

## Parvalbumin promoter hypermethylation in postmortem brain in schizophrenia

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### Published version

FACHIM, Helene, SRISAWAT, Umarat, DALTON, Caroline and REYNOLDS, Gavin (2018). Parvalbumin promoter hypermethylation in postmortem brain in schizophrenia. *Epigenomics*, 10 (5), 519-524.

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## Parvalbumin promoter hypermethylation in post-mortem brain in schizophrenia

Journal:	<i>Epigenomics</i>
Manuscript ID	EPI-2017-0159.R1
Manuscript Type:	Short Communication
Keywords:	DNA methylation, schizophrenia, parvalbumin

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1 **Abstract**

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3 Deficits of brain parvalbumin (PV) are a consistent finding in schizophrenia and  
4 models of psychosis. We investigated whether this is associated with abnormal PV  
5 gene (PVALB) methylation in the brain in schizophrenia. Bisulfite pyrosequencing  
6 was used to determine cytosine (CpG) methylation in a PVALB promoter sequence.  
7 Greater PVALB methylation was found in schizophrenia hippocampus, while no  
8 differences were observed in prefrontal cortex. LINE-1 methylation, a measure of  
9 global methylation, was also elevated in both regions in schizophrenia, although the  
10 PVALB change was independent of this effect. These results provide the first  
11 evidence that PVALB promoter methylation is abnormal in schizophrenia and suggest  
12 that this epigenetic finding may relate to the reduction of PV expression seen in the  
13 disease.

14

15 **Keywords:** Schizophrenia, parvalbumin, DNA methylation, post-mortem brain,  
16 LINE-1.

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## 38 1. Introduction

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40 It is now well established that there is a dysfunction of GABAergic systems in  
41 the brain in schizophrenia. Early post-mortem studies have shown deficits in  
42 interneurons in the neocortex and hippocampus [1] reflected by lower density of  
43 hippocampal GABA uptake sites [2]. Subsequent confirmation has come from  
44 observations of deficits in the GABAergic marker glutamic acid decarboxylase  
45 (GAD)-1 mRNA and GAD-67 protein throughout the cortex [3]. These deficits  
46 appear to be selective for subtypes of GABAergic neurons, most notably those  
47 containing parvalbumin (PV), immunostaining for which is reduced in frontal cortex  
48 and hippocampus in schizophrenia [4,5]. It seems likely that these deficits contribute  
49 to the cognitive disturbances in schizophrenia [6], although it is conceivable that  
50 hippocampal parvalbumin/GABA deficits may result in dopaminergic hyperfunction  
51 [2] and thereby contribute to positive symptoms.

52 The pathogenic mechanisms underlying the PV deficit in schizophrenia are  
53 also unclear, although it has been suggested that the GABAergic cells are intact but  
54 hypofunctioning [7]. This is consistent with the fact that the PV deficit in certain  
55 animal models of the disease appears to be related to a reversible effect of oxidative  
56 stress [8]. We have speculated whether the PV deficit might relate to epigenetic  
57 changes that could be induced by such environmental influences. One epigenetic  
58 factor is that of DNA methylation occurring at cytosine residues in CpG sequences;  
59 within promoter sequences this methylation can have major effects on gene  
60 expression [9]. There is some evidence for dynamic effects on methylation of the PV  
61 gene (PVALB) promoter sequence associated with manganese-induced neurotoxic  
62 damage in the mouse hippocampus [10]; also, we recently found a PVALB  
63 hypermethylation in the hippocampus of rats undergoing subchronic phencyclidine  
64 administration [11], in which a PV immunostaining and mRNA deficits are well-  
65 established [12–16]. Additionally, a specific association between elevated PVALB  
66 methylation and methamphetamine (METH)-induced psychosis was reported in  
67 METH-dependent subjects compared to controls with no history of drug abuse or  
68 psychiatric diagnosis [17].

69 We hypothesise that changes in methylation of the PVALB promoter might  
70 relate to PV deficits in schizophrenia. Thus we have determined the methylation  
71 status of several CpG methylation sites within this sequence in frontal cortical and

72 hippocampal tissue taken post-mortem from patients with schizophrenia and control  
73 subjects. The results were compared with a global measure of DNA methylation, that  
74 of LINE-1.

75

## 76 2. Material and Methods

77

### 78 2.1. *Post-mortem human brain tissue*

79 A post-mortem brain tissue sample from 15 schizophrenia subjects and 16  
80 age-matched controls was collected at the University of Nottingham; this sample was  
81 previously investigated for glutamatergic and GABAergic markers (e.g. Reynolds et  
82 al., 1990). Tissues were taken and stored at -70°C in compliance with the UK Human  
83 Tissue Act. Details of the sample subjects are provided in Table 1.

84

### 85 2.2. *DNA extraction, Bisulphite Conversion and Pyrosequencing*

86 Genomic DNA from human samples was extracted from PFC and  
87 hippocampus, using QIAamp DNA Blood Mini Kit (Qiagen, Valencia, CA), and was  
88 bisulphite-modified to convert unmethylated cytosine residues to uracil using the  
89 EpiTec Fast DNA Bisulphite Kit (Qiagen) with a calculated mean conversion of 99%.  
90 We identified an equivalent DNA sequence to that chosen previously in an animal  
91 study [11], in the 5' regions of the human PVALB gene and developed a  
92 pyrosequencing method for determination of methylation at each CpG sites within  
93 this sequence following bisulphite reaction. The sequence was amplified by PCR  
94 using primers, including a biotinylated reverse primer, as follows: 5'-  
95 AGTGGAGAGAGAAAGGGAGTA-3' (forward) and 5'-  
96 [btn]AACACCAAAAAAAAAACCACCTCTAAAATT-3' (reverse) (Eurofins MWG  
97 Operon).

98 PyroMark Q24 CpG LINE-1 sequence-based pyrosequencing was used to quantify  
99 methylation at four CpG sites in positions 331 to 318 of LINE-1 (GenBank accession  
100 number X58075) (Qiagen).

101 PCR reactions, amplification conditions and the methylation profile were  
102 carried out according to our previous study [11]. The sequencing primer used for  
103 PVALB studies was as follows: 5'- ATTAGTTAAGGTTTTAGATTGA -3'  
104 (Eurofins MWG Operon). Pyrosequence setup and data reading were conducted by  
105 PyroMark Q24 2.0.6.20 software (UK). Samples underwent PCR and pyrosequencing

106 in duplicate; any inconsistencies between samples were resolved following further  
107 repetition.

108

### 109 2.3. Statistical Analysis

110 Data obtained from the pyrosequencing were compared by unpaired t test and  
111 were considered significantly relevant when  $p \leq 0.05$ . All the analysis was done using  
112 SPSS 20.0 (IBM Corp: Armonk, NY, USA). Variance analysis was used to evaluate  
113 possible associations of age and sex of the patients with the methylation levels found.

114

## 115 3. Results

116

117 A series of samples of both frontal cortex and hippocampus from 16 control  
118 subjects and 15 schizophrenia subjects (Table 1) successfully underwent bisulphite  
119 conversion, PCR and pyrosequencing to determine methylation in the LINE-1 and  
120 PVALB sequences. All samples demonstrated single PCR bands with no evidence of  
121 DNA degradation. A significant effect of diagnostic category on PVALB methylation  
122 was found in the hippocampus ( $F=3.465$ ;  $p=0.021$ ) but not in the frontal cortex  
123 ( $F=0.715$ ;  $p=0.591$ ). Figure 1 shows that the effect in the hippocampus reflected  
124 increases in methylation in schizophrenia at CpG2 ( $F=8.250$ ;  $p=0.008$ ) and CpG4  
125 ( $F=12.195$ ;  $p=0.002$ ).

126 The mean methylation of LINE-1 was highly significantly increased in both  
127 frontal cortex ( $t=2.995$ ;  $p=0.006$ ) and hippocampus ( $t=2.786$ ;  $p=0.009$ ) in  
128 schizophrenia (Figure 2). Including the respective LINE-1 methylation results as a  
129 covariate in the PVALB analyses above, there were no qualitative differences in the  
130 statistical results: methylation at CpG2 and CpG4 in the hippocampus remained  
131 significantly elevated in schizophrenia.

132 Age was significantly different between the two groups but showed no  
133 significant correlation with any methylation measure; including it as a covariate also  
134 had no substantial influence on the results of the analyses above; differences in LINE-  
135 1 methylation remained significant as did hippocampal PVALB methylation at CpG2  
136 and CpG4.

137

## 138 4. Discussion

139           The major findings from our study indicate a specific increase in PVALB  
140 promoter methylation in the hippocampus in schizophrenia which is independent of  
141 increases in a measure of global methylation in the brain.

142           To the best of our knowledge, this is the first study reporting hypermethylation  
143 in PVALB promoter in schizophrenia patients; these data are supported by previously  
144 identified evidence suggesting that hyperfunctional DNA methylation may be  
145 responsible for deficiencies in GABAergic neurotransmission [18,19]. As DNA  
146 promoter hypermethylation can contribute to reduced gene expression, we suggest  
147 that the well-established reduction in PV expression in the brain in schizophrenia may  
148 be related to increased methylation of CpG sites within the gene promoter region.  
149 The deficit in PV expression is much greater in the hippocampus [5] than in the cortex  
150 [4], which may relate to the fact that a statistically significant hypermethylation was  
151 only observed in the hippocampus. We have reported elevated methylation of an  
152 equivalent sequence in the PVALB promoter of the hippocampus of rats which have  
153 undergone a sub-chronic phencyclidine (PCP) regime, modelling some symptoms of  
154 schizophrenia [11] and also find an elevation of CpG 2 methylation in blood-derived  
155 DNA in subjects with methamphetamine-induced psychosis [17].

156           The PVALB promoter region selected spans many transcription factor (TF)  
157 binding sites including those for paired box domain gene 5 (PAX5) and cyclic AMP-  
158 responsive element (CREB). At CpG2 there is a recognition site for PAX5, which has  
159 an important role in regulate the mid-hindbrain organisation during neurodevelopment  
160 [20–22], while at CpG4 is spanned by the binding site for CREB which possesses  
161 intrinsic histone acetyltransferase activity [23,24] important for gene regulation. It has  
162 been demonstrated that methylation can block this binding [25]. Additionally,  
163 genome-wide association studies have demonstrated that this TF is associated with  
164 schizophrenia [26] and interestingly, increases in DNA methylation of the CREB  
165 binding protein gene following clozapine treatment were significantly correlated with  
166 clinical improvements in treatment-resistant schizophrenia [23].

167           Our results reveal hypermethylation in LINE-1 in brain tissue of schizophrenia  
168 patients compared to controls. These repetitive elements play an important role in  
169 gene expression and may be involved in the regulation of diverse biological  
170 processes, including DNA damage and repair, inflammation, immune function,  
171 embryogenesis, cell differentiation, cell response to external stimuli and hormonal

172 responses [27], so epigenetic dysfunction in these elements in the brain might be  
173 involved in neurodegenerative and psychiatric diseases [28].

174         The increase in LINE-1 methylation indicates that there may be a global  
175 elevation in brain DNA methylation in schizophrenia. It has been reported that in the  
176 brain in schizophrenia there is an upregulation of DNA-methyltransferases (DNMT)  
177 [29,30], so a LINE-1 hypermethylation found in our study could well be a  
178 consequence of this DNMT upregulation. Abnormalities in LINE-1 methylation are  
179 seen in association with early life trauma in schizophrenia [31] and in PTSD [32],  
180 although such studies inevitably rely on blood-derived DNA.

181         However, we found that the increase in PVALB methylation was unrelated to  
182 the change in LINE-1 methylation, and thus it would appear that the finding in  
183 PVALB is an independent effect, perhaps selective to this gene and potentially related  
184 to the specific deficit in PV in the brain in schizophrenia. It was not possible to  
185 determine PV expression in the samples used in the current study, and thus a direct  
186 assessment of the correlation between DNA methylation and gene expression could  
187 not be performed.

188         This study has some further limitations; the sample size was not large and, as  
189 it is a post-mortem study, the patients were inevitably mostly elderly. There are other  
190 variables associated with post-mortem studies that are difficult to control; these  
191 include the post-mortem interval, although modelling this with rat brains at room  
192 temperature over 96 hours has demonstrated no effect on DNA methylation [33].  
193 Furthermore, the patients were not drug free and, as it is known that antipsychotic  
194 drug administration may have effects on gene methylation [34], we cannot distinguish  
195 relationships with disease from effects of drug treatment.

196

## 197 **5. Conclusions and Future Perspectives**

198         This is the first evidence for an elevation of DNA methylation in the promoter  
199 sequence of PVALB in schizophrenia, consistent with recent findings in both drug-  
200 induced psychosis and in an animal model of the disease. This epigenetic effect may  
201 underlie the PV deficits seen in both the disease and the PCP model. The PVALB  
202 hypermethylation occurs in conjunction with, but independent of, increases in a  
203 measure of global DNA methylation in the brain in schizophrenia. Much more needs  
204 to be investigated in order to determine the effects of PVALB promoter methylation  
205 in schizophrenia and related diseases and animal models. It would be important to



206 determine if the hypermethylation seen in these specific CpG sites is directly related  
207 to decreased PV expression in the disease.

208

### 209 **Executive Summary**

- 210 • A deficit of parvalbumin (PV) expression in GABAergic neurons of the  
211 hippocampus and frontal cortex is a feature common to schizophrenia.
- 212 • Increased methylation of the promoter region of the PV gene (PVALB) is  
213 associated with methamphetamine psychosis.
- 214 • Equivalent sequence in rat brain DNA also shows increased methylation in the  
215 phencyclidine model of schizophrenia.
- 216 • We found greater PVALB promoter DNA methylation in hippocampus of  
217 post-mortem schizophrenia patients compared to control subjects.
- 218 • This increase in methylation is specific to a site within a transcription factor  
219 binding sequence.
- 220 • We found hypermethylation in LINE-1 in hippocampus and prefrontal cortex  
221 of schizophrenia post-mortem brains.
- 222 • The changes in PVALB methylation were independent of those in LINE-1.
- 223 • This hypermethylation may, through effects on transcription, contribute to the  
224 enduring reduction in PV in schizophrenia.

225

### 226 **Ethical Conduct of Research**

227

228 The authors state that they have obtained appropriate institutional review  
229 board approval or have followed the principles outlined in the Declaration of Helsinki  
230 for all human or animal experimental investigations.

231

### 232 **Conflict of Interest**

233 The authors report no biomedical financial interests or potential conflicts of  
234 interest.

235

### 236 **Funding**

237 H.A. Fachim has received fellowships from CNPq (Conselho Nacional de  
238 Desenvolvimento Científico e Tecnológico) and FAPESP (proc. no. 2017/00624-5).  
239 G.P. Reynolds has received honoraria for lectures and/or advisory panel membership  
240 from Janssen, Lundbeck, Otsuka, Sumitomo and Sunovion, and a research grant from  
241 Sunovion. The authors have no other relevant affiliations or financial involvement  
242 with any organization or entity with a financial interest in or financial conflict with  
243 the subject matter or materials discussed in the manuscript apart from those disclosed.

244 No writing assistance was utilized in the production of this manuscript.

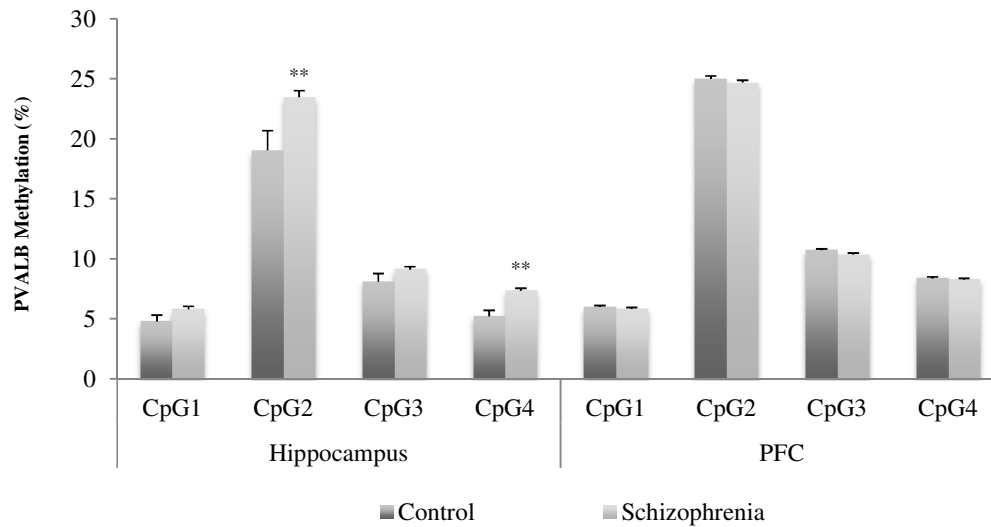
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## 246 6. References

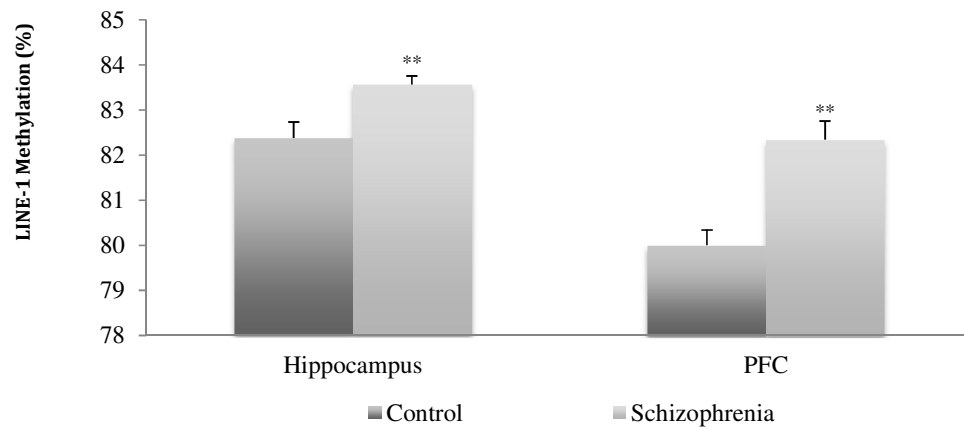
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- 350

**Figure 1.**

**Figure 1.** Percentage methylation PVALB in the hippocampus and prefrontal cortex (PFC) in post mortem brains in schizophrenia and controls of Nottingham Series. Values are expressed as the mean  $\pm$  SEM. (Student's t test, n = 15 schizophrenia and n= 16 controls). \*\*p<0.01.

**Figure 2.**

**Figure 2.** LINE-1 global methylation in the hippocampus and prefrontal cortex (PFC) in post mortem brains in schizophrenia and controls of Nottingham Series. Values are expressed as the mean  $\pm$  SEM. (Student's t test, n = 15 schizophrenia and n = 16 controls). \*\*p < 0.01.

**Table 1.** Description of demographic data of schizophrenia patients and controls

	<b>Control (n = 16)</b>	<b>Schizophrenia (n = 15)</b>	<b>P value</b>
<b>Age (Mean ±SD)</b>	67.25 ± 12.73	52.60 ± 18.18	0.016
<b>Men (%)</b>	11 (68%)	11 (73%)	
<b>PM Hrs (Mean ±SD)</b>	27.68 ± 11.42	27.08 ± 14.05	0.906

\*PM Hrs: *Post-mortem* interval of collection of the samples in hours

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