* by patulin. *Journal of Agricultural and Food Chemistry* 17: 454-456.
- McKinley, E.R. and Carlton, W.W., 1980a. Patulin mycotoxicosis in the Syrian Hamster. *Food and Cosmetics Toxicology* 18: 173-179.
- McKinley, E.R. and Carlton, W.W., 1980b. Patulin mycotoxicosis in Swiss ICR mice. *Food and Cosmetics Toxicology* 18: 181-187.

- McKinley, E.R. and Carlton, W.W., 1991. Patulin. In: Sharma, R.P. and Salunkhe, D.K. (eds.) Mycotoxins and phytoalexins. CRC Press, Boca Raton, FL, USA, pp. 191-236.
- McKinley, E.R., Carlton, W.W. and Boon, G.D., 1982. Patulin mycotoxicosis in the rat: toxicology, pathology and clinical pathology. Food and Cosmetics Toxicology 20: 289-300.
- Micco, C., Miraglia, M., Onori, R., Libanori, A., Brera, C., Montaovani, A. and Macri, C., 1989. Long-term administration of low doses of mycotoxins in poultry – synergistic effects of ochratoxin A and penicillic acid on residues and toxicity in broilers. In: Institut National de Recherche Agronomique (ed.) Agriculture, food chemistry and the consumer, vols 1 and 2: the consumer and food, analytical food chemistry and agricultural production, biotechnology and bioassays – impact of technology on food quality, food chemistry and quality assurance. 5th European Conference on Food Chemistry: agriculture, food chemistry and the consumer. 27-29 September, 1989. Versailles, France, pp. 121-125.
- Mikami, A., Okazawa, T., Sakai, N., Hanada, K. and Mizoue, K., 1996. A new derivative patulin produced by *Penicillium vulpinum* F-4148. Taxonomy, isolation, physico-chemical properties, structure and biological properties. Journal of Antibiotics 49: 985-989.
- Miljkovic, A., Pfohl-Leszkowicz, A., Drobota, M. and Mantle, P.G., 2003. Comparative responses to mode of oral administration and dos of ochratoxin A or nephrotoxic extract of *Penicillium polonicum* in rats. Experimental and Toxicologic Pathology 54: 305-312.
- Miller, J.D., Savard, M.E., Schaafma, A.E., Seifert, K.A. and Reid, L.M., 1995. Mycotoxin prioduction of *Fusarium moniliforme* and *Fusarium proliferatum* from Ontario and occurrence of fumonisins in the 1993 corn crop. Canadian Journal of Plant Pathology 17: 233-239.
- Mills, J.T., Frisvad, J.C., Seifert, K.A. and Abramson, D., 1995a. Identification of nephrotoxic *Penicillium* species from cereal grains. Mycotoxin Research 11: 25-36.
- Mills, J.T., Seifert, K.A., Frisvad, J.C. and Abramson, D., 1995b. Nephrotoxicogenic *Penicillium* species occurring on farm-stored cereal grains in Western Canada. Mycopathologia 130: 23-28.
- Mintzlaff, H.J., Ciegler, A. and Leistner, L., 1972. Potential mycotoxin problems in mould fermented sausages. Zeitschrift für Lebensmittel-Untersuchung und -Forschung 150: 133-137.
- Mioso, R., Marante, F.T. and De Laguna, I.H.B., 2015. *Penicillium roqueforti*: a multifunctional cell factory of high value-added molecules. Journal of Applied Microbiology 118: 781-791.
- Moake, M.M., Padilla-Zakour, O.I. and Worobo, R.W., 2005. Comprehensive review of patulin control methods in foods. Comprehensive Reviews in Food Science and Food Safety 4: 8-21.
- Mohan, L., Collins, G., Maher, S., Walsch, E.G. Winter, D.C., O'Brian, P.J., Brayden, D.J. and Baird, A.W., 2012. The mycotoxin patulin increases colonic epithelial permeability *in vitro*. Food and Chemical Toxicology 50: 4097-4102.
- Montenegro, T.G.C., Rodrigues, F.A.R., Jimenez, P.C., Angelim, A.L., Melo, V.M.M., Filho, E.R., De Oliveira, M. and Costa-Lotufo, L.V., 2012. Cytotoxic activity of fungal strains isolated from the Ascidian *Eudistoma vannamei*. Chemistry & Biodiversity 9: 2203-2209.
- Morgavi, D.P., Boudra, H., Jouany, J.P. and Graviou, D., 2003. Prevention of patulin toxicity on rumen microbial fermentation by SH-containing reducing agents. Journal of Agricultural and Food Chemistry 51: 6906-6910.
- Mosbach, K., 1960. Die Biosynthese der Orsellinsäure und Penicillinsäure (1). Acta Chemica Scandinavica 14: 457-464.
- Moslem, M., Abd-Elsalam, K., Yassin, M. and Bahkali, A., 2010. First morphomolecular identification of *Penicillium griseofulvum* and *Penicillium aurantiogriseum* toxicogenic isolates associated with blue mold on apple. Foodborne Pathogens and Disease 7: 857-861.
- Moslem, M.A., Yassin, M.A., El-Samawaty, A.M.A., Abd El-Rahim, M.A., Sayed, S.R.M. and Amer, O.E., 2013. Mycotoxin-producing *Penicillium* species involved in apple blue mold rot. Journal of Pure and Applied Microbiology 7: 187-193.
- Moslem, M.A., Yassin, M.A., El-Samawaty, A.R.M.A. and Sayed, S.R.M., 2011. New toxicogenic *Penicillium* species associated with apple blue mold in Saudi Arabia. Fresenius Environmental Bulletin 20: 3194-3198.
- Moubasher, A.H., Abdel-Kader, M.I.A. and El-Kady, I.A., 1978. Toxigenic fungi isolated from Roquefort cheese. Mycopathologia 66: 187-190.
- Müller, H.M. and Amend, R., 1997. Formation and disappearance of mycophenolic acid, patulin, penicillic acid and PR toxin in maize silage inoculated with *Penicillium roqueforti*. Archives of Animal Nutrition 50: 213-225.
- Murnaghan, M.F., 1946. The pharmacology of penicillic acid. Journal of Pharmacology and Experimental Therapeutics 88: 119-132.
- Myrchink, T.G., 1967. Production of patulin by a group of fungi *Penicillium lapidosum* Raper & Fennell. Antibiotiki 12: 762-766.
- Namikoshi, M., Negishi, R., Nagai, H., Dmitrenok, A. and Kobayashi, H., 2003. Three new chlorine containing antibiotics from a marine-derived fungus *Aspergillus ostianus* collected in Pohnpei. Journal of Antibiotics 56: 755-761.
- Nasser, L.A., 2008. Mycoflora, mycotoxins, bacteriological analysis and molecular assay of some bacterial species from coffee beans in Saudi Arabia. Bulletin of Pharmaceutical Sciences (Assiut University) 31: 345-373.
- Nelson, P.E., Toussoun, T.A. and Marasas, W.F.O., 1983. *Fusarium* species. An illustrated manual for identification. The Pennsylvania State University Press, University Park, PA, USA.
- Neme, K. and Mohammed, A., 2017. Mycotoxin occurrence in grains and the role of postharvest management as a mitigation strategies. A review. Food Control 78: 412-425.
- Nielsen, J.C., Grijseels, S., Prigent, S., Ji, B., Dainat, J., Nielsen, K.F., Frisvad, J.C., Workman, M. and Nielsen, J., 2017. Global analysis of secondary metabolite gene clusters in *Penicillium* species. Nature Microbiology 2, Article number: 17044.
- Nielsen, K.F., Frisvad, J.C., Sumarah, M. and Miller, J.D., 2006. Production of metabolites from the *Penicillium roqueforti* complex. Journal of Agricultural and Food Chemistry 54: 3756-3763.
- Nonaka, K., Chiba, T., Suga, T., Asami, Y., Iwatsuki, M., Msauuma, R., Omura, S. and Shiomi, K., 2015. Coclunol, a new penicillic acid produced by a co-culture of *Fusarium solani* FKI-6853 and *Talaromyces* sp. FKI-65. Journal of Antibiotics 68: 530-532.
- Northolt, M.D., Van Egmond, H.P. and Paulsch, W.E., 1978. Patulin production by some fungal species in relation to water activity and temperature. Journal of Food Protection 41: 885-890.
- Northolt, M.D., Van Egmond, H.P. and Paulsch, W.E., 1979. Penicillic acid production by some fungal species in relation to water activity and temperature. Journal of Food Protection 42: 476-484.

- Nozawa, K., Udagawa, S., Nakajima, S. and Kawai, K., 1989. A dioxopiperazine derivative from *Penicillium megasporum*. *Phytochemistry* 28: 929-931.
- Nukina, M., 1988. Terrestric acid as a phytotoxic metabolite from *Pyricularia oryzae* Cavara. *Agricultural and Biological Chemistry* 52: 2357-2358.
- Obana, H., Kumeda, Y. and Nishimune, T., 1995a. Mutagenicity of 5,6-dihydropenicillic acid (DHPA) in *Drosophila melanogaster* and bacterial conversion of penicillic acid to DHPA. *Journal of Food Protection* 58: 1375-1378.
- Obana, H., Kumeda, Y. and Nishimune, T., 1995b. *Aspergillus ochraceus* production of 5,6-dihydropenicillic acid in culture and foods. *Journal of Food Protection* 58: 519-523.
- O'Brien, M., Nielsen, K.F., O'Kiely, P., Forristal, P.D., Fuller, H.T. and Frisvad, J.C., 2006. Mycotoxins and other secondary metabolites produced *in vitro* by *Penicillium paneum* Frisvad and *Penicillium roqueforti* Thom isolated from baled grass silage in Ireland. *Journal of Agricultural and Food Chemistry* 54: 9268-9276.
- Oh, S., Chung, I., Paik, S. and Yu, S., 1998. Survey and control of the occurrence of mycotoxins from postharvest cereals. 1. Mycotoxins produced by *Penicillium* isolates from corn and wheat. *Korean Journal of Plant Pathology* 14: 700-704.
- Oh, S.-Y., Cedergreen, N., Yiannikouris, A., Swamy, H.V.L.N. and Karrow, N.A., 2015. Assessing interactions of binary mixtures of *Penicillium* mycotoxins (PMs) by using a bovine macrophage cell line (BoMacs). *Toxicology and Applied Pharmacology* 318: 33-40.
- Okeke, B., Seigle-Murandi, F., Steiman, R., Benoit-Guyod, J.-L. and Kaouadji, M., 1993. Identification of mycotoxin-producing fungal strains: a step in the isolation of compounds active against rice fungal diseases. *Journal of Agricultural and Food Chemistry* 41: 1731-1735.
- Olivigni, F.J. and Bullerman, L.B., 1977. Simultaneous production of penicillic acid and patulin by a *Penicillium* species isolated from Cheddar cheese. *Journal of Food Science* 42: 1654-1658.
- Olivigni, F.J. and Bullerman, L.B., 1978. Production of penicillic acid and patulin by an atypical *Penicillium roqueforti* isolate. *Journal of Applied Microbiology* 35: 435-438.
- Overy, D.P. and Frisvad, J.C., 2003. New *Penicillium* species associated with bulbs and root vegetables. *Systematic and Applied Microbiology* 26: 631-639.
- Overy, D.P., Valdez, J.G. and Frisvad, J.C., 2005. Revisions to the *Penicillium* ser. *Corymbifera*: agents responsible for the blue mould storage rot of various flower and vegetable bulbs. *Canadian Journal of Botany* 83: 1422-1433.
- Oxford, A.E., Raistrick, H. and Smith, G., 1942. Antibacterial substances from moulds. Part II. Penicillic acid, a metabolic product of *Penicillium puberulum* Bainier and *Penicillium cyclopium* Westling. *Chemistry & Industry* 61: 22-24.
- Oyero, O.G. and Oyefolu, A.O.B., 2009. Fungal contamination of crude herbal remedies as a possible source of mycotoxin exposure in man. *Asian Pacific Journal of Tropical Medicine* 2: 328-343.
- Pandian, V., Nayeem, M., Nanjappan, K. and Ramamurti, R., 1990. Penicillic acid as Na⁺, K⁺ and Ca²⁺ channel blocker in isolated frog heart at toxic levels. *Indian Journal of Experimental Biology* 28: 295-296.
- Paster, N., Huppeert, D. and Barkai-Golan, R., 1995. Production of patulin by different strains of *Penicillium expansum* in pear and apple cultivars stored at different temperatures and modified atmospheres. *Food Additives & Contaminants* 12: 51-58.
- Paterson, R.R.M., 2004. The isoepoxydon dehydrogenase gene of patulin biosynthesis in cultures and secondary metabolites as candidate PCR inhibitors. *Mycological Research* 108: 1431-1437.
- Paterson, R.R.M., Kozakiewicz, Z., Locke, T., Brayford, D. and Jones, S.C.B., 2003. Novel use of isoepoxydon dehydrogenase gene probe of the patulin metabolic pathway and chromatography to test penicillia isolated from apple production systems for the potential to contaminate apple juice with patulin. *Food Microbiology* 20: 359-364.
- Paterson, R.R.M., Simmonds, M.S.J. and Blaney, W.M., 1987. Mycopesticidal effects of characterized extracts of *Penicillium* isolates and purified secondary metabolites (including mycotoxins) on *Drosophila melanogaster* and *Spodoptora littoralis*. *Journal of Invertebrate Pathology* 50: 124-133.
- Patterson, M.F. and Damoglou, A.D., 1985. Identification and potential toxicity of fungi isolated from mould spoiled foods. *Record of Agricultural Research* 33: 49-55.
- Paucod, J.C., Krivobok, S. and Vidal, D., 1990. Immunotoxicity testing of T-2 and patulin on Balb/C mice. *Acta Microbiologica Hungarica* 37: 331-339.
- Pazhavinel, N., Balachandran, C., Muralimanohar, B., Dhinakarraj, G., Balahrishnan, V., Kirubuharan, J.J. and Raja, A., 2015. Alleviate effect of gingerol on cell mediated and humoral immunity and immune organs against penicillic acid mycotoxicosis in broiler chickens. *International Journal of Life science and Pharma Research* 5: L28-L34.
- Peltonen, K., Jestoi, M. and Eriksen, G.S., 2010. Health effects of moniliformin a poorly understood *Fusarium* mycotoxin. *World Mycotoxin Journal* 3: 403-414.
- Phainuphong, P., Rukachaisirikul, V., Tedpetch, K., Sukpondma, Y., Saithong, S., Phongpichit, S., Preedanon, S. and Sakayaroj, J., 2017. γ-butenolide and furanone derivatives from the soil-derived fungus *Aspergillus sclerotiorum* PSU-RSPG178. *Phytochemistry* 137: 165-173.
- Pitt, J.I., 1979. The genus *Penicillium* and its teleomorphic states *Eupenicillium* and *Talaromyces*. Academic Press, London, UK.
- Priest, J.W., 1989. Patulin biosynthesis – epoxydation of toluquinol and gentisyl alcohol by particulate preparations from *Penicillium patulum*. *Biochemistry* 28: 9192-9200.
- Puel, O., Galtier, P. and Oswald, I.P., 2010. Biosynthesis and toxicological effects of patulin. *Toxins* 2: 613-631.
- Puel, O., Tadrist, S., Delaforge, M., Oswald, I.P. and Lebrihi, A., 2007. The inability of *Byssochlamys fulva* to produce patulin is related to absence of 6-methylsalicylic acid synthase and isoepoxydon dehydrogenase genes. *International Journal of Food Microbiology* 115: 131-139.
- Qi, S., Nong, S., Zhang, X. and Xu, X., 2015. Butenolide compounds for antifouling agent use in marine coatings. Faming Zhuanli Shengqing, CN 104356730 A 20150218 [patent].

- Quintanilla, J.A., 1982. Cuatro nuevas especies de *Penicillium* aisladas en centeno: *P. mariae crucis*, sp. nov., *P. castellae*, sp. nov., *P. cieglerii*, sp. nov., y *P. smithii*, sp. nov. Avances en Alimentación y Mejora Animal 23: 333-343.
- Rabie, C.J., Lübben, A., Louw, A.I., Rathbone, E.B., Steyn, P.S. and Vleggaar, R., 1978. Moniliformin, a mycotoxin from *Fusarium fusaroides*. Journal of Agricultural and Food Chemistry 26: 375-379.
- Rabie, C.J., Marasas, W.F.O., Thiel, P.G., Lübben, A. and Vleggaar, R., 1982. Moniliformin production and toxicity of different *Fusarium* species from Southern Africa. Applied and Environmental Microbiology 43: 517-521.
- Raja, H.A., Miller, A.N., Pearce, C.J. and Oberlies, N.H., 2017. Fungal identification using molecular tools: a primer for the natural products research community. Journal of Natural Products 80: 756-770.
- Raphael, R.A., 1947a. Synthesis of the antibiotic, penicillic acid. Nature 160: 261-262.
- Raphael, R.A., 1947b. Compounds related to penicillic acid. Part. II. Synthesis of dihydropenicillic acid. J. Chem. Soc. 1947: 805-808.
- Rasmussen, T.B., Skinderoe, M.E., Bjarnsholt, T., Phipps, R.K., Christensen, K.B., Jensen, P.O., Andersen, J.B., Koch, B., Larsen, T.O., Hentzer, M., Eberl, L., Hoiby, N. and Givskov, M., 2005. Identification and effects of quorum-sensing inhibitors produced by *Penicillium* species. Microbiology-SGM 151: 1325-1340.
- Reddy, A.S. and Reddy, S.M., 1984. Evaluation of certain volatile compounds against *Aspergillus terreus* and patulin production. National Academy Science Letters (India) 7: 239-241.
- Reddy, V.K. and Reddy, S.M., 1988. Biochemical changes and patulin and terreic acid production by *Aspergillus terreus* in different cultivars of maize (*Zea mays* Linn). Journal of Food Science and Technology 25: 247-248.
- Reio, L., 1958. A method for the paper-chromatographic separation and identification of phenol derivates, mould metabolites and related compounds of biochemical interest, using a reference system. Journal of Chromatography 1: 338-373.
- Rice, S.L., Beuchat, L.R. and Worthington, R.E., 1977. Patulin production by *Byssochlamys* spp. in fruit juices. Applied and Environmental Microbiology 34: 791-796.
- Samadha, M.K. and Balachandran, C., 2008. Serum electrolyte changes in penicillic acid toxicosis. Indian Veterinary Journal 85: 248-250.
- Samson, R.A., Houbraken, J., Varga, J. and Frisvad, J.C., 2009. Polyphasic taxonomy of the heat resistant ascomycete genus *Byssochlamys* and its *Paecilomyces* anamorphs. Persoonia 22: 14-27.
- Samson, R.A., Hubka, V., Varga, J., Houbraken, J., Hong, S.B., Klaassen, C.H.W., Perrone, G., Seifert, K.A., Magistá, D., Visagie, C.M., Kocsbáé, S., Szegedi, G., Yaguchi, T., Peterson, S.W. and Frisvad, J.C., 2017. Response to Pitt and Taylor 2016: Conservation of *Aspergillus* with *A. niger* as the conserved type is unnecessary and potentially disruptive. Taxon 66: 1439-1446.
- Samson, R.A., Peterson, S.W., Frisvad, J.C. and Varga, J., 2011a. New species in *Aspergillus* section *Terrei*. Studies in Mycology 69: 39-55.
- Samson, R.A., Varga, J., Meijer, M. and Frisvad, J.C., 2011b. New species in *Aspergillus* section *Usti*. Studies in Mycology 69: 81-97.
- Samson, R.A., Visagie, C.M., Houbraken, J., Hong, S.-B., Hubka, V., Klaassen, C.H.W., Perrone, G., Seifert, K.A., Susca, A., Tanney, J.B., Varga, J., Kocsbáé, S., Szegedi, G., Yaguchi, T. and Frisvad, J.C., 2014. Phylogeny, identification and nomenclature of the genus *Aspergillus*. Studies in Mycology 78: 141-173.
- Sanders, A.G., 1946. Effect of some antibiotics on pathogenic fungi. Lancet 1946: 44-46.
- Sanderson, P.G. and Spotts, R.A., 1995. Postharvest decay of winter pear and apple fruit caused by species of *Penicillium*. Phytopathology Journal 85: 103-110.
- Sant'Ana, A.S., Simas, R.C., Almeida, C.A.A., Cabral, E.C., Rauber, R.H., Mallmann, C.A., Eberlin, M.N., Rosenthal, A. and Massaguer, P.R., 2010. Influence of package, type of apple juice and temperature on the production of patulin by *Byssochlamys nivea* and *Byssochlamys fulva*. International Journal of Food Microbiology 142: 156-163.
- Sarmadha, M.K., Balachandran, C. and Manohar, B.M., 2008a. Haematological changes in penicillic acid mycotoxicosis in the broiler chicken. Indian Veterinary Journal 85: 246-247.
- Scarpino, V., Reyneri, A., Vanara, F., Scopel, C., Causin, R. and Blandino, M., 2015. Relationship between European cork borer injury, *Fusarium proliferatum* and *F. subglutinans* and moniliformin contamination in maize. Field Crops Research 183: 69-78.
- Scauflaire, J., Gourgue, M. and Munaut, F., 2011. *Fusarium temperatum* sp. nov. from maize, an emergent species closely related to *Fusarium subglutinans*. Mycologia 103: 586-597.
- Schurman, B.T.M., Sallum, W.S.T. and Takahashi, J.A., 2010. Austin, dehydroaustin and other metabolites from *Penicillium brasiliianum*. Química Nova 33: 1044-1046.
- Schütt, F., Nirenberg, H. and Demi, G., 1998. Moniliformin production in the genus *Fusarium*. Mycotoxin Research 14: 35-40.
- Scott, A.I., Kennedy, B. and Van Walbeek, W., 1972. Desoxypatulinic acid from a patulin-producing strain of *Penicillium patulum*. Experientia 28: 1252.
- Scott, A.I., Phillips, G.T. and Kirscheis, U., 1971. Biosynthesis of polyketides. The synthesis of 6-methyl salicylic acid and triacetic acid lactone in *Penicillium patulum*. Bioorganic Chemistry 1: 380-399.
- Scott, P.M., 1974. Patulin. In: Purchase, I.F.H. (ed.) Mycotoxins. Elsevier, Amsterdam, the Netherlands, pp. 383-403.
- Scott, P.M., Abbas, H.K., Mirocha, C.J., Lawrence, G.A. and Weber, D., 1987. Formation of moniliformin by *Fusarium sporotrichioides* and *Fusarium culmorum*. Applied and Environmental Microbiology 53: 196-197.
- Scurti, J.C., Codignola, A., Nobili, G. and Caputo, O., 1973. Un ceppo di *Byssochlamys nivea* Westl., isolata da insilato di mais integrale, producente patulina. Allionia 19: 39-42.
- Seifert, K.A., Samson, R.A., Deward, J.R., Houbraken, J., Levesque, C.A., Moncalvo, J.M., Louiz-Seize, G. and Hebert P.D.N., 2007. Prospects of fungal identification using CO1 DNA barcodes, with *Penicillium* as a test case. Proceedings of the National Academy of Sciences of the USA 104: 3901-3906.
- Sekeguchi, J. and Gaucher, G.M., 1978. Identification of phyllostone as an intermediate of the patulin pathway in *Penicillium urticae*. Biochemistry 17: 1785-1791.
- Sekiguchi, J., 1983. The biosynthesis of the mycotoxin, patulin. Hakkokogaku Kaishi 61: 129-137 [In Japanese].

- Sekiguchi, J., Gaucher, G.M. and Yamada, Y., 1979. Biosynthesis of patulin in *Penicillium urticae*: identification of isopatulin as a new intermediate. *Tetrahedron Letters* 20: 41-42.
- Şenyuva, H.Z., Gilbert, J. and Öztürkoğlu, S., 2008. Rapid analysis of fungal cultures and figs for secondary metabolites by LC/TOF-MS. *Analytica Chimica Acta* 617: 97-106.
- Sewram, V., Nieuwoudt, T.W., Marasas, W.F.O., Shephard, G.S. and Ritieni, A., 1999. Determination of the mycotoxin moniliformin in cultures of *Fusarium subglutinans* and in naturally contaminated maize by HPLC-atmospheric pressure chemical ionization mass spectrometry. *Journal of Chromatography A* 848: 185-191.
- Sharma, D., Asrani, R.K., Ledoux, D.R., Jindal, N., Rottinghaus, G.E. and Gupta, V.K., 2008. Individual and combined effects of fumonisins B1 and moniliformin in clinicopathological and cell-mediated immune response in Japanese quail. *Poultry Science* 87: 1039-1051.
- Sharma, R.P., 1993. Immunotoxicity of mycotoxins. *Journal of Dairy Science* 76: 892-897.
- Shiono, Y., Murayama, T., Takahashi, K., Okada, K., Katohda, S. and Ikeda, M., 2005. Three oxygenated cyclohexanone derivatives produced by an endophytic fungus. *Bioscience, Biotechnology, and Biochemistry* 69: 287-292.
- Simonart, P. and De Lathouwer, R., 1956. Formation de patuline par *Penicillium griseofulvum* Dierckx. *Zentralblatt fur Bakteriologie, Parasitenkunde, Infektionskrankheiten und Hygiene Abt. II* 110: 107-109.
- Singh, J., 1967. Patulin. In: Gottlieb, D. and Shaw, P.D. (eds.) *Antibiotics*. Vol. 1. Springer Verlag, New York, NY, USA, pp. 621-630.
- Sklenář, F., Jurjević, Ž., Zalar, P., Frisvad, J.C., Visagie, C., Kolařík, M., Houbraken, J., Chen, A.J., Yilmaz, N., Seifert, K.A., Coton, M., Deniel, F., Gunde-Cimerman, N. and Samson, R.A., Peterson, S.W. and Hubka, V., 2017. Phylogeny of xerophilic aspergilli (subgenus *Aspergillus*) and taxonomic revision of section *Restricti*. *Studies in Mycology* 88: 161-236.
- Skóra, J., Sylyok, M., Otlewska, A. and Gutarowska, B., 2017. Toxinogenicity and cytotoxicity of *Alternaria*, *Aspergillus* and *Penicillium* moulds isolated from working environments. *International Journal of Environmental Science and Technology* 14: 595-608.
- Snini, S.P., Tadriest, S., Lafitte, J., Jamin, E.L., Oswald, I.P. and Puel, O., 2014. The gene *patG* involved in the biosynthesis pathway of patulin, a food-borne mycotoxin, encodes a 6-methylsalicylic acid decarboxylase. *International Journal of Food Microbiology* 171: 77-83.
- Snini, S.P., Tannous, J., Heuillard, P., Bailly, S., Lippi, Y., Zehraoui, E., Barreau, C., Oswald, I.P. and Puel, O., 2015. Patulin is a cultivar-dependent aggressiveness factor favouring the colonization of apples by *Penicillium expansum*. *Molecular Plant Pathology* 17: 920-930.
- Sommer, N.F., Buchanan, J.R. and Fortlage, R.J., 1974. Production of patulin by *Penicillium expansum*. *Journal of Applied Microbiology* 28: 589-593.
- Sonjak, S., Frisvad, J.C. and Gunde-Cimerman, N., 2005. Comparison of secondary metabolite production by *Penicillium crustosum* strains, isolated from Arctic and other various ecological niches. *FEMS Microbiology Ecology* 53: 51-60.
- Sorenson, W.G. and Simpson, J., 1986. Toxicity of penicillic acid for rat alveolar macrophages *in vitro*. *Environmental Research* 41: 505-513.
- Steiman, R., Seigle-Murandi, F., Sage, L. and Krivobok, S., 1989. Production of patulin by micromycetes. *Mycopathologia* 105: 129-133.
- Stoev, S., QAnguelov, G., Pavlov, D. and Pirovski, L., 1999. Some antidotes and paraclinical investigations in experimental intoxication with ochratoxin A and penicillic acid in chicks. *Veterinary Archives* 69: 179-189.
- Stoev, S.D. and Denev, S.A., 2013. Porcine/chicken or human nephropathy as a result of joint mycotoxins interaction. *Toxins* 5: 1503-1530.
- Stoev, S.D., 2013. Food safety and increasing hazard of mycotoxin occurrence in foods and feeds. *Critical Reviews in Food Science and Nutrition* 53: 887-901.
- Stoev, S.D., 2017. Balkan Endemic Nephropathy – still continuing enigma, risk assessment and underestimated hazard of joint mycotoxin exposure of animals or humans. *Chemico-Biological Interactions* 261: 63-79.
- Stoev, S.D., Dutton, M.F., Njobeh, P.B., Mosonik, J.S. and Steenkamp, P.A., 2010. Mycotoxic nephropathy in Bulgarian pigs and chickens: complex aetiology and similarity to Balkan Endemic Nephropathy. *Food Additives and Contaminants Part A* 27: 72-88.
- Stoev, S.D., Vitanov, S., Anguelov, G., Petkova-Bocharova, T. and Creppy, E.E., 2001. Experimental mycotoxin nephropathy in pigs provoked by a diet containing ochratoxin A and penicillic acid. *Veterinary Research Communications* 25: 205-223.
- Svendsen, A. and Frisvad, J.C., 1994. A chemotaxonomic study of the terverticillate penicillia based on high performance liquid chromatography of secondary metabolites. *Mycological Research* 98: 1317-1328.
- Szebiotko, K., Chełkowski, J., Dopierała, G., Godlewska, B. and Radomyska, W., 1981. Mycotoxins in cereal grain. I. Ochratoxin, citrinin, sterigmatocystin, penicillic acid and toxigenic fungi in cereal grain. *Die Nahrung* 25: 415-421.
- Tachibana, M., Matsui, C., Takeuchi, Y., Suzuki, E. and Umezawa, K., 2008. Inhibition of NF-κappa B activation by penicillic acid and dihydropenicillic acid isolated from fungi. *Heterocycles* 76: 1561-1569.
- Takahashi, J.A., De Castro, M.C.M., Souza, G.G., Lucas, E.M.F., Bracarense, A.A.P., Abreau, L.M., Marriel, I.E., Oliveira, M.S., Floreano, M.B. and Oliveira, T.S., 2008. Isolation and screening of fungal species isolated from Brazilian cerrado soil for antibacterial activity against *Escherichia coli*, *Staphylococcus aureus*, *Streptococcus pyogenes* and *Listeria monocytogenes*. *Journal de Mycologie Médicale* 18: 198-204.
- Tanenbaum, S.W. and Bassett, E.W., 1959. The biosynthesis of patulin. III. Evidence for a molecular rearrangement of the aromatic ring. *Journal of Biological Chemistry* 234: 1861-1866.
- Tangni, E.K. and Pussemier, L., 2007. Ergosterol and mycotoxins in grain dusts from fourteen Belgian cereal storages: a preliminary screening survey. *Journal of the Science of Food and Agriculture* 87: 1263-1270.
- Tannous, J., El Khoury, R., Snini, S.P., Lippi, Y., El Khoury, A., Atoui, A., Lteif, R., Oswald, I.P. and Puel, O., 2014. Sequencing, physical organization and kinetic expression of the patulin biosynthetic gene cluster from *Penicillium expansum*. *International Journal of Food Microbiology* 189: 51-60.

- Thiel, P.G., 1978. A molecular mechanism for the toxic action of moniliformin, a mycotoxin produced by *Fusarium moniliforme*. *Biochemical Pharmacology* 27: 483-486.
- Tseng, T.C., 1993. Mycotoxins produced by *Fusarium* spp. of Taiwan. *Botanical Bulletin of Academia Sinica* 34: 261-269.
- Tsunoda, H., Kishi, K., Okubo, K., Tatsuno, T. and Ohtsubo, K., 1978. Morphology and culture of *Penicillium ochraceum* and *P. carneolutescens* in *Penicillium*. *Proceedings of the Japanese Association of Mycotoxicology* 5-6: 11-13.
- Tuthill, D.E. and Frisvad, J.C., 2002. *Eupenicillium bovinosum*, a new species from dry cow manure in Wyoming. *Mycologia* 94: 240-246.
- Ueno, I., 1994. Toxicities of the mycotoxins produced by the fungi of *Penicillium* species. *Mycotoxins* 40: 15-17.
- Uhlig, S., Torp, M., Jarup, J., Parich, A., Gutleb, A.C. and Kraska, R., 2004. Moniliformin in Norwegian grain. *Food Additives and Contaminants* 21: 598-606.
- Ukai, T., Yamamoto, Y. and Yamamoto, T., 1954. Studies on the poisonous substance from a strain of *Penicillium* (Hori-Yamamoto strain) II. Culture method of Hori-Yamamoto strain and chemical structure of its poisonous substance. *Journal of the Pharmaceutical Society of Japan* 74: 450-454.
- Umezawa, M., Mizuhara, Y., Uekane, K. and Hagihara, M., 1947. A crystalline antibacterial substance from *Penicillium leucopus* and four other strains of *Penicillium* sp. and *Aspergillus clavatus* and its probable identity with patulin. *Journal of Penicillin* 1: 6-13.
- Van Eijk, G.W., 1969. Isolation and identification of orsellinic acid and penicillic acid produced by *Penicillium fennelliae* Stolk. *Antonie van Leeuwenhoek* 35: 497-504.
- Van Luijk, A., 1938. Antagonism between various micro-organisms and different species of the genus *Pythium*, parasitizing upon grasses and lucerne. *Mededelingen Phytopathologisch Laboratorium, Baarn, Scholten* 14: 43-89.
- Vansteelandt, M., Kerzaon, I., Blanchet, E., Du Pont, T.R., Jouibert, Y., Frisvad, J.C., Pouchus, Y.F. and Grovel, O., 2012. Patulin and secondary metabolite production by marine-derived *Penicillium* strains. *Fungal Biology* 116: 954-961.
- Varga, J., Due, M., Frisvad, J.C. and Samson, R.A., 2007a. Taxonomic revision of *Aspergillus* section *Clavati* based on molecular, morphological and physiological data. *Studies in Mycology* 59: 89-106.
- Varga, J., Frisvad, J.C. and Samson, R.A., 2007b. Polyphasic taxonomy of *Aspergillus* section *Candidi* based on molecular, morphological and physiological data. *Studies in Mycology* 59: 75-88.
- Varga, J., Frisvad, J.C. and Samson, R.A., 2011b. Two new aflatoxin producing species, and an overview of *Aspergillus* section *Flavi*. *Studies in Mycology* 69: 57-80.
- Varga, J., Frisvad, J.C., Kocsimbé, S., Brankovics, B., Tóth, B., Szigeti, G. and Samson, R.A., 2011a. New and revisited species in *Aspergillus* section *Nigri*. *Studies in Mycology* 69: 1-17.
- Veselá, D. and Veselý, D., 1995. Determination of *Penicillium* species using the production of mycotoxins. In: Kubátová, A. and Prášil, K. (eds.) Present state, modern methods and perspectives in *Penicillium* study. *Proceedings of the Penicillium seminar*. June 9, 1994. Czech Scientific Society for Mycology Praha, Prague, Czech Republic, pp. 98-102.
- Vesely, D. and Veselá, D., 1991. The use of chick-embryo for prediction of some embryotoxic effects of mycotoxins in mammals. *Veterinary Medicine* 36: 175-182.
- Visagie, C.M., Houbraken, J., Frisvad, J.C., Hong, S.-B., Klaassen, C.H.W., Perrone, G., Seifert, K.A., Varga, J., Yaguchi, T. and Samson, R.A., 2014b. Identification and nomenclature of the genus *Penicillium*. *Studies in Mycology* 78: 343-371.
- Visagie, C.M., Varga, J., Houbraken, J., Meijer, M., Kocsimbé, S., Yilmaz, N., Fotedar, R., Seifert K.A., Frisvad, J.C. and Samson, R.A., 2014a. Ochratoxin production and taxonomy of the yellow aspergilli (*Aspergillus* section *Circumdati*). *Studies in Mycology* 78: 1-61.
- Visagie, C.M., Yilmaz, N., Renaud, J.B., Sumarah, M.W., Hubka, V., Frisvad, J.C., Chen, A.J., Meijer, M. and Seifert, K.A., 2017. A survey of xerophilic *Aspergillus* from indoor environment, including descriptions of two new section *Aspergillus* species xerophilic species producing eurotium-like sexual states. *MycoKeys* 19: 1-30.
- Vismer, H.F., Sydenham, E.W., Schlechter, M., Brown, N.L., Hocking, A.D., Rheeder, J.P. and Marasas, W.F.O., 1996. Patulin producing *Penicillium* species isolated from naturally infected apples in South Africa. *South African Journal of Science* 92: 530-534.
- Wareing, P., Westly, A., Gibbs, J.A., Allotey, L.T. and Halm, M., 2001. Consumer preferences and fungal and mycotoxin contamination of dried cassava products from Ghana. *International Journal of Food Science & Technology* 36: 1-10.
- Welke, J.E., Hoeltz, M., Dottori, H.A. and Noll, I.B., 2011. Patulin accumulation in apples during storage by *Penicillium expansum* and *Penicillium griseofulvum* strains. *Brazilian Journal of Microbiology* 42: 172-180.
- Wiesner, B.P., 1942. Bactericidal effect of *Aspergillus clavatus*. *Nature* 149: 356-357.
- Wilson, D.M., 1976. Patulin and penicillic acid. *Advances in Chemistry* 149: 90-109.
- Wirth, J. and Klosek, R., 1972. Fungi. *Moniliales*. Relationships in *Penicillium aurantiovinorens*. *Phytochemistry* 11: 2615.
- Wirth, J.C., Gilmore, T.E. and Noval, J.J., 1956. Penicillic acid, the antibiotic responsible for the activity of a culture filtrate of a strain of *Penicillium martensii* Biourge. *Archives of Biochemistry and Biophysics* 63: 452-453.
- Wouters, M.F.A. and Speijers, G.J.A., 1996. Patulin. In: World Health Organisation (eds.) *Toxicological evaluation of certain food additives and contaminants*. WHO, Geneva, Switzerland, pp. 337-402.
- Xie, F.-C., Liu, C.-L., Yang, X.-Z. and Chen, Y.-L., 2011. Isolation of main patulin-producing strains from storage pears and their ITS sequences analysis and identification. *Food Fermentation in India* 37: 54-57.
- Yamaji, K., Fukushi, Y., Hashidoko, Y. and Tahara, S., 2005. *Penicillium frequentans* isolated from *Picea glehnii* seedling roots as a possible biological control agent against damping-off. *Ecological Research* 20: 103-107.
- Yamamoto, T., 1954. Studies on the poison producing mold isolated from dry malt. VII. Variation of patulin during culture of *Penicillium urticae* Bainier. *Journal of the Pharmaceutical Society of Japan* 76: 1377-1381.
- Yamamoto, W., Yoshitani, K. and Maeda, M., 1955. Studies on the *Penicillium* and *Fusarium* rots of Chinese yam and their control. *Science Reports of the Hyogo University of Agriculture* 1: 69-79.

- Yang, J., Li, J., Jioang, Y., Duan, X., Qu, H., Yang, B., Chen, F. and Sivakumar, D., 2014. Natural occurrence, analysis and prevention of mycotoxins in fruits and their processed products. Critical Reviews in Food Science and Nutrition 54: 64-83.
- Yassin, M.A., El-Samawaty, A.R., Bahkali, A., Moslem, M., Abd-Elsalam, K.A. and Hyde, K.D., 2010. Mycotoxin-producing fungi occurring in sorghum grains from Saudi Arabia. Fungal Diversity 44: 45-52.
- Yeulet, S.E., Mantle, P.G., Rudge, M.S. and Greig, J.B., 1988. Nephrotoxicity of *Penicillium aurantiogriseum*, a possible factor in the etiology of balkan Endemic Nephropathy. Mycopathologia 102: 21-30.
- Yin, G., Zhang, Y., Sheng, S., Hua, S.S.T., Yu, J., Bu, L., Pennerman, K.K., Huang, Q., Guo, A. and Bennett, J.W., 2017. Genome sequencing and analysis of the postharvest fungus *Penicillium expansum* R21. Genome Announcements 5: e01516-16.
- Yu, J., Jurick, W.M., Cao, H., Yin, Y., Gaskins, V.L., Losada, L., Zafar, N., Kim, M., Bennett, J.W. and Nierman, W.C., 2014. Draft genome sequence of *Penicillium expansum* strain R19, which causes postharvest decay of apple fruit. Genome Announcements 2: e00635-14.
- Yu, K., Ren, B.A.S., Wei, J.L., Chen, C.X., Sun, J.S., Song, F.H., Dai, H.Q. and Zhang, L.X., 2010. Verrucicinol and verrucosidinol acetate, two pyrone-type polyketides isolated from a marine derived fungus, *Penicillium aurantiogriseum*. Marine Drugs 8: 2744-2754.
- Yun, H., Lim, S., Chung, J., Jo, C., Park, J., Kwon, J. and Kim, D., 2006. Isolation and characterization of *Penicillium crustosum*, a patulin producing fungus, from apples. Food Science and Biotechnology 15: 896-901.
- Zhang, D., Yang, X., Kang, J.S., Choi, H.D. and Son, B.W., 2008. Chlorohydroaspyrones A and B, antibacterial aspyrone derivatives from the marine-derived Fungus *Exophiala* sp. Journal of Natural Products 71: 1458-1460.
- Zheng, J., Xu, Z., Wang, Y., Kong, K., Liu, P. and Zhu, W., 2010. Cyclic tripeptides from the halotolerant fungus *Aspergillus sclerotiorum* PT06-1. Journal of Natural Products 73: 1133-1137.