



FSN-FP 0004

## EFFECT OF SMOKING DURATION ON THE MICROBIOLOGICAL QUALITY OF COLD-SMOKED ATLANTIC COD, *GADUS MORHUA* (LINNAEUS, 1758)

KESTER, C.T., DARAMOLA, J.A.\*, ONI, P.I., AND OSHINAYA, S.O.

Department of Biological Sciences, Bells University of Technology, Ota.

Copyright 2010, Fisheries Society of Nigeria.

This paper was prepared for presentation at the 25<sup>th</sup> Annual International Conference and Exhibition in Administrative Staff College of Nigeria (ASCON), Topo-Badagry, Lagos, Nigeria, 25<sup>th</sup> – 29<sup>th</sup> October, 2010.

This paper was selected for presentation by an FISON Program Committee following review of information contained in an abstract submitted by the author(s). Contents of the paper, as presented, have not been reviewed by the Fisheries Society of Nigeria and are subject to correction by the author(s). The material, as presented, does not necessarily reflect any position of the Fisheries Society of Nigeria, its officers, or members. Papers presented at FISON meetings are subject to publication review by Editorial Committees of the Fisheries Society of Nigeria. Electronic reproduction, distribution, or storage of any part of this paper for commercial purposes without the written consent of the Fisheries Society of Nigeria is prohibited. Permission to reproduce in print is restricted to an abstract of not more than 300 words; illustrations may not be copied. The abstract must contain conspicuous acknowledgement of where and by whom the paper was presented. Write Librarian, Fisheries Society of Nigeria (FISON), P. O. Box 2607 Apapa, Lagos.

### ABSTRACT

*This study evaluated the effect of varying smoking durations of 6, 6.5, 7 and 7.5 hours on the microbiological quality and percentage moisture content of cold-smoked Atlantic Cod, Gadus morhua. The fish samples were cold-smoked using the traditional drum oven. Four batches of the smoked fish were stored in metal baskets at ambient temperatures for a period of 12 days and assessed for Staphylococcus aureus and Escherichia coli loads, Mould count, Total Plate Count and Total Coliform Count. Analyses of the smoked fish samples were carried out at the initial stage (day 0) and subsequently every alternate days (i.e. days 2, 4, 6, 8, 10 and 12). Significant variations ( $P < 0.05$ ) were obtained for all the microbial counts on the four smoked fish samples. The best microbiologically stable cold-smoked samples were that smoked at the longest duration of 7.5 hours. This produced the least mean microbial load range of TPC ( $1.50 \times 10^3 - 2.00 \times 10^5$  cfu/g); TCC varied from 0.0MPN/g to 12.0 MPN/g ; Mould count ( $1.32 \times 10^3 - 2.55 \times 10^5$  cfu/g) and Staphylococcus aureus ( $3.0 \times 10^3 - 1.35 \times 10^5$  cfu/g) while percentage moisture content ranged between 25.3% to 15.2%. All the samples tested negative to Escherichia coli.*

### INTRODUCTION

In Nigeria, fish is either eaten fresh, preserved or processed. Smoking is carried out in fishermen camps in chambers of traditional kilns made of clay, cement blocks, or drums. The traditionally smoked fish is typical of the Lake Chad region of Nigeria (Abobarin, 2000). According to Shimang (1990), in the 1970's, most of the fishes caught in the Lake Chad were smoked before the advent of drought. Technically, the duration of smoking determines the moisture content of fish product. Also, the moisture (or water activity) of a smoked fish has been found to determine the rate of microbial growth and invariably the shelf life or keeping qualities of the fish. In cold smoking, the temperature of the smoke does not exceed 30°C (Eyo, 2001). Cold smoking is uncommon preservation technique in Nigeria, due to the need for a supporting or alternative method of preservation (Waterman, 1976). Also, study has revealed that cold-smoked fish products do not last long (Moses, 1983). However, Eyo (1981) compared cold smoked and hot smoked fish and observed that cold-smoked fish possess higher nutritive value than hot-smoked fish which became cooked in the process.

### MATERIALS AND METHOD

For this study, a 20 kg carton of Atlantic Cod (*Gadus morhua*) ranging between 150-200g were purchased from a reputable frozen foods dealer in Ota and taken to an artisanal fish smoking site located at Bells Junction, Ota, Ogun State. Following standard processing steps, 68 pieces of the frozen fish were sorted out for cold smoking at temperature around 30°C. They

were thawed, sorted into four batches of 17 pieces per batch and rinsed in clean water. Each piece of fish in the batches were folded into a round shape and held in place with a sharp stick. The smoking was done on a drum-type smoking kiln with a single smoking rack which was fabricated from a 44-gallon drum. The batches of folded *Gadus morhua* were arranged on the smoking rack and were subsequently subjected to different smoking durations of 6, 6.5, 7 and 7.5 hours at temperature ranging between 30<sup>0</sup> - 35<sup>0</sup>C. The temperature was controlled by regulating the burning of the fuelwood. Smoking temperature was monitored at intervals by checking the heat that gets to the fish at the rack and dipping a thermometer into the flesh of the fish. After smoking, the fish samples A-D were cooled and separately stored in four labelled metal baskets each containing 17 pieces of the cold-smoked fish. The samples of cold-smoked *Gadus morhua*, were coded based on the different smoking durations they were subjected to. The cold-smoked fish were then taken to the Microbiology Laboratory of Bells University of Technology, Ota for analyses. The samples of the smoked fish were stored in metal baskets on the shelf at an ambient temperature for 12 days. Analyses of the smoked fish samples were carried out at the initial stage (day 0) and subsequently every alternate days (i.e. days 2, 4, 6, 8, 10 and 12); making up seven testing days. The methods of microbiological analysis described by Lyne, (1976), were adopted for the analysis of samples of the cold-smoked *Gadus morhua*. The parameters determined were Total Plate Count (TPC), Total Coliform Count (TCC), Mould count, *Staphylococcus aureus* count and *Escherichia coli*. Percentage moisture content was also determined by A.O.A.C. 1990 method.

Data obtained from the different smoking durations against the storage days were subjected to Analysis of variance (ANOVA) while Duncan's Multiple

Range Test was used to determine significant differences between the means.

## RESULTS AND DISCUSSIONS

The values of the microbial loads of the four batches of *Gadus morhua* cold-smoked at different durations and expressed in colony forming per gram (cfu/g) were significantly different ( $P < 0.05$ ). The initial 2 days of analyses recorded very low microbial loads in all the four smoked samples, A–D. Sample D, which was smoked for the longest duration of 7.5 hours had the lowest TPC of  $1.50 \times 10^3$  cfu/g at day 0 which rose to  $2.0 \times 10^5$  cfu/g at day 12. This shows the usefulness of TPC in measuring the effectiveness of time profile and heat treatment as processing procedures (FNB/NRC, 1985). In addition to this, the phenolic fraction of wood smoke has been said to possess the highest inhibiting ability on bacteria. The dual effects of heat treatment and phenolic fraction of wood smoke as displayed in sample D are made evident in the enumerated TPC values of the smoked fish samples. This result is in agreement with the findings of Ikeme and Gugnani, (1985), who, in a comparative assessment of the effect on varying periods (i.e 4, 5, 5.5 and 6.5 hours) of smoking on the acceptability and storage stability of mackerel, reported that the samples of mackerel smoked for the longest period of 6.5 hours ( lowest moisture content), were the most stable. The TPC of the frozen sample of *Gadus morhua* was recorded to be  $4.3 \times 10^4$  cfu/g.

Total Coliform Counts are particularly useful as indicators of contamination when they occur in small numbers. Their occurrence in large numbers indicates mishandling such as temperature abuse (Mossel 1967, Silliker and Gabis, 1976). The differences in the values of TCC of the four cold-smoked samples were significant ( $P < 0.05$ ). At day 0, *Gadus morhua* cold-smoked at 6, 6.5, 7 and 7.5 hours had TCC values of 3.0, 1.0, 1.0 and 0.0MPN/g respectively. Subsequently, by

the 12th day, these have increased to 17.0, 17.0, 14.0 and 12.0MPN/g. All the samples recorded fairly consistent TCC values on initial 2 days of analyses. The occurrence of these microorganisms in the four study samples almost throughout the storage days can then be attributed to contamination from processing utensils, water, handlers and probably storage materials.

The values of mould count showed that variations of all the four cold-smoked samples were significant ( $P < 0.05$ ). Mould count of the frozen sample of *Gadus morhua* was  $1 \times 10^3$ cfu/g which was lower than the values of the four cold-smoked samples. The four cold-smoked study samples A–D recorded low levels of mould growth in the first 2 days of analysis. Sample D recorded the lowest values of mould count throughout the storage days, compared to sample A that recorded the highest mould count. Hence, the effectiveness of heat treatment and time profile in processing procedure is again established. According to Olsen (1976), yeast and mould are more resistant to inhibitory influence of smoke even up to concentration of 60mg/kg. Despite the relatively low percentage moisture content of sample D, these food borne moulds were able to grow on the low moisture fish samples, because of their relatively low moisture requirements (A.O.A.C. 1990).

The values of *Staphylococcus aureus* count showed that variations for the four cold-smoked *Gadus morhua* samples were significant ( $P < 0.05$ ). *Staphylococcus aureus* count of the frozen sample of *G. morhua* was  $3.7 \times 10^5$ cfu/g which was higher than all the values obtained for the four cold-smoked samples at the initial stage (day 0) of analyses. Hence, on the subjection of the fish to heat treatment, the values of the *S. aureus* count of the raw material dropped. This establishes inhibitory effects of phenolic fraction of smoke on *Staphylococcus aureus*. The bactericidal effect of smoke as established by Olsen, (1976), is associated

with the smoke constituents especially the phenols as well as the combined heating and drying process during smoke curing. *Staphylococcus aureus* grows poorly in competition with large numbers of other microorganisms. Small numbers are to be expected in products handled by humans. Therefore, the presence of large numbers in any food material indicates possible faulty sanitary or production practice (ICMSF, 1986). According to Huss *et al.*(1995), any handling of fish, and the associated sanitary practices from the point of harvesting, however, has the potential to contribute to the microflora on the final product.

All the four batches of cold-smoked *Gadus morhua* stored for 12 days tested negative to *Escherichia coli* throughout the storage days. The resistance of *E. coli* to adverse physical and chemical conditions is low which makes *E. coli* less useful as indicator organisms in examination of frozen or otherwise preserved fish products. There was significant difference ( $P < 0.05$ ) between the values of the percentage moisture contents of the four smoked fish samples. The highest percentage moisture content of 43.9 was recorded for sample A at day 0 which reduced to 16.0 by day 12; this can be attributed to the fact that this sample was exposed to the least duration of smoking (6 hours). This agrees with the submission of Eyo, (2001) that the moisture retention of cold-smoked fish product is usually high and may be in the order of 35-45%. Sample D that recorded the least percentage moisture content of 25.3 at day 0 was cold-smoked for the longest study duration of 7.5 hours. Despite the consistent decrease in moisture content of the cold-smoked study samples, the susceptibility to infestation by microorganisms increased with storage period, such that, the microbial load picked up from day 4 and remained increasingly high till the last day. This also agrees with of Eyo, (2001) that cold-smoked fish, being not well cooked, has

shorter shelf life and is easily infested by microorganisms such as bacteria and moulds if not properly stored. The dry season in which the study was carried out

could have contributed to the decrease in moisture content of the cold-smoked *Gadus morhua* stored for 12 days.

**Table 1: Microbial load of the frozen sample of *Gadus morhua* before smoking**

Frozen fish sample	Total plate count(cfu/g)	<i>Staph. aureus</i> (cfu/g)	Mould (cfu/g)	Moisture content (%)	Total Coliform count (MPN/g)
<b>1st reading</b>	5.0 x 10 <sup>4</sup>	4.5 x 10 <sup>4</sup>	1 x 10 <sup>3</sup>	77.73	9
<b>2nd reading</b>	3.8 x 10 <sup>4</sup>	3.5 x 10 <sup>4</sup>	1 x 10 <sup>3</sup>	78.88	7
<b>3rd reading</b>	4.2 x 10 <sup>4</sup>	3.1 x 10 <sup>4</sup>	0.9 x 10 <sup>3</sup>	76.92	6
<b>Average</b>	4.3 x 10 <sup>4</sup>	3.7 x 10 <sup>5</sup>	1 x 10 <sup>3</sup>	77.84	7.3

**Table 2: Microbial and Moisture Content (%) of Cold-smoked *Gadus morhua* stored for 12 days.**

Microbial Parameters	Storage days	Smoking durations			
		6 hours (A)	6.5 hours (B)	7 hours (C)	7.5 hours (D)
Total Plate Count (cfu/g)	0	$3.05 \times 10^4 \pm 0.05$	$2.55 \times 10^4 \pm 0.45$	$1.25 \times 10^4 \pm 0.25$	$1.50 \times 10^3 \pm 0.05$
	2	$6.15 \times 10^4 \pm 0.85$	$3.35 \times 10^4 \pm 0.35$	$3.20 \times 10^4 \pm 0.20$	$2.15 \times 10^3 \pm 0.35$
	4	$1.35 \times 10^6 \pm 1.50$	$2.95 \times 10^5 \pm 1.25$	$4.40 \times 10^4 \pm 0.40$	$4.40 \times 10^5 \pm 0.40$
	6	$1.65 \times 10^6 \pm 0.75$	$1.55 \times 10^6 \pm 1.50$	$6.80 \times 10^5 \pm 0.20$	$3.90 \times 10^5 \pm 0.10$
	8	$2.20 \times 10^6 \pm 1.00$	$1.95 \times 10^6 \pm 0.50$	$7.90 \times 10^5 \pm 0.40$	$4.05 \times 10^5 \pm 0.95$
	10	$2.50 \times 10^6 \pm 0$	$2.35 \times 10^6 \pm 1.50$	$1.20 \times 10^6 \pm 1.00$	$1.90 \times 10^5 \pm 0.60$
	12	$2.65 \times 10^6 \pm 0.50$	$3.50 \times 10^6 \pm 1.50$	$1.70 \times 10^6 \pm 1.00$	$2.00 \times 10^5 \pm 0.40$
Total Coliform Count (MPN/g)	0	$3.0 \pm 0.5$	$1.0 \pm 1.0$	$1.0 \pm 1.0$	$0.0 \pm 0$
	2	$5.0 \pm 0.5$	$3.0 \pm 2.5$	$2.0 \pm 1.5$	$2.0 \pm 0.0$
	4	$5.0 \pm 1.5$	$4.0 \pm 1.0$	$3.0 \pm 2.5$	$2.0 \pm 2.0$
	6	$7.0 \pm 2.5$	$7.0 \pm 2.0$	$5.0 \pm 4.5$	$5.0 \pm 2.0$
	8	$14.0 \pm 4.5$	$10.0 \pm 2.5$	$6.0 \pm 1.0$	$5.0 \pm 2.0$
	10	$17.0 \pm 1.0$	$12.0 \pm 6.5$	$11.0 \pm 1.5$	$8.0 \pm 3.5$
	12	$17.0 \pm 1.0$	$17.0 \pm 1.0$	$14.0 \pm 4.5$	$12.0 \pm 5.5$
Mould count (cfu/g)	0	$1.65 \times 10^4 \pm 0.05$	$1.40 \times 10^4 \pm 0.10$	$1.05 \times 10^4 \pm 0.25$	$1.32 \times 10^3 \pm 0$
	2	$3.10 \times 10^4 \pm 0.20$	$3.15 \times 10^4 \pm 0.35$	$2.53 \times 10^4 \pm 0.20$	$2.15 \times 10^4 \pm 0.15$
	4	$3.00 \times 10^5 \pm 0$	$1.55 \times 10^5 \pm 0.35$	$1.00 \times 10^5 \pm 0$	$3.5 \times 10^4 \pm 0.15$
	6	$3.00 \times 10^5 \pm 0$	$3.50 \times 10^5 \pm 0.50$	$2.15 \times 10^5 \pm 0.45$	$1.45 \times 10^5 \pm 0.15$
	8	$4.40 \times 10^5 \pm 0.40$	$4.20 \times 10^5 \pm 0.20$	$2.25 \times 10^5 \pm 0.75$	$1.55 \times 10^5 \pm 0.25$
	10	$6.90 \times 10^5 \pm 0.20$	$5.80 \times 10^5 \pm 0.30$	$5.4 \times 10^5 \pm 0.47$	$1.65 \times 10^5 \pm 0.25$
	12	$9.5 \times 10^5 \pm 0.15$	$8.5 \times 10^5 \pm 0.25$	$5.60 \times 10^5 \pm 0.40$	$2.55 \times 10^5 \pm 0.25$
<i>Staphylococcus aureus</i> (cfu/g)	0	$5.70 \times 10^4 \pm 3.70$	$3.35 \times 10^4 \pm 0.55$	$4.0 \times 10^3 \pm 0$	$3.0 \times 10^3 \pm 0$
	2	$8.45 \times 10^4 \pm 0.35$	$3.50 \times 10^4 \pm 0.20$	$1.30 \times 10^4 \pm 0.20$	$3.3 \times 10^3 \pm 0.05$
	4	$5.50 \times 10^6 \pm 0.50$	$5.30 \times 10^5 \pm 3.70$	$1.10 \times 10^5 \pm 1.90$	$7.0 \times 10^4 \pm 0.10$
	6	$5.50 \times 10^6 \pm 0.46$	$1.35 \times 10^6 \pm 0.15$	$1.20 \times 10^5 \pm 0.10$	$1.00 \times 10^5 \pm 0.20$
	8	$6.0 \times 10^6 \pm 0$	$5.0 \times 10^6 \pm 0$	$1.35 \times 10^5 \pm 1.34$	$1.10 \times 10^5 \pm 0$
	10	$6.0 \times 10^6 \pm 1.00$	$5.15 \times 10^6 \pm 0.85$	$3.95 \times 10^5 \pm 2.05$	$1.20 \times 10^5 \pm 0.10$
	12	$6.5 \times 10^6 \pm 0.45$	$5.50 \times 10^6 \pm 0.50$	$5.00 \times 10^5 \pm 4.00$	$1.35 \times 10^5 \pm 0.15$
E. coli (MPN/g)	0	0	0	0	0
	2	0	0	0	0
	4	0	0	0	0
	6	0	0	0	0
	8	0	0	0	0
	10	0	0	0	0
	12	0	0	0	0

Parameter	Storage days	Smoking durations			
		6 hours (A)	6.5 hours (B)	7 hours (C)	7.5 hours (D)
% Moisture content	0	$43.9 \pm 0.20$	$41.1 \pm 0.60$	$38.10 \pm 0.70$	$25.30 \pm 0.25$
	2	$23.95 \pm 1.75$	$20.90 \pm 0.70$	$23.81 \pm 0.21$	$22.97 \pm 0.15$
	4	$20.42 \pm 0.32$	$20.19 \pm 0.32$	$21.05 \pm 0.15$	$22.13 \pm 0.07$
	6	$19.95 \pm 0.85$	$17.47 \pm 0.27$	$19.34 \pm 0.26$	$17.25 \pm 0.55$
	8	$19.61 \pm 1.29$	$17.25 \pm 0.35$	$18.15 \pm 0.05$	$16.21 \pm 0.19$
	10	$18.59 \pm 0.71$	$16.38 \pm 0.26$	$14.99 \pm 0.29$	$15.22 \pm 0.92$
	12	$16.0 \pm 0.70$	$15.09 \pm 0.95$	$14.30 \pm 0.70$	$15.20 \pm 0.20$

Mean  $\pm$  standard deviation of triplicate experiments of each sample

Mean values with different superscripts are significantly different ( $P < 0.05$ ).

## CONCLUSION

At the end of the study, fish smoked for 7.5 hours emerged the most durable. Microbiologically, the samples cold-smoked for 7.5 hours were the most stable. The other three smoking durations, though not as effective in processing fish products can be supported with refrigeration storage in other to extend the shelf-stability of the product.

## REFERENCES

- A.O.A.C, (1990). Official methods of Analysis of the Association of Official Analytical Chemists, 15th edition. Virginia. 1298pp.
- Abobarin, A. (2000). A diagnostic study of the export of traditional fishery products in Nigeria, July edition.
- Eyo, A.A. (1981). The construction and operation of a new mechanical gas (kainji lake gas klin) KLR1 technical report series 7.
- Eyo, A. A. (2001). Fish Processing Technology in the Tropics. National Institute for Freshwater Fisheries Research, New Bussa. University of Ilorin Press, Nigeria. ISBN 978 177 0457. 403 pp.
- FNB/NRC (Food and Nutrition Board, National Research Council, USA) 1985. *An evaluation of the role of microbiological criteria for foods and food ingredients* (Sub-committee on Microbiological Criteria, Committee on Food Protection). National Academy Press, Washington D.C., USA.
- Huss, H.H. Embarck, P.K.B. and Jeppesen, V.F. (1995). Control of biological hazards in cold smoked salmon production. *Food Control* 6 (6): 335-340.
- Ikeme, A.I., Gugnani, H.C. (1988). Effect of smoking time on product quality of hot-smoked mackerel In: *Proceedings of FAO Fisheries Report (FAO), no. 400 (Suppl.); Expert Consultation on Fish Technology in Africa*, Abidjan (Cote d'Ivoire), 25-28 Apr 1988 / FAO, Rome (Italy). Fishery Industries Div. 1989, p. 124-130
- International Commission on Microbial Specifications for Foods (1986). *Microorganisms in Foods. 2. Sampling for microbiological analysis: Principles and specific applications*. 2nd ed. Blackwell Scientific Publications.
- Lyne, P.M., (1976). *Microbiological methods* Edf. Butter worms, London, 4: 169-195.
- Moses, B.S. (1983). *Introduction to tropical fisheries*, Ibadan University Press. Pg. 94-95
- Mossel, D.A.A. (1967). Ecological principles and methodological aspects of the examination of foods and feeds for indicator microorganisms. *J. Assoc. Agric. Chem.* 50, 91-104.
- Olsen, C.Z. (1976). Smoke flavour and its bacteriological effect. (UFOST-IUPAC) symposium in *Advances in smoking of foods* Sept. 8-10th
- Shimang, G. N (1990). Post-harvest losses in inland fisheries in Nigeria with emphasis on lake chad and lake kainji pg. 78-83. *In: proceedings of the symposium of post-harvest fish technology CIFA. 83. Technical paper 19, FAO, Rome. Italy.*
- Silliker, J.H. and D.A. Gabis (1976). ICMSF method studies VII. Indicator tests as substitutes for direct testing of dried foods and feeds for *Salmonella*. *Can. J. Microbiol.* 22, 971-974.
- Waterman, J.J. (1976). *Production of Dried Fish*, FAO fisheries Technical Paper No. 169 pg. 39-58.