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OPEN Demographic associations for autoantibodies in diseasefree individuals of a European population

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The presence of autoantibodies usually precedes autoimmune disease, but is sometimes considered an incidental finding with no clinical relevance. The prevalence of immune-mediated diseases was studied in a group of individuals from the Estonian Genome Project (n = 51,862), and 6 clinically significant autoantibodies were detected in a subgroup of 994 (auto)immune-mediated disease-free individuals. The overall prevalence of individuals with immune-mediated diseases in the primary cohort was 30.1%. Similarly, 23.6% of the participants in the disease-free subgroup were seropositive for at least one autoantibody. Several phenotypic parameters were associated with autoantibodies. The results suggest that (i) immune-mediated diseases are diagnosed in nearly one-third of a random European population, (ii) 6 common autoantibodies are detectable in almost one-third of individuals without diagnosed autoimmune diseases, (iii) tissue non-specific autoantibodies, especially at high levels, may reflect preclinical disease in symptom-free individuals, and (iv) the incidental positivity of anti-TPO in men with positive familial anamnesis of maternal autoimmune disease deserves further medical attention. These results encourage physicians to evaluate autoantibodies in addition to treating a variety of patient health complaints to detect autoimmune-mediated disease early.

Autoantibodies are immunoglobulins (Ig) produced by activated autoreactive B cells. The immune response towards self-antigens usually involves activation of both T and B cells, but the detection of autoantibodies in sera is technically simpler than detection of T-cell reactions. Therefore, autoantibodies can be used to guide clinical management of certain diseases. These markers of disease activity and severity help to define and classify diseases and can be used to predict and diagnose specific autoimmune diseases1.

Autoimmune diseases affect at least 5% of the population¹, while the prevalence of diseases that involve immune reactions, including connective tissue diseases (CTD) and diseases with hypersensitivity reactions, is much higher². In reality, the actual burden of various (auto)immune reactions in different populations is unknown.

Some autoantibodies are functional and are therefore considered clinically significant, while the others are bystanders in disease pathogenesis (or their function has not yet been discovered). For example, IgG-type autoantibodies to the 100 kDa membrane bound glycoprotein thyroid peroxidase (anti-TPO) interrupt the production of thyroid hormones and cause autoimmune hypothyroiditis3. In addition, anti-TPO IgGs have been detected in cases of Graves' disease and postpartum thyroid dysfunction, but they have also been detected in control individuals without thyroid disease¹. Therefore, anti-TPO represents an autoantibody with tissue-specificity

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and clinical significance unspecific to thyroiditis. On the contrary to tissue-specific autoantibodies which are produced against antigens expressed in single tissue, tissue non-specific antigens recognise antigens expressed ubiquitously or at least in several tissues. IgA-type autoantibodies against the 78 kDa tissue transglutaminase (anti-tTG) are highly specific to coeliac disease⁴, making them clinically significant but not tissue specific. Although tTG belongs to a family of multifunctional transglutaminases, in coeliac disease, the anti-tTG IgAs produced in the small-intestinal mucosa interrupt the conversion of a glutamine residue into glutamic acid during gluten digestion⁵.

Depending on the disease, time of testing, and the number and detection level of autoantibodies, the sensitivity of predicting autoimmune disease is rarely 100%¹. In other words, there are always individuals who test positive for autoantibodies but have no clinical signs of autoimmune disease for years. Similarly, there are cases where autoimmune disease develops without prior clinical indication. Therefore, the interpretation of positive autoantibody tests can be challenging in diseases such as thyroiditis, as anti-thyroid autoantibodies may precede disease manifestation by two decades, and some individuals (10%) stay disease-free despite the presence of autoantibodies⁶. Data interpretation is further complicated in diseases with complex pathologies, such as CTD, an autoimmune-inflammatory disease⁷. Furthermore, autoantibodies may be produced temporarily to facilitate communication between immune cells and molecules or between immune cells and other tissues, particularly during immune challenges such as viral infections⁸.

The prevalence and relevance of autoantibodies in healthy individuals are poorly studied, and most data found in the literature are derived from assessing autoantibodies in patients with autoimmune diseases^{1,4}. Two outstanding questions that remain unanswered are how often autoantibodies can be detected in clinically healthy individuals and whether the presence of autoantibodies predicts the future onset of autoimmune disease. A prospective follow-up survey of selected individuals would be the gold standard to study these questions⁶, but needs highly synchronized medical efforts for organizing such studies and significant financial resources.

In this study, we aimed to determine the prevalence of selected clinically significant autoantibodies in (auto) immune-mediated disease-free individuals and to carry out an association study to explain the existence of autoantibodies in these healthy individuals. Namely, the data from a population-based registry of 51,862 adults from the Estonian Genome Center at the University of Tartu, Estonia (EGCUT) was used to assess (auto) immune-mediated diseases in the general population of Estonia. The study group of healthy individuals representative of the population was randomly selected from that registry. Study individuals were tested for anti-TPO IgG and 5 tissue non-specific autoantibodies diagnostic of major (auto)immune-mediated diseases. The presence of autoantibodies was assessed in relation to phenotypic characteristics in disease-free individuals.

Materials and Methods

Study population. This cross-sectional study comprised 994 individuals who lacked autoimmune or other immune-mediated diseases. Participants were selected from the 51,862 individuals aged 18 years or older in the EGCUT biopank, which is a population-based biopank compiled from 2002 to 2014 (www.biobank.ee)⁹. All study protocols were conducted in accordance with the Estonian Gene Research Act.

The EGCUT cohort closely reflected the age, sex, and geographical distribution of the Estonian population⁹, and all subjects were recruited voluntarily and randomly by general practitioners and hospital physicians. A computer-assisted personal interview, which included personal, genealogical, educational and occupational history, and lifestyle data, was completed. Anthropometric measurements, blood pressure, and resting heart rate were measured, and venous blood was drawn during the visit. Medical history and current health status were recorded according to International Classification of Diseases -10 (ICD-10) codes. The individuals excluded from the study of diagnoses with (auto)immune-mediated diseases are listed in Supplementary Table S.1. Random selection of the individuals representative of the age and gender distribution in the Estonian population resulted in 1,000 individuals. However, following a secondary survey of medical records of these 1,000 individuals, an additional 6 individuals possessing at least one of the (auto)immune-mediated diseases were excluded from the study group. Thus, the total number of study participants was 994 (491 men and 503 women ranging from 18 to 86 years old). Phenotypic characteristics and ethylenediaminetetracetic-acid-treated (EDTA) plasma samples from each individual were used. Each participant signed an informed consent prior to enrolment, and ethical approval for the study was obtained from the Ethics Review Committee on Human Research at the University of Tartu.

Phenotypic data, including socio-demographic data, smoking and alcohol consumption, medication use, parents' diseases, and female reproductive health-related data were used. Socio-demographic data consisted of age at agreement (age at time of study), nationality, city or rural residence at birth, occupation, and body mass index (BMI). Medication data consisted of medications that were used on a regular basis for the last two months. Medications were classified according to the Anatomical Therapeutic Chemical Classification System level. A study participant was considered to be using a hazardous drug if he/she used a medication associated with drug-induced lupus as described in Chang *et al.*¹⁰.

Antibody tests. All participants were tested for anti-TPO and 5 tissue non-specific autoantibodies - antinuclear antibodies (ANA), which were measured with CTD IgG screening test (anti-CTD), anti-tTG IgA and IgG, cyclic citrullinated peptide IgG (anti-CCP), and antibodies of all isotypes against glutamic acid decarboxylase with molecular weight of 65 kDa (GADA). Anti-CTD was further specified with the eight most common IgG-type autoantibodies: antibodies against double-stranded deoxyribonucleic acid (anti-dsDNA), Sjögren's syndrome type A antigen (anti-SS-A/Ro) protein, Sjögren's syndrome type B antigen (anti-SS-B/La) protein, centromere protein (anti-CENP), histidyl-tRNA synthetase (anti-Jo-1) protein, scleroderma-associated autoantigen of 70 kDa (anti-Scl-70) protein, Smith (anti-Sm) protein, and ribonucleoprotein U1 (anti-U1RNP) protein.

		% (95% confidence interval)		
Disease category	ICD-10 ² category or subcategory	Men N = 17,821	Women N = 34,041	Total N = 51,862
Hypo-, hyperthyroidism or thyroiditis	E03.4, E03.5, E03.8, E03.9, E05.0, E05.9, E06.2, E06.3	0.5% (0.4-0.6)	3.1% (2.9-3.3)	2.2% (2.1-2.3)
Coeliac disease	K90.0	0.04% (0.02-0.08)	0.05% (0.03-0.08)	0.04% (0.03-0.07)
Rheumatoid arthritis	M05.0, M05.1, M05.3, M05.4, M05.8, M05.9, M06.0, M06.1, M06.2, M06.3, M06.4, M06.8, M06.9	0.6% (0.5-0.7)	1.9% (1.7-2.0)	1.4% (1.3-1.5)
Insulin-dependent (type 1) diabetes mellitus	E10	0.5% (0.4-0.6)	0.3% (0.2-0.3)	0.4% (0.3-0.4)
Other diabetes mellitus	E11-14	5.4% (5.1-5.8)	4.5% (4.3-4.7)	4.8% (4.6-5.0)
Diseases of the skin, subcutaneous tissue, musculoskeletal system and connective tissue	L40, L63, L80, L93, M05-09, M12, M30-36, M45, M46	5.3% (5.0-5.7)	7.1% (6.8-7.4)	6.5% (6.3-6.7)
Total ¹		26.4% (25.7-27.0)	32.1% (31.6-32.6)	30.1% (29.7–30.5)

Table 1. Prevalence of common (auto)immune-mediated diseases in Estonian adults. ¹Includes all immune mediated diseases listed in Supplementary Table S.1. The prevalence in each gender was compared with the Proportion test. Bold indicates p < 0.05. ²ICD-10 — International Classification of Diseases-10

GADA were measured with commercial enzyme-linked immunosorbent assay (ELISA) kit (RSR Ltd., Cardiff, UK). Plasma from EDTA-vacutainers was treated with calcium¹¹. According to the manufacturer's instructions, values 5 U/ml or grater were considered positive.

Anti-TPO, anti-CCP, anti-tTG IgA and IgG, anti-CTD, and autoantibodies from CTD tests were analysed with a fully automated fluoro-enzyme immunoassay (FEIA) on an ImmunoCAP 100 (Phadia, Thermo Scientific Corporation, Vantaa, Finland). Positive cut-off values were chosen according to the manufacturer's recommendations: $\geq 100\,\text{IU/ml}$ for anti-TPO, $\geq 10\,\text{U/ml}$ for anti-CCP, $\geq 10\,\text{U/ml}$ for anti-tTG, and an autoantibody ratio $\geq 1.0\,\text{U}$ for anti-CTD. To specify anti-CTD, all positive and grey zone (0.7–1.0) test results were reassessed with the eight most common ANAs: anti-dsDNA, anti-SS-A/Ro, anti-SS-B/La, anti-CENP, anti-Jo-1, anti-Scl-70, anti-Sm, and anti-U1RNP IgG. Anti-dsDNA values 15 IU/ml or more were considered positive, while the cut-off value for the remaining 7 autoantibodies was set to $10\,\text{U/ml}$, according to the manufacturer's instructions.

Statistical analyses. The Welch Two Sample t-test and Proportion test with continuity correction were used to compare the reported phenotypic characteristics and prevalence of autoantibodies between men and women, and p values < 0.05 were considered significant. Multiple logistic regression analyses adjusted for age and stratified by gender were used to find associations between the presence of autoantibodies and phenotypic data. For association analyses, autoantibodies were considered binomial variables for logistic regression - (i) the presence of anti-TPO IgG as a tissue-specific autoantibody and (ii) the presence of at least one of the 5 tissue non-specific autoantibodies: GADA, anti-CCP IgG, anti-tTG IgG and IgA, or anti-CTD IgG. The age at time of study was considered a continuous variable, and a categorical variable of age groups was formed based on the values of the first and third quantiles and median age and compared with the youngest group (18–26 years). Adjusted odds ratios (adORs) were calculated, and corrected p values < 0.05 were considered statistically significant, while corrected p values > 0.05 and < 0.1 were considered tendencies for a significant association. The R3.1.0 Language and Environment was used for statistical analyses.

The data set supporting the results of this article is included within the article and it's Supplementary Table.

Results

Health parameters of the study population. According to data registered by EGCUT by May 2014, approximately 30% of the adult population in Estonia has been diagnosed with some type of (auto)immune-mediated disease (Supplementary Table S.1). As expected, women were diagnosed more frequently than men (32.1% vs. 26.4%), except in the case of insulin-dependent (type 1) diabetes, which occurred more frequently among men (0.5%) than women (0.3%, Table 1). Coeliac disease was diagnosed very rarely in the population (0.04%), and the frequency was not different in men and women. More important in the context of the current study is the fact that 2/3 of the adult population was registered healthy and free from (auto)immune-mediated diseases at the time of recruitment into the EGCUT registry and formed the basis for the selection of our study group – 994 (auto) immune disease-free individuals representative of the entire Estonian population of adults in the distribution of age and gender.

The patient characteristics and health parameters of the study group (994 individuals) are provided in Table 2. The mean age of the participants was 39.9 years. The study population also contained approximately equal numbers of men and women who originated from the city or the countryside. The prevalence of employment and student status was similar between genders, but more men were retired, and more women were unemployed due to reasons other than retirement. Smoking was greater among men. Men also started smoking at younger ages. If men did consume alcohol, they did it more frequently than women. Seventy-three percent of women reported that they had been pregnant, and 68% of all women had at least one child. More women in the study population were normal weight than men, who were more often overweight. The prevalence of both extremes, underweight (BMI <18.5 kg/m²) and obesity (BMI \ge 30 kg/m²), were similar between genders. Regular medication use in the most recent 2 months was also similar between men and women. The prevalence of individuals taking drug(s)

Characteristics ¹	Men (n = 491)	Women (n = 503)	Total (n = 994)			
Age at time of study (yr)	39.8 ± 16.5	39.9 ± 16.1	39.9 ± 16.3			
Place of birth						
City	302 (63.2%, 58.7–67.5)	310 (64.2%, 59.7–68.4)	612 (63.7%, 60.5–66.7)			
Countryside	176 (36.8%, 32.5-41.3)	173 (35.8%, 31.6–40.3)	349 (36.3%, 33.3–39.5)			
Occupation						
Employed	320 (66.8%, 62.4–71.0)	330 (68.3%, 63.9–72.4)	650 (67.6%, 64.5–70.5)			
Pension insurance ²	43 (9.0%, 6.6-12.0)	28 (5.8%, 4.0-8.4)	71 (7.4%, 5.8–9.3)			
Student or serviceman	56 (11.7%, 9.0-15.0)	46 (9.5%, 7.1–12.6)	102 (10.6%, 8.8–12.8)			
Unemployed ³	60 (12.5%, 9.8-15.9)	79 (16.4%, 13.2-20.0)	139 (14.4%, 12.3–16.9)			
Smoking status						
Never smoker	230 (46.8%, 42.4-51.4)	312 (62.0%, 57.6-66.3)	542 (54.5%, 51.4–57.6)			
Former smoker	89 (18.1%, 14.9-21.9)	50 (9.9%, 7.5-13.0)	139 (14.0%, 11.9–16.3)			
Current smoker	172 (35.0%, 30.8–39.5)	142 (28.2%, 24.4-32.4)	313 (31.5%, 28.6–34.5)			
Frequency of alcohol consumption						
Rare – up to few times a year	32 (9.9%, 7.0-13.8)	67 (21.3%, 17.0-26.3)	99 (15.5%, 12.8–18.6)			
Seldom – up to every month	48 (14.8%, 11.2-19.3)	74 (23.5%, 19.0-28.6)	122 (19.1%, 16.2–22.4)			
Moderate or frequent - up to every 2nd day	244 (75.3%, 70.2-79.8)	174 (55.2%, 49.6-60.8)	418 (65.4%, 61.6-69.1)			
Age when started smoking (yr)	18.0 ± 4.0	20.6 ± 6.5	19.7 ± 5.4			
Has been pregnant	_	365 (72.6%, 68.4–76.4)	_			
Age at first pregnancy (yr)	_	22.0 ± 3.8	_			
Live birth(s)						
No children	_	138 (27.4%, 23.6–31.6)	_			
1		97 (19.3%, 16.0–23.1)				
2	_	158 (31.4%, 27.4–35.7)				
≥3		89 (17.7%, 14.5–21.4)				
Active ovarian hormones ⁴	_	349 (69.7%, 65.4–73.6)	_			
Body mass index (kg/m²)						
Normal 18.5-24.9	221 (45.0%, 40.6-49.5)	271 (54.0%, 49.5-58.4)	492 (49.5%, 46.4–52.7)			
Underweight <18.5	8 (1.6%, 0.8–3.3)	16 (3.2%, 1.9–5.2)	24 (2.4%, 1.6-3.6)			
Overweight 25.0–29.9	186 (37.9%, 33.6-42.4)	129 (25.7%, 22.0-29.8)	315 (31.7%, 28.9–34.7)			
Obesity ≥30	76 (15.5%, 12.5–19.1)	86 (17.1%, 14.0–20.8)	162 (16.3%, 14.1–18.8)			
Recently used drugs ⁵						
Cardiovascular system (C)	82 (16.7%, 13.6–20.4)	77 (15.3%, 12.3–18.8)	159 (16.0%, 13.8–18.5)			
Urogenital tract (G)	6 (1.2%, 0.5–2.8)	7 (1.4%, 0.6–3.0)	13 (1.3%, 0.7-2.3)			
Central nervous system (N)	41 (8.4%, 6.1–11.2)	40 (8.0%, 5.8–10.8)	81 (8.1%, 6.6–10.1)			
Respiratory tract (R)	14 (2.9%, 1.6-4.9)	11 (2.3%, 1.1-4.0)	25 (2.5%, 1.7–3.7)			
Immune system (H, J, L)	16 (3.3%, 1.9-5.4)	25 (5.0%, 3.3–7.4)	41 (4.1%, 3.0-5.6)			
Subgroup of immunity-affecting drugs ⁶	68 (13.8%, 11.0-17.3)	78 (15.5%, 12.5–19.0)	146 (14.7%, 12.6–17.1)			
Maternal autoimmune disease ⁷	56 (11.4%, 8.8-14.6)	84 (16.7%, 13.6–20.3)	140 (14.1%, 12.0–16.4)			
Paternal autoimmune disease ⁸	62 (12.6%, 9.9–16.0)	39 (7.8%, 5.6–10.5)	101 (10.2%, 8.4–12.2)			

Table 2. Reported phenotypic characteristics of the study population. ¹Participants lacking data were excluded from this data calculation. ²Pension insurance includes retired persons and unemployed persons with insurance for disability. ³Unemployed includes women on childcare leave. ⁴Women with active ovarian hormones includes women who menstruate and women with primary or secondary amenorrhea who are currently receiving hormone replacement therapy. ⁵Drugs used regularly during last 2 months were classified according to the Anatomical Therapeutic Chemical Classification System. ⁶Drugs that increase risk of developing lupus or other autoimmune diseases¹⁰. ⁷Participants who reported that their mother had at least one of the following autoimmune diseases (ICD-10): thyroiditis (E06), insulin dependent diabetes (E10), other adrenal diseases (E27), sclerosis multiplex (G35), sleep disorders (G47), rhinitis (J30), asthma (J45), atopic dermatitis (L20), psoriasis (L40), urticaria (L50), vitiligo (L80), arthropathies (M00-25), seropositive rheumatoid arthritis (M05), other rheumatoid arthritis (M06), or systemic sclerosis (M34). 8Participants who reported that their father had at least one of the following autoimmune diseases (ICD-10): vitamin B12 deficiency (D51), thyrotoxicosis (E05), insulin-dependent diabetes (E10), sclerosis multiplex (G35), sleep disorders (G47), rhinitis (J30), asthma (J45), atopic dermatitis (L20), allergic contact dermatitis (23), psoriasis (L40), seropositive rheumatoid arthritis (M05), other rheumatoid arthritis (M06), systemic sclerosis (M34), other systemic involvement of connective tissue (M35), ankylosing spondylitis (M45). Statistically significant differences (p < 0.05) are shown in bold. Numeric data are provided as means \pm standard deviation and were compared between men and women with t-test. Non-parametric data are provided as count; the percentage of a row and 95% confidence intervals of percentage were compared between genders with a proportion test.

		% (95% confidence intervals)		
		Men	Women	Total
Autoantibodies	Population	n=491	n=503	n=994
(1) Anti-tTG IgA	994	0.0	1.2 (0.5-2.7)	0.6 (0.2-1.4)
(2) Anti-tTG IgG	994	0.0	0.4 (0.1-1.6)	0.2 (0.0-0.8)
(3) Anti-CCP IgG	994	0.0	1.0 (0.4-2.4)	0.5 (0.2-1.2)
(4) Anti-CTD IgG test	994	3.1 (1.8-5.1)	6.6 (4.6-9.2)	4.8 (3.6-6.4)
Anti-dsDNA IgG	851	1.0 (0.4-2.5)	2.2 (1.2-4.0)	1.6 (1.0-2.7)
Anti-SS-B/La IgG	85¹	0.2 (0.0-1.3)	0.0	0.1 (0.0-0.7)
Anti-SS-A/Ro IgG	85 ¹	0.4 (0.1-1.6)	1.0 (0.4-2.4)	0.7 (0.3-1.5)
Anti-Sm IgG	85 ¹	0.0	0.0	0.0
Anti-U1RNP IgG	85¹	0.0	0.2 (0.0-1.3)	0.1 (0.0-0.7)
Anti-CENP IgG	85¹	0.0	0.6 (0.2-1.9)	0.3 (0.1-1.0)
Anti-Jo-1 IgG	85¹	0.0	0.0	0.0
Anti-Scl-70 IgG	85 ¹	0.0	0.0	0.0
(5) GADA Ig ²	994	7.9 (5.8–10.8)	9.5 (7.2–12.5)	8.8 (7.1–10.7)
≥1 tissue non-specific autoantibodies³	994	14.7 (11.7–18.2)	19.9 (16.5-23.7)	17.3 (15.0–19.8)
(6) Anti-TPO IgG	994	3.7 (2.3-5.8)	10.7 (8.2-13.9)	7.2 (5.7–9.1)
≥1 any autoantibodies ⁴	994	17.7 (14.5-21.5)	29.4 (25.5-33.6)	23.6 (21.1-26.4)

Table 3. Prevalence of autoantibodies in the study population. ¹Antigen-specifying tests were performed on participants who were borderline positive for antinuclear autoantibodies, and the test results (prevalence) were expanded to entire study population. ²Ca²⁺-treated peripheral blood plasma. ³Presence of at least one of five tissue non-specific autoantibodies. ⁴Presence of at least one of six measured autoantibodies. Data are provided as percentage and 95% CI of percentage and prevalence was compared between genders with a proportion test. CCP - cyclic citrullinated peptide, CENP - centromere protein, dsDNA – double-stranded deoxyribonucleic acid, GADA - autoantibodies against glutamic acid decarboxylase (molecular weight of 65 kDa), Jo-1 – histidyltRNA synthetase, SS-A/Ro - Sjögren's syndrome type A antigen, SS-B/La - Sjögren's syndrome type B antigen, Scl-70 - scleroderma-associated autoantigen of 70k Da, Sm - Smith protein, TPO – thyroid peroxidase, tTG - tissue transglutaminase, U1RNP - ribonucleoprotein U1. Statistically significant differences (*p* < 0.05) are shown in bold.

included in the Anatomical Therapeutic Chemical Classification System or capable of inducing autoimmune diseases¹⁰, ranged from 1.3–16.0%, and did not differ between men and women. More women than men (16.7% vs. 11.4%) reported that their mothers had at least one (auto)immune-mediated disease and men reported more frequently same diseases in their father's (Table 2).

Prevalence of and associations with autoantibodies. The most prevalent autoantibodies detected in this study were GADA (8.8%), anti-TPO IgG (7.2%), and anti-CTD (4.8%). The total prevalence of at least one autoantibody was 23.6% (Table 3), while the co-existence of two or more autoantibodies was found in 2% of participants (95% confidence interval (CI): 0.0–8.0). The prevalence of detected autoantibodies in comparison of data from literature is presented in Table 4.

Autoantibodies were divided into two groups for further association analyses according to specificity. Anti-TPO IgG was classified as a tissue-specific autoantibody, and the tissue non-specific autoantibodies included GADA, anti-CCP IgG, anti-tTG IgG and IgA, and anti-CTD IgG. After adjusting for age and stratifying by gender, linear and logistic regression analyses revealed the phenotypic characteristics that were significantly associated with autoantibodies (Table 5). These characteristics were used as adjustments for association analyses of autoantibodies.

Multivariate regression model simultaneously controlled for all significant parameters from Table 5 was used to calculate corrected p value from multiple comparisons (corrected p value < 0.05 was considered statistically significant). These risk factors with the adjusted ORs are shown in Fig. 1. The presence of anti-TPO IgG was assessed in the gender-stratified logistic regression model adjusted for age, maternal autoimmune disease, and occupational status (significant phenotypic parameters for anti-TPO, Table 5). The analyses revealed that the presence of anti-TPO autoantibodies in men was independently associated with (i) the presence of maternal autoimmune disease (adOR = 5.51, p = 0.001 compared to men whose mothers were not suffering from autoimmune diseases), (ii) older age (adOR = 1.05, p = 0.008 for one year of age, data not shown in Fig. 1), and (iii) tended to be associated with occupational status of being student or serviceman (adOR = 3.86, p = 0.065). The same model for women showed an increased risk of anti-TPO for age group older than 52 years (adOR = 3.54, p = 0.014 for ages >52 years compared to the youngest women of 18–26 years of age) and risk for anti-TPO was not increased at younger age groups. The presence of at least one tissue non-specific autoantibody in men was assessed in association with the phenotypic parameters of age, frequency of alcohol consumption, and need for cardiovascular treatment (significant phenotypic characteristics for current autoantibodies, Table 5). This model revealed that the likelihood of having one tissue non-specific autoantibody in men tended to be increased by (i) age (adORs for age groups 27–37 years, 38–52 years, and older than 52 years were 2.15, p = 0.072; 2.21, p = 0.059;

Autoantibody						
Name	Prevalence % (95% CI)	Population	Method	Corresponding disease and its prevalence % (95% CI)	Comments	
	0.6% (0.2-1.4)	Immune-mediated disease-free adults, Estonia	FEIA*	Coeliac disease: Reported in 0.04% (0.03–0.07) of general population, Estonia ³⁴ , Estimated	According to the prevalence	
Anti-tTG IgA 0.3% (0	0.3% (0.1-0.9)	School children, Estonia	FEIA ³⁴	in 0.5–1.0% of general population, USA, UK, and other European countries (for review see	and specificity of anti-tTG IgA ^{37,38} , coeliac disease is clearly	
	0.8%	General population, Natrona County, USA	ELISA ³⁹	ref. 35). Estimated in 0.5% (0.2–0.8) of general population of adults, Denmark ³⁶	underdiagnosed at least in Estonia	
	0.5% (0.2–1.2)	Immune-mediated disease-free adults, Estonia	FEIA*		Logical difference between	
	1% (0-5)	General population, Salt Lake City, Utah, USA	ELISA ⁴¹	Rheumatoid arthritis: Reported in 1.4% (1.3–1.5) of general	autoantibody-prevalence in disease-free individuals and	
Anti-CCP IgG	1%	General population, Southern Brazil	NA ⁴²	population, Estonia, ³⁴ Systemic sclerosis:	two times high prevalence in general population. Autoantibody	
	65% (56–73)	Patients with rheumatoid arthritis, Salt Lake City, Utah, USA	ELISA ⁴¹	Estimated in 0.024% of general population, Australia, Spain, USA (for review see ref. 40)	prevalence is comparable to reported cases of rheumatoid arthritis in general population.	
	11.5%	Patients with systemic sclerosis, Southern Brazil	NA ⁴²			
	4.8% (3.6-6.4)	Immune-mediated disease-free adults, Estonia	FEIA*	Various CTDs: Reported diseases of the skin, subcutaneous tissue, musculoskeletal system	Antinuclear autoantibodies detected by CTD panel have simila	
Anti-CTD IgG:	<4%	General population, Belgium	FEIA ²⁰	and connective tissue in 6.5% (6.3-6.7) of	prevalence in different population and reflect general prevalence of	
	2.0%	General population, Australia	ELISA ¹⁹	general population, Estonia ³⁴	CTDs.	
Anti-dsDNA	1.6%(1.0-2.7)	Immune-mediated disease-free adults, Estonia	FEIA*	Lupus erythematosus ⁴³ : Reported in 0.06% (0.05–0.07) in general population, Buenos	Specified panel of anti-CTD has been provided in diseased persons	
Aliu-usDIVA	33.7%	Patients with lupus erythematosus, Brazil	NA ⁴⁷	Aires, Argentina ⁴⁴	mostly with systemic lupus erythematosus or has been detecte by other than FEIA ^{45,46} . Also, since	
A. C. C. D.II.	0.1% (0.0-0.7)	Immune-mediated disease-free adults, Estonia	FEIA*	Will Cir	particular ANA are not specific to just one disease, the prevalence of ANAs in disease-free individuals and particular CTD in general population relate reasonably.	
Anti-SS-B/La	36%	Primary Sjögren 's syndrome, Italy	ELISA ⁴⁸	Mild Sjögren´s syndrome ⁴³		
	0.7% (0.3-1.5)	Immune-mediated disease-free adults, Estonia	FEIA*			
	1.9% (0-6.0)	General population, Denmark	IIF ⁴⁹			
Anti-SS-A/Ro	68%	Patients with primary Sjögren's syndrome, Italy	ELISA ⁴⁸	Vasculitis and nephritis of CTD, Sjögren´s syndrome ⁴³		
	SS-A/SS-B 16.5%	Disease-free individuals with HLA-DR3 or/and DR11 allele and autoantibodies to enteroviruses, Estonia	ELISA ¹⁸			
Anti-Sm	0.0%	Immune-mediated disease-free adults, Estonia	FEIA*	I		
Anti-Sm	0-45%	Patients with lupus erythematosus, Hong Kong, China	ELISA ⁵⁰	- Lupus erythematosus ⁵⁰		
A C LIADAD	0.1% (0.0-0.7)	Immune-mediated disease-free adults, Estonia	FEIA*	Mixed connective tissue disease and mild		
Anti-U1RNP	0-84%	Patients with lupus erythematosus, Hong Kong, China	ELISA ⁵⁰	Sjögren´s syndrome ⁴³		
Anti-CENP	0.3% (0.1–1.0)	Immune-mediated disease-free adults, Estonia	FEIA*	CREST (calcinosis, Raynaud's, oesophageal dysmotility, sclerodactyly, and telangiectasia) syndrome ⁴³		
Anti-Jo-1	0%	Immune-mediated disease-free adults, Estonia	FEIA*	Pulmonary fibrosis and poor prognosis of CTD ⁴³		
Anti-Scl-70	0%	Immune-mediated disease-free adults, Estonia	FEIA*	Pulmonary fibrosis and poor prognosis of CTD ⁴³		
	8.8% (7.1–10.7) ≥ 5 U/ml	Immune-mediated disease-free adults, Estonia	Ca-treated plasma ELISA*	Insulin dependent diabetes mellitus: Reported in	GADA in disease-free individual reflect the prevalence of autoimmune diabetes in general population. Calcium treatment of plasma has been suggested prior to detection GADA from	
	3.4% (2.4–4.8) ≥ 10 U/ml	Immune-mediated disease-free adults, Estonia	Ca-treated plasma ELISA*	0.4% (0.3-0.4) of general population, Estonia ¹⁷ Reported and estimated 1.1-9% in general population, Europe ⁵¹ Type 1 diabetes:		
GADA	0.4% (0.1−1.1) ≥ 50 U/ml	Immune-mediated disease-free adults, Estonia	Ca-treated plasma ELISA*	Estimated in 0.6-0.8% of general population, Europe ⁵¹ Latent autoimmune diabetes of adults:	ÉDTA-plasma ¹¹ , but may still be unsuitable for detecting GADA by ELISA. Alternatively, higher cut-off values may suit for GADA ELISA from plasma, leaving out GADA with low affinity and insignificance ⁵³ .	
	0.9%	General population, Southern Spain	NA ¹⁶	Estimated in 0.5–0.7% of general population, Europe ⁵²		
	2.0% (0.6-5.4)	Disease-free adults from rural town, Estonia	Serum RIA ¹⁷	Lutope		

Autoantibody						
Name	Prevalence % (95% CI)	Population	Method	Corresponding disease and its prevalence % (95% CI)	Comments	
	7.2% (5.7–9.1)	Immune-mediated disease-free individuals, Estonia	FEIA*			
3.3%		Disease-free children and adolescents, Leipzig, Germany	ECLIA ⁵⁶	Hypo- and hyperthyroidism: Reported in 2.2% (2.1–2.3) of general population, Estonia* and	Anti-TPO prevalence in disease- free individuals seems to be	
Anti-TPO IgG	10% (5-15)	General population, Denmark	Plasma RIA ⁴⁹	in 5% of women population, California, USA ⁵⁴ <i>Hypothyroidism:</i> Screened in 12% of complaint-	comparable to the estimated prevalence of thyroid diseases in general population but higher than reported cases. Thyroid diseases are	
	8.6%	General population, Australia	ELISA ¹⁹	free individuals, but reported in <2% of general		
10.7%(8.2-13.9)		Immune-mediated disease-free women, Estonia	FEIA*	population, Michigan, USA ⁵⁵	underdiagnosed.	
	17%	General population, Iran	Serum RIA ⁵⁷			
	0.6% (0.2-1.4)	Immune-mediated disease-free adults, Estonia	FEIA and ELISA*		Taken the prevalence of type 1 diabetes ⁵¹ , the calculated co-	
Co-existed anti-TPO and GADA	0.11-0.14%	Patients with type 1 diabetes and anti-TPO, Turkey	ELISA ⁵⁹	Thyrogastric autoimmune disease ⁵⁸ : 17.8% of type 1 diabetes patients had anti-TPO, Turkey ⁵⁹	existence of anti-TPO and GADA in Turkey ⁵⁹ is lower than detected among disease-free individuals in Estonia*. The prevalence of co- existence of these autoantibodies is likely population-specific.	

Table 4. Extended commented presentation of autoantibody prevalence of current study and from literature data. ANA – antinuclear autoantibodies, ACLIA – electroluminescence assay, ELISA – enzyme linked immunosorbent assay, CTD – connective tissue diseases, CCP - cyclic citrullinated peptide, CENP - centromere protein, dsDNA - double stranded deoxyribonucleic acid, FEIA - fluoro-enzyme immunoassay, GADA - autoantibodies against glutamic acid decarboxylase (molecular weight 65 kDa), IIF – indirect immunofluorescent test, Jo-1 - histidyl tRNA synthetize, NA – not available, RIA – radioimmunoassay, SS-A/Ro - Sjögren's syndrome type A antigen, SS-B/La - Sjögren's syndrome type B antigen, Scl-70 - scleroderma-associated autoantigen of 70 kDa, Sm - Smith protein, TPO – thyroid peroxidase, tTG - tissue transglutaminase, U1RNP - ribonucleoprotein U1. *Current article by Haller-Kikkatalo K., Alnek K., Metsküla K., Kisand K., Pisarev H., Salumets A., Uibo R.

and 2.31, p = 0.054, respectively, compared to the youngest group, 18–26 years of age), (ii) the odds were higher if a man consumed alcohol regularly (adOR = 3.33, p = 0.004 compared to non-consumers), and required cardiovascular treatment (adOR = 2.31, p = 0.011). The age adjusted and gender stratified model for women revealed that women were less likely positive for tissue non-specific autoantibodies in case of active ovarian hormones (adOR = 0.45, p = 0.036 for women who were either menstruating or were receiving HRT compared to women in menopause and not receiving HRT) (Fig. 1).

The most prevalent autoantibody GADA was detected in 8.8% of individuals of disease-free population, if the cut-off value for GADA positivity was selected >5 U/ml. However, a proportion of individuals were tested GADA greater than 10, 30 or even 50 U/ml (Table 6). Multivariate logistic regression analyses revealed that women with positive anti-CTD IgG test were associated with increased chance for GADA >10 U/ml and >30 U/ml (adORs 4.27, p=0.017 and 5.65, p=0.047, respectively) when the model was controlled by the ovarian hormonal activity. At the same time, active ovarian hormonal status decreased the likelihood of GADA >10 U/ml and >30 U/ml (adORs 0.34, p=0.027, and 0.17, p=0.038, respectively). The results were similar, if models were adjusted by the age of older and younger than 45 years instead of hormonal activity (data not shown). The age 45 was selected to unify data presentation between men and women. These results suggest that finding of either high level of GADA or anti-CTD IgG in disease-free women may reflect preclinical stage of immune-mediated disease, but the onset of clinical disease may be postponed by active ovarian hormonal status.

Age adjusted and gender stratified model for men revealed that GADA (>5 U/ml) coexisted more likely with anti-TPO IgG low level (60–100 U/ml) when statistical analyses was controlled by age over 45 years (adOR = 12.18, p = 0.015). Similarly, GADA >30 U/ml coexisted more likely with anti-TPO IgG high level (>100 U/ml, adOR = 22.11, p = 0.003), when controlled by age. The presence of GADA >10 U/ml showed association with age over 45 years in men (adOR = 4.14, p = 0.010 when adjusted for the presence of anti-CTD IgG, and adOR = 3.82, p = 0.015 when adjusted for anti-TPO IgG). These results suggest that finding of anti-TPO IgG or GADA, regardless of level may reflect activation of adverse immune reaction which aggravates with age in men.

Discussion

The data here represent a cross-sectional study of the prevalence of (auto)immune-mediated diseases in the general population of Estonia and the prevalence and association of 6 clinically significant autoantibodies in 994 (auto)immune-disease-free individuals from the general population. Most importantly, the prevalence of anti-TPO IgG and 5 tissue non-specific autoantibodies, anti-tTG IgG, and IgA, anti-CCP IgG, anti-CTD IgG, and GADA, in disease-free individuals was 23.6%. This was comparable to the prevalence of (auto)immune-mediated diseases registered in the general population of Estonia. In addition, several phenotypic parameters were associated with the presence of autoantibodies. These associations were gender specific and distinct for anti-TPO and tissue non-specific autoantibodies.

The surveying of a population-based registry of 51,862 individuals revealed that a third of the Estonian adult population had been diagnosed with some type of (auto)immune-mediated disease. The list of (auto) immune-mediated diseases was selected to include most diseases' locations in the body and covered 90

	Association with phenotypic characteristics					
		on-specific tibody ⁶	Anti-TPO			
Phenotypic characteristics ¹	Men	Women	Men	Women		
Age at time of study (years)	OR = 1.03*	OR = 1.01	OR = 1.02	OR = 1.03*		
Age groups ²						
18-26 years	OR=1	OR = 1	OR=1	OR=1		
27-37 years	OR = 2.08#	OR = 0.86	OR = 0.67	OR = 1.76		
38-52 years	OR = 2.47*	OR = 1.21	OR = 1.69	OR = 1.84		
>52 years	OR = 3.39*	OR = 1.40	OR = 1.58	OR = 3.88*		
Occupation		'				
Employed	adOR=1	adOR=1	adOR=1	adOR=1		
Pension insurance ³	adOR = 2.48	adOR = 0.82	adOR = 0.49	adOR = 2.19		
Student or serviceman	adOR = 0.98	adOR = 1.56	adOR = 3.72#	adOR = 1.08		
Unemployed ⁴	adOR = 1.64	adOR = 1.10	adOR = 0.46	adOR = 0.99		
Frequency of alcohol consumption						
No consumption	adOR=1	adOR=1	adOR=1	adOR=1		
Rare – up to few times per year	adOR = 1.74	adOR = 1.31	adOR = 0.62	adOR = 0.81		
Seldom – up to every month	adOR=2.92*	adOR = 1.44	adOR = 2.05	adOR = 1.75		
Moderate or frequent – up to every 2 nd day	adOR = 1.45	adOR = 0.93	adOR = 1.10	adOR = 0.69		
Active ovarian hormones ⁵	_	adOR = 0.50*	_	adOR = 0.55		
Recently used cardiovascular drugs	adOR=1.85#	adOR = 1.19	adOR = 1.12	adOR = 0.85		
Maternal autoimmune disease	adOR = 0.38	adOR = 1.11	adOR = 5.50*	adOR = 1.17		

Table 5. Associations between autoantibodies and phenotypic characteristics. ¹Characteristics with any association with either anti-TPO or tissue non-specific autoantibodies are presented. ²Age groups were formed according to the values of the 1st and 3rd quantiles and median of the age and compared with the youngest group (18-26 years). ³Pension insurance includes retired persons and individuals who are unemployed but covered by health insurance. ⁴Unemployed also includes women on childcare leave. ⁵Women with active ovarian hormones includes women who menstruate and women with primary or secondary amenorrhea who are currently receiving hormone replacement therapy. ⁶The presence of at least one of five tissue non-specific autoantibodies (antinuclear autoantibodies detected with a connective tissue disease screening test and autoantibodies against glutamic acid decarboxylase (molecular weight of 65 kDa), thyroid peroxidase, tissue transglutaminase IgG and IgA, or cyclic citrullinated peptide). adOR – adjusted odds ratio. *Statistically significant association (p < 0.05), # statistical tendency towards association (0.05 < p < 0.1). Associations between phenotypic characteristics and autoantibody prevalence were analysed with logistic regression analyses stratified by gender and adjusted for age.

ICD-10 categories (Supplementary Table S.1). Overall, men were diagnosed less frequently than women; 26.4% of men and 32.1% of women had at least one diagnosis (Table 1). Data indicating the prevalence of so many immune-mediated diseases in a population is rarely available in the scientific literature, which makes this study unique. According to one previous study, subjects with chronic diseases account for one-third of the general population in Germany², although the list of studied diseases was different from that in this study.

The prevalence of disease was high, as was the prevalence of at least 1 of 6 autoantibodies in the population of disease-free individuals, especially among women (diseases occurred in 32.1% (31.6–32.6) of women in the general population and autoantibodies were present in 29.4% (25.5–33.6) of diseases-free women (proportion test, p > 0.05). Autoantibodies measured here in healthy individuals are presented with previously published data in Table 4. The prevalence of anti-tTG IgA, anti-CCP IgG, anti-CTD IgG and anti-TPO IgG in our study is comparable to the autoantibodies in general population of other countries ^{12–15}. The prevalence of GADA ^{16,17} or autoantibodies from CTD IgG panel ^{17–20} is not comparable due to differences either in laboratory tests or study populations.

Importantly, the autoantibodies detected in this study, anti-tTG IgG and IgA, anti-CTD IgG, GADA, and anti-TPO IgG, have diagnostic value for common autoimmune diseases, and at least some of these may play roles in disease pathogenesis. Although autoantibodies were assessed in healthy individuals, the positive cut-off value for each autoantibody test was chosen to match the level used to diagnose the corresponding autoimmune disease. Logically, it could be hypothesized that the group of seropositive but healthy individuals will develop autoimmune diseases, and the presence of autoantibodies is prognostic marker that precedes clinical manifestation. On the other hand, some of autoantibody-positive, disease-free individuals may never develop clinical disease, and the presence of autoantibodies may have no clinical significance for them. Although it is not known which of the above scenarios occurs, the current study provides important information. Long-term follow-up would be the best way to resolve this question.

In men, the presence of anti-TPO was strongly associated with familial anamnesis of maternal autoimmune disease. The same association was not revealed in women, although maternal autoimmune disease was reported more often in women than men (16.7% vs. 11.4%). Maternal thyroid disease, in particular, accounted for a

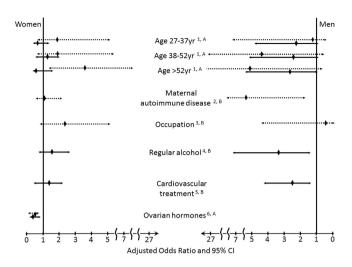


Figure 1. Risk factors for autoantibodies detected in men and women. Autoantibodies against thyroid peroxidase (anti-TPO, dashed line) and any tissue non-specific autoantibody (solid line) groups are shown separately. The tissue non-specific autoantibody group comprised individuals who tested positive for at least one of the 5 tested tissue non-specific autoantibodies. ¹age groups were formed according to the values of 1st and 3rd quantiles and the median age compared to the youngest group (18-26 years); ²the presence of maternal autoimmune disease compared to individuals without maternal autoimmune disease; 3 individuals with an occupational status of unemployed but being covered by pension insurance (includes retired individuals and unemployed individuals with insurance for disability) compared to employed persons; ⁴seldom but regular alcohol consumption (once a month) compared to non-consumers; 5the need for cardiovascular treatment on a regular basis at least 2 months prior to the study; 6women with active ovarian hormones includes women who menstruate and women with primary or secondary amenorrhea who receive hormone replacement therapy compared to women in menopause. Association values are given as odds ratios considered for multiple comparisons from multivariate logistic regression models stratified by gender and adjusted for the following confounders selected from Table 5: A age groups; B age groups (or ovarian hormonal activity in women), maternal autoimmune disease, occupational status, alcohol consumption, cardiovascular treatment. CI – confidence interval.

negligible proportion of maternal autoimmune diseases in our cohort (reported in 4 cases, 1 woman and 3 men). Therefore, it seems likely that maternal autoimmunity, not specifically thyroid disease, leads to anti-thyroid autoimmunity in sons. This mother-to-son inheritance relies on several phenomena. First, the X-chromosome contains immunologically important genes, associated with a variety of autoimmune diseases²¹. Therefore, because male offspring inherit maternal X-chromosome, autoimmune susceptibility may be inherited if the mother is affected. Second, an increased frequency of skewed X-chromosome inactivation (XCI) has been found in many autoimmune diseases²² and may result from chance or genetic factors²³. These sons may inherit the defective X-chromosome, which cannot be balanced because of the lack of spare X-chromosome in males, in contrast to the case in daughters. In this case, the mechanism of XCI to favour autoimmunity has been attributed to the potential escape of X-linked self-antigens in the thymus or other peripheral sites that are involved in tolerance²⁴. However, these two explanations do not explain why the thyroid gland is preferentially targeted in sons of mothers suffering from autoimmune diseases. The third possible explanation involves maternal microchimerism, which has been $detected \ in \ the \ newborn \ thyroid \ gland^{25}. \ Regardless \ of \ the \ reason \ for \ anti-TPO \ antibodies \ in \ male \ participants,$ anti-TPO IgG can activate complement and cause damage to thyroid cells via antibody dependent cell cytotoxicity¹³ eventually leading to thyroid disease. Accordingly, men with anti-thyroid autoantibodies have a 5 times greater risk of progressing to overt thyroid gland disease than women⁶. Indeed, women may be protected from autoimmune diseases until menopause, because active ovarian hormones postpone the production of anti-TPO and tissue non-specific autoantibodies in women²⁶.

Multivariate association analyses detected several phenotypic associations, suggesting the presence of at least one tissue non-specific autoantibody associated with moderate but regular alcohol intake and the need for cardiovascular treatment. These associations were independent of age and they were present in men but not women. The impacts of alcohol consumption on health are complex and modulated by several factors such as pattern and amount of drinking, genetics, the organ system studied, and the sex and age of the user²⁷. Regular heavy consumption (\geq 3 drinks a day in men) causes suppression of innate immunity²⁸, whereas moderate consumption (\leq 2 drinks a day for men)²⁹ may enhance the effects of vaccines²⁷ and increase intestinal permeability³⁰, which can lead to immune recognition of self-antigens³¹. Here, the effects of alcohol were not considered beneficial, since autoantibodies can mark future autoimmunity. Similarly, most of the benefits of moderate alcohol consumption have been described in the context of cardiovascular disease³². Here, we revealed that cardiovascular diseases were associated with increased prevalence of tissue non-specific autoantibodies, regardless of whether the person was consuming alcohol or not, which has been suggested previously³³. Surprisingly, these associations were revealed in men and were not significant in women. Since women were tested for risk factors by statistical

	GADA positive individuals (%, 95% confidential interval)		
GADA cut-off level	Men (N=491)	Women (N = 503)	
>5 U/ml	39/491 (7.9%, 5.8–10.8)	48/503 (9.5%, 7.2–12.5)	
<45 years	19/39 (48.7%, 32.7–65.0)	23/48 (47.9%, 33.5–62.6)	
>45 years	20/39 (51.3, 35.0-67.3)	25/48 (52.1%, 37.4–66.5)	
Active ovarian hormones		21/48 (43.8%, 29.8–58.7)	
Non-active ovarian hormones		26/48 (54.2%, 39.3–68.4)	
>10 U/ml	16/491 (3.3%, 1.9–5.4)	18/503 (3.6%, 2.2-5.7)	
<45 years	5/16 (31.3%, 12.1–58.5)*	8/18 (44.4%, 22.4–68.7)	
>45 years	11/16 (68.8%, 41.5-87.9)*	10/18 (55.6%, 31.3-77.6)	
Active ovarian hormones		8/18 (44.4%, 22.4–68.7)	
Non-active ovarian hormones		10/18 (55.6%, 31.3–77.6)	
>30 U/ml	4/491 (0.8%, 0.3–2.2)	7/503 (1.4%, 0.6–3.0)	
<45 years	1/4 (25.0%, 1.3–78.1)	3/7 (42.9%, 11.8–79.8)	
>45 years	3/4 (75.0%, 21.9–98.7)	4/7 (57.1%, 20.2–88.2)	
Active ovarian hormones		2/7 (28.6%, 5.1–69.7)*	
Non-active ovarian hormones		5/7 (71.4%, 30.3–94.9)*	
>50 U/ml	1/491 (0.2%, 0-1.3)	3/503 (0.6%, 0.2–1.9)	
<45 years	0/1 (0%, 0-94.5)*	1/3 (33.3%, 1.8–87.5)	
>45 years	1/1 (100%, 5.4–100)*	2/3 (66.7%, 12.5–98.2)	
Active ovarian hormones		1/3 (33.3%, 1.8–87.5)	
Non-active ovarian hormones		2/3 (66.7%, 12.5–98.2)	

Table 6. GADA autoantibodies of different level. GADA prevalence are provided among study population and not extended to entire Estonian population (compare Table 3 and 4). *proportion test p < 0.05 between rows.

models adjusted for active ovarian hormones, other phenotypic parameters seem less important than ovarian hormonal profiles in women.

Childbearing age seemed to be the major protective factor from autoimmune disease in women. Indeed, women with active ovarian hormones (either because they were in their fertile years or receiving HRT)¹² were less likely to be seropositive for anti-TPO and tissue non-specific autoantibodies regardless of confounders. Age related protection against autoimmunity can be explained by age related acquired X-chromosome loss or monosomy (XCM)¹⁴. Because the X-chromosome has immunologically important genes, XCM contributes to the speed of development and the number of overlapping autoimmune diseases¹⁵. In addition, the X-chromosome contains genes that appear to be crucial in the maintenance of physiological sex hormone levels¹⁵. Commonly, a substantial decline of sex hormone levels in women occurs at menopause. However, female sex hormones have various immunomodulatory effects²⁶. These factors may explain why women were found to be likely autoantibody-positive in this study.

In conclusion, approximately one-third of the adult population of Estonia, individuals who lacked (auto) immune-mediated diseases, tested positive for at least one of six clinically significant autoantibodies. At the same time, the documented prevalence of the corresponding (auto)immune-mediated diseases in the general population was also one-third. However, some of these diseases, including thyroid disease and coeliac disease, were diagnosed less frequently than estimated. The results suggest that the presence of tissue non-specific autoantibodies may serve as prognostic markers for future diseases in currently disease-free individuals. The presence of autoantibodies in men depends on general health, age or health manners. In contrast, the protection of active ovarian hormones in women decrease any putative risk of health parameters studied here. Different from anti-TPO IgG in women, incidental finding of anti-TPO IgG in men with positive familial anamnesis of maternal autoimmune disease deserves further medical intention. The wider implications of these findings suggest that physicians should be encouraged to look for autoimmune markers in addition to treating a variety of patient health complaints.

References

- 1. Leslie, D., Lipsky, P. & Notkins, A. L. Autoantibodies as predictors of disease. J Clin Invest 108, 1417–1422, doi: 10.1172/JCI14452 (2001).
- Bussing, A., Ostermann, T. & Koenig, H. G. Relevance of religion and spirituality in German patients with chronic diseases. Int J Psychiatry Med 37, 39–57 (2007).
- 3. Eschler, D. C., Hasham, A. & Tomer, Y. Cutting edge: the etiology of autoimmune thyroid diseases. Clin Rev Allergy Immunol 41, 190–197, doi: 10.1007/s12016-010-8245-8 (2011).
- 4. Husby, S. et al. European Society for Pediatric Gastroenterology, Hepatology, and Nutrition guidelines for the diagnosis of coeliac disease. J Pediatr Gastroenterol Nutr 54, 136–160, doi: 10.1097/MPG.0b013e31821a23d0 (2012).
- 5. Klock, C., Diraimondo, T. R. & Khosla, C. Role of transglutaminase 2 in celiac disease pathogenesis. Semin Immunopathol 34, 513–522, doi: 10.1007/s00281-012-0305-0 (2012).
- 6. Vanderpump, M. P. et al. The incidence of thyroid disorders in the community: a twenty-year follow-up of the Whickham Survey. Clin Endocrinol (Oxf) 43, 55–68 (1995).
- Pereira, D. A., Kawassaki Ade, M. & Baldi, B. G. Interpretation of autoantibody positivity in interstitial lung disease and lungdominant connective tissue disease. J Bras Pneumol 39, 728–741, doi: 10.1590/S1806-37132013000600012 (2013).

- Cohen, I. R. Autoantibody repertoires, natural biomarkers, and system controllers. Trends Immunol 34, 620–625, doi: 10.1016/j. it.2013.05.003 (2013).
- 9. Leitsalu, L. et al. Cohort Profile: Estonian Biobank of the Estonian Genome Center, University of Tartu. Int J Epidemiol, doi: 10.1093/ije/dyt268 (2014).
- 10. Chang, C. & Gershwin, M. E. Drugs and autoimmunity-a contemporary review and mechanistic approach. *J Autoimmun* 34, J266–275, doi: 10.1016/j.jaut.2009.11.012 (2010).
- 11. Nilson, E., Ekholm, B., Rees Smith, B., Torn, C. & Hillman, M. Calcium addition to EDTA plasma eliminates falsely positive results in the RSR GADAb ELISA. *Clin Chim Acta* 388, 130–134, doi: 10.1016/j.cca.2007.10.021 (2008).
- 12. Bulow Pedersen, I. et al. Lack of association between thyroid autoantibodies and parity in a population study argues against microchimerism as a trigger of thyroid autoimmunity. Eur J Endocrinol 154, 39–45, doi: 10.1530/eje.1.02070 (2006).
- 13. McLachlan, S. M. & Rapoport, B. Why measure thyroglobulin autoantibodies rather than thyroid peroxidase autoantibodies? *Thyroid* 14, 510–520, doi: 10.1089/1050725041517057 (2004).
- 14. Russell, L. M., Strike, P., Browne, C. E. & Jacobs, P. A. X chromosome loss and ageing. Cytogenet Genome Res 116, 181–185, doi: 10.1159/000098184 (2007).
- 15. Svyryd, Y. et al. X chromosome monosomy in primary and overlapping autoimmune diseases. Autoimmun Rev 11, 301–304, doi: 10.1016/j.autrev.2010.03.001 (2012).
- 16. Soriguer-Escofet, F. et al. Prevalence of latent autoimmune diabetes of adults (LADA) in Southern Spain. Diabetes Res Clin Pract 56, 213–220 (2002).
- 17. Uibo, R. et al. Comparison of the prevalence of glutamic acid decarboxylase (GAD65) and gliadin antibodies (AGA) in a randomly selected adult estonian population. Horm Metab Res 33, 564–567, doi: 10.1055/s-2001-17215 (2001).
- 18. Metskula, K., Salur, L., Mandel, M. & Uibo, R. Demonstration of high prevalence of SS-A antibodies in a general population: association with HLA-DR and enterovirus antibodies. *Immunol Lett* 106, 14–18, doi: 10.1016/j.imlet.2006.03.005 (2006).
- Napthali, K. et al. Thyroid antibodies, autoimmunity and cognitive decline: is there a population-based link? Dement Geriatr Cogn Dis Extra 4, 140–146, doi: 10.1159/000362716 (2014).
- Op De Beeck, K. et al. Detection of antinuclear antibodies by indirect immunofluorescence and by solid phase assay. Autoimmun Rev 10, 801–808, doi: 10.1016/j.autrev.2011.06.005 (2011).
- Chang, D. et al. Accounting for eXentricities: analysis of the X chromosome in GWAS reveals X-linked genes implicated in autoimmune diseases. PLoS One 9, e113684, doi: 10.1371/journal.pone.0113684 (2014).
- Knudsen, G. P. Gender bias in autoimmune diseases: X chromosome inactivation in women with multiple sclerosis. J Neurol Sci 286, 43–46, doi: 10.1016/j.jns.2009.04.022 (2009).
- Belmont, J. W. Genetic control of X inactivation and processes leading to X-inactivation skewing. Am J Hum Genet 58, 1101–1108 (1996).
- 24. Kyewski, B. & Derbinski, J. Self-representation in the thymus: an extended view. *Nat Rev Immunol* 4, 688–698, doi: 10.1038/nri1436 (2004).
- Srivatsa, B., Srivatsa, S., Johnson, K. L. & Bianchi, D. W. Maternal cell microchimerism in newborn tissues. J Pediatr 142, 31–35, doi: 10.1067/mpd.2003.mpd0327 (2003).
- Tan, I. J., Peeva, E. & Zandman-Goddard, G. Hormonal modulation of the immune system A spotlight on the role of progestogens. *Autoimmun Rev*, doi: 10.1016/j.autrev.2015.02.004 (2015).
- 27. Messaoudi, I., Pasala, S. & Grant, K. Could moderate alcohol intake be recommended to improve vaccine responses ?Expert Rev Vaccines 13, 817–819, doi: 10.1586/14760584.2014.924405 (2014).
- *vaccines* 13, 817–819, aoi: 10.1586/14/60584.2014.924405 (2014).

 28. Curtis, B. J., Zahs, A. & Kovacs, E. J. Epigenetic targets for reversing immune defects caused by alcohol exposure. *Alcohol Res* 35,
- 97–113 (2013).

 29. Nova, E., Baccan, G. C., Veses, A., Zapatera, B. & Marcos, A. Potential health benefits of moderate alcohol consumption: current perspectives in research. *Proc Nutr Soc* **71**, 307–315, doi: 10.1017/S0029665112000171 (2012).
- 30. Bhonchal, S. *et al.* Functional and morphological alterations in small intestine mucosa of chronic alcoholics. *J Gastroenterol Hepatol* 23, e43–48, doi: 10.1111/j.1440-1746.2007.05080.x (2008).
- 31. Lerner, A. & Matthias, T. Changes in intestinal tight junction permeability associated with industrial food additives explain the rising incidence of autoimmune disease. *Autoimmun Rev*, doi: 10.1016/j.autrev.2015.01.009 (2015).
- 32. Howie, E. K. et al. Alcohol consumption and risk of all-cause and cardiovascular disease mortality in men. J Aging Res 2011, 805062, doi: 10.4061/2011/805062 (2011).
- 33. Majka, D. S. & Chang, R. W. Is preclinical autoimmunity benign? The case of cardiovascular disease. *Rheum Dis Clin North Am* 40, 659–668, doi: 10.1016/j.rdc.2014.07.006 (2014).
- 34. Ress, K. *et al.* High prevalence of coeliac disease: need for increasing awareness among physicians. *Dig Liver Dis* **39**, 136–139, doi: 10.1016/j.dld.2006.07.012 (2007).
- 35. Evans, K. E. & Sanders, D. S. Celiac disease. Gastroenterol Clin North Am 41, 639-650, doi: 10.1016/j.gtc.2012.06.004 (2012).
- 36. Horwitz, A. et al. Screening for celiac disease in Danish adults. Scand J Gastroenterol, 1–7, doi: 10.3109/00365521.2015.1010571 (2015).
- 37. Di Pisa, M. et al. Epitope mapping of the N-terminal portion of tissue transglutaminase protein antigen to identify linear epitopes in celiac disease. J Pept Sci 20, 689–695, doi: 10.1002/psc.2650 (2014).
- Lammi, A. et al. Antibodies to Deamidated Gliadin Peptide in Diagnosis of Celiac Disease in Children. J Pediatr Gastroenterol Nutr, doi: 10.1097/MPG.0000000000000666 (2014).
- 39. Katz, K. D. et al. Screening for celiac disease in a North American population: sequential serology and gastrointestinal symptoms. Am J Gastroenterol 106, 1333–1339, doi: 10.1038/ajg.2011.21 (2011).
- 40. Saketkoo, L. A., Magnus, J. H. & Doyle, M. K. The primary care physician in the early diagnosis of systemic sclerosis: the cornerstone of recognition and hope. *Am J Med Sci* 347, 54–63, doi: 10.1097/MAJ.0b013e3182a55d24 (2014).
- Jaskowski, T. D. et al. Relationship between rheumatoid factor isotypes and IgG anti-cyclic citrullinated peptide antibodies. J Rheumatol 37, 1582–1588, doi: 10.3899/jrheum.091236 (2010).
- 42. Polimeni, M., Feniman, D., Skare, T. S. & Nisihara, R. M. Anti-cyclic citrullinated peptide antibodies in scleroderma patients. Clin Rheumatol 31, 877–880, doi: 10.1007/s10067-011-1930-z (2012).
- Hang, L. M. & Nakamura, R. M. Current concepts and advances in clinical laboratory testing for autoimmune diseases. Crit Rev Clin Lab Sci 34, 275–311, doi: 10.3109/10408369708998095 (1997).
- 44. Scolnik, M. et al. Incidence and prevalence of lupus in Buenos Aires, Argentina: a 11-year health management organisation-based study. Lupus Sci Med 1, e000021, doi: 10.1136/lupus-2014-000021 (2014).
- 45. Freire, P. V. et al. Distinct antibody profile: a clue to primary antiphospholipid syndrome evolving into systemic lupus erythematosus? Clin Rheumatol 33, 349–353, doi: 10.1007/s10067-013-2472-3 (2014).
- 46. Hanly, J. G., Su, L., Farewell, V. & Fritzler, M. J. Comparison between multiplex assays for autoantibody detection in systemic lupus erythematosus. *J Immunol Methods* **358**, 75–80, doi: 10.1016/j.jim.2010.04.005 (2010).
- 47. Skare, T., Damin, R. & Hofius, R. Prevalence of the American College of Rheumatology hematological classification criteria and associations with serological and clinical variables in 460 systemic lupus erythematosus patients. *Rev Bras Hematol Hemoter* 37, 115–119, doi: 10.1016/j.bjhh.2015.01.006 (2015).

- 48. Bartoloni, E. et al. Cardiovascular disease risk burden in primary Sjogren's syndrome: results of a population-based multicentre cohort study. J Intern Med, doi: 10.1111/joim.12346 (2015).
- 49. Andersen-Ranberg, K., M, H. O.-M., Wiik, A., Jeune, B.& Hegedus, L.High prevalence of autoantibodies among Danish centenarians. Clin Exp Immunol 138, 158–163, doi: 10.1111/j.1365-2249.2004.02575.x (2004).
- 50. Li, P. H. et al. Relationship between autoantibody clustering and clinical subsets in SLE: cluster and association analyses in Hong Kong Chinese. Rheumatology (Oxford) 52, 337–345, doi: 10.1093/rheumatology/kes261 (2013).
- 51. Kim, C., Newton, K. M. & Knopp, R. H. Gestational diabetes and the incidence of type 2 diabetes: a systematic review. *Diabetes Care* **25**, 1862–1868 (2002).
- 52. Tuomi, T. *et al.* Antibodies to glutamic acid decarboxylase reveal latent autoimmune diabetes mellitus in adults with a non-insulindependent onset of disease. *Diabetes* **42**, 359–362 (1993).
- 53. Krause, S. et al. GAD autoantibody affinity in adult patients with latent autoimmune diabetes, the study participants of a GAD65 vaccination trial. *Diabetes Care* 37, 1675–1680, doi: 10.2337/dc13-1719 (2014).
- Wang, C. & Crapo, L. M. The epidemiology of thyroid disease and implications for screening. Endocrinol Metab Clin North Am 26, 189–218 (1997).
- 55. Canaris, G. J., Tape, T. G. & Wigton, R. S. Thyroid disease awareness is associated with high rates of identifying subjects with previously undiagnosed thyroid dysfunction. *BMC Public Health* 13, 351, doi: 10.1186/1471-2458-13-351 (2013).
- 56. Taubner, K. et al. Serum concentrations of anti-thyroid peroxidase and anti-thyroglobulin antibodies in children and adolescents without apparent thyroid disorders. Clin Biochem 47, 3–7, doi: 10.1016/j.clinbiochem.2013.09.017 (2014).
- 57. Karimi, F., Kalantarhormozi, M. R., Dabbaghmanesh, M. H. & Ranjbar Omrani, G. Thyroid disorders and the prevalence of antithyroid antibodies in Shiraz population. *Arch Iran Med* 17, 347–351, doi: 0141705/AIM.008 (2014).
- 58. Abrams, P., De Leeuw, I. & Vertommen, J. In new-onset insulin-dependent diabetic patients the presence of anti-thyroid peroxidase antibodies is associated with islet cell autoimmunity and the high risk haplotype HLA DQA1*0301-DQB1*0302. Belgian Diabetes Registry. Diabet Med 13, 415–419, doi: 10.1002/(SICI)1096-9136(199605)13:5<415::AID-DIA96>3.0.CO;2-X (1996).
- 59. Erten, G., Gurol, A. O., Deniz, G., Satman, I. & Yilmaz, M. T. Organ specific autoantibodies in preclinical and early clinical type 1 diabetes in Turkey. *Ups J Med Sci* 112, 231–243 (2007).

Author Contributions

A.S., R.U., K.K., E.M. and A.M. contributed to the design of the study, K.A. and K.M. carried out the immunoassays, and K.H.K. and H.P. performed the statistical analysis. K.H.K. and K.A. conceived the study and participated in its coordination. K.H.K. drafted the manuscript. K.H.K., A.S. and R.U. helped to draft the manuscript. All authors read and approved the final manuscript.

Additional Information

Supplementary information accompanies this paper at http://www.nature.com/srep

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