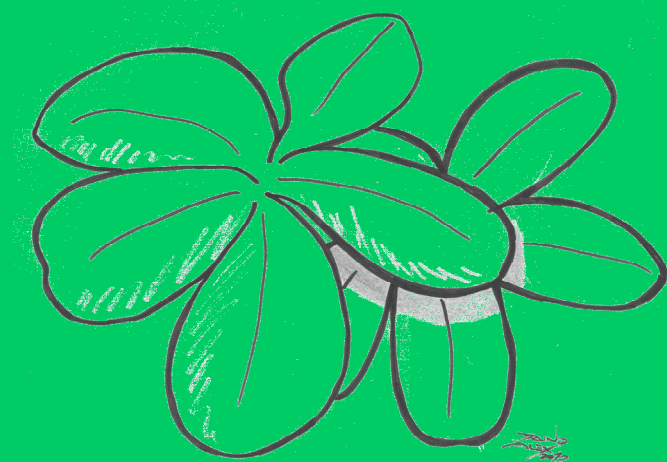


RESPONSES OF HIGHER PLANTS TO BORON DEFICIENCY



Marta Alexandra Marques Alves

June 2010
Oeiras, Portugal

Dissertation presented to obtain the degree of Doctorate in Biochemistry by
Instituto de Tecnologia Química e Biológica of Universidade Nova de Lisboa



RESPONSES OF HIGHER PLANTS TO BORON DEFICIENCY

Marta Alexandra Marques Alves



Dissertation presented to obtain the degree of Doctorate in Biochemistry by
Instituto de Tecnologia Química e Biológica of Universidade Nova de Lisboa

Supervisor: Dr. Cândido Pinto Ricardo

Co-supervisor: Dr. Phillip Jackson

June 2010

Oeiras, Portugal

The investigation was co-financed by Fundação da Ciência e Tecnologia (FCT) POCI 2010 and Fundo Social Europeu (FSE) through the PhD fellowship SFRH/BD/18273/2004.



Programa Operacional Ciência e Inovação 2010

MINISTÉRIO DA CIÊNCIA, TECNOLOGIA E ENSINO SUPERIOR

Work performed at:

Plant Biochemistry Laboratory
Instituto de Tecnologia Química e Biológica
ITQB-UNL
Av. da República - Estação Agronómica Nacional
2780-157 Oeiras
Portugal

Supervisor

Cândido Pinto Ricardo

Full Professor of Plant Physiology at *Departamento de Botânica e Engenharia Biológica*
(Instituto Superior de Agronomia, Universidade Técnica de Lisboa)

Head of the Plant Biochemistry group at *Instituto de Tecnologia Química e Biológica*
(Universidade Nova de Lisboa)

Co-supervisor

Phil Jackson

Head of the Plant Cell Wall group at *Instituto de Tecnologia Química e Biológica*
(Universidade Nova de Lisboa)

To my best friend, husband and father of my child,
Paulo Oliveira

“Although nature commences with reason and ends in experience it is necessary for us to do the opposite that is to commence with experience and from this to proceed to investigate the reason.”

Leonardo da Vinci

(15th April 1452 – 2nd May 1519)

ACKNOWLEDGMENTS

Primeiro que tudo queria agradecer à minha família! Em primeiro lugar, quero agradecer à minha Mãe por me ter dado a oportunidade de fazer as minhas conquistas e pelo seu apoio incondicional. Foi, em grande parte, graças a ti que cheguei até aqui! Muito, muito obrigada! Ao Paulo, quero apenas agradecer por ser quem é, um excelente Amigo, maravilhoso Companheiro e sempre Pai atento! É um privilégio poder partilhar todos os momentos da minha vida contigo, incluindo este! Apesar de ser ainda uma criança quero também agradecer ao Ricardo a constante alegria com que enche o meu dia a dia! Finalmente, quero agradecer ao Zé e à Fernanda, pelos pedacinhos de tranquilidade que proporcionam...

A nível de trabalho quero agradecer principalmente ao Prof. Pinto Ricardo pela oportunidade que me deu, por me ter aberto as portas do laboratório, onde desenvolvi todo este trabalho. Aprendi muito consigo, não só a nível profissional, como também a nível pessoal! Quero também agradecer ao Phil, pelas suas sugestões científicas e espírito crítico. Para o Nelson Saibo, tenho um obrigada muito especial, não só pela ajuda no desenvolvimento do trabalho de bancada, mas também pelo teu 'dom' da assertividade! Deste-me conselhos em alturas cruciais da minha carreira que fizeram toda a diferença. Gostei muito de trabalhar contigo!

Já é sabido que grande parte do nosso dia é passado no trabalho... Apesar de ser trabalho, as pessoas que nos rodeiam fazem a diferença para que o dia corra melhor. De facto, tenho que agradecer à Carla Pinheiro, à Inês Chaves, à Catarina Rodrigues e à Isa Ribeiro por me terem proporcionado bons momentos, cada uma à sua maneira! Obrigada a todas! Não podia deixar de agradecer à Carla as pertinentes discussões científicas, e à Inês o profícuo trabalho que desenvolvemos em conjunto.

Neste meu percurso científico tive oportunidade de me cruzar com diversas pessoas. Uma muito especial com quem trabalhei na quantificação de boro foi a Diana Martins. Não podia deixar de agradecer à Helena Matias pelo apoio dado no NMR, à Paula Chicau na análise das amostras por HPLC e à Ana Fortunato pelo apoio dado com o aparelho de RT-qPCR. Outras duas pessoas a quem quero deixar um obrigada especial são o Jörg Becker, pela sua paciência e claro, profissionalismo na análise de resultados dos microarrays, e a Sofia Pereira que me guiou na análise dos resultados do RT-qPCR.

I would like to thank to two lovely people that I had the opportunity to meet, Paul Jenö and Suzette Moes, for their excellent work on protein MS analysis and ready answers for my doubts.

Não me posso esquecer de agradecer a disponibilidade e prontidão na resolução de pequenos contratempos que, se não fosse pela

Eugénia Santos e pelo Alexandre Maia, se tornariam em grandes entaves ao trabalho.

Por fim, queria também agradecer ao Bruno Gouveia pelo apoio na parte gráfica da tese. Obrigada!

Aqui no ITQB, consegui não só desenvolver trabalho com qualidade científica, mas também grandes Amizades, que tornaram todo este processo bem mais interessante! Não podia deixar de agradecer à Rita Francisco pelos bons momentos conversa científica e não só... MUITO OBRIGADA, Isabel Martins e Sónia Negrão por todo o vosso apoio, não há palavras que cheguem... As conversas das montanhadas, são contigo, Cláudia Santos, MUITO OBRIGADA pelo teu acompanhamento!... Estas são Amizades que espero guardar por muitos e bons anos.

SUMMARY

Despite being known for more than 80 years that B is essential for plant growth, the role of this element in plant metabolism remains elusive. Until now, B involvement in the plant cell wall structure is the only well-documented participation of B. However, this participation can not explain all the symptoms observed in B-deficient plants nor the requirement of B for early animal developmental processes.

The aim of this study was to acquire additional data on the role(s) of B in the growth and development of higher plants. For that purpose, *Lupinus albus*, an important crop plant, was used to study long-term B deficiency responses, characterizing the proteomes of the leaf apoplast (Chapter 2) and of the root system (Chapter 3). A metabolite analysis was also performed in the different organs (leaf-blades, petioles, apexes, hypocotyls and roots) of *L. albus* plants (Chapter 4). Some consistent responses were observed with these different approaches. The analysis of the metabolite content of *Lupinus albus* plants, showed minor changes in the content of sugars and absence of variation in malate suggests that the central carbohydrate metabolism is being little affected by B deficiency (Chapter 4). The proteome study of the root, revealed the differentially expression of UDP-glucose pyrophosphorylase and of several ATPases (Chapter 3), what appears to be related with the

activation of alternative energy sources. Thus, the carbon flow seems to be able to continue even under stressful conditions. Proteins related with the cell wall biosynthesis, namely of polysaccharide components, were suppressed due to B deficiency (Chapter 3), probably as a consequence of the structural cell wall damage. Some proteins related to protein folding and proteolytic processes were suppressed while others were *de novo* expressed (Chapter 3), pointing for a shift in the protein metabolism, probably to allow the plant to partially cope with B deficiency. Several proteins related with cytoskeleton biosynthesis were also affected by B deficiency, as for instance, tubulins and actins, that were *de novo* expressed (Chapter 3). The additional increased content of the branched-chain amino acids (Chapter 4), that could be related with cytoskeleton biosynthesis, points for some active involvement of B with cytoskeleton biosynthetic processes.

The long-term effects observed could result from both direct and indirect stress responses, but the indirect effects could have some degree of specificity to the stress, as suggested by the leaf apoplast study (Chapter 2), since the majority of the proteins that were commonly responsive to both B and water-deficit showed different patterns of expression.

Short-term B deficiency responses were also investigated, performing a transcriptional analysis with the model plant *Arabidopsis thaliana*

(Chapter 5). Altered expression of genes related with the cell wall was observed, as well as genes related to branched-chain amino acids. In analogy to what was discussed in the long-term responses, the increased expression of these genes could be related with impaired cytoskeleton biosynthesis. Genes related with sulphur metabolism were also found to be differentially expressed due to B deficiency. In particular, the expression of the three genes that encode for 5'-adenylylsulfate reductase, a key enzyme of the sulphate assimilation pathway, was decreased, what is a response with similarity to sulphur excess in the plant.

Thus, altered cytoskeleton biosynthesis seems to be a common response in short and long-term B deficiency studies, suggesting that a direct interaction of B with cytoskeleton may exist. Such B participation in plant metabolism is consistent with higher B requirement in the earlier animal development and reproductive stage in plants, where cytoskeleton is actively required for cell division processes.

SUMÁRIO

Apesar de ser conhecido há mais de 80 anos que o B é essencial para o crescimento das plantas, o seu papel no metabolismo das plantas continua por elucidar. Até agora, o envolvimento na estrutura da parede celular das plantas é a única e bem documentada participação do B que, no entanto, não parece ser suficiente para explicar todos os sintomas observados nas plantas devido à deficiência de B, nem o requisito de B nos processos iniciais do desenvolvimento animal.

O objectivo deste estudo foi o de reunir informação adicional sobre o papel do B no crescimento e desenvolvimento das plantas superiores. Para isso utilizámos uma importante cultura agrícola, o *Lupinus albus*, para os estudos das respostas à deficiência prolongada de B, caracterizando o proteoma do apoplasto da folha (Capítulo 2) e do sistema radicular (Capítulo 3). Foi também efectuada uma análise metabólica nos diferentes órgãos desta planta (folhas, pecíolos, ápices, hipocótilos e raízes). Estas diferentes abordagens permitiram obter algumas respostas consistentes. Observou-se uma pequena variação nos teores de açúcares e a ausência de variação nos teores de malato, o que sugere que o metabolismo central dos hidratos de carbono está a ser pouco afectado pela deficiência (Capítulo 4). O estudo do proteoma da raiz revelou uma expressão diferenciada da UDP-glucose fosforilase

e de várias ATPases (Capítulo 3), proteínas essas envolvidas na activação de vias metabólicas alternativas para a produção de energia. A planta consegue, desta forma, manter o fluxo de carbono apesar das condições de stress.

A biossíntese da parede celular parece ter sido afectada, dada a supressão de proteínas relacionadas com síntese de polissacáridos (Capítulo 3), provavelmente como consequência de danos causados na estrutura da parede celular. O rearranjo tridimensional das proteínas e certos processos proteolíticos foram modificados (capítulo 3), indicando uma provável alteração do metabolismo das proteínas na deficiência de B.

Várias proteínas relacionadas com a biossíntese do citosqueleto foram também afectadas pela deficiência de B, como por exemplo tubulinas e actinas (Capítulo 3).

O aumento dos níveis de aminoácidos de cadeia ramificada que foi observado (Capítulo 4) poderá ser relacionado com o citosqueleto, o que poderá reforçar a hipótese da participação do B na biossíntese do citosqueleto.

Os efeitos da deficiência de B a longo prazo podem resultar de respostas directas ou indirectas ao stress, mas, todavia, mesmo sendo indirectos podem ter um certo grau de especificidade, como sugere o estudo do proteoma do apoplasto da folha (Capítulo 2). Com efeito, a maioria das proteínas que responderam tanto à

deficiência de B como à seca mostraram diferentes padrões de expressão.

A análise dos transcriptos da planta modelo *Arabidopsis thaliana* (Capítulo 5) foi usada para investigar as respostas à deficiência de B a curto prazo. Verificou-se a alteração da expressão de genes relacionados com a parede celular, bem como de genes envolvidos com os aminoácidos de cadeia ramificada. Analogamente com o que foi discutido na resposta a longo prazo, o aumento da expressão destes genes poderá estar relacionada com danos na biossíntese do citosqueleto. Genes relacionados com o metabolismo do enxofre foram também diferencialmente expressos devido à deficiência de B. Em particular a expressão dos três genes que codificam para 5'-adenililsulfato redutase, a enzima chave da via da assimilação do enxofre, diminuiu, uma resposta que se assemelha a excesso de enxofre nas plantas.

Assim, os resultados obtidos tanto a longo como a curto prazo apontam para uma possível relação entre a deficiência de B e o citosqueleto. Esta sugestão é consistente com o elevado requisito de B tanto na fase reprodutiva das plantas como na fase inicial do desenvolvimento animal, nas quais o citosqueleto é activamente solicitado para processos de divisão celular.

TABLE OF CONTENTS

ABBREVIATIONS

xxiii

| | | |
|------------------|---|----|
| CHAPTER 1 | General Introduction | 1 |
| | Introduction | 3 |
| | The chemistry of boron | 3 |
| | Boron in plants | 5 |
| | <i>Uptake and transport</i> | 6 |
| | <i>Cell wall</i> | 8 |
| | <i>Cellular membrane</i> | 10 |
| | <i>Oxidative damage</i> | 11 |
| | <i>Photosynthesis</i> | 12 |
| | <i>Signalling</i> | 12 |
| | Boron in other living organisms | 13 |
| | <i>Animals</i> | 13 |
| | <i>Unicellular Organisms</i> | 15 |
| | Scope of this Thesis | 17 |
| | References | 18 |
| CHAPTER 2 | Analysis of <i>Lupinus albus</i> leaf apoplastic proteins in response to boron deficiency | 27 |
| | Summary | 30 |
| | Introduction | 30 |
| | Materials and Methods | 32 |
| | <i>Plant material</i> | 32 |
| | <i>Boron determination</i> | 33 |
| | <i>Extraction of soluble apoplastic proteins</i> | 33 |
| | <i>Two-dimensional gel electrophoresis (2-DE)</i> | 34 |
| | <i>Protein identification by MS/MS</i> | 35 |
| | Results | 36 |
| | <i>Determination of suitable B concentration for L. albus growth</i> | 36 |
| | <i>Morphological and physiological effects of B deficiency</i> | 37 |
| | <i>Protein analysis by 2-DE</i> | 39 |
| | <i>Protein identification by MS/MS</i> | 42 |
| | Discussion | 45 |
| | Acknowledgements | 47 |
| | References | 48 |

| | | |
|------------------|---|-----|
| CHAPTER 3 | The analysis of <i>Lupinus albus</i> root proteome revealed cytoskeleton altered features due to long-term boron deficiency | 55 |
| <hr/> | | |
| | Summary | 58 |
| | Introduction | 58 |
| | Materials and Methods | 59 |
| | <i>Plant Material</i> | 59 |
| | <i>Protein extraction and solubilisation</i> | 60 |
| | <i>Two-dimensional gel electrophoresis</i> | 60 |
| | <i>Two-dimensional gel analysis</i> | 61 |
| | <i>In-gel digestion</i> | 62 |
| | <i>MS/MS analysis</i> | 62 |
| | Results and Discussion | 64 |
| | <i>Morphological effects of B deficiency</i> | 64 |
| | <i>Quantitative variations of the root proteome</i> | 65 |
| | <i>Metabolic changes associated with B deficiency</i> | 67 |
| | Conclusions | 85 |
| | Acknowledgements | 85 |
| | References | 86 |
| | | |
| CHAPTER 4 | Metabolic analysis revealed altered amino acid profiles in <i>Lupinus albus</i> organs due to boron deficiency | 93 |
| <hr/> | | |
| | Summary | 96 |
| | Introduction | 97 |
| | Materials and Methods | 98 |
| | <i>Plant Material</i> | 98 |
| | <i>Metabolite analysis</i> | 98 |
| | <i>Statistical analysis</i> | 100 |
| | Results and Discussion | 101 |
| | <i>Biomass analysis of L. albus plants grown under B deficiency</i> | 101 |
| | <i>Metabolite analysis</i> | 102 |
| | Conclusions | 110 |
| | Acknowledgements | 111 |
| | References | 111 |

| | | |
|------------------|--|-----|
| CHAPTER 5 | Transcriptional analysis of short-term boron deficiency in Arabidopsis revealed altered sulphur metabolism | 117 |
| | Summary | 120 |
| | Introduction | 121 |
| | Materials and Methods | 123 |
| | <i>Plant material and growth conditions</i> | 123 |
| | <i>Boron quantification</i> | 123 |
| | <i>RNA Isolation, Target Synthesis and Hybridization to Affymetrix GeneChips</i> | 124 |
| | <i>GeneChip Data Analysis</i> | 125 |
| | <i>Reverse transcriptase quantitative real-time PCR (RT-qPCR) analysis</i> | 127 |
| | <i>Gene annotation</i> | 128 |
| | Results and Discussion | 129 |
| | <i>Morphological symptoms of B deficiency</i> | 129 |
| | <i>Boron levels in the different treatments</i> | 129 |
| | <i>Gene expression analysis and functional categorization</i> | 129 |
| | Conclusions | 147 |
| | Acknowledgements | 148 |
| | References | 148 |
| | Supplementary data | 159 |
| CHAPTER 6 | General discussion and Future Perspectives | 243 |
| | General Discussion | 245 |
| | <i>Long-term B deficiency effects</i> | 245 |
| | <i>Short-term B deficiency effects</i> | 248 |
| | <i>A plausible hypothesis for B participation in plant metabolism</i> | 249 |
| | <i>Conclusions</i> | 252 |
| | Future perspectives | 253 |
| | References | 254 |

ABBREVIATIONS

| ABBREVIATION | FULL FORM |
|--------------|---|
| ψ pd | Water potential at pre-dawn |
| 2-DE | Two-dimensional gel electrophoresis |
| A | Apexes |
| A4SA | Aspartate 4-semialdehyde |
| ABA | Absciscic acid |
| ACT | Actin |
| AI | Auto-inducer |
| APR | 5'-Adenylyl sulphate reductase |
| APS | Ammonium persulphate |
| ATGH9CE | Glycosyl hydrolase 9C2 |
| ATMGL | Methionine gamma-lyase |
| ATP | Adenosine triphosphate |
| B | Boron |
| BCAA | Branched-chain amino acid |
| BCAT | Branched-chain amino acid aminotransferase |
| BD | Boron deficiency |
| Ca | Calcium |
| CCR | Cinnamoyl-CoA reductase |
| cDNA | Complementary DNA |
| CHAPS | 3-[3-Cholamidopropyl (dimethylammonio)]-1-propanesulphonate |
| CND | Chloroplast nucleoid DNA |
| CoA | Coenzyme A |
| cRNA | Complementary RNA |

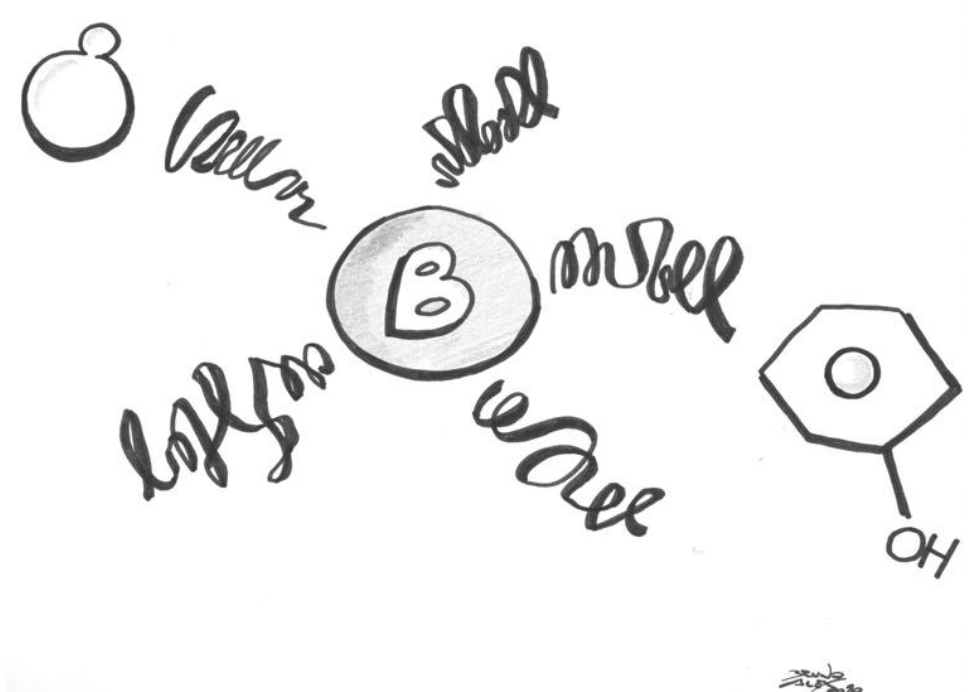
| ABBREVIATION | FULL FORM |
|---------------------|---|
| Ct | Cycle threshold |
| DNA | Deoxyribonucleic acid |
| DTT | Dithiothreitol |
| EXP5 | Expasin5 |
| FDR | False discovery rate |
| FMO GS-OX | Flavin-containing monooxygenases |
| FW | Fresh weight |
| GABA | γ -Aminobutyric acid |
| GO | Gene ontology |
| GTP | Guanosine triphosphate, |
| H | Hypocotyls |
| HPLC | High liquid performance chromatography |
| Hsp | Heat shock protein |
| ICP-AES | Inductively coupled plasma-atomic emission spectrometry |
| IEF | Isoelectric-focusing |
| IPG | Immobilized pH gradient |
| IVT | <i>In vitro</i> transcription |
| L | Leaf-blades |
| LSU | Low sulphur upregulated |
| MALDI-TOF-TOF | Matrix assisted laser desorption/ionization-time of flight-time of flight |
| MCCA | 3-Methylcrotonyl-CoA carboxylase |
| MDH | Malate dehydrogenase |
| MIOX | <i>myo</i> -Inositol oxygenase |
| MIP | Major intrinsic protein |
| mRNA | Messenger RNA |

| ABBREVIATION | FULL FORM |
|---------------------|---|
| MS | Mass spectrometry |
| <i>MS5</i> | <i>Male sterility5</i> |
| MS/MS | Tandem mass spectrometry |
| MW | Molecular weight |
| NAC | Nascent polypeptide-associated complex |
| NAD | Nicotinamide adenine dinucleotide |
| NADH | Reduced form of NAD |
| NADP | Nicotinamide adenine dinucleotide phosphate |
| NADPH | Reduced form of NADP |
| NL | Non linear |
| NMR | Nuclear magnetic resonance |
| NS1 | Nicotianamine synthase |
| ODP | Ovule development protein |
| P | Petioles |
| PAGE | Polyacrylamide gel electrophoresis |
| PAR | Photosynthetic active radiation |
| pI | Isoelectric point |
| PME | Pectin methylesterase |
| PMEI | Pectin methylesterase inhibitor |
| PP2A | Protein phosphatase 2A subunit A3 |
| PR | Pathogenesis Related |
| PVP | Polyvinylpyrrolidone |
| R | Roots |
| RG-II | Rhamnogalacturonan-II |
| RhaT | Rhamnosyltransferase |
| RNA | Ribonucleic acid |

| ABBREVIATION | FULL FORM |
|---------------------|--|
| RT-qPCR | Reverse transcriptase quantitative real-time polymerase chain reaction |
| RuBisCO | Ribulose-1,5-bisphosphate carboxylase-oxygenase |
| SDR | Short-chain dehydrogenase/reductase |
| SDS | Sodium dodecyl sulphate |
| ST | Sulfotransferase |
| TAIR | The Arabidopsis information resource |
| TCA | Trichloroacetic acid |
| TF | Transcription factor |
| TRE | Trehalase |
| Tris | Tris (hydroxymethyl) aminomethane |
| TUB | Tubulin |
| UBC10 | Ubiquitin-conjugating enzyme E2 |
| UDP | Uridine diphosphate |
| USP | Universal stress protein |
| WD | Water-deficit |

CHAPTER 1

GENERAL INTRODUCTION



INTRODUCTION

The importance of boron (B) for living organisms is quite puzzling. The dramatic effect of its deficiency in plants was first noticed almost one century ago, and current evidence suggests that B might be necessary for most organisms. Despite this, its precise functions in the metabolism of plants and other organisms remain to be elucidated. The study of B roles in metabolism is therefore, a subject of central interest in biology. In particular, in agriculture, B deficiency is a widespread problem and large yield losses are reported to occur annually. In Portugal, Olive (*Olea europaea*), Apple (*Malus domestica*) and Pear (*Pyrus communis*) are the fruit trees most affected by B deficiency. In the Douro region, grapevine “maromba” anomaly was found to be due to B deficiency, while in the Alentejo pastures this deficiency is known to causes a significant decrease in growth of legume plants.

THE CHEMISTRY OF BORON

The chemistry of B is unique and, after carbon, B might be the most intriguing and complex of elements (Greenwood and Earnshaw 1984). Along with other light elements, like lithium and berilium, B originates from the Big Bang nucleosynthesis or galactic cosmic-ray spallation (Revees 2001, Prochaska et al. 2003), and its

abundance is extremely low: only about 10^{-9} times that of hydrogen and about 10^{-6} that of carbon. However, in spite of its low abundance in the Universe, B is widely distributed in the Earth's crust (from 5mg kg^{-1} in basalts to 100mg kg^{-1} in shales), in the sea water ($1\text{-}10\text{mg L}^{-1}$) and in the rivers ($3\text{-}30\mu\text{g L}^{-1}$) (Power and Woods 1997). Boron is a small atom with only three valence electrons that belongs to the semiconductor group of elements with properties intermediate between metals and non-metals. In the industry, B is widely used and the main commercial productions and materials are colemanite, ulexite, borax, borax-pentahydrate, sodium perborates as well as boric acid. The major usage of B includes insulators, fire prevention materials, enamel coating, laundry bleach (sodium perborate), glass fibers, borosilicate glass, frit, ceramic glazes, chemical fertilizers (as a trace element for agriculture) and herbicides (WHO 1998). Tetrahedral borate or boronate complexes have been shown to be involved in enzyme inhibition. Serine proteases were proposed to be inhibited by boronic acid compounds (Yang et al. 2003) and simple borates have been patented as protease stabilizers in liquid detergent formulations (Hora and Kivits 1981, Severson 1985).

At a near-neutral pH found in most biological fluids, B exists primarily as boric acid, B(OH)_3 and a small amount as borate

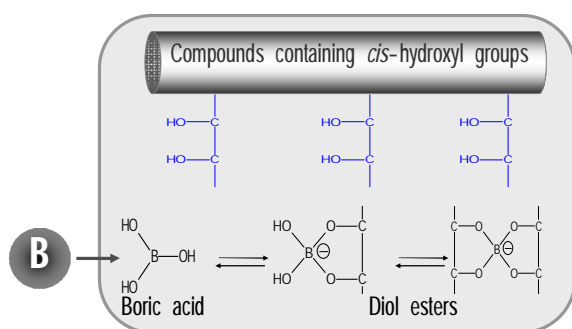


Figure 1.1. Esterification reactions of boric acid, B(OH)₃ with compounds containing *cis*-hydroxyl groups to form diol esters.

anion, B(OH)⁴⁻
(Woods 1996). Both boric acid and borate readily form complexes with a wide variety of sugars and other compounds containing *cis*-hydroxyl groups

(Loomis and Durst 1992) such as phenols, organic acids and some polymers (Figure 1.1; Boesekem 1949, Raven 1980). Except for complexes with polyols little is known about B binding inside the cell.

BORON IN PLANTS

Boron was established as an essential micronutrient for all vascular plants in the early 1920s (Warington 1923), but only recent research findings have improved our understanding of B interaction in plants for B uptake and transport processes, cell-wall formation and cellular membrane functions (Goldbach and Wimmer 2007, Bonilla et al. 2009). Boron participation in other processes was also proposed, including anti-oxidative systems (Cakmak and Römheld 1997, Koshiba et al. 2009) and signalling (Redondo-Nieto et al. 2001, González-Fontes et al. 2008).

Uptake and Transport

In soils, B occurs as borosilicates, which are resistant to weathering and not readily available to plants. Mobilisation of immobile forms of rock B occurs by acid-base, oxidation-reduction and dissolution-precipitation reactions. The dominant species in the soil, when B from primary silicates goes into solution is boric acid, $B(OH)_3$. This form of B is mobile and easily lost by leaching, but in this form it can be taken up by plants, held by organic matter or temporarily adsorbed on fine fractions (Nable et al. 1997). Depending on B availability, boric acid uptake by roots can be carried out by several molecular mechanisms such as passive diffusion across the lipid bilayers, facilitated transport by major intrinsic protein (MIP) channels, and also by energy-dependent high affinity transport systems. Whereas the first two represent passive uptake systems, which are constitutively present, the latter is induced by low B supply (Figure 1.2; Tanaka et al. 2008, Takano et al. 2008). Different proportions of sterols and longer chain fatty acids in the plasma membrane of root cells significantly altered B uptake in *Arabidopsis thaliana* mutants (Dordas and Brown 2000). Increased membrane rigidity, as a response to chilling temperature, results in a reduction of the membrane fluidity and permeability of root cells, possibly contributing to the inhibition of B uptake in chilling-sensitive species (Ye et al. 2003). Two members of the MIP family were

identified as being up-regulated in B-deficient *Arabidopsis* plants. The *NIP5;1* gene was identified in roots (Takano et al. 2006) and the *NIP6;1* gene in shoots. The boric acid/borate exporter BOR1 is responsible for loading B into the xylem under both normal and B-deficient conditions (Takano et al. 2002) and its accumulation is controlled by post-transcriptional mechanisms according to B availability (Takano et al. 2005).

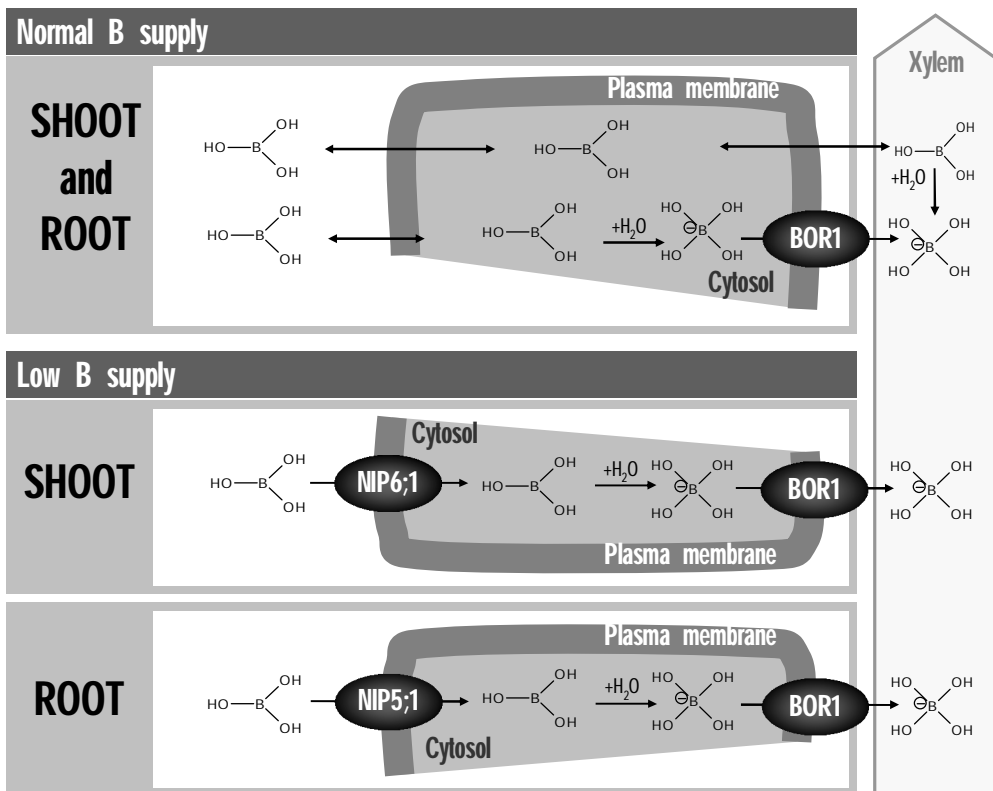


Figure 1.2. Model of B transport mechanisms in plants. Under normal B supply, B is passively transported as boric acid [B(OH)_3] across the plasma membrane. Under low B conditions, B is transported by boric channels, NIP5;1 in roots and NIP6;1 in young developing tissues. Under both normal and low B conditions the boric acid/borate exporter BOR1 is responsible for B loading into the xylem (Tanaka et al. 2008, Takano et al. 2008).

Two potential BOR1 homologues were already found in a monocot plant, *Oryza sativa* (Nakagawa et al. 2007) and in eucalypt, a tree (Domingues et al. 2005).

Boron moves from the roots with the transpiration stream and accumulates in growing points of leaves and stems (Blevins and Lukaszewski 1998). Nevertheless, B transport in the phloem depends on the sugar or polyol molecules used by a particular plant. Several B transport molecules were isolated and characterized, such as sorbitol-B-sorbitol complexes from the floral nectar of peach and mannitol-B-mannitol complexes from phloem sap of celery. There is also evidence that dulcitol is a B transport molecule in some species (Blevins and Lukaszewski 1998). Although it seems that the B retranslocation is limited to sugar alcohol-producing plants, recently it was demonstrated that B is retranslocated from old tissues to younger tissues in response to short-term B deficiency in lupin, which does not produce sugar alcohols (Huang et al. 2008).

Cell wall

In the plant cell up to 90% of the cellular B is localized in the cell wall (Martini and Thellier 1993), an important structure for determining cell size and shape during plant development. The cross-linking of the pectic polysaccharide components by B was

proposed to have an important contribution to the cell wall architecture, hence influencing the mechanical properties of growing cell walls (Kobayashi et al. 1996, O'Neill et al. 1996, 2001). Boron cross-links through the diol bridges of the rhamnogalacturonan-II (RG-II) molecules creating binding sites for

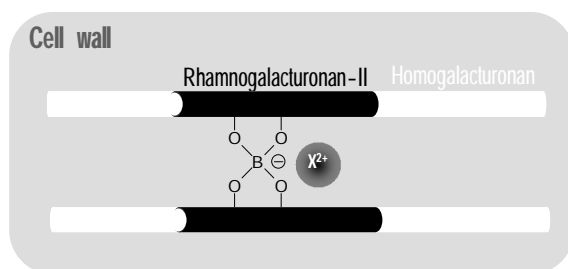


Figure 1.3. Model showing pectin rhamnogalacturonan-II molecules, of the plant cell wall, cross-linked by B. Through this cross-linking B contributes to the stabilization of the cell wall structure, additionally allowing a metal ion (X^{2+}) such as Ca, Mg, Pb, Ba or Sr to be bound to the B-RG-II complex (Ishii and Matsunaga 2001).

calcium ions (Figure 1.3), which will further increase the strength of the B-RG-II complexes (Kobayashi et al. 1999).

Genetic studies employing the *Arabidopsis thaliana mur1* mutant and the haploid *Nicotiana plumbaginifolia* callus mutant nolac-H18 indicated that B-RG-II cross-linking is essential for normal plant growth (O'Neill et al. 2001, Iwai et al. 2002). The correct establishing of an effective symbiosis between legume-*Rhizobium* is also dependent on B availability that seems to be an important component for the maintenance of the nodule cell wall structure (Bolaños et al. 1994, Bonilla et al. 1997).

Cellular membrane

A structural role for B in membranes has also been proposed due to the ability of B for linking or binding to hydroxyl-containing constituents, such as phosphoinositides, glycoproteins, glycolipids (Bonilla et al. 1997, Brown et al. 2002). Thus, B may play a role in the maintenance of membrane rheology, by modulating the hydration and fluidity of lipid bilayers (Verstraeten et al. 2005), and might explain the altered membrane composition observed in B-deficient plants. So, the lipid composition of membranes determines the magnitude and direction of B deficiency effects (Brown et al. 2002). Other observations that point to a structural role of B in membrane stabilization are modified permeability for potassium and sugars, altered membrane-bound calcium levels and damage of the peribacteroid membrane in nodules detected as a consequence of B deficiency (Goldbach and Wimmer 2007). Indeed, B is described to be an indispensable element for infection thread development and nodule invasion (Bolaños et al. 1996). In *Rhizobium*-plant interactions B seems to act as a modulator between the glycoprotein matrix of the plant and the bacterial cell surface. In the absence of B the glycoproteins can attach to the rhizobium cell surface, preventing the interaction with the plant cell membrane, thereby inhibiting cell invasion (Bolaños et al. 2004).

Several studies have indicated that B interferes with the functioning of enzymes and other proteins of the plasma membrane. For example, B deficiency alters the membrane potential, reduces the activity of Fe-reductase and proton-pumping ATPase, and thus the proton gradient across the plasma membrane. The plasma membrane-bound oxidoreductase activity was also inhibited within minutes of B deprivation (Goldbach and Wimmer 2007).

Despite the several observations for membrane damage upon B deficiency, there are not yet concluding remarks for a structural role for B in cellular membranes.

Oxidative damage

The accumulation of oxidative free radicals in cells, which is one of the indirect consequences of B deficiency in root and leaf cells, could be responsible for the impairment of membrane functions (Cakmak and Römheld 1997). In tobacco BY-2 cells grown under B deprivation, the accumulation of reactive oxygen species, higher lipid peroxidation and decreased cellular ascorbate levels were observed. The addition of lipophilic antioxidants suppressed the death of B-deprived cells, suggesting that oxidative damage is the immediate and major cause of cell death under B deficiency (Koshiba et al. 2009).

Photosynthesis

Photosynthesis has been shown to be affected by B deficiency. The existing evidence has been mostly obtained from *in vivo* experiments with plants grown under extended periods of B deficiency (Kastori et al. 1995, El-Shintinawy 1999), which raises the question whether such effects reflect a requirement for B in photosynthesis, or are related with the effects of B deficiency on other areas of plant metabolism. In fact, preliminary experiments with isolated spinach chloroplasts showed only weak effects, if any, under B-deficient conditions. It is therefore quite feasible that the effects observed in chloroplasts are secondary and caused by growth-inhibition, which could indirectly cause a reduced sink activity and an oversaturation of the electron acceptors of photosystems. These possible effects may increase the rate of photo-oxidative damage in response to further stresses (Goldbach and Wimmer 2007).

Signalling

The capacity of root exudates to induce the activity of nodulation genes is modulated by B nutrition in the host plant. Exudates derived from B-deficient pea plants led to a low level of *nod*-gene expression. Infection thread development was arrested at very early stages, and cell invasion by endocytosis was precluded, leading to

B-deficient nodules almost empty of bacteria (Redondo-Nieto et al. 2001).

González-Fontes et al. (2008) had proposed that B can have a role as a cellular signal through its direct or indirect interaction with transcription factors (TFs). The TF interaction with the target gene might be modulated by intercellular B levels due to an eventual direct linkage of B to the TF or to molecules that will then link to a TF. Recently, the WRKY6 was described to be a TF essential for the normal development of the roots of *A. thaliana* plants grown under low-B conditions (Kasajima et al. 2010).

BORON IN OTHER LIVING ORGANISMS

Only some 20 years ago it was considered that B was essential for animal development, including humans (Nielson 1997). Nowadays B is considered of nutritional importance for a broad spectrum of organisms, not only in plants and animals but also in bacteria and yeast.

Animals

The identification of a mammalian homolog of the *A. thaliana* BOR1 transporter, the NaBC1, suggests the need for minimum B levels in animal cells (Park et al. 2005). The finding that low

concentrations of borate activate the MAPK pathway and that the knockdown of NaBC1 halted cell growth and proliferation provide further evidence for an essential functional role of B in animal metabolism. Several other reports showed that B was essential for embryonic development, at least for vertebrates, where B deprivation resulted in a high percentage of necrotic eggs and abnormal development of the gut in *Xenopus laevis* (Fort et al. 1999). The early stage of development is especially sensitive to B deficiency as described for zebrafish (Rowe and Eckhert 1999). It is interesting that Behrendt and Zoglauer (1996) also demonstrated that B was required during the early phase of somatic embryogenesis in the plant species, *Larix decidua*, where suspensor development formation was blocked without a B supply.

It has been found that physiological concentrations of B reduced the amount of insulin required to maintain plasma glucose (Bakken and Hunt 2003), but in a number of nutritional studies, the beneficial effects of B in animals seems to be related with bone metabolism. For example, B supplementation of a low B diet reduced gross-bone abnormalities in the vitamin D-deficient chick (Bai and Hunt 1996) and enhanced the apparent absorption and retention of calcium and phosphorus, and increased femur magnesium concentrations in vitamin D-deficient rats (Hegsted et al. 1991). In male pigs, bone lipid was lower and the bending moment higher

when B was supplemented to a low-B diet (Armstrong et al. 2000). Thus, the effect of B on bone metabolism could be one of the essential functions of B in animals and humans.

Unicellular Organisms

Boron essentiality has been established for the growth of specific types of bacteria, such as heterocystous cyanobacteria (Bonilla et al. 1990) and actinomycetes of the genus *Frankia* (Bolaños et al. 1993). Both types of microorganisms require B for the stability of the envelopes that prevent access of nitrogenase-poisoning oxygen when grown under N₂-fixing conditions. Considering their different chemical composition, the stabilization of these envelopes by B is quite intriguing. In the heterocysts, B is present in an inner laminated layer formed of specific glycolipids. In contrast, in *Frankia*, B is found in a multilaminate vesicle wall composed of glycolipids and neutral lipids with a very high proportion of long-chain polyhydroxy, fatty acids or alcohols (Berry et al. 1991).

The discovery of a B-containing bacterial signal molecule, the auto-inducer AI-2, revealed a role for B in bacterial quorum sensing (Chen et al. 2002). Quorum sensing allows bacterial populations to monitor cell density, leading to coordinated alteration of gene expression, as the population increases (Coulthurst et al. 2002). The AI-2, identified as a furanosyl borate diester, is a signalling

molecule for both, structure and function. The gene encoding the AI-2 synthase (*luxS*) is widely conserved, raising the possibility that AI-2 might serve as a universal bacterial signal for communication among species (Chen et al. 2002, Winans 2002). This molecule could also serve as a B transporter, in a way that B can be moved in or out of the cell, depending on growth or environmental conditions (Coulthurst et al. 2002).

Several B containing compounds with antibiotic activity are produced by several bacteria, such as *Sorangium cellulosum* and *Streptomyces* sp. For instance, the *Streptomyces antibioticus* is responsible for the production of boromycin, an antibiotic that inhibits the growth of Gram-positive bacteria (Rezanka and Sigler 2008) and the replication of HIV-1 (Kohno et al. 1996). Boron deficiency significantly decreases *Saccharomyces cerevisiae* growth rate as well as population size. By analyzing log-phase mRNA of yeast grown without B, several genes were seen to be differentially expressed, namely those with functions in cell growth and maintenance (Becker et al. 2000, Bennett et al. 1999).

SCOPE OF THIS THESIS

The aim of this thesis is to increase the knowledge on the functions of B in higher plants. Two different plants, an important legume crop highly susceptible to B deficiency, *Lupinus albus*, and the model plant, *Arabidopsis thaliana*, were studied and several approaches were tested.

Since it is known that B participates in the cell wall structure, a first study consisted in the analysis of the protein profile of the leaf apoplast of B-deficient *L. albus* (Chapter 2).

Nutrient imbalance in soils is primary sensed by the root system, the organ responsible for sending signals to the shoot for growth modulation. Alterations in the protein pattern of *L. albus* roots due to B suppression were then analysed (Chapter 3).

Mineral stress can cause specific metabolic responses in the different plant organs. The analysis of metabolites of several organs of *L. albus* grown under B deficiency is described in Chapter 4.

To provide the insight into the early plant responses to B deficiency, a transcriptional analysis of *A. thaliana* was performed, as shown in Chapter 5.

The main conclusions of this work and proposed future perspectives are discussed in the last Chapter of this thesis (Chapter 6).

REFERENCES

- Armstrong T, Spears J, Crenshaw T, Nielsen F (2000) Boron supplementation of a semi purified diet for weanling pigs improves feed efficiency and bone strength characteristics and alters plasma lipid metabolites. *J Nutr* 130: 2575–2581.
- Bai Y, Hunt C (1996) Dietary boron enhances efficacy of cholecalciferol in broiler chicks. *J Trace Elem Exp Med* 9: 117–132.
- Bakken N, Hunt C (2003) Dietary boron decreases peak pancreatic *in situ* insulin release in chicks and plasma insulin concentrations in rats regardless of vitamin D or magnesium status. *J Nutr* 133: 3577–3583.
- Becker K, Bennett A, Eckhert C (2000) Identification by gene arrays of boron responsive genes in yeast, pp 1078. In: Trace elements in man and animals 10. Eds: Springer U, Roussel A, Anderson R, Favrier A. Kluwer Academic Publishers, NY.
- Berry A, Moreau R, Jones A (1991) Bacteriohopanetetrol: abundant lipid in *Frankia* cells and in nitrogen-fixing nodule tissue. *Plant Physiol* 95: 111–115.
- Behrendt U, Zoglauer K (1996) Boron controls suspensor development in embryogenic cultures of *Larix decidua*. *Physiol Plant* 97: 321–326.
- Bennett A, Rowe R, Soch N, Eckhert C (1999) B stimulates yeast (*Saccharomyces cerevisiae*) growth. *J Nutr* 129: 2236–2238.

Blevins D, Lukaszewski K (1998) Boron in plant structure and function. *Annu Rev Plant Physiol Plant Mol Biol* 49: 481–500.

Boesekem J (1949) The use of boric acid for the determination of the configuration of carbohydrates. *Adv Carbohydr Chem* 4: 189–210.

Bolaños L, Brevin N, Bonilla I (1996) Effects of B on Rhizobium–legume cell–surface interactions and nodule development. *Plant Physiol* 110: 1249–1256.

Bolaños L, Esteban E, Lorenzo C, Fernández–Pascual M, Felipe M, Gárate A, Bonilla I (1994) Essentiality of B for symbiotic dinitrogen fixation in pea (*Pisum sativum*)–Rhizobium nodules. *Plant Physiol* 104: 85–90.

Bolaños L, Lukaszewski K, Bonilla I, Blevins D (2004) Why boron? *Plant Physiol Biochem* 42: 907–912.

★ A review about B chemistry and deficiency symptoms.

Bolaños L, Mateo P, Bonilla I (1993) Calcium–mediated recovery of B deficient *Anabaena* sp. PCC 7119 grown under nitrogen fixing conditions. *J Plant Physiol* 142: 513–517.

Bonilla I, Blevins D, Bolaños L (2009) Boron Functions in Plants: Looking Beyond the Cell Wall. In: *Plant Physiology*. Eds: Taiz L, Zeiger E. 4th Ed. IOP Publishing physics web. <http://4e.plantphys.net>. Accessed Dec 2009.

★ A concise review about B deficiency symptoms.

Bonilla I, García–González M, Mateo P (1990) B requirement in Cyanobacteria. Its possible role in the early evolution of photosynthetic organisms. *Plant Physiol* 94: 1554–1560.

Bonilla I, Mergold-Villasenor C, Campos M E, Sánchez N, Pérez H, López L, Castrejon L, Sanchez F (1997) The aberrant cell walls of B-deficient bean root nodules have no covalently bound hydroxyproline/proline rich proteins. *Plant Physiol* 115: 1329-1340.

Brown P, Bellaloui N, Wimmer M, Bassil E, Ruiz J, Hu H, Pfeiffer H, Dannel F, Romheld V (2002) Boron in plant biology. *Plant Biol* 4: 203-223.

★★ An excellent review about B deficiency symptoms.

Cakmak I, Römheld V (1997) Boron deficiency-induced impairments of cellular functions in plants. *Plant Soil* 193: 71-83.

Chen X, Schauder S, Potier N, Van Dorsselaer A, Pelczer I, Bassler B, Hughson F M (2002) Structural identification of a bacterial quorum-sensing signal containing B. *Nature* 415: 545-549.

Coulthurst S, Whitehead N, Welch M, Salmond G (2002) Can B get bacteria talking? *Trends Biochem Sci* 27: 217-219.

Domingues D, Leite S, Farro A, Coscrato V, Mori E, Furtado E, Wilcken C, Velini E, Guerrini I, Maia I, Marino C (2005) Boron transport in Eucalyptus. 2. Identification *in silico* of a putative boron transporter for xylem loading in eucalypt. *Genet Mol Biol* 28: 625-629.

Dordas C, Brown P (2000) Permeability of boric acid across lipid bilayers and factors affecting it. *J Membrane Biol* 175: 95-105.

El-Shintinawy F (1999) Structural and Functional Damage Caused by Boron Deficiency in Sunflower Leaves. *Photosynthetica* 36: 565-573.

- Fort D, Stover E, Strong P, Murray F, Keen C (1999) Chronic feeding of a low B diet adversely affects reproduction and development in *Xenopus laevis*. J Nut 129: 2055-2060.
- Goldbach H, Wimmer M (2007) Boron in plants and animals: Is there a role beyond cell-wall structure? J Plant Nutr Soil Sci 170: 39-48.
- González-Fontes A, Rexach J, Navarro-Gochicoa M, Herrera-Rodríguez M, Beato V, Maldonado J, Camacho-Cristóbal J (2008). Is boron involved solely in structural roles in vascular plants? Plant Signal Behav 3: 24-26.
- Greenwood N, Earnshaw A (1984) Chemistry of the Elements, Pergamon Press, NY.
- Hegsted M, Keenan M, Siver F, Wozniak P (1991) Effect of boron on vitamin D deficient rats. Biol Trace Elem Res 28: 243-255.
- Hora J, Kivits G (1981) Stabilized enzymatic liquid detergent composition containing a polyalkanolamine and a boron compound. US Patent 4261868.
- Huang L, Bell R, Dell B (2008) Evidence of phloem boron transport in response to interrupted boron supply in white lupin (*Lupinus albus* L. cv. Kiev Mutant) at the reproductive stage. J Exp Bot 59: 575-583.
- Ishii T, Matsunaga T (2001) Pectic polysaccharide rhamnogalacturonan II is covalently linked to homogalacturonan. Phytochem 57: 969-974.
- Iwai H, Masaoka N, Ishii T, Satoh S (2002) A pectin glucuronyltransferase gene is essential for intercellular attachment in the plant meristem. PNAS 99: 16319-16324.

- Kasajima I, Ide Y, Hirai M, Fujiwara T (2010) WRKY6 is involved in the response to boron deficiency in *Arabidopsis thaliana*. *Physiol Plant* (*in press*).
- Kastori R, Plesnicar M, Pankovic D, Sakac Z (1995) Photosynthesis, chlorophyll fluorescence and soluble carbohydrates in sunflower leaves as affected by boron deficiency. *J Plant Nutr* 18: 1751-1763.
- Kobayashi M, Matoh T, Azuma J (1996) Two chains of rhamnogalacturonan II are cross-linked by borate-diol ester bonds in higher plant cell walls, *Plant Physiol* 110: 1017-1020.
- Kobayashi M, Nakagawa H, Asaka T, Matoh T (1999) Borate rhamnogalacturonan II bonding reinforced by Ca^{2+} retains pectic polysaccharides in higher plant cell walls. *Plant Physiol* 119: 199-204.
- Kohno J, Kawahata T, Otake T, Morimoto M, Mori H, Ueba N, Nishio M, Kinumaki A, Komatsubara S, Kawashima K (1996) Boromycin, an anti-HIV antibiotic. *Biosci Biotechnol Biochem* 60: 1036-1037.
- Koshiba T, Kobayashi M, Matoh T (2009) Boron Nutrition of Tobacco BY-2 Cells. V. Oxidative Damage is the Major Cause of Cell Death Induced by Boron Deprivation. *Plant Cell Physiol* 50: 26-36.
- Loomis W, Durst R (1992) Chemistry and biology of boron. *Biofactors* 3: 229-239.
- Martini F, Thellier M (1993) Boron distribution in parenchyma cells of clover leaves. *Plant Physiol Biochem* 31: 777-786.
- Nable R, Bañuelos G, Paul J (1997) Boron toxicity. *Plant Soil* 193: 181-198.

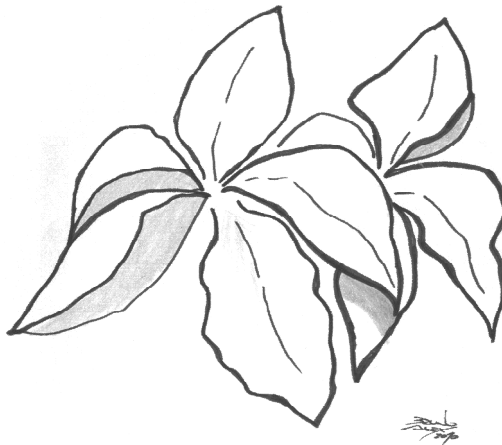
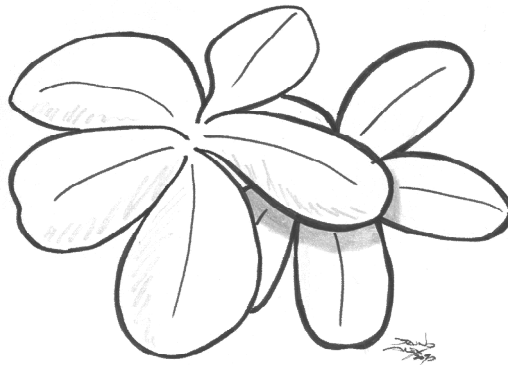
- Nakagawa Y, Hanaoka H, Kobayashi M, Miyoshi K, Miwa K, Fujiwara T (2007) Cell-type specificity of the expression of Os BOR1, a rice efflux boron transporter gene, is regulated in response to boron availability for efficient boron uptake and xylem loading. *Plant Cell* 19: 2624-2635.
- Nielsen F (1997) Boron in human and animal nutrition. *Plant Soil* 193: 199-208.
- O'Neill M, Eberhard S, Albersheim P, Darvill A (2001) Requirement of borate cross-linking of cell wall rhamnogalacturonan II for *Arabidopsis* growth. *Science* 294: 846-849.
- O'Neill M, Warrenfeltz D, Kates K, Pellerin P, Doco T, Darvill A, Albersheim P (1996) Rhamnogalacturonan-II, a pectic polysaccharide in the walls of growing plant cell, forms a dimer that is covalently cross linked by a borate ester. *J Biol Chem* 271: 22923-22930.
- Park M, Li Q, Shcheynikov N, Muallem S, Zeng W (2005): Borate transport and cell growth and proliferation: not only in plants. *Cell Cycle* 4: 24-26.
- Power P, Woods W (1997) The chemistry of boron and its speciation in plants. *Plant Soil* 193: 1-13.
- Prochaska J, Howk J, Wolfe A (2003) The elemental abundance pattern in a galaxy at $z = 2.626$. *Nature* 423: 57-59.
- Raven J (1980) Short-distance and long-distance transport of boric acid in plants. *New Phytol* 84: 231-249.
- Redondo-Nieto M, Rivilla R, El-Hamdaoui A, Bonilla I, Bolaños L (2001) B deficiency affects early infection events in the pea-*Rhizobium* symbiotic interaction. *Aust J Plant Physiol* 28: 819-823.

- Reeves H (2001) The origin of the light elements in the early Universe, pp 423–440. In: The Century of Space Science, Kluwer Academic Publishers, Dordrecht.
- Rezanka T, Sigler K (2008) Biologically active compounds of semi-metals, *Phytochem* 69: 585–606.
- Rowe R, Eckhert C (1999) Boron is required for zebrafish embryogenesis. *J Exp Bot* 202: 1649–1654.
- Severson R (1985) Liquid detergents containing boric acid and formate to stabilize enzymes. US Patent 4537707.
- Takano J, Miwa K, Fujiwara T (2008) Boron transport mechanisms: collaboration of channels and transporters. *Trends Plant Sci.* 13: 451–457.
- Takano J, Miwa K, Yuan LX, von Wirén N, Fujiwara T (2005). Endocytosis and degradation of BOR1, a boron transporter of *Arabidopsis thaliana*, regulated by boron availability. *PNAS* 102: 12276–12281.
- Takano J, Noguchi K, Yasumori M, Kobayashi M, Gajdos Z, Miwa K, Hayashi H, Yoneyama T, Fujiwara T (2002) Arabidopsis boron transporter for xylem loading. *Nature* 420: 337–340.
- Takano J, Wada M, Ludewig U, Schaaf G, von Wirén N, Fujiwara T (2006). The Arabidopsis major intrinsic protein NIP5;1 is essential for efficient boron uptake and plant development under boron limitation. *Plant Cell* 18, 1498–1509.
- Tanaka M, Wallace I, Takano J, Roberts D, Fujiwara T (2008) NIP6;1 is a boric acid channel for preferential transport of boron to growing shoot tissues in Arabidopsis. *Plant Cell* 20: 2860–2875.

- Verstraeten S, Lanoue L, Keen C, Oteiza P (2005) Relevance of lipid polar headgroups on boron-mediated changes in membrane physical properties. Arch Biochem Biophys 438: 103–110.
- Warington K (1923) The effect of boric acid and borax on the broad bean and certain other plants. Ann Bot 37: 629–672.
- WHO (1998) Boron. In: International Programme on Chemical Safety (Environmental Health Criteria 204). World Health Organization, Geneva (<http://www.inchem.org/documents/ehc/ehc/ehc204.htm>).
- Winans S (2002) Bacterial Esperanto. Nat Struct Biol 9: 83–84.
- Woods W (1996) A review of possible boron speciation relating to its essentiality. J Tr Elem Exp Med 9: 153–163.
- Yang W, Gao X, Wang B (2003) Boronic acid compounds as potential pharmaceutical agents. Med Res Rev 23: 346–368.
- Ye Z, Huang L, Bell R, Dell D (2003) Low root zone temperature favours shoot B partitioning in young leaves of oilseed rape (*Brassica napus*). Physiol Plant 118: 213–220.

CHAPTER 2

ANALYSIS OF *Lupinus albus* LEAF APOPLASTIC PROTEINS IN RESPONSE TO BORON DEFICIENCY



Analysis of *Lupinus albus* leaf apoplastic proteins in response to boron deficiency

Alves M^{1,@}, Francisco R^{1,@}, Martins I^{1,@}, Ricardo CP^{1, *}

¹Instituto de Tecnologia Química e Biológica, Universidade Nova de Lisboa,
Av. da República – EAN, 2780-157, Oeiras, Portugal.

@These authors contributed equally to this paper

*Corresponding author: ricardo@itqb.unl.pt

Reformatted version of the article in: Plant Soil (2006) 279: 1–11.

CONTRIBUTION: The three first authors contributed equally for the plant growth, harvesting, electrophoresis technique and writing.

KEYWORDS: Apoplast, B deficiency, *Lupinus albus*, proteomics, PR-proteins.

SUMMARY

Boron (B) is essential for plant development and although its precise functions are not fully understood there is evidence for its importance in the extracellular matrix. So we have analysed by two-dimensional gel electrophoresis the effect of B deficiency in the soluble apoplast proteins of *Lupinus albus* leaf. Twenty-three polypeptide spots varied significantly between the control and the B deficiency patterns. Of these polypeptides only 9 could be identified by mass spectrometry techniques: PR-1 like protein, β -1,3-glucanases, class III chitinases, thaumatin like proteins and an expansin-like protein, all of them being involved in plant defence mechanisms. Only PR-1 like protein was *de novo* expressed under B deficiency, while the remaining proteins also responded to water stress. Although general response mechanisms seem to be triggered by both B and water stress, the pattern of protein expression was distinct, suggesting that under B deficiency specific regulatory mechanisms may be induced.

INTRODUCTION

Boron is an essential micronutrient for vascular plants and some green algae, but apparently is not required by fungi and bacteria (Marschner 1995). The role of this element in plant nutrition is still

the least understood of all the mineral nutrients, which is surprising since on a molar basis its requirement is, at least for dicotyledons, higher than that of any other micronutrient (Marschner 1995). Boron is mainly localized in the plant cell wall, in fact, more than 90% of the total B in the cell is in that compartment (Blevins and Lukaszewski 1998), where it acts as a cross-linker (Matoh 1997, O'Neill et al. 1996, 2001). Matoh et al. (1993) isolated B-polysaccharide complexes from radish cell walls revealing that B was linked to rhamnogalacturonan-II molecules. This specific role of B is considered to be required for normal leaf expansion, but does not seem to explain all the observed effects of B deficiency in plant physiology (Dell and Huang 1997, Brown et al. 2002). Indeed, there is growing evidence showing that B plays essential roles in a number of membrane features and in the metabolism of carbohydrates, nucleic acids, indolacetic acid and phenols (Cakmak and Römheld 1997, Blevins and Lukaszewski 1998). It has been suggested that under B deficiency, phenolics concentrate through the stimulation of the enzymes phenylalanine-ammonium lyase and polyphenoloxidase (Cakmak and Römheld, 1997). Boron is also involved in lignification and nodulation processes (Bolaños et al. 1994).

The apoplast, that includes the cell wall, the intercellular spaces and other components external to the plasma membrane, is a dynamic

compartment continuously changing during growth and development (Dietz 1997, Pignocchi and Foyer 2003). Due to the participation of B in the cell wall structure, the plant response to B deficiency is likely to induce changes in the protein profile of the apoplast. Such apoplastic response was observed in a number of stress conditions, such as osmotic stress (Marshall et al. 1999), heavy metal toxicity (Fecht-Christoffers et al. 2003, Kataoka et al. 2003), or cold stress (Marents et al. 1993). To get an insight into the apoplastic soluble proteins that react to B deficiency in *L. albus* leaves, we have analysed them by two-dimensional electrophoresis (2-DE) and mass spectrometry techniques. The identification of these proteins may give valuable information for understanding the metabolic processes that may be affected by B deficiency.

MATERIALS AND METHODS

Plant material

Lupin seeds (*Lupinus albus* cv. Rio Maior) pre-germinated in distilled water for 48h were sown in white sand and grown for 28 days under controlled temperature (19/25°C, night/day), photoperiod (12h) and light intensity ($250\mu\text{mol m}^{-2} \text{s}^{-1}$, PAR). The plants were watered every other day with a nutrient solution (Arnon 1938) containing 0, 0.05, 0.46, 11.5, 23.1 or $46.1\mu\text{M}$ B.

The water-deficit was induced in 25 day old plants by suppressing watering for 3 days. The control plants were grown for 28 days with a suitable B concentration ($23.1\mu\text{M B}$). The leaf water potential was measured with a Sholander pressure chamber (PMS instrument Co, Corvallis, Oregon, USA) at pre-dawn (ψ_{pd}). At least 10 plants per experiment were analysed, in two independent experiments. Dry weight of lupin seeds was determined after oven drying for 48h at 80°C .

Boron determination

Boron content in lupin seeds and in several substrates for plant growth (white sand, peat, vermiculite and perlite) was analysed by inductively coupled plasma-atomic emission spectrometry (ICP-AES) at RAIZ (Instituto de Investigação da Floresta e Papel, Eixo, Portugal). The seeds were ashed overnight in an oven at 500°C and the ashes dissolved in 3N HCl. The substrates were extracted with 3N HCl (1:1) by shaking for 48h.

Extraction of soluble apoplastic proteins

Expanded leaves from 28 day old plants were used for the extraction of the soluble apoplastic proteins. This extraction was made using a vacuum infiltration/centrifugation technique based on the method of Parent and Asselin (1984) and Regalado and

Ricardo (1996). The leaf pieces were washed with distilled water, infiltrated in vacuum with 0.025M Tris-HCl pH 8.0 buffer, for 5 periods of 30s each and carefully blotted and centrifuged at 1480 \times g for 15min at 4°C to collect the infiltrate. The protein samples were concentrated on a speed-vac and stored at -20°C before use. Malate dehydrogenase (MDH, EC 1.1.1.37) was used as a cytosolic contamination marker of the apoplastic fluid, according to López-Millán et al. (2000).

Two-dimensional gel electrophoresis (2-DE)

Protein concentration in the extracts was determined according to Bradford (1976) and modified by Ramagli (1999). For isoelectric focusing (IEF) electrophoresis, the IPGphor system was used (Amersham Biosciences) with non linear (NL) pH gradient gels of 3–10 and 3–5.6 (IPGstrips, Amersham Biosciences) loaded with 25 μ g or 60 μ g of protein, respectively. Proteins were solubilized in 8M urea, 2% (w/v) CHAPS, 0.04M DTT and 0.5% (v/v) IPG buffer 3–10NL or/and 3.5–5NL (Amersham Biosciences). IEF was carried out at 30V for 12h, followed by 200V for 1h, 500V for 1.5h, 1000V for 1.5h, and 8000V for 6.5h, at 20°C. Prior to SDS-PAGE the IPGstrips were equilibrated for 2x15min in a buffer solution containing 0.05M Tris-HCl pH 8.8, 6M urea, 30% (v/v) glycerol, 2% (w/v) SDS and a trace of Bromophenol Blue. To the

first equilibration step DTT at 1% (w/v) was added and to the second one iodoacetamide at 4% (w/v). The SDS-PAGE was performed on slab gels (Laemmli 1970). The 2-DE gels were stained with colloidal Coomassie Blue (Neuhoff et al. 1985) or silver stained (Blum et al. 1987) and scanned using the ImageQuant v3.3 densitometer (Molecular Dynamics). After imaging, the gels were analysed by ImageMaster 2D Platinum software v5.0 (Amersham Biosciences). This analysis allowed to do spot detection, spot measurement, background subtraction and spot matching. The amount of a protein spot was expressed as the volume of that spot defined as the sum of the intensities of all the pixels of a spot. In order to correct the variability due to gel staining and to reflect the quantitative variations of protein spots intensity, the spot volumes were normalized as a percentage of the total volume in all of the spots present in the gel. To compare differences in protein abundance among the different samples, a one-way ANOVA ($p < 0.05$) was performed for each spot, using the Ludesi software 2-DE interpreter (<http://www.ludesi.com>).

Protein identification by MS/MS

Spots were excised from colloidal Coomassie Blue stained gels and analysed by MALDI-TOF-TOF (matrix assisted laser desorption/ionization time of flight-time of flight) at the Technology

Facility, Department of Biology, University of York (UK). Database searching Similarity searching with the obtained partial amino acid sequences was performed at the EMBL-EBI server (<http://www.dove.embl-heidelberg.de/Blast2/>).

RESULTS

Determination of suitable B concentration for L. albus growth

Since it was intended to induce B deficiency in the lupin plant, it was necessary to choose an adequate substrate with low B content to grow the plants. Vermiculite, peat, perlite and white sand were analysed and their B content (nmol B/ g dry weight), were found to be respectively, 530 ± 30 , 30 ± 4 , 17 ± 2 and 1.8 ± 0.1 . So, white sand was considered the best substrate. The Arnon (1938) nutrient solution was used in the experiments after checking that its B concentration ($46.1 \mu\text{M}$) was adequate for *L. albus* growth. Considering that lupin seed contained $1.31 \pm 0.06 \mu\text{mol B/g dry weight}$, an unavoidable source of B, for seeds with a mean dry weight of 0.5g, even when grown at $0 \mu\text{M B}$, the lupine plant would have at its disposal *ca.* $0.65 \mu\text{mol B}$. Several experiments were performed in which B concentration in the nutrient solution was varied: 0, 0.05, 0.46, 11.5, 23.1 and $46.1 \mu\text{M}$. The total biomass and the root/shoot ratio were determined under those conditions

(Figure 2.1). The B concentration in a range of 11.5–23.1 μM was not limiting *L. albus* growth and 46.1 μM B was marginally excessive. So we chose half the B concentration of Arnon's solution (23.1 μM B) as a suitable concentration for *L. albus* growth.

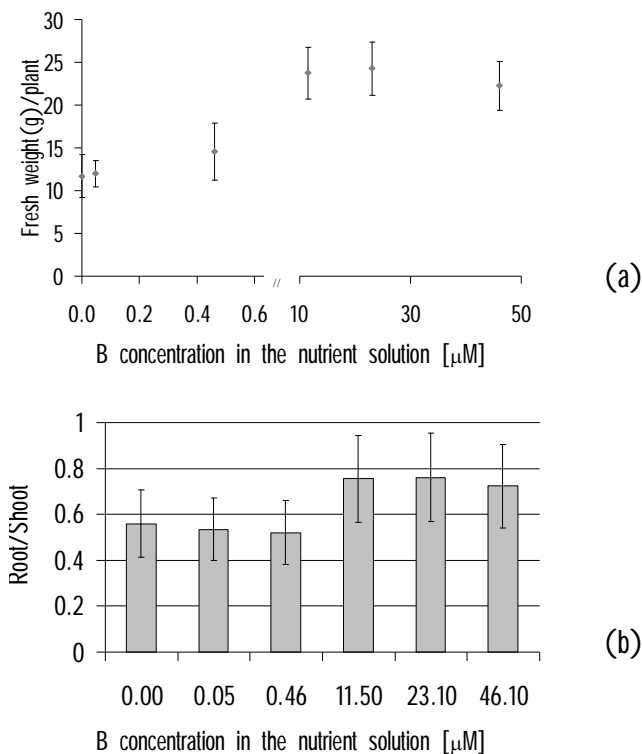


Figure 2.1. Effects of B concentrations in the nutrient solution on *L. albus* biomass production; (a) total biomass, (b) root/shoot ratio. At least ten plants per experiment were analysed, in two independent experiments and bars indicate standard errors.

Morphological and physiological effects of B deficiency

Morphological differences between the plants grown with 0 and 23.1 μM B started to be evident from the 3rd to the 4th week from



(a)

sowing (Figure 2.2). The most evident deficiency symptoms were necrosis of the terminal buds, darkened



(b)

Figure 2.2. Morphological differences of *L. albus* plants grown in white sand culture for 28 days, with (a) 23.1 μ M B or (b) 0 μ M B in the nutrient solution.

and structurally deformed leaves, cracking and breaking stems and petioles, and the absence of secondary and proteoid roots. It was also verified that the cotyledons became thicker and dark green and that their life span was extended. Despite the dramatic morphological effects, only a small decrease in ψ_{pd} was observed in B deficient plants when compared to that of water-deficit plants (Figure 2.3).

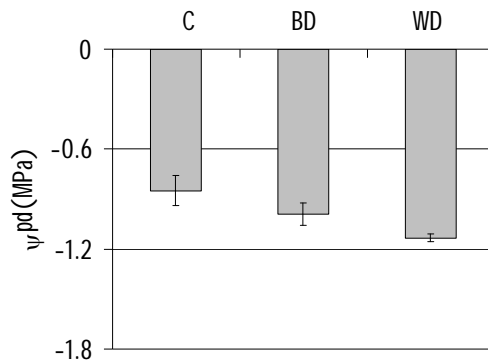


Figure 2.3. Leaf water potential (ψ_{pd}) of 28 days old *L. albus*: control plants (Control), B deficient (BD) and water-deficit (WD) plants (bars indicating standard errors). The water potential was determined by Sholander pressure chamber measurements on the 5th leaf before the beginning of the illumination period.

Protein analysis by 2-DE

The activity of MDH was used as a marker of the contamination of the apoplastic samples with cytoplasm proteins. It was found that in

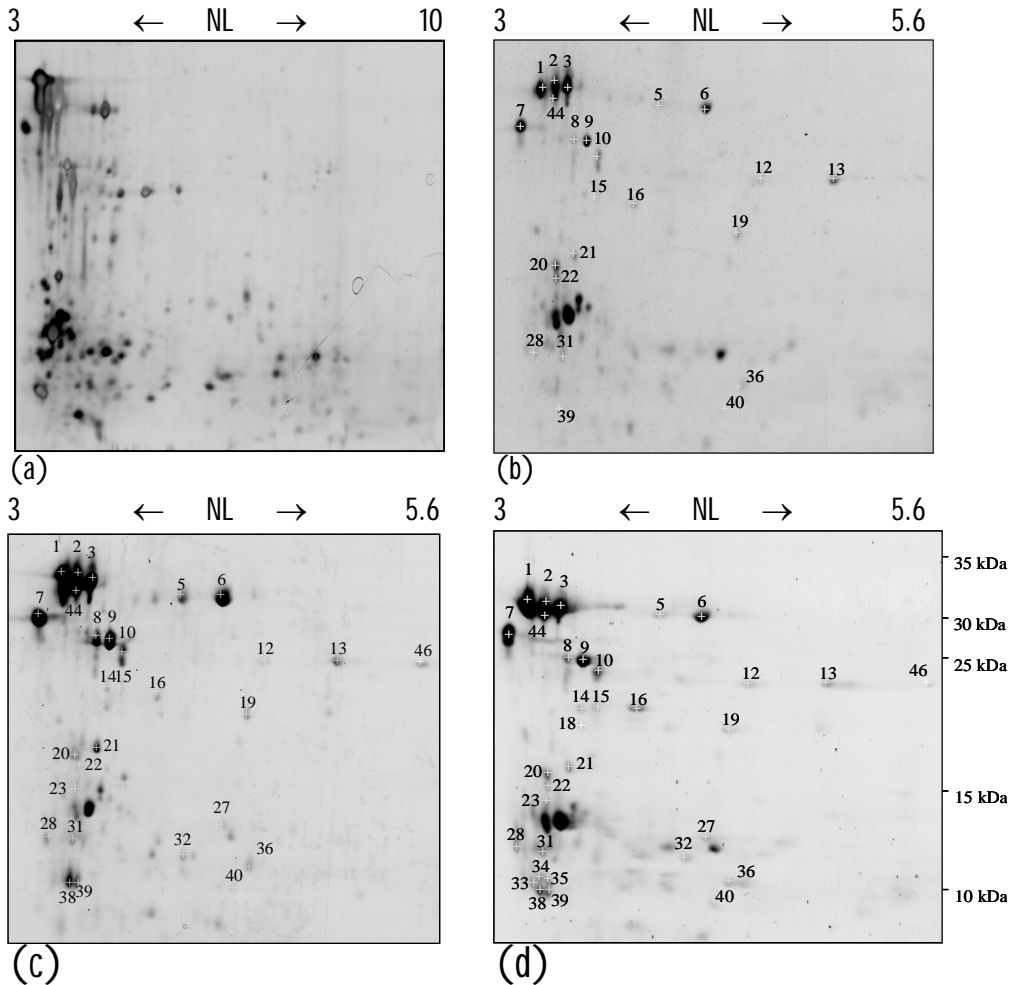


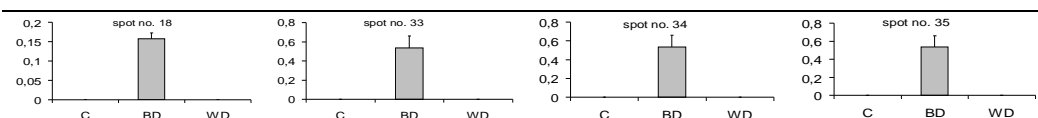
Figure 2.4. 2-DE gels from *L. albus* apoplast leaf proteins obtained from control (a, b), water deficit (c) and boron deficiency (d) plants. Gel (a) was loaded with 25 μ g of proteins which were resolved using a NL pH gradient 3-10 and silver stained; gels (b, c, d) were loaded with 60 μ g proteins which were resolved using a NL pH gradient 3.0-5.6 and colloidal Coomassie Blue stained. Labelled proteins shown in the gels were selected for MS/MS identification.

the apoplastic samples this enzymatic activity was less than 3% (mean value of 4 replicates) of the activity of leaf homogenates. This indicates a negligible apoplastic contamination with cytoplasmatic components. 2-DE was carried out using IPGstrips with a pH gradient of 3-10NL and it was observed that the major soluble apoplastic proteins have an acidic pI (Figure 2.4a). Consequently a narrow pH gradient of 3-5.6NL was used to perform all the subsequent studies. The apoplastic soluble proteins collected from plants under water-deficit were used for a comparative purpose in order to identify the proteins that responded to general stress conditions (Pinheiro et al. 2005). The obtained protein profiles were analysed with ImageMaster 2D Platinum Software v5.0 for detection and matching of the protein spots and it was observed that 51 spots were present in the control and water-deficit gels and 55 spots in the B deficiency gels (Figure 4b-d). The statistical analysis made with Ludesi software 2-DE interpreter identified the polypeptides which were significantly altered by the imposition of the stress conditions ($p < 0.05$) (Table 2.1).

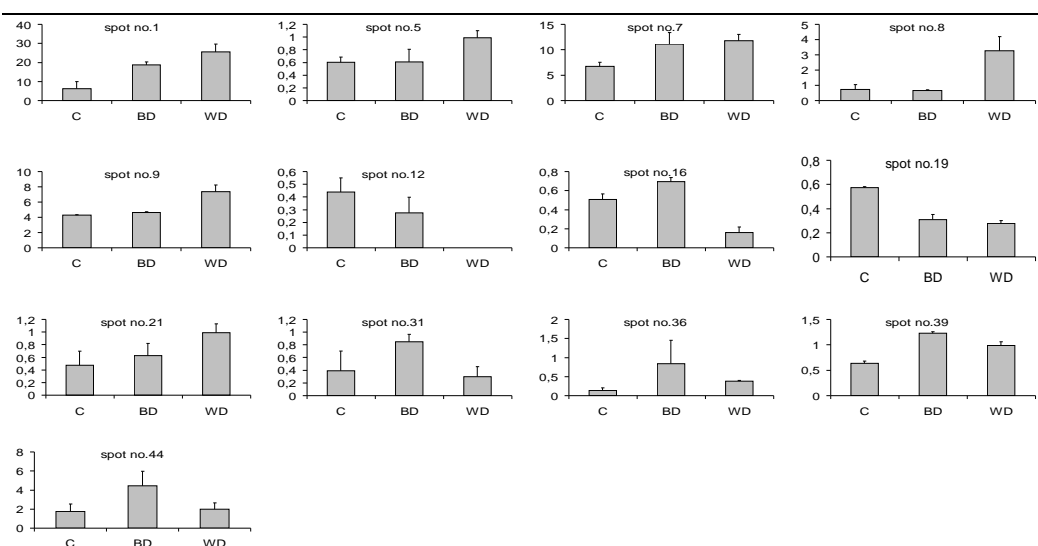
Three groups of polypeptides with particular expression patterns relative to the control could be defined: i) *de novo* expressed under B deficiency (4 polypeptides), ii) *de novo* expressed under B deficiency and/or water-deficit (6 polypeptides) and iii) already

Table 2.1. Relative abundances (% of normalized volume) of the individual apoplast spots whose levels were significantly altered in the stress conditions (bars indicating standard errors). C, control; BD, B deficiency; WD, Water-deficit.

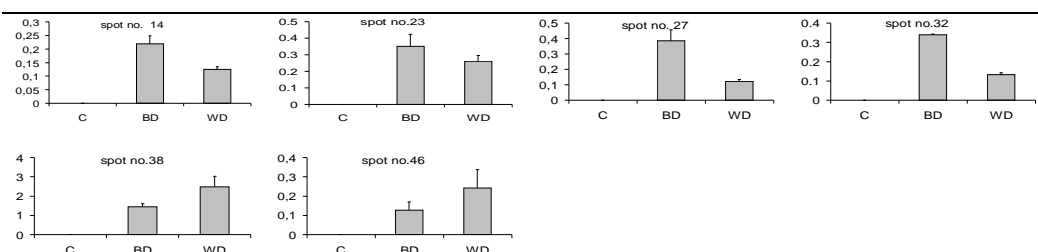
***De novo* expressed under BD**



Responsive to BD and/or WD



***De novo* expressed under BD and WD**



present in the control but responsive to B deficiency and water-deficit (13 polypeptides). MS/MS methods were used to try to

identify these spots and others unaffected by the imposed stresses (spots 2, 3, 6, 10, 13, 15, 22 and 40).

Protein identification by MS/MS

Thirty one spots were selected for protein identification by MS/MS. We were able to identify proteins of 12 spots: two glucanase isoforms (Q01412), three acidic chitinase isoforms (O49876), four thaumatin-like proteins (P83332, P25096, Q8LKA8, O80327), one PR-1 like protein (Q9S9G8), one expansin-like protein (Q6K4C6) and one putative secretory protein (Q9FWU3) (Table 2.2). All these identified proteins are extracellular, since the presence of the signal peptide was predicted when using SignalP software (<http://www.cbs.dtu.dk/services/SignalP>). The spot 13 which is an exception, since it had no predicted signal peptide, was nonetheless identified as an extracellular protein. The polypeptides of the remaining 19 spots, although sequenced were not identified, which may be explained by the scarcity of information existing in lupin sequence databases.

Table 2.2. MS/MS identifications of *L. albus* leaf apoplast proteins resolved by 2-DE. C, control; BD, B deficiency; WD, water-deficit.

| Spot | Sequence ^b (TrEMBL Ac No.) ^c | pI ^d | MW ^d |
|--|---|-----------------|-----------------|
| <i>De novo</i> expressed under BD | | | |
| 33 ions^a | PR-1 like protein (Q9S9G8) | 9.45 | 10.8 |
| | 1126.56 LLTHGPGFNGN | | |
| | 1244.51 EPSKADYLGKH | | |
| | 1656.73 DSKADYLNHNAAR | | |
| | 1731.81 KGMSDGKDKGVDPFGHG | | |
| Responsive to BD and/or WD | | | |
| 1 ions^a | Endo- β -1,3-glucanase (Q01412) | 6.67 | 35.2 |
| | 1433.65 LYSPDEGTLOALK | | |
| | 1543.70 YLAVGNEVEPNDPK | | |
| | 2168.98 (LFA)RFETYLFAMFDENR | | |
| 7 ions^a | Class III chitinase (O49876) | 4.48 | 28.2 |
| | 1110.50 FNDLQTGYSDALK | | |
| | 1470.65 YGGVMLWNR | | |
| 8 ions^a | Class III chitinase (O49876) | 4.48 | 28.2 |
| | 1126.55 LITHPGGFNGN | | |
| | 1471.70 FNDLKTGYSDALK | | |
| | 2891.35 SSKGGADLVLCAGGAELDFNNGPDDAGKGA | | |
| 9 ions^a | Class III chitinase (Q945U2) | 9.18 | 30.9 |
| | 952.467 ALDGFSSKK | | |
| | 1755.87 AAGTFGDTKGEPLNLAHG | | |
| | 2891.35 RYPNFHGLDFDLEAGGMRLVDENPE | | |
| | 3336.95 SRTLLVVCVAAAAPSGGFMWAHLLSKVLLTPK | | |
| 16 ions^a | Thaumatococin-like protein (P25096) | 4.84 | 21.5 |
| | 1562.71 GCTADINGKGCPSVLK | | |
| | 1819.74 DDKTFGGMFGPGGTNYR | | |
| | 2198.81 DTPLLHVPSLPYKSEPYSR | | |
| 31 ions^a | Osmotin-like protein (Q8LKA8) | 4.58 | 39.6 |
| | 1100.46 CGPDAYSYPK | | |
| | 1562.72 GCTADLNGKGCSPVLK | | |
| | 1801.74 VLKTSTFFGGPPECYR | | |
| | 1819.75 VMKTSTFTCVDPATPYR | | |
| 39 ions^a | Putative blight-associated protein/ Expansin like protein (Q6K4C6) | 8.25 | 13.4 |
| | 1731.81 LTCTGATNKGVPKPKCK | | |
| | 1747.81 DDCTKTNKGVPKPKCK | | |
| | 959.503 TGLADLNAGK | | |
| | 2807.32 DCLLWLASMAEKSLDLTGVSVSDDPP | | |

Table 2.2 (cont) MS/MS identifications of *L. albus* leaf apoplast proteins resolved by 2-DE.

C, control; BD, B deficiency; WD, water-deficit.

| Spot | Sequence ^b (TrEMBL Ac No.) ^c | pI ^d | MW ^d |
|-----------------------------------|--|-----------------|-----------------|
| 44 | Thaumatococcus-like protein (O80327) | 5.01 | 22.9 |
| ^a ions | 882.381 STEMEGGR | | |
| | 979.402 TGCSNNNGR | | |
| | 2119.01 STSCPANLNTVCLDVLKGGI | | |
| | 2327.99 CGNSGEKKCAKPGYMDCHK | | |
| Non responsive to BD or WD | | | |
| 2 | Thaumatococcus-like protein (P83332) | 8.14 | 23.3 |
| ^a ions | 1230.61 LSDLPAPAFSGR | | |
| | 1458.60 GTGCLAFNPKPKYCG | | |
| | 2119.01 ARAHPANLNTVGCNGGVKLGIG | | |
| | 2133.02 VCSAGEARGKTVELMNKVVA | | |
| 6 | Endo- β -1,3-glucanase (Q01412) | 6.67 | 35.2 |
| ^a ions | 1384.70 ASPTKDTCAGRFM | | |
| | 1544.76 YLAVGNEVEPGGDPK | | |
| | 2724.54 AKYVLPVLAGNLKALSSANIGAVGRK | | |
| | 2746.48 AKYVLPVNSSLANKLNAGLKLGLAYE | | |
| 13 | Putative secretory protein (Q9FWU3) | 7.89 | 28.7 |
| ^a ions | 1061.18 AGMDAKLNNAG | | |
| | 1172.35 KGYAPSHSVVK | | |
| | 1295.46 AEDKLWKDYK | | |
| | 1367.54 RMDELGEKYAR | | |
| | 1735.99 LTYCGTPPAMPTDGGVR | | |
| | 1767.96 APYANPKVWHSPSGTR | | |
| | 2567.97 CDDPNKLLGHRTMKNNLVTR | | |

^a Ion precursor (mass/charge unit)

^b Proteins were sequenced by TOF/TOF at University of York

^c Proteins were identified by <http://www.dove.embl-heidelberg.de/Blast2/>

^d pI and MW were calculated by <http://www.exapsy.org/>

DISCUSSION

The apoplast, as the prime barrier between the plant and its environment is a critical compartment for studying plant responses to stressful conditions. In fact our analysis of that compartment in *L. albus* leaves revealed marked alterations in response to B deficiency and water-deficit. Apoplastic protein alterations were also reported in tobacco under salt stress (Dani et al. 2005) and in cowpea subjected to manganese toxicity (Fecht-Christoffers et al. 2003). Although the majority of *L. albus* proteins analysed gave good MS/MS spectra, only a few could be identified. This result may reflect the reduced number of studies concerning the apoplast and the scarcity of information available for *L. albus* proteins in databases.

The identified proteins can all be related to stress and plant defence responses, namely of the pathogenesis related type. Both PR and PR-like proteins (Van Loon and Van Strien 1999) are known to be induced in plant tissues by several biotic and abiotic stresses, such as fungal infection (Niderman et al. 1995), heavy metal toxicity (Rakwal et al. 1999), salinity (Esaka et al. 1994), high temperature (Margis-Pinheiro et al. 1994) and wounding (Ruperti et al. 2002). In previous studies it was observed that in the *L. albus* leaf apoplast PR-like proteins are constitutively expressed (Pinto and Ricardo 1995, Regalado and Ricardo 1996). We now

found that endo- β -1,3-glucanases, thaumatin-like proteins and class III chitinases, were significantly increased under B deficiency as well as under water-deficit. The increased incorporation of glucose into the cell wall β -1,3-glucans, as a result of B deficiency may suggest a higher involvement of endo- β -1,3-glucanase in the cell wall metabolism under this stress (Dugger and Palmer 1985). Kobayashi *et al.* (2004) also observed an enhancement of endo- β -1,3-glucanase gene expression in tobacco cells under low B supply. In our study, of the 4 proteins *de novo* expressed under B deficiency only one could be identified, a PR-1-like protein belonging to a PR-1 gene family to which no putative function has been assigned (Green and Fluhr 1995).

Kobayashi *et al.* (2004) observed that B deficiency induces genes, namely of PR-like families, that are involved in the general response of plants to stressful conditions. The precise role of these several PR proteins in stress responses, and in particularly in B deficiency, is not fully understood. However it has been shown that some of them participate in specific processes such as antimicrobial activity (Broekaert *et al.* 1995, Epple *et al.* 1997) and antifreeze activity (Hon *et al.* 1995).

A putative blight associated protein, p12, described as an expansin-like protein (EXPL) soluble in the extracellular space (Kayim *et al.* 2004), was shown to increase under B deficiency. Expression of

expansin genes correlates with growth of cells, and its increase also occurs during fruit ripening (Cosgrove 1997). However, the functional and potential wall-loosening activity of the EXPL proteins has not been established (Cosgrove 2003).

The presence in *L. albus* apoplast of three chitinase isoforms and two glucanase isoforms, that could result from post-translational modifications of the proteins (Dani *et al.* 2005), appears to be of some relevance. Salzer *et al.* (2000) when studying fungal infection and symbiosis in *Medicago truncatula* concluded that protein isoforms may have a crucial role in plant response to stress. It is also well documented that in addition to stress-specific adaptive responses plants have some other types of responses that protect them from more than one environmental stress (Chinnusamy *et al.* 2003). In this connection, it is relevant to note that the proteins that responded to both B deficiency and water-deficit in *L. albus* apoplast had a different pattern of expression. This seems to suggest that specific regulatory mechanisms are involved in those distinct stress conditions.

ACKNOWLEDGEMENTS

The authors would like to thank Dr. Sérgio Fabres (RAIZ, Instituto de Investigação da Floresta e Papel) for the ICP-AES analyses,

Dr. Ana Rodrigues (ISA, Instituto Superior de Agronomia, Lisboa, Portugal) for the Sholander pressure chamber measurements. Financial support from Plant IBET project 2003-A is also acknowledged.

REFERENCES

- Arnon D (1938) Microelements in culture-solution experiment with higher plants. *Amer J Bot* 25: 322-325.
- Blevins D, Lukaszewski K (1998) Boron in plant structure and function. *Annu Rev Plant Physiol Plant Mol Biol* 49: 481-500.
- Blum H, Beier H, Gross H (1987) Improved silver staining of plant proteins, RNA and DNA in polyacrylamide gels. *Electrophoresis* 8: 93-99.
- Bolaños L, Esteban E, Lorenzo C, Pascual M, Felipe M, Gárate A, Bonilla I (1994) Essentiality of Boron for symbiotic dinitrogen fixation in pea (*Pisum sativum*) rhizobium nodules. *Plant Physiol* 104: 85-90.
- Bradford M (1976) A Rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem* 72: 248-254.
- Broekaert W, Terras F, Cammue B, Osborn R (1995) Plant defensins: novel antimicrobial peptides as components of the host defense system. *Plant Physiol* 108: 1353-1358.

- Brown P, Bellaloui N, Wimmer M, Bassil E, Ruiz J, Hu H, Pfeffer H, Dannel F, Römhel V (2002) Boron in Plant Biology. *Plant Biol* 4: 205-223.
- Cakmak I, Römhel V (1997) Boron deficiency-induced impairments of cellular functions in plants. *Plant Soil* 193: 71-83.
- Chinnusamy V, Schumaker K, Zhu J-K (2003) Molecular genetic perspectives on cross-talk and specificity in abiotic stress signalling in plants. *J Exp Bot* 55: 1-12.
- Cosgrove D (1997) Assembly and enlargement of the primary cell wall in plants. *Annu Rev Cell Dev Biol* 13: 171-201.
- Cosgrove D (2003) Expansion of the plant cell wall, pp 237-263. In: *The Plant Cell Wall*, Annual Plant Reviews Vol. 8. Eds: Rose J. Blackwell, Oxford.
- Dani V, Simon W, Duranti M, Croy R (2005) Changes in the tobacco leaf apoplast proteome in response to salt stress. *Proteomics* 5: 737-745.
- Dell B, Huang L (1997) Physiological response of plants to low boron. *Plant Soil* 193: 103-120.
- Dietz K-J (1997) Functions and responses of the leaf apoplast under stress. *Prog Bot* 58: 221-254.
- Dugger W, Palmer R (1985) Effect of boron on the incorporation glucose by cotton fibers grown *in vitro*. *J Plant Nutr* 8: 311-325.
- Epple P, Apel K, Bohlmann H (1997) Overexpression of an endogenous thionin enhances resistance of *Arabidopsis* against *Fusarium oxysporum*. *Plant Cell* 9: 509-520.

- Esaka M, Toyota A, Hayakawa H (1994) Secretion of basic and acidic chitinase from salt-adapted and -unadapted winged bean cells. *Physiol Plant* 92: 90-96.
- Fecht-Christoffers M, Braun H, Lemaitre-Guillier C, VanDorsseleer A, Horst W (2003) Effect of Manganese Toxicity on the Proteome of the Leaf Apoplast in Cowpea. *Plant Physiol* 133: 1935-1946.
- Green R, Fluhr R (1995) UV-B induced PR-1 accumulation is mediated by active oxygen species. *Plant Cell* 7: 203-212.
- Hon W, Griffith M, Mlynarz A, Kwok Y, Yang D (1995) Antifreeze proteins in winter rye are similar to pathogenesis-related proteins. *Plant Physiol* 190: 879-889.
- Kataoka T, Furukawa J, Nakanishi, T (2003) The decrease of extracted apoplast protein in soybean root tip by aluminium treatment. *Biol Plant* 36: 445-449.
- Kayim M, Ceccardi T, Berretta M, Barthe G, Derrick K (2004) Introduction of a citrus blight-associated gene into Carrizo citrange [*Citrus sinensis* (L.) Osbc. × *Poncirus trifoliata* (L.) Raf.] by *Agrobacterium*-mediated transformation. *Plant Cell Rep* 23: 377-385.
- Kobayashi M, Mutoh T, Matoh T (2004) Boron nutrition of cultured tobacco BY-2 cells IV. Genes induced under low boron supply. *J Exp Bot* 55: 1441-1443.
- Laemmli U (1970) Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* 227: 680-685.
- López-Millán A, Morales F, Abadía A, Abadía J (2000) Effects of Iron Deficiency on the Composition of the Leaf Apoplastic

Fluid and Xylem Sap in Sugar Beet. Implications for Iron and Carbon Transport. *Plant Physiol* 124: 873–884.

Marents E, Griffith M, Mlynarz A and Brush R (1993) Protein accumulate in the apoplast of winter rye leaves during cold acclimatation. *Physiol Plant* 87: 499–507.

Margis-Pinheiro M, Marivet J, Burkard G (1994) Bean class IV chitinase gene: structure, developmental expression and induction by heat stress. *Plant Sci* 98: 163–173.

Marschner H (1995) Boron, pp 379–396. In: Mineral nutrition of higher plants, Academic Press, London.

Marshall J, Dumbroff E, Thatcher B, Martin B, Rutledge R, Blumwald E (1999) Synthesis and oxidative insolubilization of cell-wall proteins during osmotic stress. *Planta* 208: 401–408.

Matoh T, Ishigaki K, Ohno K, Azuma J (1993) Isolation and characterization of a boron-polysaccharide complex from radish roots. *Plant Cell Physiol* 34: 639–642.

Matoh T (1997) Boron in plant cell walls. *Plant Soil* 193: 59–70.

Neuhoff V, Stamm R, Eibl H (1985) Clear background and highly sensitive protein staining with Coomassie Blue dyes in polyacrylamide gels: a systematic analysis. *Electrophoresis* 6: 427–448.

Niderman T, Genetet I, Bruyere T, Gees R, Stintzi A, Legrand M, Fritig B, Mosinger E (1995) Pathogenesis-related PR-1 proteins are antifungal (isolation and characterization of three 14-kilodalton proteins of tomato and of a basic PR-1 of tobacco with inhibitory activity against *Phytophthora infestans*). *Plant Physiol* 108: 17–27.

- O'Neill M, Eberhard S, Albersheim P, Darvill A (2001) Requirement of borate cross-linking of cell wall rhamnogalacturonan-II for Arabidopsis growth. *Science* 294: 846-849.
- O'Neill M, Warrenfeltz D, Kates K, Pellerin P, Doco T, Darvill A, Albersheim P (1996) Rhamnogalacturonan-II a pectic polysaccharide in walls of growing plant cell, forms a dimer that is covalently cross-linked by a borate ester. *J Biol Chem* 271: 22923-22930.
- Parent J-G, Asselin A (1984) Detection of pathogenesis-related proteins (PR or b) and of the other protein in the intercellular fluid of hypersensitive plants infected with tobacco mosaic virus. *Can J Bot* 62: 564-569.
- Pignocchi C, Foyer C (2003) Apoplastic ascorbate metabolism and its role in the regulation of cell signalling. *Curr Opin Plant Biol* 6: 379-389.
- Pinheiro C, Kehr J, Ricardo C (2005) Effects of water stress on lupin stem protein analysed by two-dimensional gel electrophoresis. *Planta* 221: 716-728.
- Pinto P, Ricardo C (1995) *Lupinus albus* L. pathogenesis-related proteins that show similarity to PR-10 proteins. *Plant Physiol* 109: 1345-1351.
- Rakwal R, Agrawal G, Yonekura M (1999) Separation of proteins from stressed rice (*Oryza sativa* L.) leaf tissues by two-dimensional polyacrylamide gel electrophoresis: induction of pathogenesis-related and cellular protectant proteins by jasmonic acid, UV irradiation and copper chloride. *Electrophoresis* 20: 3472-3478.
- Ramagli L (1999) Quantifying protein in 2D PAGE solubilization buffers, pp 95-105. In: *Methods in Molecular Biology - 2D*

Proteome Analysis Protocols. Eds. Andrew J. Link Humana Press, Totowa, New Jersey.

- Regalado A, Ricardo C (1996) Study of the intercellular fluid of healthy *Lupinus albus* organs. *Plant Physiol* 10: 227-232.
- Rupert B, Cattivelli L, Pagni S, Ramina A (2002) Ethylene-responsive genes are differentially regulated during abscission, organ senescence and wounding in peach (*Prunus persica*). *J Exp Bot* 53: 429-437.
- Salzer P, Bonanami A, Beyer K, Vögeli-Lange R, Aeschbacher R, Lange J, Wiemken A, Kim D, Cook D, Boller T (2000) Differential Expression of eight chitinase genes in *Medicago truncatula* roots during mycorrhiza formation, nodulation and pathogen infection. *Mol Plant-Microbe Interact* 13: 763-777.
- Van Loon L, Van Strien E (1999) The families of pathogenesis-related proteins, their activities, and comparative analysis of PR-1 type proteins. *Physiol Mol Plant Pathol* 55: 85-97.

CHAPTER 3

THE ANALYSIS OF *Lupinus albus* ROOT PROTEOME REVEALED CYTOSKELETON ALTERED FEATURES DUE TO LONG-TERM BORON DEFICIENCY



The analysis of *Lupinus albus* root proteome revealed cytoskeleton altered features due to long-term boron deficiency

Alves M¹, Moes S², Jenö P², Pinheiro C¹, Passarinho J³, Ricardo CP^{1,*}

¹Instituto de Tecnologia Química e Biológica, Universidade Nova de Lisboa, Av. da República, 2780-157 Oeiras, Portugal

²Biozentrum of the University of Basel, Klingelbergstrasse 50-70, CH-4056 Basel, Switzerland.

³Instituto Nacional de Recursos Biológicos/L-INIA, Av. da República, 2784-505 Oeiras, Portugal.

*Corresponding author: ricardo@itqb.unl.pt

CONTRIBUTION: The first author performed all the experimental work and writing, with the exception of the technical MS analysis.

KEYWORDS: Boron deficiency, cytoskeleton, *Lupinus albus*, proteomics, root.

SUMMARY

Boron (B) deficiency in plants greatly limits their growth and development. Since the role of B in plant biology is not completely understood, we have analysed the adaptive responses of *Lupinus albus* roots to a long-term B deficiency. For this purpose, two-dimensional electrophoresis and mass spectrometry techniques were used to compare the root proteome of plants treated with or without B. Despite the large morphological differences observed, from the more than 790 protein spots detected, only 262 were responsive to B deficiency. The suppression of the majority of these polypeptides reflects the reduction of the biosynthetic fluxes. The cytoskeletal biosynthesis was also affected by B deficiency, probably indicating an important contribution of B in this process.

INTRODUCTION

Boron (B) has long been known to be an essential micronutrient for higher plants (Warington 1923), but to date, its precise roles in essential plant biological processes remain elusive. So far, B is known to have a structural role in the cell wall, through the borate cross-linking of rhamnogalacturonan-II (Kobayashi et al. 1996, O'Neill et al. 1996, 2001), but a plethora of biochemical, physiological and anatomical effects due to B suppression are not

well explained by this role alone (Brown et al. 2002). Several authors proposed that B may be involvement in membrane features and cytoskeleton structural processes (Bassil et al. 2004, Bonilla et al. 2009). Since nutrient imbalance is primary sensed by the root system and because this is the organ responsible for sending signals to the shoots to modify growth and developmental processes (Fitter 2002), we have analysed the *Lupinus albus* L. plant root adaptive responses to long-term B deficiency. For this purpose, the root proteome of plants grown with and without B were analysed by two-dimensional electrophoresis (2-DE) and mass spectrometry (MS) techniques.

MATERIALS AND METHODS

Plant Material

Lupin seeds (*Lupinus albus* cv. Rio Maior) were pre-germinated in distilled water for 48h, sown in white sand and grown under controlled conditions of temperature (19/25°C, night/day), photoperiod (12h) and light intensity ($250\mu\text{mol m}^{-2} \text{s}^{-1}$, PAR). The plants were watered every other day with a nutritive solution (Arnon 1938) containing either 0 or $23.1\mu\text{M}$ B (Alves et al. 2006). The roots of *Lupinus albus* were harvested 28 days after sowing. Fresh weight statistical analysis was made with a Student's

t-test ($p < 0.05$) by using the SigmaStat v3.10 software (Systat Software, Inc.).

Protein extraction and solubilisation

Roots were ground to a fine powder in liquid nitrogen, resuspended in a cold acetone solution containing 60 mM DTT and 10% (w/v) TCA (12.5mL/g) and kept at -20°C for 1h. After a 15min centrifugation at $27,200\times g$ and 4°C , the pellet was resuspended in cold acetone with 60 mM DTT (25mL/g) and kept at -20°C for 1h. After centrifugation, the pellet was dried under vacuum and resuspended (0.05g/mL) in a solubilization buffer containing 2M thiourea, 7M urea, 4% (w/v) CHAPS, 0.4% (v/v) Triton X-100, 60 mM DTT and 1% (v/v) IPG Buffer 3–10 NL (GE). After 2h dissolution at room temperature, the protein extracts were centrifuged at $15,000\times g$ for 10min and the supernatant collected and stored at -80°C until further use. The Protein concentration was determined according to the Bradford method as modified by Ramagli (Ramagli 1999).

Two-dimensional gel electrophoresis

For isoelectric focusing (IEF) electrophoresis, the IPGphor system was used (Amersham Biosciences) with a non-linear pH gradient gel of 3–10 (IPGstrips, GE) loaded with $200\mu\text{g}$ of protein

resolubilized in 8M urea, 4% (w/v) CHAPS, 60mM DTT and 0.5% (v/v) IPG buffer 3–10NL (GE). The IEF was carried out at 30V for 12h, followed by 200V for 1h, 500V for 1.5h, 1000V for 1.5h, and 8000V for 6.5h, at 20°C. Prior to SDS-PAGE the IPGstrips were equilibrated for 2x15min in a buffer solution containing 0.05M Tris-HCl pH 8.8, 6M urea, 30% (v/v) glycerol and 2% (w/v) SDS. The DTT at 1% (w/v) was added to the first equilibration step and 4% (w/v) iodoacetamide to the second one. The SDS-PAGE was performed on slab gels (Laemmli 1970) and run at constant temperature of 15°C. The 2-DE gels were stained with colloidal Coomassie Blue (Neuhoff et al. 1985) and scanned using the ImageScanner (Amersham Biosciences).

Two-dimensional gel analysis

The gels were analysed in the ImageMaster 2D Platinum software v5.0 (GE) for spot detection, measurement and matching. In order to correct for variation in gel staining and for quantitative variations of protein spot intensities, the total spot volumes of each gel were normalized to 100, and % spot volumes calculated. The protein abundance analysis was performed according to Meunier et al. (2005), with minor changes. For each differentially expressed spot a Student's *t*-test ($p < 0.05$) analysis was performed with SigmaStat v3.10 (Systat Software, Inc.).

In-gel digestion

The protein spots were rinsed with a washing solution of 50% (v/v) acetonitrile and 0.1M ammonium bicarbonate for 4h. Prior to digestion the dried spots were reduced with 0.01M DTT for 2h at 37°C, then alkylated with 0.05M iodoacetamide for 15min at room temperature, in the dark. The gel spots were again rinsed with the washing solution for 2h. The digestion was made overnight, with 125ng of trypsin (Sequencing Grade, Promega) in 0.05M ammonium bicarbonate at 37°C. The peptides in the supernatant were collected and the gel pieces were extracted with a solution of 0.1% (v/v) acetic acid and 50% (v/v) acetonitrile. The extract was pooled with the tryptic peptides and dried in a speed vac. The pellet redissolved in 0.1% (v/v) acetic acid and 2% (v/v) acetonitrile solution was used for mass spectrometric analysis.

MS/MS analysis

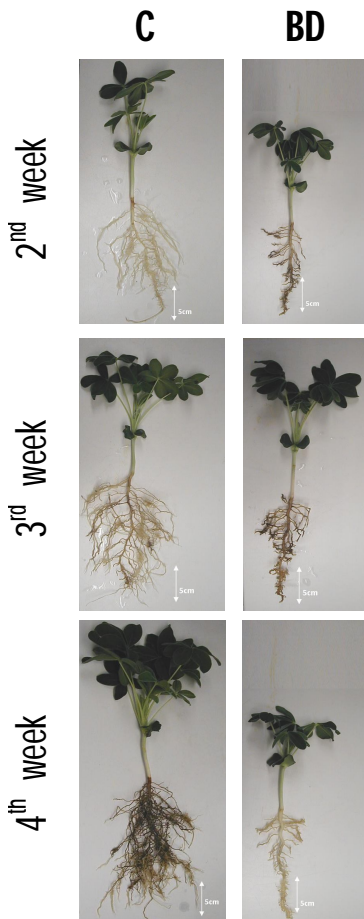
The trypsin digested proteins were analysed by capillary liquid chromatography tandem MS (LC/MS/MS) using a set up of a trapping 300SB C-18 column (0.3 x 50mm) (Agilent Technologies, Basel, Switzerland) and a separating column (0.1mm x 10cm) that had been packed with Magic 300Å C18 reverse-phase material (5µm particle size, Michrom Bioresources, Inc., Auburn, CA, USA). The columns were connected on line to an

Orbitrap FT hybrid instrument (Thermo Finnigan, San Jose, CA, USA). A linear gradient from 2 to 80% of solvent B [0.1% (v/v) acetic acid and 80% (v/v) acetonitrile] in solvent A [0.1% (v/v) acetic acid and 2% (v/v) acetonitrile] was delivered with a Rheos 2200 pump (Flux Instruments, Basel, Switzerland) for 85min at a flow rate of 100 μ L/min. A pre-column split was used to reduce the flow to approximately 100nL/min. The injection of 10 μ L of peptide digest was made by an auto-sampler thermostatted to 4°C and the eluting peptides ionized at 1.7kV. The mass spectrometer was operated in a data-dependent fashion. The precursor scan was done in the Orbitrap set to 60,000 resolution, while the fragment ions were mass analysed in the LTQ instrument. A top five method was run so that the five most intense precursors were selected for fragmentation. The MS/MS spectra were then searched against the NCBI non-redundant database, version August 15th 2008, using TurboSequest software (Gatlin et al. 2000). The databank was searched with Bioworks version 3.3.1. SP1 by setting the precursor ion tolerance to 10ppm, while the fragment ion tolerance was set to 0.5Da. Cleavage rules were set to Fully enzymatic – cleaves at both ends, allowing 2 missed cleavages. Post filtering was set to the following parameters: Δ CN, 0.1; Xcorr versus charge state was 1.50 (1+), 2.00 (2+), 2.50 (3+); peptide probability, 0.01; protein probability 0.01.

RESULTS AND DISCUSSION

Morphological effects of B deficiency

As dicots, *Lupinus albus* plants are quite sensitive to B suppression. Plants grown under B deficiency for two weeks already showed morphological differences, with shorter and darker roots lacking secondary and proteoid roots, despite that no significant difference in the biomass were observed. The morphological differences observed



in the roots indicate that the initial B present in the lupin seed is not enough for this early stage of development. Three weeks from sowing, the morphological differences in roots become more evident, accompanied by a significant difference in the biomass. In shoots, the

Figure 3.1. Morphological differences of *Lupinus albus* plants grown under B deficiency. The development of *L. albus* plants grown with (C) and without B (BD) in the nutrient solution was monitored two, three and four weeks after germination.

morphological differences due to B deficiency become visible, with darker and deformed leaves, but without significant biomass differences. After four weeks the morphological differences were much more marked, with significant reduced biomass in both shoots and roots (Figure 3.1, Figure 3.2).

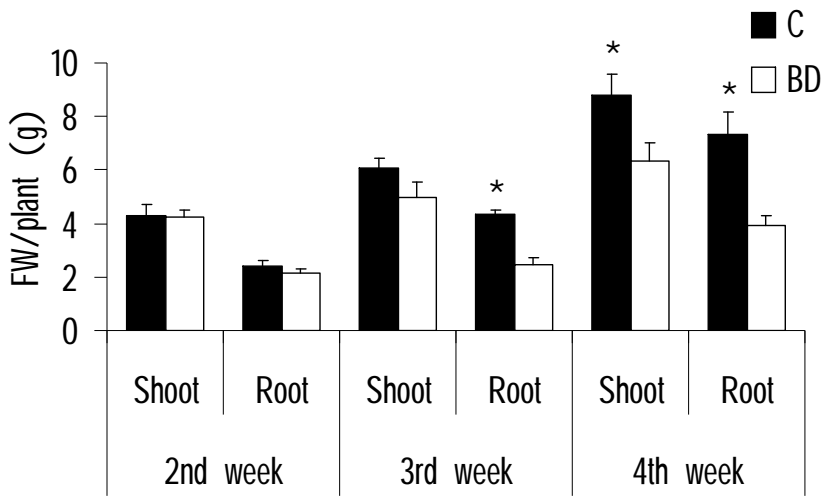


Figure 3.2. Fresh weight of *Lupinus albus* shoots and roots grown with (C) and without B (BD) for two, three and four weeks after seedling. Bars indicate standard errors. Significant changes were evaluated by the Student's *t*-test (*, $p < 0.05$).

Quantitative variations of the root proteome

Considering the importance of the root system and the ability of the lupin plant to maintain a reduced growth under B deficiency, the root proteome of plants treated with or without B were analysed by 2-DE (Figure 3.3). This fine resolution system, coupled to MS

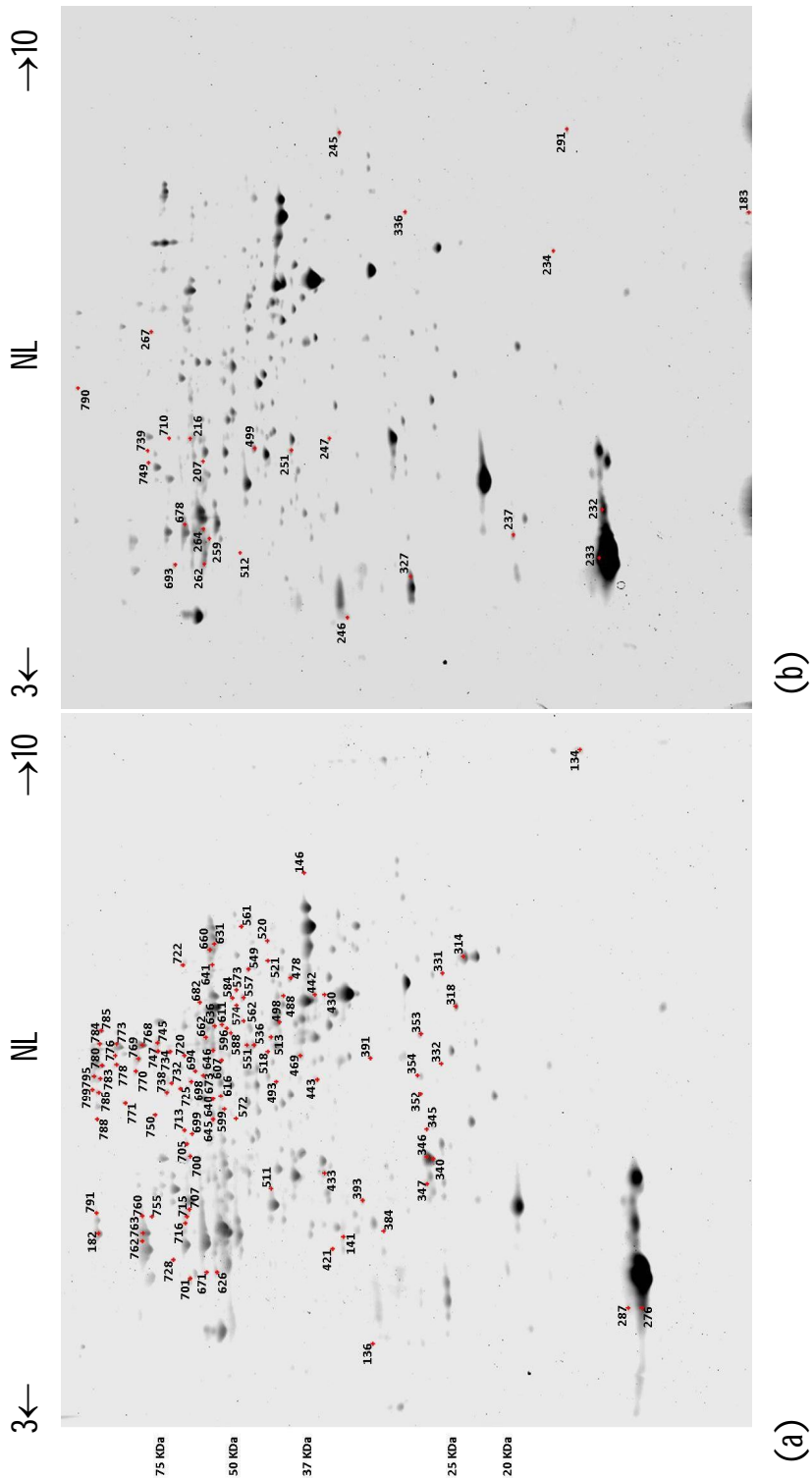


Figure 3.3. Representative 2-DE gels of *Lupinus albus* roots grown with (a) and without B (b). The gels were Coomassie Blue stained. Numbered proteins were identified by MS/MS (Table 3.1). Labelled proteins in gel a) are those suppressed due to B deficiency, while those labelled in gel b) are differentially expressed or de novo expressed due to B deficiency.

techniques and adequate statistical analysis, is a powerful tool to investigate the proteins that were differential expressed due to a long-term B deficiency.

Despite the large morphological changes observed due to B deficiency, the analysis of the root proteome revealed that 532 spots (67%) were non-responsive to B deficiency and that only 262 spots (33%) were responsive to B deficiency, either being suppressed (26%), expressed *de novo* (5%) or differentially expressed (2%) (Figure 3.4). The 262 spots reproducibly detected as

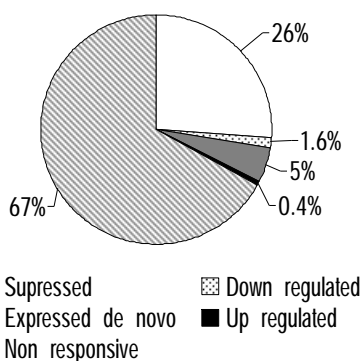


Figure 3.4. Percentage of polypeptides that was responsive to B deficiency in the *Lupinus albus* plant roots.

responsive to B deficiency were analysed by MS/MS techniques, and, 213 were identified. However, 83 spots gave ambiguous identification (i.e. spots that had homology with two or more non-homologous proteins) and, thus only the remainder 130 spots were considered for secure quantification.

Metabolic changes associated with B deficiency

In an attempt to identify the metabolic events associated with the plant root ability to cope with long-term B deficiency the identified proteins were grouped according to their biological functions (Figure 3.5).

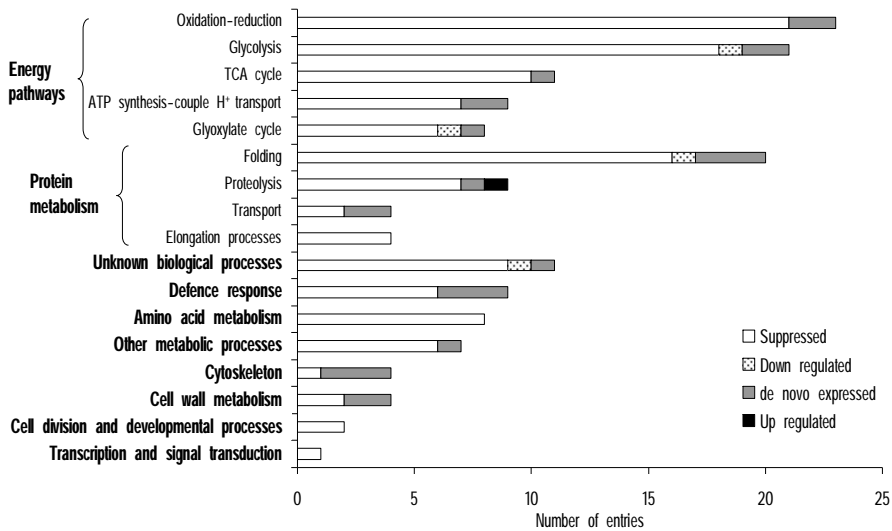


Figure 3.5. Functional classification of the proteins that had responded to B deficiency. The proteins of *Lupinus albus* plant roots identified were group according to the biological functions described in databases.

The majority of the suppressed proteins are from classes related with normal plant growth and developmental processes. This higher number of suppressed proteins is an indication of reduced biosynthetic flux that may be directly related with the reduced growth rates observed due to B deficiency. The fact that the up regulated or expressed *de novo* proteins belong to some of the same classes

of the suppressed or down regulated proteins (energy pathways, protein metabolism, defence and cytoskeleton biosynthesis) can be an indication that the plant metabolism was not just impaired as a whole, but instead, was modified in a controlled manner.

The most relevant biological processes where the identified proteins (Table 3.1) are involved will be discussed and related with B role in plant metabolism.

Energy metabolism

Changes in several proteins that are related to cellular energy metabolism were observed in B deficient roots. Key enzymes of respiration, such as glucose-6-phosphate-1-dehydrogenase (spot 700) from the pentose phosphate pathway, and the glyceraldehyde-3-phosphate dehydrogenase (spot 146) from the glycolysis pathways were suppressed. Also suppressed were several kinases, namely frutokinase (spot 421) and phosphoglycerate kinase (spots 493, 499 and 511), while the UDP-glucose pyrophosphorylase (spot 216) was expressed *de novo*. This enzyme is part of an alternative biochemical pathway for sucrose degradation that requires

Table 3.1. Protein identification of the spots selected as responsive to B deficiency. Proteins were grouped according to the most relevant biological process where they are involved. In the column of expression level, the de novo expressed proteins are marked as N and the suppressed as S. The differentially expressed proteins have their percentage of volume represented.


| ID ^a | Energy pathways | Protein identification ^b | Species | pI/MW (kDa) | | Expr. level |
|-----------------|-----------------|---|----------------------|------------------------|--------|--|
| | | | | Predicted ^c | Exp | |
| 146 | Energy pathways | Glyceraldehyde-3-phosphate dehydrogenase | Hordeum vulgare | --- | 8.2/38 | S |
| 216 | | UDP-glucose pyrophosphorylase | Amorpha fruticosa | 6.07/51.6 | 5.6/54 | N |
| 232 | | Aconitate hydratase | Cucurbita maxima | 5.74/98.0 | 5.1/15 | N |
| 245 | | Ascorbate peroxidase | Vigna unguiculata | 5.64/27.0 | 8.6/31 | N |
| 251 | | Pyruvate dehydrogenase E1 component subunit β | Arabidopsis thaliana | 5.11/35.9 | 5.5/36 | N |
| 264 | | V-type proton ATPase subunit B2 | Hordeum vulgare | 5.12/53.7 | 5.0/53 | N |
| 340 | | Ascorbate peroxidase | Vigna unguiculata | 5.64/27.0 | 6.3/26 | S |
| 346 | | Ascorbate peroxidase | Glycine max | 5.51/27.1 | 6.4/27 | S |
| 347 | | Ascorbate peroxidase | Retana raelam | 5.88/23.6 | 5.7/27 | S |
| 421 | | Fructokinase-1 | Oryza sativa | 5.07/34.7 | 5.1/35 | S |
| 442 | | Malate dehydrogenase | Lupinus albus | 6.10/35.6 | 6.4/37 | S |
| 443 | | Malate dehydrogenase | Lupinus albus | 6.10/35.6 | 5.9/36 | S |
| 469 | | Fructose-bisphosphate aldolase | Oryza sativa | 6.55/38.7 | 6.0/38 | S |
| 478 | | Formate dehydrogenase 1, mitochondrial | Oryza sativa | 6.20/39.3 | 6.6/40 | S |
| 488 | | Pyruvate dehydrogenase E1 component subunit α -1 | Arabidopsis thaliana | 6.42/39.6 | 6.4/41 | S |
| 493 | | Phosphoglycerate kinase | Arabidopsis thaliana | 5.04/42.6 | 5.9/41 | S |
| 498 | | Pyruvate dehydrogenase E1 component subunit α | Pisum sativum | --- | 6.2/42 | S |
| 499 | | Phosphoglycerate kinase | Arabidopsis thaliana | 5.49/42.1 | 6.1/41 |  |
| 511 | | Phosphoglycerate kinase | Arabidopsis thaliana | 5.04/42.6 | 5.5/43 | |
| 536 | | NADP-specific isocitrate dehydrogenase | Lupinus albus | 6.13/46.1 | 6.0/46 | |
| 551 | | NADP-specific isocitrate dehydrogenase | Lupinus albus | 5.99/46.0 | 6.0/47 | |
| | | | | | | |

Table 3.1 (cont) Protein identification of the spots selected as responsive to B deficiency. Proteins were grouped according to the most relevant biological process where they are involved. In the column of expression level, the de novo expressed proteins are marked as N and the suppressed as S. The differentially expressed proteins have their percentage of volume represented.

| ID ^a | Protein identification ^b | Species | pI/MW (kDa) | | Expr. level | |
|-----------------|-------------------------------------|---|------------------------|-----------|-------------|---|
| | | | Predicted ^c | Exp | | |
| 557 | BOFGG5 | Monodehydroascorbate reductase | Vaccinium corymbosum | 5.78/47.4 | 6.3/48 | S |
| 561 | P93033 | Fumarate hydratase 1 | Arabidopsis thaliana | 6.65/49.9 | 7.3/48 | S |
| 562 | BOFGG5 | Monodehydroascorbate reductase | Vaccinium corymbosum | 5.78/47.4 | 6.1/49 | S |
| 572 | A9SGH3 | Endase | Physcomitrella patens | 5.22/46.1 | 5.7/50 | S |
| 588 | O9FFR3 | 6-phosphogluconate dehydrogenase, decarboxylating | Arabidopsis thaliana | 5.62/53.3 | 6.1/50 | S |
| 599 | O9FFR3 | 6-phosphogluconate dehydrogenase, decarboxylating | Arabidopsis thaliana | 5.62/53.3 | 5.7/52 | S |
| 607 | O9LI00 | 6-phosphogluconate dehydrogenase, decarboxylating | Oryza sativa | 5.85/52.7 | 6.0/53 | S |
| 611 | O9LEJ0 | Endase 1 | Hevea brasiliensis | 5.57/47.8 | 6.1/73 | S |
| 626 | Q40079 | V-type proton ATPase subunit B2 | Hordeum vulgare | 5.12/53.7 | 4.9/54 | S |
| 631 | O9FNM5 | Subunit of complex I | Arabidopsis thaliana | 8.46/53.4 | 7.0/55 | S |
| 636 | P12862 | ATP synthase subunit alpha | Triticum aestivum | 5.70/55.3 | 6.1/55 | S |
| 640 | P12862 | ATP synthase subunit alpha | Triticum aestivum | 5.70/55.3 | 5.8/55 | S |
| 641 | P49357 | Serine hydroxymethyltransferase 1 | Flaveria pringlei | 8.15/53.6 | 6.8/55 | S |
| 646 | P12862 | ATP synthase subunit alpha | Triticum aestivum | 5.70/55.3 | 5.0/43 | S |
| 660 | O23254 | Serine hydroxymethyltransferase | Arabidopsis thaliana | 6.80/51.7 | 7.0/56 | S |
| 662 | B9SH74 | Aldehyde dehydrogenase, putative | Ricinus communis | 5.87/42.0 | 6.1/57 | S |
| 700 | Q42919 | Glucose-6-phosphate 1-dehydrogenase | Medicago sativa | 5.85/58.9 | 6.5/63 | S |
| 738 | O82663 | Succinate dehydrogenase [ubiquinone] flavoprotein subunit 1 | Arabidopsis thaliana | 5.58/66.0 | 5.8/71 | S |
| 739 | P31405 | V-type proton ATPase catalytic subunit A | Gossypium hirsutum | 5.36/68.5 | 5.5/77 | N |
| 771 | O9FGI6 | NADH-ubiquinone oxidoreductase 75 kDa subunit | Arabidopsis thaliana | 5.72/77.9 | 8.1/29 | S |
| 780 | P49608 | Aconitate hydratase | Cucurbita maxima | 5.74/98.0 | 5.9/135 | S |
| 783 | P49608 | Aconitate hydratase | Cucurbita maxima | 5.74/98.0 | 5.9/138 | S |
| 784 | P49608 | Aconitate hydratase | Cucurbita maxima | 5.74/98.0 | 6.0/138 | S |

Table 3.1 (cont) Protein identification of the spots selected as responsive to B deficiency. Proteins were grouped according to the most relevant biological process where they are involved. In the column of expression level, the de novo expressed proteins are marked as N and the suppressed as S. The differentially expressed proteins have their percentage of volume represented.

| ID ^a | Protein identification ^b | Species | pI/MW (kDa) | | Expr. Level |
|---------------------------|--|-----------------------|------------------------|---------|-------------|
| | | | Predicted ^c | Exp | |
| 785 | P49608 Aconitate hydratase | Cucurbita maxima | 5.74/98.0 | 6.1/136 | S |
| 786 | P49608 Aconitate hydratase | Cucurbita maxima | 5.74/98.0 | 6.0/138 | S |
| 788 | P49608 Aconitate hydratase | Cucurbita maxima | 5.74/98.0 | 5.7/143 | S |
| 790 | P49608 Aconitate hydratase | Cucurbita maxima | 5.74/98.0 | 5.8/140 | S |
| Protein metabolism | | | | | |
| 134 | O49886 Peptidyl-prolyl cis-trans isomerase | Lupinus luteus | 8.71/18.3 | 9.6/17 | S |
| 136 | O40682 Elongation factor 1-delta 2 | Oryza sativa | 4.40/24.5 | 4.2/32 | S |
| 247 | Q94JX9 Nascent polypeptide-associated complex subunit alpha-like protein 2 | Arabidopsis thaliana | 4.37/23.7 | 5.6/32 | N |
| 259 | Q9LS40 CND41, chloroplast nucleoid DNA binding protein-like | Arabidopsis thaliana | 5.27/53.2 | 5.0/48 | N |
| 291 | O49886 Peptidyl-prolyl cis-trans isomerase | Lupinus luteus | 8.71/18.3 | 8.7/17 | N |
| 332 | Q3HVM0 Proteasome subunit alpha type | Solanum tuberosum | 5.40/28.1 | 5.9/26 | S |
| 345 | A9TVH1 Proteasome subunit alpha type | Physcomitrella patens | 5.92/27.2 | 5.7/27 | S |
| 352 | A5AXI5 Proteasome subunit alpha type | Vitis vinifera | 6.11/27.2 | 5.8/27 | S |
| 354 | A5ALB2 Proteasome subunit alpha type | Vitis vinifera | 5.91/27.3 | 5.9/27 | S |
| 512 | A9PEP6 Predicted protein | Populus trichocarpa | 4.94/45.3 | 5.0/43 | S |
| 513 | O9ZRUE Elongation factor Tu | Catharanthus roseus | -- | 6.0/43 | S |
| 573 | A0FH76 EBPI | Solanum tuberosum | 6.26/42.8 | 6.4/50 | S |
| 584 | O0DDX2 26S protease regulatory subunit 7 | Oryza sativa | 6.03/47.7 | 6.3/50 | S |
| 645 | Q6K669 Leucine aminopeptidase 2 | Oryza sativa | 5.50/55.0 | 5.7/55 | S |



Table 3.1 (cont) Protein identification of the spots selected as responsive to B deficiency. Proteins were grouped according to the most relevant biological process where they are involved. In the column of expression level, the de novo expressed proteins are marked as N and the suppressed as S. The differentially expressed proteins have their percentage of volume represented.


| ID ^a | Protein identification ^b | | Species | pI/MW (kDa) | | Expr. Level |
|-----------------|-------------------------------------|--|----------------------|------------------------|---------|---|
| | | | | Predicted ^c | Exp | |
| 693 | P21239 | RuBisCO large subunit-binding protein subunit α | Brassica napus | 4.78/57.0 | 4.9/61 |  |
| 694 | P21239 | RuBisCO large subunit-binding protein subunit α | Brassica napus | 4.78/57.0 | 5.9/61 | S |
| 699 | O940P8 | Putative uncharacterized protein | Arabidopsis thaliana | 5.59/57.3 | 5.6/63 | S |
| 701 | P21239 | RuBisCO large subunit-binding protein subunit α | Brassica napus | 4.78/57.0 | 4.9/63 | S |
| 705 | O93ZM7 | Chaperonin CPN60-like 2 | Arabidopsis thaliana | 5.32/57.1 | 6.0/64 | S |
| 707 | O05045 | Chaperonin CPN60-1 | Cucurbita maxima | 5.09/57.4 | 5.3/63 | S |
| 710 | P21240 | RuBisCO large subunit-binding protein subunit β | Arabidopsis thaliana | 5.26/58.1 | 5.6/64 | N |
| 713 | P21240 | RuBisCO large subunit-binding protein subunit β | Arabidopsis thaliana | 5.26/58.1 | 5.6/65 | S |
| 715 | P21240 | RuBisCO large subunit-binding protein subunit β | Arabidopsis thaliana | 5.26/58.1 | 5.3/65 | S |
| 716 | P21240 | RuBisCO large subunit-binding protein subunit β | Arabidopsis thaliana | 5.26/58.1 | 5.3/65 | S |
| 720 | A5BFM5 | Putative uncharacterized protein | Vitis vinifera | 6.03/61.2 | 6.0/65 | S |
| 722 | A5BFM5 | Putative uncharacterized protein | Vitis vinifera | 6.03/61.2 | 6.7/66 | S |
| 725 | A5BFM5 | Putative uncharacterized protein | Vitis vinifera | 6.03/61.2 | 5.8/67 | S |
| 728 | A5BFM5 | Putative uncharacterized protein | Vitis vinifera | 6.03/61.2 | 5.0/69 | S |
| 732 | O9M888 | Putative uncharacterized protein | Arabidopsis thaliana | 5.83/58.9 | 5.9/70 | S |
| 749 | P37900 | Heat shock 70 kDa protein | Pisum sativum | 5.18/66.7 | 5.8/66 | N |
| 750 | Q43468 | Heat shock protein ST1 | Glycine max | 5.81/63.6 | 5.7/77 | S |
| 755 | P37900 | Heat shock 70 kDa protein | Pisum sativum | 5.18/66.7 | 5.3/80 | S |
| 760 | P11143 | Heat shock 70 kDa protein | Zea mays | 5.22/70.6 | 5.3/89 | S |
| 762 | O39043 | Luminal-binding protein 2 | Arabidopsis thaliana | 5.08/71.1 | 5.2/87 | S |
| 763 | O39043 | Luminal-binding protein 2 | Arabidopsis thaliana | 5.08/71.1 | 5.2/88 | S |
| 791 | O9LZF6 | Cell division control protein 48 homolog E | Arabidopsis thaliana | 5.08/90.0 | 5.3/142 | S |

Table 3.1 (cont) Protein identification of the spots selected as responsive to B deficiency. Proteins were grouped according to the most relevant biological process where they are involved. In the column of expression level, the de novo expressed proteins are marked as N and the suppressed as S. The differentially expressed proteins have their percentage of volume represented.

| ID ^a | Protein identification ^b | Species | pI/MW (kDa) | | Expr. Level |
|-------------------------------|---|-------------------------------|------------------------|---------|-------------|
| | | | Predicted ^c | Exp | |
| 795 | O23755 Elongation factor 2 | Beta vulgaris | 5.93/93.8 | 5.9/146 | S |
| 799 | O23755 Elongation factor 2 | Beta vulgaris | 5.93/93.8 | 5.8/150 | S |
| Defence response | | | | | |
| 233 | P52779 Protein LIR18B | Lupinus luteus | 5.35/16.6 | 4.7/15 | N |
| 234 | Q93X10 Pathogenesis-related 10 | Lupinus albus | 4.87/16.9 | 6.8/18 | N |
| 276 | Q93X10 Pathogenesis-related 10 | Lupinus albus | 4.87/16.9 | 4.6/15 | S |
| 287 | Q93X10 Pathogenesis-related 10 | Lupinus albus | 4.87/16.9 | 4.6/15 | S |
| 318 | O0PN10 Glutathione S-transferase | Caragana korshinskii | 6.86/25.8 | 6.3/25 | S |
| 327 | Q9SXM5 Acidic chitinase | Glycine max | 5.01/31.9 | 4.7/26 | N |
| 430 | P23535 Glucan endo-1,3- β -glucosidase, basic isoform | Phaseolus vulgaris | 8.75/35.2 | 6.4/36 | S |
| Transcription | | | | | |
| 314 | O5E163 Quinone reductase 2 | Triticum monococcum | 5.95/21.7 | 6.7/24 | S |
| Amino acids metabolism | | | | | |
| 393 | A3RM06 Cysteine synthase | Glycine max | 5.29/34.7 | 5.4/32 | S |
| 520 | P54260 Aminomethyltransferase | Solanum tuberosum | 7.28/40.9 | 7.1/44 | S |
| 521 | Q40108 Aspartate aminotransferase | Lupinus angustifolius | 8.36/45.8 | 6.8/44 | S |
| 616 | Q9SP37 Adenosylhomocysteinase | Lupinus luteus | 5.64/53.3 | 5.8/53 | S |
| 773 | P93263 5-methyltetrahydropteroyltriglutamate-homocysteine methyltransferase | Mesembryanthemum crystallinum | 5.90/84.8 | 6.0/114 | S |
| 776 | P93263 5-methyltetrahydropteroyltriglutamate-homocysteine methyltransferase | Mesembryanthemum crystallinum | 5.90/84.8 | 6.0/119 | S |
| 778 | P93263 5-methyltetrahydropteroyltriglutamate-homocysteine methyltransferase | Mesembryanthemum crystallinum | 5.90/84.8 | 5.9/116 | S |

Table 3.1 (cont) Protein identification of the spots selected as responsive to B deficiency. Proteins were grouped according to the most relevant biological process where they are involved. In the column of expression level, the de novo expressed proteins are marked as N and the suppressed as S. The differentially expressed proteins have their percentage of volume represented.

| ID ^a | Protein identification ^b | | Species | pI/MW (kDa) | | Expr. Level |
|---------------------------|-------------------------------------|--|----------------------|------------------------|---------|-------------|
| | | | | Predicted ^c | Exp | |
| Cell wall metabolism | | | | | | |
| 267 | P34105 | NADP-dependent malic enzyme | Populus trichocarpa | 6.50/65.2 | 6.4/61 | N |
| 518 | Q93VR3 | GDP-mannose 3,5-epimerase | Arabidopsis thaliana | 5.85/42.8 | 6.0/44 | S |
| 673 | Q96558 | UDP-glucose 6-dehydrogenase | Glycine max | 5.74/52.9 | 5.9/58 | S |
| 734 | A9PGL9 | Malic enzyme | Populus trichocarpa | 7.61/54.6 | 6.0/70 | S |
| 745 | P34105 | NADP-dependent malic enzyme | Populus trichocarpa | 6.50/65.2 | 6.0/75 | S |
| 747 | P34105 | NADP-dependent malic enzyme | Populus trichocarpa | 6.50/65.2 | 6.0/75 | S |
| Cytoskeleton biosynthesis | | | | | | |
| 182 | O1G0Z1 | Putative spindle disassembly related protein CDC48 | Nicotiniana tabacum | 5.13/89.9 | 5.2/149 | S |
| 183 | O1G0Z1 | Putative spindle disassembly related protein CDC48 | Nicotiniana tabacum | 5.13/89.9 | 7.4/10 | N |
| 207 | P20363 | α -3/ α -5 tubulin chain | Arabidopsis thaliana | 4.95/49.7 | 5.4/49 | N |
| 262 | O9STD0 | β -tubulin | Zinnia elegans | 4.75/50.1 | 4.9/49 | N |
| 331 | P41916 | GTP-binding nuclear protein Ran-1 | Arabidopsis thaliana | 6.39/25.3 | 6.7/26 | S |
| 336 | P41916 | GTP-binding nuclear protein Ran-1 | Arabidopsis thaliana | 6.39/25.3 | 7.4/26 | N |
| Other metabolic processes | | | | | | |
| 391 | O8LCJ6 | Ethylene-responsive protein 2-like | Oryza sativa | 10.86/12.6 | 6.0/31 | S |
| 574 | O7M1Z8 | Globulin-2 | Zea mays | 6.16/49.9 | 6.3/50 | S |
| 596 | P19595 | UTP-glucose-1-phosphate uridylyltransferase | Solanum tuberosum | 5.71/51.7 | 6.1/52 | S |
| 682 | P15590 | Globulin-1 S allele | Zea mays | 6.75/55.1 | 6.3/60 | S |
| 768 | O7SIC9 | Transketolase | Zea mays | 5.47/73.0 | 6.0/88 | S |
| 769 | O7SIC9 | Transketolase | Zea mays | 5.47/73.0 | 6.0/32 | S |
| 770 | O7SIC9 | Transketolase | Zea mays | 5.47/73.0 | 5.9/95 | S |

Table 3.1 (cont) Protein identification of the spots selected as responsive to B deficiency. Proteins were grouped according to the most relevant biological process where they are involved. In the column of expression level, the de novo expressed proteins are marked as N and the suppressed as S. The differentially expressed proteins have their percentage of volume represented.

| ID ^a | Protein identification ^b | | Species | pI/MW (kDa) | | Expr. Level |
|------------------------------|-------------------------------------|--|----------------------|------------------------|--------|-------------|
| | | | | Predicted ^c | Exp | |
| Unknown biological processes | | | | | | |
| 141 | Q8LPE5 | Fructokinase-like protein | Cicer arietinum | --- | 5.1/35 | S |
| 237 | Q9M328 | Putative uncharacterized protein T18D12.90 | Arabidopsis thaliana | 5.66/17.8 | 5.1/20 | N |
| 246 | Q8GY98 | Putative germin | Arabidopsis thaliana | 8.39/23.5 | 4.4/30 | N |
| 353 | Q2V987 | Transcription factor APFI-like | Solanum tuberosum | 7.05/29.1 | 5.8/27 | S |
| 384 | Q9SMK5 | Plasma membrane intrinsic polypeptide | Cicer arietinum | 4.95/23.3 | 5.2/30 | S |
| 549 | A5CB20 | Putative uncharacterized protein | Vitis vinifera | 8.42/54.2 | 6.7/47 | S |
| 671 | Q7XCL2 | Ubiquitin domain containing protein | Oryza sativa | 4.71/59.3 | 4.9/58 | S |
| 678 | Q94IC1 | Betaine aldehyde dehydrogenase | Hordeum vulgare | 5.47/54.5 | 5.2/58 | |

| ID | % spot volume |
|-----|---------------|
| 141 | 1.5 |
| 237 | 2.5 |
| 246 | 3.5 |
| 353 | 4.5 |
| 384 | 5.5 |
| 549 | 6.5 |
| 671 | 7.5 |
| 678 | 8.5 |



^a Spot numbers are corresponding to the numbers in Figure 3.3.

^b Protein identification according to the UniProt database (<http://www.uniprot.org>)

^c Predicted pI and MW (kDa) were calculated by using an ExPASy tool (<http://www.expasy.org>)

inorganic pyrophosphate, whereas the breakdown involving kinases requires two molecules of ATP (Stitt 1998). The activation of bypass pathways allows the carbon flow to continue under stressful conditions by using alternative energy sources and thereby reducing ATP demand. Indeed, several ATPases were affected by B deficiency as it will be discussed below.

For the V-type proton ATPase some subunits were expressed *de novo* (subunits A, spot 739 and B2, spot 264) and others suppressed (a subunit B2 isoform, spot 626) as well as some ATPase synthases subunit isoforms (spots 636, 640 and 646). The reduced activity of proton-pumping ATPase, as previously observed in sunflower cells, was related with possible membrane damage caused by B deficiency (Ferrol and Donaine 1992). The expression *de novo* of pyruvate dehydrogenase E1 component subunit beta (spot 251) is also relevant since E1 protein levels correlate with mitochondrial pyruvate dehydrogenase complex activity (Luethy et al. 2001), and higher expression of this enzyme correlates with metabolic and structural changes that accompany membrane expansion and remodelling (Tovar-Méndez et al. 2003).

Therefore, these results indicate an effect of B deficiency on membranes, and it should be referred, that a physiological role for B in membranes has been proposed by a wealth of information in which B deficiency has been shown to disrupt membrane-associated

processes, including membrane potential and electron transport (Brown et al. 2002).

Several aconitate hydratase isoforms were suppressed (spots 780, 783, 784, 785, 786 and 788) or down regulated (spot 790) due to B deficiency and another form (spot 232) was expressed *de novo*. Since this protein spot has a considerable lower molecular mass than that annotated in the database, it may have been targeted for degradation during B deficiency. Considering that the aconitase cluster is lost under oxidative stress (Moeder et al. 2007) and that oxidative damage is the major cause of cell death induced by B-deprivation in tobacco cells (Koshihara et al. 2009), the aconitase suppression or degradation probably results from the oxidative damage caused by B deficiency.

Protein metabolism

The Rubisco large subunit-binding protein subunit β (spot 710) belongs to the heat shock protein 60 (Hsp60) family. Both this Hsp60 and a Hsp70 (spot 749) were expressed *de novo* under B deficiency and are molecular chaperones responsible for preventing irreversible aggregation of non-native proteins under both normal and stressful conditions (Timperio et al. 2008). Another protein involved in the protein folding processes, that was also expressed *de novo* under B deficiency, is a peptidylprolyl *cis-trans* isomerase (spot

291), that can additionally play important roles in protein degradation, signal transduction and mRNA processing (Nuc et al. 2001).

The nascent polypeptide-associated complex (NAC) is a heterodimeric complex of α - and β - chains that is postulated to be involved in protein transport for an appropriate targeting of ribosome-nascent polypeptide complexes (Rospert et al. 2002). The expression *de novo* of a NAC subunit α -like protein 2 (spot 247), might suggest altered protein translation and targeting, important for adaptive stress responses.

Both the CND41 (spot 259) that was expressed *de novo*, and the predicted protein (A9PEP6; spot 512) that was up regulated under B deficiency have aspartic-type endopeptidase activity. This activity is apparently related with proteolytic processes implicated in post-mortem proteolysis of the 7S globular storage protein and the degradation of extracellular pathogenesis-related (PR) proteins (Voigt et al. 1997). Indeed, a 7S seed storage protein, the globulin-1S allele (spot 682) and two pathogenesis-related (PR)-10 proteins (spots 276 and 287) were suppressed due to B deficiency.

The protein degradation is an ATP-demanding process, so its repression will decrease protein synthesis and turnover, thereby reducing ATP demand (Hochachka and Lutz 2001). The fact that

suppressed and *de novo* expressed proteins belong to the same metabolic class, points out for a shift in protein folding and proteolysis processes, that may lead to a redirected protein metabolism towards plant survival under stressful conditions.

Amino acid metabolism

Some proteins related with amino acid metabolism were suppressed by B deficiency. The majority of these proteins (Adenosylhomocysteinase, spot 616; Cysteine synthase, spot 393 and 5-methyltetrahydropteroyltriglutamate-homocysteine methyltransferase, spots 773, 776 and 778) are related with sulphur containing amino acids. Studies on the transcriptional profile of *Arabidopsis thaliana* plants grown under B deficiency suggested altered sulphur metabolism (Chapter 5), but the relationship between B role in plants and sulphur metabolism remains unclear.

Cell wall metabolism

Some proteins associated with the cell wall metabolism were suppressed due to B deficiency. Both the UDP-glucose 6-dehydrogenase (spot 673) and the GDP-mannose 3,5-epimerase (spot 518) are involved in the carbohydrate metabolism directed for the cell wall biosynthesis. Several malic enzyme isoforms (spots 734, 745 and 747), that were suppressed under B deficiency, are

described to provide the NADPH used for the production of H_2O_2 in lignin biosynthesis (Martinoia and Rentsch 1994). Lignin production that results from the hydroxycinnamyl alcohols polymerization by peroxidases (Higuchi 1990) could be compromised by B deficiency, as observed by the lower degree of lignification of some trees growing in a low B content soil (Dell and Malajczuk 1994). The suppression of several ascorbate peroxidases isoforms (spots 340, 346 and 347), and the previously observation of ascorbate peroxidase inhibition in B deficient squash roots (Cakmak and Römheld 1997) could be related with the phenol increased content described for B-deficient plants (Blevins and Lukaszewski 1998).

In a transcriptional analysis of B deficient *Arabidopsis thaliana* roots, it was found that several genes related with cell wall metabolism were down regulated (Camacho-Cristóbal et al. 2008). The known participation of B in cell wall structure may explain the impaired cell wall metabolism observed in *L. albus*, however this is not enough to explain all the symptoms observed due to B deficiency.

Defence responses

Several proteins related with defence responses were expressed *de novo* due to B deficiency, namely PR-10 (spot 234) and LIR18B protein (spot 233), both belongs to the PR-10 family, and an acidic chitinases (spot 327) that belongs to the PR-8 family

(Watanabe et al. 1999). The PR family proteins are known to be induced by several biotic and abiotic stresses (Van Loon and Van Strien 1999). An uncharacterized T18D12.90 protein (spot 237) and a putative germin-like protein (spot 246), that were expressed *de novo* under B deficiency, have unknown biological functions however, they have been also associated with various stress responses. For example, in *A. thaliana* the protein T18D12.90 is described to be part of the universal stress protein (USP) family (Kvint et al. 2003). Germin and germin-like proteins, besides their involvement in stress responses, have been described to participate in a wide range of activities related to developmental processes and cell wall biosyntheses (Bernie and Berna 2001). Several other studies had already reported increased defence proteins in association with B deficiency (Alves et al. 2006, Kobayashi et al. 2004, Reguera et al. 2010). Updated evidence shows that PR proteins and other stress responsive proteins may display additional functions in growth and developmental processes by modulating signal molecules (Kasprzewska 2003, Liu and Ekramoddoullah 2006), however the association of these defence proteins with B is not yet understood.

Cytoskeleton biosynthesis

Several proteins related with cytoskeleton biosynthesis were affected by B deficiency. Under B deficiency we detected *de novo* expression of tubulins (spots 207 and 262), which are major components of microtubules, and a putative actin from *Dictyostelium discoideum* (Q553U6). Another expressed *de novo* protein, which could be involved in the regulation of cytoskeletal assembly and organization, was a GTP-binding nuclear protein Ran-1 (spot 336) (Haizel et al. 1997, Vernoud et al. 2003). The *de novo* expression of a putative spindle disassembly-related protein CDC48 (spot 183) is probably the result of protein degradation processes since this protein spot has a considerable lower molecular mass than the annotated in the database. Beside this last protein, we have detected several others, which were suppressed due to B deficiency, related with cytoskeleton biosynthetic process. The putative uncharacterized proteins from *A. thaliana* (spots 699 and 732) and from *V. vinifera* (spots 725, 720, 722 and 728) belong to the TCP-1 chaperonin family that, in association with the Hsp70 molecular chaperones, can interact with cytoskeletal components (Gatenby and Viitanen 1994). Elongation factors (spots 136 and 513), besides their participation in the translational apparatus, appear to have a second role as a regulator of cytoskeleton rearrangements (Gromadski et al. 2002).

Previous studies on altered cytoskeleton features were reported as increased levels of tubulins, actins and altered polymerization patterns of these cytoskeleton proteins in higher plants subjected to B deficiency (Yu et al. 2001, 2003). Since cytoskeleton is involved in diverse important cellular aspects, such as mitotic spindle formation, intracellular transport and control of cell shape (Gunning and Hardham 1982), a modified cytoskeleton biosynthesis can be a possible explanation for the missing part of B role in higher plants. Indeed, a possible role for B in cytoskeleton is supported by previous findings that B deficiency primarily disrupts processes where active cytoskeleton remodelling is required, such as the initial phases of differentiation, including pollen tube growth, anther development (Loomis and Durst 1992, Rawson 1996), somatic embryo formation (Behrendt and Zoglauer 1996) and early nodulation processes (Bonilla et al. 1994).

Protein isoforms in stress responses

One striking feature of this proteomic study is the detection of several changes in protein isoforms as a consequence of B deficiency. A numerous group is that of the proteins that showed slight changes in pI and/or MW in response to the deficiency. Different protein isoforms were already found to have a crucial role in fungal infection and symbiosis studies in *Medicago truncatula*

(Salzer et al. 2000). This may indicate that isoforms could have an active participation in regulatory processes, determinant for plants to cope with stressful conditions.

CONCLUSIONS

There was a clear metabolic adjustment of the biosynthetic fluxes of the lupin root in response to B deficiency. The adaptive responses to the deficiency resulted in a reduction of important metabolic processes, namely in energy and protein metabolic processes. Other common adaptive stress responses are related with defence proteins. Several other metabolic processes were affected by the deficiency, such as cell wall metabolism, as expected since B is known to participate in plant cell wall structure, and cytoskeleton biosynthesis. The high requirement of B in active cytoskeleton remodelling, such as in initial phases of differentiation (Behrendt and Zoglauer 1996), in reproductive processes (Loomis and Durst 1992, Rawson 1996) and in nodulation processes, is consistent with a possible role for B in cytoskeleton biosynthetic processes.

ACKNOWLEDGMENTS

We gratefully acknowledge financial support from the Fundação de Ciência e Tecnologia (grant nº. FCT SFRH/BD/18273/2004)

REFERENCES

- Alves M, Francisco R, Martins I, Ricardo C (2006) Proteomic analysis of the extracellular domain of *Lupinus albus* leaves in response to boron deficiency. *Plant Soil* 279: 1–11.
- Arnon D (1938) Microelements in culture–solution experiment with higher plants. *Amer J Bot* 25: 322–325.
- Bassil E, Hu H, Brown P (2004) Use of Phenylboronic Acids to Investigate Boron Function in Plants. Possible Role of Boron in Transvacuolar Cytoplasmic Strands and Cell-to-Wall Adhesion. *Plant Physiol* 136: 3383–3395.
- Behrendt U, Zoglauer K (1996) Boron controls suspensor development in embryogenic cultures of *Larix decidua*. *Physiol Plant* 97: 321–326.
- Bernier F, Berna A (2001) Germins and germin-like proteins: Plant do-all proteins. But what do they do exactly? *Plant Physiol Biochem* 39: 545–554.
- Blevins D, Lukaszewski K (1998) Boron in plant structure and function. *Annu Rev Plant Physiol Plant Mol Biol* 49: 481–500.
- Bollaños L, Esteban E de Lorenzo C, Fernández-Pascual M, de Felipe M, Gárate A, Bonilla I (1994) Essentiality of boron

for symbiotic dinitrogen fixation in pea (*Pisum sativum*)-*Rhizobium* nodules. Plant Physiol 104: 85-90.

- Bonilla I, Blevins D, Bolaños L (2009) Boron Functions in Plants: Looking Beyond the Cell Wall. In: Plant Physiology. Eds: Taiz L, Zeiger E. 4th Ed. IOP Publishing physics web. <http://4e.plantphys.net>. Accessed Dec 2009.
- Brown P, Bellaloui N, Wimmer M, Bassil E, Ruiz J, Hu H, Pfeiffer H, Dannel F, Romheld V (2002) Boron in plant biology. Plant Biol 4: 203-223.
- Cakmak I, Römheld V (1997) Boron deficiency-induced impairments of cellular functions in plants. Plant Soil 193: 71-83.
- Camacho-Cristóbal J, Herrera-Rodríguez M, Beato V, Rexach J, Navarro-Gochicoa M, Maldonado J, González-Fontes A (2008) The expression of several cell wall-related genes in Arabidopsis roots is down-regulated under boron deficiency. Env Exp Bot 63: 351-358.
- Dell B, Malajczuk N (1994) Boron deficiency in eucalypt plantations in China. Can J For Res 24: 2409-2416.
- Ferrol N, Donaire J (1992) Effect of boron on plasma membrane proton extrusion and redox activity in sunflower cells. Plant Sci 86: 41-47.
- Fitter A (2002) Characteristics and functions of root systems, pp 3-25. In: Plant Roots: the hidden half, Eds: Waisel Y, Eshel A, Kafkafi U. CRC Press, NY.
- Gatenby A, Viitanen P (1994) Structural and functional aspects of chaperonin mediated protein folding. Annu Rev Plant Physiol Plant Mol Biol 45: 469-491.

- Gatlin C, Eng J, Cross S, Detter J, Yates J (2000) 3rd: Automated identification of amino acid sequence variations in proteins by HPLC/microspray tandem mass spectrometry. *Anal Chem* 72: 757-763.
- Gromadski K, Wieden H, Rodnina M (2002) Kinetic Mechanism of Elongation Factor Ts-Catalyzed Nucleotide Exchange in Elongation Factor Tu. *Biochem* 41: 162-169.
- Gunning B, Hardham A (1982) Microtubules. *Annu Rev Plant Physiol* 33: 651-698.
- Haizel T, Merkle T, Pay A, Fejes E, Nagy F (1997) Characterization of proteins that interact with the GTP-bound form of the regulatory GTPase Ran in Arabidopsis. *Plant J* 11: 93-103.
- Higuchi T (1990) Lignin biochemistry: biosynthesis and biodegradation. *Wood Sci Technol* 24: 23-63.
- Hochachka P, Lutz P (2001) Mechanism, origin and evolution of anoxia tolerance in animals. *Comp Biochem Physiol* 130: 435-459.
- Kasprzewska A (2003) Plant chitinases – regulation and function. *Cell Mol Biol Lett* 8: 809-824.
- Kobayashi M, Mutoh T, Matoh T (2004) Boron nutrition of cultured tobacco BY-2 cells. IV. Genes induced under low boron supply. *J Exp Bot* 55: 1441-1443.
- Kobayashi M, Nakagawa H, Asaka T, Matoh T (1999) Borate rhamnogalacturonan II bonding reinforced by Ca²⁺ retains pectic polysaccharides in higher plant cell walls. *Plant Physiol* 119: 199-204.

- O'Neill M, Eberhard S, Albersheim P, Darvill A (2001) Requirement of borate cross-linking of cell wall rhamnogalacturonan II for *Arabidopsis* growth. *Science* 294: 846-849.
- O'Neill M, Warrenfeltz D, Kates K, Pellerin P, Doco T, Darvill A, Albersheim P (1996) Rhamnogalacturonan-II, a pectic polysaccharide in the walls of growing plant cell, forms a dimer that is covalently cross linked by a borate ester. *J Biol Chem* 271: 22923-22930.
- Kobayashi M, Match T, Azuma J (1996) Two chains of rhamnogalacturonan II are cross-linked by borate-diol ester bonds in higher plant cell walls. *Plant Physiol* 110: 1017-1020.
- Koshiba T, Kobayashi M, Match T (2009) Boron nutrition of tobacco BY-2 cells. V. Oxidative damage is the major cause of cell death induced by boron deprivation. *Plant Cell Physiol* 50: 26-36.
- Kvint K, Nachin L, Diez A, Nyström T (2003) The bacterial universal stress protein: function and regulation. *Curr Opin Microbiol* 6: 140-145.
- Laemmli U (1970) Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* 227: 680-685.
- Liu J-J, Ekramoddoullah A (2006) The family 10 of plant pathogenesis-related proteins: Their structure, regulation, and function in response to biotic and abiotic stresses. *Physiol Mol Plant Pathol* 68: 3-13.
- Loomis W, Durst R (1992) Chemistry and biology of boron. *BioFactors* 3: 229-239.

- Luethy M, Gemel J, Johnston M, Mooney B, Miernyk J, Randall D (2001) Developmental expression of the mitochondrial pyruvate dehydrogenase complex in pea (*Pisum sativum*) seedlings. *Physiol Plant* 112: 559–566.
- Martinoia E, Rentsch D (1994) Malate compartmentation-responses to a complex metabolism. *Annu Rev Plant Physiol Plant Mol Biol* 45:447–467.
- Meunier B, Bouley J, Piec I, Bernard C, Picard B, Hocquette J-F (2005) Data analysis methods for detection of differential protein expression in two-dimensional gel electrophoresis. *Anal Biochem* 340: 226–230.
- Moeder W, Pozo O, Navarre D, Martin G, Klessig D (2007) Aconitase plays a role in regulating resistance to oxidative stress and cell death in *Arabidopsis* and *Nicotiana benthamiana*. *Plant Mol Biol* 63: 273–287.
- Neuhoff V, Stamm R, Eibl H (1985) Clear background and highly sensitive protein staining with Coomassie Blue dyes in polyacrylamide gels: a systematic analysis. *Electrophoresis* 6: 427–448.
- Nuc K, Nuc P, Slomski R (2001) Yellow lupine cyclophilin transcripts are highly accumulated in the nodule meristem zone. *MPMI* 14: 1384–1394.
- Ramagli L (1999) Quantifying protein in 2D PAGE solubilization buffers, pp 95–105. In: *Methods in Molecular Biology - 2D Proteome Analysis Protocols*. Eds. Andrew J. Link Humana Press, Totowa, New Jersey.
- Rawson H (1996) The developmental stage during which boron limitation causes sterility in wheat genotypes and the recovery of fertility. *Aust J Plant Physiol* 23: 709–717.

- Reguera M, Bonilla I, Bolaños L (2010) Boron deficiency results in induction of pathogenesis-related proteins from the PR-10 family during the legume-rhizobia interaction. *J Plant Physiol* (*in press*).
- Rospert S, Dubaquié Y, Gautschi M (2002) Nascent-polypeptide associated complex. *Cell Mol Life Sci* 59: 1632-1639.
- Salzer P, Bonanami A, Beyer K, Vögeli-Lange R, Aeschbacher R, Lange J, Wiemken A, Kim D, Cook D and Boller T (2000) Differential Expression of eight chitinase genes in *Medicago truncatula* roots during mycorrhiza formation, nodulation and pathogen infection. *MPMI* 13: 763-777.
- Stitt M (1998) Pyrophosphate as an alternative energy donor in the cytosol of plant cells: an enigmatic alternative to ATP. *Bot Acta* 111: 167-175.
- Timperio A, Giulia M, Zolla L (2008) Proteomics applied on plant abiotic stress: role of heat shock proteins (HSP). *J Prot* 71: 391-411.
- Tovar-Méndez A, J Miernyk, Randall D (2003) Regulation of pyruvate dehydrogenase complex activity in plant cells. *FEBS* 270: 1043-1049.
- Van Loon L, Van Strien E (1999) The families of pathogenesis-related proteins, their activities, and comparative analysis of PR-1 type proteins. *Physiol Mol Plant Pathol* 55: 85-97.
- Vernoud V, Horton A, Yang Z, Nielsen E (2003) Analysis of the small GTPase gene superfamily of Arabidopsis. *Plant Physiol* 131: 1191-1208.
- Voigt G, Biehl B, Heinrichs H, Voigt J (1997) Aspartic proteinase levels in seeds of different angiosperms. *Phytochem* 44: 389-392.

- Warington K (1923) The effect of boric acid and borax on the broad bean and certain other plants. *Ann Bot* 37: 629–672.
- Watanabe A, Nong V, Zhang D, Arahira M, Yeboah N, Udaka K, Fukazawa C (1999) Molecular cloning and ethylene-inducible expression of Chib1 chitinase from soybean. *Biosci Biotechnol Biochem* 63: 251–256.
- Yu Q, Baluška F, Jasper F, Menzel D, Goldbach H (2003) Short-term boron deprivation enhances levels of cytoskeletal proteins in maize, but not zucchini, root apices. *Physiol Plant* 117: 270 – 278.
- Yu Q, Wingender R, Schulz M, Baluska F, Goldbach H (2001) Short-term boron deprivation induces increased levels of cytoskeletal proteins in *Arabidopsis* roots. *Plant Biol* 3: 335–340.

CHAPTER 4

METABOLIC ANALYSIS REVEALED ALTERED AMINO ACID PROFILES IN *Lupinus albus* ORGANS DUE TO BORON DEFICIENCY



Metabolic analysis revealed altered amino acid profiles in *Lupinus albus* organs due to boron deficiency

Alves M¹, Chicau P¹, Matias H¹, Passarinho J^{1,2}, Pinheiro C¹, Ricardo CP^{1,*}

¹Instituto de Tecnologia Química e Biológica, Universidade Nova de Lisboa, Av. da República, 2780-157 Oeiras, Portugal.

²Instituto Nacional de Recursos Biológicos/L-INIA, Av. da República, 2784-505 Oeiras, Portugal.

*Corresponding author: ricardo@itqb.unl.pt

CONTRIBUTION: The first author had the major contribution for all the work described, with the exception of free amino acid HPLC analysis and in handling of the NMR spectrometer.

KEYWORDS: *Lupinus albus*, B deficiency, metabolite, branched-chain amino acids, glycine.

SUMMARY

In B-deficient *Lupinus albus*, marked changes in the free amino acid content of leaf-blades, petioles, apices, hypocotyls and roots were observed while the absence of variation in malate, and the minor changes in the content of sugars, can be an indication that the central carbohydrate metabolism is little affected by B deficiency. The changes observed in the different organs were qualitatively different, with increases in some amino acids while others decreased, suggesting that the observed alteration in the free amino acid content is not the result of an indiscriminate effect on protein metabolism. Increased branched-chain amino acids content was observed in all the studied organs, which supports the hypothesis of the involvement of B with the cytoskeleton. Glycine decreased in leaf-blades and active growing organs (apexes and roots), but was not detected in petioles and hypocotyls. Considering the proposed role of glycine in plant signalling processes, there could also be an association of this amino acid with the decreased growth rates observed in the active growing organs of *L. albus* plants due to B deficiency.

INTRODUCTION

Boron is an essential microelement for higher plants (Warington 1923) that is considered to be important for plant cell wall structure through its formation of diester bridges between rhamnogalacturonan-II molecules (O'Neill et al. 1996, 2001, Kobayashi et al. 1999). However, this participation does not seem to explain all the plant symptoms due to B deprivation, or the recent evidence of B requirement in animal metabolism (Nielson 1997). Several recent reviews propose the additional participation of B in metabolic supporting activities (Brown et al. 2002, Goldbach and Wimmer 2007, Bonilla et al. 2009).

Free amino acids, besides their obvious role of nitrogen storage molecules and participation in protein synthesis, can also act as signalling molecules. Recently, amino acids were proposed to regulate multiple processes related to gene expression, not only on a global scale, but also by inducing preferential changes in the translation of mRNA encoding particular proteins or family of proteins (Kimball and Jefferson 2006).

Despite the increasing recognition of the importance of free amino acids in plant metabolism this matter has been little studied and there is also a lack of information on the effect of stress on the free amino acids of distinct plant organs. The fact that B deficiency virtually affects every aspect of plant metabolism, prompted us to

investigate the effects of B deficiency on the free amino acid content of several organs (leaf-blades, petioles, apices, hypocotyls and roots) of *Lupinus albus* plants.

MATERIALS AND METHODS

Plant material

Lupin seeds (*Lupinus albus* cv. Rio Maior) were pre-germinated in distilled water for 48h and then sown in white sand and grown under controlled conditions of temperature (19/25°C, night/day) and light (intensity of $250\mu\text{mol m}^{-2} \text{s}^{-1}$ PAR and 12h photoperiod). The plants were watered every other day with a nutritive solution (Arnon 1938) containing either 0 or $23.1\mu\text{M}$ B (Alves et al. 2006). The different organs of the control and B-deficient *L. albus*: leaf-blade, petiole, apex, hypocotyl and root were harvested 28 days after germination, frozen in liquid nitrogen and stored at -80°C.

Metabolite analysis

For the analysis of metabolites, the frozen samples (3-10g, in triplicate) were ground to a fine powder and then boiled in ddH₂O (4mL/g fresh weight) for 5min before centrifugation (15min; 14,000xg) and recovery of the supernatant. The supernatant was

lyophilized to dryness and resuspended in 2mL of an aqueous solution containing D₂O (5.8M), Na₂EDTA (2.5mM) and NaN₃ (2.5mM).

The major soluble metabolites were analysed with ¹³C-nuclear magnetic resonance (¹³C-NMR) in a Bruker Avance II NMR spectrometer (400MHz) at 100MHz using a broadband probe head (5mm diameter), with the following parameters: number of scans 1500; repetition delay 1.86s; sweep width 24kHz and a 30° pulse flip angle. Proton decoupling was applied during the acquisition only. The temperature of the probe head was kept at 25°C. Data was processed with 4Hz exponential line broadening. Unequivocally identified resonances from each metabolite were integrated using the Mnova software v5.1.1 (Mestrelab Research S.L.). Fully relaxed spectra, with a 31.36s repetition delay, were acquired and used to calculate area saturation correction factors of the relevant resonances. Chemical shifts are expressed in ppm relative to dioxan (66.66ppm) that was also used as an internal concentration standard (0.194M) for the metabolite quantification. Resonances due to asparagine, malate, *myo*-inositol, sucrose, glucose and fructose were identified by adding a 100mM solution of the pure compound to a tissue extract.

Free amino acids were analysed by high performance liquid chromatography (HPLC). The samples were previously cleaned-up

with solid phase extraction (Sep-Pak® C-18 Cartridges) to remove low molecular weight contaminants. After derivatization with phenylisothiocyanate, the free amino acids were analyzed by HPLC (Alliance 2695, Waters) using a RP C18 (Nova-Pak, 60Å, 4µm, 3.9 x 300mm) at an absorbance of 254nm (PDA 2996, Waters). The eluents used for the chromatographic separation were (A) 0.07M sodium acetate, pH 6.55 with 2.5% acetonitrile, and (B) 45% acetonitrile and 15% methanol (Cohen et al. 1986). The free amino acids analysed were alanine, β-alanine, aminoadipic acid, α-aminobutyric acid, γ-aminobutyric acid, β- aminoisobutyric acid, arginine, aspartic acid, carnosine, citrulline, cysteic acid, cystine, glycine, glutamic acid, glutamine, histidine, hydroxylysine, hydroxyproline, isoleucine, leucine, lysine, methionine, ornithine, phenylalanine, phosphoethanolamine, phosphoserine, proline, taurine, threonine, tryptophan, tyrosine and valine.

Statistical analysis

Significant changes in metabolite concentrations between stress and control were evaluated by the Student's *t*-test ($p < 0.05$) using the software SigmaStat v3.10 (Systat Software, Inc.) and correlation analysis performed with Microsoft Excel analysis ToolPak.

RESULTS AND DISCUSSION

Biomass analysis of L. albus plants grown under B deficiency

The morphological symptoms of B deficiency in *Lupinus albus* plants are quite evident four weeks after germination, and are expressed as necrosis of the terminal bud, darkened and deformed leaf-blades, cracked petioles and hypocotyls and marked reduction in root development (Alves et al. 2006). Roots showed the greatest decrease in biomass (47%), followed by petioles and leaf-blades (43% and 37%, respectively; Figure 4.1). No statistically significant differences were found for apices (16% decrease) or hypocotyls (15% increase).

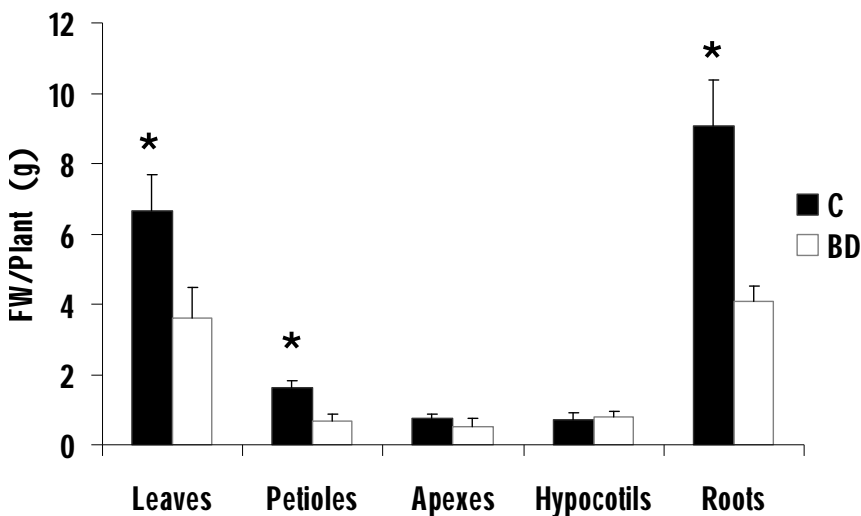


Figure 4.1. Biomass analysis of *Lupinus albus* plants grown under B deficiency. Fresh weight (FW) of the leaf-blades, petioles, apices, hypocotyls and roots of the *L. albus* grown with (C) or without B (BD). Significant changes were evaluated by the Student's *t*-test (*, $p < 0.05$).

Metabolite analysis

The metabolite changes observed in the different organs of *Lupinus albus* plants grown for four weeks under B deficiency are shown in Table 4.1. Concerning carbohydrates, sucrose content of B-deficient plants was increased in all organs, while fructose and glucose increased in leaf-blades, but decreased in the other organs. This observation seems to indicate a reduced sucrose consumption that could be due to stress resulting from B deficiency. In the leaf-blades the excess of sucrose was probably hydrolysed, leading to the increased content of the two reducing sugars, while in the remaining organs sucrose could accumulate and not be hydrolysed. Therefore, the reduced plant growth observed due to B deficiency does not seem to result from sugar limitation.

The *myo*-inositol content showed no change in leaves of B-deficient plants. A high correlation was observed between *myo*-inositol and sucrose ($r=0.99$) and with glucose ($r=0.90$), indicating that these compounds have similar metabolic behaviour under B deficiency.

Malate is a key product of plant metabolism that is thought to be the ultimate product of glycolysis, and is seen as an important marker of this pathway (Schulze et al. 2002). In *L. albus*, malate is known to accumulate in large amounts in shoots (Peñaloza et

Table 4.1. The concentration (nmol/g fresh weight) of the major soluble compounds identified by ^{13}C -NMR and of the free amino acids identified by HPLC in the different organs of *L. albus* grown with or without B. Data are the means \pm sd. Statistical analysis was made with Student's t-test ($p < 0.05$) and statistical significant changes resulting from B deficiency are marked for decreases (■) or increases (■) in the metabolite.

| Metabolism | Compound | Treatment | Leaf-blades | Petioles | Apex | Hypocotyls | Roots |
|------------------------------------|--------------|-----------|-------------------------------|-------------------------------|-------------------------------|-------------------------------|-------------------------------|
| Carbohydrates | Sucrose | BD | (1.0 \pm 0.3) $\times 10^6$ | (1.4 \pm 0.1) $\times 10^6$ | (6.5 \pm 0.5) $\times 10^5$ | (1.4 \pm 0.2) $\times 10^6$ | (5 \pm 2) $\times 10^5$ |
| | | C | <1 $\times 10^5$ | (6.4 \pm 0.9) $\times 10^5$ | (6.1 \pm 0.3) $\times 10^5$ | (8 \pm 2) $\times 10^5$ | (2.2 \pm 0.9) $\times 10^5$ |
| | Fructose | BD | (7 \pm 3) $\times 10^5$ | <1 $\times 10^5$ | <1 $\times 10^5$ | <1 $\times 10^5$ | <1 $\times 10^5$ |
| | | C | (3 \pm 2) $\times 10^5$ | (3.2 \pm 0.5) $\times 10^5$ | <1 $\times 10^5$ | <1 $\times 10^5$ | (3 \pm 1) $\times 10^5$ |
| | Glucose | BD | (9 \pm 3) $\times 10^5$ | (1.4 \pm 0.4) $\times 10^6$ | (3 \pm 1) $\times 10^5$ | <1 $\times 10^5$ | 0.1 \pm 0.1 |
| | | C | (3 \pm 2) $\times 10^5$ | (2.4 \pm 0.8) $\times 10^6$ | (7.4 \pm 0.3) $\times 10^5$ | <1 $\times 10^5$ | (1.9 \pm 0.6) $\times 10^5$ |
| TCA cycle | myo-Inositol | BD | (3 \pm 2) $\times 10^5$ | <1 $\times 10^5$ | <1 $\times 10^5$ | <1 $\times 10^5$ | <1 $\times 10^5$ |
| | | C | (1.9 \pm 0.6) $\times 10^5$ | <1 $\times 10^5$ | <1 $\times 10^5$ | <1 $\times 10^5$ | <1 $\times 10^5$ |
| | Malate | BD | (4 \pm 1) $\times 10^6$ | (5.6 \pm 0.6) $\times 10^6$ | (2.6 \pm 0.7) $\times 10^6$ | (2.4 \pm 0.6) $\times 10^6$ | (1.7 \pm 0.4) $\times 10^6$ |
| | | C | (6 \pm 1) $\times 10^6$ | (5.6 \pm 0.5) $\times 10^6$ | (4.1 \pm 0.6) $\times 10^6$ | (3.4 \pm 0.9) $\times 10^6$ | (1.6 \pm 0.4) $\times 10^6$ |
| Aromatic amino acids | Tyrosine | BD | 0.14 \pm 0.02 | <4 $\times 10^{-3}$ | <4 $\times 10^{-3}$ | 0.12 \pm 0.06 | <4 $\times 10^{-3}$ |
| | | C | <4 $\times 10^{-3}$ | <4 $\times 10^{-5}$ | <4 $\times 10^{-5}$ | <4 $\times 10^{-5}$ | <4 $\times 10^{-5}$ |
| Branched-chain amino acids (BCCAs) | Isoleucine | BD | 0.4 \pm 0.1 | 0.16 \pm 0.04 | 0.29 \pm 0.04 | 0.24 \pm 0.07 | 0.11 \pm 0.03 |
| | | C | 0.17 \pm 0.07 | 0.07 \pm 0.01 | 0.17 \pm 0.04 | <4 $\times 10^{-3}$ | <4 $\times 10^{-3}$ |
| | Leucine | BD | 0.17 \pm 0.02 | 0.11 \pm 0.05 | 0.14 \pm 0.04 | 0.16 \pm 0.03 | 0.07 \pm 0.03 |
| | | C | 0.08 \pm 0.03 | <4 $\times 10^{-3}$ | <4 $\times 10^{-3}$ | <4 $\times 10^{-3}$ | <4 $\times 10^{-3}$ |
| GABA shunt | Valine | BD | 0.5 \pm 0.1 | 0.5 \pm 0.2 | 1.3 \pm 0.2 | 0.6 \pm 0.1 | 0.22 \pm 0.08 |
| | | C | 0.24 \pm 0.04 | 0.19 \pm 0.05 | 0.4 \pm 0.1 | 0.23 \pm 0.08 | 0.11 \pm 0.02 |
| | Alanine | BD | 1.0 \pm 0.3 | 0.48 \pm 0.05 | 1.2 \pm 0.1 | 0.8 \pm 0.3 | 0.4 \pm 0.2 |
| | | C | 1.0 \pm 0.2 | 0.23 \pm 0.05 | 0.6 \pm 0.2 | 0.30 \pm 0.06 | 0.25 \pm 0.04 |
| | GABA | BD | 1.2 \pm 0.4 | 0.73 \pm 0.07 | 1.6 \pm 0.6 | 0.9 \pm 0.2 | 0.3 \pm 0.1 |
| | | C | 2.0 \pm 0.4 | 0.33 \pm 0.03 | 0.7 \pm 0.4 | 0.27 \pm 0.04 | 0.13 \pm 0.01 |

Table 4.1 (cont) The concentration (nmol/g fresh weight) of the major soluble compounds identified by ¹³C-NMR and of the free amino acids identified by HPLC in the different organs of *L. albus* grown with or without B. Data are the means ± sd. Statistical analysis was made with Student's t-test (p<0.05) and statistical significant changes resulting from B deficiency are marked for decreases (■) or increases (■) in the metabolite.

| Metabolism | Compound | Treatment | Leaf-blades | Petioles | Apex | Hypocotyls | Roots |
|------------------------------|---------------|-----------|-----------------------|---------------------------|---------------------------|---------------------------|---------------------|
| Glutamate family amino acids | Glutamate | BD | 0.44±0.03 | 0.64±0.05 | 1.2±0.2 | 0.6±0.2 | 0.57±0.04 |
| | | C | 0.42±0.07 | 0.5±0.1 | 1.1±0.4 | 0.55±0.05 | 0.48±0.06 |
| | Arginine | BD | 0.24±0.03 | 0.27±0.05 | 0.5±0.2 | 0.1±0.2 | 0.20±0.02 |
| | | C | 0.25±0.04 | 0.20±0.02 | 0.31±0.07 | 0.22±0.05 | 0.196±0.003 |
| | Histidine | BD | 0.17±0.03 | 0.3±0.1 | 0.7±0.3 | 0.62±0.04 | 0.13±0.06 |
| | | C | <4x10 ⁻⁵ | 0.05±0.04 | 0.13±0.04 | 0.13±0.06 | 0.074±0.005 |
| Asparate family amino acids | Proline | BD | 0.3±0.1 | 0.8±0.3 | 0.8±0.4 | 0.5±0.1 | 0.2±0.1 |
| | | C | 0.15±0.04 | 0.07±0.01 | 0.12±0.05 | <4x10 ⁻⁵ | 0.15±0.08 |
| | Aspartic acid | BD | 0.9±0.2 | 1.26±0.05 | 1.8±0.2 | 1.0±0.2 | 0.6±0.2 |
| | | C | 1.45±0.07 | 0.88±0.09 | 1.4±0.5 | 0.6±0.2 | 0.37±0.02 |
| | Lysine | BD | 0.14±0.05 | 0.13±0.02 | 0.17±0.04 | 0.14±0.01 | 0.08±0.02 |
| | | C | 0.20±0.03 | 0.07±0.02 | 0.13±0.02 | 0.08±0.01 | 0.07±0.02 |
| Others | Threonine | BD | 0.23±0.07 | 0.28±0.08 | 0.44±0.03 | 0.28±0.08 | 0.10±0.04 |
| | | C | 0.16±0.04 | 0.09±0.01 | 0.18±0.07 | 0.09±0.04 | 0.07±0.01 |
| | Asparagine | BD | (7±3)x10 ⁵ | (3.0±0.5)x10 ⁶ | (2.4±0.7)x10 ⁶ | (2.3±0.4)x10 ⁶ | <1x10 ⁵ |
| | | C | (2±2)x10 ⁵ | (6±3)x10 ⁵ | (3.5±0.9)x10 ⁵ | <1x10 ⁵ | <1x10 ⁵ |
| | Glycine | BD | <4x10 ⁻⁵ | <4x10 ⁻⁵ | <4x10 ⁻⁵ | <4x10 ⁻⁵ | <4x10 ⁻⁵ |
| | | C | 0.21±0.08 | <4x10 ⁻⁵ | 0.13±0.03 | <4x10 ⁻⁵ | 0.06±0.01 |

al. 2002), but under stressful conditions, such as water-deficit, the malate content was decreased in all organs, but most markedly in the leaves and roots (Pinheiro et al. 2004). Under B deficiency, the absence of variation in malate, and the minor changes in the content of sugars, can be an indication that the central carbohydrate metabolism is little affected by B deficiency (Figure 4.2).

Regarding the amino acids, we observed marked changes in their content due to B deficiency, but distinct patterns of variation were observed between them and also between the several organs (Table 4.1, Figure 4.2). The fact that in every organ some amino acids increased while others decreased, suggests that the observed alterations are not the result of indiscriminate protein degradation. For instance, in roots, despite the small number of amino acids changed due to B deficiency the content of leucine, valine, isoleucine and GABA increased but the content of glycine decreased. In the leaf-blades, the additional changes were observed: tyrosine, leucine, valine, isoleucine and histidine increased, while aspartic acid and glycine decreased. Such an increase in tyrosine could be related to the known increase in phenol content of leaves as a consequence of B deficiency (Blevins and Lukaszewski, 1998). Changes in tyrosine could also be important due to the participation of this amino acid in the synthesis of signalling molecules responsible for growth modulation processes (Lea and Forde 1994).

The increase of the branched-chain amino acids (BCAAs), leucine, isoleucine and valine, in leaf-blades appears to be an interesting observation since, in animal tissues BCAAs are known to participate in the regulation of skeletal muscle protein synthesis, leucine being

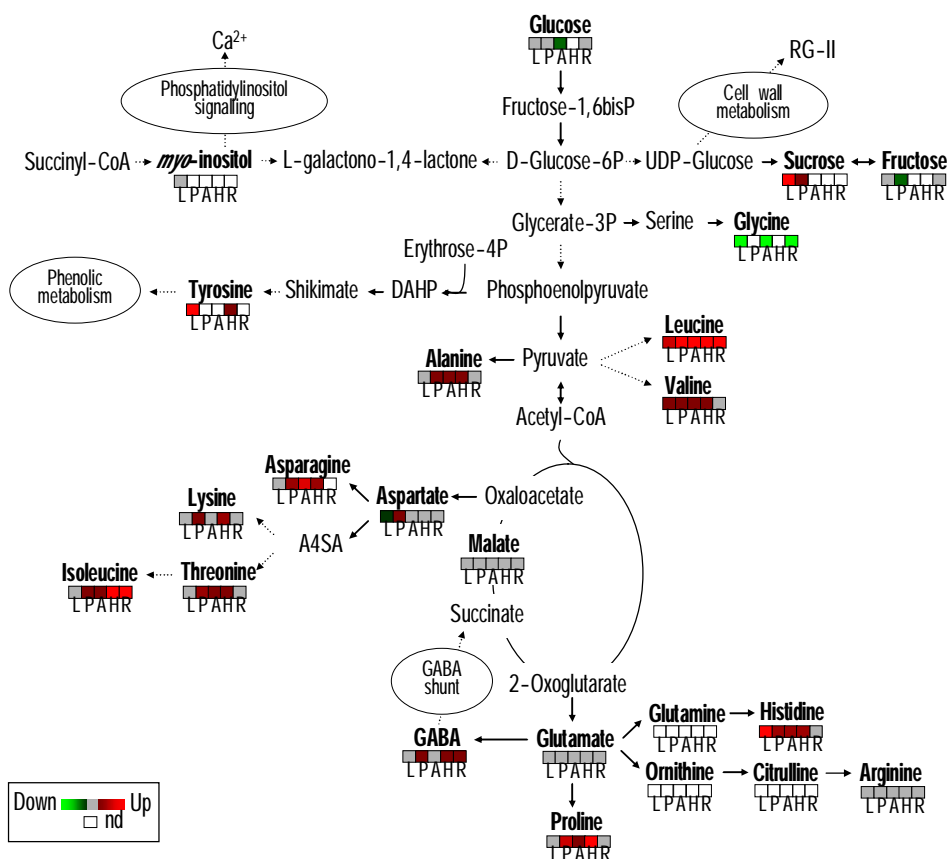


Figure 4.2. Metabolic sequences affected by B deficiency in *L. albus* plants, as predicted by the analysed metabolites of leaf-blades (L), petioles (P), apices (A), hypocotyls (H) and roots (R). Decreased metabolite contents are in green and increased changes in red. Non altered content is in grey, whereas white represents non detected metabolites. Abbreviations: A4SA, aspartate 4-semialdehyde; DAHP, 3-Deoxy-D-arabino-heptulosonate-7-phosphate; RG-II, rhamnogalacturonan-II.

the most effective with regard to the regulation of these proteins (Kimball and Jefferson 2006, Rhoads and Wu 2009). The importance of actin in the cytoskeleton and the observation that B deficiency affects the morphology of leaf-blades and roots, may suggest some association between the increase in BCAAs and some developmental processes affected by B deficiency. Enhanced levels of cytoskeletal proteins and abnormal polymerization patterns, in *Arabidopsis* and maize roots, were observed in short-term B deficiencies (Yu et al. 2001, 2003), which could be related to the postulated hypothesis of BCAAs influencing cytoskeleton structure. The BCAAs also markedly increased in *Arabidopsis* when subjected to drought stress (Urano et al. 2009).

Histidine, which increased in leaf-blades, was also found to increase in heavy metal stress and its participation as a chelator or transporter of metal ions was proposed. In addition, histidine is involved in several biosynthetic pathways, such as that of purines, pyrimidines, pyridine nucleotides, folates and tryptophan (Stepansky and Leustek 2006), all of which could have been affected by the B-deficiency.

The decrease in the leaf-blades of aspartic acid and glycine could be significant to the B deficiency response. Aspartic acid serves as the nitrogen donor in numerous aminotransferase reactions and could be related with decreased nitrate content due to B deficiency, an

observation described in tobacco leaves (Camacho-Cristóbal and González-Fontes 1999).

In what concerns glycine, this amino acid was also decreased in the apexes and roots (active growing organs), but was not detected in petioles and hypocotyls.

Glycine has a well established role in the signalling processes of the mammalian central system and in plants there is also evidence for glycine participation in signalling processes. The postulated binding of glycine to glutamate receptors could explain why the increased levels of glycine induced higher cytosolic calcium concentrations in *Arabidopsis* (Dubos et al. 2003). It is noteworthy to refer the observed amelioration of some B deficiency symptoms after the addition of calcium (Bolaños et al. 2004).

Comparing amino acid changes in leaf-blades to those occurring in petioles, apexes and hypocotyls, it is evident that a higher number of amino acids were statistically affected by B deficiency in these three organs. In addition to the specific metabolic changes that might be occurring in each organ, the possibility of some amino acids being translocated from the leaf-blades (Fischer et al. 1998) should not be excluded. For instance, asparagine, which increased only slightly in the leaf-blades under B deficiency, is one form of N-transport from source to sink tissues (Lea et al. 2007) and it did markedly increase in apexes, petioles and hypocotyls. Such an

accumulation in the petioles and hypocotyls may be an indication of temporary functioning of these organs as transient stores, in response to B deficiency, similarly to what was previously suggested for *L. albus* hypocotyls in response to drought stress (Pinheiro et al. 2004).

The asparagine increased content is highly correlated with the proline increase ($r=0.98$) under B-deficient conditions, and the two amino acids are known to accumulate under conditions of stress. Proline, in particular, is associated with a wide range of biotic and abiotic stresses (Verbruggen and Hermans 2008). The most common proposed hypothesis is that proline acts as an osmotic and a protective agent for cytosolic enzymes and membrane structures, but it may also be part of stress signalling processes responsible for adaptive stress responses (Verbruggen and Hermans 2008). The observed proline significant increase in the petioles, apexes and hypocotyls can be envisaged as an important protective role in the apexes, but it is less clear why this amino acid should also highly accumulate in both petioles and hypocotyls.

GABA is another amino acid commonly associated with biotic and abiotic stresses (Fait et al. 2007). GABA and alanine, both increased in the petioles, apexes and hypocotyls, showing a high correlation value ($r=0.90$), and possibly their production resulted from a common metabolic process, known as the GABA shunt. The

accumulation of alanine and GABA in low oxygen stressed plants has been reported to be partially associated with the GABA shunt (Miyashita and Good 2008). When GABA production was inhibited, the accumulation of reactive oxygen species was prevented (Fait et al. 2005). Thus, it could be considered that increased GABA levels could be related to the described accumulation of reactive oxygen species in B-deficient plants (Cakmak and Römheld 1997, Koshiba et al. 2009).

CONCLUSIONS

The plant responses to stress constitute a complex framework of metabolic events that also result from complex plant organs interactions. This complexity is well expressed in the free amino acids profile that was quite distinct in the different organs of *Lupinus albus* grown under B deficiency. The observed changes in the several amino acids indicate that B deficiency is affecting many aspects of the plant metabolism including certain signalling processes. The increase in BCAAs content in all the studied organs supports the hypothesis of damaged cytoskeleton structure due to B deficiency. Boron requirement in dynamic microtubule rearrangements, like in the early phases of tissue differentiation in zebra fish (Rowe and Eckert 1999) and in *Larix* embryogenesis (Behrendt and

Zoglauer 1996) is consistent with B involvement in the organization of the cytoskeleton structure.

Thus, amino acid biosynthesis seems to be, directly or indirectly, associated with various aspects of plant response to B deficiency.

ACKNOWLEDGMENTS

We acknowledge the facilities of the National Nuclear Magnetic Resonance Network - CERMAX (ITQB, Oeiras, Portugal) and the financial support (grant n°. FCT SFRH/BD/18273/2004) from FCT. We also thank Phil Jackson for revising the manuscript.

REFERENCES

- Alves M, Francisco R, Martins I, Ricardo CPP (2006) Analysis of *Lupinus albus* leaf apoplastic proteins in response to boron deficiency. *Plant Soil* 279: 1–11.
- Arnon D (1938) Microelements in culture–solution experiment with higher plants. *Amer J Bot* 25: 322–325.
- Behrendt U, Zoglauer K (1996) Boron controls suspensor development in embryogenic cultures of *Larix decidua*. *Physiol Plant* 97: 321–326.
- Blevins D, Lukaszewski K (1998) Boron in plant structure and function. *Annu Rev Plant Physiol Plant Mol Biol* 49: 481–500.

- Bolaños L, Lukaszewski K, Bonilla I, Blevins D (2004) Why boron? *Plant Physiol Biochem* 42: 907-912.
- Bonilla I, Blevins D, Bolaños L (2009) Boron Functions in Plants: Looking Beyond the Cell Wall. In: *Plant Physiology*. Eds: Taiz L, Zeiger E. 4th Ed. IOP Publishing physics web. <http://4e.plantphys.net>. Accessed Dec 2009.
- Brown P, Bellaloui N, Wimmer M, Bassil E, Ruiz J, Hu H, Pfeiffer H, Dannel F, Romheld V (2002) B in plant biology. *Plant Biol* 4: 203-223.
- Cakmak I, Römheld V (1997) Boron deficiency-induced impairments of cellular functions in plants. *Plant Soil* 193: 71-83.
- Camacho-Cristóbal J, González-Fontes A (1999) Boron deficiency causes a drastic decrease in nitrate content and nitrate reductase activity, and increases the content of carbohydrates in leaves from tobacco plants. *Planta* 209: 528-536.
- Cohen S, Bidlingmeyer B, Tarvin T (1986) PITC derivatives in amino acid analysis. *Nature* 320: 769-770.
- Dubos C, Huggins D, Grant G, Knight M, Campbell M (2003) A role for glycine in the gating of plant NMDA-like receptors. *Plant J* 35: 800-810.
- Fait A, Fromm H, Walter D, Galili G, Fernie A (2007) Highway or byway: the metabolic role of the GABA shunt in plants. *Trends Plant Sci* 13: 14-19.
- Fait A, Yellin A, Fromm H (2005) GABA shunt deficiencies and accumulation of reactive oxygen intermediates: insight from *Arabidopsis* mutants. *FEBS Lett* 579: 415-420.

- Fischer W, André B, Rentsch D, Krolkiewicz S, Tegeder M, Breitzkreuz k Frommer W (1998) Amino acid transport in plants. Trends Plant Sci 3: 188-194.
- Goldbach H, Wimmer M (2007) Boron in plants and animals: Is there a role beyond cell-wall structure? J Plant Nutr Soil Sci 170: 39-48.
- Kimball S, Jefferson L (2006) New functions for amino acids: effects on gene transcription and translation. Am J Clin Nutr 83: 500S-507S.
- Kobayashi M, Nakagawa H, Asaka T, Matoh T (1999) Borate-rhamnogalacturonan II bonding reinforced by Ca^{2+} retains pectic polysaccharides in higher-plant cell walls. Plant Physiol 119: 199-203.
- Koshiba T, Kobayashi M, Matoh T (2009) Boron Nutrition of Tobacco BY-2 Cells. V. Oxidative Damage is the Major Cause of Cell Death Induced by Boron Deprivation. Plant Cell Physiol 50: 26-36.
- Lea P, Forde B (1994) The use of mutants and transgenic plants to study amino acid metabolism Plant Cell Environ 17: 541-556.
- Lea P, Sodek L, Parry M, Shewry P, Halford N (2007) Asparagine in plants. Ann Appl Biol 150: 1-26.
- Miyashita Y, Good A (2008) Contribution of the GABA shunt to hypoxia-induced alanine accumulation in roots of *Arabidopsis thaliana*. Plant Cell Physiol 49: 92-102.
- Nielson F (1997) Boron in human and animal nutrition. Plant Soil 193: 199-208.

- O'Neill MA, Warrenfeltz D, Kates K, Pellerin P, Doco T, Darvill A, Albersheim P (1996) Rhamnogalacturonan-II, a pectic polysaccharide in the walls of growing plant cell, forms a dimer that is covalently cross linked by a borate ester. *J Biol Chem.* 271: 22923–22930.
- O'Neill M, Eberhard S, Albersheim P, Darvill A (2001) Requirement of Borate Cross-Linking of Cell Wall Rhamnogalacturonan II for *Arabidopsis* Growth. *Science* 294: 846–849.
- Peñaloza E, Corcuera L, Martinez J (2002) Spatial and temporal variation in citrate and malate exudation and tissue concentration as affected by P stress in roots of white lupin. *Plant Soil* 241: 209–221.
- Pinheiro C, Passarinho J, Ricardo CP (2004) Effect of drought and rewatering on the metabolism of *Lupinus albus* organs. *J Plant Physiol* 161: 1203–1210.
- Rhoads J, Wu G (2009) Glutamine, arginine, and leucine signaling in the intestine. *Amino Acids* 37: 111–122.
- Rowe R, Eckhert C (1999) Boron is required for zebrafish embryogenesis. *J Exp Biol* 202: 1649–1654.
- Schulze J, Tesfaye M, Litjens R, Bucciarelli B, Trepp G, Miller S, Samac D, Allan D, Vance C (2002) Malate plays a central role in plant nutrition. *Plant Soil* 247: 133–139.
- Stepansky A, Leustek T (2006) Histidine biosynthesis in plants. *Amino Acids* 30: 127–142.
- Urano K, Maruyama K, Ogata Y, Morishita Y, Takeda M, Sakurai N, Suzuki H, Saito K, Shibats D, Kobayashi M, Yamaguchi-Shinozaki K, Shinozaki K (2009) Characterization of ABA-

regulated global responses to dehydration in *Arabidopsis* by metabolomics. *Plant J* 57: 1065–1078.

Verbruggen N, Hermans C (2008) Proline accumulation in plants: a review. *Amino Acids* 35:753–759.

Warington K (1923) The effect of boric acid and borax on the broad bean and certain other plants. *Ann Bot* 37: 629–672.

Yu Q, Baluška F, Jasper F, Menzel D, Goldbach H (2003) Short-term boron deprivation enhances levels of cytoskeletal proteins in maize, but not zucchini, root apices. *Physiol Plant* 117: 270 – 278.

Yu Q, Wingender R, Schulz M, Baluska F, Goldbach H (2001) Short-term boron deprivation induces increased levels of cytoskeletal proteins in *Arabidopsis* roots. *Plant Biol* 3: 335–340.

CHAPTER 5

TRANSCRIPTIONAL ANALYSIS OF SHORT-TERM BORON DEFICIENCY IN ARABIDOPSIS REVEALED ALTERED SULPHUR METABOLISM



Transcriptional analysis of short-term boron deficiency in *Arabidopsis* indicates altered sulphur metabolism

Alves M¹, Saibo NJM¹, Gonçalves D^{1,2}, Ricardo CP^{1,*}

¹Instituto de Tecnologia Química e Biológica, Av. República, Estação Agronómica Nacional, 2780-157 Oeiras, Portugal.

²Instituto Superior Técnico Av. Rovisco Pais, 1049-001 Lisboa, Portugal.

*Corresponding author: ricardo@itqb.unl.pt

CONTRIBUTION: The first author performed all the experimental work and writing with the exception of the GeneChip hybridization, GeneChip data analysis and boron quantification.

KEYWORDS: *Arabidopsis thaliana*, Boron deficiency, Calcium deficiency, cytoskeleton, sulphur metabolism.

SUMMARY

For more than 80 years, boron (B) has been recognized as an essential microelement for higher plants, but in spite of this its precise role remains elusive. In order to gain insight into the participation of B in the metabolism of higher plants, we have analysed *Arabidopsis thaliana* for transcriptional alterations caused by short-term B deficiency. Given that several studies suggest an interaction between B and calcium (Ca) in plants, the transcriptional profile of *A. thaliana* plants under Ca-deficiency was also analysed. The microarray analysis revealed that the expression of 102 genes was altered 2 days after withholding B and 208 by day 4. For the Ca deficiency, 2 days after Ca withholding morphological symptoms were already evident and more than 2,500 genes were altered, and at 4 days the plants showed necrotic symptoms and only 270 genes were differentially expressed. Of the 298 B-responsive genes, 37 were identified as having the same expression trend at 2 and 4 days of deficiency (15 up-regulated and 22 down-regulated) and, so, were selected for further analysis. Thirteen genes that had a fold-change higher than 2 were chosen for RT-qPCR analysis and since we were able to validate all of them we included all the 37 genes for the detailed discussion. In this set, we found genes related with cell wall biosynthesis (24%), sulphur metabolism (24%), transcriptional factors and hormones

(18%), while genes related with miscellaneous and unknown functions represent 13% and 16%, respectively. Altered cell wall biosynthesis is a process already described for B-deficient plants, whereas altered sulphur metabolism due to B deficiency is a rather novel observation.

INTRODUCTION

Boron (B) is known to be an essential micronutrient for higher plants since 1923 (Warington 1923), but only in the 90's it was reported to participate in the cell wall structure as a cross linker of the cell wall pectin, rhamnogalacturonan-II (RG-II) (Kobayashi et al. 1996, O'Neill et al. 1996, 2001). However, this participation does not seem enough to explain all the symptoms of B deficiency in plants.

There are symptoms of short-term B deficiency that remain unexplained, such as the quantitative and qualitative changes observed in the phenolic metabolism of tobacco (Camacho-Cristóbal et al. 2002) and the increased levels of cytoskeletal proteins in both *Arabidopsis* and maize roots (Yu et al. 2001, 2003). The enhancement of cytoskeleton protein levels and the altered polymerization patterns were hypothesized to be involved in the mechanical reinforcement of the cell periphery complex. Another

observed symptom that results from short-term B deprivation is the inhibition of pectin endocytosis from the cell walls of meristematic cells in maize and wheat root apices, which could be related to endocytosis-mediated pectin signalling (Yu et al. 2002). Additionally, it has been suggested that the metabolic functions of calcium (Ca) and B in plants may be inter-linked. For instance, in nodulation processes some effects of B deficiency were ameliorated by Ca addition (Redondo-Nieto et al. 2003, Koshiba et al. 2010), and genetic studies showed that the expression of some genes affected by B deficiency could be reversed by Ca supplementation (Bolaños et al. 2004). Stability studies of different B fractions suggested that B cross-link of RG-II may result in conformational changes that can create binding sites for Ca ions, which increase pectin association and rigidity (Kobayashi et al. 1999). Since little is known about the roles of B in higher plants, and given the possible interactions between B and Ca, we have analysed the transcriptional profile of *Arabidopsis thaliana* plants subjected to 2 and 4 days of B or Ca deficiency. Our results provide insight into the early responses to B deprivation in *Arabidopsis*.

MATERIALS AND METHODS

Plant material and growth conditions

Arabidopsis thaliana cv. Columbia seeds were sown in 1L plastic containers filled with perlite, which has a low B content (Alves et al. 2006). After a period of 2 days at 4°C, the containers were transferred to a growth chamber (Fitoclima 700 EDT4, Aralab) at a temperature of 16/22°C (night/day), with a 16h photoperiod and a light intensity of $150\mu\text{mol m}^{-2} \text{s}^{-1}$, PAR. The plants were watered every other day with a complete nutrient solution (Arnon 1938) in which the B concentration was $25\mu\text{M}$ and the Ca concentration 3mM. Boron or Ca deficiencies were imposed 17 days after sowing by daily watering the plants with a B-free nutrient solution or with a Ca-free nutrient solution corrected for the nitrate content with magnesium and potassium salts.

The *Arabidopsis* plants were harvested 2 and 4 days after B or Ca suppression and kept frozen at -80°C until further use. The *Arabidopsis* plants grown for longer than 12 days under B or Ca deficiency were only used for morphological analysis. For dry weight determinations, the plants were oven-dried for 48h at 80°C.

Boron quantification

Boron was quantified following the Azomethine-H method (Sungur and Okur 2009) adapted for micro-assays. The plants were ashed

overnight in an oven at 550°C and the ashes dissolved in 3N HCl. The solution was filtered through a 0.45µm filter and 20 µL were used for analysis. To each well were added 20µL of water, 80µL of azomethine-H solution [0.45% (w/v) azometinhe-H and 2% (w/v) ascorbic acid], and 80µL of buffer solution at pH 5.7 [48% (w/v) ammonium acetate, 3% (w/v) EDTA disodium salt and 24% (v/v) acetic acid]. The mixture was kept at room temperature for 40min. The absorbance was measured in a UV/Vis 96-well plate reader (Powerwave xs, Biotek) at 420nm. A calibration curve was prepared using different concentrations of aqueous B solutions. All analyses were performed in triplicate.

RNA Isolation, Target Synthesis and Hybridization to Affymetrix GeneChips

Total RNA was extracted using the RNeasy Plant Mini Kit (Qiagen, Hilden, Germany). Concentration and purity were determined by spectrophotometry and integrity was confirmed using an Agilent 2100 Bioanalyzer with a RNA 6000 Nano Assay (Agilent Technologies, Palo Alto, CA). The RNA was processed for use on Affymetrix (Santa Clara, CA, USA) *Arabidopsis* ATH1 Genome Arrays, according to the manufacturer's One-Cycle Target Labelling Assay. Briefly, 5µg of total RNA spiked with Poly-A RNA controls (GeneChip Expression GeneChip Eukaryotic Poly-A RNA Control Kit;

Affymetrix) was used in a reverse transcription reaction (One-Cycle DNA synthesis kit; Affymetrix) to generate first-strand cDNA. After second-strand synthesis, double-stranded cDNA was used in an *in vitro* transcription (IVT) reaction to generate biotinylated cRNA (GeneChip Expression 3'-Amplification Reagents for IVT-Labeling; Affymetrix). Size distribution of the cRNA and fragmented cRNA, respectively, was assessed using an Agilent 2100 Bioanalyzer with a RNA 6000 Nano Assay. From the fragmented cRNA 15µg were used in a 300µL hybridization containing added hybridization controls. From the mixture, 200µL were hybridized on arrays for 16h at 45°C. Standard post hybridization wash and double-stain protocols (EukGE-WS2v4) were used on an Affymetrix GeneChip Fluidics Station 450. Arrays were scanned on an Affymetrix GeneChip scanner 3000 7G.

GeneChip Data Analysis

Scanned arrays were analyzed first with GCOS 1.4 software to obtain Absent/Present calls and for subsequent analysis with DNA-Chip Analyzer (dChip; release Jan 24, 2008) (<http://www.dchip.org>, Wong Lab, Harvard). The arrays were normalized to a baseline array with median CEL intensity of 160 by applying an Invariant Set Normalization Method (Li and Wong 2001a). Normalized CEL intensities of the 12 arrays (2 per

treatment) were used to obtain model-based gene expression indices based on a Perfect Match-only model (Li and Hung Wong, 2001b). Replicate data for the same sample type were weighted gene-wise by using inverse squared standard error as weights. Only genes called Present in at least one of the 12 arrays were kept for downstream analysis (17,797 genes). Thus, genes called Absent in all arrays were excluded. Li and Hung Wong (2001b) have shown that using the lower confidence bound of the fold change as a conservative estimate of the fold change is a more reliable as a ranking statistic for changes in gene expression. Therefore, all genes compared were considered to be differentially expressed if the lower confidence bound of the fold-change was greater than 90% and if above 1.2, between experiment and baseline, in both data sets. The lower confidence bound criterion means that we can be 90% confident that the fold-change is a value between the lower confidence bound and a variable upper confidence bound.

The data set for the 2 and 4 days of deficiency were analysed separately or jointly. In the separated analysis, a cut-off by FDR of 7.8 or 1.9 for 2 days of B or Ca deficiency, and 33.7 or 55.2 for 4 days of B or Ca deficiency, respectively was used.

Annotations for the 22,392 *Arabidopsis thaliana* transcripts that are represented on the GeneChip Arabidopsis ATH1 Genome Array were obtained from the NetAffx database (<http://www.affymetrix.com>) as

Arabidopsis thaliana transcriptional profile of January 2008 and imported into dChip using ChipInfo software (Zhong et al. 2003).

Reverse transcriptase quantitative real-time PCR (RT-qPCR) analysis

The mRNA from duplicate samples was isolated from *Arabidopsis* plant as previously described. A first-strand cDNA was synthesized using the iScript cDNA synthesis kit (Bio-Rad) according to the manufacturer's instructions. The primers used for the RT-qPCR were designed using the Roche Applied Science software (ProbeFinder v. 2.44 at <http://qpcr.probefinder.com>) for the thirteen genes: At5g24660, At1g62560, At1g65860, At4g04610, At1g64390, At4g15920, At4g26260, At1g10070, At1g79700, At1g03090, At2g19800, At1g21400, At1g76410 and At1g64660. The specificity of each primer pair was checked against the *Arabidopsis* genome using WU-BLAST 2.0 (<http://www.arabidopsis.org>). The RT-qPCR was performed with an iQ5 detection system (Bio-Rad) by using the SYBR Green supermix reagent (Bio-Rad) according to the manufacturer's instructions. The following standard thermal profile was used for all PCRs: Stage 1, 1 cycle at 95°C for 3min; Stage 2, 55 cycles at 95°C for 10s, primer specific annealing temperature for 20s and 72°C for 30s; Stage 3 (melting curve), from 65°C to

95°C with increments of 0.5°C/min. For primer sequences and annealing temperatures, see Supplementary Table 5.2.

A pool of cDNA from the control, B deficiency, and Ca deficiency samples was used for primer efficiency assays and the Ct values generated for each primer pair set over a range of dilutions were used to calculate the primer efficiency. The relative quantification method ($\Delta\Delta Ct$) was used to assess the relative expression of each gene analysed (Livak and Schmittgen, 2001) with two housekeeping genes, At5g53300, an *ubiquitin-conjugating enzyme E2 10 (UBC10)* and At1g13320, *protein phosphatase 2A subunit A3 (PP2A)* (Czechowski et al. 2005).

Gene annotation

For gene annotation, we have used the updated TAIR9 (The Arabidopsis Information Resource at <http://www.arabidopsis.org>) annotation. Genes were classified using the Gene Ontology categories to group functionally-related genes.

RESULTS AND DISCUSSION

Morphological symptoms of B deficiency

Seventeen-day-old *Arabidopsis thaliana* plants were subjected to B or Ca deficiency. The morphological symptoms of the B-deficient

plants were not visible 2 days after withholding B, but after 4 days, the plants showed slightly darker leaves. In contrast, the morphological symptoms of Ca-deficient plants were already visible 2 days after withholding Ca. Plants showed yellowish and curled leaves and reduced growth rates, while after 4 days of imposed deficiency, some symptoms of leaf necrosis were also visible. The plants grown under B deficiency for a few more weeks seemed to have arrested development, while the plants subjected to Ca deficiency died (data not shown).

Boron levels in the different treatments after 4 days of deficiency

Control plants were seen to contain $27 \pm 3 \mu\text{g B/g}$ dry weight, whereas the B content of plants grown under B deficiency for 4 days was reduced to $18 \pm 3 \mu\text{g B/g}$ dry weight (t -test $p < 0.05$). In Ca-deficient plants, no effect on B content was observed $27 \pm 2 \mu\text{g B/g}$ dry weight (t -test $p < 0.05$).

Gene expression analysis and functional categorization

The Arabidopsis GeneChip ATH1 oligonucleotide array, representing 22,392 annotated genes of the Arabidopsis genome, was used to study the gene expression profile of plants subjected to either B or Ca deficiency for 2 and 4 days. Two days after B deficiency, 102 genes were found to be differentially expressed while 4 days after B

deficiency twice as many (208) were differentially expressed. A different behaviour was observed for the Ca deficiency treatment. Two days after Ca deficiency 2,975 genes were found to be differentially expressed, while 4 days after Ca deficiency, only one tenth of the genes (270) were differentially expressed (Supplementary Table 5.1). The different sets of genes were categorized into biological processes (Figure 5.1), according to annotations present in *A. thaliana* database (TAIR9; <http://www.arabidopsis.org>).

The description of Ca as a versatile messenger mediating a variety of responses related with growth and stress conditions (Hepler 2005), might explain the higher number of the differentially expressed genes observed after only 2 days of Ca deficiency. It is likely that this results from a complexity of down-stream responses (secondary effects) which could also include adaptive responses towards the management of Ca deficiency. The reduced number of genes expressed 4 days after Ca deficiency (Figure 5.1a) could possibly be the expression of necrotic processes taking place. This dramatic effects observed in Ca-deficient plants precluded any direct comparison of the transcriptomics of Ca and B deficiency.

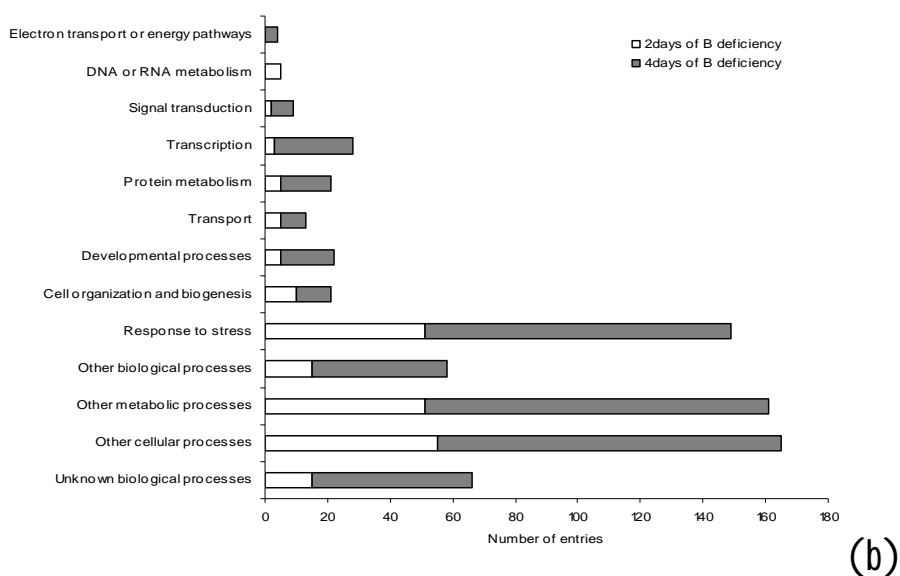
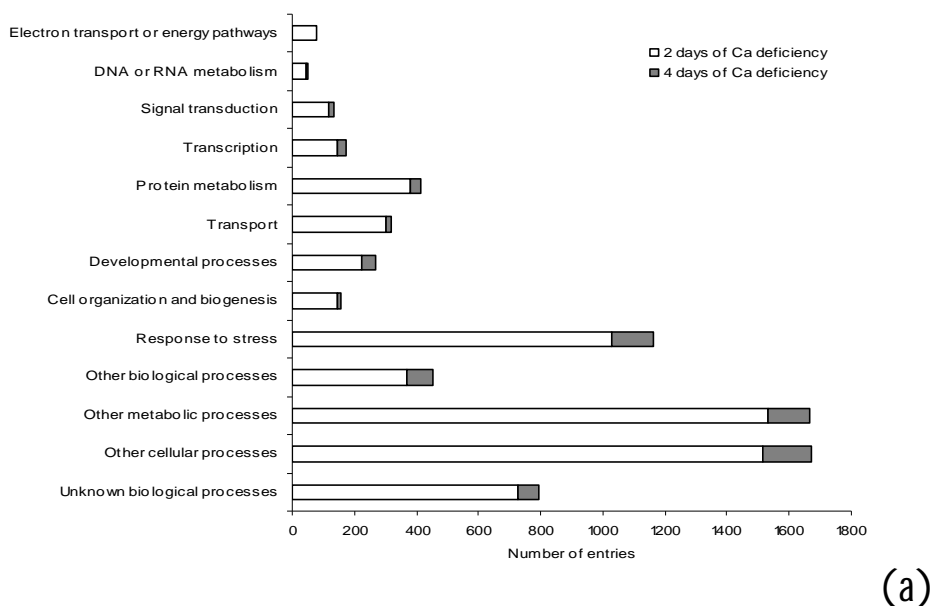


Figure 5.1. Biological functional categorization of the differentially expressed genes due to Ca or B deficiency (Supplementary Table 5.1). The genes that were differentially expressed 2 (□) and 4 days (■) after Ca (a) or B deficiency (b) were grouped according to gene ontology (GO) classification annotated in TAIR9 database (<http://www.arabidopsis.org>).

Boron is reported to cross-link cell wall pectin (RG-II) molecules, and its deficiency results in damaged cell wall structure (Kobayashi et al. 1996, O'Neill et al. 1996, 2001). In our study, genes related with cell organization and biogenesis (Figure 5.1b), namely cell elongation, cell wall loosening, xyloglucan biosynthesis and lignin biosynthetic process were altered by B deficiency. A recent report on B-deficient *Arabidopsis thaliana* root revealed that the majority of the cell wall genes that were down-regulated are associated with the cell elongation process, cell wall loosening and xyloglucan biosynthesis (Camacho-Cristóbal et al. 2008).

A structural role for B in membranes has also been proposed and it can explain a large number of reported effects of B deficiency in processes that are related with cellular membrane damage (Brown et al. 2002). In our study we observed that B deficiency altered the expression of genes related with lipid processing, namely lipid transport-related processes (that is partially represented by altered transport 2 days after B deficiency; Figure 5.1b) and lipid metabolic processing (that is partially represented in other metabolic processes 4 days after B deficiency; Figure 5.1b). Altered expression of genes related with K^+ transport was also observed 4 days after the imposition of B deficiency. Altered lipid biosynthesis and increased effluxes of K^+ due to B deficiency have already been reported (Belver et al. 1983, Schon et al. 1990, Cakmak et al. 1995). An

important plasma membrane boric channel, NIP5;1, previously described to be crucial for B import into cell roots under B limitation (Takano et al. 2006), was also observed to be up regulated 4 days after B deficiency (Supplementary Table 5.1).

Boron has also been associated with cytoskeleton biosynthesis, reproductive stage and organ morphogenesis, three events represented in the developmental processes (Figure 5.1b). In our study, we observed the up-regulation of *Actin3* (*ACT3*) and *Tubulin1* (*TUB1*) after 2 days of B deficiency. Yu et al. (2001) observed increased levels of cytoskeleton proteins and altered polymerization patterns in *A. thaliana* roots subjected to short-term B deficiency. An alteration in the expression of genes related with the reproductive stage and organ morphogenesis is in agreement with the referred higher requirement of B in plant reproductive structures (Loomis and Durst 1992, Rawson 1996).

We also detected changes into the expression of genes related with responses to stress (Figure 5.1b) in particular with oxidative stress, other stresses (e. g. cadmium stress) and defence responses. In a recent study, the major cause of cell death in B-deficient tobacco cells was associated to oxidative damage (Koshiba et al. 2009), and several defence-related responses have been reported in B-deficient plants (Kobayashi et al. 2004, Alves et al. 2006, Reguera et al. 2010). Additionally, the expression of genes related

with proteolysis and protein folding processes were also altered as a consequence of B deficiency as represented by altered protein metabolism in Figure 5.1b.

In the other metabolic processes (Figure 5.1b) we also observed the altered expression of several genes related with amino acid (mainly the branched-chain amino acids, methionine and threonine) and with glucosinolate metabolism. Free amino acids were suggested to regulate processes related to gene expression, not only on a global scale, but also in relation to translation of a particular group of proteins (Kimball and Jefferson 2006). Glucosinolate biosynthetic processing is modulated by either normal or stressful conditions, for example, wounding (Grubb and Abel 2006).

We detected several genes encoding transcription factors, which are included in the transcription class (Figure 5.1b), that were altered by B deficiency. Previously, Kasajima et al. (2010) reported that the *A. thaliana* transcription factor WRKY6, regulates several genes under B deficiency.

The increased number of differentially expressed genes from the 2nd to the 4th day of B deficiency (Figure 5.1 b) could reflect the transition from early or immediate responses to B deficiency (primary), to down-stream responses (secondary effects) which could include adaptive responses towards the management of B deficiency, but also non-specific responses to general stress.

Therefore, we have preferentially selected genes which were similarly differentially expressed after 2 and 4 days of B deficiency (Table 5.1).

Among the 37 genes selected (Supplementary Table 5.1), only the differential expression of one (At3g44450) was common to both B and Ca deficiencies, being down- and up-regulated, respectively. The remaining 36 genes were specifically responsive to B deficiency. Considering that the differential regulation of a chosen set of genes (13 genes) was validated by RT-qPCR analysis (Supplementary Table 5.2), the 37 genes selected are discussed in more detail below.

Cell wall biosynthesis

Several genes related with cell wall biosynthesis were differentially expressed under B deficiency. It has been reported that B is an essential element for the cell wall structure, through the cross-linking of rhamnogalacturonan molecules (Kobayashi et al. 1996, O'Neill et al. 1996, 2001). Polygalacturonans are abundant constituents of pectin and several enzymes are responsible for pectin modifications, including pectin methylesterases (Giovane et al. 2004). We observed the increased gene expression of a *pectin methylesterase*

Table 5.1. Functional classification of genes differentially expressed due to B deficiency. The thirty seven genes selected were expressed 2 and 4 days after B deficiency and were classified according to their biological functions.

| Probe Set | AGI ID ² | Curator Summary | Expr. | Biological Processes |
|--|---------------------|--|-------|---|
| Cell wall biosynthesis | | | | |
| 266693_at | At2g19800 | Myo-inositol oxygenase 2 (MIOX2) | Up | L-ascorbic acid biosynthesis; inositol catabolism; ox/red |
| 254001_at | At4g26260 | Myo-inositol oxygenase 4 (MIOX4) | Up | L-ascorbic acid biosynthesis; inositol catabolism; ox/red |
| 264898_at | At1g23205 | Pectin methylesterase inhibitor | Up | Pectinesterase inhibitor activity |
| 259736_at | At1g64390 | Glycosyl hydrolase 9C2 (ATGH9C2) | Down | Cell wall organization; cellulose catabolism |
| 258003_at | At3g29030 | Expansin (EXP5) | Down | Cell wall modification during multidimensional cell growth |
| 263477_at | At2g31790 | UDP-glucuronosyl/UDP-glucosyl transferase | Down | Metabolic process |
| 260955_at | At1g06000 | Flavonol-7-O-rhamnosyltransferase (7RhaT) | Down | Flavonol biosynthetic process |
| 264931_at | At1g60590 | Pectinase, putative | Down | Carbohydrate metabolic process |
| 253638_at | At4g30470 | Cinnamoyl-CoA reductase-related (CCR) | Down | Lignin biosynthetic process |
| Sulphur metabolism | | | | |
| 261957_at | At1g64660 | Methionine gamma-lyase (ATMGL) | Up | Methionine catabolic process via 2-oxobutanolate |
| 265121_at | At1g65860 | Flavin-containing monooxygenase 1 (FMO GS-OX1) | Down | Glucosinolate biosynthesis from homMet; Ox/Red |
| 261913_at | At1g62560 | Flavin-containing monooxygenase 3 (FMO GS-OX3) | Down | Glucosinolate biosynthesis from homMet; Ox/Red |
| 254343_at | At4g04610 | 5'-adenylylsulfate reductase (APR1) | Down | Sulfate assimilation and reduction; Cys biosynthesis; Ox/Red |
| 264745_at | At1g62180 | 5'-adenylylsulfate reductase 2 (APR2) | Down | Sulfate assimilation and reduction; Cys biosynthesis; Ox/Red |
| 255284_at | At4g21990 | 5'-adenylylsulfate reductase 3 (APR3) | Down | Sulfate assimilation and reduction; Cys biosynthesis; Ox/Red |
| 249752_at | At5g24660 | Response to low sulfur 2 (LSU2) | Down | Unknown |
| 260385_at | At5g07010 | Sulfotransferase (AIST2a) | Down | Jasmonic acid metabolic process, response to jasmonic acid stimulus |
| 250662_at | At1g74090 | Desulphoglucosinolate sulfotransferase (AIST5b) | Down | Glucosinolate biosynthetic process |
| Branched-chain amino acid (BCAA) catabolism | | | | |
| 264524_at | At1g10070 | Branched-chain amino acid transaminase 2 (BCAT2) | Up | BCAA biosynthesis/catabolism |
| 263118_at | At1g03090 | 3-methylcrotonyl-CoA carboxylase 1 (MCCAT1) | Up | Leucine catabolic process |

Table 5.1 (cont) Functional classification of genes differentially expressed due to B deficiency. The thirty seven genes selected were expressed 2 and 4 days after B deficiency and were classified according to their biological functions.

| Probe Set ¹ | AGI ID ² | Curator Summary | Expr. | Biological Processes |
|---|---------------------|--|-------|---|
| Transcriptional factors and hormones | | | | |
| 261395_at | At1g79700 | Ovule development protein (ODP), putative | Up | Regulation of transcription, DNA-dependent; transcription |
| 259982_at | At1g76410 | ATL1R | Up | Protein binding, zinc ion binding |
| 259230_at | At3g07780 | OBE1 | Up | Regulation of gene expression |
| 266015_at | At2g24190 | Short-chain dehydrogenase/reductase 1 (SDR1) | Up | Metabolic process |
| 263739_at | At2g21320 | DBB1a | Down | Regulation of transcription |
| 254231_at | At4g23810 | WRKY53 | Down | Regulation of transcription |
| 248353_at | At5g52320 | Cytochrome P450 (CYP96A4) | Down | Unknown |
| Miscellaneous | | | | |
| 254197_at | At4g24040 | Trehalase 1 (TRE1) | Up | Trehalose catabolic process |
| 261177_at | At1g04770 | Male sterility 5 (MS5) | Up | Unknown |
| 249527_at | At5g38710 | Proline oxidase, putative | Up | Glutamate biosynthetic process, proline catabolic process |
| 250832_at | At5g04950 | Nicotianamine synthase 1 (NAS1) | Down | Nicotianamine biosynthetic process |
| 249063_at | At5g44110 | ABC transporter AINAP2 | Down | Response to red light |
| Unknown | | | | |
| 264467_at | At1g10140 | Expressed protein | Up | Unknown |
| 247374_at | At5g63190 | MA3 domain-containing protein | Up | Unknown |
| 252478_at | At3g46540 | Epsin N-terminal homology (ENTH) domain-containing protein | Down | Unknown |
| 245524_at | At4g15920 | Expressed protein | Down | Unknown |
| 252661_at | At3g44450 | Expressed protein | Down | Unknown |

¹Affymetrix microarray ATH1

²From the TAIR9 (The Arabidopsis Information Resource) database (<http://www.arabidopsis.org>)

inhibitor (*PMEI*) and the decreased gene expression of a putative *pectinase*. Previous work (Camacho-Cristóbal et al. 2008) reported the decreased gene expression of two pectin methylesterases (*PME*) in B-deficient *Arabidopsis* roots, which suggests impaired pectin demethylation due to B deficiency.

The cellulose framework is thought to be embedded in the pectic polysaccharide matrix. We also detected decreased expression of genes involved in cellulose biosynthesis, namely the *glycosyl hydrolase 9C2* (*ATGH9CE*), involved in cellulose degradation and in an *expansin5* (*EXP5*). This data indicates that cell elongation was impaired or at least reduced due to B deficiency. Indeed, endoglycosidases and expansins are thought to be needed to modify the cell wall architecture in order to allow the incorporation of new material and consequent cell elongation (Gibeaut and Carpita 1994, Cosgrove et al. 2002).

Boron deficiency also induced several changes that interfere with the formation of cell wall precursors. *myo*-Inositol oxygenase (*MIOX*) is a key enzyme for the oxygenative cleavage of inositol into glucuronic acid, and has been associated with the control of carbohydrates flux to the cell wall (Kanter et al. 2005). From this multigene family, two (*MIOX2* and 4) were up-regulated due to B deficiency. This observation suggests an increase in the synthesis of glucuronic acid, the main precursor for important residues of plant cell wall polymers,

and an important substrate for nucleotide sugar inter-conversions (Kanter et al. 2005). The converted nucleotide sugars can then be used as substrates for glycosyltransferases, which are involved in the transfer of glycosyl residues to a wide range of acceptor molecules, and are responsible for regulating properties, such as bioactivity or solubility (Ross et al. 2001). Two genes, encoding a *glucosyltransferase* and a *rhamnosyltransferase* (*7RhaT*), were down-regulated due to B deficiency, reinforcing the observation that indicates perturbations in the cell wall metabolism. Furthermore, the gene expression of *trehalase 1* (*TRE1*), an enzyme responsible for trehalose degradation, was up-regulated under B deficiency. Trehalase is a plasma membrane-bound enzyme with its catalytic domain oriented towards the cell wall (Frison et al. 2007). The possible decreased content of trehalose, a recognized important sugar, could be related with carbohydrate partitioning and allocation (Müller et al. 2001). It seems that regulation of endogenous trehalose levels requires the transport of this sugar out of the cell (Frison et al. 2007). Thus, B deficiency, by altering the cell wall structure could also be interfering with trehalose metabolism and signalling in the plant.

Lignin is an important constituent of the secondary cell wall, and is composed of a complex network of aromatic polymers derived from the phenylpropanoid pathway. The *cinnamoyl-CoA reductase* (*CCR*)

gene that encodes a key enzyme for the formation of lignin monomers (Lacombe et al. 1997) was also down regulated due to B deficiency. A reduction in *CCR* gene expression was reported to decrease lignin biosynthesis (Piquemal et al. 1998), while the fraction of soluble phenols increases (van der Rest et al. 2006). Indeed, an increase in phenol content of B-deficient plants has been widely described (Brown et al. 2002).

Boron deficiency thus seems to affect cell wall biosynthesis at different levels (Figure 5.2). Probably, the damage of the pectin network compromises the stability of the embedded and adjacent matrixes. It seems that on one hand the plant is reducing cell wall related processes by decreasing the expression of the cell wall related genes, but on the other hand is trying to rescue the wall damage by increasing glucuronate biosynthesis.

Sulphur metabolism

Sulphur, in its reduced form, is involved in the biosynthesis of compounds of primary and secondary metabolism (Schmidt and Jäger 1992). The expression of an unknown gene, described as *Low sulphur up-regulated 2* gene (*LSU2*) (Lewandowska et al. 2010), was down-regulated due to B deficiency. Additionally, the three genes (*APR1*, 2 and 3) that encode 5'-adenylylsulfate reductases (APRs) were also down-regulated by B deficiency. APR

is encoded by a multigene family (Gutierrez-Marcos et al 1996) and plays a key role in the regulation of the sulphate assimilation pathway (Vauclare et al. 2002). Taken together, these results indicate that sulphur metabolism is being significantly affected by B

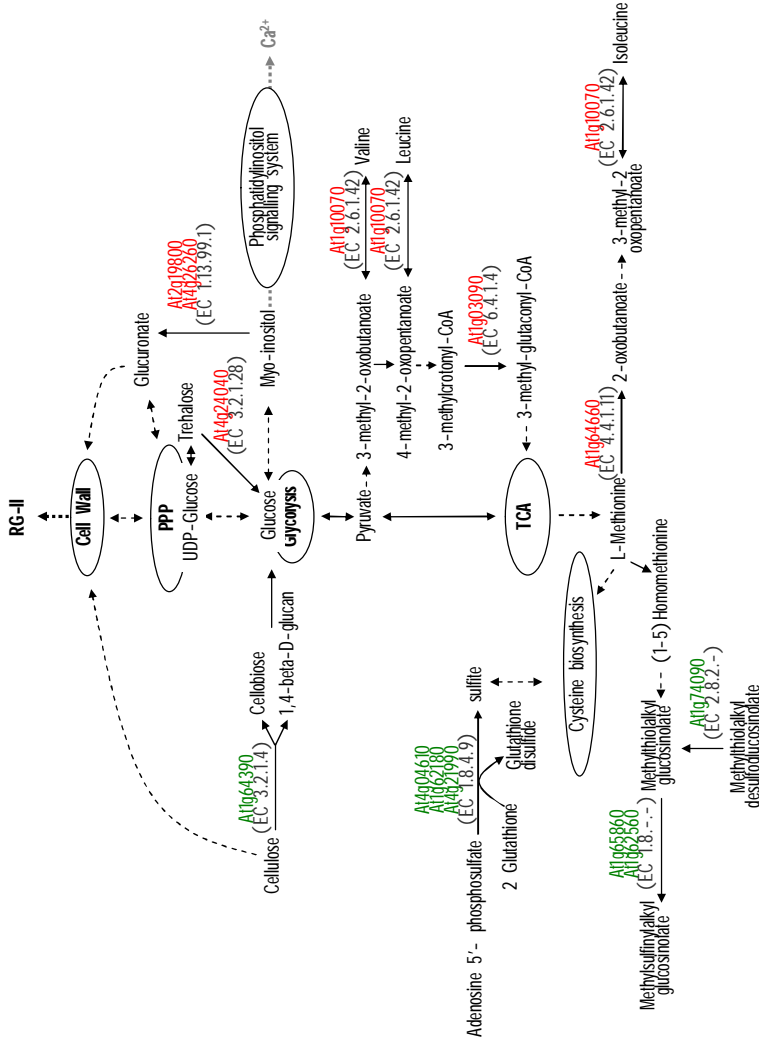


Figure 5.2. Metabolic overview of the pathways affected by B deficiency. Genes are marked either red (up regulated) or green (down regulated); see Table 5.1. For enzymes the respective EC number is indicated.

deficiency. Once inside the plant, sulphur can be fixed into cysteine and thereafter interconverted in methionine (Saito 2004). The catabolism of methionine is often related with homeostatic management of the excess of sulphur-containing amino acids. Thus, our observation of increased expression of the gene encoding *methionine gamma-lyase* (*ATMGL*) responsible for the degradation of methionine, can be directly related with the decrease in the sulphate assimilation pathway (Rennenberg 1984). It appears that B deficiency induces an effect similar to that of sulphur excess in plants. The expression of other genes related with sulphur metabolism was also decreased by B deficiency (Table 5.1), namely two flavin-containing monooxygenases and two sulfotransferases. These enzymes have been described as participants in sulphur-related steps in glucosinolate and jasmonate biosyntheses. The two *flavin-containing monooxygenases* (*FMO GS-OX1* and *3*) and a *desulphoglucosinolate sulfotransferase* (*AtST5b*) are required for glucosinolate biosynthesis; the FMO in the sulphur redox reactions (Grubb and Abel 2006), and the ST in the sulfation of desulphoglucosinolates, preferentially those derived from methionine (Piotrowski et al. 2004, Klein and Papenbrock 2006). Glucosinolates are sulphur-rich compounds that are differentially regulated at different developmental stages and conditions of stress, e. g. altered mineral nutrition (Halkier and Gershenzon 2006,

Grubb and Abel 2006). The *sulfotransferase AtST5b* is a close relative of the previous *AtST2a* that has been described as a hydroxyjasmonate sulfotransferase (Klein and Papenbrock 2006), proposed to regulate the levels of jasmonate and/or hydroxyjasmonate in plants (Gidda et al. 2003). Therefore, B may have an important impact on jasmonate biosynthesis.

Our work provides an indication that B deficiency affects several aspects of sulphur metabolism in plants (Figure 5.2). However, in our research of the literature, we only found a reference to decreased glutathione levels and increased gene expression of glutathione-S-transferase due to B deficiency (Lukaszewski and Blevins 1996, Kobayashi et al. 2004).

A link between sulphur and nitrogen metabolism has been known for many years, the deprivation of one leading to a disruption of the metabolism of the other (Koprivova et al. 2000). Our observation of the decreased expression of a nitrogen-related gene, *nicotianamine synthase (NS1)*, an enzyme reported to be induced by nitrate, (Wang et al. 2003) suggests a decrease in nitrogen metabolism. Previous studies on B deficiency also indicate a decrease in nitrogen metabolism (Camacho-Cristóbal and González-Fontes 1999, Beato et al. 2010).

Branched-chain amino acid catabolism

The branched-chain amino acid aminotransferase (BCAT) is responsible for the translocation of amino groups for both the biosynthesis and catabolism of the branched-chain amino acids (BCAAs), isoleucine, leucine and valine (Schuster and Binder 2005). The increased expression of *BCAT2*, a member of this gene family, and the repression of the *3-methylcrotonyl-CoA carboxylase 1 (MCCAT)*, which is described to participate in leucine catabolism (Anderson et al. 1998), may indicate that *BCAT2* is operating towards BCAAs degradation, in particular of leucine (Figure 5.2). In a recent study with *A. thaliana*, another member of the branched-chain amino acid aminotransferase, *BCAT4*, was also found to be up-regulated by B deficiency (Kasajima et al. 2010). In animals, the BCAA were described to stimulate muscle protein synthesis, leucine being the most effective with regard to the regulation of these proteins (Rhoads and Wu 2009). Since actin is an important muscle and also a cytoskeleton component (Hussey et al. 2002), changes in BCAAs content, in particular leucine, could interfere with cytoskeleton rearrangement. Under short-term B deficiency, increased levels of cytoskeletal proteins and altered polymerization patterns were already reported (Yu et al. 2001, 2003). Cytoskeleton altered biosynthesis is a hypothesis consistent with the knowledge that B plays a critical role in reproduction where

an actively changing cytoskeleton structure is required, as for instance during meiosis. We also found that the expression of a meiosis associated gene, the *male sterility 5 (MS5)*, important for male fertility (Glover et al. 1998) was decreased due to B deficiency.

Transcriptional factors and hormones

Regarding transcription related processes, we detected the differential expression of five transcription factors during B deficiency. A putative *ovule development protein (ODP)*, a RING-H2 finger protein, *ATL1R*, and a nuclear PHD finger, *OBE1*, were up-regulated due to B deficiency, while *DBB1a* and *WRKY53* were down regulated.

Three of these transcription factors, *ODP*, *OBE1* and *WRKY53*, have been associated to both biotic and abiotic stress and to hormonal responses. It has been reported that *ODP* and *OBE1* genes are up-regulated by cytokinins (Feng et al 2005) and auxins, respectively (Saiga et al. 2008), and that *WRKY53* is down-regulated by jasmonic acid (Thomas et al. 2009). Concerning this last, we also observed that B deficiency caused the down-regulation of another gene related with jasmonic acid biosynthesis, *CYP96A4*, which is involved in the metabolism of oxylipins (Benveniste et al. 2006). These observations suggest that B deficiency may have some involvement with auxins, cytokinins and jasmonate.

Another gene involved with hormone biosynthesis, *short-chain dehydrogenase/reductase 1 (SDR1)*, was up-regulated due to B deficiency. SDR1 participates in ABA biosynthesis (Lee et al. 2007) and was described to function as a molecular link between nutrient signalling and plant hormone biosynthesis (Cheng et al. 2002).

The literature contains little information concerning B and hormones. In relation to auxin, there are contradictory reports on the effect of B deficiency on auxin levels (Blevins and Lukaszewski 1998, Wang et al. 2006). Regarding cytokinins, it was shown by Wang and colleagues (2006) that their levels were decreased in the apices of B-deficient pea.

Of the other two transcription factors affected by B deficiency, a defect in the *DBB1a* protein was recently described to cause abnormal floral development in *Arabidopsis* (Wang et al. 2009). Thus, the decreased expression of *DBB1a* that we observed under B deficiency might be related with the described abnormal development of the plant reproductive structures caused by B deficiency (Brown et al. 2002). Finally, the transcription factor *ATL1R*, whose gene was up-regulated upon B deficiency, is related with RNA-binding proteins, thus being important for RNA processing (Wang et al. 2008).

Miscellaneous

We observed that the *proline oxidase* gene was induced by B deficiency. This gene encodes an enzyme involved in glutamate biosynthetic processes and proline catabolic processes. However, in *L. albus* we detected an increase in proline content during B deficiency (Chapter 4). We also observed that *AtNAP2*, which encodes an ABC transporter, was down-regulated by B deficiency. ABC transporters are responsible for the transport of a broad range of substances across membranes (Marin et al. 2006) and might therefore be involved in the plant adaptation to B deficiency.

CONCLUSIONS

A transcriptional analysis of *Arabidopsis thaliana* grown under B deficiency for 2 and 4 days was performed. Thirteen seven genes, which demonstrated similar differentially regulation after both 2 and 4 days of B deficiency were selected for discussion. In agreement with the well-documented importance of B in the cell wall structure, we found evidence for alteration in transcription of genes related with cell wall biosynthesis (9). In addition, genes related with transcriptional factors and hormones (7) were also seen to be affected by B deficiency. Interestingly, we find alterations in genes

related with sulphur metabolism (9), indicating an important, novel link between sulphur metabolism and B deficiency.

ACKNOWLEDGMENTS

We are grateful to Dr. Jörg Becker (Instituto Gulbenkian de Ciência, Oeiras, Portugal) for his help with the microarrays analyses, Dr. Ana Ribeiro and Dr. J. Cochicho Ramalho for the RT-qPCR machine loan (Instituto Investigação Científica Tropical, Oeiras, Portugal) and Dr. Sofia Pereira for the support with RT-qPCR data analysis. We also thank Phil Jackson for revising the manuscript. The financial support from the FCT (SFRH/BD/18273/2004) is also acknowledged.

REFERENCES

- Alves M, Francisco R, Martins I, Ricardo CPP (2006) Analysis of *Lupinus albus* leaf apoplastic proteins in response to boron deficiency. *Plant Soil* 279, 1–11.
- Anderson M, Che P, Song J, Nikolau B, Wurtele E (1998) 3-Methylcrotonyl-Coenzyme A Carboxylase Is a Component of the Mitochondrial Leucine Catabolic Pathway in Plants. *Plant Physiol* 118: 1127–1138.
- Amon D (1938) Microelements in culture–solution experiment with higher plants. *Amer J Bot* 25: 322–325.

- Beato V, Rexach J, Navarro-Gochicoa M, Camacho-Cristóbal J, Herrera-Rodríguez M, Maldonado J, González-Fontes J (2010) A tobacco asparagine synthetase gene responds to carbon and nitrogen status and its root expression is affected under boron stress. *Plant Sci* 178: 289-298.
- Belver A, Donaire J (1983) Partial purification of soluble lipoxygenase of sunflower cotyledons: action of boron on the enzyme and lipid constituents. *Z Pflanzenphysiol* 109: 309-317.
- Benveniste I, Saito T, Wang Y, Kandel S, Huang H, Pinot F, Kahn R, Salaun J, Shimoji M (2006) Evolutionary relationship and substrate specificity of *Arabidopsis thaliana* fatty acid omegahydroxylase. *Plant Sci* 170: 326-338.
- Blevins D, Lukaszewski K (1998) Boron in plant structure and function. *Annu Rev Plant Physiol Plant Mol Biol* 49: 481-500.
- Bolaños L, Lukaszewski K, Bonilla I, Blevins D (2004) Why boron? *Plant Physiol Biochem* 42: 907-912.
- Brown P, Bellaloui N, Wimmer M, Bassil E, Ruiz J, Hu H, Pfeiffer H, Dannel F, Romheld V (2002) Boron in plant biology. *Plant Biol* 4: 203-223.
- Cakmak I, Kurz H, Marschner H (1995) Short-term effects of boron, germanium and high light intensity on membrane permeability in boron deficient sunflower leaves. *Physiol Plant* 95: 11-18.
- Camacho-Cristóbal J, Anzellotti D, González-Fontes A (2002) Changes in phenolic metabolism of tobacco plants during short-term boron deficiency. *Plant Physiol Biochem* 12: 997-1002.

- Camacho-Cristóbal J, González-Fontes A (1999) Boron deficiency causes a drastic decrease in nitrate content and nitrate reductase activity, and increases the content of carbohydrates in leaves from tobacco plants. *Planta* 209: 528-536.
- Camacho-Cristóbal J, Herrera-Rodríguez M, Beato V, Rexach J, Navarro-Gochicoa M, Maldonado J, González-Fontes A (2008) The expression of several cell wall-related genes in *Arabidopsis* roots is down-regulated under boron deficiency. *Env Exp Bot* 63: 351-358.
- Cheng W, Endo A, Zhou L, Penney J, Chen H, Arroyo A, Leon P, Nambara E, Asami T, Seo M, Koshiba T, Sheen J (2002) A unique short-chain dehydrogenase/reductase in *Arabidopsis* glucose signaling and abscisic acid biosynthesis and functions. *Plant Cell* 14: 2723-2743.
- Cosgrove D, Li L, Cho H-T, Hoffmann-Benning S, Moore R, Blecker D (2002) The Growing World of Expansins. *Plant Cell Physiol* 43: 1436-1444.
- Czechowski T, Stitt M, Altmann T, Udvardi M, Scheible W-R (2005) Genome-Wide Identification and Testing of Superior Reference Genes for Transcript Normalization in *Arabidopsis*. *Plant Physiol* 139: 5-17.
- Feng J, Liu D, Pan Y, Gong W, Ma L, Luo J, Deng X, Zhu Y (2005) An annotation update via cDNA sequence analysis and comprehensive profiling of developmental, hormonal or environmental responsiveness of the *Arabidopsis* AP2/EREBP transcription factor gene family. *Plant Mol Biol* 59: 853-868.
- Frison M, Parrou J, Guillaumot D, Masquelier D, François J, Chaumont F, Batoko H (2007) The *Arabidopsis thaliana* trehalase is a plasma membrane-bound enzyme with extracellular activity. *FEBS Lett* 581: 4010-4016.

- Gibeaut D, Carpita N (1994) Biosynthesis of plant cell wall polysaccharides. *FASEB J* 8: 904–915.
- Gidda S, Miersch O, Levitin A, Schmidt J, Wasternack C, Varin L (2003) Biochemical and Molecular Characterization of a Hydroxyjasmonate Sulfotransferase from *Arabidopsis thaliana*. *JBC* 278: 17895–17900.
- Giovane A, Servillo L, Balestrieri C, Raiola A, D'Avino R, Tamburrini M, Ciardiello M, Camardella L (2004) Pectin methylesterase inhibitor. *BBA* 1696: 245–252.
- Glover J, Grelon M, Craig S, Chaudhury A, Dennis E (1998) Cloning and characterization of MS5 from *Arabidopsis*: a gene critical in male meiosis. *Plant J* 15: 345–356.
- Grubb C, Abel S (2006) Glucosinolate metabolism and its control. *Trends Plant Sci* 11: 89–100.
- Gutierrez-Marcos J, Roberts M, Campbell E, Wray J (1996) Three members of a novel small gene-family from *Arabidopsis thaliana* able to complement functionally an *Escherichia coli* mutant defective in PAPS reductase activity encode proteins with a thioredoxin-like domain and "APS reductase" activity. *PNAS* 93: 13377–13382.
- Halkier B, Gershenzon J (2006) Biology and biochemistry of glucosinolates. *Annu Rev Plant Biol* 57: 303–333.
- Hepler P (2005) Calcium: A Central Regulator of Plant Growth and Development. *Plant Cell* 17: 2142–2155.
- Hussey P, Allwood E, Smertenko A (2002) Actin-binding proteins in the *Arabidopsis* genome database: properties of functionally distinct plant actin-depolymerizing factors/cofilins. *Phil Trans R Soc Lond B* 357: 791–798.

- Kanter U, Usadel B, Guerineau F, Li Y, Pauly M, Tenhaken R (2005) The inositol oxygenase gene family of *Arabidopsis* is involved in the biosynthesis of nucleotide sugar precursors for cell-wall matrix polysaccharides. *Planta* 221: 243–254.
- Kasajima I, Ide Y, Hirai M, Fujiwara T (2010) WRKY6 is involved in the response to boron deficiency in *Arabidopsis thaliana*. *Physiol Plant* (*in press*).
- Kimball S, Jefferson L (2006) New functions for amino acids: effects on gene transcription and translation. *Am J Clin Nutr* 83: 500–507.
- Klein M, Papenbrock J (2004) The multi-protein family of *Arabidopsis* sulphotransferases and their relatives in other plant species. *J Exp Bot* 55: 1809–1820.
- Kobayashi M, Matoh T, Azuma J (1996) Two chains of rhamnogalacturonan II are cross-linked by borate-diol ester bonds in higher plant cell walls, *Plant Physiol* 110: 1017–1020.
- Kobayashi M, Mutoh T, Matoh T (2004) Boron nutrition of cultured tobacco BY-2 cells. IV. Genes induced under low boron supply. *J. Exp. Bot.* 55: 1441–1443.
- Kobayashi M, Nakagawa H, Asaka T, Matoh T (1999) Borate-Rhamnogalacturonan II Bonding Reinforced by Ca^{2+} Retains Pectic Polysaccharides in Higher-Plant Cell Walls. *Plant Physiol* 119: 199–203.
- Koprivova A, Suter M, Op den Camp R, Brunold C, Kopriva S (2000) Regulation of sulfate assimilation by nitrogen in *Arabidopsis*. *Plant Physiology* 122, 737–746.
- Koshiba T, Kobayashi M, Ishihara A, Matoh T (2010) Boron Nutrition of Cultured Tobacco BY-2 Cells. VI. Calcium is

Involved in Early Responses to Boron Deprivation. *Plant Cell Physiol* 51: 323-327.

Koshiba T, Kobayashi M, Matoh T (2009) Boron Nutrition of Tobacco BY-2 Cells. V. Oxidative Damage is the Major Cause of Cell Death Induced by Boron Deprivation. *Plant Cell Physiol* 50: 26-36.

Lacombe E, Hawkins S, Van Doorselaere J, Piquemal J, Goffner D, Poeydomenge O, Boudet AM, Grima-Pettenati J (1997) Cinnamoyl CoA reductase, the first committed enzyme of the lignin branch biosynthetic pathway: cloning, expression and phylogenetic relationships. *Plant J* 11: 429-441.

Lee J, Woodward A, Chen Z (2007) Gene Expression Changes and Early Events in Cotton Fibre Development. *Ann Bot* 100: 1391-1401.

Lewandowska M, Wawrzynska A, Moniuszko G, Lukomska J, Zientara K, Piecho M, Hodureka P, Zhukov I, Liszewska F, Nikiforova V, Sirko A (2010) A Contribution to Identification of Novel Regulators of Plant Response to Sulfur Deficiency: Characteristics of a Tobacco Gene UP9C, Its Protein Product and the Effects of UP9C Silencing. *Mol Plant* 3: 347-360.

Li C, Wong W (2001a) Model-based analysis of oligonucleotide arrays: model validation, design issues and standard error application. *Genome Biol* 2: RESEARCH0032.

Li C, Wong W (2001b) Model-based analysis of oligonucleotide arrays: expression index computation and outlier detection. *PNAS* 98: 31-36.

Livak K, Schmittgen T (2001) Analysis of relative gene expression data using real-time quantitative PCR and the $2^{-\Delta\Delta C(T)}$ Method. *Methods* 25: 402-408.

- Loomis W, Durst R (1992) Chemistry and biology of boron. *Biofactors* 3: 229-239.
- Lukaszewski K, Blevins D (1996) Root Growth Inhibition in Boron-Deficient or Aluminum-Stressed Squash May Be a Result of Impaired Ascorbate Metabolism. *Plant Physiol* 112: 1135-1140.
- Marin E, Divol F, Bechtold N, Vavasseur A, Nussaume L, Forestier C (2006) Molecular characterization of three *Arabidopsis* soluble ABC proteins which expression is induced by sugars. *Plant Sci* 171: 84-90.
- Mosblech A, Ivo Feussner, Ingo Heilmann (2009) Oxylipins: Structurally diverse metabolites from fatty acid oxidation. *Plant Physiol Biochem* 47: 511-517.
- Müller J, Aeschbacher R, Wingler A, Boller T, Wiemken A (2001) Trehalose and Trehalase in *Arabidopsis*. *Plant Physiol* 125: 1086-1093.
- O'Neill M, Eberhard S, Albersheim P, Darvill A (2001) Requirement of borate cross-linking of cell wall rhamnogalacturonan II for *Arabidopsis* growth, *Science* 294: 846-849.
- O'Neill M, Warrenfeltz D, Kates K, Pellerin P, Doco T, Darvill A, Albersheim P (1996) Rhamnogalacturonan-II, a pectic polysaccharide in the walls of growing plant cell, forms a dimer that is covalently cross linked by a borate ester. *J Biol Chem* 271: 22923-22930.
- Piotrowski M, Schemenewitz A, Lopukhina A, Müller A, Janowitz T, Weiler E, Oecking C (2004) Desulfoglucosinolate Sulfotransferases from *Arabidopsis thaliana* Catalyze the Final Step in the Biosynthesis of the Glucosinolate Core Structure. *JBC* 279: 50717-50725.

- Piquemal J, Lapierre C, Myton K, O'Connell A, Schuch W, Grima-Pettenati J, Boudet A (1998) Down regulation of cinnamoylCoA reductase induces significant changes of lignin profiles in transgenic tobacco plants. *Plant J* 13: 71–83.
- Rawson H (1996) The developmental stage during which boron limitation causes sterility in wheat genotypes and the recovery of fertility. *Aust J Plant Physiol* 23: 709–717.
- Reddy V, Reddy A (2004) Proteomics of calcium-signaling components in plants. *Phytochem* 65: 1745–1776.
- Redondo-Nieto M, Wilmot A, El-Hamdaoui A, Bonilla I, Bolaños L (2003) Relationship between boron and calcium in the N₂-fixing legume-rhizobia symbiosis. *Plant Cell Environ* 26: 1905–1915.
- Reguera M, Bonilla I, Bolaños L (2010) Boron deficiency results in induction of pathogenesis-related proteins from the PR-10 family during the legume-rhizobia interaction. *J Plant Physiol* (*in press*).
- Rennenberg H (1984) The fate of excess sulfur in higher plants. *Ann. Rev. Plant Physiol.* 35: 121–153.
- Rhoads J, Wu G (2009) Glutamine, arginine and leucine signaling in the intestine. *Amino Acids* 37: 111–122.
- Ross J, Li Y, Lim E, Bowles D (2001) Higher plant glycosyltransferases. *Genome Biol* 2: 3004.1–3004.6.
- Saiga S, Furumizu C, Yokoyama R, Kurata T, Sato S, Kato T, Tabata S, Suzuki M, Komeda Y (2008) The *Arabidopsis* OBERON1 and OBERON2 genes encode plant homeodomain finger proteins and are required for apical meristem maintenance. *Development* 135:1751–1759.

- Saito K (2004) Sulfur assimilatory metabolism. The long and smelling road. *Plant Physiol* 136: 2443–2450.
- Schmidt A, Jäger K (1992) Open questions about sulfur metabolism in plants. *Plant Mol Biol* 43: 325–349.
- Schon M, Novacky A, Blevins D (1990) Boron Induces Hyperpolarization of Sunflower Root Cell Membranes and Increases Membrane Permeability to K. *Plant Physiol* 93: 566–571.
- Schuster S, Binder S (2005) The mitochondrial branched-chain aminotransferase (AtBACT-1) is capable of initiate degradation of leucine, isoleucine and valine in almost all tissues in *Arabidopsis thaliana*. *Plant Mol Biol* 57: 241–254.
- Sungur S, Okur R (2009) Using Azomethine-H method determination of boron contents of various foods consumed in Hatay Region in Turkey. *Food Chem* 115: 711–714.
- Takano J, Wada M, Ludewig U, Schaaf G, von Wirén N, Fujiwara T (2006). The *Arabidopsis* major intrinsic protein NIP5;1 is essential for efficient boron uptake and plant development under boron limitation. *Plant Cell* 18, 1498–1509.
- Thomas C, Schmidt D, Bayer E, Dreos R, Maule A (2009) *Arabidopsis* plant homeodomain finger proteins operate downstream of auxin accumulation in specifying the vasculature and primary root meristem. *Plant J* 59: 426–436.
- van der Rest B, Danoun S, Boudet A-M, Rochange S (2006) Down-regulation of cinnamoyl-CoA reductase in tomato (*Solanum lycopersicum* L.) induces dramatic changes in soluble phenolic pools. *J Exp Bot*, 57: 1399–1411.
- Vauclare P, Kopriva S, Fell D, Suter M, Sticher L, von Ballmoos O, Krähenbuhl U, Op den Camp R, Brunold C (2002) Flux

- control of sulphate assimilation in *Arabidopsis thaliana*: adenosine 5'-phosphosulphate reductase is more susceptible to negative control by thiols than ATP sulphurylase. *Plant J* 31: 729-740.
- Wang D, Guo Y, Wu C, Yang G, Li Y, Zheng C (2008) Genome-wide analysis of CCCH zinc finger family in *Arabidopsis* and rice. *BMC Genomics* 9: 44.
- Wang G, Römheld V, Li C, Bangerth F (2006) Involvement of auxin and CKs in boron deficiency induced changes in apical dominance of pea plants (*Pisum sativum* L.) *J Plant Physiol* 163: 591-600.
- Wang Q, Tu X, Deng K, Zeng J, Zhao X, Tang D, Liu X (2009) A Defect in Zinc Finger Protein Double B-box 1a (DBB1a) Causes Abnormal Floral Development in *Arabidopsis*. *J Plant Biol* 52: 543-549.
- Wang R, Okamoto M, Xing X, Crawford N (2003) Microarray Analysis of the Nitrate Response in *Arabidopsis* Roots and Shoots Reveals over 1,000 Rapidly Responding Genes and New Linkages to Glucose, Trehalose-6-Phosphate, Iron, and Sulfate Metabolism. *Plant Physiol* 132: 556-567.
- Warington K (1923) The effect of boric acid and borax on the broad bean and certain other plants. *Ann Bot* 37: 629-672.
- Yu Q, Baluška F, Jasper F, Menzel D, Goldbach H (2003) Short-term boron deprivation enhances levels of cytoskeletal proteins in maize, but not zucchini, root apices. *Physiol Plant* 117: 270-278.
- Yu Q, Hlavacka A, Matoh T, Volkmann D, Menzel D, Goldbach H, Baluska F (2002) Short-term boron deprivation inhibits endocytosis of cell wall pectins in meristematic cells of maize and wheat root apices. *Plant Physiol* 130: 415-421.

- Yu Q, Wingender R, Schulz M, Baluska F, Goldbach H (2001) Short-term boron deprivation induces increased levels of cytoskeletal proteins in *Arabidopsis* roots. *Plant Biol* 3: 335–340.
- Zhong S, Li C, Wong W (2003) ChiplInfo: Software for extracting gene annotation and gene ontology information for microarray analysis. *Nucleic Acids Res* 31: 3483–3486.

SUPPLEMENTARY DATA

Supplementary Table 5.1. Genes differentially expressed 2 days (102) and 4 days (208) after B suppression and genes differentially expressed 2 days (2,975) and 4 days (270) after Ca suppression, organized according to the fold change (FC) observed in the Affymetrix ATH1 geneChip array.

| Probe Set ¹ | AGI ID ² | Curatory Summary | FC |
|-----------------------------------|---------------------|--|-------|
| <i>2 days after B suppression</i> | | | |
| 252269_at | At3g49580 | expressed protein | 15.64 |
| 259842_at | At1g73600 | phosphoethanolamine N-methyltransferase 3, putative (NMT3) | 7 |
| 248844_s_at | At5g46900 | protease inhibitor/seed storage/lipid transfer protein | 6.81 |
| 259813_at | At1g49860 | glutathione S-transferase, putative | 6.17 |
| 255689_at | At4g00670 | remorin | 5.75 |
| 261684_at | At1g47400 | expressed protein | 5.72 |
| 249752_at | At5g24660 | expressed protein | 5.38 |
| 249867_at | At5g23020 | 2-isopropylmalate synthase 2 (IMS2) | 5.02 |
| 260745_at | At1g78370 | glutathione S-transferase, putative | 4.85 |
| 250661_at | At5g07030 | aspartyl protease | 4.76 |
| 248139_at | At5g54970 | expressed protein | 4.62 |
| 266395_at | At2g43100 | aconitase C-terminal domain-containing protein | 4.44 |
| 262978_at | At1g75780 | tubulin beta-1 chain (TUB1) | 4.26 |
| 249836_at | At5g23420 | high mobility group (HMG1/2) | 4.22 |
| 249167_at | At5g42860 | expressed protein | 4.22 |
| 256125_at | At1g18250 | thaumatin, putative | 4.16 |
| 265121_at | At1g62560 | flavin-containing monooxygenase | 3.98 |
| 261913_at | At1g65860 | flavin-containing monooxygenase | 3.93 |
| 258675_at | At3g08770 | lipid transfer protein 6 (LTP6) | 3.89 |
| 262717_s_at | At1g16410 | cytochrome P450 | 3.73 |
| 255517_at | At4g02290 | glycosyl hydrolase family 9 protein | 3.67 |
| 251524_at | At3g58990 | aconitase C-terminal domain-containing protein | 3.66 |
| 250892_at | At5g03760 | glycosyl transferase family 2 protein | 3.64 |
| 264262_at | At1g09200 | histone H3 | 3.63 |
| 254789_at | At4g12880 | plastocyanin-like domain-containing protein | 3.49 |
| 263941_at | At2g35870 | At2g35870 | 3.48 |
| 258851_at | At3g03190 | glutathione S-transferase, putative | 3.43 |
| 258470_at | At3g06035 | expressed protein | 3.32 |
| 263431_at | At2g22170 | lipid-associated | 3.3 |
| 247651_at | At5g59870 | histone H2A, putative | 3.28 |
| 247268_at | At5g64080 | protease inhibitor/seed storage/lipid transfer protein | 3.28 |
| 252442_at | At3g46940 | deoxyuridine 5'-triphosphate nucleotidohydrolase | 3.25 |
| 259681_at | At1g77760 | nitrate reductase 1 (NR1) | 3.24 |
| 253040_at | At4g37800 | xyloglucan:xyloglucosyl transferase, putative | 3.16 |
| 250434_at | At5g10390 | histone H3 | 3.15 |
| 254862_at | At4g12030 | bile acid:sodium symporter | 3.14 |
| 255874_at | At2g40550 | expressed protein | 3.13 |
| 261330_at | At1g44900 | DNA replication licensing factor, putative | 3.07 |

| | | | |
|-------------|-----------|--|-------|
| 245196_at | At1g67750 | pectate lyase | 3.07 |
| 267590_at | At2g39700 | expansin, putative (EXP4) | 3.06 |
| 245343_at | At4g15830 | expressed protein | 3.05 |
| 247192_at | At5g65360 | histone H3 | 3.04 |
| 266223_at | At2g28790 | osmotin-like protein, putative | 3.03 |
| 263628_at | At2g04780 | fasciclin-like arabinogalactan-protein (FLA7) | 3.03 |
| 257021_at | At3g19710 | branched-chain amino acid transaminase (BCAT4) | 2.99 |
| 261080_at | At1g07370 | proliferating cell nuclear antigen 1 (PCNA1) | 2.97 |
| 249916_at | At5g22880 | histone H2B, putative | 2.94 |
| 252911_at | At4g39510 | cytochrome P450 | 2.92 |
| 258859_at | At3g02120 | hydroxyproline-rich glycoprotein | 2.88 |
| 262232_at | At1g68600 | expressed protein | 2.88 |
| 248891_at | At5g46280 | DNA replication licensing factor, putative | 2.8 |
| 258480_at | At3g02640 | expressed protein | 2.79 |
| 260902_at | At1g21440 | mutase | 2.76 |
| 263882_at | At2g21790 | ribonucleotide reductase, putative | 2.76 |
| 247812_at | At5g58390 | peroxidase, putative | 2.74 |
| 262109_at | At1g02730 | cellulose synthase | 2.72 |
| 256237_at | At3g12610 | DNA-damage-repair/toleration protein, putative (DRT100) | 2.71 |
| 265656_at | At2g13820 | protease inhibitor/seed storage/lipid transfer protein | 2.71 |
| 261921_at | At1g65900 | expressed protein | 2.67 |
| 252148_at | At3g51280 | male sterility MS5, putative | 2.64 |
| 263535_at | At2g24970 | expressed protein | 2.63 |
| 254687_at | At4g13770 | cytochrome P450 | 2.61 |
| 261309_at | At1g48600 | phosphoethanolamine N-methyltransferase 2, putative (NMT2) | 2.56 |
| 253340_s_at | At4g33260 | WD-40 repeat | 2.56 |
| 254119_at | At4g24780 | pectate lyase | 2.56 |
| 264377_at | At2g25060 | plastocyanin-like domain-containing protein | 2.55 |
| 267175_s_at | At2g37620 | actin 3 (ACT3) | 2.55 |
| 249866_at | At5g23010 | 2-isopropylmalate synthase 3 (IMS3) | 2.47 |
| 264061_at | At2g27970 | cyclin-dependent kinase, putative | 2.38 |
| 246962_s_at | At5g24800 | bZIP transcription factor | -2.71 |
| 266267_at | At2g29460 | glutathione S-transferase, putative | -2.86 |
| 266590_at | At2g46240 | IQ domain-containing protein | -2.96 |
| 260015_at | At1g67980 | caffeoyl-CoA 3-O-methyltransferase, putative | -2.98 |
| 247095_at | At5g66400 | dehydrin (RAB18) | -3 |
| 259982_at | At1g76410 | zinc finger (C3HC4-type RING finger) | -3.08 |
| 250580_at | At5g07440 | glutamate dehydrogenase 2 (GDH2) | -3.08 |
| 260662_at | At1g19540 | isoflavone reductase, putative | -3.13 |
| 266799_at | At2g22860 | phytosulfokines 2 (PSK2) | -3.18 |
| 260248_at | At1g74310 | heat shock protein 101 (HSP101) | -3.2 |
| 266841_at | At2g26150 | heat shock transcription factor | -3.25 |
| 266693_at | At2g19800 | expressed protein | -3.49 |
| 264280_at | At1g61820 | glycosyl hydrolase family 1 protein | -3.54 |

| | | | |
|-------------|-----------|---|--------|
| 256965_at | At3g13450 | branched-chain alpha-keto acid dehydrogenase E1 beta subunit (DIN4) | -3.61 |
| 266294_at | At2g29500 | 17.6 kDa class I small heat shock protein (HSP17.6B-CI) | -3.69 |
| 263118_at | At1g03090 | 3-methylcrotonyl-CoA carboxylase 1 (MCCA) | -3.78 |
| 260900_s_at | At1g21400 | 2-oxoisovalerate dehydrogenase, putative | -3.8 |
| 252570_at | At3g45300 | isovaleryl-CoA-dehydrogenase (IVD) | -3.82 |
| 261395_at | At1g79700 | ovule development protein, putative | -3.83 |
| 251755_at | At3g55790 | expressed protein | -4.13 |
| 252515_at | At3g46230 | 17.4 kDa class I heat shock protein (HSP17.4-CI) | -4.57 |
| 262047_at | At1g80160 | lactoylglutathione lyase / glyoxalase I | -4.82 |
| 252415_at | At3g47340 | asparagine synthetase 1 (ASN1) | -4.92 |
| 264524_at | At1g10070 | branched-chain amino acid transaminase 2 (BCAT2) | -4.95 |
| 260668_at | At1g19530 | expressed protein | -5.03 |
| 260741_at | At1g15045 | glutamine amidotransferase-related | -5.45 |
| 262307_at | At1g71000 | DNAJ heat shock N-terminal domain-containing protein | -5.59 |
| 261957_at | At1g64660 | Cys/Met metabolism pyridoxal-phosphate-dependent enzyme | -6.67 |
| 253829_at | At4g28040 | nodulin MIN21 | -6.88 |
| 264777_at | At1g08630 | L-allo-threonine aldolase-related | -7.2 |
| 250351_at | At5g12030 | II heat shock protein 17.6A (HSP17.7-CII) | -7.98 |
| 264514_at | At1g09500 | cinnamyl-alcohol dehydrogenase | -8.54 |
| 254001_at | At4g26260 | expressed protein | -9.29 |
| 262325_at | At1g64160 | disease resistance-responsive / dirigent | -12.87 |

4 days after B suppression

| | | | |
|-----------|-----------|---|------|
| 261177_at | At1g04770 | male sterility MS5 | 3.13 |
| 265665_at | At2g27420 | cysteine proteinase, putative | 3.09 |
| 249752_at | At5g24660 | expressed protein | 2.7 |
| 255604_at | At4g01080 | expressed protein | 2.6 |
| 250533_at | At5g08640 | flavonol synthase 1 (FLS1) | 2.43 |
| 258133_at | At3g24500 | ethylene-responsive transcriptional coactivator, putative | 2.32 |
| 256999_at | At3g14200 | DNAJ heat shock N-terminal domain-containing protein | 2.21 |
| 248676_at | At5g48850 | male sterility MS5 | 2.2 |
| 247447_at | At5g62730 | proton-dependent oligopeptide transport (POT) | 2.14 |
| 260248_at | At1g74310 | heat shock protein 101 (HSP101) | 2.11 |
| 264898_at | At1g23205 | invertase/pectin methylesterase inhibitor | 2.11 |
| 254343_at | At4g21990 | 5'-adenylylsulfate reductase (APR3) | 2.05 |
| 255284_at | At4g04610 | 5'-adenylylsulfate reductase (APR1) | 2.03 |
| 264931_at | At1g60590 | polygalacturonase, putative / pectinase, putative | 2 |
| 261907_at | At1g65060 | 4-coumaroyl-CoA synthase 3 (4CL3) | 1.97 |
| 248607_at | At5g49480 | sodium-responsive calcium-binding protein (ACP1) | 1.94 |
| 264331_at | At1g04130 | tetratricopeptide repeat (TPR)-containing protein | 1.9 |
| 263374_at | At2g20560 | DNAJ heat shock | 1.88 |
| 248448_at | At5g51190 | AP2 domain-containing transcription factor, putative | 1.87 |
| 249837_at | At5g23480 | expressed protein | 1.86 |
| 261953_at | At1g64440 | UDP-glucose 4-epimerase, putative | 1.85 |
| 258560_at | At3g06020 | expressed protein | 1.84 |

| | | | |
|-------------|-----------|--|------|
| 260955_at | At1g06000 | UDP-glucuronosyl/UDP-glucosyl transferase | 1.83 |
| 266841_at | At2g26150 | heat shock transcription factor | 1.82 |
| 255511_at | At4g02075 | zinc finger (C3HC4-type RING finger) | 1.82 |
| 252661_at | At3g44450 | expressed protein | 1.82 |
| 265892_at | At2g15020 | expressed protein | 1.81 |
| 256518_at | At1g66080 | expressed protein | 1.81 |
| 264745_at | At1g62180 | 5'-adenylylsulfate reductase 2, chloroplast (APR2) | 1.81 |
| 263739_at | At2g21320 | zinc finger (B-box type) | 1.81 |
| 265121_at | At1g62560 | flavin-containing monooxygenase | 1.81 |
| 261913_at | At1g65860 | flavin-containing monooxygenase | 1.78 |
| 245777_at | At1g73540 | MutT/nudix | 1.77 |
| 267053_s_at | At2g38390 | peroxidase 22 (PER22) | 1.75 |
| 248190_at | At5g54120 | calcium-binding EF hand | 1.74 |
| 253382_at | At4g33040 | glutaredoxin | 1.74 |
| 248191_at | At5g54130 | calcium-binding EF hand | 1.74 |
| 252958_at | At4g38620 | myb family transcription factor (MYB4) | 1.74 |
| 247877_at | At5g57740 | zinc finger (C3HC4-type RING finger) | 1.7 |
| 249774_at | At5g24150 | squalene epoxidase 1,1 (SQP1,1) | 1.7 |
| 252363_at | At3g48460 | GDSL-motif lipase/hydrolase | 1.69 |
| 266720_s_at | At2g46790 | timing of CAB expression 1-like protein, putative | 1.68 |
| 247543_at | At5g61600 | ethylene-responsive element-binding | 1.67 |
| 245025_at | atpF | atpF | 1.67 |
| 263477_at | At2g31790 | UDP-glucuronosyl/UDP-glucosyl transferase | 1.66 |
| 258830_at | At3g07090 | expressed protein | 1.66 |
| 259432_at | At1g01520 | myb family transcription factor | 1.65 |
| 266015_at | At2g24190 | short-chain dehydrogenase/reductase (SDR) | 1.65 |
| 254231_at | At4g23810 | WRKY family transcription factor | 1.65 |
| 257421_at | At1g12030 | expressed protein | 1.65 |
| 266278_at | At2g29300 | tropinone reductase, putative | 1.65 |
| 244934_at | ndhG | ndhG | 1.64 |
| 259580_at | At1g28030 | oxidoreductase, 2OG-Fe(II) oxygenase | 1.63 |
| 258505_at | At3g06530 | BAP28-related | 1.62 |
| 258979_at | At3g09440 | heat shock cognate 70kDa protein 3 (HSC70-3) | 1.62 |
| 261597_at | At1g49780 | U-box domain-containing protein | 1.62 |
| 255622_at | At4g01070 | UDP-glucuronosyl/UDP-glucosyl transferase | 1.62 |
| 258003_at | At3g29030 | expansin, putative (EXP5) | 1.62 |
| 263150_at | At1g54050 | heat shock protein (HSP17.4-CIII) | 1.61 |
| 263122_at | At1g78510 | solaneyl diphosphate synthase (SPS) | 1.6 |
| 259364_at | At1g13260 | DNA-binding protein RAV1 | 1.6 |
| 266219_at | At2g28880 | para-aminobenzoate (PABA) synthase | 1.6 |
| 249288_at | At5g41050 | expressed protein | 1.6 |
| 248353_at | At5g52320 | cytochrome P450, putative | 1.59 |
| 262416_at | At1g49390 | oxidoreductase, 2OG-Fe(II) oxygenase | 1.58 |
| 266299_at | At2g29450 | glutathione S-transferase (103-1A) | 1.57 |
| 247323_at | At5g64170 | dentin sialophosphoprotein-related | 1.57 |

| | | | |
|-------------|-----------|---|------|
| 251744_at | At3g56010 | expressed protein | 1.55 |
| 262656_at | At1g14200 | zinc finger (C3HC4-type RING finger) | 1.55 |
| 248964_at | At5g45340 | cytochrome P450 | 1.55 |
| 245524_at | At4g15920 | nodulin MIN3 | 1.55 |
| 254878_at | At4g11660 | heat shock transcription factor 7 (HSTF7) | 1.54 |
| 250315_at | At5g12130 | integral membrane TerC | 1.54 |
| 258687_at | At3g07860 | expressed protein | 1.54 |
| 260696_at | At1g32520 | expressed protein | 1.54 |
| 260385_at | At1g74090 | sulfotransferase | 1.54 |
| 250758_at | At5g06000 | eukaryotic translation initiation factor 3G, putative (eIF3g) | 1.53 |
| 258468_at | At3g06070 | expressed protein | 1.53 |
| 255028_at | At4g09890 | expressed protein | 1.53 |
| 252483_at | At3g46600 | scarecrow transcription factor | 1.53 |
| 267336_at | At2g19310 | expressed protein | 1.52 |
| 252011_at | At3g52720 | carbonic anhydrase | 1.52 |
| 261754_at | At1g76130 | alpha-amylase, putative | 1.52 |
| 254971_at | At4g10380 | major intrinsic protein (MIP) | 1.52 |
| 257710_at | At3g27350 | expressed protein | 1.51 |
| 266447_at | At2g43290 | calmodulin-like protein (MSS3) | 1.51 |
| 261911_at | At1g80750 | 60S ribosomal protein L7 (RPL7A) | 1.51 |
| 258111_at | At3g14630 | cytochrome P450, putative | 1.51 |
| 256442_at | At3g10930 | expressed protein | 1.51 |
| 257323_at | orf294 | orf294 | 1.51 |
| 261139_at | At1g19700 | homeobox-leucine zipper | 1.5 |
| 260380_at | At1g73870 | zinc finger (B-box type) | 1.5 |
| 247678_at | At5g59520 | zinc transporter (ZIP2) | 1.5 |
| 254795_at | At4g12990 | expressed protein | 1.5 |
| 247367_at | At5g63290 | coproporphyrinogen oxidase-related | 1.49 |
| 252478_at | At3g46540 | clathrin assembly protein-related | 1.49 |
| 249415_at | At5g39660 | Dof-type zinc finger domain-containing protein | 1.49 |
| 262911_s_at | At1g59860 | 17.8 kDa class I heat shock protein (HSP17.8-CI) | 1.49 |
| 254146_at | At4g24260 | endo-1,4-beta-glucanase, putative | 1.49 |
| 247780_at | At5g58770 | dehydrodichyl diphosphate synthase, putative (DPS) | 1.48 |
| 261084_at | At1g07440 | tropinone reductase, putative | 1.48 |
| 259736_at | At1g64390 | endo-1,4-beta-glucanase, putative | 1.47 |
| 253313_at | At4g33870 | 1.11.1.7: peroxidase, putative | 1.47 |
| 261914_at | At1g65870 | disease resistance-responsive | 1.46 |
| 248304_at | At5g53180 | polypyrimidine tract-binding protein, putative | 1.46 |
| 251492_at | At3g59280 | signaling molecule-related | 1.46 |
| 253061_at | At4g37610 | TAZ zinc finger | 1.46 |
| 249583_at | At5g37770 | calmodulin-related protein 2, touch-induced (TCH2) | 1.45 |
| 246502_at | At5g16240 | acyl-(acyl-carrier-protein) desaturase, putative | 1.45 |
| 264850_at | At2g17340 | pantothenate kinase-related | 1.45 |
| 265339_at | At2g18230 | inorganic pyrophosphatase (PPA) | 1.45 |
| 246884_at | At5g26220 | ChaC-like | 1.45 |

| | | | |
|-------------|-----------|---|-------|
| 256245_at | At3g12580 | heat shock protein 70, putative (HSP70) | 1.44 |
| 250099_at | At5g17300 | myb family transcription factor | 1.44 |
| 249247_at | At5g42310 | pentatricopeptide (PPR) repeat-containing protein | 1.44 |
| 248799_at | At5g47230 | ethylene-responsive element-binding factor 5 (ERF5) | 1.44 |
| 247293_at | At5g64510 | expressed protein | 1.44 |
| 248270_at | At5g53450 | protein kinase | 1.44 |
| 246597_at | At5g14760 | L-aspartate oxidase | 1.43 |
| 248028_at | At5g55620 | expressed protein | 1.43 |
| 254850_at | At4g12000 | expressed protein | 1.43 |
| 249770_at | At5g24110 | WRKY family transcription factor | 1.43 |
| 250256_at | At5g13650 | elongation factor | 1.43 |
| 249344_at | At5g40770 | prohibitin | 1.43 |
| 264519_at | At1g10000 | expressed protein | 1.43 |
| 252606_at | At3g45010 | serine carboxypeptidase III, putative | 1.42 |
| 262693_at | At1g62780 | expressed protein | 1.42 |
| 265327_at | At2g18210 | expressed protein | 1.42 |
| 249063_at | At5g44110 | ABC transporter | 1.41 |
| 249575_at | At5g37670 | 15.7 kDa class I-related small heat shock protein-like (HSP15.7-CI) | 1.41 |
| 248153_at | At5g54250 | cyclic nucleotide-gated channel (CNGC4) | 1.41 |
| 267069_at | At2g41010 | VQ motif-containing protein | 1.41 |
| 247467_at | At5g62130 | Per1-like protein-related | 1.41 |
| 260653_at | At1g32440 | pyruvate kinase, putative | 1.41 |
| 260266_at | At1g68520 | zinc finger (B-box type) | 1.4 |
| 261081_at | At1g07350 | transformer serine | 1.4 |
| 267235_at | At2g43940 | expressed protein | 1.4 |
| 261166_s_at | At1g34570 | expressed protein | 1.4 |
| 250667_at | At5g07090 | 40S ribosomal protein S4 (RPS4B) | 1.4 |
| 262170_at | At1g74940 | senescence-associated protein-related | 1.39 |
| 267305_at | At2g30070 | potassium transporter (KUP1) | 1.38 |
| 267063_at | At2g41120 | expressed protein | 1.38 |
| 253638_at | At4g30470 | cinnamoyl-CoA reductase-related | 1.37 |
| 262483_at | At1g17220 | translation initiation factor IF-2, chloroplast, putative | 1.36 |
| 251800_at | At3g55510 | expressed protein | 1.36 |
| 245993_at | At5g20700 | senescence-associated protein-related | 1.35 |
| 253922_at | At4g26850 | expressed protein | 1.34 |
| 255908_s_at | At1g18010 | expressed protein | 1.34 |
| 255749_at | At1g31970 | DEAH box helicase, putative | 1.34 |
| 250832_at | At5g04950 | nicotianamine synthase, putative | 1.33 |
| 246308_at | At3g51820 | chlorophyll synthetase, putative | 1.29 |
| 259230_at | At3g07780 | expressed protein | -1.28 |
| 257350_x_at | At2g19040 | rapid alkalization factor (RALF) | -1.29 |
| 248766_at | At5g47580 | expressed protein | -1.34 |
| 248279_at | At5g52910 | timeless | -1.35 |
| 251996_at | At3g52840 | beta-galactosidase, putative | -1.36 |

| | | | |
|-------------|-----------|---|-------|
| 261467_at | At1g28520 | expressed protein | -1.36 |
| 245696_at | At5g04190 | phytochrome kinase substrate-related | -1.38 |
| 267202_s_at | At2g31030 | oxysterol-binding | -1.4 |
| 252088_at | At3g52100 | expressed protein | -1.4 |
| 264501_at | At1g09390 | GDSL-motif lipase | -1.41 |
| 245947_at | At5g19530 | spermine | -1.42 |
| 264021_at | At2g21200 | auxin-responsive protein, putative | -1.42 |
| 246178_s_at | At5g28430 | hypothetical protein | -1.43 |
| 247374_at | At5g63190 | MA3 domain-containing protein | -1.44 |
| 255281_at | At4g04970 | callose synthase, putative | -1.44 |
| 253660_at | At4g30140 | GDSL-motif lipase | -1.45 |
| 257964_at | At3g19850 | phototropic-responsive NPH3 | -1.46 |
| 259163_at | At3g01490 | protein kinase, putative | -1.46 |
| 252751_at | At3g43430 | zinc finger (C3HC4-type RING finger) | -1.49 |
| 261567_at | At1g33055 | expressed protein | -1.51 |
| 266571_at | At2g23830 | vesicle-associated membrane protein, putative (VAMP) | -1.52 |
| 264264_at | At1g09250 | expressed protein | -1.54 |
| 254197_at | At4g24040 | trehalase, putative | -1.55 |
| 261957_at | At1g64660 | Cys/Met metabolism pyridoxal-phosphate-dependent enzyme | -1.56 |
| 261395_at | At1g79700 | ovule development protein, putative | -1.59 |
| 256469_at | At1g32540 | zinc finger protein, putative | -1.61 |
| 260914_at | At1g02640 | glycosyl hydrolase | -1.61 |
| 249384_at | At5g39890 | expressed protein | -1.61 |
| 249383_at | At5g39860 | bHLH protein | -1.62 |
| 252068_at | At3g51440 | strictosidine synthase | -1.62 |
| 261026_at | At1g01240 | expressed protein | -1.65 |
| 247284_at | At5g64410 | oligopeptide transporter (OPT) | -1.67 |
| 263118_at | At1g03090 | 3-methylcrotonyl-CoA carboxylase 1 (MCCA) | -1.67 |
| 249527_at | At5g38710 | proline oxidase, putative | -1.68 |
| 264846_at | At2g17850 | senescence-associated | -1.69 |
| 252173_at | At3g50650 | scarecrow-like transcription factor 7 (SCL7) | -1.69 |
| 264529_at | At1g30820 | CTP synthase, putative | -1.69 |
| 247474_at | At5g62280 | expressed protein | -1.72 |
| 259982_at | At1g76410 | zinc finger (C3HC4-type RING finger) | -1.73 |
| 259977_at | At1g76590 | zinc-binding | -1.73 |
| 264788_at | At2g17880 | DNAJ heat shock protein, putative | -1.74 |
| 260221_at | At1g74670 | gibberellin-responsive protein, putative | -1.77 |
| 266693_at | At2g19800 | expressed protein | -1.78 |
| 264467_at | At1g10140 | expressed protein | -1.78 |
| 250327_at | At5g12050 | expressed protein | -1.8 |
| 263096_at | At2g16060 | non-symbiotic hemoglobin 1 (HB1) | -1.85 |
| 245276_at | At4g16780 | homeobox-leucine zipper protein 4 (HAT4) | -1.87 |
| 254001_at | At4g26260 | expressed protein | -1.88 |
| 245353_at | At4g16000 | expressed protein | -1.94 |
| 265481_at | At2g15960 | expressed protein | -1.97 |

| | | | |
|-------------|-----------|--|-------|
| 260900_s_at | At1g21400 | 2-oxoisovalerate dehydrogenase, putative | -1.97 |
| 245076_at | At2g23170 | auxin-responsive GH3 | -2.03 |
| 266814_at | At2g44910 | homeobox-leucine zipper protein 4 (HB-4) | -2.13 |
| 267461_at | At2g33830 | dormancy | -2.39 |
| 253423_at | At4g32280 | auxin-responsive AUX/IAA | -2.63 |
| 250662_at | At5g07010 | sulfotransferase | -2.75 |
| 264524_at | At1g10070 | branched-chain amino acid transaminase 2 (BCAT2) | -3.01 |

2 days after Ca suppression

| | | | |
|-------------|-----------|--|-------|
| 256593_at | At3g28510 | AAA-type ATPase | 17.76 |
| 245035_at | At2g26400 | acireductone dioxygenase (ARD/ARD') | 17.19 |
| 267345_at | At2g44240 | expressed protein | 13.7 |
| 259925_at | At1g75040 | pathogenesis-related protein 5 (PR-5) | 12.45 |
| 260116_at | At1g33960 | avirulence induced gene (AIG1) | 10.99 |
| 253301_at | At4g33720 | pathogenesis-related protein, putative | 10.61 |
| 266385_at | At2g14610 | pathogenesis-related protein 1 (PR-1) | 10.37 |
| 263852_at | At2g04450 | MutT/nudix | 9.79 |
| 255341_at | At4g04500 | protein kinase | 9.34 |
| 246401_at | At1g57560 | myb transcription factor (MYB50) | 8.29 |
| 254975_at | At4g10500 | oxidoreductase, 2OG-Fe(II) oxygenase | 7.88 |
| 259388_at | At1g13420 | sulfotransferase | 7.54 |
| 257099_s_at | At3g24982 | leucine-rich repeat, 5' fragment | 7.4 |
| 251625_at | At3g57260 | glycosyl hydrolase 17 protein | 7.38 |
| 257591_at | At3g24900 | disease resistance | 7.3 |
| 255689_at | At4g00670 | remorin | 7.27 |
| 251612_at | At3g57950 | expressed protein | 7.19 |
| 249890_at | At5g22570 | WRKY transcription factor | 7.12 |
| 249814_at | At5g23840 | MD-2-related lipid recognition domain-containing protein | 7.01 |
| 257100_at | At3g25010 | disease resistance | 6.99 |
| 263539_at | At2g24850 | aminotransferase, putative | 6.55 |
| 265058_s_at | At1g52040 | myrosinase-binding protein, putative (F-ATMBP) | 5.92 |
| 251928_at | At3g53980 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 5.86 |
| 254828_at | At4g12550 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 5.86 |
| 251673_at | At3g57240 | beta-1,3-glucanase (BG3) | 5.81 |
| 266989_at | At2g39330 | jacalin lectin | 5.77 |
| 249812_at | At5g23830 | MD-2-related lipid recognition domain-containing protein | 5.75 |
| 255940_at | At1g20380 | prolyl oligopeptidase, putative | 5.72 |
| 248728_at | At5g48000 | cytochrome P450 | 5.65 |
| 255912_at | At1g66960 | lupeol synthase, putative | 5.6 |
| 255254_at | At4g05030 | heavy-metal-associated domain-containing protein | 5.54 |
| 248844_s_at | At5g46900 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 5.39 |
| 263161_at | At1g54020 | myrosinase-associated protein, putative | 5.33 |
| 262838_at | At1g14960 | major latex protein-related | 5.31 |
| 262516_at | At1g17190 | glutathione S-transferase, putative | 5.31 |
| 260869_at | At1g43800 | acyl- (acyl-carrier-protein) desaturase, putative | 5.29 |
| 255807_at | At4g10270 | wound-responsive | 5.26 |

| | | | |
|-------------|-----------|---|------|
| 252269_at | At3g49580 | expressed protein | 5.23 |
| 248227_at | At5g53820 | expressed protein | 5.16 |
| 249364_at | At5g40590 | DC1 domain-containing protein | 5.14 |
| 259507_at | At1g43910 | AAA-type ATPase | 5.13 |
| 249438_at | At5g40010 | AAA-type ATPase | 5.04 |
| 257774_at | At3g29250 | short-chain dehydrogenase/reductase (SDR) | 5.03 |
| 254385_s_at | At4g21830 | methionine sulfoxide reductase domain-containing protein | 4.94 |
| 248169_at | At5g54610 | ankyrin repeat | 4.94 |
| 260117_at | At1g33950 | avirulence induced gene (AIG1) | 4.93 |
| 259655_at | At1g55210 | disease resistance response protein-related | 4.85 |
| 246825_at | At5g26260 | meprin and TRAF homology domain-containing protein | 4.78 |
| 250764_at | At5g05960 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 4.77 |
| 246125_at | At5g19875 | expressed protein | 4.76 |
| 258218_at | At3g18000 | phosphoethanolamine N-methyltransferase 1 (NMT1) | 4.75 |
| 252938_at | At4g39190 | expressed protein | 4.75 |
| 254907_at | At4g11190 | disease resistance-responsive | 4.73 |
| 263096_at | At2g16060 | non-symbiotic hemoglobin 1 (HB1) | 4.72 |
| 259550_at | At1g35230 | arabinogalactan-protein (AGP5) | 4.69 |
| 260130_s_at | At1g66280 | beta-glucosidase (PSR3.2) | 4.65 |
| 259813_at | At1g49860 | glutathione S-transferase, putative | 4.61 |
| 263594_at | At2g01880 | purple acid phosphatase (PAP7) | 4.6 |
| 263854_at | At2g04430 | MutT/nudix | 4.56 |
| 254271_at | At4g23150 | protein kinase | 4.56 |
| 266376_at | At2g14620 | xyloglucan:xyloglucosyl transferase, putative | 4.56 |
| 248727_at | At5g47990 | cytochrome P450 | 4.51 |
| 250062_at | At5g17760 | AAA-type ATPase | 4.51 |
| 247755_at | At5g59090 | subtilase | 4.5 |
| 250942_at | At5g03350 | legume lectin | 4.46 |
| 249096_at | At5g43910 | pfkB-type carbohydrate kinase | 4.43 |
| 267567_at | At2g30770 | cytochrome P450 71A13, putative (CYP71A13) | 4.41 |
| 266336_at | At2g32270 | zinc transporter (ZIP3) | 4.34 |
| 255630_at | At4g00700 | C2 domain-containing protein | 4.27 |
| 249167_at | At5g42860 | expressed protein | 4.27 |
| 249867_at | At5g23020 | 2-isopropylmalate synthase 2 (IMS2) | 4.26 |
| 256833_at | At3g22910 | calcium-transporting ATPase, plasma membrane-type, putative (ACA13) | 4.24 |
| 249491_at | At5g39130 | germin-like protein, putative (GER2) | 4.24 |
| 256368_at | At1g66800 | cinnamyl-alcohol dehydrogenase | 4.22 |
| 247337_at | At5g63660 | plant defensin-fusion protein, putative (PDF2.5) | 4.19 |
| 255110_at | At4g08770 | peroxidase, putative | 4.18 |
| 252605_s_at | At3g45070 | sulfotransferase | 4.16 |
| 249743_at | At5g24540 | glycosyl hydrolase 1 protein | 4.15 |
| 248889_at | At5g46230 | expressed protein | 4.09 |
| 248048_at | At5g56080 | nicotianamine synthase, putative | 4.09 |
| 253608_at | At4g30290 | xyloglucan:xyloglucosyl transferase, putative | 4.06 |

| | | | |
|-------------|-----------|--|------|
| 260462_at | At1g10970 | metal transporter, putative (ZIP4) | 4.06 |
| 245555_at | At4g15390 | transferase | 4.05 |
| 259681_at | At1g77760 | nitrate reductase 1 (NR1) | 4.02 |
| 257066_at | At3g18280 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 4.01 |
| 246149_at | At5g19890 | peroxidase, putative | 4 |
| 259173_at | At3g03640 | glycosyl hydrolase 1 protein | 3.96 |
| 256596_at | At3g28540 | AAA-type ATPase | 3.96 |
| 264953_at | At1g77120 | alcohol dehydrogenase (ADH) | 3.94 |
| 260904_at | At1g02450 | NIM1-interacting protein 1 (NIMIN-1) | 3.93 |
| 249203_at | At5g42590 | cytochrome P450 71A16, putative (CYP71A16) | 3.89 |
| 266383_at | At2g14580 | pathogenesis-related protein, putative | 3.84 |
| 256265_at | At3g12220 | serine carboxypeptidase S10 | 3.84 |
| 250646_at | At5g06720 | peroxidase, putative | 3.82 |
| 260713_at | At1g17615 | disease resistance protein (TIR-NBS class), putative | 3.81 |
| 248062_at | At5g55450 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 3.81 |
| 256289_s_at | At3g12230 | serine carboxypeptidase S10 | 3.8 |
| 248139_at | At5g54970 | expressed protein | 3.77 |
| 264998_at | At1g67330 | expressed protein | 3.76 |
| 261986_s_at | At1g33720 | cytochrome P450, putative | 3.69 |
| 261763_at | At1g15520 | ABC transporter | 3.69 |
| 251232_at | At3g62780 | C2 domain-containing protein | 3.68 |
| 257101_at | At3g25020 | disease resistance | 3.67 |
| 250455_at | At5g09980 | expressed protein | 3.66 |
| 259065_at | At3g07520 | glutamate receptor (GLR1.4) | 3.65 |
| 259559_at | At1g21240 | wall-associated kinase, putative (WAK3) | 3.64 |
| 265050_at | At1g52070 | jacalin lectin | 3.62 |
| 259040_at | At3g09270 | glutathione S-transferase, putative | 3.62 |
| 266363_at | At2g41250 | haloacid dehalogenase-like hydrolase | 3.62 |
| 246855_at | At5g26280 | meprin and TRAF homology domain-containing protein | 3.61 |
| 250661_at | At5g07030 | aspartyl protease | 3.57 |
| 257763_s_at | At3g23110 | disease resistance | 3.57 |
| 257473_at | At1g33840 | hypothetical protein | 3.54 |
| 266415_at | At2g38530 | nonspecific lipid transfer protein 2 (LTP2) | 3.54 |
| 248178_at | At5g54370 | late embryogenesis abundant protein-related | 3.53 |
| 253413_at | At4g33020 | metal transporter, putative (ZIP9) | 3.52 |
| 256431_s_at | At3g11010 | disease resistance | 3.51 |
| 249752_at | At5g24660 | expressed protein | 3.51 |
| 258377_at | At3g17690 | cyclic nucleotide-binding transporter 2 (CNBT2) | 3.5 |
| 265161_at | At1g30900 | vacuolar sorting receptor, putative (AtELP6) | 3.49 |
| 254361_at | At4g22212 | expressed protein | 3.47 |
| 251970_at | At3g53150 | UDP-glucuronosyl/UDP-glucosyl transferase | 3.46 |
| 248330_at | At5g52810 | ornithine cyclodeaminase | 3.46 |
| 249004_at | At5g44570 | hypothetical protein | 3.45 |
| 259276_at | At3g01190 | peroxidase 27 (PER27) | 3.44 |
| 255575_at | At4g01430 | nodulin MtN21 | 3.43 |

| | | | |
|-------------|-----------|--|------|
| 257673_at | At3g20370 | meprin and TRAF homology domain-containing protein | 3.42 |
| 253684_at | At4g29690 | type I phosphodiesterase | 3.38 |
| 257592_at | At3g24954 | leucine-rich repeat , 5' fragment | 3.37 |
| 252607_at | At3g44990 | xyloglucan:xyloglucosyl transferase, putative | 3.36 |
| 255516_at | At4g02270 | pollen Ole e 1 allergen and extensin | 3.35 |
| 248676_at | At5g48850 | male sterility (MS5) | 3.35 |
| 249732_at | At5g24420 | glucosamine | 3.35 |
| 254741_s_at | At4g13900 | pseudogene, similar to NL0D | 3.34 |
| 266743_at | At2g02990 | ribonuclease 1 (RNS1) | 3.34 |
| 256781_at | At3g13650 | disease resistance response protein-related | 3.34 |
| 261684_at | At1g47400 | expressed protein | 3.33 |
| 257952_at | At3g21770 | peroxidase 30 (PER30) | 3.32 |
| 260623_at | At1g08090 | high-affinity nitrate transporter (ACH1) | 3.32 |
| 250798_at | At5g05340 | peroxidase, putative | 3.32 |
| 260568_at | At2g43570 | chitinase, putative | 3.32 |
| 266142_at | At2g39030 | GCN5-related N-acetyltransferase (GNAT) | 3.32 |
| 252421_at | At3g47540 | chitinase, putative | 3.31 |
| 264958_at | At1g76960 | expressed protein | 3.31 |
| 262671_at | At1g76040 | calcium-dependent protein kinase, putative | 3.29 |
| 250724_at | At5g06330 | hairpin-responsive protein, putative (HIN1) | 3.29 |
| 252888_at | At4g39210 | glucose-1-phosphate adenylyltransferase large subunit 3 (APL3) | 3.29 |
| 254232_at | At4g23600 | coronatine-responsive tyrosine aminotransferase | 3.28 |
| 247717_at | At5g59320 | lipid transfer protein 3 (LTP3) | 3.27 |
| 267121_at | At2g23540 | GDSL-motif lipase/hydrolase | 3.26 |
| 249836_at | At5g23420 | high mobility group (HMG1/2) | 3.25 |
| 255111_at | At4g08780 | peroxidase, putative | 3.25 |
| 259609_at | At1g52410 | caldesmon-related | 3.24 |
| 254820_s_at | At4g12510 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 3.23 |
| 261335_at | At1g44800 | nodulin MtN21 | 3.22 |
| 264567_s_at | At1g05250 | peroxidase, putative | 3.22 |
| 262427_s_at | At1g47600 | glycosyl hydrolase family 1 protein | 3.22 |
| 254265_s_at | At4g23140 | receptor-like protein kinase 5 (RLK5) | 3.22 |
| 254805_at | At4g12480 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 3.21 |
| 257363_at | At2g45760 | BON1-associated protein (BAP1)-related | 3.21 |
| 250445_at | At5g10760 | aspartyl protease | 3.21 |
| 247314_at | At5g64000 | 3' (2'),5'-bisphosphate nucleotidase | 3.2 |
| 245689_at | At5g04120 | phosphoglycerate | 3.2 |
| 261157_at | At1g34510 | peroxidase, putative | 3.2 |
| 253298_at | At4g33560 | expressed protein | 3.18 |
| 247210_at | At5g65020 | annexin 2 (ANN2) | 3.17 |
| 263153_s_at | At1g54010 | myrosinase-associated protein, putative | 3.16 |
| 253667_at | At4g30170 | peroxidase, putative | 3.16 |
| 252060_at | At3g52430 | phytoalexin-deficient 4 protein (PAD4) | 3.15 |
| 259403_at | At1g17745 | D-3-phosphoglycerate dehydrogenase (3-PGDH) | 3.14 |

| | | | |
|-------------|-----------|--|------|
| 245257_at | At4g14640 | calmodulin-8 (CAM8) | 3.14 |
| 267546_at | At2g32680 | disease resistance | 3.13 |
| 256933_at | At3g22600 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 3.13 |
| 245889_at | At5g09480 | hydroxyproline-rich glycoprotein | 3.12 |
| 250689_at | At5g06610 | expressed protein | 3.12 |
| 256647_at | At3g13610 | oxidoreductase, 2OG-Fe(II) oxygenase | 3.11 |
| 266967_at | At2g39530 | integral membrane protein, putative | 3.11 |
| 266292_at | At2g29350 | tropinone reductase, putative | 3.1 |
| 253687_at | At4g29520 | expressed protein | 3.1 |
| 264648_at | At1g09080 | luminal binding protein 3 (BiP-3) | 3.09 |
| 263063_s_at | At2g18140 | peroxidase, putative | 3.08 |
| 257421_at | At1g12030 | expressed protein | 3.08 |
| 250832_at | At5g04950 | nicotianamine synthase, putative | 3.07 |
| 247684_at | At5g59670 | leucine-rich repeat protein kinase, putative | 3.07 |
| 251422_at | At3g60540 | sec61beta | 3.06 |
| 261562_at | At1g01750 | actin-depolymerizing factor, putative | 3.06 |
| 255622_at | At4g01070 | UDP-glucuronosyl/UDP-glucosyl transferase | 3.05 |
| 249052_at | At5g44420 | plant defensin protein, putative (PDF1.2a) | 3.04 |
| 251045_s_at | At5g02360 | pseudogene, CHP-rich zinc finger protein, putative | 3.04 |
| 258470_at | At3g06035 | expressed protein | 3.02 |
| 257432_at | At2g21850 | DC1 domain-containing protein | 3.01 |
| 249599_at | At5g37990 | S-adenosyl-L-methionine:carboxyl methyltransferase | 3 |
| 264262_at | At1g09200 | histone H3 | 2.99 |
| 266223_at | At2g28790 | osmotin-like protein, putative | 2.98 |
| 253073_at | At4g37410 | cytochrome P450, putative | 2.96 |
| 250302_at | At5g11920 | glycosyl hydrolase family 32 protein | 2.96 |
| 247765_at | At5g58860 | cytochrome P450 86A1 (CYP86A1) | 2.96 |
| 261443_at | At1g28480 | glutaredoxin | 2.96 |
| 249061_at | At5g44550 | integral membrane | 2.95 |
| 254606_at | At4g19030 | major intrinsic protein (MIP) | 2.95 |
| 255795_at | At2g33380 | calcium-binding RD20 protein (RD20) | 2.95 |
| 248848_at | At5g46520 | disease resistance protein (TIR-NBS-LRR class), putative | 2.95 |
| 247318_at | At5g63990 | 3'(2'),5'-bisphosphate nucleotidase, putative | 2.94 |
| 257185_at | At3g13100 | ABC transporter | 2.94 |
| 252511_at | At3g46280 | protein kinase-related | 2.94 |
| 259870_at | At1g76780 | expressed protein | 2.93 |
| 259553_x_at | At1g21310 | proline-rich extensin-like | 2.92 |
| 249576_at | At5g37690 | GDSL-motif lipase/hydrolase | 2.91 |
| 267496_at | At2g30550 | lipase class 3 | 2.9 |
| 251419_at | At3g60470 | hypothetical protein | 2.9 |
| 253998_at | At4g26010 | peroxidase, putative | 2.9 |
| 254190_at | At4g23885 | expressed protein | 2.89 |
| 246481_s_at | At5g15960 | stress-responsive protein (KIN1) | 2.89 |
| 246228_at | At4g36430 | peroxidase, putative | 2.88 |
| 254326_at | At4g22610 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 2.88 |

| | | | |
|-------------|-----------|--|------|
| 265049_at | At1g52060 | jacalin lectin | 2.88 |
| 249195_s_at | At5g42500 | disease resistance-responsive | 2.88 |
| 265658_at | At2g13810 | aminotransferase class I and II | 2.87 |
| 247268_at | At5g64080 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 2.86 |
| 266070_at | At2g18660 | expansin (EXPR3) | 2.86 |
| 247965_at | At5g56540 | arabinogalactan-protein (AGP14) | 2.86 |
| 256125_at | At1g18250 | thaumatin, putative | 2.85 |
| 266271_at | At2g29440 | glutathione S-transferase, putative | 2.85 |
| 251060_at | At5g01820 | CBL-interacting protein kinase 14 (CIPK14) | 2.84 |
| 249051_at | At5g44390 | FAD-binding domain-containing protein | 2.84 |
| 254603_at | At4g19050 | mob1/phocein | 2.83 |
| 247477_at | At5g62340 | invertase/pectin methylesterase inhibitor | 2.83 |
| 250433_at | At5g10400 | histone H3 | 2.83 |
| 250517_at | At5g08260 | serine carboxypeptidase S10 | 2.83 |
| 259721_at | At1g60890 | phosphatidylinositol-4-phosphate 5-kinase | 2.83 |
| 262050_at | At1g80130 | expressed protein | 2.83 |
| 250434_at | At5g10390 | histone H3 | 2.82 |
| 257139_at | At3g28890 | leucine-rich repeat | 2.82 |
| 261125_at | At1g04990 | zinc finger (CCCH-type) | 2.82 |
| 265102_at | At1g30870 | cationic peroxidase, putative | 2.82 |
| 254387_at | At4g21850 | methionine sulfoxide reductase domain-containing protein | 2.82 |
| 258675_at | At3g08770 | lipid transfer protein 6 (LTP6) | 2.82 |
| 245401_at | At4g17670 | senescence-associated protein-related | 2.81 |
| 260955_at | At1g06000 | UDP-glucuronosyl/UDP-glucosyl transferase | 2.81 |
| 260068_at | At1g73805 | calmodulin-binding protein | 2.81 |
| 258930_at | At3g10040 | expressed protein | 2.8 |
| 248248_at | At5g53120 | spermidine synthase, putative | 2.79 |
| 248729_at | At5g48010 | pentacyclic triterpene synthase, putative (ATPEN1) | 2.79 |
| 258859_at | At3g02120 | hydroxyproline-rich glycoprotein | 2.78 |
| 253842_at | At4g27860 | integral membrane | 2.78 |
| 252618_at | At3g45140 | lipoxygenase (LOX2) | 2.78 |
| 251457_s_at | At3g60160 | ABC transporter | 2.78 |
| 245385_at | At4g14020 | rapid alkalization factor (RALF) | 2.78 |
| 262939_s_at | At1g79530 | glyceraldehyde 3-phosphate dehydrogenase, putative | 2.77 |
| 253024_at | At4g38080 | hydroxyproline-rich glycoprotein | 2.77 |
| 248823_s_at | At5g46960 | invertase/pectin methylesterase inhibitor | 2.77 |
| 265943_at | At2g19570 | cytidine deaminase (CDD) | 2.76 |
| 254226_at | At4g23690 | disease resistance-responsive | 2.76 |
| 255591_at | At4g01630 | expansin, putative (EXP17) | 2.74 |
| 253879_s_at | At4g27570 | glycosyltransferase | 2.74 |
| 251065_at | At5g01870 | lipid transfer protein, putative | 2.73 |
| 251418_at | At3g60440 | expressed protein | 2.73 |
| 261606_at | At1g49570 | peroxidase, putative | 2.73 |
| 253060_at | At4g37710 | VQ motif-containing protein | 2.72 |
| 252448_at | At3g47050 | glycosyl hydrolase 3 protein | 2.71 |

| | | | |
|-----------|-----------|---|------|
| 251226_at | At3g62680 | proline-rich | 2.71 |
| 263535_at | At2g24970 | expressed protein | 2.7 |
| 258080_at | At3g25930 | universal stress protein (USP) | 2.7 |
| 249046_at | At5g44400 | FAD-binding domain-containing protein | 2.7 |
| 248309_at | At5g52540 | expressed protein | 2.69 |
| 256829_at | At3g22850 | expressed protein | 2.69 |
| 259910_at | At1g72700 | haloacid dehalogenase-like hydrolase | 2.69 |
| 265837_at | At2g14560 | expressed protein | 2.69 |
| 266358_at | At2g32280 | expressed protein | 2.69 |
| 259632_at | At1g56430 | nicotianamine synthase, putative | 2.69 |
| 254252_at | At4g23310 | receptor-like protein kinase, putative | 2.68 |
| 257715_at | At3g12750 | zinc transporter (ZIP1) | 2.68 |
| 254736_at | At4g13820 | disease resistance | 2.67 |
| 252487_at | At3g46660 | UDP-glucuronosyl/UDP-glucosyl transferase | 2.67 |
| 259640_at | At1g52400 | glycosyl hydrolase family 1 protein | 2.66 |
| 258942_at | At3g09960 | calcineurin-like phosphoesterase | 2.66 |
| 251952_at | At3g53650 | histone H2B, putative | 2.66 |
| 254044_at | At4g25820 | endo-xyloglucan transferase (XTR9) | 2.66 |
| 247594_at | At5g60800 | heavy-metal-associated domain-containing protein | 2.66 |
| 249545_at | At5g38030 | MATE efflux | 2.66 |
| 250172_at | At5g14330 | expressed protein | 2.65 |
| 256489_at | At1g31550 | GDSL-motif lipase, putative | 2.65 |
| 260884_at | At1g29240 | expressed protein | 2.65 |
| 256569_at | At3g19550 | expressed protein | 2.65 |
| 253657_at | At4g30110 | ATPase E1-E2 type | 2.65 |
| 264005_at | At2g22470 | arabinogalactan-protein (AGP2) | 2.64 |
| 248968_at | At5g45280 | pectinacetyltransferase, putative | 2.64 |
| 256911_at | At3g24090 | hexosephosphate aminotransferase, putative (GLCN6P) | 2.64 |
| 263536_at | At2g25000 | WRKY transcription factor | 2.64 |
| 262275_at | At1g68710 | haloacid dehalogenase-like hydrolase | 2.63 |
| 263250_at | At2g31390 | pfkB-type carbohydrate kinase | 2.63 |
| 264386_at | At1g12000 | pyrophosphate-fructose-6-phosphate 1-phosphotransferase beta subunit, putative | 2.62 |
| 262281_at | At1g68570 | proton-dependent oligopeptide transport (POT) | 2.62 |
| 247192_at | At5g65360 | histone H3 | 2.61 |
| 252403_at | At3g48080 | lipase class 3 | 2.61 |
| 253334_at | At4g33360 | terpene cyclase | 2.61 |
| 255342_at | At4g04510 | protein kinase | 2.61 |
| 262504_at | At1g21750 | protein disulfide isomerase, putative | 2.61 |
| 255406_at | At4g03450 | ankyrin repeat | 2.6 |
| 246055_at | At5g08380 | alpha-galactosidase, putative | 2.6 |
| 251739_at | At3g56170 | Ca(2+)-dependent nuclease | 2.59 |
| 246842_at | At5g26731 | expressed protein | 2.59 |
| 247727_at | At5g59490 | haloacid dehalogenase-like hydrolase | 2.59 |
| 260745_at | At1g78370 | glutathione S-transferase, putative | 2.59 |

| | | | |
|-------------|-----------|---|------|
| 247151_at | At5g65640 | basic helix-loop-helix (bHLH) | 2.58 |
| 266364_at | At2g41230 | expressed protein | 2.58 |
| 260560_at | At2g43590 | chitinase, putative | 2.57 |
| 266776_at | At2g29070 | ubiquitin fusion degradation UFD1 | 2.56 |
| 257714_at | At3g27360 | histone H3 | 2.56 |
| 255842_at | At2g33530 | serine carboxypeptidase S10 | 2.55 |
| 257295_at | At3g17420 | protein kinase | 2.55 |
| 251372_at | At3g60520 | expressed protein | 2.55 |
| 260582_at | At2g47200 | expressed protein | 2.55 |
| 255595_at | At4g01700 | chitinase, putative | 2.55 |
| 264832_at | At1g03660 | expressed protein | 2.55 |
| 252417_at | At3g47480 | calcium-binding EF hand | 2.54 |
| 248692_s_at | At4g15070 | DC1 domain-containing protein | 2.54 |
| 265962_at | At2g37460 | nodulin MtN21 | 2.54 |
| 247651_at | At5g59870 | histone H2A, putative | 2.53 |
| 265117_at | At1g62500 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 2.53 |
| 257064_at | At3g18260 | reticulon (RTNLB9) | 2.53 |
| 254166_at | At4g24190 | shepherd protein (SHD) | 2.53 |
| 260392_at | At1g74030 | enolase, putative | 2.52 |
| 250702_at | At5g06730 | peroxidase, putative | 2.52 |
| 266581_at | At2g46140 | late embryogenesis abundant protein, putative / LEA protein, putative | 2.5 |
| 259120_at | At3g02240 | expressed protein | 2.5 |
| 251840_at | At3g54960 | thioredoxin | 2.5 |
| 256736_at | At3g29410 | terpene synthase/cyclase | 2.5 |
| 256252_at | At3g11340 | UDP-glucuronosyl/UDP-glucosyl transferase | 2.49 |
| 266395_at | At2g43100 | aconitase C-terminal domain-containing protein | 2.49 |
| 248499_at | At5g50400 | calcineurin-like phosphoesterase | 2.49 |
| 251287_at | At3g61820 | aspartyl protease | 2.49 |
| 252921_at | At4g39030 | enhanced disease susceptibility 5 (EDS5) | 2.48 |
| 251298_at | At3g62040 | haloacid dehalogenase-like hydrolase | 2.48 |
| 254791_at | At4g12910 | serine carboxypeptidase S10 | 2.48 |
| 250286_at | At5g13320 | auxin-responsive GH3 | 2.47 |
| 255340_at | At4g04490 | protein kinase | 2.47 |
| 246390_at | At1g77330 | 1-aminocyclopropane-1-carboxylate oxidase, putative | 2.47 |
| 254718_at | At4g13580 | disease resistance-responsive | 2.47 |
| 246524_at | At5g15860 | expressed protein | 2.46 |
| 253103_at | At4g36110 | auxin-responsive protein, putative | 2.46 |
| 245738_at | At1g44130 | nucellin protein, putative | 2.46 |
| 265354_at | At2g16700 | actin-depolymerizing factor 5 (ADF5) | 2.46 |
| 251517_at | At3g59370 | expressed protein | 2.46 |
| 261167_at | At1g04980 | thioredoxin | 2.46 |
| 265025_at | At1g24575 | expressed protein | 2.46 |
| 264071_at | At2g27920 | serine carboxypeptidase S10 | 2.45 |
| 260234_at | At1g74460 | GDSL-motif lipase/hydrolase | 2.45 |

| | | | |
|-------------|-----------|--|------|
| 247618_at | At5g60280 | lectin protein kinase | 2.45 |
| 264886_at | At1g61120 | terpene synthase/cyclase | 2.45 |
| 256560_s_at | At3g31415 | terpene synthase/cyclase | 2.45 |
| 252495_at | At3g46770 | transcriptional factor B3 | 2.45 |
| 266270_at | At2g29470 | glutathione S-transferase, putative | 2.45 |
| 261020_at | At1g26390 | FAD-binding domain-containing protein | 2.44 |
| 263882_at | At2g21790 | ribonucleoside-diphosphate reductase small chain, putative / ribonucleotide reductase, putative | 2.44 |
| 260077_at | At1g73620 | thaumatin-like protein, putative / pathogenesis-related protein, putative | 2.44 |
| 267590_at | At2g39700 | expansin, putative (EXP4) | 2.44 |
| 265659_at | At2g25440 | leucine-rich repeat | 2.44 |
| 254229_at | At4g23610 | expressed protein | 2.44 |
| 262978_at | At1g75780 | tubulin beta-1 chain (TUB1) | 2.44 |
| 246197_at | At4g37010 | caltractin, putative | 2.44 |
| 266464_at | At2g47800 | glutathione-conjugate transporter (MRP4) | 2.44 |
| 253697_at | At4g29700 | type I phosphodiesterase | 2.44 |
| 265048_at | At1g52050 | jacalin lectin | 2.43 |
| 255874_at | At2g40550 | expressed protein | 2.43 |
| 261363_at | At1g41830 | multi-copper oxidase type I | 2.43 |
| 261999_at | At1g33800 | expressed protein | 2.43 |
| 247600_at | At5g60890 | receptor-like protein kinase (ATR1) (MYB34) | 2.43 |
| 263941_at | At2g35870 | At2g35870 | 2.43 |
| 253113_at | At4g35985 | senescence/dehydration-associated protein-related | 2.42 |
| 264846_at | At2g17850 | senescence-associated | 2.42 |
| 246652_at | At5g35190 | proline-rich extensin-like | 2.41 |
| 266669_at | At2g29750 | UDP-glucuronosyl/UDP-glucosyl transferase | 2.41 |
| 265656_at | At2g13820 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 2.4 |
| 259228_at | At3g07720 | kelch repeat-containing protein | 2.4 |
| 256962_at | At3g13560 | glycosyl hydrolase family 17 protein | 2.4 |
| 254789_at | At4g12880 | plastocyanin-like domain-containing protein | 2.4 |
| 249474_s_at | At5g39190 | germin-like protein, putative | 2.4 |
| 261269_at | At1g26690 | emp24/gp25L/p24 | 2.39 |
| 264260_at | At1g09210 | calreticulin 2 (CRT2) | 2.39 |
| 253416_at | At4g33070 | pyruvate decarboxylase, putative | 2.39 |
| 254740_s_at | At4g13890 | glycine hydroxymethyltransferase, putative | 2.39 |
| 264263_at | At1g09155 | SKP1 interacting partner 3-related | 2.39 |
| 255127_at | At4g08300 | nodulin MtN21 | 2.39 |
| 246991_at | At5g67400 | peroxidase 73 (PER73) (P73) (PRXR11) | 2.38 |
| 262646_at | At1g62800 | aspartate aminotransferase, cytoplasmic isozyme 2 / transaminase A (ASP4) | 2.37 |
| 256712_at | At2g34020 | calcium-binding EF hand | 2.37 |
| 265572_at | At2g28210 | carbonic anhydrase | 2.37 |
| 252661_at | At3g44450 | expressed protein | 2.37 |
| 260179_at | At1g70690 | kinase-related | 2.37 |
| 258037_at | At3g21230 | 4-coumaroyl-CoA synthase, putative (4CL) | 2.37 |

| | | | |
|-------------|-----------|--|------|
| 259878_at | At1g76790 | O-methyltransferase family 2 protein | 2.37 |
| 257844_at | At3g28480 | oxidoreductase, 2OG-Fe(II) oxygenase | 2.36 |
| 265464_at | At2g37080 | myosin heavy chain-related | 2.36 |
| 249916_at | At5g22880 | histone H2B, putative | 2.36 |
| 249600_s_at | At5g38000 | NADP-dependent oxidoreductase, putative | 2.36 |
| 253228_at | At4g34630 | expressed protein | 2.36 |
| 248820_at | At5g47060 | senescence-associated protein-related | 2.36 |
| 256428_at | At3g11080 | disease resistance | 2.36 |
| 260693_at | At1g32450 | proton-dependent oligopeptide transport (POT) | 2.36 |
| 258751_at | At3g05890 | hydrophobic protein (RC12B) | 2.36 |
| 262832_s_at | At1g14870 | expressed protein | 2.35 |
| 254243_at | At4g23210 | protein kinase | 2.35 |
| 267472_at | At2g02850 | plastocyanin-like domain-containing protein | 2.35 |
| 250596_at | At5g07780 | formin homology 2 domain-containing protein | 2.35 |
| 261080_at | At1g07370 | proliferating cell nuclear antigen 1 (PCNA1) | 2.35 |
| 258217_at | At3g17990 | phosphoethanolamine N-methyltransferase 1 (NMT1) | 2.35 |
| 259723_at | At1g60960 | metal transporter, putative (IRT3) | 2.35 |
| 267053_s_at | At2g38390 | peroxidase 22 (PER22) | 2.34 |
| 260784_at | At1g06180 | myb family transcription factor | 2.34 |
| 261366_at | At1g53100 | glycosyltransferase family 14 protein | 2.34 |
| 246230_at | At4g36710 | scarecrow transcription factor | 2.34 |
| 264144_at | At1g79320 | latex abundant protein, putative (AMC5) | 2.34 |
| 267388_at | At2g44450 | glycosyl hydrolase family 1 protein | 2.34 |
| 258894_at | At3g05650 | disease resistance | 2.34 |
| 248971_at | At5g45000 | Toll-Interleukin-Resistance (TIR) domain-containing protein | 2.33 |
| 248263_at | At5g53370 | pectinesterase | 2.33 |
| 250344_at | At5g11930 | glutaredoxin | 2.33 |
| 265053_at | At1g52000 | jacalin lectin | 2.32 |
| 245315_at | At4g14800 | 20S proteasome beta subunit D2 (PBD2) | 2.32 |
| 264044_at | At2g22480 | phosphofructokinase | 2.32 |
| 267457_at | At2g33790 | pollen Ole e 1 allergen and extensin | 2.32 |
| 261692_at | At1g08450 | calreticulin 3 (CRT3) | 2.32 |
| 261305_at | At1g48470 | glutamine synthetase, putative | 2.32 |
| 257500_s_at | At1g73300 | serine carboxypeptidase S10 | 2.32 |
| 260941_at | At1g44970 | peroxidase, putative | 2.31 |
| 254832_at | At4g12490 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 2.31 |
| 256216_at | At1g56340 | calreticulin 1 (CRT1) | 2.31 |
| 246532_at | At5g15870 | glycosyl hydrolase 81 protein | 2.31 |
| 262930_at | At1g65690 | harpin-induced protein-related | 2.3 |
| 256543_at | At1g42480 | expressed protein | 2.3 |
| 254894_at | At4g11840 | phospholipase D gamma 3 (PLDGAMMA3) | 2.3 |
| 254085_at | At4g24960 | ABA-responsive protein (HVA22d) | 2.3 |
| 253268_s_at | At4g34135 | UDP-glucuronosyl/UDP-glucosyl transferase | 2.3 |
| 260919_at | At1g21520 | expressed protein | 2.3 |
| 247009_at | At5g67600 | expressed protein | 2.29 |

| | | | |
|-------------|-----------|--|------|
| 251182_at | At3g62600 | DNAJ heat shock | 2.29 |
| 259757_at | At1g77510 | protein disulfide isomerase, putative | 2.29 |
| 249306_at | At5g41400 | zinc finger (C3HC4-type RING finger) | 2.29 |
| 267307_at | At2g30210 | laccase, putative | 2.29 |
| 249375_at | At5g40730 | arabinogalactan-protein (AGP24) | 2.28 |
| 264968_at | At1g67360 | rubber elongation factor (REF) | 2.28 |
| 253040_at | At4g37800 | xyloglucan:xyloglucosyl transferase, putative | 2.28 |
| 250438_at | At5g10580 | expressed protein | 2.28 |
| 261576_at | At1g01070 | nodulin MtN21 | 2.28 |
| 260408_at | At1g69880 | thioredoxin, putative | 2.28 |
| 253627_at | At4g30650 | hydrophobic protein, putative | 2.28 |
| 245688_at | At1g28290 | pollen Ole e 1 allergen and extensin | 2.27 |
| 261240_at | At1g32940 | subtilase | 2.27 |
| 259251_at | At3g07600 | heavy-metal-associated domain-containing protein | 2.27 |
| 249477_s_at | At5g38930 | germin-like protein, putative | 2.27 |
| 247604_at | At5g60950 | phytochelatin synthetase-related | 2.27 |
| 255587_at | At4g01480 | inorganic pyrophosphatase, putative (soluble) | 2.27 |
| 248252_at | At5g53250 | arabinogalactan-protein, putative (AGP22) | 2.27 |
| 245151_at | At2g47550 | pectinesterase | 2.26 |
| 266596_at | At2g46150 | expressed protein | 2.26 |
| 263963_at | At2g36080 | DNA-binding protein, putative | 2.26 |
| 251368_at | At3g61380 | expressed protein | 2.26 |
| 264537_at | At1g55610 | protein kinase | 2.26 |
| 264579_at | At1g05205 | expressed protein | 2.26 |
| 249934_at | At5g22410 | peroxidase, putative | 2.26 |
| 261921_at | At1g65900 | expressed protein | 2.26 |
| 257153_at | At3g27220 | kelch repeat-containing protein | 2.26 |
| 250778_at | At5g05500 | pollen Ole e 1 allergen and extensin | 2.26 |
| 252098_at | At3g51330 | aspartyl protease | 2.25 |
| 255517_at | At4g02290 | glycosyl hydrolase family 9 protein | 2.25 |
| 255632_at | At4g00680 | actin-depolymerizing factor, putative | 2.25 |
| 251304_at | At3g61990 | O-methyltransferase family 3 protein | 2.24 |
| 257644_at | At3g25780 | allene oxide cyclase, putative | 2.24 |
| 256243_at | At3g12500 | basic endochitinase | 2.24 |
| 250059_at | At5g17820 | peroxidase 57 (PER57) | 2.24 |
| 267432_at | At2g35020 | UTP--glucose-1-phosphate uridylyltransferase | 2.24 |
| 267483_at | At2g02810 | UDP-galactose/UDP-glucose transporter | 2.24 |
| 245944_at | At5g19520 | mechanosensitive ion channel domain-containing protein | 2.24 |
| 256927_at | At3g22550 | senescence-associated protein-related | 2.24 |
| 260046_at | At1g73800 | calmodulin-binding protein | 2.24 |
| 262717_s_at | At1g16410 | cytochrome P450 | 2.24 |
| 249910_at | At5g22630 | prephenate dehydratase | 2.23 |
| 256989_at | At3g28580 | AAA-type ATPase | 2.23 |
| 253238_at | At4g34480 | glycosyl hydrolase family 17 protein | 2.23 |
| 252450_s_at | At3g47090 | leucine-rich repeat transmembrane protein kinase, putative | 2.22 |

| | | | |
|-------------|-----------|--|------|
| 245932_at | At5g09290 | 3'(2'),5'-bisphosphate nucleotidase, putative | 2.21 |
| 250625_at | At5g07340 | calnexin, putative | 2.21 |
| 250633_at | At5g07460 | peptide methionine sulfoxide reductase, putative | 2.21 |
| 254914_at | At4g11290 | peroxidase, putative | 2.21 |
| 245196_at | At1g67750 | pectate lyase | 2.21 |
| 251063_at | At5g01850 | protein kinase, putative | 2.21 |
| 265682_at | At2g24390 | avirulence-responsive protein-related | 2.21 |
| 248941_s_at | At5g45460 | expressed protein | 2.21 |
| 246373_at | At1g51860 | leucine-rich repeat protein kinase, putative | 2.21 |
| 248021_at | At5g56500 | chaperonin, putative | 2.2 |
| 254630_at | At4g18360 | (S)-2-hydroxy-acid oxidase, peroxisomal, putative | 2.2 |
| 261330_at | At1g44900 | DNA replication licensing factor, putative | 2.2 |
| 262507_at | At1g11330 | S-locus lectin protein kinase | 2.2 |
| 249273_at | At5g41850 | expressed protein | 2.2 |
| 265695_at | At2g24490 | replication protein, putative | 2.2 |
| 252437_at | At3g47380 | invertase | 2.2 |
| 266827_at | At2g22920 | serine carboxypeptidase S10 | 2.2 |
| 262616_at | At1g06620 | 2-oxoglutarate-dependent dioxygenase, putative | 2.19 |
| 256012_at | At1g19250 | flavin-containing monooxygenase (FMO) | 2.19 |
| 259185_at | At3g01550 | triose phosphate/phosphate translocator, putative | 2.19 |
| 252596_at | At3g45330 | lectin protein kinase | 2.19 |
| 267595_at | At2g32990 | glycosyl hydrolase family 9 protein | 2.19 |
| 261979_at | At1g37130 | nitrate reductase 2 (NR2) | 2.18 |
| 252200_at | At3g50280 | transferase | 2.18 |
| 254390_at | At4g21940 | calcium-dependent protein kinase, putative (CDPK) | 2.18 |
| 259105_at | At3g05500 | rubber elongation factor (REF) | 2.18 |
| 247912_at | At5g57480 | AAA-type ATPase | 2.17 |
| 262137_at | At1g77920 | bZIP family transcription factor | 2.17 |
| 250435_at | At5g10380 | zinc finger (C3HC4-type RING finger) | 2.17 |
| 246157_at | At5g20080 | NADH-cytochrome b5 reductase, putative | 2.17 |
| 248861_at | At5g46700 | senescence-associated protein, putative | 2.17 |
| 260553_at | At2g41800 | expressed protein | 2.17 |
| 260405_at | At1g69930 | glutathione S-transferase, putative | 2.17 |
| 262694_at | At1g62790 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 2.17 |
| 245302_at | At4g17695 | myb family transcription factor (KAN3) | 2.16 |
| 263715_at | At2g20570 | golden2-like transcription factor (GLK1) | 2.16 |
| 251668_at | At3g57010 | strictosidine synthase | 2.16 |
| 260902_at | At1g21440 | mutase | 2.15 |
| 260648_at | At1g08050 | zinc finger (C3HC4-type RING finger) | 2.15 |
| 256935_at | At3g22570 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 2.15 |
| 252562_s_at | At3g45930 | histone H4 | 2.14 |
| 257761_at | At3g23090 | expressed protein | 2.14 |
| 249441_at | At5g39730 | avirulence-responsive protein-related | 2.14 |
| 255578_at | At4g01450 | nodulin MtN21 | 2.14 |
| 249188_at | At5g42830 | transferase | 2.14 |

| | | | |
|-------------|-----------|--|------|
| 246272_at | At4g37150 | esterase, putative | 2.14 |
| 248891_at | At5g46280 | DNA replication licensing factor, putative | 2.14 |
| 246948_at | At5g25130 | cytochrome P450 | 2.14 |
| 249979_s_at | At5g18860 | inosine-uridine preferring nucleoside hydrolase | 2.14 |
| 254735_at | At4g13810 | disease resistance | 2.14 |
| 258023_at | At3g19450 | cinnamyl-alcohol dehydrogenase (CAD) | 2.13 |
| 248118_at | At5g55050 | GDSL-motif lipase/hydrolase | 2.13 |
| 258813_at | At3g04060 | no apical meristem (NAM) | 2.13 |
| 254283_s_at | At4g22870 | leucoanthocyanidin dioxygenase, putative | 2.13 |
| 262664_at | At1g13970 | expressed protein | 2.13 |
| 249058_at | At5g44510 | disease resistance protein (TIR-NBS-LRR class), putative | 2.12 |
| 245746_at | At1g51070 | basic helix-loop-helix (bHLH) | 2.12 |
| 247791_at | At5g58710 | peptidyl-prolyl cis-trans isomerase, putative | 2.12 |
| 247586_at | At5g60660 | major intrinsic protein (MIP) | 2.12 |
| 251370_at | At3g60450 | expressed protein | 2.12 |
| 247494_at | At5g61790 | calnexin 1 (CNX1) | 2.12 |
| 245267_at | At4g14060 | major latex protein-related | 2.12 |
| 265260_at | At2g43000 | no apical meristem (NAM) | 2.11 |
| 262666_at | At1g14080 | xyloglucan fucosyltransferase, putative (FUT6) | 2.11 |
| 252058_at | At3g52470 | harpin-induced (HIN1) | 2.11 |
| 254819_at | At4g12500 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 2.1 |
| 260028_at | At1g29980 | expressed protein | 2.1 |
| 245343_at | At4g15830 | expressed protein | 2.1 |
| 259410_at | At1g13340 | expressed protein | 2.1 |
| 253276_at | At4g34050 | caffeoyl-CoA 3-O-methyltransferase, putative | 2.1 |
| 260051_at | At1g78210 | hydrolase, alpha/beta fold | 2.1 |
| 266356_at | At2g32300 | uclacyanin I | 2.1 |
| 263552_x_at | At2g24980 | proline-rich extensin-like | 2.1 |
| 257708_at | At3g13330 | expressed protein | 2.09 |
| 259385_at | At1g13470 | expressed protein | 2.09 |
| 249276_at | At5g41880 | DNA primase small subunit family | 2.09 |
| 260083_at | At1g63220 | C2 domain-containing protein | 2.09 |
| 261285_at | At1g35720 | annexin 1 (ANN1) | 2.09 |
| 264382_at | At2g25110 | MIR domain-containing protein | 2.09 |
| 245956_s_at | At5g28540 | luminal binding protein 1 (BiP-1) | 2.09 |
| 249214_at | At5g42720 | glycosyl hydrolase family 17 protein | 2.09 |
| 260601_at | At1g55910 | metal transporter, putative (ZIP11) | 2.08 |
| 254909_at | At4g11210 | disease resistance-responsive | 2.08 |
| 266780_at | At2g29110 | glutamate receptor (GLR2.8) | 2.08 |
| 264543_at | At1g55780 | heavy-metal-associated domain-containing protein | 2.08 |
| 247293_at | At5g64510 | expressed protein | 2.08 |
| 263628_at | At2g04780 | fasciclin-like arabinogalactan-protein (FLA7) | 2.08 |
| 250083_at | At5g17220 | glutathione S-transferase, putative | 2.08 |
| 252911_at | At4g39510 | cytochrome P450 | 2.08 |
| 267605_at | At2g32920 | thioredoxin | 2.08 |

| | | | |
|-------------|-----------|--|------|
| 264859_at | At1g24280 | glucose-6-phosphate 1-dehydrogenase, putative | 2.08 |
| 251400_at | At3g60420 | expressed protein | 2.07 |
| 248908_at | At5g45800 | leucine-rich repeat transmembrane protein kinase, putative | 2.07 |
| 259534_at | At1g12290 | disease resistance protein (CC-NBS-LRR class), putative | 2.07 |
| 258473_s_at | At3g02610 | acyl- (acyl-carrier-protein) desaturase, putative | 2.07 |
| 247812_at | At5g58390 | peroxidase, putative | 2.07 |
| 254500_at | At4g20110 | vacuolar sorting receptor, putative (AtELP3) | 2.07 |
| 249021_at | At5g44820 | expressed protein | 2.07 |
| 245575_at | At4g14760 | M protein repeat-containing protein | 2.07 |
| 260146_at | At1g52770 | phototropic-responsive NPH3 | 2.07 |
| 266941_at | At2g18980 | peroxidase, putative | 2.07 |
| 246145_at | At5g19880 | peroxidase, putative | 2.07 |
| 247265_at | At5g64550 | loricrin-related | 2.07 |
| 257173_at | At3g23810 | adenosylhomocysteinase, putative | 2.07 |
| 261500_at | At1g28400 | expressed protein | 2.06 |
| 259198_at | At3g03610 | phagocytosis and cell motility protein ELMO1-related | 2.06 |
| 248100_at | At5g55180 | glycosyl hydrolase family 17 protein | 2.06 |
| 249417_at | At5g39670 | calcium-binding EF hand | 2.06 |
| 265853_at | At2g42360 | zinc finger (C3HC4-type RING finger) | 2.06 |
| 261930_at | At1g22440 | alcohol dehydrogenase, putative (ADH) | 2.06 |
| 258160_at | At3g17820 | glutamine synthetase (GS1) | 2.05 |
| 256512_at | At1g33265 | expressed protein | 2.05 |
| 260386_at | At1g74010 | strictosidine synthase | 2.05 |
| 262881_at | At1g64890 | integral membrane transporter | 2.05 |
| 246429_at | At5g17450 | heavy-metal-associated domain-containing protein | 2.05 |
| 259548_at | At1g35260 | Bet v I allergen | 2.05 |
| 261485_at | At1g14360 | UDP-galactose/UDP-glucose transporter, putative | 2.04 |
| 261224_at | At1g20160 | subtilase | 2.04 |
| 254915_s_at | At4g11310 | cysteine proteinase, putative | 2.04 |
| 264577_at | At1g05260 | peroxidase 3 (PER3) | 2.04 |
| 257154_at | At3g27210 | expressed protein | 2.03 |
| 248192_at | At5g54140 | IAA-amino acid hydrolase, putative (ILL3) | 2.03 |
| 267261_at | At2g23120 | expressed protein | 2.03 |
| 261826_at | At1g11580 | pectin methylesterase, putative | 2.03 |
| 260731_at | At1g17500 | haloacid dehalogenase-like hydrolase | 2.03 |
| 252442_at | At3g46940 | deoxyuridine 5'-triphosphate nucleotidohydrolase family | 2.03 |
| 250142_at | At5g14650 | polygalacturonase, putative | 2.03 |
| 250472_at | At5g10210 | expressed protein | 2.02 |
| 261806_at | At1g30510 | ferredoxin-NADP(+) reductase, putative | 2.02 |
| 259489_at | At1g15790 | expressed protein | 2.02 |
| 254392_at | At4g21600 | bifunctional nuclease, putative | 2.02 |
| 256803_at | At3g20960 | cytochrome P450 | 2.02 |
| 250449_at | At5g10830 | embryo-abundant protein-related | 2.02 |
| 267380_at | At2g26170 | thromboxane-A synthase, putative | 2.02 |
| 262133_at | At1g78000 | sulfate transporter (Sultr1:2) | 2.02 |

| | | | |
|-------------|-----------|---|------|
| 260590_at | At1g53310 | phosphoenolpyruvate carboxylase, putative (PPC1) | 2.02 |
| 253227_at | At4g35030 | protein kinase | 2.02 |
| 253480_at | At4g31840 | plastocyanin-like domain-containing protein | 2.01 |
| 265561_s_at | At2g05510 | glycine-rich protein | 2.01 |
| 251503_at | At3g59140 | ABC transporter | 2.01 |
| 246522_at | At5g15830 | bZIP transcription factor | 2.01 |
| 255676_at | At4g00490 | beta-amylase, putative | 2.01 |
| 258047_at | At3g21240 | 4-coumarate-CoA ligase 2 (4CL2) | 2.01 |
| 245175_at | At2g47470 | thioredoxin | 2.01 |
| 254468_at | At4g20460 | NAD-dependent epimerase/dehydratase | 2 |
| 247989_at | At5g56350 | pyruvate kinase, putative | 2 |
| 267026_at | At2g38340 | AP2 domain-containing transcription factor, putative (DRE2B) | 2 |
| 252077_at | At3g51720 | expressed protein | 2 |
| 247056_at | At5g66750 | SNF2 domain-containing protein | 2 |
| 261774_at | At1g76260 | transducin | 2 |
| 249897_at | At5g22550 | expressed protein | 2 |
| 252619_at | At3g45210 | expressed protein | 2 |
| 266613_at | At2g14900 | gibberellin-regulated | 1.99 |
| 266517_at | At2g35120 | glycine cleavage system H protein, mitochondrial, putative | 1.99 |
| 264372_at | At1g11840 | lactoylglutathione lyase, putative | 1.99 |
| 261309_at | At1g48600 | phosphoethanolamine N-methyltransferase 2, putative (NMT2) | 1.99 |
| 259472_at | At1g18910 | zinc finger (C3HC4-type RING finger) | 1.99 |
| 254056_at | At4g25250 | invertase | 1.99 |
| 261868_s_at | At1g11460 | nodulin MTN21 | 1.99 |
| 249055_at | At5g44460 | calcium-binding protein, putative | 1.99 |
| 253609_at | At4g30190 | ATPase 2, plasma membrane-type, putative | 1.99 |
| 262286_at | At1g68585 | expressed protein | 1.99 |
| 247606_at | At5g61000 | replication protein, putative | 1.99 |
| 252712_at | At3g43800 | glutathione S-transferase, putative | 1.99 |
| 259447_s_at | At1g02430 | ADP-ribosylation factor, putative | 1.98 |
| 263584_at | At2g17040 | no apical meristem (NAM) | 1.98 |
| 250644_at | At5g06750 | protein phosphatase (PP2C) | 1.98 |
| 247945_at | At5g57150 | basic helix-loop-helix (bHLH) | 1.98 |
| 245998_at | At5g20830 | sucrose synthase (SUS1) | 1.98 |
| 265334_at | At2g18370 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 1.98 |
| 262806_at | At1g20950 | pyrophosphate-fructose-6-phosphate 1-phosphotransferase-related | 1.98 |
| 260874_at | At1g21390 | expressed protein | 1.98 |
| 267084_at | At2g41180 | sigA-binding protein-related | 1.98 |
| 252537_at | At3g45710 | proton-dependent oligopeptide transport (POT) | 1.98 |
| 265955_at | At2g37280 | ABC transporter | 1.98 |
| 262788_at | At1g10690 | expressed protein | 1.98 |
| 256319_at | At1g35910 | trehalose-6-phosphate phosphatase, putative | 1.98 |
| 257151_at | At3g27200 | plastocyanin-like domain-containing protein | 1.97 |

| | | | |
|-------------|-----------|--|------|
| 263841_at | At2g36870 | xyloglucan:xyloglucosyl transferase, putative | 1.97 |
| 249469_at | At5g39320 | UDP-glucose 6-dehydrogenase, putative | 1.97 |
| 252010_at | At3g52740 | expressed protein | 1.97 |
| 265439_at | At2g21045 | senescence-associated | 1.97 |
| 261016_at | At1g26560 | glycosyl hydrolase 1 protein | 1.97 |
| 267382_at | At2g44300 | lipid transfer protein-related | 1.96 |
| 255065_s_at | At4g08870 | arginase, putative | 1.96 |
| 260076_at | At1g73630 | calcium-binding protein, putative | 1.96 |
| 266139_at | At2g28085 | auxin-responsive | 1.96 |
| 259307_at | At3g05230 | signal peptidase subunit | 1.96 |
| 264061_at | At2g27970 | cyclin-dependent kinase, putative (CDK) | 1.95 |
| 251273_at | At3g61960 | protein kinase | 1.95 |
| 248945_at | At5g45510 | leucine-rich repeat | 1.95 |
| 252984_at | At4g37990 | mannitol dehydrogenase, putative (ELI3-2) | 1.95 |
| 248890_at | At5g46270 | disease resistance protein (TIR-NBS-LRR class), putative | 1.95 |
| 264010_at | At2g21100 | disease resistance-responsive protein-related | 1.95 |
| 263181_at | At1g05720 | selenoprotein | 1.95 |
| 261904_at | At1g65040 | zinc finger (C3HC4-type RING finger) | 1.95 |
| 259018_at | At3g07390 | auxin-induced protein (AIR12) | 1.95 |
| 259596_at | At1g28130 | auxin-responsive GH3 | 1.95 |
| 259009_at | At3g09260 | glycosyl hydrolase family 1 protein | 1.95 |
| 258658_at | At3g09820 | adenosine kinase 1 (ADK1) | 1.95 |
| 249794_at | At5g23530 | expressed protein | 1.95 |
| 258275_at | At3g15760 | expressed protein | 1.95 |
| 248322_at | At5g52760 | heavy-metal-associated domain-containing protein | 1.95 |
| 255240_at | At4g05530 | short-chain dehydrogenase/reductase (SDR) | 1.95 |
| 251633_at | At3g57460 | expressed protein | 1.94 |
| 261729_s_at | At1g47840 | hexokinase, putative | 1.94 |
| 245164_at | At2g33210 | chaperonin, putative | 1.94 |
| 263706_s_at | At1g31180 | 3-isopropylmalate dehydrogenase, chloroplast, putative | 1.94 |
| 245089_at | At2g45290 | transketolase, putative | 1.94 |
| 263576_at | At2g17080 | hypothetical protein | 1.94 |
| 260254_at | At1g74210 | glycerophosphoryl diester phosphodiesterase | 1.94 |
| 259328_at | At3g16440 | jacalin lectin | 1.94 |
| 249894_at | At5g22580 | expressed protein | 1.94 |
| 262396_at | At1g49470 | expressed protein | 1.94 |
| 254521_at | At5g44810 | expressed protein | 1.93 |
| 248674_at | At5g48800 | phototropic-responsive NPH3 | 1.93 |
| 267012_at | At2g39220 | patatin | 1.93 |
| 247030_at | At5g67210 | expressed protein | 1.93 |
| 258941_at | At3g09940 | monodehydroascorbate reductase, putative | 1.93 |
| 254818_at | At4g12470 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 1.93 |
| 262317_at | At2g48140 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 1.93 |
| 256942_at | At3g23290 | expressed protein | 1.93 |
| 245523_at | At4g15910 | drought-induced protein (Di21) | 1.92 |

| | | | |
|-------------|-----------|---|------|
| 245840_at | At1g58420 | expressed protein | 1.92 |
| 258480_at | At3g02640 | expressed protein | 1.92 |
| 254389_s_at | At4g21910 | MATE efflux | 1.92 |
| 257213_at | At3g15020 | malate dehydrogenase (NAD), mitochondrial, putative | 1.92 |
| 264647_at | At1g09090 | respiratory burst oxidase protein B (RbohB) | 1.92 |
| 249848_at | At5g23220 | isochorismatase hydrolase | 1.92 |
| 254932_at | At4g11120 | translation elongation factor Ts (EF-Ts), putative | 1.92 |
| 266566_at | At2g24040 | hydrophobic protein, putative | 1.91 |
| 262756_at | At1g16370 | transporter-related | 1.91 |
| 258089_at | At3g14740 | PHD finger | 1.91 |
| 258646_at | At3g08040 | MATE efflux | 1.91 |
| 254202_at | At4g24140 | hydrolase, alpha/beta fold | 1.91 |
| 253274_at | At4g34200 | D-3-phosphoglycerate dehydrogenase, putative / 3-PGDH, putative | 1.91 |
| 266326_at | At2g46650 | cytochrome b5, putative | 1.91 |
| 246093_at | At5g20550 | oxidoreductase, 2OG-Fe(II) oxygenase | 1.91 |
| 254690_at | At4g17830 | peptidase M20/M25/M40 | 1.91 |
| 257755_at | At3g18760 | ribosomal protein S6 | 1.91 |
| 266203_at | At2g02230 | SKP1 interacting partner 3-related | 1.9 |
| 250234_at | At5g13420 | transaldolase, putative | 1.9 |
| 256053_at | At1g07260 | UDP-glucuronosyl/UDP-glucosyl transferase | 1.9 |
| 262232_at | At1g68600 | expressed protein | 1.9 |
| 265204_at | At2g36650 | expressed protein | 1.9 |
| 249675_at | At5g35940 | jacalin lectin | 1.9 |
| 259758_s_at | At1g77530 | O-methyltransferase family 2 protein | 1.9 |
| 253191_at | At4g35350 | cysteine endopeptidase, papain-type (XCP1) | 1.89 |
| 248298_at | At5g53110 | expressed protein | 1.89 |
| 256425_at | At1g33560 | disease resistance protein (CC-NBS-LRR class), putative | 1.89 |
| 249178_at | At5g42890 | sterol carrier protein 2 (SCP-2) | 1.89 |
| 248092_at | At5g55170 | small ubiquitin-like modifier 3 (SUMO) | 1.89 |
| 252988_at | At4g38410 | dehydrin, putative | 1.89 |
| 256409_at | At1g66620 | seven in absentia (SINA) protein, putative | 1.89 |
| 257814_at | At3g25110 | acyl- (acyl carrier protein) thioesterase | 1.89 |
| 246144_at | At5g20110 | dynein light chain, putative | 1.89 |
| 261660_at | At1g18370 | kinesin motor (NACK1) | 1.89 |
| 258017_at | At3g19370 | expressed protein | 1.89 |
| 253622_at | At4g30560 | cyclic nucleotide-regulated ion channel, putative | 1.89 |
| 257056_at | At3g15350 | glycosyltransferase family 14 protein | 1.89 |
| 264895_at | At1g23100 | 10 kDa chaperonin, putative | 1.89 |
| 260002_at | At1g67940 | ABC transporter | 1.89 |
| 259382_s_at | At3g16430 | jacalin lectin | 1.88 |
| 255308_at | At4g04910 | AAA-type ATPase | 1.88 |
| 254318_at | At4g22530 | embryo-abundant protein-related | 1.88 |
| 258977_s_at | At3g02020 | aspartate kinase, lysine-sensitive | 1.88 |
| 259247_at | At3g07570 | membrane protein, putative | 1.88 |

| | | | |
|-------------|-----------|--|------|
| 261481_at | At1g14260 | zinc finger (C3HC4-type RING finger) | 1.88 |
| 255513_at | At4g02060 | proliferin protein (PRL) | 1.88 |
| 262533_at | At1g17090 | expressed protein | 1.88 |
| 260896_at | At1g29310 | protein transport protein sec61, putative | 1.88 |
| 256018_at | At1g58300 | heme oxygenase, putative | 1.88 |
| 255243_at | At4g05590 | expressed protein | 1.88 |
| 254231_at | At4g23810 | WRKY family transcription factor | 1.88 |
| 263498_at | At2g42610 | expressed protein | 1.88 |
| 247333_at | At5g63600 | flavonol synthase, putative | 1.88 |
| 251336_at | At3g61190 | BON1-associated protein 1 (BAP1) | 1.88 |
| 263227_at | At1g30750 | expressed protein | 1.88 |
| 262913_at | At1g59960 | aldo/keto reductase, putative | 1.87 |
| 253990_at | At4g26160 | thioredoxin | 1.87 |
| 254458_at | At4g21180 | DNAJ heat shock N-terminal domain-containing protein | 1.87 |
| 245750_at | At1g51060 | histone H2A, putative | 1.87 |
| 261530_at | At1g63460 | glutathione peroxidase, putative | 1.87 |
| 257522_at | At3g08990 | yippee | 1.87 |
| 266761_at | At2g47130 | short-chain dehydrogenase/reductase (SDR) | 1.87 |
| 254346_at | At4g21980 | autophagy 8a (APG8a) | 1.87 |
| 252179_at | At3g50760 | glycosyl transferase family 8 protein | 1.87 |
| 258341_at | At3g22790 | kinase interacting | 1.87 |
| 265338_at | At2g18400 | ribosomal protein L6 | 1.87 |
| 256451_s_at | At1g75170 | SEC14 cytosolic factor | 1.87 |
| 253866_at | At4g27480 | glycosyltransferase family 14 protein | 1.87 |
| 250978_at | At5g03080 | phosphatidic acid phosphatase-related | 1.87 |
| 251487_at | At3g59760 | cysteine synthase, mitochondrial, putative | 1.87 |
| 245052_at | At2g26440 | pectinesterase | 1.87 |
| 262181_at | At1g78060 | glycosyl hydrolase family 3 protein | 1.87 |
| 258768_at | At3g10880 | hypothetical protein | 1.87 |
| 266479_at | At2g31160 | expressed protein | 1.87 |
| 246366_at | At1g51850 | leucine-rich repeat protein kinase, putative | 1.87 |
| 246620_at | At5g36220 | cytochrome P450 81D1 (CYP81D1) | 1.86 |
| 262177_at | At1g74710 | isochorismate synthase 1 (ICS1) | 1.86 |
| 248932_at | At5g46050 | proton-dependent oligopeptide transport (POT) | 1.86 |
| 263047_at | At2g17630 | phosphoserine aminotransferase, putative | 1.86 |
| 252587_at | At3g45620 | transducin | 1.86 |
| 256890_at | At3g23830 | glycine-rich RNA-binding protein, putative | 1.86 |
| 265243_at | At2g43040 | calmodulin-binding protein | 1.86 |
| 249063_at | At5g44110 | ABC transporter | 1.86 |
| 259424_at | At1g13830 | beta-1,3-glucanase-related | 1.86 |
| 258167_at | At3g21560 | UDP-glucosyltransferase, putative | 1.86 |
| 263134_at | At1g78570 | NAD-dependent epimerase/dehydratase | 1.86 |
| 248664_at | At5g48580 | FK506-binding protein 2-2 (FKBP15-2) | 1.86 |
| 255284_at | At4g04610 | 5'-adenylylsulfate reductase (APR1) | 1.86 |
| 254559_at | At4g19200 | proline-rich | 1.85 |

| | | | |
|-------------|-----------|---|------|
| 253992_at | At4g26060 | expressed protein | 1.85 |
| 251531_at | At3g58550 | protease inhibitor/seed storage/lipid transfer protein (LTP) | 1.85 |
| 251846_at | At3g54560 | histone H2A.F/Z | 1.85 |
| 260009_at | At1g67950 | RNA recognition motif (RRM)-containing protein | 1.85 |
| 259854_at | At1g72200 | zinc finger (C3HC4-type RING finger) | 1.85 |
| 253646_at | At4g29810 | mitogen-activated protein kinase kinase (MAPKK) | 1.85 |
| 249060_at | At5g44560 | SNF7 | 1.85 |
| 252148_at | At3g51280 | male sterility MS5, putative | 1.85 |
| 252155_at | At3g50920 | phosphatidic acid phosphatase-related | 1.85 |
| 249581_at | At5g37600 | glutamine synthetase, putative | 1.85 |
| 264301_at | At1g78780 | pathogenesis-related | 1.85 |
| 263544_at | At2g21590 | glucose-1-phosphate adenylyltransferase large subunit, putative | 1.85 |
| 245293_at | At4g16660 | heat shock protein 70, putative (HSP70) | 1.85 |
| 267076_at | At2g41090 | calmodulin-like calcium-binding protein, 22 kDa (CaBP-22) | 1.85 |
| 254343_at | At4g21990 | 5'-adenylylsulfate reductase (APR3) | 1.85 |
| 265339_at | At2g18230 | inorganic pyrophosphatase (soluble) (PPA) | 1.85 |
| 262658_at | At1g14220 | ribonuclease T2 | 1.85 |
| 258040_at | At3g21190 | expressed protein | 1.85 |
| 258630_at | At3g02820 | zinc knuckle (CCHC-type) | 1.85 |
| 254824_s_at | At4g12620 | replication control protein, putative | 1.85 |
| 252622_at | At3g45310 | cysteine proteinase, putative | 1.84 |
| 251107_at | At5g01610 | expressed protein | 1.84 |
| 249817_at | At5g23820 | MD-2-related lipid recognition domain-containing protein | 1.84 |
| 247255_at | At5g64780 | expressed protein | 1.84 |
| 258655_at | At3g09805 | isocitrate dehydrogenase, putative | 1.84 |
| 261933_at | At1g22410 | 2-dehydro-3-deoxyphosphoheptonate aldolase, putative | 1.84 |
| 256869_at | At3g26430 | glycine-rich RNA-binding protein | 1.84 |
| 265188_at | At1g23800 | aldehyde dehydrogenase, mitochondrial (ALDH3) | 1.84 |
| 257382_at | At2g40750 | WRKY family transcription factor | 1.84 |
| 258710_s_at | At3g09800 | clathrin adaptor complex small chain | 1.84 |
| 259842_at | At1g73600 | phosphoethanolamine N-methyltransferase 3, putative (NMT3) | 1.84 |
| 257798_at | At3g15950 | DNA topoisomerase-related | 1.84 |
| 262704_at | At1g16530 | lateral organ boundaries domain protein 3 (LBD3) | 1.84 |
| 265183_at | At1g23750 | DNA-binding protein-related | 1.83 |
| 252671_at | At3g44190 | pyridine nucleotide-disulphide oxidoreductase | 1.83 |
| 251106_at | At5g01500 | mitochondrial substrate carrier | 1.83 |
| 258102_at | At3g23600 | dienelactone hydrolase | 1.83 |
| 257466_at | At1g62840 | expressed protein | 1.83 |
| 264466_at | At1g10380 | expressed protein | 1.83 |
| 249227_at | At5g42180 | peroxidase 64 (PER64) | 1.83 |
| 247983_at | At5g56630 | phosphofructokinase | 1.83 |
| 254673_at | At4g18430 | Ras-related GTP-binding protein, putative | 1.83 |
| 247575_at | At5g61030 | RNA-binding protein, putative | 1.83 |

| | | | |
|-------------|-----------|--|------|
| 263787_at | At2g46420 | expressed protein | 1.83 |
| 254299_at | At4g22920 | expressed protein | 1.82 |
| 260551_at | At2g43510 | trypsin inhibitor, putative | 1.82 |
| 258437_at | At3g16560 | protein phosphatase 2C-related (PP2C) | 1.82 |
| 262344_at | At1g64060 | respiratory burst oxidase protein F (RbohF) (RbohAp108) | 1.82 |
| 259365_at | At1g13300 | myb family transcription factor | 1.82 |
| 248833_at | At5g47120 | Bax inhibitor-1 putative | 1.82 |
| 254341_at | At4g22130 | protein kinase | 1.82 |
| 261574_at | At1g01190 | cytochrome P450, putative | 1.82 |
| 249274_at | At5g41860 | expressed protein | 1.82 |
| 259850_at | At1g72240 | expressed protein | 1.82 |
| 256673_at | At3g52370 | beta-Ig-H3 domain-containing protein | 1.82 |
| 266334_at | At2g32380 | expressed protein | 1.82 |
| 253515_at | At4g31320 | auxin-responsive protein, putative | 1.82 |
| 266687_at | At2g19670 | protein arginine N-methyltransferase, putative | 1.82 |
| 253720_at | At4g29270 | acid phosphatase class B | 1.82 |
| 246988_at | At5g67340 | armadillo/beta-catenin repeat | 1.82 |
| 245874_at | At1g26250 | proline-rich extensin, putative | 1.82 |
| 248943_s_at | At5g45490 | disease resistance protein-related | 1.81 |
| 258469_at | At3g06050 | alkyl hydroperoxide reductase (AhpC/TSA) | 1.81 |
| 246302_at | At3g51860 | cation exchanger, putative (CAX3) | 1.81 |
| 253177_s_at | At4g35150 | O-methyltransferase family 2 protein | 1.81 |
| 266059_at | At2g40765 | expressed protein | 1.81 |
| 258588_s_at | At3g04120 | glyceraldehyde-3-phosphate dehydrogenase (GAPC) | 1.81 |
| 259575_at | At1g35320 | expressed protein | 1.81 |
| 249552_s_at | At5g38240 | serine/threonine protein kinase, putative | 1.81 |
| 247706_at | At5g59480 | haloacid dehalogenase-like hydrolase | 1.81 |
| 249481_at | At5g38900 | DSBA oxidoreductase | 1.8 |
| 249490_s_at | At5g39110 | germin-like protein, putative | 1.8 |
| 249494_at | At5g39050 | transferase | 1.8 |
| 247656_at | At5g59890 | actin-depolymerizing factor 4 (ADF4) | 1.8 |
| 259939_s_at | At1g71300 | Vps52/Sac2 | 1.8 |
| 249377_at | At5g40690 | expressed protein | 1.8 |
| 249187_at | At5g43060 | cysteine proteinase, putative | 1.8 |
| 250407_at | At5g10870 | chorismate mutase, cytosolic (CM2) | 1.8 |
| 256766_at | At3g22231 | expressed protein | 1.8 |
| 245951_at | At5g19550 | aspartate aminotransferase, cytoplasmic isozyme 1 (ASP2) | 1.8 |
| 248333_at | At5g52390 | photoassimilate-responsive protein, putative | 1.8 |
| 253101_at | At4g37430 | cytochrome P450 81F1 (CYP81F1) | 1.8 |
| 255627_at | At4g00955 | expressed protein | 1.8 |
| 262615_at | At1g13950 | eukaryotic translation initiation factor 5A-1 (eIF-5A-1) | 1.79 |
| 261398_at | At1g79610 | sodium proton exchanger, putative (NHX6) | 1.79 |
| 248615_at | At5g49570 | transglutaminase-like | 1.79 |
| 263797_at | At2g24570 | WRKY family transcription factor | 1.79 |
| 254869_at | At4g11890 | protein kinase | 1.79 |

| | | | |
|-------------|-----------|--|------|
| 254234_at | At4g23680 | major latex protein-related | 1.79 |
| 264355_at | At1g03210 | phenazine biosynthesis PhzC/PhzF | 1.79 |
| 262081_at | At1g59540 | kinesin motor protein-related | 1.79 |
| 248230_at | At5g53830 | VQ motif-containing protein | 1.79 |
| 246600_at | At5g14930 | leaf senescence-associated protein (SAG101) | 1.79 |
| 263640_at | At2g25270 | expressed protein | 1.79 |
| 252691_at | At3g44050 | kinesin motor protein-related | 1.79 |
| 252309_at | At3g49340 | cysteine proteinase, putative | 1.79 |
| 257925_at | At3g23170 | expressed protein | 1.79 |
| 252094_at | At3g51260 | 20S proteasome alpha subunit D (PAD1) | 1.79 |
| 264766_at | At1g61420 | S-locus lectin protein kinase | 1.79 |
| 254227_at | At4g23630 | reticulon (RTNLB1) | 1.79 |
| 249806_at | At5g23850 | expressed protein | 1.79 |
| 246511_at | At5g15490 | UDP-glucose 6-dehydrogenase, putative | 1.79 |
| 260605_at | At2g43780 | expressed protein | 1.79 |
| 250892_at | At5g03760 | glycosyl transferase family 2 protein | 1.79 |
| 252880_at | At4g39730 | lipid-associated | 1.79 |
| 263478_at | At2g31880 | leucine-rich repeat transmembrane protein kinase, putative | 1.78 |
| 258162_at | At3g17810 | dihydroorotate dehydrogenase | 1.78 |
| 253776_at | At4g28390 | ADP, ATP carrier protein, mitochondrial, putative | 1.78 |
| 253779_at | At4g28490 | leucine-rich repeat transmembrane protein kinase, putative | 1.78 |
| 263180_at | At1g05620 | inosine-uridine preferring nucleoside hydrolase | 1.78 |
| 251733_at | At3g56240 | copper chaperone (CCH) | 1.78 |
| 259360_at | At1g13310 | expressed protein | 1.78 |
| 264022_at | At2g21185 | expressed protein | 1.78 |
| 252827_at | At4g39950 | cytochrome P450 79B2, putative (CYP79B2) | 1.78 |
| 250236_at | At5g13450 | ATP synthase delta chain, mitochondrial, putative | 1.78 |
| 262324_at | At1g64170 | cation/hydrogen exchanger, putative (CHX16) | 1.78 |
| 266226_at | At2g28740 | histone H4 | 1.78 |
| 259096_at | At3g04840 | 40S ribosomal protein S3A (RPS3aA) | 1.78 |
| 265920_s_at | At2g15120 | secretory protein, putative | 1.78 |
| 261129_at | At1g04820 | tubulin alpha-2/alpha-4 chain (TUA4) | 1.78 |
| 255546_at | At4g01910 | DC1 domain-containing protein | 1.78 |
| 258105_at | At3g23605 | UBX domain-containing protein | 1.78 |
| 248236_at | At5g53870 | plastocyanin-like domain-containing protein | 1.78 |
| 247577_at | At5g61290 | flavin-containing monooxygenase (FMO) | 1.78 |
| 258984_at | At3g08970 | DNAJ heat shock N-terminal domain-containing protein | 1.78 |
| 248512_at | At5g50460 | protein transport protein SEC61 gamma subunit, putative | 1.77 |
| 258288_at | At3g23295 | expressed protein | 1.77 |
| 247522_at | At5g61340 | expressed protein | 1.77 |
| 260506_at | At1g47210 | cyclin | 1.77 |
| 249152_s_at | At5g43370 | inorganic phosphate transporter (PHT1) | 1.77 |
| 266312_at | At2g27020 | 20S proteasome alpha subunit G (PAG1) | 1.77 |
| 263845_at | At2g37040 | phenylalanine ammonia-lyase 1 (PAL1) | 1.77 |
| 263441_at | At2g28620 | kinesin motor protein-related | 1.77 |

| | | | |
|-------------|-----------|--|------|
| 258259_s_at | At3g26840 | esterase/lipase/thioesterase | 1.77 |
| 247024_at | At5g66985 | expressed protein | 1.77 |
| 247794_at | At5g58670 | phosphoinositide-specific phospholipase C (PLC1) | 1.77 |
| 263183_at | At1g05570 | callose synthase 1 (CALS1) | 1.77 |
| 250431_at | At5g10440 | cyclin | 1.77 |
| 253088_at | At4g36220 | cytochrome P450 84A1 (CYP84A1) | 1.76 |
| 248896_at | At5g46350 | WRKY family transcription factor | 1.76 |
| 254233_at | At4g23800 | high mobility group (HMG1/2) | 1.76 |
| 257277_at | At3g14470 | disease resistance protein (NBS-LRR class), putative | 1.76 |
| 250540_at | At5g08580 | calcium-binding EF hand | 1.76 |
| 250365_at | At5g11410 | protein kinase | 1.76 |
| 259604_at | At1g56450 | 20S proteasome beta subunit G1 (PBG1) | 1.76 |
| 266419_at | At2g38760 | annexin 3 (ANN3) | 1.76 |
| 263612_at | At2g16440 | DNA replication licensing factor, putative | 1.76 |
| 249726_at | At5g35480 | expressed protein | 1.76 |
| 250090_at | At5g17330 | glutamate decarboxylase 1 (GAD 1) | 1.76 |
| 246645_at | At5g35080 | expressed protein | 1.75 |
| 257026_at | At3g19200 | hypothetical protein | 1.75 |
| 259383_at | At3g16470 | jacalin lectin | 1.75 |
| 257334_at | orf111d | orf111d | 1.75 |
| 255547_at | At4g01920 | DC1 domain-containing protein | 1.75 |
| 247776_at | At5g58700 | phosphoinositide-specific phospholipase C | 1.75 |
| 266297_at | At2g29570 | proliferating cell nuclear antigen 2 (PCNA2) | 1.75 |
| 260683_at | At1g17560 | ribosomal protein L14 | 1.75 |
| 264903_at | At1g23190 | phosphoglucomutase, putative | 1.75 |
| 252068_at | At3g51440 | strictosidine synthase | 1.75 |
| 251062_at | At5g01840 | ovate | 1.75 |
| 245324_at | At4g17260 | L-lactate dehydrogenase, putative | 1.75 |
| 260325_at | At1g63940 | monodehydroascorbate reductase, putative | 1.75 |
| 267592_at | At2g39710 | aspartyl protease | 1.74 |
| 245736_at | At1g73330 | protease inhibitor, putative (DR4) | 1.74 |
| 264802_at | At1g08560 | syntaxin-related protein KNOLLE (KN) | 1.74 |
| 264969_at | At1g67320 | DNA primase, large subunit family | 1.74 |
| 251644_at | At3g57540 | remorin | 1.74 |
| 256150_at | At1g55120 | beta-fructosidase, putative | 1.74 |
| 265935_at | At2g19580 | senescence-associated protein-related | 1.74 |
| 249397_at | At5g40230 | nodulin-related | 1.74 |
| 250965_at | At5g03020 | kelch repeat-containing F-box | 1.74 |
| 250901_at | At5g03530 | Ras-related GTP-binding | 1.74 |
| 247962_at | At5g56580 | mitogen-activated protein kinase kinase (MKK6) | 1.74 |
| 262482_at | At1g17020 | oxidoreductase, 2OG-Fe(II) oxygenase | 1.74 |
| 262109_at | At1g02730 | cellulose synthase | 1.74 |
| 249809_at | At5g23910 | kinesin motor protein-related | 1.74 |
| 258890_at | At3g05690 | CCAAT-binding transcription factor (CBF-B/NF-YA) | 1.74 |
| 257678_at | At3g20420 | ribonuclease III | 1.74 |

| | | | |
|-------------|-----------|---|------|
| 247570_at | At5g61250 | glycosyl hydrolase family 79 N-terminal domain-containing protein | 1.74 |
| 265717_at | At2g03510 | band 7 | 1.74 |
| 259366_at | At1g13280 | allene oxide cyclase | 1.74 |
| 264592_at | At2g17720 | oxidoreductase, 2OG-Fe(II) oxygenase | 1.73 |
| 249374_at | At5g40580 | 20S proteasome beta subunit B (PBB2) (PRCFC) | 1.73 |
| 262942_at | At1g79450 | LEM3 (ligand-effect modulator 3) | 1.73 |
| 267175_s_at | At2g37620 | actin 3 (ACT3) | 1.73 |
| 253600_at | At4g30810 | serine carboxypeptidase S10 | 1.73 |
| 248967_at | At5g45350 | proline-rich | 1.73 |
| 255773_at | At1g18590 | sulfotransferase | 1.73 |
| 251108_at | At5g01620 | expressed protein | 1.73 |
| 258666_at | At3g08550 | elongation defective 1 protein | 1.73 |
| 260503_at | At1g47250 | 20S proteasome alpha subunit F2 (PAF2) | 1.73 |
| 252114_at | At3g51450 | strictosidine synthase | 1.73 |
| 259669_at | At1g52340 | short-chain dehydrogenase/reductase (SDR) | 1.73 |
| 261599_at | At1g49700 | expressed protein | 1.73 |
| 257021_at | At3g19710 | branched-chain amino acid transaminase, putative (BCAT4) | 1.73 |
| 266170_at | At2g39050 | hydroxyproline-rich glycoprotein | 1.73 |
| 254512_at | At4g20230 | terpene synthase | 1.73 |
| 265435_s_at | At2g21020 | major intrinsic protein(MIP) | 1.73 |
| 267165_at | At2g37710 | lectin protein kinase, putative | 1.72 |
| 264622_at | At2g17790 | vacuolar protein sorting-associated protein 35 (VPS35) | 1.72 |
| 254351_at | At4g22300 | phospholipase | 1.72 |
| 262619_at | At1g06550 | enoyl-CoA hydratase | 1.72 |
| 264377_at | At2g25060 | plastocyanin-like domain-containing protein | 1.72 |
| 264219_at | At1g60420 | DC1 domain-containing protein | 1.72 |
| 261368_at | At1g53070 | legume lectin | 1.72 |
| 253629_at | At4g30450 | glycine-rich protein | 1.72 |
| 247354_at | At5g63590 | flavonol synthase, putative | 1.72 |
| 264574_at | At1g05300 | metal transporter, putative (ZIP5) | 1.72 |
| 253105_at | At4g35840 | zinc finger (C3HC4-type RING finger) | 1.72 |
| 263892_at | At2g36890 | myb family transcription factor (MYB38) | 1.72 |
| 267428_at | At2g34840 | coatamer protein epsilon subunit (COPE) | 1.72 |
| 265097_at | At1g04020 | zinc finger (C3HC4-type RING finger) | 1.72 |
| 259678_at | At1g77750 | 30S ribosomal protein S13, chloroplast, putative | 1.72 |
| 247215_at | At5g64905 | expressed protein | 1.72 |
| 247094_at | At5g66280 | GDP-D-mannose 4,6-dehydratase, putative | 1.72 |
| 256529_at | At1g33260 | protein kinase | 1.72 |
| 256169_at | At1g51800 | leucine-rich repeat protein kinase, putative | 1.71 |
| 262826_at | At1g13080 | cytochrome P450 | 1.71 |
| 247999_at | At5g56150 | ubiquitin-conjugating enzyme, putative | 1.71 |
| 258913_at | At3g06450 | anion exchange | 1.71 |
| 266625_at | At2g35380 | peroxidase 20 (PER20) | 1.71 |
| 246593_at | At5g14790 | expressed protein | 1.71 |

| | | | |
|-------------|-----------|--|------|
| 254761_at | At4g13195 | expressed protein | 1.71 |
| 248528_at | At5g50760 | auxin-responsive | 1.71 |
| 257377_at | At2g28890 | protein phosphatase 2C (PP2C) | 1.71 |
| 256969_at | At3g21080 | ABC transporter-related | 1.71 |
| 248715_at | At5g48290 | heavy-metal-associated domain-containing protein | 1.71 |
| 247190_at | At5g65420 | cyclin, putative | 1.71 |
| 264458_at | At1g10410 | expressed protein | 1.7 |
| 246993_at | At5g67450 | zinc finger (C2H2 type) protein 1 (AZF1) | 1.7 |
| 252606_at | At3g45010 | serine carboxypeptidase III, putative | 1.7 |
| 262644_at | At1g62710 | vacuolar processing enzyme beta | 1.7 |
| 255302_at | At4g04830 | methionine sulfoxide reductase domain-containing protein | 1.7 |
| 263582_at | At2g17120 | peptidoglycan-binding LysM domain-containing protein | 1.7 |
| 253203_at | At4g34710 | arginine decarboxylase 2 (SPE2) | 1.7 |
| 252677_at | At3g44320 | nitrilase 3 (NIT3) | 1.7 |
| 267439_at | At2g19060 | GDSL-motif lipase/hydrolase | 1.7 |
| 266418_at | At2g38750 | annexin 4 (ANN4) | 1.7 |
| 246092_at | At5g20500 | glutaredoxin, putative | 1.7 |
| 256875_at | At3g26330 | cytochrome P450 | 1.7 |
| 246203_at | At4g36610 | hydrolase, alpha/beta fold | 1.7 |
| 245253_at | At4g15440 | hydroperoxide lyase (HPL1) | 1.7 |
| 258371_at | At3g14410 | transporter-related | 1.7 |
| 266928_at | At2g45790 | eukaryotic phosphomannomutase | 1.7 |
| 259102_at | At3g11660 | harpin-induced (HIN1) | 1.7 |
| 260030_at | At1g68880 | bZIP transcription factor | 1.7 |
| 259735_at | At1g64405 | expressed protein | 1.7 |
| 253361_at | At4g33100 | expressed protein | 1.69 |
| 259937_s_at | At1g71330 | ABC transporter | 1.69 |
| 254850_at | At4g12000 | expressed protein | 1.69 |
| 259671_at | At1g52290 | protein kinase | 1.69 |
| 259928_at | At1g34380 | 5'-3' exonuclease | 1.69 |
| 257365_x_at | At2g26020 | plant defensin-fusion protein, putative (PDF1.2b) | 1.69 |
| 264028_at | At2g03680 | expressed protein | 1.69 |
| 254380_at | At4g21865 | expressed protein | 1.69 |
| 245200_at | At1g67850 | expressed protein | 1.69 |
| 249659_s_at | At5g36710 | expressed protein | 1.69 |
| 256131_at | At1g13600 | bZIP transcription factor | 1.69 |
| 249011_at | At5g44670 | expressed protein | 1.69 |
| 263431_at | At2g22170 | lipid-associated | 1.69 |
| 262161_at | At1g52600 | signal peptidase, putative | 1.69 |
| 266244_at | At2g27740 | expressed protein | 1.69 |
| 265740_at | At2g01150 | zinc finger (C3HC4-type RING finger) | 1.69 |
| 259852_at | At1g72280 | endoplasmic reticulum oxidoreductin 1 (ERO1) | 1.69 |
| 253172_at | At4g35060 | heavy-metal-associated domain-containing protein | 1.69 |
| 252188_at | At3g50860 | clathrin adaptor complex small chain | 1.69 |
| 254093_at | At4g25110 | latex-abundant (AMC2) | 1.69 |

| | | | |
|-------------|-----------|--|------|
| 250129_at | At5g16450 | dimethylmenaquinone methyltransferase | 1.69 |
| 250493_at | At5g09800 | U-box domain-containing protein | 1.69 |
| 251393_at | At3g60640 | autophagy 8g (APG8g) | 1.68 |
| 257535_at | At3g09490 | chloroplast lumen common | 1.68 |
| 247602_at | At5g60900 | lectin protein kinase | 1.68 |
| 262039_at | At1g80050 | adenine phosphoribosyltransferase 2 (APT2) | 1.68 |
| 263406_at | At2g04160 | subtilisin-like protease (AIR3) | 1.68 |
| 255259_at | At4g05020 | NADH dehydrogenase-related | 1.68 |
| 259104_at | At3g02170 | expressed protein | 1.68 |
| 249385_at | At5g39950 | thioredoxin H-type 2 (TRX-H-2) (Gif2) | 1.68 |
| 265538_at | At2g15860 | expressed protein | 1.68 |
| 252692_at | At3g43960 | cysteine proteinase, putative | 1.68 |
| 260225_at | At1g74590 | glutathione S-transferase, putative | 1.68 |
| 259561_at | At1g21250 | wall-associated kinase 1 (WAK1) | 1.68 |
| 264001_at | At2g22420 | peroxidase 17 (PER17) | 1.68 |
| 255879_at | At1g67000 | protein kinase | 1.68 |
| 264635_at | At1g65500 | expressed protein | 1.68 |
| 247264_at | At5g64530 | no apical meristem (NAM) | 1.68 |
| 249777_at | At5g24210 | lipase class 3 | 1.67 |
| 252652_at | At3g44720 | prephenate dehydratase | 1.67 |
| 250299_at | At5g11910 | esterase | 1.67 |
| 255629_at | At4g00860 | stress-related ozone-induced protein (OZ1) | 1.67 |
| 256899_at | At3g24660 | leucine-rich repeat transmembrane protein kinase, putative | 1.67 |
| 255319_at | At4g04220 | disease resistance | 1.67 |
| 261410_at | At1g07610 | metallothionein-like protein 1C (MT-1C) | 1.67 |
| 253340_s_at | At4g33260 | WD-40 repeat (FZR) | 1.67 |
| 263496_at | At2g42570 | expressed protein | 1.67 |
| 248049_at | At5g56090 | cytochrome oxidase assembly | 1.67 |
| 267264_at | At2g22970 | serine carboxypeptidase S10 | 1.67 |
| 264685_at | At1g65610 | endo-1,4-beta-glucanase, putative | 1.67 |
| 262180_at | At1g78050 | phosphoglycerate/bisphosphoglycerate mutase | 1.67 |
| 248972_at | At5g45010 | DSS1/SEM1 | 1.67 |
| 245791_at | At1g32210 | defender against cell death 1 (DAD1) | 1.67 |
| 259891_at | At1g72730 | eukaryotic translation initiation factor 4A, putative (eIF-4A) | 1.67 |
| 259169_at | At3g03520 | phosphoesterase | 1.67 |
| 251761_at | At3g55700 | UDP-glucuronosyl/UDP-glucosyl transferase | 1.67 |
| 248320_at | At5g52720 | heavy-metal-associated domain-containing protein | 1.67 |
| 262131_at | At1g02900 | rapid alkalization factor (RALF) | 1.67 |
| 256427_at | At3g11090 | lateral organ boundaries domain (LBD21) | 1.67 |
| 247692_s_at | At5g59690 | histone H4 | 1.66 |
| 251222_at | At3g62580 | expressed protein | 1.66 |
| 262744_at | At1g28680 | transferase | 1.66 |
| 262426_s_at | At1g47630 | expressed protein | 1.66 |
| 265383_at | At2g16780 | WD-40 repeat protein (MSI2) | 1.66 |
| 265214_at | At1g05000 | tyrosine specific protein phosphatase | 1.66 |

| | | | |
|-------------|-----------|--|------|
| 249514_at | At5g38480 | 14-3-3 protein GF14 psi (GRF3) | 1.66 |
| 252291_s_at | At3g49120 | peroxidase 33 (PER33) | 1.66 |
| 247448_at | At5g62770 | expressed protein | 1.66 |
| 247338_at | At5g63680 | pyruvate kinase, putative | 1.66 |
| 252324_at | At3g48565 | protein transport protein SEC61 gamma subunit, putative | 1.66 |
| 250316_at | At5g12140 | cysteine protease inhibitor, putative | 1.66 |
| 259936_at | At1g71220 | UDP-glucose:glycoprotein glucosyltransferase, putative | 1.66 |
| 258332_at | At3g16180 | proton-dependent oligopeptide transport (POT) | 1.66 |
| 245395_at | At4g17420 | expressed protein | 1.66 |
| 251661_at | At3g56950 | small basic membrane integral | 1.66 |
| 246073_at | At5g20180 | ribosomal protein L36 | 1.66 |
| 246714_at | At5g28220 | expressed protein | 1.66 |
| 259563_s_at | At1g20590 | cyclin, putative | 1.66 |
| 252022_at | At3g52930 | fructose-bisphosphate aldolase, putative | 1.66 |
| 248732_at | At5g48070 | xyloglucan:xyloglucosyl transferase, putative | 1.65 |
| 259293_at | At3g11580 | DNA-binding protein, putative | 1.65 |
| 257705_at | At3g12760 | expressed protein | 1.65 |
| 256883_at | At3g26440 | expressed protein | 1.65 |
| 266757_at | At2g46940 | expressed protein | 1.65 |
| 265130_at | At1g30890 | integral membrane HRF1 | 1.65 |
| 247740_at | At5g58940 | protein kinase | 1.65 |
| 250212_at | At5g13960 | SET domain-containing protein (SUVH4) | 1.65 |
| 266820_at | At2g44940 | AP2 domain-containing transcription factor TINY, putative | 1.65 |
| 264439_at | At1g27450 | adenine phosphoribosyltransferase 1 (APT1) | 1.65 |
| 261415_at | At1g07750 | cupin | 1.65 |
| 257436_s_at | At2g24650 | transcriptional factor B3 | 1.65 |
| 262711_at | At1g16500 | expressed protein | 1.65 |
| 257643_at | At3g25730 | AP2 domain-containing transcription factor, putative | 1.65 |
| 253382_at | At4g33040 | glutaredoxin | 1.65 |
| 266037_at | At2g05940 | protein kinase, putative | 1.65 |
| 267518_at | At2g30500 | kinase interacting | 1.65 |
| 258270_at | At3g15650 | phospholipase | 1.65 |
| 248509_at | At5g50335 | expressed protein | 1.65 |
| 248951_at | At5g45550 | mob1/phocein | 1.65 |
| 263179_at | At1g05710 | ethylene-responsive protein, putative | 1.65 |
| 266401_s_at | At2g38620 | cell division control protein, putative | 1.65 |
| 262452_at | At1g11210 | expressed protein | 1.65 |
| 252549_at | At3g45860 | receptor-like protein kinase, putative | 1.65 |
| 259911_at | At1g72680 | cinnamyl-alcohol dehydrogenase, putative (CAD) | 1.64 |
| 261339_at | At1g35710 | leucine-rich repeat transmembrane protein kinase, putative | 1.64 |
| 260933_at | At1g02470 | expressed protein | 1.64 |
| 248942_at | At5g45480 | expressed protein | 1.64 |
| 248144_at | At5g54800 | glucose-6-phosphate/phosphate translocator, putative | 1.64 |
| 254759_at | At4g13180 | short-chain dehydrogenase/reductase (SDR) | 1.64 |
| 264669_at | At1g09630 | Ras-related GTP-binding protein, putative | 1.64 |

| | | | |
|-----------|-----------|--|------|
| 255499_at | At4g02730 | transducin | 1.64 |
| 245301_at | At4g17190 | farnesyl pyrophosphate synthetase 2 (FPS2) | 1.64 |
| 265510_at | At2g05630 | APG8d: autophagy 8d (APG8d) | 1.64 |
| 265539_at | At2g15830 | expressed protein | 1.64 |
| 263120_at | At1g78490 | cytochrome P450 | 1.64 |
| 255726_at | At1g25530 | lysine and histidine specific transporter, putative | 1.64 |
| 258982_at | At3g08870 | lectin protein kinase, putative | 1.64 |
| 261258_at | At1g26640 | aspartate | 1.64 |
| 256426_at | At1g33420 | PHD finger | 1.64 |
| 253148_at | At4g35620 | cyc2b: cyclin 2b (CYC2b) | 1.64 |
| 265382_at | At2g16790 | shikimate kinase | 1.64 |
| 249092_at | At5g43710 | glycoside hydrolase family 47 protein | 1.64 |
| 264091_at | At1g79110 | expressed protein | 1.64 |
| 258898_at | At3g05740 | DNA helicase (RECQ1) | 1.64 |
| 250560_at | At5g08020 | replication protein, putative | 1.64 |
| 266802_at | At2g22900 | galactosyl transferase (GMA12) | 1.64 |
| 258487_at | At3g02550 | lateral organ boundaries domain protein 41 (LBD41) | 1.63 |
| 262408_at | At1g34750 | protein phosphatase 2C, putative (PP2C) | 1.63 |
| 245394_at | At4g16450 | expressed protein | 1.63 |
| 261177_at | At1g04770 | male sterility MS5 | 1.63 |
| 249974_at | At5g18780 | F-box | 1.63 |
| 267212_at | At2g44060 | late embryogenesis abundant (LEA) | 1.63 |
| 260738_at | At1g14990 | expressed protein | 1.63 |
| 256237_at | At3g12610 | DNA-damage-repair (DRT100) | 1.63 |
| 246281_at | At4g36940 | nicotinate phosphoribosyltransferase | 1.63 |
| 257724_at | At3g18510 | expressed protein | 1.63 |
| 254687_at | At4g13770 | cytochrome P450 | 1.63 |
| 246320_at | At1g16560 | Per1-like | 1.63 |
| 249644_at | At5g37010 | expressed protein | 1.63 |
| 257711_at | At3g27430 | 20S proteasome beta subunit B (PBB1) | 1.63 |
| 253999_at | At4g26200 | 1-aminocyclopropane-1-carboxylate synthase, putative | 1.63 |
| 260728_at | At1g48210 | serine/threonine protein kinase, putative | 1.63 |
| 259429_at | At1g01600 | cytochrome P450, putative | 1.63 |
| 258362_at | At3g14280 | expressed protein | 1.63 |
| 251705_at | At3g56400 | WRKY family transcription factor | 1.63 |
| 260023_at | At1g30040 | gibberellin 2-oxidase (GA2OX2) | 1.63 |
| 258067_at | At3g25980 | mitotic spindle checkpoint protein, putative (MAD2) | 1.63 |
| 248426_at | At5g51740 | peptidase M48 | 1.62 |
| 251337_at | At3g60820 | 20S proteasome beta subunit F1 (PBF1) | 1.62 |
| 261504_at | At1g71692 | MADS-box protein (AGL12) | 1.62 |
| 249025_at | At5g44720 | molybdenum cofactor sulfuryase | 1.62 |
| 251886_at | At3g54260 | expressed protein | 1.62 |
| 247275_at | At5g64370 | beta-ureidopropionase, putative | 1.62 |
| 267618_at | At2g26760 | cyclin, putative | 1.62 |
| 251689_at | At3g56500 | serine-rich protein-related | 1.62 |

| | | | |
|-------------|-----------|---|------|
| 248147_at | At5g54900 | RNA-binding protein 45 (RBP45), putative | 1.62 |
| 264588_at | At2g17730 | zinc finger (C3HC4-type RING finger) | 1.62 |
| 252572_at | At3g45290 | seven transmembrane MLO (MLO3) | 1.62 |
| 256415_at | At3g11210 | GDSSL-motif lipase/hydrolase | 1.62 |
| 245160_at | At2g33080 | leucine-rich repeat | 1.62 |
| 257176_s_at | At3g23510 | cyclopropane fatty acid synthase, putative (CPA-FA) | 1.62 |
| 260262_at | At1g68470 | exostosin | 1.62 |
| 259264_at | At3g01260 | aldose 1-epimerase | 1.62 |
| 255566_s_at | At4g01780 | XH/XS domain-containing protein | 1.62 |
| 253712_at | At4g29330 | Der1-like | 1.61 |
| 254833_s_at | At4g12280 | copper amine oxidase, putative | 1.61 |
| 258295_at | At3g23400 | plastid-lipid associated protein (PAP) | 1.61 |
| 254355_at | At4g22380 | ribosomal protein L7Ae/L30e/S12e/Gadd45 | 1.61 |
| 266639_at | At2g35520 | defender against cell death 2 (DAD2) | 1.61 |
| 248829_at | At5g47130 | Bax inhibitor-1 | 1.61 |
| 248200_at | At5g54160 | 5-hydroxyferulic acid O-methyltransferase (OMT1) | 1.61 |
| 247077_at | At5g66420 | expressed protein | 1.61 |
| 262096_at | At1g56010 | transcription activator NAC1 (NAC1) | 1.61 |
| 261153_at | At1g04850 | ubiquitin-associated (UBA) | 1.61 |
| 246920_at | At5g25090 | plastocyanin-like domain-containing protein | 1.61 |
| 265913_at | At2g25625 | expressed protein | 1.61 |
| 263776_s_at | At2g46440 | cyclic nucleotide-gated channel (CNGC3) | 1.61 |
| 249801_at | At5g23580 | calcium-dependent protein kinase 9 (CDPK9) | 1.61 |
| 264089_at | At2g31200 | actin-depolymerizing factor 6 (ADF6) | 1.61 |
| 251229_at | At3g62740 | glycosyl hydrolase family 1 protein | 1.61 |
| 246478_at | At5g15980 | pentatricopeptide (PPR) repeat-containing protein | 1.61 |
| 259680_at | At1g77690 | amino acid permease, putative | 1.61 |
| 246580_at | At1g31770 | ABC transporter | 1.61 |
| 258837_at | At3g07110 | 60S ribosomal protein L13A (RPL13aA) | 1.61 |
| 252313_at | At3g49390 | RNA-binding protein, putative | 1.6 |
| 248286_at | At5g52870 | expressed protein | 1.6 |
| 245292_at | At4g15093 | catalytic LigB subunit of aromatic ring-opening dioxygenase | 1.6 |
| 254262_at | At4g23480 | hydroxyproline-rich glycoprotein | 1.6 |
| 266873_at | At2g44740 | cyclin | 1.6 |
| 253987_at | At4g26270 | phosphofructokinase | 1.6 |
| 260908_at | At1g02580 | maternal embryogenesis control protein (MEA) | 1.6 |
| 267237_s_at | At2g44040 | dihydrodipicolinate reductase | 1.6 |
| 262444_at | At1g47480 | expressed protein | 1.6 |
| 257701_at | At3g12710 | methylenadenine glycosylase | 1.6 |
| 256110_at | At1g16900 | curculin-like (mannose-binding) lectin | 1.6 |
| 267600_at | At2g33040 | ATP synthase gamma chain, mitochondrial (ATPC) | 1.6 |
| 263284_at | At2g36100 | integral membrane | 1.6 |
| 261025_at | At1g01225 | NC domain-containing protein-related | 1.6 |
| 260684_at | At1g17590 | CCAAT-binding transcription factor (CBF-B/NF-YA) | 1.6 |
| 256322_at | At1g54990 | expressed protein | 1.6 |

| | | | |
|-----------|-----------|---|------|
| 253957_at | At4g26320 | arabinogalactan-protein (AGP13) | 1.6 |
| 245197_at | At1g67800 | copine-related | 1.6 |
| 266234_at | At2g02350 | SKIP3: SKP1 interacting partner 3 (SKIP3) | 1.6 |
| 252763_at | At3g42725 | expressed protein | 1.6 |
| 261794_at | At1g16060 | ovule development protein, putative | 1.6 |
| 256617_at | At3g22240 | expressed protein | 1.6 |
| 254333_at | At4g22753 | sterol desaturase | 1.6 |
| 257617_at | At3g26550 | DC1 domain-containing protein | 1.6 |
| 246974_at | At5g24980 | expressed protein | 1.6 |
| 254386_at | At4g21960 | peroxidase 42 (PER42) | 1.6 |
| 262134_at | At1g77990 | sulfate transporter | 1.6 |
| 252527_at | At3g46440 | NAD-dependent epimerase/dehydratase | 1.59 |
| 247469_at | At5g62165 | MADS-box protein (AGL42) | 1.59 |
| 266416_at | At2g38710 | AMMECR1 family | 1.59 |
| 250550_at | At5g07870 | transferase | 1.59 |
| 250324_at | At5g12040 | carbon-nitrogen hydrolase | 1.59 |
| 246968_at | At5g24870 | zinc finger (C3HC4-type RING finger) | 1.59 |
| 257148_at | At3g27240 | cytochrome c1, putative | 1.59 |
| 256255_at | At3g11280 | myb family transcription factor | 1.59 |
| 248835_at | At5g47250 | disease resistance protein (CC-NBS-LRR class), putative | 1.59 |
| 259116_at | At3g01350 | proton-dependent oligopeptide transport (POT) | 1.59 |
| 249465_at | At5g39720 | avirulence-responsive protein-related | 1.59 |
| 263785_at | At2g46390 | expressed protein | 1.59 |
| 259599_at | At1g28110 | serine carboxypeptidase S10 | 1.59 |
| 264581_at | At1g05210 | expressed protein | 1.59 |
| 253804_at | At4g28230 | expressed protein | 1.59 |
| 260530_at | At2g47320 | peptidyl-prolyl cis-trans isomerase cyclophilin-type | 1.59 |
| 254216_at | At4g23710 | vacuolar ATP synthase subunit G 2 (VATG2) | 1.59 |
| 245911_at | At5g19690 | oligosaccharyl transferase STT3 subunit | 1.59 |
| 261835_at | At1g16050 | expressed protein | 1.59 |
| 251535_at | At3g58540 | expressed protein | 1.59 |
| 259701_at | At1g77770 | expressed protein | 1.59 |
| 263595_at | At2g01890 | purple acid phosphatase, putative | 1.59 |
| 259964_at | At1g53680 | glutathione S-transferase, putative | 1.59 |
| 245463_at | At4g17030 | expansin-related | 1.59 |
| 266298_at | At2g29590 | thioesterase | 1.58 |
| 254532_at | At4g19660 | ankyrin repeat | 1.58 |
| 265244_at | At2g43020 | amine oxidase | 1.58 |
| 257072_at | At3g14220 | GDSL-motif lipase | 1.58 |
| 256682_at | At3g52200 | dihydrolipoamide S-acetyltransferase, putative | 1.58 |
| 265170_at | At1g23730 | carbonic anhydrase, putative | 1.58 |
| 246042_at | At5g19440 | cinnamyl-alcohol dehydrogenase, putative (CAD) | 1.58 |
| 260545_at | At2g43350 | glutathione peroxidase, putative | 1.58 |
| 254423_at | At4g21610 | zinc finger protein, putative | 1.58 |
| 254328_at | At4g22570 | adenine phosphoribosyltransferase, putative | 1.58 |

| | | | |
|-------------|-----------|---|------|
| 259520_at | At1g12320 | expressed protein | 1.58 |
| 252328_at | At3g48570 | protein transport protein SEC61 gamma subunit, putative | 1.58 |
| 263758_s_at | At2g21260 | mannose 6-phosphate reductase (M6PR) | 1.58 |
| 260189_at | At1g67550 | urease, putative | 1.58 |
| 257228_at | At3g27890 | NADPH-dependent FMN reductase | 1.58 |
| 252117_at | At3g51430 | strictosidine synthase, putative (YLS2) | 1.58 |
| 259016_at | At3g07480 | expressed protein | 1.58 |
| 257797_at | At3g15940 | glycosyl transferase family 1 protein | 1.58 |
| 253437_at | At4g32460 | expressed protein | 1.58 |
| 252713_at | At3g43810 | calmodulin-7 (CAM7) | 1.58 |
| 262716_at | At1g16470 | 20S proteasome alpha subunit B (PAB1) | 1.58 |
| 258414_at | At3g17380 | meprin and TRAF homology domain-containing protein | 1.58 |
| 248944_at | At5g45500 | expressed protein | 1.57 |
| 267103_at | At2g41490 | dolichol phosphate N-acetylglucosamine-1-phosphate transferase (UDP-GlcNAc) | 1.57 |
| 252373_at | At3g48090 | disease resistance protein (EDS1) | 1.57 |
| 258966_at | At3g10690 | DNA gyrase subunit A | 1.57 |
| 246724_at | At5g29000 | myb family transcription factor | 1.57 |
| 262751_at | At1g16310 | cation efflux | 1.57 |
| 246378_at | At1g57620 | emp24 | 1.57 |
| 257422_at | At1g11940 | expressed protein | 1.57 |
| 267500_s_at | At2g45510 | cytochrome P450 | 1.57 |
| 255633_at | At4g00585 | expressed protein | 1.57 |
| 261436_at | At1g07870 | protein kinase | 1.57 |
| 254242_at | At4g23200 | protein kinase | 1.57 |
| 248597_at | At5g49160 | DNA (cytosine-5-)-methyltransferase (ATHIM) | 1.57 |
| 261221_at | At1g19960 | expressed protein | 1.57 |
| 266036_s_at | At2g05840 | 20S proteasome alpha subunit A1 (PAA1) | 1.57 |
| 247881_at | At5g57700 | BNR/Asp-box repeat | 1.57 |
| 260943_at | At1g45145 | thioredoxin H-type 5 (TRX-H-5) | 1.57 |
| 262575_at | At1g15210 | ABC transporter | 1.57 |
| 254958_at | At4g11010 | nucleoside diphosphate kinase 3, mitochondrial (NDK3) | 1.57 |
| 246029_at | At5g21090 | leucine-rich repeat protein, putative | 1.57 |
| 251992_at | At3g53350 | myosin heavy chain-related | 1.57 |
| 245623_s_at | At4g14103 | F-box | 1.57 |
| 251438_s_at | At3g59930 | expressed protein | 1.56 |
| 257252_at | At3g24170 | glutathione reductase, putative | 1.56 |
| 262237_at | At1g48320 | thioesterase | 1.56 |
| 263553_at | At2g16430 | purple acid phosphatase (PAP10) | 1.56 |
| 257623_at | At3g26210 | cytochrome P450 71B23, putative (CYP71B23) | 1.56 |
| 265341_at | At2g18360 | hydrolase, alpha/beta fold | 1.56 |
| 260156_at | At1g52880 | no apical meristem (NAM) | 1.56 |
| 245832_at | At1g48850 | chorismate synthase, putative | 1.56 |
| 256900_at | At3g24670 | pectate lyase | 1.56 |
| 254416_at | At4g21380 | S-locus protein kinase, putative (ARK3) | 1.56 |

| | | | |
|-------------|-----------|--|------|
| 252833_at | At4g40090 | arabinogalactan-protein (AGP3) | 1.56 |
| 252385_at | At3g47810 | calcineurin-like phosphoesterase | 1.56 |
| 249523_at | At5g38630 | cytochrome B561 | 1.56 |
| 246061_at | At5g19320 | RAN GTPase activating protein 2 (RanGAP2) | 1.56 |
| 249487_at | At5g39040 | ABC transporter (TAP2) | 1.56 |
| 249176_at | At5g42980 | thioredoxin H-type 3 (TRX-H-3) | 1.56 |
| 245873_at | At1g26260 | basic helix-loop-helix (bHLH) | 1.56 |
| 247425_at | At5g62550 | expressed protein | 1.56 |
| 255590_at | At4g01610 | cathepsin B-like cysteine protease, putative | 1.56 |
| 255527_at | At4g02360 | expressed protein | 1.56 |
| 251090_at | At5g01340 | mitochondrial substrate carrier | 1.56 |
| 248800_at | At5g47320 | 30S ribosomal protein S19, mitochondrial (RPS19) | 1.56 |
| 255230_at | At4g05390 | ferredoxin-NADP(+) reductase, putative | 1.56 |
| 252478_at | At3g46540 | clathrin assembly protein-related | 1.56 |
| 256674_at | At3g52360 | expressed protein | 1.56 |
| 261944_at | At1g64650 | expressed protein | 1.56 |
| 257971_at | At3g27530 | vesicle tethering | 1.55 |
| 259255_at | At3g07690 | NAD-dependent glycerol-3-phosphate dehydrogenase | 1.55 |
| 260611_at | At2g43670 | glycosyl hydrolase 17 | 1.55 |
| 264657_at | At1g09100 | 26S protease regulatory subunit 6A, putative | 1.55 |
| 250823_at | At5g05180 | expressed protein | 1.55 |
| 248576_at | At5g49810 | methionine S-methyltransferase | 1.55 |
| 266992_at | At2g39200 | seven transmembrane MLO (MLO12) | 1.55 |
| 255257_at | At4g05050 | polyubiquitin (UBQ11) | 1.55 |
| 267040_at | At2g34300 | dehydration-responsive protein-related | 1.55 |
| 255412_at | At4g02980 | auxin-binding protein 1 (ABP1) | 1.55 |
| 247929_at | At5g57330 | aldose 1-epimerase | 1.55 |
| 262378_at | At1g72830 | CCAAT-binding transcription factor (CBF-B/NF-YA) | 1.55 |
| 262001_at | At1g33790 | jacalin lectin | 1.55 |
| 254109_at | At4g25240 | multi-copper oxidase type I | 1.55 |
| 267181_at | At2g37760 | aldo/keto reductase | 1.55 |
| 255462_at | At4g02940 | oxidoreductase, 2OG-Fe(II) oxygenase | 1.55 |
| 259381_s_at | At3g16390 | jacalin lectin | 1.55 |
| 250423_s_at | At5g10610 | cytochrome P450 | 1.55 |
| 265386_at | At2g20930 | expressed protein | 1.54 |
| 256118_at | At1g16970 | Ku70-like protein | 1.54 |
| 264708_at | At1g09740 | ethylene-responsive protein, putative | 1.54 |
| 249791_at | At5g23810 | amino acid transporter | 1.54 |
| 245428_at | At4g17570 | zinc finger (GATA type) | 1.54 |
| 255554_at | At4g01895 | expressed protein | 1.54 |
| 258188_at | At3g17800 | expressed protein | 1.54 |
| 263681_at | At1g26840 | origin recognition complex subunit 6 (ORC6) | 1.54 |
| 247403_at | At5g62740 | band 7 | 1.54 |
| 249816_at | At5g23880 | cleavage and polyadenylation specificity factor | 1.54 |
| 267419_at | At2g35010 | thioredoxin | 1.54 |

| | | | |
|-----------|-----------|--|------|
| 266926_at | At2g46000 | expressed protein | 1.54 |
| 256184_at | At1g51650 | ATP synthase epsilon chain, mitochondrial | 1.54 |
| 263399_at | At2g31490 | expressed protein | 1.54 |
| 263865_at | At2g36910 | multidrug resistance P-glycoprotein (PGP1) | 1.53 |
| 253967_at | At4g26550 | expressed protein | 1.53 |
| 260257_at | At1g74340 | dolichol phosphate-mannose biosynthesis regulatory protein-related | 1.53 |
| 265649_at | At2g27510 | ferredoxin, putative | 1.53 |
| 265219_at | At2g02050 | NADH-ubiquinone oxidoreductase B18 subunit, putative | 1.53 |
| 257745_at | At3g29240 | expressed protein | 1.53 |
| 246505_at | At5g16250 | expressed protein | 1.53 |
| 263589_at | At2g25280 | expressed protein | 1.53 |
| 258489_at | At3g02520 | 14-3-3 protein GF14 nu (GRF7) | 1.53 |
| 266752_at | At2g47000 | multidrug resistant (MDR) ABC transporter, putative | 1.53 |
| 254375_at | At4g21800 | ATP-binding | 1.53 |
| 262657_at | At1g14210 | ribonuclease T2 | 1.53 |
| 265858_at | At2g01720 | ribophorin I | 1.53 |
| 258072_at | At3g26090 | expressed protein | 1.53 |
| 248769_at | At5g47730 | SEC14 cytosolic factor, putative | 1.53 |
| 266221_at | At2g28760 | NAD-dependent epimerase/dehydratase | 1.53 |
| 249515_at | At5g38530 | tryptophan synthase-related | 1.53 |
| 262277_at | At1g68650 | expressed protein | 1.53 |
| 259832_at | At1g69580 | myb family transcription factor | 1.53 |
| 252870_at | At4g39940 | adenylylsulfate kinase 2 (AKN2) | 1.53 |
| 248288_at | At5g52840 | NADH-ubiquinone oxidoreductase-related | 1.53 |
| 245619_at | At4g13990 | exostosin | 1.53 |
| 254921_at | At4g11300 | expressed protein | 1.52 |
| 262814_at | At1g11660 | heat shock protein, putative | 1.52 |
| 264059_at | At2g31305 | expressed protein | 1.52 |
| 253183_at | At4g35220 | cyclase | 1.52 |
| 247076_at | At5g66510 | bacterial transferase hexapeptide repeat-containing protein | 1.52 |
| 251847_at | At3g54640 | tryptophan synthase, alpha subunit (TSA1) | 1.52 |
| 262105_at | At1g02810 | pectinesterase | 1.52 |
| 250377_at | At5g11560 | PQQ enzyme repeat-containing protein | 1.52 |
| 257005_at | At3g14190 | expressed protein | 1.52 |
| 267153_at | At2g30860 | glutathione S-transferase, putative | 1.52 |
| 249627_at | At5g37510 | NADH-ubiquinone dehydrogenase, mitochondrial, putative | 1.52 |
| 262155_at | At1g52420 | glycosyl transferase family 1 protein | 1.52 |
| 250117_at | At5g16440 | isopentenyl diphosphate:dimethylallyl diphosphate isomerase I (IPP1) | 1.52 |
| 264787_at | At2g17840 | senescence/dehydration-associated protein-related (ERD7) | 1.52 |
| 247359_at | At5g63560 | transferase | 1.51 |
| 261723_at | At1g76200 | expressed protein | 1.51 |
| 257208_at | At3g14910 | expressed protein | 1.51 |
| 265459_at | At2g46540 | expressed protein | 1.51 |

| | | | |
|-------------|-----------|--|------|
| 264523_at | At1g10030 | integral membrane | 1.51 |
| 267239_at | At2g02510 | expressed protein | 1.51 |
| 246422_at | At5g17060 | ADP-ribosylation factor, putative | 1.51 |
| 265868_at | At2g01650 | zinc finger (C2H2 type) | 1.51 |
| 264003_at | At2g22475 | GRAM domain-containing protein | 1.51 |
| 260944_at | At1g45130 | beta-galactosidase, putative | 1.51 |
| 256524_at | At1g66200 | glutamine synthetase, putative | 1.51 |
| 251210_at | At3g62810 | complex 1 | 1.51 |
| 260153_at | At1g52760 | esterase | 1.51 |
| 261086_at | At1g17460 | myb family transcription factor | 1.51 |
| 250994_at | At5g02490 | heat shock cognate 70 kDa protein 2 (HSC70-2) | 1.51 |
| 247527_at | At5g61480 | leucine-rich repeat transmembrane protein kinase, putative | 1.51 |
| 266457_at | At2g22780 | malate dehydrogenase, glyoxysomal, putative | 1.51 |
| 252361_at | At3g48490 | expressed protein | 1.51 |
| 249711_at | At5g35680 | eukaryotic translation initiation factor 1A, putative (eIF-1A) | 1.51 |
| 246214_at | At4g36990 | heat shock factor protein 4 (HSF4) | 1.51 |
| 265065_at | At1g03980 | phytochelatin synthase 2 (PCS2) | 1.51 |
| 256263_at | At3g12290 | tetrahydrofolate dehydrogenase | 1.5 |
| 248787_at | At5g47420 | expressed protein | 1.5 |
| 250448_at | At5g10820 | integral membrane transporter | 1.5 |
| 259256_at | At3g07680 | emp24 | 1.5 |
| 245524_at | At4g15920 | nodulin MtN3 | 1.5 |
| 260852_at | At1g21900 | emp24 | 1.5 |
| 252099_at | At3g51250 | senescence/dehydration-associated protein-related | 1.5 |
| 259529_at | At1g12400 | expressed protein | 1.5 |
| 251269_at | At3g62360 | expressed protein | 1.5 |
| 265104_at | At1g31020 | thioredoxin o (TRXO2) | 1.5 |
| 253277_at | At4g34230 | cinnamyl-alcohol dehydrogenase, putative (CAD) | 1.5 |
| 247634_at | At5g60520 | late embryogenesis abundant protein-related (LEA) | 1.5 |
| 247011_at | At5g67590 | NADH-ubiquinone oxidoreductase-related | 1.5 |
| 252080_at | At3g51670 | SEC14 cytosolic factor | 1.5 |
| 255104_at | At4g08685 | pollen Ole e 1 allergen and extensin | 1.5 |
| 252754_at | At3g43510 | copa-like retrotransposon family | 1.5 |
| 267565_at | At2g30750 | cytochrome P450 71A12, putative (CYP71A12) | 1.5 |
| 264609_at | At1g04530 | expressed protein | 1.5 |
| 250820_at | At5g05160 | leucine-rich repeat transmembrane protein kinase, putative | 1.5 |
| 245341_at | At4g16447 | expressed protein | 1.5 |
| 262255_at | At1g53790 | F-box | 1.5 |
| 252681_at | At3g44350 | no apical meristem (NAM) | 1.5 |
| 247644_s_at | At5g60390 | elongation factor 1-alpha | 1.5 |
| 261139_at | At1g19700 | homeobox-leucine zipper | 1.49 |
| 266413_at | At2g38740 | haloacid dehalogenase-like hydrolase | 1.49 |
| 257193_at | At3g13160 | pentatricopeptide (PPR) repeat-containing protein | 1.49 |
| 247039_at | At5g67270 | microtubule-associated EB1 | 1.49 |
| 257849_at | At3g13060 | expressed protein | 1.49 |

| | | | |
|-------------|-----------|---|------|
| 259824_at | At1g66240 | copper homeostasis factor, putative (CCH) | 1.49 |
| 258563_at | At3g05970 | long-chain acyl-CoA synthetase (LACS6) | 1.49 |
| 250676_at | At5g06320 | harpin-induced (HIN1) | 1.49 |
| 259014_at | At3g07320 | glycosyl hydrolase family 17 protein | 1.49 |
| 255459_at | At4g02890 | polyubiquitin (UBQ14) | 1.49 |
| 256981_at | At3g13380 | leucine-rich repeat | 1.49 |
| 254834_at | At4g12300 | cytochrome P450 | 1.49 |
| 263707_at | At1g09300 | metallopeptidase M24 | 1.49 |
| 248259_at | At5g53330 | expressed protein | 1.48 |
| 257612_at | At3g26600 | armadillo/beta-catenin repeat | 1.48 |
| 258108_at | At3g23570 | dienelactone hydrolase | 1.48 |
| 251186_at | At3g62790 | NADH-ubiquinone oxidoreductase-related | 1.48 |
| 260550_at | At2g43420 | 3-beta hydroxysteroid dehydrogenase/isomerase | 1.48 |
| 252357_at | At3g48410 | hydrolase, alpha/beta fold | 1.48 |
| 259297_at | At3g05360 | disease resistance | 1.48 |
| 254776_at | At4g13360 | enoyl-CoA hydratase | 1.48 |
| 249636_at | At5g36890 | glycosyl hydrolase family 1 protein | 1.48 |
| 261318_at | At1g53035 | expressed protein | 1.48 |
| 256611_at | At3g29270 | expressed protein | 1.48 |
| 249860_at | At5g22860 | serine carboxypeptidase S28 | 1.48 |
| 255827_at | At2g40600 | appr-1-p processing enzyme | 1.48 |
| 251688_at | At3g56480 | myosin heavy chain-related | 1.48 |
| 254594_at | At4g18930 | cyclic phosphodiesterase | 1.47 |
| 253948_at | At4g26940 | galactosyltransferase | 1.47 |
| 252389_at | At3g47833 | expressed protein | 1.47 |
| 257004_s_at | At3g14150 | (S)-2-hydroxy-acid oxidase, peroxisomal, putative | 1.47 |
| 254891_at | At4g11740 | ara4-interacting protein, putative (SAY1) | 1.47 |
| 246479_at | At5g16060 | expressed protein | 1.47 |
| 249161_at | At5g42790 | 20S proteasome alpha subunit F1 (PAF1) | 1.47 |
| 259560_at | At1g21270 | wall-associated kinase 2 (WAK2) | 1.47 |
| 249866_at | At5g23010 | 2-isopropylmalate synthase 3 (IMS3) | 1.47 |
| 258852_at | At3g06300 | oxidoreductase, 2OG-Fe(II) oxygenase | 1.47 |
| 250603_at | At5g07820 | expressed protein | 1.47 |
| 256179_at | At1g51710 | ubiquitin-specific protease 6, putative (UBP6) | 1.47 |
| 254027_at | At4g25835 | AAA-type ATPase | 1.47 |
| 245854_at | At5g13490 | adenine nucleotide translocator 2 (ANT2) | 1.47 |
| 253436_at | At4g32470 | ubiquinol-cytochrome C reductase complex 14 kDa protein, putative | 1.47 |
| 248088_at | At5g55070 | 2-oxoacid dehydrogenase | 1.47 |
| 254429_at | At4g21105 | expressed protein | 1.47 |
| 253705_at | At4g29130 | hexokinase 1 (HXK1) | 1.47 |
| 262637_at | At1g06640 | 2-oxoglutarate-dependent dioxygenase, putative | 1.46 |
| 257713_at | At3g27380 | sdh2-1: succinate dehydrogenase, iron-sulphur subunit, mitochondrial (SDH2-1) | 1.46 |
| 251975_at | At3g53230 | cell division cycle protein 48, putative (CDC48) | 1.46 |

| | | | |
|-------------|-----------|---|------|
| 251922_at | At3g54030 | protein kinase | 1.46 |
| 251499_at | At3g59100 | glycosyl transferase family 48 protein | 1.46 |
| 245501_at | At4g15620 | integral membrane | 1.46 |
| 253303_at | At4g33780 | expressed protein | 1.46 |
| 249796_at | At5g23540 | 26S proteasome regulatory subunit, putative | 1.46 |
| 262781_s_at | At1g13060 | 20S proteasome beta subunit E, putative | 1.46 |
| 252877_at | At4g39630 | expressed protein | 1.46 |
| 250775_at | At5g05460 | glycosyl hydrolase family 85 protein | 1.46 |
| 245085_at | At2g23350 | polyadenylate-binding protein, putative (PABP) | 1.46 |
| 265769_at | At2g48090 | expressed protein | 1.46 |
| 252246_at | At3g49730 | pentatricopeptide (PPR) repeat-containing protein | 1.46 |
| 249372_at | At5g40760 | glucose-6-phosphate 1-dehydrogenase (ACG12) | 1.46 |
| 266187_at | At2g38970 | zinc finger (C3HC4-type RING finger) | 1.46 |
| 258614_at | At3g02770 | dimethylmenaquinone methyltransferase | 1.45 |
| 262769_at | At1g13180 | actin-related protein 3 (ARP3) | 1.45 |
| 251076_at | At5g01970 | expressed protein | 1.45 |
| 252864_at | At4g39740 | electron transport SCO1/SenC | 1.45 |
| 257809_at | At3g27060 | ribonucleoside-diphosphate reductase small chain, putative | 1.45 |
| 254280_at | At4g22756 | sterol desaturase | 1.45 |
| 259708_at | At1g77420 | hydrolase, alpha/beta fold | 1.45 |
| 246189_at | At5g20910 | zinc finger (C3HC4-type RING finger) | 1.45 |
| 255671_at | At4g00355 | expressed protein | 1.45 |
| 247725_at | At5g59410 | expressed protein | 1.45 |
| 263663_at | At1g04410 | malate dehydrogenase, cytosolic, putative | 1.45 |
| 261831_at | At1g10630 | ADP-ribosylation factor, putative | 1.45 |
| 267055_at | At2g38360 | prenylated rab acceptor (PRA1) | 1.44 |
| 254083_at | At4g24920 | protein transport protein SEC61 gamma subunit, putative | 1.44 |
| 262481_at | At1g17080 | expressed protein | 1.44 |
| 249354_at | At5g40480 | expressed protein | 1.44 |
| 264724_at | At1g22920 | COP9 signalosome subunit 5B (CSN5B) | 1.44 |
| 256268_at | At3g12280 | RB: retinoblastoma-related protein (RBR1) | 1.44 |
| 248127_at | At5g54750 | transport protein particle (TRAPP) component Bet3, putative | 1.44 |
| 247257_at | At5g64760 | 26S proteasome regulatory subunit, putative (RPN5) | 1.44 |
| 262314_at | At1g70810 | C2 domain-containing protein | 1.43 |
| 254275_at | At4g22670 | tetratricopeptide repeat (TPR)-containing protein | 1.43 |
| 265446_at | At2g37110 | expressed protein | 1.43 |
| 256186_at | At1g51680 | 4-coumaroyl-CoA synthase 1 (4CL1) | 1.43 |
| 267001_at | At2g34470 | urease accessory protein (UREG) | 1.43 |
| 266206_at | At2g27730 | expressed protein | 1.43 |
| 246562_at | At5g15580 | expressed protein | 1.43 |
| 245891_at | At5g09220 | amino acid permease 2 (AAP2) | 1.43 |
| 245412_at | At4g17280 | auxin-responsive | 1.43 |
| 260673_at | At1g19330 | expressed protein | 1.43 |
| 256787_at | At3g13790 | beta-fructosidase (BFRUCT1) | 1.43 |
| 266874_at | At2g44760 | expressed protein | 1.43 |

| | | | |
|-------------|-----------|--|------|
| 262918_at | At1g65000 | expressed protein | 1.42 |
| 253052_at | At4g37310 | cytochrome P450, putative | 1.42 |
| 263919_at | At2g36470 | expressed protein | 1.42 |
| 260003_at | At1g68100 | IAA-alanine resistance protein 1, putative | 1.42 |
| 255728_at | At1g25500 | choline transporter-related | 1.42 |
| 255236_at | At4g05520 | calcium-binding EF hand | 1.42 |
| 248760_at | At5g47570 | expressed protein | 1.42 |
| 253642_at | At4g29960 | expressed protein | 1.42 |
| 250553_at | At5g07960 | expressed protein | 1.42 |
| 250471_at | At5g10170 | inositol-3-phosphate synthase, putative | 1.42 |
| 258391_at | At3g15420 | expressed protein | 1.42 |
| 255596_at | At4g01720 | WRKY family transcription factor | 1.42 |
| 255261_s_at | At4g05110 | equilibrative nucleoside transporter, putative (ENT6) | 1.42 |
| 254053_s_at | At4g25300 | oxidoreductase, 2OG-Fe(II) oxygenase | 1.42 |
| 245202_at | At1g67720 | leucine-rich repeat | 1.42 |
| 262366_at | At1g72890 | disease resistance protein (TIR-NBS class), putative | 1.41 |
| 258457_at | At3g22422 | imidazoglycerol-phosphate dehydratase 1 (IGPD1) | 1.41 |
| 267355_at | At2g39900 | LIM domain-containing protein | 1.41 |
| 262946_at | At1g79390 | expressed protein | 1.41 |
| 255291_at | At4g04700 | calcium-dependent protein kinase, putative (CDPK) | 1.41 |
| 251034_at | At5g02040 | prenylated rab acceptor (PRA1) | 1.41 |
| 261227_at | At1g20200 | 26S proteasome regulatory subunit S3, putative (RPN3) | 1.41 |
| 254462_at | At4g20150 | expressed protein | 1.4 |
| 252955_at | At4g38630 | 26S proteasome regulatory subunit S5A (RPN10) | 1.4 |
| 263224_at | At1g30580 | expressed protein | 1.4 |
| 252027_at | At3g52850 | vacuolar sorting receptor, putative | 1.4 |
| 246824_at | At5g26990 | drought-responsive | 1.4 |
| 260274_at | At1g80460 | glycerol kinase, putative | 1.4 |
| 259978_at | At1g76540 | cell division control protein, putative | 1.4 |
| 252723_at | At3g43520 | expressed protein | 1.4 |
| 264249_at | At1g78920 | vacuolar-type H+-translocating inorganic pyrophosphatase (AVPL1) | 1.39 |
| 263419_at | At2g17220 | protein kinase, putative | 1.39 |
| 247080_at | At5g66140 | 20S proteasome alpha subunit D2 (PAD2) | 1.39 |
| 247521_at | At5g61450 | 2-phosphoglycerate kinase-related | 1.39 |
| 263986_at | At2g42790 | citrate synthase, glyoxysomal, putative | 1.39 |
| 263314_at | At2g05760 | xanthine/uracil permease | 1.39 |
| 261087_at | At1g17350 | auxin-induced-related | 1.38 |
| 250863_at | At5g04750 | F1FO-ATPase inhibitor protein, putative | 1.38 |
| 251828_at | At3g55070 | expressed protein | 1.38 |
| 248778_at | At5g47940 | expressed protein | 1.38 |
| 261862_at | At1g50410 | SNF2 domain-containing protein | 1.38 |
| 260887_at | At1g29160 | Dof-type zinc finger domain-containing protein | 1.38 |
| 257431_at | At2g36360 | kelch repeat-containing protein | 1.38 |
| 267189_at | At2g44180 | methionyl aminopeptidase, putative | 1.38 |

| | | | |
|-------------|-----------|---|-------|
| 265965_at | At2g37500 | arginine biosynthesis protein ArgJ family | 1.38 |
| 257444_at | At2g12550 | ubiquitin-associated (UBA) | 1.38 |
| 265343_at | At2g22570 | isochorismatase hydrolase | 1.38 |
| 264506_at | At1g09560 | germin-like protein (GLP4) | 1.38 |
| 249095_at | At5g43900 | myosin heavy chain (MYA2) | 1.38 |
| 250406_at | At5g10810 | enhancer of rudimentary protein, putative | 1.37 |
| 264303_s_at | At1g78870 | ubiquitin-conjugating enzyme, putative | 1.37 |
| 251954_at | At3g53670 | expressed protein | 1.37 |
| 248276_at | At5g53550 | transporter, putative | 1.37 |
| 266553_at | At2g46170 | reticulon (RTNLB5) | 1.37 |
| 260772_at | At1g49050 | aspartyl protease | 1.37 |
| 251932_at | At3g54010 | peptidyl-prolyl cis-trans isomerase, putative (PAS1-D) | 1.37 |
| 265912_at | At2g25570 | expressed protein | 1.36 |
| 261144_s_at | At1g19660 | wound-responsive | 1.36 |
| 265738_at | At2g01350 | quinolinate phosphoribosyl transferase | 1.36 |
| 256042_at | At1g07220 | expressed protein | 1.36 |
| 247971_at | At5g56730 | peptidase M16 | 1.36 |
| 257634_s_at | At3g26170 | cytochrome P450 71B19, putative (CYP71B19) | 1.35 |
| 257893_at | At3g17000 | ubiquitin-conjugating enzyme, putative | 1.35 |
| 257926_at | At3g23280 | zinc finger (C3HC4-type RING finger) | 1.35 |
| 253317_at | At4g33960 | expressed protein | 1.35 |
| 247330_at | At5g63510 | bacterial transferase hexapeptide repeat-containing protein | 1.35 |
| 266090_at | At2g38000 | chaperone protein dnaJ-related | 1.35 |
| 245046_at | At2g26510 | xanthine/uracil permease | 1.35 |
| 264871_at | At1g24180 | pyruvate dehydrogenase E1 component alpha subunit, putative | 1.35 |
| 250193_at | At5g14540 | proline-rich | 1.34 |
| 259342_at | At3g03890 | expressed protein | 1.33 |
| 254547_at | At4g19860 | lecithin:cholesterol acyltransferase (LACT) | 1.32 |
| 251787_at | At3g55410 | 2-oxoglutarate dehydrogenase E1 component, putative | 1.31 |
| 259230_at | At3g07780 | expressed protein | -1.28 |
| 260127_at | At1g36320 | expressed protein | -1.33 |
| 246045_at | At5g19430 | zinc finger (C3HC4-type RING finger) | -1.33 |
| 262468_at | At1g50200 | aminoacyl-tRNA synthetase | -1.33 |
| 251237_at | At3g62420 | bZIP transcription factor | -1.33 |
| 245042_at | At2g26540 | uroporphyrinogen-III synthase | -1.33 |
| 249181_at | At5g42920 | expressed protein | -1.33 |
| 247103_at | At5g66610 | LIM domain-containing protein | -1.33 |
| 260610_at | At2g43680 | calmodulin-binding | -1.33 |
| 267000_at | At2g34310 | expressed protein | -1.34 |
| 266606_at | At2g46310 | AP2 domain-containing transcription factor, putative | -1.34 |
| 263688_at | At1g26920 | expressed protein | -1.34 |
| 247043_at | At5g66880 | serine/threonine protein kinase, putative | -1.34 |
| 263334_at | At2g03820 | nonsense-mediated mRNA decay NMD3 | -1.34 |
| 259768_at | At1g29390 | stress-responsive protein, putative | -1.35 |

| | | | |
|-------------|-----------|--|-------|
| 250385_at | At5g11520 | aspartate aminotransferase (ASP3) | -1.35 |
| 253756_at | At4g28980 | cyclin-dependent kinase-activating kinase 1At (CAK1) | -1.35 |
| 249138_at | At5g43070 | MFP1 attachment factor, putative | -1.35 |
| 264750_at | At1g22870 | protein kinase | -1.35 |
| 256765_at | At3g22200 | 4-aminobutyrate aminotransferase | -1.35 |
| 260800_at | At1g78240 | dehydration-responsive protein-related | -1.35 |
| 261887_at | At1g80780 | CCR4-NOT transcription complex protein, putative | -1.36 |
| 263750_at | At2g21530 | forkhead-associated domain-containing protein | -1.36 |
| 259092_at | At3g04870 | zeta-carotene desaturase (ZDS1) | -1.36 |
| 263760_at | At2g21280 | expressed protein | -1.36 |
| 262882_at | At1g64900 | cytochrome P450, putative | -1.36 |
| 261535_at | At1g01725 | expressed protein | -1.36 |
| 267092_at | At2g38120 | amino acid permease, putative (AUX1) | -1.37 |
| 262959_at | At1g54290 | eukaryotic translation initiation factor SUI1, putative | -1.37 |
| 252922_at | At4g39040 | expressed protein | -1.37 |
| 265413_s_at | At2g16650 | expressed protein | -1.37 |
| 263053_at | At2g13440 | glucose-inhibited division family A protein | -1.37 |
| 256747_at | At3g29180 | expressed protein | -1.37 |
| 245572_at | At4g14720 | expressed protein | -1.37 |
| 247000_at | At5g67380 | casein kinase II alpha chain 1 | -1.37 |
| 251845_at | At3g54540 | ABC transporter | -1.37 |
| 259258_at | At3g07670 | SET domain-containing protein | -1.38 |
| 251403_at | At3g60300 | RWD domain-containing protein | -1.38 |
| 250705_at | At5g06340 | diadenosine 5',5'''-P1,P4-tetraphosphate hydrolase, putative | -1.38 |
| 266335_at | At2g32440 | ent-kaurenoic acid hydroxylase, putative | -1.38 |
| 265318_at | At2g22650 | FAD-dependent oxidoreductase | -1.38 |
| 255376_x_at | At4g03790 | At4g03790 | -1.38 |
| 265305_at | At2g20340 | tyrosine decarboxylase, putative | -1.38 |
| 264828_at | At1g03380 | expressed protein | -1.38 |
| 247653_at | At5g59950 | RNA and export factor-binding protein, putative | -1.38 |
| 264970_at | At1g67280 | lactoylglutathione lyase, putative | -1.38 |
| 260705_at | At1g32400 | senescence-associated | -1.38 |
| 258052_at | At3g16190 | isochorismatase hydrolase | -1.38 |
| 260903_at | At1g02460 | glycoside hydrolase family 28 protein | -1.38 |
| 258643_at | At3g08010 | expressed protein | -1.38 |
| 258585_at | At3g04340 | FtsH protease | -1.38 |
| 253440_at | At4g32570 | expressed protein | -1.39 |
| 246611_at | At5g35330 | methyl-CpG-binding domain-containing protein | -1.39 |
| 264996_at | At1g67230 | expressed protein | -1.39 |
| 260510_at | At1g51580 | KH domain-containing protein | -1.39 |
| 248181_at | At5g54290 | cytochrome c biogenesis protein family | -1.39 |
| 257810_at | At3g27070 | translocase of outer membrane 20kDa subunit 1 (TOM20-1) | -1.39 |
| 263898_at | At2g21950 | SKP1 interacting partner 6 (SKIP6) | -1.39 |
| 251450_at | At3g60030 | squamosa promoter-binding protein-like 12 (SPL12) | -1.39 |

| | | | |
|-------------|-----------|--|-------|
| 267492_at | At2g30620 | histone H1.2 | -1.4 |
| 259862_at | At1g72650 | myb family transcription factor | -1.4 |
| 254612_at | At4g19100 | expressed protein | -1.4 |
| 266728_at | At2g03140 | CAAX amino terminal protease | -1.4 |
| 263946_at | At2g36000 | mitochondrial transcription termination factor-related | -1.4 |
| 245165_at | At2g33180 | expressed protein | -1.4 |
| 251211_s_at | At3g62470 | pentatricopeptide (PPR) repeat-containing protein | -1.4 |
| 255720_at | At1g32060 | phosphoribulokinase (PRK) | -1.4 |
| 259038_at | At3g09210 | KOW domain-containing transcription factor | -1.4 |
| 253834_at | At4g27800 | protein phosphatase 2C PPH1 (PPH1) | -1.4 |
| 265707_at | At2g03390 | uvrB/uvrC motif-containing protein | -1.4 |
| 262502_at | At1g21600 | expressed protein | -1.4 |
| 258206_at | At3g14010 | hydroxyproline-rich glycoprotein | -1.41 |
| 266923_at | At2g45980 | expressed protein | -1.41 |
| 258602_at | At3g02750 | protein phosphatase 2C (PP2C) | -1.41 |
| 254057_at | At4g25170 | expressed protein | -1.41 |
| 252759_at | At3g42630 | pentatricopeptide (PPR) repeat-containing protein | -1.41 |
| 263365_at | At2g20550 | DNAJ chaperone C-terminal domain-containing protein | -1.41 |
| 262592_at | At1g15400 | expressed protein | -1.41 |
| 261179_at | At1g04985 | expressed protein | -1.41 |
| 250711_at | At5g06110 | DNAJ heat shock N-terminal domain-containing protein / cell division protein-related | -1.41 |
| 249827_at | At5g23330 | riboflavin biosynthesis protein-related | -1.41 |
| 259242_at | At3g33520 | actin-related protein 6 (ARP6) | -1.41 |
| 249288_at | At5g41050 | expressed protein | -1.41 |
| 247108_at | At5g66160 | protease-associated zinc finger (C3HC4-type RING finger) | -1.41 |
| 260490_at | At1g51500 | ABC transporter | -1.41 |
| 261628_at | At1g50000 | hypothetical protein | -1.41 |
| 261190_at | At1g32990 | ribosomal protein L11 | -1.41 |
| 253355_at | At4g33380 | expressed protein | -1.41 |
| 262988_at | At1g23310 | glutamate:glyoxylate aminotransferase 1 (GGT1) | -1.41 |
| 245795_at | At1g32160 | expressed protein | -1.42 |
| 245396_at | At4g14870 | expressed protein | -1.42 |
| 259207_at | At3g09050 | expressed protein | -1.42 |
| 251855_at | At3g54690 | sugar isomerase (SIS) domain-containing protein / CBS domain-containing protein | -1.42 |
| 262110_at | At1g02840 | pre-mRNA splicing factor SF2 (SF2) / SR1 protein | -1.42 |
| 252105_at | At3g51470 | protein phosphatase 2C, putative / PP2C, putative | -1.42 |
| 261320_at | At1g53120 | RNA-binding S4 domain-containing protein | -1.42 |
| 260283_at | At1g80480 | PRLI-interacting factor L, putative | -1.42 |
| 257908_at | At3g25410 | bile acid:sodium symporter | -1.42 |
| 261844_at | At1g15940 | expressed protein | -1.42 |
| 256855_at | At3g15190 | chloroplast 30S ribosomal protein S20, putative | -1.42 |
| 254764_at | At4g13250 | short-chain dehydrogenase/reductase (SDR) | -1.42 |
| 245285_s_at | At4g14040 | selenium-binding protein, putative | -1.42 |

| | | | |
|-------------|-----------|---|-------|
| 265142_at | At1g51360 | expressed protein | -1.42 |
| 262599_at | At1g15350 | expressed protein | -1.43 |
| 255015_at | At4g09980 | methyltransferase MT-A70 | -1.43 |
| 252612_at | At3g45160 | expressed protein | -1.43 |
| 261197_at | At1g12900 | glyceraldehyde 3-phosphate dehydrogenase, chloroplast, putative | -1.43 |
| 259514_at | At1g12480 | C4-dicarboxylate transporter | -1.43 |
| 266229_at | At2g28840 | ankyrin repeat | -1.43 |
| 262235_at | At1g48350 | ribosomal protein L18 | -1.43 |
| 256892_at | At3g19000 | oxidoreductase, 2OG-Fe(II) oxygenase | -1.43 |
| 247617_at | At5g60270 | lectin protein kinase | -1.43 |
| 255829_at | At2g40540 | potassium transporter, putative (KT2) | -1.43 |
| 246294_at | At3g56910 | expressed protein | -1.43 |
| 246856_at | At5g26210 | PHD finger | -1.43 |
| 266929_at | At2g45850 | DNA-binding | -1.43 |
| 266716_at | At2g46820 | expressed protein | -1.43 |
| 250430_at | At5g10460 | haloacid dehalogenase-like hydrolase | -1.43 |
| 245008_at | ycf3 | ycf3 | -1.43 |
| 263906_at | At2g36250 | chloroplast division protein FtsZ (FtsZ2-1) | -1.43 |
| 256680_at | At3g52230 | expressed protein | -1.43 |
| 258161_at | At3g17930 | expressed protein | -1.43 |
| 255710_at | At4g00030 | plastid-lipid associated protein PAP | -1.43 |
| 253001_at | At4g38490 | expressed protein | -1.43 |
| 248774_at | At5g47830 | expressed protein | -1.44 |
| 266575_at | At2g24060 | translation initiation factor 3 (IF-3) | -1.44 |
| 260930_at | At1g02620 | GTP-binding protein (SAR1A) | -1.44 |
| 260260_at | At1g68540 | oxidoreductase | -1.44 |
| 254697_at | At4g17970 | expressed protein | -1.44 |
| 255217_s_at | At4g07680 | pseudogene, Ulp1 protease famiy | -1.44 |
| 254504_at | At4g20030 | RNA recognition motif (RRM)-containing protein | -1.44 |
| 267089_at | At2g38300 | myb family transcription factor | -1.44 |
| 260898_at | At1g29070 | ribosomal protein L34 | -1.44 |
| 248713_at | At5g48180 | kelch repeat-containing protein | -1.44 |
| 245952_at | At5g28500 | expressed protein | -1.44 |
| 264977_at | At1g27090 | glycine-rich protein | -1.44 |
| 264342_at | At1g12080 | expressed protein | -1.44 |
| 254680_at | At4g18130 | phytochrome E (PHYE) | -1.44 |
| 265732_at | At2g01300 | expressed protein | -1.44 |
| 259123_at | At3g02200 | proteasome | -1.44 |
| 252199_at | At3g50270 | transferase | -1.44 |
| 263046_at | At2g05380 | glycine-rich protein (GRP3S) | -1.44 |
| 256402_at | At3g06130 | heavy-metal-associated domain-containing protein | -1.44 |
| 248838_at | At5g46800 | mitochondrial carnitine | -1.44 |
| 247079_at | At5g66055 | ankyrin repeat protein (AKR) | -1.44 |
| 264956_at | At1g76990 | ACT domain containing protein | -1.44 |

| | | | |
|-------------|-----------|--|-------|
| 267553_s_at | At2g32650 | expressed protein | -1.45 |
| 262295_at | At1g27650 | U2 snRNP auxiliary factor small subunit, putative | -1.45 |
| 246215_at | At4g37180 | myb family transcription factor | -1.45 |
| 255413_at | At4g03140 | short-chain dehydrogenase/reductase (SDR) | -1.45 |
| 246339_at | At3g44890 | 50S ribosomal protein L9, chloroplast (CL9) | -1.45 |
| 265247_at | At2g43030 | ribosomal protein L3 | -1.45 |
| 260467_at | At1g10890 | F-box | -1.45 |
| 254073_at | At4g25500 | arginine/serine-rich splicing factor RSP40 (RSP40) | -1.45 |
| 251810_at | At3g55250 | expressed protein | -1.45 |
| 266642_at | At2g35410 | 33 kDa ribonucleoprotein, chloroplast, putative | -1.45 |
| 262480_at | At1g11340 | S-locus lectin protein kinase | -1.45 |
| 257953_at | At3g21865 | expressed protein | -1.45 |
| 261141_at | At1g19740 | ATP-dependent protease La (LON) | -1.45 |
| 261196_at | At1g12860 | basic helix-loop-helix (bHLH) | -1.45 |
| 258696_at | At3g09650 | pentatricopeptide (PPR) repeat-containing protein | -1.45 |
| 246588_at | At5g14840 | expressed protein | -1.45 |
| 261668_at | At1g18500 | 2-isopropylmalate synthase, putative | -1.45 |
| 246548_at | At5g14910 | heavy-metal-associated domain-containing protein | -1.45 |
| 266121_at | At2g02160 | zinc finger (CCCH-type) | -1.45 |
| 259160_at | At3g05410 | expressed protein | -1.46 |
| 253235_at | At4g34350 | LytB | -1.46 |
| 247146_at | At5g65610 | expressed protein | -1.46 |
| 259991_at | At1g68040 | S-adenosyl-L-methionine:carboxyl methyltransferase | -1.46 |
| 259773_at | At1g29500 | auxin-responsive protein, putative | -1.46 |
| 264613_at | At1g04640 | biotin/lipoate A/B protein ligase | -1.46 |
| 260044_at | At1g73655 | immunophilin | -1.46 |
| 256796_at | At3g22210 | expressed protein | -1.46 |
| 256021_at | At1g58270 | meprin and TRAF homology domain-containing protein | -1.46 |
| 249283_at | At5g41800 | amino acid transporter | -1.46 |
| 266856_at | At2g26910 | ABC transporter | -1.46 |
| 262721_at | At1g43560 | thioredoxin | -1.46 |
| 250786_at | At5g05540 | exonuclease | -1.46 |
| 248634_at | At5g49030 | tRNA synthetase class I (I, L, M and V) | -1.46 |
| 256033_at | At1g07250 | UDP-glucuronosyl/UDP-glucosyl transferase | -1.46 |
| 261230_at | At1g20010 | tubulin beta-5 chain (TUB5) | -1.46 |
| 260409_at | At1g69935 | expressed protein | -1.46 |
| 247307_at | At5g63860 | UVB-resistance protein (UVR8) | -1.46 |
| 254769_at | At4g13330 | expressed protein | -1.47 |
| 254388_at | At4g21860 | methionine sulfoxide reductase domain-containing protein | -1.47 |
| 246454_at | At5g16710 | dehydroascorbate reductase, putative | -1.47 |
| 265415_at | At2g20890 | expressed protein | -1.47 |
| 265284_at | At2g20230 | expressed protein | -1.47 |
| 254188_at | At4g23920 | UDP-glucose 4-epimerase, putative | -1.47 |
| 262945_at | At1g79510 | expressed protein | -1.47 |
| 261218_at | At1g20020 | ferredoxin-NADP(+) reductase, putative | -1.47 |

| | | | |
|-------------|-----------|--|-------|
| 257789_at | At3g27020 | oligopeptide transporter OPT | -1.47 |
| 252823_at | At4g40045 | expressed protein | -1.47 |
| 259436_at | At1g01500 | expressed protein | -1.47 |
| 258512_at | At3g06510 | glycosyl hydrolase family 1 protein | -1.47 |
| 247750_at | At5g58950 | protein kinase | -1.47 |
| 244996_at | rps2 | rps2 | -1.47 |
| 255332_at | At4g04340 | early-responsive to dehydration protein-related (ERD) | -1.47 |
| 261264_at | At1g26710 | expressed protein | -1.48 |
| 252694_at | At3g43630 | nodulin, putative | -1.48 |
| 247972_at | At5g56740 | histone acetyltransferase | -1.48 |
| 264991_s_at | At3g43400 | phagocytosis and cell motility protein ELMO1-related | -1.48 |
| 246829_at | At5g26570 | glycoside hydrolase starch-binding domain-containing protein | -1.48 |
| 246463_at | At5g16970 | NADP-dependent oxidoreductase, putative (P1) | -1.48 |
| 257477_at | At1g10660 | expressed protein | -1.48 |
| 251218_at | At3g62410 | CP12 domain-containing protein | -1.48 |
| 248854_at | At5g46580 | pentatricopeptide (PPR) repeat-containing protein | -1.48 |
| 249993_at | At5g18570 | GTP1/OBG | -1.48 |
| 267220_at | At2g02500 | expressed protein | -1.48 |
| 250763_at | At5g06060 | tropinone reductase, putative | -1.48 |
| 248828_at | At5g47110 | lil3 protein, putative | -1.48 |
| 246613_at | At5g35360 | acetyl-CoA carboxylase, biotin carboxylase subunit (CAC2) | -1.48 |
| 253307_at | At4g33670 | L-galactose dehydrogenase (L-GalDH) | -1.48 |
| 264121_at | At1g02280 | GTP-binding protein (TOC33) | -1.48 |
| 248603_at | At5g49430 | transducin | -1.49 |
| 245718_at | At5g04100 | DNA topoisomerase II | -1.49 |
| 260205_at | At1g70700 | expressed protein | -1.49 |
| 256063_at | At1g07130 | OB-fold nucleic acid binding domain-containing protein | -1.49 |
| 244997_at | rpoC2 | rpoC2 | -1.49 |
| 261483_at | At1g14270 | CAAX amino terminal protease | -1.49 |
| 254659_at | At4g18240 | starch synthase-related protein | -1.49 |
| 263014_at | At1g23400 | expressed protein | -1.49 |
| 247855_at | At5g58210 | hydroxyproline-rich glycoprotein | -1.49 |
| 267504_at | At2g45530 | zinc finger (C3HC4-type RING finger) | -1.49 |
| 255572_at | At4g01050 | hydroxyproline-rich glycoprotein | -1.49 |
| 250146_at | At5g14660 | peptide deformylase, chloroplast 1B (PDF1B) | -1.49 |
| 264584_at | At1g05140 | membrane-associated zinc metalloprotease, putative | -1.49 |
| 263920_at | At2g36410 | expressed protein | -1.49 |
| 259296_at | At3g05350 | aminopeptidase P, cytosolic, putative | -1.49 |
| 248238_at | At5g53900 | expressed protein | -1.49 |
| 264624_at | At1g08930 | early-responsive to dehydration stress protein (ERD6) | -1.49 |
| 256502_at | At1g36730 | eukaryotic translation initiation factor 5, putative (eIF-5) | -1.49 |
| 245905_at | At5g11090 | serine-rich protein-related | -1.49 |
| 266624_s_at | At2g35390 | ribose-phosphate pyrophosphokinase 2 (PRS2) | -1.49 |
| 263296_at | At2g38800 | calmodulin-binding protein-related | -1.49 |
| 251929_at | At3g53920 | RNA polymerase sigma subunit SigC (SIG3) | -1.49 |

| | | | |
|-------------|-----------|--|-------|
| 246090_at | At5g20520 | expressed protein | -1.49 |
| 257827_at | At3g26630 | pentatricopeptide (PPR) repeat-containing protein | -1.5 |
| 260086_at | At1g63240 | expressed protein | -1.5 |
| 248250_at | At5g53130 | cyclic nucleotide-regulated ion channel (CNGC1) | -1.5 |
| 245354_at | At4g17600 | lil3 protein | -1.5 |
| 263412_at | At2g28720 | histone H2B, putative | -1.5 |
| 265139_at | At1g51310 | tRNA methyl transferase | -1.5 |
| 245117_at | At2g41560 | calcium-transporting ATPase 4, plasma membrane-type (ACA4) | -1.5 |
| 246007_at | At5g08410 | ferredoxin-thioredoxin reductase, putative | -1.5 |
| 254760_at | At4g13200 | expressed protein | -1.5 |
| 245198_at | At1g67700 | expressed protein | -1.5 |
| 247163_at | At5g65685 | soluble glycogen synthase-related | -1.5 |
| 264521_at | At1g10020 | expressed protein | -1.5 |
| 264045_at | At2g22450 | riboflavin biosynthesis protein, putative | -1.5 |
| 247826_at | At5g58480 | glycosyl hydrolase family 17 protein | -1.5 |
| 262174_at | At1g74910 | ADP-glucose pyrophosphorylase | -1.5 |
| 246019_at | At5g10690 | pentatricopeptide (PPR) repeat-containing protein | -1.5 |
| 262104_at | At1g02910 | tetratricopeptide repeat (TPR)-containing protein | -1.5 |
| 258060_at | At3g26030 | serine/threonine protein phosphatase 2A (PP2A) | -1.51 |
| 248029_at | At5g55700 | glycosyl hydrolase family 14 protein | -1.51 |
| 262572_at | At1g15140 | oxidoreductase NAD-binding domain-containing protein | -1.51 |
| 248094_at | At5g55220 | trigger factor type chaperone | -1.51 |
| 264845_at | At1g03675 | thioredoxin M-type 1, chloroplast (TRX-M1) | -1.51 |
| 264640_at | At1g65680 | beta-expansin, putative (EXBP2) | -1.51 |
| 262483_at | At1g17220 | translation initiation factor IF-2, chloroplast, putative | -1.51 |
| 261457_at | At1g21065 | expressed protein | -1.51 |
| 258071_s_at | At3g26070 | plastid-lipid associated protein PAP | -1.51 |
| 255788_at | At2g33310 | IAA13: auxin-responsive protein | -1.51 |
| 247095_at | At5g66400 | RAB18: dehydrin (RAB18) | -1.51 |
| 261053_at | At1g01320 | tetratricopeptide repeat (TPR)-containing protein | -1.51 |
| 253421_at | At4g32340 | expressed protein | -1.51 |
| 255636_at | At4g00730 | anthocyaninless2 (ANL2) | -1.51 |
| 246031_at | At5g21160 | La domain-containing protein | -1.51 |
| 245592_at | At4g14540 | CCAAT-box binding transcription factor subunit B (NF-YB) | -1.51 |
| 263705_at | At1g31190 | inositol monophosphatase | -1.51 |
| 251055_at | At5g01710 | expressed protein | -1.51 |
| 248140_at | At5g54980 | integral membrane | -1.51 |
| 247524_at | At5g61440 | thioredoxin | -1.51 |
| 267196_at | At2g30950 | FtsH protease (VAR2) | -1.51 |
| 252978_at | At4g38590 | glycosyl hydrolase family 35 protein | -1.51 |
| 249519_at | At5g38660 | expressed protein | -1.51 |
| 248906_at | At5g46420 | 16S rRNA processing protein RimM family | -1.51 |
| 257611_at | At3g26580 | expressed protein | -1.51 |
| 249109_at | At5g43700 | auxin-responsive protein (AUX2-11) | -1.52 |

| | | | |
|-----------|-----------|---|-------|
| 250649_at | At5g06690 | thioredoxin | -1.52 |
| 267511_at | At2g45670 | calcineurin B subunit-related | -1.52 |
| 253009_at | At4g37930 | glycine hydroxymethyltransferase (SHM1) | -1.52 |
| 247543_at | At5g61600 | ethylene-responsive element-binding | -1.52 |
| 263000_at | At1g54350 | ABC transporter | -1.52 |
| 262127_at | At1g52550 | expressed protein | -1.52 |
| 259379_at | At3g16350 | myb family transcription factor | -1.52 |
| 259181_at | At3g01690 | expressed protein | -1.52 |
| 266207_at | At2g27680 | aldo/keto reductase | -1.52 |
| 256058_at | At1g07240 | UDP-glucuronosyl/UDP-glucosyl transferase | -1.52 |
| 260730_at | At1g48030 | dihydrolipoamide dehydrogenase 1, mitochondrial (MTLPD1) | -1.52 |
| 258963_at | At3g10550 | expressed protein | -1.52 |
| 258708_at | At3g09580 | amine oxidase | -1.52 |
| 247386_at | At5g63430 | metallo-beta-lactamase | -1.52 |
| 259858_at | At1g68400 | leucine-rich repeat transmembrane protein kinase, putative | -1.52 |
| 250884_at | At5g03940 | signal recognition particle 54 kDa protein, chloroplast (FFC) | -1.52 |
| 246053_at | At5g08340 | riboflavin biosynthesis protein-related | -1.52 |
| 265392_at | At2g20860 | lipoic acid synthase (LIP1) | -1.52 |
| 261211_at | At1g12780 | UDP-glucose 4-epimerase | -1.53 |
| 250058_at | At5g17870 | plastid-specific ribosomal protein-related | -1.53 |
| 262503_at | At1g21670 | expressed protein | -1.53 |
| 262065_at | At1g56180 | expressed protein | -1.53 |
| 246154_at | At5g19940 | plastid-lipid associated protein PAP-related | -1.53 |
| 266099_at | At2g38040 | acetyl co-enzyme A carboxylase carboxyltransferase alpha subunit family | -1.53 |
| 258149_at | At3g18110 | pentatricopeptide (PPR) repeat-containing protein | -1.53 |
| 257706_at | At3g12685 | expressed protein | -1.53 |
| 259308_at | At3g05180 | GDSL-motif lipase/hydrolase | -1.53 |
| 247126_at | At5g66080 | protein phosphatase 2C (PP2C) | -1.53 |
| 261534_at | At1g01820 | peroxisomal biogenesis factor 11 (PEX11) | -1.53 |
| 253858_at | At4g27600 | pfkB-type carbohydrate kinase | -1.53 |
| 265790_at | At2g01170 | amino acid permease | -1.53 |
| 258782_at | At3g11750 | dihydroneopterin aldolase, putative | -1.53 |
| 267471_at | At2g30390 | ferrochelatase II | -1.53 |
| 264328_at | At1g04100 | indoleacetic acid-induced protein 10 (IAA10) | -1.53 |
| 246194_at | At4g37000 | accelerated cell death 2 (ACD2) | -1.53 |
| 260638_at | At1g62390 | Bem1p (PB1) domain-containing protein | -1.53 |
| 250218_at | At5g14170 | SWIB complex BAF60b domain-containing protein | -1.53 |
| 262600_at | At1g15340 | methyl-CpG-binding domain-containing protein | -1.53 |
| 253823_at | At4g28030 | GCN5-related N-acetyltransferase (GNAT) | -1.53 |
| 261206_at | At1g12800 | S1 RNA-binding domain-containing protein | -1.54 |
| 257519_at | At3g01210 | RNA recognition motif (RRM)-containing protein | -1.54 |
| 246311_at | At3g51880 | high mobility group protein alpha (HMGalpha) | -1.54 |
| 261507_at | At1g71720 | S1 RNA-binding domain-containing protein | -1.54 |
| 255456_at | At4g02920 | expressed protein | -1.54 |

| | | | |
|-----------|-----------|---|-------|
| 263947_at | At2g35820 | expressed protein | -1.54 |
| 253517_at | At4g31390 | ABC1 | -1.54 |
| 256653_at | At3g18870 | mitochondrial transcription termination factor-related | -1.54 |
| 249810_at | At5g23920 | expressed protein | -1.54 |
| 245532_at | At4g15110 | cytochrome P450 97B3, putative (CYP97B3) | -1.54 |
| 252391_at | At3g47860 | apolipoprotein D-related | -1.54 |
| 264781_at | At1g08540 | RNA polymerase sigma subunit SigB (sigB) | -1.54 |
| 251193_at | At3g62910 | peptide chain release factor, putative | -1.54 |
| 258797_at | At3g04730 | indoleacetic acid-induced protein 16 (IAA16) | -1.54 |
| 264963_at | At1g60600 | UbiA prenyltransferase | -1.54 |
| 247813_at | At5g58330 | malate dehydrogenase (NADP) putative | -1.54 |
| 255078_at | At4g09010 | L-ascorbate peroxidase putative | -1.54 |
| 253477_at | At4g32320 | peroxidase | -1.54 |
| 261272_at | At1g26665 | expressed protein | -1.54 |
| 264841_at | At1g03740 | protein kinase | -1.55 |
| 263889_at | At2g37010 | ABC transporter | -1.55 |
| 259523_at | At1g12500 | phosphate translocator-related | -1.55 |
| 247637_at | At5g60600 | 1-hydroxy-2-methyl-2-(E)-butenyl 4-diphosphate synthase, putative | -1.55 |
| 248404_at | At5g51460 | trehalose-6-phosphate phosphatase (TPPA) | -1.55 |
| 261728_at | At1g76160 | multi-copper oxidase type I | -1.55 |
| 258920_at | At3g10520 | non-symbiotic hemoglobin 2 (HB2) | -1.55 |
| 251987_at | At3g53280 | cytochrome P450 71B5 (CYP71B5) | -1.55 |
| 249849_at | At5g23230 | isochorismatase hydrolase | -1.55 |
| 263460_at | At2g31810 | acetolactate synthase small subunit, putative | -1.55 |
| 260783_at | At1g06160 | ethylene-responsive factor, putative | -1.55 |
| 266402_at | At2g38780 | expressed protein | -1.55 |
| 245229_at | At4g25620 | hydroxyproline-rich glycoprotein | -1.55 |
| 250128_at | At5g16540 | zinc finger (CCCH-type) | -1.55 |
| 244979_at | rps11 | rps11 | -1.55 |
| 261559_at | At1g01780 | LIM domain-containing protein | -1.55 |
| 252993_at | At4g38540 | monooxygenase, putative (MO2) | -1.55 |
| 261570_at | At1g01120 | fatty acid elongase 3-ketoacyl-CoA synthase 1 (KCS1) | -1.56 |
| 250378_at | At5g11570 | proton-dependent oligopeptide transport (POT) | -1.56 |
| 247613_at | At5g60740 | ABC transporter | -1.56 |
| 255625_at | At4g01120 | G-box binding factor 2 (GBF2) | -1.56 |
| 246999_at | At5g67440 | phototropic-responsive NPH3 | -1.56 |
| 248547_at | At5g50280 | pentatricopeptide (PPR) repeat-containing protein | -1.56 |
| 259980_at | At1g76520 | auxin efflux carrier | -1.56 |
| 249007_at | At5g44650 | expressed protein | -1.56 |
| 267581_at | At2g41980 | seven in absentia (SINA) | -1.56 |
| 258081_at | At3g26085 | CAAX amino terminal protease | -1.56 |
| 258923_at | At3g10450 | serine carboxypeptidase S10 | -1.56 |
| 246633_at | At1g29720 | protein kinase | -1.56 |
| 255866_at | At2g30350 | endo/excinuclease amino terminal domain-containing protein | -1.56 |

| | | | |
|-------------|-----------|--|-------|
| 246612_at | At5g35320 | expressed protein | -1.56 |
| 257965_at | At3g19900 | expressed protein | -1.56 |
| 248899_at | At5g46390 | peptidase S41 (ctpA) | -1.56 |
| 250061_at | At5g17710 | co-chaperone grpE | -1.56 |
| 261118_at | At1g75460 | ATP-dependent protease La (LON) domain-containing protein | -1.56 |
| 256088_at | At1g20810 | immunophilin | -1.56 |
| 255482_at | At4g02510 | chloroplast outer membrane protein, putative | -1.56 |
| 252272_at | At3g49670 | leucine-rich repeat transmembrane protein kinase, putative | -1.57 |
| 260431_at | At1g68190 | zinc finger (B-box type) | -1.57 |
| 258494_at | At3g02450 | cell division protein ftsH, putative | -1.57 |
| 258375_at | At3g17470 | RelA/SpoT domain-containing protein | -1.57 |
| 255635_at | At4g00720 | shaggy-related protein kinase theta (ASK8) | -1.57 |
| 250730_at | At5g06490 | zinc finger (C3HC4-type RING finger) | -1.57 |
| 247139_at | At5g66090 | expressed protein | -1.57 |
| 256223_at | At1g56200 | expressed protein | -1.57 |
| 251120_at | At3g63490 | ribosomal protein L1 | -1.57 |
| 245278_at | At4g17730 | syntaxin 23 (SYP23) | -1.57 |
| 257769_at | At3g23050 | indoleacetic acid-induced protein 7 (IAA7) | -1.57 |
| 266971_at | At2g39580 | expressed protein | -1.57 |
| 259591_at | At1g28150 | expressed protein | -1.57 |
| 256655_at | At3g18890 | expressed protein | -1.57 |
| 262712_at | At1g16460 | mercaptopyruvate sulfurtransferase (MST2) | -1.57 |
| 245593_at | At4g14550 | auxin-responsive AUX/IAA | -1.58 |
| 266120_at | At2g02070 | zinc finger (C2H2 type) | -1.58 |
| 250694_at | At5g06710 | homeobox-leucine zipper protein 14 (HAT14) | -1.58 |
| 264901_at | At1g23090 | sulfate transporter, putative | -1.58 |
| 256728_at | At3g25660 | glutamyl-tRNA(Gln) amidotransferase, putative | -1.58 |
| 258927_at | At3g10160 | folypolyglutamate synthetase (DHFS/FPGS3) | -1.58 |
| 255440_at | At4g02530 | chloroplast thylakoid lumen protein | -1.58 |
| 264844_at | At1g03520 | glycosyltransferase family 14 protein | -1.58 |
| 262693_at | At1g62780 | expressed protein | -1.58 |
| 265673_at | At2g32090 | lactoylglutathione lyase | -1.58 |
| 266882_at | At2g44670 | senescence-associated protein-related | -1.58 |
| 263420_at | At2g17240 | expressed protein | -1.58 |
| 258281_at | At3g26900 | shikimate kinase | -1.58 |
| 246158_at | At5g19855 | expressed protein | -1.58 |
| 251701_at | At3g56650 | thylakoid lumenal 20 kDa protein | -1.58 |
| 251227_at | At3g62700 | glutathione-conjugate transporter, putative | -1.58 |
| 264985_at | At1g27150 | expressed protein | -1.58 |
| 266922_s_at | At2g45950 | SKP1 | -1.58 |
| 265918_at | At2g15090 | fatty acid elongase, putative | -1.58 |
| 263556_at | At2g16350 | F-box | -1.58 |
| 266224_at | At2g28800 | chloroplast membrane protein (ALBINO3) | -1.58 |
| 252975_s_at | At4g38430 | expressed protein | -1.58 |

| | | | |
|-----------|-----------|---|-------|
| 251117_at | At3g63390 | expressed protein | -1.58 |
| 251122_at | At5g01020 | protein kinase | -1.58 |
| 261611_at | At1g49730 | protein kinase | -1.59 |
| 255193_at | At4g07400 | F-box (FBL8) | -1.59 |
| 253251_at | At4g34730 | ribosome-binding factor A | -1.59 |
| 261519_at | At1g71810 | ABC1 | -1.59 |
| 259859_at | At1g68410 | protein phosphatase 2C-related | -1.59 |
| 247130_at | At5g66180 | expressed protein | -1.59 |
| 244975_at | psbH | psbH | -1.59 |
| 263699_at | At1g31120 | potassium transporter | -1.59 |
| 252659_at | At3g44430 | expressed protein | -1.59 |
| 255830_at | At2g33340 | transducin | -1.59 |
| 263980_at | At2g42770 | peroxisomal membrane 22 kDa | -1.59 |
| 256115_at | At1g16880 | uridylyltransferase-related | -1.59 |
| 251744_at | At3g56010 | expressed protein | -1.59 |
| 257773_at | At3g29185 | expressed protein | -1.59 |
| 250371_at | At5g11450 | oxygen-evolving complex-related | -1.59 |
| 247216_at | At5g64860 | 4-alpha-glucanotransferase, putative | -1.59 |
| 246308_at | At3g51820 | chlorophyll synthetase, putative | -1.59 |
| 264300_at | At1g78670 | gamma-glutamyl hydrolase, putative | -1.59 |
| 245020_at | petA | petA | -1.6 |
| 267549_at | At2g32640 | expressed protein | -1.6 |
| 254041_at | At4g25830 | integral membrane | -1.6 |
| 247979_at | At5g56750 | Ndr | -1.6 |
| 245668_at | At1g28330 | dormancy-associated protein, putative (DRM1) | -1.6 |
| 255088_at | At4g09350 | DNAJ heat shock N-terminal domain-containing protein | -1.6 |
| 263763_at | At2g21385 | expressed protein | -1.6 |
| 257647_at | At3g25805 | expressed protein | -1.6 |
| 248459_at | At5g51020 | expressed protein | -1.6 |
| 252409_at | At3g47650 | bundle-sheath defective protein 2 family | -1.6 |
| 247320_at | At5g64040 | photosystem I reaction center subunit PSI-N, chloroplast, putative (PSAN) | -1.6 |
| 257966_at | At3g19800 | expressed protein | -1.6 |
| 267061_at | At2g32480 | membrane-associated zinc metalloprotease, putative | -1.6 |
| 262505_at | At1g21680 | expressed protein | -1.6 |
| 259118_at | At3g01310 | expressed protein | -1.6 |
| 249224_at | At5g42130 | mitochondrial substrate carrier | -1.6 |
| 256530_at | At1g33290 | sporulation protein-related | -1.6 |
| 267088_at | At2g38140 | chloroplast 30S ribosomal protein S31 (PSRP4) | -1.6 |
| 247889_at | At5g57930 | expressed protein | -1.6 |
| 245244_at | At1g44350 | IAA-amino acid hydrolase 6, putative (ILL6) | -1.61 |
| 246011_at | At5g08330 | TCP family transcription factor, putative | -1.61 |
| 254370_at | At4g21750 | L1 specific homeobox gene (ML1) | -1.61 |
| 262539_at | At1g17200 | integral membrane | -1.61 |
| 256873_at | At3g26310 | cytochrome P450 | -1.61 |

| | | | |
|-------------|-----------|--|-------|
| 249583_at | At5g37770 | calmodulin-related protein 2, touch-induced (TCH2) | -1.61 |
| 258800_at | At3g04550 | expressed protein | -1.61 |
| 255077_at | At4g09150 | T-complex protein 11 | -1.61 |
| 258468_at | At3g06070 | expressed protein | -1.61 |
| 252412_at | At3g47295 | expressed protein | -1.61 |
| 263442_at | At2g28605 | expressed protein | -1.61 |
| 257666_at | At3g20270 | lipid-binding serum glycoprotein | -1.61 |
| 245561_at | At4g15500 | UDP-glucuronosyl/UDP-glucosyl transferase | -1.61 |
| 250293_s_at | At5g13360 | auxin-responsive GH3 | -1.61 |
| 262089_s_at | At1g55980 | expressed protein | -1.61 |
| 261081_at | At1g07350 | transformer serine/arginine-rich ribonucleoprotein, putative | -1.61 |
| 260089_at | At1g73170 | expressed protein | -1.61 |
| 251391_at | At3g60910 | expressed protein | -1.61 |
| 265495_at | At2g15695 | expressed protein | -1.62 |
| 262355_at | At1g72820 | mitochondrial substrate carrier | -1.62 |
| 267477_at | At2g02710 | PAC motif-containing protein | -1.62 |
| 264700_at | At1g70100 | expressed protein | -1.62 |
| 258133_at | At3g24500 | ethylene-responsive transcriptional coactivator, putative | -1.62 |
| 266219_at | At2g28880 | para-aminobenzoate (PABA) synthase | -1.62 |
| 259316_at | At3g01175 | expressed protein | -1.62 |
| 261635_at | At1g50020 | expressed protein | -1.62 |
| 262884_at | At1g64720 | expressed protein | -1.62 |
| 254580_at | At4g19390 | expressed protein | -1.62 |
| 257895_at | At3g16950 | lipamide dehydrogenase 1 (PTLPD1) | -1.62 |
| 244950_at | cox2 | cox2 | -1.62 |
| 260872_at | At1g21350 | expressed protein | -1.62 |
| 263726_at | At2g13610 | ABC transporter | -1.62 |
| 262287_at | At1g68660 | expressed protein | -1.62 |
| 251969_at | At3g53130 | cytochrome P450 | -1.62 |
| 265472_at | At2g15580 | zinc finger (C3HC4-type RING finger) | -1.62 |
| 263374_at | At2g20560 | DNAJ heat shock | -1.62 |
| 254298_at | At4g22890 | expressed protein | -1.62 |
| 267294_at | At2g23670 | expressed protein | -1.62 |
| 252957_at | At4g38680 | cold-shock DNA-binding | -1.62 |
| 263755_at | At2g21340 | salicylic acid induction deficient protein, putative | -1.63 |
| 259625_at | At1g42970 | glyceraldehyde-3-phosphate dehydrogenase B, chloroplast (GAPB) | -1.63 |
| 257800_at | At3g15900 | expressed protein | -1.63 |
| 256700_at | At3g52260 | pseudouridine synthase | -1.63 |
| 245397_at | At4g14560 | indoleacetic acid-induced protein 1 (IAA1) | -1.63 |
| 256697_at | At3g20660 | organic cation transporter | -1.63 |
| 264930_at | At1g60800 | leucine-rich repeat | -1.63 |
| 245449_at | At4g16870 | At4g16870 | -1.63 |
| 254117_at | At4g24750 | expressed protein | -1.63 |
| 251328_at | At3g61600 | BTB/POZ domain-containing protein | -1.63 |

| | | | |
|-------------|-----------|--|-------|
| 254545_at | At4g19830 | immunophilin | -1.63 |
| 263471_at | At2g31890 | expressed protein | -1.63 |
| 264673_at | At1g09795 | ATP phosphoribosyl transferase 2 (ATP-PRT2) | -1.63 |
| 256156_at | At3g08510 | phosphoinositide-specific phospholipase C (PLC2) | -1.63 |
| 254187_at | At4g23890 | expressed protein | -1.63 |
| 252325_at | At3g48560 | acetohydroxy-acid synthase (ALS) | -1.63 |
| 257171_at | At3g23760 | expressed protein | -1.63 |
| 246284_at | At4g36780 | brassinosteroid signalling positive regulator-related | -1.63 |
| 250262_at | At5g13410 | immunophilin | -1.63 |
| 262598_at | At1g15260 | expressed protein | -1.64 |
| 253585_at | At4g30720 | expressed protein | -1.64 |
| 256458_at | At1g75220 | integral membrane protein, putative | -1.64 |
| 251330_at | At3g61550 | zinc finger (C3HC4-type RING finger) | -1.64 |
| 247734_at | At5g59400 | expressed protein | -1.64 |
| 244982_at | rpl14 | rpl14 | -1.64 |
| 249775_at | At5g24160 | squalene monooxygenase 1,2 (SQP1,2) | -1.64 |
| 257288_at | At3g29670 | transferase | -1.64 |
| 246509_at | At5g16715 | tRNA synthetase class I (I, L, M and V) | -1.64 |
| 246654_s_at | At5g35210 | peptidase M50 | -1.64 |
| 251395_at | At2g45470 | fasciclin-like arabinogalactan-protein (FLA8) | -1.64 |
| 264709_at | At1g09770 | myb family transcription factor | -1.64 |
| 254068_at | At4g25450 | ABC transporter | -1.64 |
| 258156_at | At3g18050 | expressed protein | -1.64 |
| 247783_at | At5g58800 | quinone reductase | -1.64 |
| 245347_at | At4g14890 | ferredoxin | -1.64 |
| 263846_at | At2g36990 | RNA polymerase sigma subunit SigF (sigF) | -1.64 |
| 260388_at | At1g74070 | peptidyl-prolyl cis-trans isomerase cyclophilin-type | -1.64 |
| 264437_at | At1g27510 | expressed protein | -1.64 |
| 250016_at | At5g18100 | copper/zinc superoxide dismutase (CSD3) | -1.64 |
| 254999_at | At4g09830 | expressed protein | -1.64 |
| 250867_at | At5g03880 | expressed protein | -1.64 |
| 256982_at | At3g13460 | expressed protein | -1.64 |
| 259822_at | At1g66230 | myb family transcription factor (MYB20) | -1.65 |
| 247439_at | At5g62670 | ATPase, plasma membrane-type, putative | -1.65 |
| 265321_at | At2g18280 | TULP2: tubby-like protein 2 (TULP2) | -1.65 |
| 259499_at | At1g15730 | PRLI-interacting factor L, putative | -1.65 |
| 261873_at | At1g11350 | S-locus lectin protein kinase | -1.65 |
| 261732_at | At1g47770 | hypothetical protein | -1.65 |
| 248285_at | At5g52960 | expressed protein | -1.65 |
| 266608_at | At2g35500 | shikimate kinase-related | -1.65 |
| 251516_s_at | At3g59310 | expressed protein | -1.65 |
| 253342_at | At4g33520 | metal-transporting P-type ATPase, putative (PAA1) | -1.65 |
| 251118_at | At3g63410 | chloroplast inner envelope membrane protein, putative (APG1) | -1.65 |
| 250824_at | At5g05200 | ABC1 | -1.65 |

| | | | |
|-----------|-----------|--|-------|
| 254126_at | At4g24770 | RNA-binding protein cp31, putative | -1.65 |
| 261351_at | At1g79790 | haloacid dehalogenase-like hydrolase | -1.65 |
| 245790_at | At1g32200 | glycerol-3-phosphate acyltransferase, chloroplast (ATS1) | -1.65 |
| 246319_at | At3g56680 | expressed protein | -1.65 |
| 262489_at | At1g21830 | expressed protein | -1.66 |
| 257236_at | At3g15095 | expressed protein | -1.66 |
| 255522_at | At4g02260 | RelA/SpoT protein, putative (RSH1) | -1.66 |
| 251720_at | At3g56160 | expressed protein | -1.66 |
| 250267_at | At5g12930 | expressed protein | -1.66 |
| 258038_at | At3g21260 | glycolipid transfer protein-related | -1.66 |
| 257485_at | At1g63580 | protein kinase-related | -1.66 |
| 255779_at | At1g18650 | glycosyl hydrolase 17 | -1.66 |
| 261821_at | At1g11530 | thioredoxin | -1.66 |
| 248104_at | At5g55250 | S-adenosyl-L-methionine:carboxyl methyltransferase | -1.66 |
| 261629_at | At1g49980 | UMUC-like DNA repair | -1.66 |
| 262634_at | At1g06690 | aldo/keto reductase | -1.66 |
| 260685_at | At1g17650 | 6-phosphogluconate dehydrogenase NAD-binding domain-containing protein | -1.66 |
| 267397_at | At1g76170 | expressed protein | -1.66 |
| 254105_at | At4g25080 | magnesium-protoporphyrin O-methyltransferase, putative | -1.66 |
| 246005_at | At5g08415 | lipoic acid synthase | -1.66 |
| 262648_at | At1g14030 | ribulose-1,5 biphosphate carboxylase oxygenase large subunit N-methyltransferase, putative | -1.66 |
| 261295_at | At1g48450 | expressed protein | -1.66 |
| 250886_at | At5g04440 | expressed protein | -1.66 |
| 265494_at | At2g15680 | calmodulin-related protein, putative | -1.67 |
| 262190_at | At1g78030 | expressed protein | -1.67 |
| 251225_at | At3g62660 | glycosyl transferase family 8 protein | -1.67 |
| 251491_at | At3g59480 | pfkB-type carbohydrate kinase | -1.67 |
| 257235_at | At3g15060 | Ras-related GTP-binding | -1.67 |
| 262370_at | At1g73090 | expressed protein | -1.67 |
| 256383_at | At1g66820 | glycine-rich protein | -1.67 |
| 246069_at | At5g20220 | zinc knuckle (CCHC-type) | -1.67 |
| 248502_at | At5g50450 | zinc finger (MYND type) | -1.67 |
| 246748_at | At5g27730 | expressed protein | -1.67 |
| 253391_at | At4g32590 | ferredoxin-related | -1.67 |
| 264307_at | At1g61900 | expressed protein | -1.67 |
| 267606_at | At2g26640 | beta-ketoacyl-CoA synthase, putative | -1.67 |
| 256516_at | At1g66150 | leucine-rich repeat protein kinase, putative (TMK1) | -1.68 |
| 256672_at | At3g52310 | ABC transporter | -1.68 |
| 264371_at | At1g12090 | protease inhibitor/seed storage/lipid transfer protein (LTP) | -1.68 |
| 256757_at | At3g25620 | ABC transporter | -1.68 |
| 258745_at | At3g05920 | heavy-metal-associated domain-containing protein | -1.68 |
| 245632_at | At1g25290 | rhomboid | -1.68 |
| 256542_at | At1g42550 | expressed protein | -1.68 |

| | | | |
|-------------|-----------|---|-------|
| 246702_at | At5g28050 | cytidine/deoxycytidylate deaminase | -1.68 |
| 265024_at | At1g24600 | expressed protein | -1.68 |
| 248283_at | At5g52920 | pyruvate kinase, putative | -1.68 |
| 255692_at | At4g00400 | phospholipid/glycerol acyltransferase | -1.68 |
| 249785_at | At5g24300 | starch synthase, putative | -1.68 |
| 249900_at | At5g22640 | MORN (Membrane Occupation and Recognition Nexus) repeat-containing protein | -1.68 |
| 266045_s_at | At2g07727 | cytochrome b (MTCYB) | -1.68 |
| 245948_at | At5g19540 | expressed protein | -1.68 |
| 258607_at | At3g02730 | thioredoxin, putative | -1.68 |
| 256792_at | At3g22150 | pentatricopeptide (PPR) repeat-containing protein | -1.68 |
| 252724_at | At3g43540 | expressed protein | -1.68 |
| 246110_at | At5g20140 | SOUL heme-binding | -1.68 |
| 262172_at | At1g74970 | ribosomal protein S9 (RPS9) | -1.68 |
| 248235_at | At5g53860 | expressed protein | -1.68 |
| 246547_at | At5g14970 | expressed protein | -1.68 |
| 253624_at | At4g30580 | phospholipid/glycerol acyltransferase | -1.68 |
| 250369_at | At5g11300 | cyclin, putative (CYC3b) | -1.69 |
| 257563_at | At3g19610 | hypothetical protein | -1.69 |
| 267027_at | At2g38330 | MATE efflux | -1.69 |
| 263912_at | At2g36390 | starch branching enzyme class II (SBE2-1) | -1.69 |
| 247044_at | At5g66850 | protein kinase | -1.69 |
| 262954_at | At1g54500 | rubredoxin | -1.69 |
| 253412_at | At4g33000 | calcineurin B-like protein 10 (CBL10) | -1.69 |
| 245797_at | At1g45261 | defective chloroplasts and leaves protein-related (DCL) | -1.69 |
| 258472_at | At3g06080 | expressed protein | -1.69 |
| 251610_at | At3g57930 | expressed protein | -1.69 |
| 256324_at | At1g66760 | MATE efflux | -1.7 |
| 258565_at | At3g04350 | expressed protein | -1.7 |
| 248336_at | At5g52420 | expressed protein | -1.7 |
| 254635_at | At4g18670 | leucine-rich repeat | -1.7 |
| 262845_at | At1g14740 | expressed protein | -1.7 |
| 265680_at | At2g32150 | haloacid dehalogenase-like hydrolase | -1.7 |
| 257672_at | At3g20300 | expressed protein | -1.7 |
| 264837_at | At1g03600 | photosystem II | -1.7 |
| 255926_at | At1g22190 | AP2 domain-containing transcription factor, putative | -1.7 |
| 253476_at | At4g32300 | lectin protein kinase | -1.7 |
| 249978_at | At5g18850 | expressed protein | -1.7 |
| 245000_at | ycf6 | ycf6 | -1.7 |
| 266267_at | At2g29460 | glutathione S-transferase, putative | -1.71 |
| 261633_at | At1g49930 | hypothetical protein | -1.71 |
| 267635_at | At2g42220 | rhodanese-like domain-containing protein | -1.71 |
| 263198_at | At1g53990 | GDSL-motif lipase/hydrolase | -1.71 |
| 246222_at | At4g36900 | AP2 domain-containing protein RAP2.10 (RAP2.10) | -1.71 |
| 262797_at | At1g20840 | transporter-related | -1.71 |

| | | | |
|-------------|-----------|---|-------|
| 266319_s_at | At2g46720 | fatty acid elongase 3-ketoacyl-CoA synthase, putative | -1.71 |
| 253688_at | At4g29590 | expressed protein | -1.71 |
| 261924_at | At1g22550 | proton-dependent oligopeptide transport (POT) | -1.71 |
| 254770_at | At4g13340 | leucine-rich repeat | -1.71 |
| 246432_at | At5g17490 | gibberellin response modulator, putative | -1.71 |
| 258929_at | At3g10060 | immunophilin, putative | -1.71 |
| 255623_at | At4g01310 | ribosomal protein L5 | -1.71 |
| 260967_at | At1g12230 | transaldolase, putative | -1.71 |
| 253302_at | At4g33660 | expressed protein | -1.71 |
| 247525_at | At5g61380 | ABI3-interacting protein 1 (AIP1) | -1.71 |
| 261577_at | At1g01080 | 33 kDa ribonucleoprotein, chloroplast, putative | -1.71 |
| 248669_at | At5g48730 | pentatricopeptide (PPR) repeat-containing protein | -1.72 |
| 251670_at | At3g57190 | peptide chain release factor, putative | -1.72 |
| 244981_at | rps8 | rps8 | -1.72 |
| 245761_at | At1g66890 | expressed protein | -1.72 |
| 265675_at | At2g32120 | heat shock protein 70 (HSP70) | -1.72 |
| 249542_at | At5g38140 | histone-like transcription factor (CBF/NF-Y) | -1.72 |
| 246170_at | At5g32450 | RNA recognition motif (RRM)-containing protein | -1.72 |
| 253548_at | At4g30993 | expressed protein | -1.72 |
| 255617_at | At4g01330 | protein kinase | -1.72 |
| 264177_at | At1g02150 | pentatricopeptide (PPR) repeat-containing protein | -1.72 |
| 246159_at | At5g20935 | expressed protein | -1.72 |
| 253160_at | At4g35760 | expressed protein | -1.72 |
| 267644_s_at | At2g32880 | meprin and TRAF homology domain-containing protein | -1.73 |
| 258269_at | At3g15690 | biotin carboxyl carrier protein of acetyl-CoA carboxylase-related | -1.73 |
| 253335_at | At4g33500 | protein phosphatase 2C-related | -1.73 |
| 264528_at | At1g30810 | zinc finger (C5HC2 type) | -1.73 |
| 260134_at | At1g66370 | myb family transcription factor (MYB113) | -1.73 |
| 245383_at | At4g17810 | zinc finger (C2H2 type) | -1.73 |
| 254638_at | At4g18740 | expressed protein | -1.73 |
| 245370_at | At4g16840 | expressed protein | -1.73 |
| 262114_at | At1g02860 | SPX domain-containing protein | -1.73 |
| 258087_at | At3g26060 | peroxiredoxin Q, putative | -1.73 |
| 255982_at | At1g34000 | light stress-responsive one-helix protein (OHP2) | -1.73 |
| 253758_at | At4g29060 | elongation factor Ts | -1.73 |
| 249101_at | At5g43580 | protease inhibitor, putative | -1.73 |
| 266483_at | At2g47910 | expressed protein | -1.73 |
| 248578_at | At5g49820 | expressed protein | -1.73 |
| 262202_at | At2g01110 | thylakoid membrane formation protein (APG2) | -1.73 |
| 249125_at | At5g43450 | 2-oxoglutarate-dependent dioxygenase, putative | -1.74 |
| 248075_at | At5g55740 | pentatricopeptide (PPR) repeat-containing protein | -1.74 |
| 258359_s_at | At3g14415 | (S)-2-hydroxy-acid oxidase, peroxisomal, putative | -1.74 |
| 248622_at | At5g49360 | glycosyl hydrolase family 3 protein | -1.74 |
| 255436_at | At4g03150 | expressed protein | -1.74 |

| | | | |
|-------------|-----------|--|-------|
| 252132_at | At3g50790 | late embryogenesis abundant protein, putative (LEA) | -1.74 |
| 255059_at | At4g09420 | disease resistance protein (TIR-NBS class), putative | -1.74 |
| 246408_at | At1g57680 | expressed protein | -1.74 |
| 256440_at | At3g10970 | haloacid dehalogenase-like hydrolase | -1.74 |
| 252473_s_at | At3g46610 | pentatricopeptide (PPR) repeat-containing protein | -1.74 |
| 261622_at | At1g01970 | pentatricopeptide (PPR) repeat-containing protein | -1.74 |
| 267130_at | At2g23390 | expressed protein | -1.75 |
| 245885_at | At5g09440 | phosphate-responsive protein, putative | -1.75 |
| 246200_at | At4g37240 | expressed protein | -1.75 |
| 261583_at | At1g01090 | pyruvate dehydrogenase E1 component alpha subunit, chloroplast | -1.75 |
| 260567_at | At2g43820 | UDP-glucuronosyl/UDP-glucosyl transferase | -1.75 |
| 252859_at | At4g39780 | AP2 domain-containing transcription factor, putative | -1.75 |
| 257723_at | At3g18500 | nocturnin-related | -1.75 |
| 262322_at | At1g27590 | expressed protein | -1.75 |
| 264158_at | At1g65260 | PspA/IM30 | -1.75 |
| 256595_x_at | At3g28530 | expressed protein | -1.75 |
| 260465_at | At1g10910 | pentatricopeptide (PPR) repeat-containing protein | -1.75 |
| 253337_at | At4g33470 | histone deacetylase | -1.75 |
| 267152_at | At2g31040 | ATP synthase protein I -related | -1.75 |
| 260856_at | At1g21910 | AP2 domain-containing transcription factor | -1.75 |
| 248398_at | At5g51970 | sorbitol dehydrogenase, putative | -1.75 |
| 260542_at | At2g43560 | immunophilin | -1.75 |
| 264920_at | At1g60550 | naphthoate synthase, putative | -1.75 |
| 253028_at | At4g38160 | mitochondrial transcription termination factor-related | -1.75 |
| 266570_at | At2g24090 | ribosomal protein L35 | -1.76 |
| 263391_at | At2g11810 | 1,2-diacylglycerol 3-beta-galactosyltransferase, putative | -1.76 |
| 248516_at | At5g50540 | expressed protein | -1.76 |
| 248585_at | At5g49640 | expressed protein | -1.76 |
| 245399_at | At4g17340 | major intrinsic protein (MIP) | -1.76 |
| 265974_at | At2g11260 | Hypothetical protein, complete cds, clone: RAFL16-43-P18 | -1.76 |
| 259965_at | At1g53670 | transcription factor-related | -1.76 |
| 267005_at | At2g34460 | flavin reductase-related | -1.76 |
| 264435_at | At1g10360 | glutathione S-transferase, putative | -1.76 |
| 267505_at | At2g45560 | cytochrome P450 | -1.76 |
| 262168_at | At1g74730 | expressed protein | -1.76 |
| 265966_at | At2g37220 | 29 kDa ribonucleoprotein, chloroplast, putative | -1.76 |
| 261666_at | At1g18440 | peptidyl-tRNA hydrolase | -1.77 |
| 259970_at | At1g76570 | chlorophyll A-B binding | -1.77 |
| 249383_at | At5g39860 | bHLH protein | -1.77 |
| 264037_at | At2g03750 | sulfotransferase | -1.77 |
| 258742_at | At3g05800 | expressed protein | -1.77 |
| 265741_at | At2g01320 | ABC transporter | -1.77 |
| 256698_at | At3g20680 | expressed protein | -1.77 |
| 245357_at | At4g17560 | ribosomal protein L19 | -1.77 |

| | | | |
|-------------|-----------|---|-------|
| 252563_at | At3g45970 | expansin (EXPL1) | -1.77 |
| 265021_at | At1g24610 | SET domain-containing protein | -1.77 |
| 266673_at | At2g29630 | thiamine biosynthesis | -1.78 |
| 260015_at | At1g67980 | caffeoyl-CoA 3-O-methyltransferase, putative | -1.78 |
| 245015_at | rbcl | rbcl | -1.78 |
| 266636_at | At2g35370 | glycine cleavage system H protein 1 (GDCSH) | -1.78 |
| 265022_at | At1g24520 | anther-specific protein agp1 | -1.78 |
| 244977_at | petD | petD | -1.78 |
| 250906_at | At5g03650 | starch branching enzyme class II (SBE2-2) | -1.78 |
| 253076_at | At4g36160 | no apical meristem (NAM) | -1.78 |
| 267201_at | At2g31010 | protein kinase | -1.78 |
| 261078_at | At1g07320 | 50S ribosomal protein L4, chloroplast (CL4) | -1.78 |
| 248798_at | At5g47190 | ribosomal protein L19 | -1.78 |
| 262850_at | At1g14920 | gibberellin response modulator (GAI) | -1.78 |
| 254727_at | At4g13670 | peptidoglycan-binding domain-containing protein | -1.78 |
| 265967_at | At2g37450 | nodulin MtN21 | -1.78 |
| 248402_at | At5g52100 | dihydrodipicolinate reductase | -1.78 |
| 247034_at | At5g67260 | cyclin | -1.78 |
| 255082_at | At4g09160 | SEC14 cytosolic factor | -1.79 |
| 259511_at | At1g12520 | superoxide dismutase copper chaperone, putative | -1.79 |
| 257794_at | At3g27050 | expressed protein | -1.79 |
| 246001_at | At5g20790 | expressed protein | -1.79 |
| 256836_at | At3g22960 | pyruvate kinase, putative | -1.79 |
| 250668_at | At5g07020 | proline-rich | -1.79 |
| 265394_at | At2g20725 | CAAX amino terminal protease | -1.79 |
| 260367_at | At1g69760 | expressed protein | -1.79 |
| 255540_at | At4g01800 | preprotein translocase secA subunit, putative | -1.79 |
| 251820_at | At3g55040 | ln2-1 protein, putative | -1.79 |
| 247347_at | At5g63780 | zinc finger (C3HC4-type RING finger) | -1.79 |
| 261767_s_at | At1g15500 | chloroplast ADP, ATP carrier protein, putative | -1.79 |
| 255447_at | At4g02790 | GTP-binding | -1.79 |
| 253537_at | At4g31560 | expressed protein | -1.79 |
| 249869_at | At5g23050 | acyl-activating enzyme 17 (AAE17) | -1.79 |
| 256754_at | At3g25690 | hydroxyproline-rich glycoprotein | -1.79 |
| 252979_at | At4g38225 | expressed protein | -1.79 |
| 256856_at | At3g15110 | expressed protein | -1.79 |
| 253208_at | At4g34820 | pentatricopeptide (PPR) repeat-containing protein | -1.8 |
| 251195_at | At3g62930 | glutaredoxin | -1.8 |
| 262115_at | At1g02813 | expressed protein | -1.8 |
| 262566_at | At1g34310 | transcriptional factor B3 | -1.8 |
| 258263_at | At3g15780 | expressed protein | -1.8 |
| 265417_at | At2g20920 | expressed protein | -1.8 |
| 264041_at | At2g03710 | MADS-box protein (AGL3) | -1.8 |
| 258860_at | At3g02050 | potassium transporter (KUP3) | -1.8 |
| 265724_at | At2g32100 | ovate protein-related | -1.8 |

| | | | |
|-------------|-----------|---|-------|
| 254794_at | At4g12970 | expressed protein | -1.8 |
| 266460_at | At2g47930 | hydroxyproline-rich glycoprotein | -1.8 |
| 265175_at | At1g23480 | glycosyl transferase family 2 protein | -1.8 |
| 262368_at | At1g73060 | expressed protein | -1.8 |
| 264424_at | At1g61740 | expressed protein | -1.8 |
| 259983_at | At1g76490 | 3-hydroxy-3-methylglutaryl-CoA reductase 1 (HMG1) | -1.8 |
| 249899_at | At5g22620 | phosphoglycerate/bisphosphoglycerate mutase | -1.8 |
| 253322_at | At4g33980 | expressed protein | -1.8 |
| 264508_at | At1g09570 | phytochrome A (PHYA) | -1.81 |
| 261926_at | At1g22530 | SEC14 cytosolic factor | -1.81 |
| 248606_at | At5g49450 | bZIP family transcription factor | -1.81 |
| 266251_s_at | At2g27540 | expressed protein | -1.81 |
| 245047_at | psbA | psbA | -1.81 |
| 263674_at | At2g04790 | expressed protein | -1.81 |
| 251719_at | At3g56140 | expressed protein | -1.81 |
| 247100_at | At5g66520 | pentatricopeptide (PPR) repeat-containing protein | -1.81 |
| 245388_at | At4g16410 | expressed protein | -1.81 |
| 250503_at | At5g09820 | plastid-lipid associated protein PAP | -1.81 |
| 263048_s_at | At2g05310 | expressed protein | -1.81 |
| 265454_at | At2g46530 | transcriptional factor B3 | -1.81 |
| 246847_at | At5g26820 | ferroportin-related | -1.81 |
| 244973_at | psbT | psbT | -1.81 |
| 262369_at | At1g73010 | expressed protein | -1.81 |
| 249148_at | At5g43260 | chaperone protein dnaJ-related | -1.82 |
| 245726_at | At1g73360 | homeobox-leucine zipper | -1.82 |
| 266892_at | At2g26080 | glycine dehydrogenase (decarboxylating), putative | -1.82 |
| 245925_at | At5g28770 | bZIP transcription factor | -1.82 |
| 261518_at | At1g71695 | peroxidase 12 (PER12) | -1.82 |
| 247880_at | At5g57780 | expressed protein | -1.82 |
| 245798_at | At1g45545 | hypothetical protein | -1.82 |
| 261265_at | At1g26800 | zinc finger (C3HC4-type RING finger) | -1.82 |
| 250268_s_at | At5g12950 | expressed protein | -1.82 |
| 257033_at | At3g19170 | peptidase | -1.82 |
| 246075_at | At5g20410 | 1,2-diacylglycerol 3-beta-galactosyltransferase, putative | -1.82 |
| 263111_s_at | At1g65190 | protein kinase | -1.82 |
| 252215_at | At3g50240 | kinesin motor protein-related | -1.83 |
| 257253_at | At3g24190 | ABC1 | -1.83 |
| 246028_at | At5g21170 | 5'-AMP-activated protein kinase beta-2 subunit, putative | -1.83 |
| 264483_at | At1g77230 | AMP-binding protein, putative | -1.83 |
| 244901_at | orf25 | orf25 | -1.83 |
| 264575_at | At1g05190 | ribosomal protein L6 | -1.83 |
| 262377_at | At1g73110 | ribulose biphosphate carboxylase/oxygenase activase, putative | -1.83 |
| 246237_at | At4g36390 | radical SAM domain-containing protein | -1.83 |
| 261338_at | At1g44920 | expressed protein | -1.83 |

| | | | |
|-------------|-----------|--|-------|
| 256417_s_at | At3g11170 | omega-3 fatty acid desaturase, chloroplast (FAD7) | -1.83 |
| 260840_at | At1g29050 | expressed protein | -1.84 |
| 245794_at | At1g32170 | xyloglucan:xyloglucosyl transferase, putative (XTR4) | -1.84 |
| 265400_at | At2g10940 | protease inhibitor/seed storage/lipid transfer protein (LTP) | -1.84 |
| 251421_at | At3g60510 | enoyl-CoA hydratase | -1.84 |
| 245005_at | rps14 | rps14 | -1.84 |
| 244965_at | ORF31 | ORF31 | -1.84 |
| 244936_at | ndhA | ndhA | -1.84 |
| 247201_at | At5g65220 | ribosomal protein L29 | -1.84 |
| 256525_at | At1g66180 | aspartyl protease | -1.84 |
| 254413_at | At4g21440 | myb family transcription factor (MYB102) | -1.84 |
| 258622_at | At3g02720 | DJ-1 | -1.84 |
| 251031_at | At5g02120 | thylakoid membrane one helix protein (OHP) | -1.84 |
| 252411_at | At3g47430 | peroxisomal biogenesis factor 11 (PEX11) | -1.84 |
| 247925_at | At5g57560 | xyloglucan:xyloglucosyl transferase (TCH4) | -1.85 |
| 246843_at | At5g26734 | expressed protein | -1.85 |
| 264280_at | At1g61820 | glycosyl hydrolase family 1 protein | -1.85 |
| 263606_at | At2g16280 | very-long-chain fatty acid condensing enzyme, putative | -1.85 |
| 250189_at | At5g14410 | expressed protein | -1.85 |
| 266925_at | At2g45740 | peroxisomal biogenesis factor 11 (PEX11) | -1.85 |
| 244940_at | rps12.2 | rps12.2 | -1.85 |
| 266165_at | At2g28190 | copper/zinc superoxide dismutase (CSD2) | -1.85 |
| 251755_at | At3g55790 | expressed protein | -1.85 |
| 248409_at | At5g51540 | peptidase M3 | -1.85 |
| 254553_at | At4g19530 | disease resistance protein (TIR-NBS-LRR class), putative | -1.85 |
| 249876_at | At5g23060 | expressed protein | -1.85 |
| 256168_at | At1g51805 | leucine-rich repeat protein kinase, putative | -1.85 |
| 248242_at | At5g53580 | aldo/keto reductase | -1.85 |
| 249244_at | At5g42270 | FtsH protease, putative | -1.85 |
| 259193_at | At3g01480 | peptidyl-prolyl cis-trans isomerase, putative | -1.85 |
| 255289_at | At4g04690 | F-box (FBX15) | -1.86 |
| 245010_at | ndhJ | ndhJ | -1.86 |
| 254424_at | At4g21510 | F-box | -1.86 |
| 247745_at | At5g59030 | copper transporter 1 (COPT1) | -1.86 |
| 254985_x_at | At4g10580 | gypsy-like retrotransposon family | -1.86 |
| 260041_at | At1g68780 | leucine-rich repeat | -1.86 |
| 259839_at | At1g52190 | proton-dependent oligopeptide transport (POT) | -1.86 |
| 261488_at | At1g14345 | expressed protein | -1.86 |
| 266355_at | At2g01400 | expressed protein | -1.86 |
| 245806_at | At1g45474 | chlorophyll A-B binding protein, putative (LHCA5) | -1.86 |
| 264546_at | At1g55805 | BolA-like | -1.86 |
| 250745_at | At5g05850 | leucine-rich repeat | -1.87 |
| 251575_at | At3g58120 | bZIP transcription factor | -1.87 |
| 266656_at | At2g25900 | zinc finger (CCCH-type) | -1.87 |
| 252463_at | At3g47070 | expressed protein | -1.87 |

| | | | |
|-------------|-----------|--|-------|
| 261165_at | At1g34430 | dihydrolipoamide S-acetyltransferase, putative | -1.87 |
| 253174_at | At4g35090 | catalase 2 | -1.87 |
| 248380_at | At5g51820 | phosphoglucumutase, chloroplast (PGM) | -1.87 |
| 249658_s_at | At5g36700 | phosphoglycolate phosphatase, putative | -1.87 |
| 247488_at | At5g61820 | expressed protein | -1.87 |
| 253489_at | At4g31780 | monogalactosyldiacylglycerol synthase, putative (MGD1) | -1.87 |
| 249759_at | At5g24380 | transporter, putative | -1.88 |
| 266901_at | At2g34600 | expressed protein | -1.88 |
| 258554_at | At3g06980 | DEAD/DEAH box helicase, putative | -1.88 |
| 263873_at | At2g21860 | violaxanthin de-epoxidase-related | -1.88 |
| 255943_at | At1g22370 | UDP-glucuronosyl/UDP-glucosyl transferase | -1.88 |
| 247278_at | At5g64380 | fructose-1,6-bisphosphatase | -1.88 |
| 256097_at | At1g13670 | expressed protein | -1.88 |
| 266518_at | At2g35170 | MORN (Membrane Occupation and Recognition Nexus) repeat-containing protein | -1.88 |
| 251146_at | At3g63520 | neoxanthin cleavage enzyme (NCED1) | -1.88 |
| 263499_at | At2g42580 | tetratricopeptide repeat (TPR)-containing protein | -1.88 |
| 264513_at | At1g09420 | glucose-6-phosphate 1-dehydrogenase, putative | -1.88 |
| 247266_at | At5g64570 | glycosyl hydrolase family 3 protein | -1.89 |
| 245025_at | atpF | atpF | -1.89 |
| 251142_at | At5g01015 | expressed protein | -1.89 |
| 258386_at | At3g15520 | peptidyl-prolyl cis-trans isomerase (TLP38) | -1.89 |
| 262397_at | At1g49380 | cytochrome c biogenesis protein family | -1.89 |
| 261353_at | At1g79600 | ABC1 | -1.9 |
| 258025_at | At3g19480 | D-3-phosphoglycerate dehydrogenase, putative | -1.9 |
| 246449_at | At5g16810 | expressed protein | -1.9 |
| 267430_at | At2g34860 | chaperone protein dnaJ-related | -1.9 |
| 253373_at | At4g33150 | lysine-ketoglutarate reductase | -1.9 |
| 262784_at | At1g10760 | starch excess protein (SEX1) | -1.9 |
| 266614_at | At2g14910 | expressed protein | -1.9 |
| 263533_at | At2g24820 | Rieske (2Fe-2S) domain-containing protein | -1.9 |
| 262059_at | At1g80030 | DNAJ heat shock protein, putative | -1.9 |
| 249847_at | At5g23210 | serine carboxypeptidase S10 | -1.91 |
| 261801_at | At1g30520 | acyl-activating enzyme 14 (AAE14) | -1.91 |
| 250498_at | At5g09660 | malate dehydrogenase, glyoxysomal | -1.91 |
| 246427_at | At5g17400 | ADP, ATP carrier protein, mitochondrial, putative | -1.91 |
| 266018_at | At2g18710 | preprotein translocase secY subunit, chloroplast (CpSecY) | -1.92 |
| 265959_at | At2g37240 | expressed protein | -1.92 |
| 245559_at | At4g15460 | glycine-rich protein | -1.92 |
| 245701_at | At5g04140 | glutamate synthase (GLU1) | -1.92 |
| 256940_at | At3g30720 | expressed protein | -1.92 |
| 250940_at | At5g03310 | auxin-responsive | -1.93 |
| 260201_at | At1g67600 | expressed protein | -1.93 |
| 245352_at | At4g15490 | UDP-glucuronosyl/UDP-glucosyl transferase | -1.93 |
| 253283_at | At4g34090 | expressed protein | -1.93 |

| | | | |
|-------------|-----------|---|-------|
| 249691_at | At5g36170 | peptide chain release factor, putative | -1.93 |
| 249685_at | At5g36120 | YGGT | -1.93 |
| 267379_at | At2g26340 | expressed protein | -1.93 |
| 265547_at | At2g28305 | expressed protein | -1.94 |
| 247474_at | At5g62280 | expressed protein | -1.94 |
| 254465_at | At4g20420 | tapetum-specific protein-related | -1.94 |
| 259373_at | At1g69160 | expressed protein | -1.94 |
| 254815_at | At4g12420 | multi-copper oxidase, putative (SKU5) | -1.94 |
| 254642_at | At4g18810 | expressed protein | -1.94 |
| 248287_at | At5g52970 | thylakoid lumen 15.0 kDa protein | -1.94 |
| 249174_at | At5g42900 | expressed protein | -1.94 |
| 249510_at | At5g38510 | rhomboid | -1.94 |
| 260547_at | At2g43550 | trypsin inhibitor, putative | -1.95 |
| 257772_at | At3g23080 | expressed protein | -1.95 |
| 246122_at | At5g20380 | transporter-related | -1.95 |
| 244990_s_at | orf77.1 | orf77.1 | -1.95 |
| 251109_at | At5g01600 | ferritin 1 (FER1) | -1.95 |
| 259738_at | At1g64355 | expressed protein | -1.95 |
| 253495_at | At4g31850 | pentatricopeptide (PPR) repeat-containing protein | -1.95 |
| 245730_at | At1g73470 | expressed protein | -1.95 |
| 244966_at | petG | petG | -1.95 |
| 249872_at | At5g23130 | peptidoglycan-binding LysM domain-containing protein | -1.96 |
| 249852_at | At5g23270 | sugar transporter, putative | -1.96 |
| 251461_at | At3g59780 | expressed protein | -1.96 |
| 252876_at | At4g39970 | haloacid dehalogenase-like hydrolase | -1.96 |
| 266509_at | At2g47940 | DegP2 protease (DEGP2) | -1.96 |
| 249378_at | At5g40450 | expressed protein | -1.96 |
| 251584_at | At3g58620 | tetratricopeptide repeat (TPR)-containing protein | -1.96 |
| 266766_at | At2g46880 | calcineurin-like phosphoesterase | -1.97 |
| 254502_at | At4g20130 | ribulose-1,5 biphosphate carboxylase | -1.97 |
| 265628_at | At2g27290 | expressed protein | -1.97 |
| 250073_at | At5g17170 | rubredoxin | -1.97 |
| 256149_at | At1g55110 | zinc finger (C2H2 type) | -1.97 |
| 247943_at | At5g57170 | macrophage migration inhibitory factor (MIF) | -1.98 |
| 246736_at | At5g27560 | expressed protein | -1.98 |
| 246838_at | At5g26675 | endonuclease, putative | -1.98 |
| 259292_at | At3g11560 | expressed protein | -1.98 |
| 265998_at | At2g24270 | NADP-dependent glyceraldehyde-3-phosphate dehydrogenase, putative | -1.98 |
| 259275_at | At3g01060 | expressed protein | -1.99 |
| 253039_at | At4g37760 | squalene monooxygenase, putative | -1.99 |
| 256544_at | At1g42560 | seven transmembrane MLO (MLO9) | -1.99 |
| 264229_at | At1g67480 | kelch repeat-containing F-box | -1.99 |
| 247709_at | At5g59250 | sugar transporter | -1.99 |
| 262473_at | At1g50250 | cell division protein ftsH homolog 1 (FTSH1) | -1.99 |

| | | | |
|-----------|-----------|---|-------|
| 245026_at | atpH | atpH | -1.99 |
| 257831_at | At3g26710 | expressed protein | -1.99 |
| 259981_at | At1g76450 | oxygen-evolving complex-related | -1.99 |
| 258736_at | At3g05900 | neurofilament protein-related | -1.99 |
| 247252_at | At5g64770 | expressed protein | -1.99 |
| 251243_at | At3g61870 | expressed protein | -1.99 |
| 262176_at | At1g74960 | 3-ketoacyl-ACP synthase, putative | -1.99 |
| 263352_at | At2g22080 | expressed protein | -1.99 |
| 251494_at | At3g59350 | serine/threonine protein kinase, putative | -1.99 |
| 260877_at | At1g21500 | expressed protein | -2 |
| 258227_at | At3g15620 | 6-4 photolyase (UVR3) | -2 |
| 261135_at | At1g19610 | plant defensin-fusion protein, putative (PDF1.4) | -2 |
| 245017_at | psal | psal | -2 |
| 245528_at | At4g15530 | pyruvate phosphate dikinase | -2 |
| 256788_at | At3g13730 | cytochrome P450, putative | -2 |
| 256423_at | At1g33540 | serine carboxypeptidase S10 | -2 |
| 262878_at | At1g64770 | expressed protein | -2 |
| 259943_at | At1g71480 | nuclear transport factor 2 (NTF2) | -2 |
| 256130_at | At1g18170 | immunophilin | -2 |
| 253522_at | At4g31290 | ChaC-like | -2 |
| 258114_at | At3g14660 | cytochrome P450, putative | -2.01 |
| 264887_at | At1g23120 | major latex protein-related / MLP-related | -2.01 |
| 253849_at | At4g28080 | expressed protein | -2.01 |
| 256503_at | At1g75250 | myb family transcription factor | -2.01 |
| 259658_at | At1g55370 | expressed protein | -2.01 |
| 248975_at | At5g45040 | cytochrome c6 (ATC6) | -2.01 |
| 251268_at | At3g62350 | hypothetical protein | -2.01 |
| 265867_at | At2g01620 | expressed protein | -2.01 |
| 247954_at | At5g56870 | beta-galactosidase, putative / lactase, putative | -2.01 |
| 254137_at | At4g24930 | thylakoid lumenal 17.9 kDa protein, chloroplast | -2.01 |
| 252181_at | At3g50685 | expressed protein | -2.01 |
| 246268_at | At1g31800 | cytochrome P450 | -2.01 |
| 253547_at | At4g30950 | omega-6 fatty acid desaturase, chloroplast (FAD6) | -2.01 |
| 257381_at | At2g37950 | zinc finger (C3HC4-type RING finger) | -2.01 |
| 249002_at | At5g44520 | ribose 5-phosphate isomerase-related | -2.01 |
| 247348_at | At5g63810 | beta-galactosidase, putative / lactase, putative | -2.01 |
| 247261_at | At5g64460 | expressed protein | -2.01 |
| 246071_at | At5g20150 | SPX domain-containing protein | -2.02 |
| 264580_at | At1g05340 | expressed protein | -2.02 |
| 259791_at | At1g29700 | expressed protein | -2.02 |
| 258935_at | At3g10120 | expressed protein | -2.02 |
| 257172_at | At3g23700 | S1 RNA-binding domain-containing protein | -2.02 |
| 263761_at | At2g21330 | fructose-bisphosphate aldolase, putative | -2.02 |
| 263350_at | At2g13360 | serine-glyoxylate aminotransferase-related | -2.02 |
| 246226_at | At4g37200 | thioredoxin | -2.02 |

| | | | |
|-----------|-----------|---|-------|
| 266357_at | At2g32290 | beta-amylase, putative | -2.02 |
| 255331_at | At4g04330 | expressed protein | -2.02 |
| 252116_at | At3g51510 | expressed protein | -2.02 |
| 253597_at | At4g30690 | translation initiation factor 3 (IF-3) | -2.03 |
| 266277_at | At2g29310 | tropinone reductase, putative | -2.03 |
| 263678_at | At1g04420 | aldo/keto reductase | -2.03 |
| 266617_at | At2g29670 | expressed protein | -2.04 |
| 247884_at | At5g57800 | CER1 protein, putative (WAX2) | -2.04 |
| 245284_at | At4g14210 | phytoene desaturase (PDS) | -2.04 |
| 251192_at | At3g62720 | galactosyl transferase (GMA12) | -2.04 |
| 264799_at | At1g08550 | violaxanthin de-epoxidase precursor, putative (AVDE1) | -2.04 |
| 255939_at | At1g12730 | cell division cycle protein-related | -2.04 |
| 256514_at | At1g66130 | oxidoreductase N-terminal domain-containing protein | -2.04 |
| 245349_at | At4g16690 | esterase | -2.04 |
| 257168_at | At3g24430 | expressed protein | -2.04 |
| 250812_at | At5g04900 | short-chain dehydrogenase/reductase (SDR) | -2.04 |
| 263953_at | At2g36050 | ovate protein-related | -2.04 |
| 253956_at | At4g26700 | fimbrin-like protein (FIM1) | -2.04 |
| 252425_at | At3g47620 | TCP family transcription factor, putative | -2.04 |
| 245011_at | psbG | psbG | -2.05 |
| 261674_at | At1g18270 | ketose-bisphosphate aldolase class-II | -2.05 |
| 257547_at | At3g13000 | expressed protein | -2.05 |
| 259037_at | At3g09350 | beta-catenin repeat | -2.05 |
| 248395_at | At5g52120 | SKP1 interacting partner 3-related | -2.05 |
| 247641_at | At5g60540 | SNO glutamine amidotransferase | -2.05 |
| 249065_at | At5g44260 | zinc finger (CCCH-type) | -2.06 |
| 250777_at | At5g05440 | expressed protein | -2.06 |
| 248273_at | At5g53500 | WD-40 repeat | -2.06 |
| 246596_at | At5g14740 | carbonate dehydratase 2 (CA2) | -2.06 |
| 246958_at | At5g24690 | expressed protein | -2.06 |
| 251306_at | At3g61260 | DNA-binding protein | -2.06 |
| 266904_at | At2g34590 | transketolase | -2.06 |
| 259161_at | At3g01500 | carbonate dehydratase 1 (CA1) | -2.07 |
| 266813_at | At2g44920 | thylakoid lumenal 15 kDa protein, chloroplast | -2.07 |
| 264653_at | At1g08980 | amidase | -2.07 |
| 264394_at | At1g11860 | aminomethyltransferase, putative | -2.07 |
| 256061_at | At1g07040 | expressed protein | -2.07 |
| 244974_at | psbN | psbN | -2.08 |
| 251885_at | At3g54050 | fructose-1,6-bisphosphatase, putative | -2.08 |
| 259015_at | At3g07350 | expressed protein | -2.09 |
| 256914_at | At3g23880 | F-box | -2.09 |
| 245440_at | At4g16680 | RNA helicase, putative | -2.09 |
| 260056_at | At1g78140 | methyltransferase-related | -2.09 |
| 260914_at | At1g02640 | glycosyl hydrolase family 3 protein | -2.1 |
| 263597_at | At2g01870 | expressed protein | -2.1 |

| | | | |
|-------------|-----------|--|-------|
| 263157_at | At1g54100 | aldehyde dehydrogenase, putative | -2.1 |
| 248335_at | At5g52450 | MATE efflux protein-related | -2.1 |
| 261658_at | At1g50040 | expressed protein | -2.1 |
| 252092_at | At3g51420 | strictosidine synthase | -2.1 |
| 265742_at | At2g01290 | expressed protein | -2.11 |
| 248205_at | At5g54300 | expressed protein | -2.11 |
| 259849_at | At1g72190 | oxidoreductase | -2.11 |
| 256786_at | At3g13740 | URF 4-related | -2.11 |
| 250563_at | At5g08050 | expressed protein | -2.11 |
| 258383_at | At3g15440 | expressed protein | -2.12 |
| 249837_at | At5g23480 | expressed protein | -2.12 |
| 251235_at | At3g62860 | esterase | -2.12 |
| 258495_at | At3g02690 | integral membrane | -2.12 |
| 262226_at | At1g53885 | senescence-associated protein-related | -2.13 |
| 245088_at | At2g39850 | subtilase | -2.13 |
| 262970_at | At1g75690 | chaperone protein dnaJ-related | -2.13 |
| 257003_at | At3g14110 | tetratricopeptide repeat (TPR)-containing protein | -2.13 |
| 266285_at | At2g29180 | expressed protein | -2.13 |
| 247162_at | At5g65730 | xyloglucan:xyloglucosyl transferase, putative | -2.14 |
| 256577_at | At3g28220 | mepirin and TRAF homology domain-containing protein | -2.14 |
| 251750_at | At3g55710 | UDP-glucuronosyl/UDP-glucosyl transferase | -2.14 |
| 258327_at | At3g22640 | cupin | -2.14 |
| 250856_at | At5g04810 | pentatricopeptide (PPR) repeat-containing protein | -2.14 |
| 265768_at | At2g48020 | sugar transporter, putative | -2.15 |
| 255381_at | At4g03510 | zinc finger (C3HC4-type RING finger) (RMA1) | -2.15 |
| 263275_at | At2g14170 | methylnalonate-semialdehyde dehydrogenase, putative | -2.15 |
| 250256_at | At5g13650 | elongation factor | -2.15 |
| 264525_at | At1g10060 | branched-chain amino acid transaminase 1 (BCAT1) | -2.15 |
| 253825_at | At4g28025 | expressed protein | -2.15 |
| 256870_at | At3g26300 | cytochrome P450 | -2.15 |
| 266963_at | At2g39450 | cation efflux | -2.16 |
| 262760_at | At1g10770 | invertase | -2.16 |
| 255774_at | At1g18620 | expressed protein | -2.16 |
| 251996_at | At3g52840 | beta-galactosidase, putative | -2.16 |
| 263184_at | At1g05560 | UDP-glucose transferase (UGT75B2) | -2.16 |
| 246959_at | At5g24700 | expressed protein | -2.16 |
| 265569_at | At2g05620 | expressed protein | -2.16 |
| 246199_at | At4g36530 | hydrolase, alpha/beta fold | -2.16 |
| 250255_at | At5g13730 | sigma-like factor (SIG4) | -2.16 |
| 257367_at | At2g25780 | hypothetical protein | -2.17 |
| 259103_at | At3g11690 | expressed protein | -2.17 |
| 264553_s_at | At1g09480 | cinnamyl-alcohol dehydrogenase family (CAD) | -2.17 |
| 250580_at | At5g07440 | glutamate dehydrogenase 2 (GDH2) | -2.18 |
| 245743_at | At1g51080 | expressed protein | -2.18 |
| 264383_at | At2g25080 | phospholipid hydroperoxide glutathione peroxidase, chloroplast | -2.18 |

| (GPX1) | | | |
|-------------|-----------|--|-------|
| 260522_x_at | At2g41730 | expressed protein | -2.18 |
| 252353_at | At3g48200 | expressed protein | -2.18 |
| 250781_at | At5g05410 | DRE-binding protein (DREB2A) | -2.18 |
| 245018_at | ycf4 | ycf4 | -2.18 |
| 264954_at | At1g77060 | mutase | -2.18 |
| 260379_at | At1g73880 | expressed protein | -2.18 |
| 267247_at | At2g30170 | expressed protein | -2.18 |
| 265182_at | At1g23740 | oxidoreductase, zinc-binding dehydrogenase | -2.19 |
| 252950_at | At4g38690 | 1-phosphatidylinositol phosphodiesterase-related | -2.19 |
| 249073_at | At5g44020 | acid phosphatase class B | -2.19 |
| 244932_at | psaC | psaC | -2.19 |
| 262897_at | At1g59840 | expressed protein | -2.19 |
| 248962_at | At5g45680 | FK506-binding protein 1 (FKBP13) | -2.19 |
| 264397_at | At1g11820 | glycosyl hydrolase family 17 protein | -2.19 |
| 259603_at | At1g56500 | haloacid dehalogenase-like hydrolase | -2.19 |
| 259896_at | At1g71500 | Rieske (2Fe-2S) domain-containing protein | -2.19 |
| 264442_at | At1g27480 | lecithin:cholesterol acyltransferase (LACT) | -2.2 |
| 251059_at | At5g01810 | CBL-interacting protein kinase 15 (CIPK15) | -2.2 |
| 261388_at | At1g05385 | photosystem II 11 kDa protein-related | -2.2 |
| 251784_at | At3g55330 | photosystem II reaction center PsbP | -2.2 |
| 262304_at | At1g70890 | major latex protein-related | -2.2 |
| 262979_s_at | At1g75610 | histone H3, putative | -2.21 |
| 260106_at | At1g35420 | dienelactone hydrolase | -2.21 |
| 261861_at | At1g50450 | expressed protein | -2.21 |
| 246211_at | At4g36730 | G-box binding factor 1 (GBF1) | -2.22 |
| 253790_at | At4g28660 | photosystem II reaction centre W (PsbW) | -2.22 |
| 248449_at | At5g51110 | expressed protein | -2.22 |
| 256304_at | At1g69523 | UbiE/COQ5 methyltransferase | -2.23 |
| 253951_at | At4g26860 | alanine racemase | -2.23 |
| 261931_at | At1g22430 | alcohol dehydrogenase, putative (ADH) | -2.23 |
| 245793_at | At1g32220 | expressed protein | -2.23 |
| 251143_at | At5g01220 | sulfolipid synthase (SQD2) | -2.23 |
| 260014_at | At1g68010 | glycerate dehydrogenase | -2.23 |
| 247931_at | At5g57040 | lactoylglutathione lyase | -2.23 |
| 265073_at | At1g55480 | expressed protein | -2.24 |
| 262645_at | At1g62750 | elongation factor Tu | -2.24 |
| 248193_at | At5g54080 | homogentisic acid oxidase (HGO) | -2.25 |
| 267497_at | At2g30540 | glutaredoxin | -2.25 |
| 261793_at | At1g16080 | expressed protein | -2.25 |
| 251647_at | At3g57770 | protein kinase, putative | -2.25 |
| 249472_at | At5g39210 | expressed protein | -2.25 |
| 251155_at | At3g63160 | expressed protein | -2.26 |
| 247924_at | At5g57655 | xylose isomerase | -2.26 |
| 249927_at | At5g19220 | ADP-glucose pyrophosphorylase (ADG2) | -2.26 |

| | | | |
|-------------|-----------|--|-------|
| 254038_at | At4g25910 | nitrogen fixation protein, putative | -2.27 |
| 260969_at | At1g12240 | beta-fructosidase (BFRUCT4) | -2.27 |
| 250752_at | At5g05690 | cytochrome P450 90A1 (CYP90A1) | -2.27 |
| 263443_at | At2g28630 | beta-ketoacyl-CoA synthase | -2.28 |
| 261536_at | At1g01790 | K ⁺ efflux antiporter, putative (KEA1) | -2.28 |
| 251230_at | At3g62750 | glycosyl hydrolase family 1 protein | -2.28 |
| 261822_at | At1g11380 | expressed protein | -2.29 |
| 250151_at | At5g14570 | transporter, putative | -2.29 |
| 259199_at | At3g08980 | signal peptidase I | -2.29 |
| 244972_at | psbB | psbB | -2.29 |
| 248419_at | At5g51550 | phosphate-responsive 1 | -2.3 |
| 253394_at | At4g32770 | sucrose export defective 1 (SXD1) | -2.3 |
| 265511_at | At2g05540 | glycine-rich protein | -2.3 |
| 263676_at | At1g09340 | expressed protein | -2.3 |
| 248224_at | At5g53490 | thylakoid lumenal 17.4 kDa protein | -2.3 |
| 246951_at | At5g04880 | expressed protein | -2.31 |
| 251762_at | At3g55800 | sedoheptulose-1,7-bisphosphatase | -2.31 |
| 260968_at | At1g12250 | thylakoid lumenal protein-related | -2.31 |
| 255543_at | At4g01870 | tolB protein-related | -2.32 |
| 247845_at | At5g58090 | glycosyl hydrolase 17 protein | -2.33 |
| 259633_at | At1g56505 | haloacid dehalogenase-like hydrolase | -2.33 |
| 245744_at | At1g51110 | plastid-lipid associated protein PAP | -2.33 |
| 244995_at | atpl | atpl | -2.33 |
| 264096_at | At1g78995 | expressed protein | -2.33 |
| 248657_at | At5g48570 | peptidyl-prolyl cis-trans isomerase, putative | -2.34 |
| 263150_at | At1g54050 | 17.4 kDa class III heat shock protein (HSP17.4-CIII) | -2.34 |
| 245291_at | At4g16155 | lipamide dehydrogenase 2 (PTLPD2) | -2.34 |
| 262603_at | At1g15380 | lactoylglutathione lyase | -2.35 |
| 266589_at | At2g46250 | myosin heavy chain-related | -2.35 |
| 252853_at | At4g39710 | FKBP-type peptidyl-prolyl cis-trans isomerase, putative | -2.35 |
| 249226_at | At5g42170 | family II extracellular lipase, putative | -2.36 |
| 257315_at | At3g30775 | proline oxidase, mitochondrial (POX) | -2.36 |
| 263881_at | At2g21820 | expressed protein | -2.37 |
| 262162_at | At1g78020 | senescence-associated protein-related | -2.37 |
| 250531_at | At5g08650 | GTP-binding protein LepA, putative | -2.37 |
| 261480_at | At1g14280 | phytochrome kinase, putative | -2.37 |
| 252441_at | At3g46780 | expressed protein | -2.37 |
| 266426_x_at | At2g07140 | F-box | -2.37 |
| 263880_at | At2g21960 | expressed protein | -2.37 |
| 265703_at | At2g03430 | ankyrin repeat | -2.38 |
| 258055_at | At3g16250 | ferredoxin-related | -2.38 |
| 253387_at | At4g33010 | glycine dehydrogenase (decarboxylating), putative | -2.38 |
| 263668_at | At1g04350 | 2-oxoglutarate-dependent dioxygenase, putative | -2.39 |
| 260704_at | At1g32470 | glycine cleavage system H protein, mitochondrial, putative | -2.39 |
| 266767_at | At2g46910 | plastid-lipid associated protein PAP | -2.39 |

| | | | |
|-------------|-----------|--|-------|
| 264209_at | At1g22740 | Ras-related protein (RAB7) | -2.39 |
| 256772_at | At3g13750 | beta-galactosidase, putative | -2.39 |
| 252991_at | At4g38470 | protein kinase | -2.4 |
| 262151_at | At1g52510 | hydrolase, alpha/beta fold | -2.4 |
| 260696_at | At1g32520 | expressed protein | -2.4 |
| 260978_at | At1g53540 | 17.6 kDa class I small heat shock protein (HSP17.6C-CI) (AA 1-156) | -2.41 |
| 266278_at | At2g29300 | tropinone reductase, putative | -2.42 |
| 253281_at | At4g34138 | UDP-glucuronosyl/UDP-glucosyl transferase | -2.43 |
| 265387_at | At2g20670 | expressed protein | -2.43 |
| 263410_at | At2g04039 | expressed protein | -2.43 |
| 260837_at | At1g43670 | fructose-1,6-bisphosphatase, putative | -2.43 |
| 258972_at | At3g01920 | yrdC | -2.43 |
| 256036_at | At1g07110 | fructose-2,6-bisphosphatase (F2KP) | -2.43 |
| 264289_at | At1g61890 | MATE efflux | -2.44 |
| 254496_at | At4g20070 | peptidase M20/M25/M40 | -2.44 |
| 250075_at | At5g17670 | expressed protein | -2.44 |
| 250167_at | At5g15310 | myb family transcription factor | -2.44 |
| 248624_at | At5g48790 | expressed protein | -2.44 |
| 250910_at | At5g03720 | heat shock transcription factor | -2.45 |
| 251919_at | At3g53800 | armadillo/beta-catenin repeat | -2.45 |
| 246962_s_at | At5g24800 | bZIP transcription factor | -2.45 |
| 252539_at | At3g45730 | expressed protein | -2.46 |
| 262605_at | At1g15170 | MATE efflux | -2.46 |
| 259694_at | At1g63180 | UDP-glucose 4-epimerase, putative | -2.46 |
| 245007_at | psaA | psaA | -2.46 |
| 246651_at | At5g35170 | adenylate kinase | -2.46 |
| 245003_at | psbC | psbC | -2.47 |
| 263142_at | At1g65230 | expressed protein | -2.47 |
| 265116_at | At1g62480 | vacuolar calcium-binding protein-related | -2.48 |
| 261948_at | At1g64680 | expressed protein | -2.49 |
| 249230_at | At5g42070 | expressed protein | -2.49 |
| 256548_at | At3g14770 | nodulin MtN3 | -2.5 |
| 267036_at | At2g38465 | expressed protein | -2.5 |
| 245016_at | accD | accD | -2.5 |
| 252081_at | At3g51910 | heat shock transcription factor | -2.51 |
| 267057_at | At2g32500 | expressed protein | -2.52 |
| 246411_at | At1g57770 | amine oxidase family | -2.52 |
| 261084_at | At1g07440 | tropinone reductase, putative | -2.52 |
| 260284_at | At1g80380 | phosphoribulokinase | -2.53 |
| 264728_at | At1g22850 | expressed protein | -2.53 |
| 267262_at | At2g22990 | sinapoylglucose:malate sinapoyltransferase (SNG1) | -2.54 |
| 267523_at | At2g30610 | BTB/POZ domain-containing protein | -2.55 |
| 256076_at | At1g18060 | expressed protein | -2.55 |
| 248491_at | At5g51010 | rubredoxin | -2.55 |

| | | | |
|-------------|-----------|---|-------|
| 255793_at | At2g33250 | expressed protein | -2.56 |
| 253217_at | At4g34970 | actin-depolymerizing factor, putative | -2.56 |
| 255962_at | At1g22335 | expressed protein | -2.57 |
| 246630_at | At1g50730 | expressed protein | -2.58 |
| 248566_s_at | At5g49730 | ferric reductase-like transmembrane component | -2.6 |
| 246034_at | At5g08350 | GRAM domain-containing protein | -2.6 |
| 246998_at | At5g67370 | expressed protein | -2.6 |
| 246792_at | At5g27290 | expressed protein | -2.6 |
| 249120_at | At5g43750 | expressed protein | -2.6 |
| 256999_at | At3g14200 | DNAJ heat shock N-terminal domain-containing protein | -2.61 |
| 259982_at | At1g76410 | zinc finger (C3HC4-type RING finger) | -2.62 |
| 251036_at | At5g02160 | expressed protein | -2.62 |
| 262569_at | At1g15180 | MATE efflux | -2.62 |
| 258250_at | At3g15850 | fatty acid desaturase | -2.62 |
| 250972_at | At5g02840 | myb family transcription factor | -2.63 |
| 251157_at | At3g63140 | mRNA-binding protein, putative | -2.63 |
| 253496_at | At4g31870 | glutathione peroxidase, putative | -2.64 |
| 253279_at | At4g34030 | 3-methylcrotonyl-CoA carboxylase 2 (MCCB) | -2.64 |
| 247919_at | At5g57650 | eukaryotic translation initiation factor-related | -2.65 |
| 262698_at | At1g75960 | AMP-binding protein, putative | -2.66 |
| 251759_at | At3g55630 | dihydrofolate synthetase/folylpolyglutamate synthetase (DHFS/FPGS4) | -2.66 |
| 261788_at | At1g15980 | expressed protein | -2.66 |
| 251642_at | At3g57520 | alkaline alpha galactosidase, putative | -2.67 |
| 249524_at | At5g38520 | hydrolase, alpha/beta fold | -2.67 |
| 263827_at | At2g40420 | amino acid transporter | -2.68 |
| 251248_at | At3g62150 | multidrug resistant (MDR) ABC transporter, putative | -2.68 |
| 254561_at | At4g19160 | expressed protein | -2.68 |
| 245002_at | psbD | psbD | -2.69 |
| 259914_at | At1g72640 | expressed protein | -2.71 |
| 253049_at | At4g37300 | expressed protein | -2.71 |
| 259707_at | At1g77490 | L-ascorbate peroxidase, thylakoid-bound (IAPX) | -2.71 |
| 249875_at | At5g23120 | photosystem II stability/assembly factor, chloroplast (HCF136) | -2.71 |
| 248207_at | At5g53970 | aminotransferase, putative | -2.72 |
| 252924_at | At4g39070 | zinc finger (B-box type) | -2.72 |
| 256015_at | At1g19150 | chlorophyll A-B binding protein, putative | -2.73 |
| 259329_at | At3g16360 | phosphotransfer | -2.73 |
| 251068_at | At5g01920 | protein kinase | -2.73 |
| 245745_at | At1g51115 | plastid-lipid associated protein PAP | -2.74 |
| 263252_at | At2g31380 | salt tolerance-like protein (STH) | -2.76 |
| 258338_at | At3g16150 | L-asparaginase, putative | -2.76 |
| 244933_at | ndhE | ndhE | -2.76 |
| 246510_at | At5g15410 | cyclic nucleotide-gated channel (CNGC2) | -2.77 |
| 266329_at | At2g01590 | expressed protein | -2.77 |

| | | | |
|-------------|-----------|--|-------|
| 267035_at | At2g38400 | alanine-glyoxylate aminotransferase, putative (AGT) | -2.79 |
| 260900_s_at | At1g21400 | 2-oxoisovalerate dehydrogenase, putative | -2.79 |
| 255012_at | At4g10030 | hydrolase, alpha/beta fold | -2.8 |
| 261942_at | At1g22590 | MADS-box | -2.81 |
| 254835_s_at | At4g12310 | cytochrome P450, putative | -2.82 |
| 254020_at | At4g25700 | beta-carotene hydroxylase | -2.82 |
| 255016_at | At4g10120 | sucrose-phosphate synthase, putative | -2.88 |
| 261814_at | At1g08310 | esterase | -2.88 |
| 254839_at | At4g12400 | stress-inducible protein, putative | -2.89 |
| 266979_at | At2g39470 | photosystem II reaction center PsbP | -2.89 |
| 266294_at | At2g29500 | 17.6 kDa class I small heat shock protein (HSP17.6B-CI) | -2.91 |
| 261422_at | At1g18730 | expressed protein | -2.91 |
| 262635_at | At1g06570 | 4-hydroxyphenylpyruvate dioxygenase (HPD) | -2.93 |
| 262911_s_at | At1g59860 | 17.8 kDa class I heat shock protein (HSP17.8-CI) | -2.93 |
| 254787_at | At4g12690 | expressed protein | -2.95 |
| 256894_at | At3g21870 | cyclin | -2.99 |
| 245876_at | At1g26230 | chaperonin, putative | -2.99 |
| 260662_at | At1g19540 | isoflavone reductase, putative | -3 |
| 248347_at | At5g52250 | transducin | -3.01 |
| 249765_at | At5g24030 | C4-dicarboxylate transporter | -3.01 |
| 247882_at | At5g57785 | expressed protein | -3.02 |
| 266590_at | At2g46240 | IQ domain-containing protein | -3.02 |
| 247816_at | At5g58260 | expressed protein | -3.04 |
| 263133_at | At1g78450 | SOUL heme-binding | -3.05 |
| 266279_at | At2g29290 | tropinone reductase, putative | -3.06 |
| 260743_at | At1g15040 | At1g15040 | -3.1 |
| 262175_at | At1g74880 | expressed protein | -3.1 |
| 258527_at | At3g06850 | branched chain alpha-keto acid dehydrogenase E2 subunit (din3) | -3.11 |
| 248793_at | At5g47240 | MutT/nudix | -3.11 |
| 266984_at | At2g39570 | ACT domain-containing protein | -3.11 |
| 252570_at | At3g45300 | isovaleryl-CoA-dehydrogenase (IVD) | -3.12 |
| 259789_at | At1g29395 | stress-responsive protein, putative | -3.15 |
| 265070_at | At1g55510 | 2-oxoisovalerate dehydrogenase, putative | -3.16 |
| 255719_at | At1g32080 | membrane protein, putative | -3.18 |
| 253161_at | At4g35770 | senescence-associated protein (SEN1) | -3.24 |
| 258956_at | At3g01440 | oxygen evolving enhancer 3 (PsbQ) | -3.24 |
| 254016_at | At4g26150 | zinc finger (GATA type) | -3.27 |
| 261663_at | At1g18330 | myb family transcription factor | -3.28 |
| 264213_at | At1g65400 | disease resistance protein (TIR class), putative | -3.29 |
| 263981_at | At2g42870 | expressed protein | -3.29 |
| 246195_at | At4g36410 | ubiquitin-conjugating enzyme 17 (UBC17) | -3.29 |
| 251353_at | At3g61080 | fructosamine kinase | -3.29 |
| 254384_at | At4g21870 | 26.5 kDa class P-related heat shock protein (HSP26.5-P) | -3.33 |
| 267524_at | At2g30600 | BTB/POZ domain-containing protein | -3.37 |

| | | | |
|-------------|-----------|---|-------|
| 245757_at | At1g35140 | phosphate-responsive protein, putative | -3.38 |
| 248434_at | At5g51440 | 23.5 kDa mitochondrial small heat shock protein (HSP23.5-M) | -3.39 |
| 266956_at | At2g34510 | expressed protein | -3.42 |
| 253197_at | At4g35250 | vestitone reductase-related | -3.45 |
| 262047_at | At1g80160 | lactoylglutathione lyase | -3.51 |
| 264561_at | At1g55810 | uracil phosphoribosyltransferase, putative | -3.52 |
| 253943_at | At4g27030 | expressed protein | -3.6 |
| 256965_at | At3g13450 | branched-chain alpha-keto acid dehydrogenase E1 beta subunit (DIN4) | -3.62 |
| 261395_at | At1g79700 | ovule development protein, putative | -3.63 |
| 259076_at | At3g02140 | expressed protein | -3.64 |
| 248050_at | At5g56100 | glycine-rich protein | -3.67 |
| 251356_at | At3g61060 | F-box | -3.68 |
| 252366_at | At3g48420 | haloacid dehalogenase-like hydrolase | -3.68 |
| 262288_at | At1g70760 | inorganic carbon transport protein-related | -3.69 |
| 247814_at | At5g58310 | hydrolase, alpha/beta fold | -3.7 |
| 267138_s_at | At2g38210 | ethylene-responsive protein, putative | -3.71 |
| 264923_s_at | At1g60740 | peroxiredoxin type 2, putative | -3.73 |
| 266693_at | At2g19800 | expressed protein | -3.79 |
| 260668_at | At1g19530 | expressed protein | -3.83 |
| 263287_at | At2g36145 | expressed protein | -3.83 |
| 252415_at | At3g47340 | glutamine-dependent asparagine synthetase 1 (ASN1) | -3.85 |
| 258856_at | At3g02040 | glycerophosphoryl diester phosphodiesterase | -3.91 |
| 256245_at | At3g12580 | heat shock protein 70, putative (HSP70) | -3.96 |
| 262612_at | At1g14150 | oxygen evolving enhancer 3 (PsbQ) | -3.98 |
| 260248_at | At1g74310 | heat shock protein 101 (HSP101) | -4 |
| 248377_at | At5g51720 | expressed protein | -4.12 |
| 261957_at | At1g64660 | Cys/Met metabolism pyridoxal-phosphate-dependent enzyme | -4.14 |
| 251196_at | At3g62950 | glutaredoxin | -4.15 |
| 264524_at | At1g10070 | branched-chain amino acid transaminase 2 (BCAT2) | -4.17 |
| 255298_at | At4g04840 | methionine sulfoxide reductase domain-containing protein | -4.22 |
| 251428_at | At3g60140 | glycosyl hydrolase family 1 protein | -4.26 |
| 266841_at | At2g26150 | heat shock transcription factor | -4.27 |
| 245362_at | At4g17460 | homeobox-leucine zipper protein 1 (HAT1) | -4.31 |
| 260536_at | At2g43400 | electron transfer flavoprotein-ubiquinone oxidoreductase | -4.32 |
| 257670_at | At3g20340 | expressed protein | -4.34 |
| 263118_at | At1g03090 | 3-methylcrotonyl-CoA carboxylase 1 (MCCA) | -4.35 |
| 250351_at | At5g12030 | 17.7 kDa class II heat shock protein 17.6A (HSP17.7-CII) | -4.51 |
| 264238_at | At1g54740 | expressed protein | -4.62 |
| 267265_at | At2g22980 | serine carboxypeptidase S10 | -4.64 |
| 258181_at | At3g21670 | nitrate transporter (NTP3) | -4.65 |
| 247478_at | At5g62360 | invertase | -4.67 |
| 253966_at | At4g26520 | fructose-bisphosphate aldolase | -4.69 |
| 260915_at | At1g02660 | lipase class 3 | -4.82 |

| | | | |
|------------------------------------|-----------|--|-------|
| 249337_at | At5g41080 | glycerophosphoryl diester phosphodiesterase | -4.86 |
| 254783_at | At4g12830 | hydrolase, alpha/beta fold | -4.92 |
| 260741_at | At1g15045 | glutamine amidotransferase-related | -5.03 |
| 250199_at | At5g14180 | lipase | -5.21 |
| 246251_at | At4g37220 | stress-responsive protein, putative | -5.23 |
| 254001_at | At4g26260 | expressed protein | -5.26 |
| 265111_at | At1g62510 | protease inhibitor/seed storage/lipid transfer protein (LTP) | -5.52 |
| 262307_at | At1g71000 | DNAJ heat shock N-terminal domain-containing protein | -5.96 |
| 264777_at | At1g08630 | L-allo-threonine aldolase-related | -6.42 |
| 264514_at | At1g09500 | cinnamyl-alcohol dehydrogenase family (CAD) | -7.02 |
| 253971_at | At4g26530 | fructose-bisphosphate aldolase, putative | -7.93 |
| 253829_at | At4g28040 | nodulin MtN21 | -8.18 |
| 256527_at | At1g66100 | thionin, putative | -8.35 |
| <i>4 days after Ca suppression</i> | | | |
| 264400_at | At1g61800 | glucose-6-phosphate/phosphate translocator, putative | 4.36 |
| 267147_at | At2g38240 | oxidoreductase, 2OG-Fe(II) oxygenase | 3.36 |
| 252265_at | At3g49620 | 2-oxoacid-dependent oxidase, putative (DIN11) | 2.75 |
| 266720_s_at | At2g46790 | pseudo-response regulator, putative | 2.53 |
| 252661_at | At3g44450 | expressed protein | 2.49 |
| 249364_at | At5g40590 | DC1 domain-containing protein | 2.46 |
| 252827_at | At4g39950 | cytochrome P450 79B2, putative (CYP79B2) | 2.44 |
| 258586_s_at | At3g04320 | trypsin and protease inhibitor | 2.36 |
| 260130_s_at | At1g66280 | beta-glucosidase (PSR3.2) | 2.36 |
| 260557_at | At2g43610 | glycoside hydrolase family 19 protein | 2.3 |
| 253608_at | At4g30290 | xyloglucan:xyloglucosyl transferase, putative | 2.29 |
| 254361_at | At4g22212 | expressed protein | 2.2 |
| 256999_at | At3g14200 | DNAJ heat shock N-terminal domain-containing protein | 2.17 |
| 265665_at | At2g27420 | cysteine proteinase, putative | 2.08 |
| 246490_at | At5g15950 | adenosylmethionine decarboxylase | 2.06 |
| 247755_at | At5g59090 | subtilase | 2.02 |
| 258133_at | At3g24500 | ethylene-responsive transcriptional coactivator, putative | 1.91 |
| 265221_s_at | At2g02010 | glutamate decarboxylase, putative | 1.9 |
| 260248_at | At1g74310 | heat shock protein 101 (HSP101) | 1.89 |
| 254550_at | At4g19690 | iron-responsive transporter (IRT1) | 1.89 |
| 253382_at | At4g33040 | glutaredoxin | 1.89 |
| 266841_at | At2g26150 | heat shock transcription factor | 1.87 |
| 245627_at | At1g56600 | galactinol synthase, putative | 1.87 |
| 252563_at | At3g45970 | expansin (EXPL1) | 1.84 |
| 262656_at | At1g14200 | zinc finger (C3HC4-type RING finger) | 1.84 |
| 256518_at | At1g66080 | expressed protein | 1.79 |
| 256619_at | At3g24470 | TMS membrane (TDE) | 1.79 |
| 265892_at | At2g15020 | expressed protein | 1.78 |
| 258830_at | At3g07090 | expressed protein | 1.78 |
| 258419_at | At3g16670 | expressed protein | 1.77 |
| 247573_at | At5g61160 | transferase | 1.77 |

| | | | |
|-------------|-----------|---|------|
| 252679_at | At3g44260 | CCR4-NOT transcription complex protein, putative | 1.74 |
| 266271_at | At2g29440 | glutathione S-transferase, putative | 1.73 |
| 252984_at | At4g37990 | mannitol dehydrogenase, putative (ELI3-2) | 1.72 |
| 260134_at | At1g66370 | myb family transcription factor (MYB113) | 1.72 |
| 248732_at | At5g48070 | xyloglucan:xyloglucosyl transferase, putative | 1.7 |
| 248138_at | At5g54960 | pyruvate decarboxylase, putative | 1.69 |
| 264787_at | At2g17840 | senescence/dehydration-associated protein-related (ERD7) | 1.68 |
| 258259_s_at | At3g26840 | esterase | 1.67 |
| 255773_at | At1g18590 | sulfotransferase | 1.67 |
| 252114_at | At3g51450 | strictosidine synthase | 1.67 |
| 258418_at | At3g16660 | expressed protein | 1.65 |
| 267411_at | At2g34930 | disease resistance | 1.65 |
| 245734_at | At1g73480 | hydrolase, alpha/beta fold | 1.63 |
| 253162_at | At4g35630 | phosphoserine aminotransferase, chloroplast (PSAT) | 1.62 |
| 252958_at | At4g38620 | myb family transcription factor (MYB4) | 1.61 |
| 256178_s_at | At1g51780 | IAA-Ala hydrolase 3 (IAR3) | 1.61 |
| 253398_at | At4g32810 | dioxygenase-related | 1.6 |
| 247877_at | At5g57740 | zinc finger (C3HC4-type RING finger) / ankyrin repeat | 1.6 |
| 247452_at | At5g62430 | Dof-type zinc finger domain-containing protein | 1.59 |
| 265897_at | At2g25680 | expressed protein | 1.58 |
| 251154_at | At3g63110 | cytokinin synthase (IPT3) | 1.58 |
| 266326_at | At2g46650 | cytochrome b5, putative | 1.58 |
| 259518_at | At1g20510 | 4-coumaroyl-CoA synthase | 1.57 |
| 252952_at | At4g38710 | glycine-rich protein | 1.56 |
| 253405_at | At4g32800 | AP2 domain-containing transcription factor TINY, putative | 1.53 |
| 248271_at | At5g53420 | expressed protein | 1.52 |
| 252255_at | At3g49220 | pectinesterase | 1.52 |
| 255700_at | At4g00200 | DNA-binding | 1.52 |
| 260712_at | At1g17550 | protein phosphatase 2C-related | 1.51 |
| 248190_at | At5g54120 | calcium-binding EF hand | 1.5 |
| 266609_at | At2g35510 | WWE domain-containing protein | 1.5 |
| 266553_at | At2g46170 | reticulon (RTNLB5) | 1.5 |
| 251234_s_at | At2g47650 | NAD-dependent epimerase/dehydratase | 1.49 |
| 254563_at | At4g19120 | early-responsive to dehydration stress protein (ERD3) | 1.49 |
| 260830_at | At1g06760 | histone H1, putative | 1.49 |
| 256529_at | At1g33260 | protein kinase | 1.49 |
| 251704_at | At3g56360 | expressed protein | 1.47 |
| 250500_at | At5g09530 | hydroxyproline-rich glycoprotein | 1.47 |
| 248469_at | At5g50820 | no apical meristem (NAM) | 1.47 |
| 265359_at | At2g16720 | myb family transcription factor | 1.47 |
| 251791_at | At3g55500 | expansin, putative (EXP16) | 1.47 |
| 257335_at | orf105b | orf105b | 1.46 |
| 267112_at | At2g14750 | adenylsulfate kinase 1 (AKN1) | 1.46 |
| 256645_at | At3g24250 | glycine-rich protein | 1.45 |
| 261749_at | At1g76180 | dehydrin (ERD14) | 1.45 |

| | | | |
|-----------|-----------|---|-------|
| 248272_at | At5g53485 | importin beta-2, putative | 1.45 |
| 248270_at | At5g53450 | protein kinase | 1.44 |
| 259894_at | At1g71430 | expressed protein | 1.43 |
| 260342_at | At1g69280 | expressed protein | 1.41 |
| 250096_at | At5g17190 | expressed protein | 1.41 |
| 256022_at | At1g58360 | amino acid permease I (AAP1) | 1.39 |
| 259366_at | At1g13280 | allene oxide cyclase | 1.38 |
| 260494_at | At2g41820 | leucine-rich repeat transmembrane protein kinase, putative | 1.37 |
| 267561_at | At2g45590 | protein kinase | -1.3 |
| 261049_at | At1g01430 | expressed protein | -1.32 |
| 267135_at | At2g23430 | kip-related protein 1 (KRP1) | -1.32 |
| 245269_at | At4g14500 | expressed protein | -1.32 |
| 245313_at | At4g15420 | PRLI-interacting factor K | -1.33 |
| 256659_at | At3g12020 | kinesin motor protein-related | -1.33 |
| 265675_at | At2g32120 | heat shock protein 70 (HSP70) | -1.34 |
| 261802_at | At1g30550 | expressed protein | -1.35 |
| 250107_at | At5g15330 | SPX domain-containing protein | -1.35 |
| 244977_at | petD | petD | -1.35 |
| 260554_at | At2g41790 | peptidase M16 | -1.35 |
| 264060_at | At2g27980 | expressed protein | -1.36 |
| 257618_at | At3g24720 | protein kinase | -1.36 |
| 260364_at | At1g70560 | alliinase C-terminal domain-containing protein | -1.38 |
| 247980_at | At5g56860 | zinc finger (GATA type) | -1.38 |
| 247713_at | At5g59330 | At5g59330 | -1.38 |
| 258544_at | At3g07040 | disease resistance protein RPM1 (CC-NBS-LRR class), putative | -1.39 |
| 255382_at | At4g03430 | pre-mRNA splicing factor-related | -1.4 |
| 253703_at | At4g29910 | origin recognition complex subunit 5-related (ORC5) | -1.4 |
| 257205_at | At3g16520 | UDP-glucuronosyl/UDP-glucosyl transferase | -1.4 |
| 266348_at | At2g01450 | mitogen-activated protein kinase, putative (MPK17) | -1.41 |
| 258931_at | At3g10010 | HhH-GPD base excision DNA repair | -1.41 |
| 246327_at | At1g16670 | protein kinase | -1.41 |
| 264649_at | At1g09060 | transcription factor jumonji (jmjC) domain-containing protein | -1.41 |
| 252972_at | At4g38840 | auxin-responsive protein, putative | -1.41 |
| 256457_at | At1g75230 | HhH-GPD base excision DNA repair | -1.42 |
| 266606_at | At2g46310 | AP2 domain-containing transcription factor, putative | -1.43 |
| 250063_at | At5g17880 | disease resistance protein (TIR-NBS-LRR class), putative | -1.43 |
| 265868_at | At2g01650 | zinc finger (C2H2 type) | -1.43 |
| 261791_at | At1g16170 | expressed protein | -1.43 |
| 263133_at | At1g78450 | SOUL heme-binding | -1.43 |
| 262394_at | At1g49510 | expressed protein | -1.43 |
| 246975_at | At5g24890 | expressed protein | -1.43 |
| 254032_at | At4g25940 | epsin N-terminal homology (ENTH) domain-containing protein | -1.43 |
| 257935_at | At3g25440 | group II intron splicing factor CRS1-related | -1.44 |

| | | | |
|-------------|-----------|---|-------|
| 250257_at | At5g13770 | pentatricopeptide (PPR) repeat-containing protein | -1.44 |
| 250549_at | At5g07860 | transferase | -1.44 |
| 264867_at | At1g24150 | formin homology 2 domain-containing protein / FH2 domain-containing protein | -1.44 |
| 248106_at | At5g55100 | SWAP (Suppressor-of-White-APricot) | -1.45 |
| 244990_s_at | orf77.1 | orf77.1 | -1.45 |
| 258647_at | At3g07870 | F-box | -1.45 |
| 249074_at | At5g44080 | bZIP transcription factor | -1.45 |
| 259560_at | At1g21270 | wall-associated kinase 2 (WAK2) | -1.46 |
| 252504_at | At3g46590 | telomere repeat-binding protein, putative | -1.46 |
| 248873_at | At5g46450 | disease resistance protein (TIR-NBS-LRR class), putative | -1.46 |
| 258351_at | At3g17700 | cyclic nucleotide-binding transporter 1 (CNGC20) | -1.46 |
| 253369_at | At4g33210 | F-box (FBL15) | -1.46 |
| 266502_at | At2g47720 | expressed protein | -1.46 |
| 249341_at | At5g41170 | pentatricopeptide (PPR) repeat-containing protein | -1.46 |
| 245506_at | At4g15700 | glutaredoxin | -1.46 |
| 263305_at | At2g01930 | expressed protein | -1.46 |
| 252126_at | At3g50950 | disease resistance protein (CC-NBS-LRR class), putative | -1.46 |
| 265023_at | At1g24440 | expressed protein | -1.47 |
| 262129_at | At1g52500 | formamidopyrimidine-DNA glycolase (MMH-1) | -1.47 |
| 256193_at | At1g30200 | F-box | -1.47 |
| 258297_at | At3g23325 | splicing factor, putative | -1.47 |
| 266361_at | At2g32450 | calcium-binding EF hand | -1.47 |
| 257751_at | At3g18690 | VQ motif-containing protein | -1.47 |
| 245821_at | At1g26270 | phosphatidylinositol 3- and 4-kinase | -1.47 |
| 259582_at | At1g28060 | small nuclear ribonucleoprotein (snRNP) | -1.47 |
| 267357_at | At2g40000 | expressed protein | -1.48 |
| 259822_at | At1g66230 | myb family transcription factor (MYB20) | -1.48 |
| 260770_at | At1g49200 | zinc finger (C3HC4-type RING finger) | -1.48 |
| 250398_at | At5g11000 | expressed protein | -1.49 |
| 262922_at | At1g79420 | expressed protein | -1.5 |
| 256790_at | At3g16857 | two-component responsive regulator | -1.5 |
| 264700_at | At1g70100 | expressed protein | -1.5 |
| 250177_at | At5g14420 | copine-related | -1.5 |
| 250004_at | At5g18750 | DNAJ heat shock N-terminal domain-containing protein | -1.51 |
| 256542_at | At1g42550 | expressed protein | -1.51 |
| 246283_at | At4g36860 | LIM domain-containing protein | -1.51 |
| 258350_at | At3g17510 | CBL-interacting protein kinase 1 (CIPK1) | -1.51 |
| 251321_at | At3g61460 | zinc finger (C3HC4-type RING finger) (BRH1) | -1.51 |
| 260431_at | At1g68190 | zinc finger (B-box type) | -1.52 |
| 254016_at | At4g26150 | zinc finger (GATA type) | -1.52 |
| 266230_at | At2g28830 | armadillo/beta-catenin repeat | -1.53 |
| 257858_at | At3g12920 | expressed protein | -1.54 |
| 256262_at | At3g12150 | expressed protein | -1.54 |
| 264200_at | At1g22650 | beta-fructofuranosidase, putative | -1.55 |

| | | | |
|-------------|-----------|--|-------|
| 255563_at | At4g01740 | DC1 domain-containing protein | -1.55 |
| 264040_at | At2g03730 | ACT domain-containing protein (ACR5) | -1.55 |
| 249319_s_at | At5g40880 | WD-40 repeat (ZFW4) | -1.55 |
| 254318_at | At4g22530 | embryo-abundant protein-related | -1.56 |
| 250821_at | At5g05190 | expressed protein | -1.56 |
| 265908_at | At4g00270 | DNA-binding storekeeper protein-related | -1.57 |
| 264339_at | At1g70290 | trehalose-6-phosphate synthase, putative | -1.57 |
| 251641_at | At3g57470 | peptidase M16 | -1.57 |
| 247413_at | At5g63020 | disease resistance protein (CC-NBS-LRR class), putative | -1.57 |
| 262552_at | At1g31350 | F-box | -1.57 |
| 261052_at | At1g01440 | extra-large G-protein-related | -1.57 |
| 249694_at | At5g35790 | glucose-6-phosphate 1-dehydrogenase (APG1) | -1.57 |
| 266882_at | At2g44670 | senescence-associated protein-related | -1.58 |
| 256914_at | At3g23880 | F-box | -1.58 |
| 252250_at | At3g49790 | expressed protein | -1.58 |
| 249485_at | At5g39020 | protein kinase | -1.58 |
| 267614_at | At2g26710 | cytochrome P450, putative | -1.59 |
| 263106_at | At2g05160 | zinc finger (CCCH-type) | -1.59 |
| 262801_at | At1g21010 | expressed protein | -1.59 |
| 248888_at | At5g46240 | inward rectifying potassium channel (KAT1) | -1.6 |
| 251621_at | At3g57700 | protein kinase, putative | -1.6 |
| 249384_at | At5g39890 | expressed protein | -1.6 |
| 257053_at | At3g15210 | ethylene-responsive element-binding factor 4 (ERF4) | -1.6 |
| 260734_at | At1g17600 | disease resistance protein (TIR-NBS-LRR class), putative | -1.6 |
| 260303_at | At1g70520 | protein kinase | -1.6 |
| 258668_at | At3g08780 | expressed protein | -1.61 |
| 256487_at | At1g31540 | disease resistance protein (TIR-NBS-LRR class), putative | -1.62 |
| 267289_at | At2g23770 | protein kinase | -1.62 |
| 254553_at | At4g19530 | disease resistance protein (TIR-NBS-LRR class), putative | -1.62 |
| 245362_at | At4g17460 | homeobox-leucine zipper protein 1 (HAT1) | -1.62 |
| 246180_at | At5g20840 | phosphoinositide phosphatase | -1.62 |
| 259332_at | At3g03830 | auxin-responsive protein, putative | -1.64 |
| 246215_at | At4g37180 | myb family transcription factor | -1.65 |
| 250012_x_at | At5g18060 | auxin-responsive protein, putative | -1.65 |
| 253075_at | At4g36150 | disease resistance protein (TIR-NBS-LRR class), putative | -1.65 |
| 254573_at | At4g19420 | pectinacetylesterase | -1.65 |
| 266803_at | At2g28930 | protein kinase (APK1b) | -1.66 |
| 252167_at | At3g50560 | short-chain dehydrogenase/reductase (SDR) | -1.66 |
| 257950_at | At3g21780 | UDP-glucuronosyl/UDP-glucosyl transferase | -1.66 |
| 249606_at | At5g37260 | myb family transcription factor | -1.67 |
| 246275_at | At4g36540 | basic helix-loop-helix (bHLH) | -1.67 |
| 260362_at | At1g70530 | protein kinase | -1.68 |
| 266313_at | At2g26980 | CBL-interacting protein kinase 3 (CIPK3) | -1.68 |
| 253181_at | At4g35180 | amino acid transporter | -1.68 |
| 255602_at | At4g01026 | expressed protein | -1.69 |

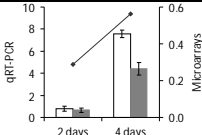
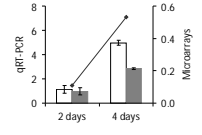
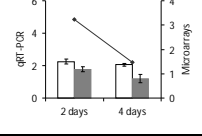
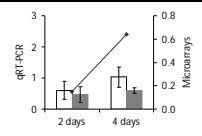
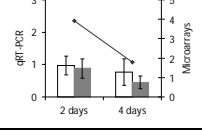
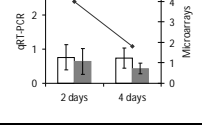
| | | | |
|-------------|-----------|--|-------|
| 245005_at | rps14 | rps14 | -1.69 |
| 245493_at | At4g16380 | At4g16380 | -1.71 |
| 259784_at | At1g29450 | auxin-responsive protein, putative | -1.71 |
| 249923_at | At5g19120 | expressed protein | -1.71 |
| 253493_at | At4g31820 | phototropic-responsive NPH3 | -1.71 |
| 252862_at | At4g39830 | L-ascorbate oxidase, putative | -1.72 |
| 252040_at | At3g52060 | expressed protein | -1.72 |
| 264445_at | At1g27290 | expressed protein | -1.73 |
| 257516_at | At1g69040 | ACT domain containing protein (ACR4) | -1.74 |
| 253626_at | At4g30640 | F-box (FBL19) | -1.75 |
| 260567_at | At2g43820 | UDP-glucuronosyl/UDP-glucosyl transferase | -1.76 |
| 259076_at | At3g02140 | expressed protein | -1.76 |
| 266259_at | At2g27830 | expressed protein | -1.77 |
| 253298_at | At4g33560 | expressed protein | -1.77 |
| 257764_at | At3g23010 | disease resistance | -1.78 |
| 259879_at | At1g76650 | calcium-binding EF hand | -1.79 |
| 259977_at | At1g76590 | zinc-binding | -1.79 |
| 256789_at | At3g13672 | seven in absentia (SINA) | -1.79 |
| 262399_at | At1g49500 | expressed protein | -1.8 |
| 267238_at | At2g44130 | kelch repeat-containing F-box | -1.81 |
| 258906_at | At3g06380 | F-box | -1.82 |
| 256098_at | At1g13700 | glucosamine | -1.83 |
| 252088_at | At3g52100 | expressed protein | -1.83 |
| 245181_at | At5g12420 | expressed protein | -1.84 |
| 258487_at | At3g02550 | lateral organ boundaries domain protein 41 (LBD41) | -1.84 |
| 250860_at | At5g04770 | amino acid permease | -1.85 |
| 257153_at | At3g27220 | kelch repeat-containing protein | -1.85 |
| 249383_at | At5g39860 | bHLH protein | -1.87 |
| 245566_at | At4g14610 | At4g14610 | -1.88 |
| 259787_at | At1g29460 | auxin-responsive protein, putative | -1.9 |
| 266140_at | At2g28120 | nodulin | -1.9 |
| 265387_at | At2g20670 | expressed protein | -1.92 |
| 254931_at | At4g11460 | protein kinase | -1.93 |
| 259790_s_at | At1g29430 | auxin-responsive | -1.94 |
| 252234_at | At3g49780 | phytosulfokines 3 (PSK3) | -1.96 |
| 265342_at | At2g18300 | basic helix-loop-helix (bHLH) | -1.97 |
| 267115_s_at | At2g32540 | cellulose synthase | -1.97 |
| 264836_at | At1g03610 | expressed protein | -1.99 |
| 256948_at | At3g18920 | zinc finger (C3HC4-type RING finger) | -2.01 |
| 267093_at | At2g38170 | calcium exchanger (CAX1) | -2.02 |
| 266695_at | At2g19810 | zinc finger (CCCH-type) | -2.02 |
| 264788_at | At2g17880 | DNAJ heat shock protein, putative | -2.04 |
| 265276_at | At2g28400 | expressed protein | -2.04 |
| 259015_at | At3g07350 | expressed protein | -2.05 |
| 264846_at | At2g17850 | senescence-associated | -2.07 |

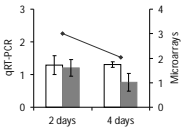
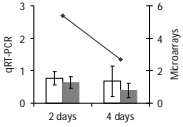
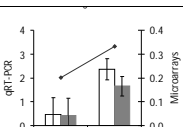
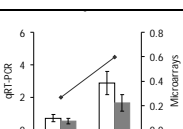
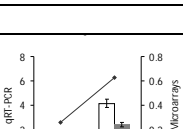
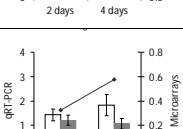
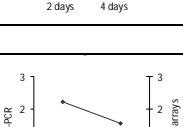
| | | | |
|-----------|-----------|--|-------|
| 254027_at | At4g25835 | AAA-type ATPase | -2.07 |
| 260126_at | At1g36370 | glycine hydroxymethyltransferase, putative | -2.1 |
| 255807_at | At4g10270 | wound-responsive | -2.2 |
| 259466_at | At1g19050 | two-component responsive regulator | -2.21 |
| 247524_at | At5g61440 | thioredoxin | -2.3 |
| 266072_at | At2g18700 | glycosyl transferase family 20 protein | -2.36 |
| 250327_at | At5g12050 | expressed protein | -2.44 |
| 251039_at | At5g02020 | expressed protein | -2.49 |
| 248606_at | At5g49450 | bZIP family transcription factor | -2.5 |
| 252367_at | At3g48360 | speckle-type POZ protein-related | -2.53 |
| 249862_at | At5g22920 | zinc finger (C3HC4-type RING finger) | -2.58 |
| 265067_at | At1g03850 | glutaredoxin | -2.61 |
| 263302_at | At2g15190 | pseudogene, Ulp1 protease family | -2.66 |
| 258930_at | At3g10040 | expressed protein | -2.83 |
| 261567_at | At1g33055 | expressed protein | -3.24 |
| 266814_at | At2g44910 | homeobox-leucine zipper protein 4 (HB-4) | -3.95 |
| 245276_at | At4g16780 | homeobox-leucine zipper protein 4 (HAT4) | -4.71 |

¹Affymetrix microarray ATH1

²From the TAIR8 (The Arabidopsis Information Resource) database (<http://www.arabidopsis.org>)

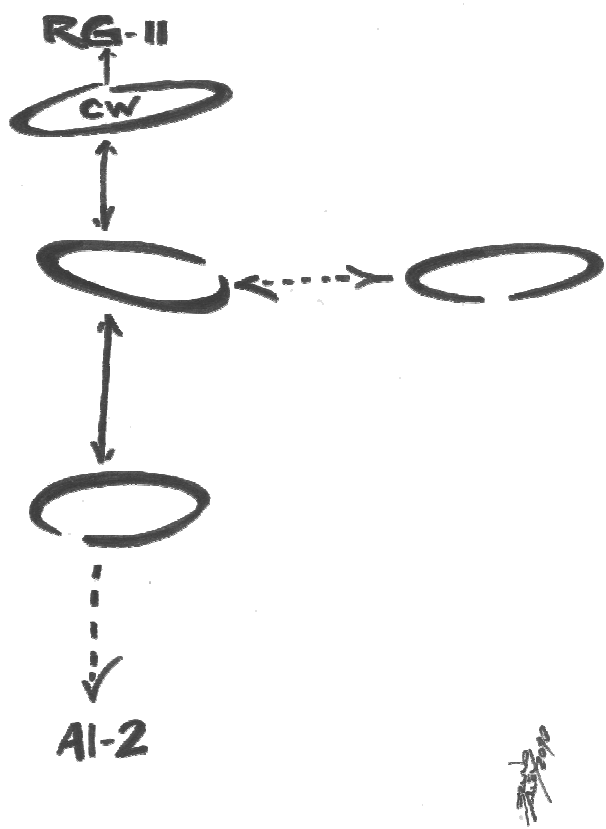
Supplementary Table 5.2. Genes selected to be validated by RT-qPCR. The thirteen genes selected were differentially expressed 2 and 4 days after B deficiency. The primers used, amplicons specifications [E(%), efficiency; AL(nt) amplicon length; Ta(°C) annealing temperature] and relative expression are described.

| AGI ID | 5'→3' Primer Sequences (Forward/Reverse) | | | Relative expression Affymetrix ATH1 microarray (◆), RT-qPCR using two house-keeping genes: UBC10 (□) and PP2A (■). |
|------------------------|--|---------|---------|---|
| | E (%) | AL (nt) | Ta (°C) | |
| Housekeeping genes | | | | |
| At1g13320 | TAACGTGGCCAAAATGATGC GTTCTCCACAACCGTTGGT | | | |
| | 90.6 | 105 | 55 | |
| At5g53300 | ATGGGTCCTTCAGAGAGTCCT CTTGGTCCTAAAGGCCACCT | | | |
| | 94.8 | 105 | 60 | |
| Cell wall biosynthesis | | | | |
| At2g19800 | CTTGTTGAACATTTGTTCTGA CAAAACCTCCATCCAACACC | | |  |
| | 90.8 | 91 | 59 | |
| At4g26260 | ATCATAAGATACCATTCTTTACCC TGCTTCCTCGTTCATAAGATGG | | |  |
| | 102.4 | 73 | 59 | |
| At1g64390 | GCTACTACTCACTCAGCCTATCA CGGTAGTATTCTGGGCAACA | | |  |
| | 105.0 | 96 | 61 | |
| Sulphur metabolism | | | | |
| At1g64660 | ACCGTGCTCAACCTCAGC GTTGCAGCATCACTGACGATA | | |  |
| | 101.7 | 103 | 61 | |
| At1g65860 | GTTGCTCATATTCCTGGGATAAA AATTTCCGATCACCACCACT | | |  |
| | 105.6 | 109 | 62 | |
| At1g62560 | GAGTTCCTGATCCATTGAAAA CACCACTCGCAAAATTCCT | | |  |
| | 96.9 | 60 | 61 | |

| | | | | |
|---|---|-----|----|--|
| At4g04610 | CTGTGAAGATGGCAATGTCTG ACCTCAACGAACCAATTTGC | | |  |
| | 92.0 | 114 | 61 | |
| At5g24660 | AGAAGCGGAGGAGCGTCT CGAGCCTGGTCTAAAGATTCTG | | |  |
| | 102.4 | 66 | 61 | |
| Branched-chain amino acid (BCAA) catabolism | | | | |
| At1g10070 | TCACAAATTATGCCCAGTT CGAGATAAAGAACGTCTGAAAACC | | |  |
| | 102.9 | 75 | 61 | |
| At1g03090 | AGAGACATGGGTGATAAAAGTGC TATCCAGGCACGAGAGGAAC | | |  |
| | 108.4 | 71 | 55 | |
| Transcription | | | | |
| At1g79700 | ATCAAAATACAGAGGCGTTGC TCTTGCGTGGCGTATGTTT | | |  |
| | 98.6 | 111 | 61 | |
| At1g76410 | TCGTGGAGTGTGCTATCTGTTT GCAACACCCTAAGCTCGTCT | | |  |
| | 105.2 | 60 | 62 | |
| Unknown | | | | |
| At4g15920 | GCCGTTTTGGTTGTCGTT ACACAAACCCCACTCCATT | | |  |
| | 97.1 | 110 | 61 | |

CHAPTER 6

GENERAL DISCUSSION AND FUTURE PERSPECTIVES



GENERAL DISCUSSION

The aim of this Thesis was to increment knowledge of the role of B in higher plant metabolism. For this purpose, long-term effects of B deficiency were studied in *Lupinus albus* and early effects were studied in *Arabidopsis thaliana* using distinct approaches. The experimental work provided clear evidence of B requirements during plant growth and development, not only by the marked morphological changes observed, but also by the significant alterations detected in the plant metabolism.

Long-term B deficiency effects

The marked alterations observed in the leaves and roots of *Lupinus albus* after 4 weeks of withholding B, gave indications on the long-term B deficiency responses. The analysis of the metabolite content of *Lupinus albus* plants revealed minor changes in the content of sugars and absence of variation in malate (Chapter 4). The fact that malate is considered to be the ultimate product of glycolysis (Schulze et al. 2002) seems to indicate that the central carbohydrate metabolism is being little affected by B deficiency. Furthermore, in the study of the root proteome it was observed the *de novo* expression of UDP-glucose pyrophosphorylase, that is part of an alternative biochemical pathway for sucrose degradation (Chapter 3) and some ATPases were suppressed. Therefore, there

appears to be an activation of alternative energy sources that will permit the carbon flow to continue under stressful conditions with a reduced ATP demand. Indeed, pyrophosphate is known to be produced in large amounts in the plant cells as a by-product of anabolic reaction and its preferential utilization as an energy donor is particularly important when cellular ATP pools become diminished during stress (e. g. cold, anoxia, phosphate starvation) (Stitt 1998). Such a metabolic flexibility that helps plants to survive stress thus appears to be operating under B deficiency.

Concomitant with the changes observed in energy metabolism we observed marked shifts in the expression of proteins related with several metabolic processes, such as those associated with the cell wall, protein and amino acid metabolism, defence responses and cytoskeleton biosynthesis. Considering the cell wall, proteins related with the synthesis of extracellular matrix polysaccharides were suppressed, supporting the observation that B deficiency causes the decreased expression of several cell wall modifying enzymes (Camacho-Cristóbal et al. 2008).

Proteins related to protein folding and proteolytic processes were markedly affected by B deficiency, some being suppressed and others expressed *de novo*. This observation points out that a shift in protein metabolism had occurred. It has been suggested that a repression of protein degradation is a cellular response to stress in

order to reduce ATP demand (Geigenberger 2003). It is not directly evident if the altered free amino acid contents (Chapter 4) could somehow be related with the described alterations in protein metabolism, and/or if they could act as signalling molecules.

Several proteins related with cytoskeleton biosynthesis were affected by B deficiency. For instance, tubulins and actins were expressed *de novo* (Chapter 3). The additional increased content of the branched-chain amino acids (Chapter 4), possibly related with cytoskeleton biosynthesis (Kimball and Jefferson 2006, Rhoads and Wu 2009), also suggests an involvement of B with cytoskeleton biosynthetic processes.

The long-term effects just discussed could result from both direct and indirect stress responses to B deficiency, and indirect effects could have a high contribution to the overall metabolic responses. However, a certain degree of specificity was suggested from studies of the leaf apoplast (Chapter 2), where the majority of the responsive proteins that were commonly responsive to both B and water-deficit showed different patterns of expression in the two stress regimes.

Several protein isoforms were observed to be responsive to B deficiency in both leaf apoplast and root proteomes (Chapters 2 and 3). The differential expression of different protein isoforms were already found to have some relevance in fungal infection and

symbiosis studies in *Medicago truncatula* (Salzer et al. 2000) and could play a determinant role in the ability of the plant to cope with stressful conditions. Amino acids could also be involved in regulatory processes (Kimball and Jefferson 2006), as referred to above for the branched-chain amino acids. Glycine, that was the only amino acid to show a strong negative effect due to B deficiency, was proposed to have a connection with free levels of Ca^{2+} in the cytosol, important for plant cell signalling processes (Dubos et al. 2003, Hepler 2005).

Short-term B deficiency effects

In *Arabidopsis thaliana* plants subjected to short-term B deficiency, no marked morphological effects were observed, although evident metabolic alterations were detected at the level of the transcriptome (Chapter 5). Altered expression of genes related with the cell wall were observed, which is in accordance with a previous report of down-regulation of cell wall related genes in short-term B-deficient roots of *Arabidopsis* (Camacho-Cristóbal et al. 2008). The other relevant results we obtained concern branched-chain amino acids, sulphur metabolism and transcription-related processes. The increased expression of branched-chain amino acid transferase gene was also recently reported by Kasajima et al. (2010) in B-deficient wild-type *Arabidopsis thaliana* plants. The increased expression of this

gene could be related with impaired cytoskeleton biosynthesis since in animal systems, branched-chain amino acids were described to stimulate muscle protein synthesis (Rhoads and Wu 2009). Furthermore, increased levels of actin and tubulin with altered polymerization patterns have been reported in *Arabidopsis* roots in response to short-term B deficiency (Yu et al. 2001, 2003).

Concerning sulphur metabolism, we observe that the three genes of 5'-adenylylsulfate reductase (APR) are repressed in response to B deficiency. APR is a key enzyme of the sulphate assimilation pathway and the three genes constitute the entire family so far described (Gutierrez-Marcos et al. 1996, Vauclare et al. 2002). Hence, B deficiency seems to trigger responses similar to sulphate excess (Saito 2004).

Of the five genes involved in transcriptional processes, which were differentially expressed in B-deficient plants, three of them are associated with hormonal responses (e. g. auxin, cytokinin and jasmonic acid). Considering that there are as yet few studies relating B deficiency and plant hormones, this observation could be a starting point for further studies.

A plausible hypothesis for B participation in plant metabolism

Several hypotheses on the participation of B in plant metabolism have been proposed (Brown et al. 2002, Goldbach and Wimmer

2007, Bonilla et al. 2009). A few authors have proposed a participation of B in binding *cis*-diol molecules. However, the cell contains high abundance of many molecules possessing this chemical characteristic, including sugar moieties such as mannose, apiose, galactose (cell wall related), ribose (a major component of ribonucleic acids and nucleotides), glycoproteins and glycolipids (minor component of membranes) and other molecules like serine or threonine (e. g. proteins), which are also capable of forming stable B complexes (Loomis and Durst 1992, Brown et al. 2002). Considering that B is a relatively scarce micronutrient in plants and that about 90% of the B is covalently linked to the cell wall structure (Martini and Thellier 1993), the amount of B available for forming potential alternative interactions with compounds other than RG-II is severely limited. It is difficult to imagine how the loss of such a minor and non-specific interaction may lead to the observed symptoms of B deficiency and to alterations in specific areas of metabolism.

However, although the specific cellular mechanisms remain unknown, there is a large body of empirical evidence that B is specifically attached to RG-II during the biosynthesis of extracellular matrix components, implying the operation of highly specific targeting mechanisms. The demonstrable importance of B in other areas of metabolism, including that in animal systems which are devoid of

cell walls, at least allows us to consider the possibility of additional mechanisms operating for the targeted delivery of B to other important acceptor molecules.

For instance, boronic acids and B containing peptides are known to be inhibitors of serine proteases (Kettner and Shenvi 1984, Nakamura et al. 2009). Serine proteases have been described as modulators of a high number of metabolic events in plants (Antão and Malcata 2005), including the infection process of plant cells during symbiosis (Ribeiro et al. 1995) and in a defence process that comprises an oxidative burst (Yano et al. 1999). The putative role of serine proteases in differentiation of specialized plant tissues was also proposed to be mediated by signalling processes associated with secondary cell wall synthesis (Groover and Jones 1999).

Some of metabolic processes affected by serine proteases are common to the processes affected by B depletion (Chapter 1), suggesting that at least part of the plant response to B deficiency may be mediated by B effects on serine proteases. This is an avenue of research which would be interesting to explore further.

Conclusions

In conclusion, the alteration of cytoskeleton biosynthetic processes appears to be a major result from both short- and long-term B deficiency.

Long-term effects of B deficiency on other areas of metabolism are also clearly observable. However, these almost certainly include those which are not directly affected by B deficiency, but result from down-stream events which are epistatic to earlier responses. These might include the later shift to alternative energy pathway, or other epistatic effects from the activation of signal pathways, including those involving, for instance ABA and jasmonate.

However, changes to the biosynthesis of the cytoskeleton might occur as a more immediate result to alterations in cell wall structure; whether this occurs as a result of perturbations in the cell wall-plasmalemma-cytoskeleton continuum, or due to other types of B interactions with the cytoskeleton remains to be elucidated. Such B participation in higher plant metabolism is consistent with the observation of higher B requirements in the early phases of animal development (Rowe and Eckhert 1999) and reproductive stages in plants (Behrendt and Zoglauer 1996), where the cytoskeleton provides an essential structure during cell division processes (Gunning and Hardham 1982).

In addition to the observed changes in cell wall metabolism that corroborate the described participation of B in this structure, an important novel link between sulphur metabolism and B deficiency was also proposed with the short-term B deficiency studies in *A. thaliana*.

FUTURE PERSPECTIVES

Although analytical instruments and procedures have steadily improved during the last decade, further progress greatly depends on the development of new methodologies with a higher capability for analytical imaging of B isotopes (^{10}B and ^{11}B) at physiological concentrations in plant tissues, in order to identify B-binding ligands. Promising biological tools are also available for an attempt to identify additional functions for B that may include for instance, the use of mutants available for *Arabidopsis thaliana* plants. Considering the previous discussion, it would seem interesting to assess if knock-out mutants in cytoskeleton components would have an ameliorated response to extra B supply, or if the over-expression of APR, the key enzyme in sulphur assimilation metabolism, would reduce symptoms of B deficiency.

REFERENCES

- Antão C, Malcata F (2005) Plant serine proteases: biochemical, physiological and molecular features. *Plant Physiol Biochem* 43: 637–650.
- Appel L, Prout M, Abu-Shumays R, Hammonds A, Garbe J, Fristrom D, Fristrom J (1993) The *Drosophila* Stubble-stubloid gene encodes an apparent transmembrane serine protease required for epithelial morphogenesis. *PNAS* 90: 4937–4941.
- Behrendt U, Zoglauer K (1996) Boron controls suspensor development in embryogenic cultures of *Larix decidua*. *Physiol Plant* 97: 321–326.
- Bonilla I, Blevins D, Bolaños L (2009) Boron Functions in Plants: Looking Beyond the Cell Wall. In: *Plant Physiology*. Eds: Taiz L, Zeiger E. 4th Ed. IOP Publishing physics web. <http://4e.plantphys.net>. Accessed Dec 2009.
- Brown P, Bellaloui N, Wimmer M, Bassil E, Ruiz J, Hu H, Pfeiffer H, Dannel F, Romheld V (2002) Boron in plant biology. *Plant Biol* 4: 203–223.
- Camacho-Cristóbal J, Herrera-Rodríguez M, Beato V, Rexach J, Navarro-Gochicoa M, Maldonado J, González-Fontes A (2008) The expression of several cell wall-related genes in *Arabidopsis* roots is down-regulated under boron deficiency. *Env Exp Bot* 63: 351–358.
- Dubos C, Huggins D, Grant G, Knight M, Campbell M (2003) A role for glycine in the gating of plant NMDA-like receptors. *Plant J* 35: 800–810.

- Geigenberger P (2003) Response of plant metabolism to too little oxygen. *Curr Opin Plant Biol* 6: 247-256.
- Goldbach H, Wimmer M (2007) Boron in plants and animals: Is there a role beyond cell-wall structure? *J Plant Nutr Soil Sci* 170: 39-48.
- Greenberg D, Mize G, Takayama T (2003) Protease-Activated Receptor Mediated RhoA Signaling and Cytoskeletal Reorganization in LNCaP Cells. *Biochem* 42: 702-709.
- Groover A, Jones A (1999) Tracheary element differentiation uses a novel mechanism coordinating programmed cell death and secondary cell wall synthesis. *Plant Physiol* 119: 375-384.
- Gunning B, Hardham A (1982) Microtubules. *Annu Rev Plant Physiol* 33: 651-698.
- Gutierrez-Marcos J, Roberts M, Campbell E, Wray J (1996) Three members of a novel small gene-family from *Arabidopsis thaliana* able to complement functionally an *Escherichia coli* mutant defective in PAPS reductase activity encode proteins with a thioredoxin-like domain and "APS reductase" activity. *PNAS* 93: 13377-13382.
- Hepler P (2005) Calcium: A Central Regulator of Plant Growth and Development. *Plant Cell* 17: 2142-2155.
- Kasajima I, Ide Y, Hirai M, Fujiwara T (2010) WRKY6 is involved in the response to boron deficiency in *Arabidopsis thaliana*. *Physiol Plant* (*in press*).
- Kettner C, Shenvi A (1984) Inhibition of the serine proteases leukocyte elastase, pancreatic elastase, cathepsin G, and chymotrypsin by peptide boronic acids. *JBC* 259, 15106-15114.

- Kimball S, Jefferson L (2006) New functions for amino acids: effects on gene transcription and translation. *Am J Clin Nutr* 83: 500S-507S.
- Loomis W, Durst R (1992) Chemistry and biology of boron. *BioFactors* 3: 229-239.
- Martini F, Thellier M (1993) Boron distribution in parenchyma cells of clover leaves. *Plant Physiol Biochem* 31: 777-786.
- Nakamura H, Watanabe M, Ban H, Nabeyama W, Asai A (2009) Synthesis and biological evaluation of boron peptide analogues of Belactosin C as proteasome inhibitors. *Bioorg Med Chem Lett* 19: 3220-3224.
- Navarro-García F, Canizalez-Roman A, Sui B, Nataro J, Azamar Y (2004) The Serine Protease Motif of EspC from Enteropathogenic *Escherichia coli* Produces Epithelial Damage by a Mechanism Different from That of Pet Toxin from Enteroaggregative *E. coli*. *Infec Immun* 72: 3609-3621.
- Navarro-García F, Sears C, Eslava C, Cravioto A, Nataro J (1999) Cytoskeletal Effects Induced by Pet, the Serine Protease Enterotoxin of Enteroaggregative *Escherichia coli*. *Infec Immun* 67: 2184-2192.
- Rhoads J, Wu G (2009) Glutamine, arginine, and leucine signaling in the intestine. *Amino Acids* 37: 111-122.
- Ribeiro A, Akkermans A, van Kammen A, Bisseling T, Pawlowski K (1995) A nodule-specific gene encoding a subtilisin-like protease is expressed in early stages of actinorhizal nodule development. *Plant Cell* 7: 785-794.
- Rowe R, Eckhert C (1999) Boron is required for zebrafish embryogenesis. *J Exp Biol* 202: 1649-1654.

- Saito K (2004) Sulfur assimilatory metabolism. The long and smelling road. *Plant Physiol* 136: 2443–2450.
- Salzer P, Bonanami A, Beyer K, Vögeli-Lange R, Aeschbacher R, Lange J, Wiemken A, Kim D, Cook D and Boller T (2000) Differential Expression of eight chitinase genes in *Medicago truncatula* roots during mycorrhiza formation, nodulation and pathogen infection. *MPMI* 13: 763–777.
- Schulze J, Tesfaye M, Litjens R, Bucciarelli B, Trepp G, Miller S, Samac D, Allan D, Vance C (2002) Malate plays a central role in plant nutrition. *Plant Soil* 247: 133–139.
- Stitt M (1998) Pyrophosphate as an alternative energy donor in the cytosol of plant cells: an enigmatic alternative to ATP. *Bot Acta* 111: 167–175.
- Tornero P, Conejero V, Vera P (1996) Primary structure and expression of a pathogen-induced protease (PR-P69) in tomato plants: similarity of functional domains to subtilisin-like endoproteases. *PNAS* 93: 6332–6337.
- Vauclare P, Kopriva S, Fell D, Suter M, Sticher L, von Ballmoos O, Krähenbuhl U, Op den Camp R, Brunold C (2002) Flux control of sulphate assimilation in *Arabidopsis thaliana*: adenosine 5'-phosphosulphate reductase is more susceptible to negative control by thiols than ATP sulphurylase. *Plant J* 31: 729–740.
- Yano A, Suzuki K, Shinshi H (1999) A signaling pathway, independent of the oxidative burst, that leads to hypersensitive cell death in cultured tobacco cells includes a serine protease. *Plant J* 18: 105–109.
- Yu Q, Baluška F, Jasper F, Menzel D, Goldbach H (2003) Short-term boron deprivation enhances levels of cytoskeletal

proteins in maize, but not zucchini, root apices. *Physiol Plant* 117: 270–278.

- Yu Q, Wingender R, Schulz M, Baluska F, Goldbach H (2001) Short-term boron deprivation induces increased levels of cytoskeletal proteins in *Arabidopsis* roots. *Plant Biol* 3: 335–340.

