

Copyright is reserved and owned by the author of this dissertation. A copy of the document can be downloaded by an individual only for the purpose of research and private study. None of this document, either in its entirety or in part, may be reproduced elsewhere unless permitted by the author.

**FUNCTIONAL CHARACTERISATION of
*CONSTITUTIVE EXPRESSER of PATHOGENESIS-
RELATED GENES 5***

A thesis presented in partial fulfilment of the requirements for the degree of

Doctor of Philosophy

in

Molecular Biology

at Massey University, Palmerston North, New Zealand.



MASSEY UNIVERSITY
TE KUNENGA KI PŪREHUROA

UNIVERSITY OF NEW ZEALAND

Muhammad Faisal

2017

iii

ABSTRACT

As reported previously, CPR5 negatively regulates the onset of leaf death, hypersensitive response, disease resistance and early leaf senescence. *cpr5* plants contain aberrant trichomes and higher levels of ROS, SA and JA. Cell-cycle, JA/ET, ABA and sugar signalling are also affected in *cpr5* plants. These results suggest that CPR5 is a master regulator of multiple processes. However, how CPR5 manages to exert pleiotropic effects is still poorly understood. The first objective of the current study was the purification of the CPR5 protein to solve its crystal structure. Extensive *in silico* analyses were carried out and the results showed that CPR5 is predicted to be a membrane protein with 4 or 5 transmembrane (TM) domains. Additionally, CPR5 contains intrinsically disordered regions (IDRs) at its N-terminus. Proteins containing IDRs and TM domains are often difficult to purify for crystallization studies. Therefore, the undesirable regions of CPR5 such as, IDR and TM domains were deleted and a set of 24 constructs were developed. Despite several efforts, none of the CPR5 recombinant proteins were isolated. In addition to predicting IDR and TM domains, *in silico* results also predicted three NLS-encoding clusters, casein kinase phosphorylation sites, multiple start codons, coiled-coil domains and glycine motifs. To find out the roles of these putative structural elements on CPR5 functions, firstly a *CPR5* cDNA was synthesised and termed as *SynCPR5*. Subsequently, predicted sites or motifs were mutated in *SynCPR5* through site-directed mutagenesis and a set of 25 mutated *CPR5* transgenes (cDNA constructs) were developed. Using a complementation strategy, all the constructs were transformed into *cpr5-2* plants. The results show that the complementation of *cpr5-2* plants with *SynCPR5*, fully restored HR-like lesions, wildtype-like trichomes and leaves on *SynCPR5* plants. Further physiological characterization such as, transcript abundance of *SynCPR5*, *PR1*, *PR5* and *PDF1.2*, leaf area measurements and ploidy levels showed that *CPR5* regulates some of its functions and phenotypes quantitatively as well as qualitatively. When compared with the wildtype, better growth (larger leaves) but enhanced disease susceptibility was found in *metCPR5* transgenic lines (in which putative start codons were mutated), indicating that CPR5 regulates a balance between growth and resistance. Functional characterization of NLS mutants (*nlsCPR5*) showed that NLS-encoding clusters are important for CPR5 proper

functions. However, current evidence is insufficient to relate their role in CPR5 localization. Moreover, *in silico* results show that putative NLS clusters are present in the region of CPR5 which were annotated as intrinsically disordered region (IDR). Similar phenotypes shown by both *nlsCPR5* and *Del63CPR5* (in which the first 63 amino acids of CPR5 including putative NLS were deleted), indicate that the putative NLS clusters could be part of IDR and may have dual functions. Loss-of-function phenotypes shown by coiled-coil domain mutants (*ccdCPR5*) reinforce the role of coiled-coil domains in CPR5 homo-dimerization. Moreover, in contrast to previous reports, the downregulation of *PDF1.2* in the majority of *CPR5* complementation lines proposes CPR5 to be a positive regulator of *PDF1.2*. Based on the results presented in the current study, putative CPR5 IDRs and coiled-coil domains are proposed to facilitate CPR5 dimerization in order to restrict the entry of deregulated cargos into the nucleus. Moreover, these results uncover a novel role of CPR5 in the regulation of balance between plant growth and resistance. Furthermore, this study, for the first time, reports evidence of the requirement of NLS clusters for CPR5 functions.

ACKNOWLEDGEMENTS

All glory and praises for **ALLAH**, The most Beneficent, The most Merciful, Who is the only source of entire and complete knowledge and wisdom endowed to mankind. It is a matter of great honour and pleasure for me to express my ineffable gratitude and profound indebtedness to the Higher Education Commission (HEC), Pakistan and Massey University, New Zealand for providing me the best educational and research facilities. I offer my humblest gratitude to my revered supervisor Dr **PAUL DIJKWEL** and mentors Drs **GILLIAN NORRIS** and **JASNA RAKONJAC** for their skilful guidance, enlightened views, valuable suggestions, constructive criticism, unfailing patience and inspiring attitude during my studies and research.

I feel proud to have nice and helpful colleagues and friends in C5.19 and X-lab and I am grateful for their contributions, moral support and valuable suggestions. I also feel highly indebted to **VICTORIA SIBLEY** for her time and efforts spent in proofreading my dissertation. I would like to extend my gratitude and the deepest appreciation to my mother for her ever encouraging and supporting role in achieving my goals of life. Words are lacking to express my obligations to my affectionate mother, Ms **NASEEM AKHTAR**, for her love, best wishes, inspirations and unceasing prayers, without which the present destination would have been a mere dream. I am highly indebted to my brother, **MUHAMMAD ASLAM MUNNA** who exhibited great patience and good will throughout this study, and looked after the whole family.

I deem it my utmost pleasure to avail myself of opportunity to express the heartiest gratitude to my wife, Dr **TINA SEHRISH** for her love, patience, encouragement, good wishes and understanding that inspired me to accomplish this humble effort. I also owe a lot of love, care and time to my children, **AYESHA** and **MUHAMMAD** who could not get the proper time, love and attention during the course of my research and studies. I have no words to express as thanks to my family except to say them aloud, I love you all and I am lucky to have you all. Thank God for blessing me with a great, loving family.

MUHAMMAD FAISAL

TABLE OF CONTENTS

ABSTRACT	I
ACKNOWLEDGEMENTS	III
LIST OF FIGURES	XI
LIST OF TABLES	XIV
LIST OF ABBREVIATIONS	XV
Chapter 1 CPR5 as a master regulator- review of literature.....	1
1.1 INTRODUCTION.....	1
1.1.1 CONSTITUTIVE EXPRESSER OF PATHOGENESIS-RELATED GENES (CPR) mutants..	2
1.1.2 CONSTITUTIVE EXPRESSER of PATHOGENESIS-RELATED GENES 5 (CPR5) and other CPR5 mutant alleles	2
1.1.3 CPR5 Localizes in nuclear membrane.....	4
1.1.4 <i>CPR5</i> is constitutively expressed in the majority of plant tissues	4
1.2 Role of CPR5 in programmed cell death, hypersensitive response and pathogen resistance	5
1.3 Leaf senescence regulation and CPR5.....	10
1.4 Cell cycle, cell growth and development, and CPR5.....	14
1.5 Potassium homeostasis in <i>cpr5</i>	17
1.6 Germination and seedling growth response in <i>cpr5</i>	18
1.7 Plant growth and resistance trade-off and <i>CPR5</i>	19
1.8 Outlook and Aims of the Research.....	25
Chapter 2 Materials and Methods	27
2.1 <i>In silico</i> computer tools used for CPR5 analyses.....	27
2.2 Primer designing, RNA extractions and cDNA synthesis	28
2.2.1 Primer designing.....	28
2.2.2 RNA Extractions and cDNA synthesis	29
2.2.3 qRT-PCR data analyses.....	30
2.2.4 PCR reactions and profiles	30

2.3 Cloning strategy and development of <i>CPR5</i> constructs for structural studies.....	31
2.4 Amplification, transformation and confirmation of 8 <i>CPR5</i> transgenes	32
2.5 Cloning techniques and protocols	33
2.5.1 DNA restriction/digestions.....	33
2.5.2 DNA elution/extractions from agarose gels	33
2.5.3 DNA ligation	37
2.6 Bacterial transformation protocol.....	38
2.6.1 Preparation of LB liquid and agar media	38
2.6.2 LB agar plates for IPTG/X-Gal for white/blue Selection	39
2.7 Bacterial plasmid (DNA) isolation protocol (Alkaline lysis)	39
2.8 Competent cells preparations for <i>E. coli</i> and <i>Agrobacterium</i> -mediated transformations.....	40
2.9 Purification of <i>CPR5</i> protein variants	40
2.9.1 Sample preparation.....	40
2.9.2 Resin (sepharose beads) preparation.....	41
2.9.3 Purification using His-tag	41
2.9.4 Purification using ion-exchange chromatography	41
2.9.5 List and recipes of buffers used for Ni ⁺ -column based purifications.....	42
2.9.6 List and Recipes of buffers used for ion-exchange purifications.....	43
2.9.7 Preparation of 12% acrylamide gels.....	44
2.10 In-gel protein digestion for mass-spectrometry.....	44
2.10.1 Excision of the protein bands of interest from Gel	44
2.10.2 Solutions and suffers used for In-gel protein digestion	46
2.11 General sowing and plant growth conditions.....	46
2.12 Trichome counting and leaf area measurements	46
2.13 <i>CPR5</i> gene synthesis, site-directed mutagenesis and transformations	47
2.13.1 Gene Synthesis	47
2.13.2 <i>SynCPR5</i> transgenes transformation into <i>Agrobacterium GV3101</i>	48
2.13.3 <i>Agrobacterium</i> -mediated transformations of <i>cpr5-2</i> plants.....	48
2.13.4 Confirmation of positive transformants (genotyping)	49

2.14 <i>Pseudomonas syringae</i> infiltrations	49
2.15 Electron Scanning Microscopy (SEM) and cell size measurements	50
2.16 Ploidy level measurement using Flow Cytometer	51
2.17 Preparation of antibiotics	53
2.17.1 Ampicillin sodium salt (Sigma)	53
2.17.2 Kanamycin sulphate (Sigma)	53
2.17.3 Tetracycline hydrochloride (Sigma)	53
2.17.4 Chloramphenicol (Duchefa)	53
2.17.5 Rifampicin (Duchefa)	53
2.17.6 Gentamycin (Duchefa)	53
Chapter 3 <i>In silico</i> Characterization of CPR5	54
3.1 INTRODUCTION	54
3.2 RESULTS	55
3.2.1 CPR5 is widely distributed throughout the plant kingdom	55
3.2.2 CPR5 <i>in plant</i> homology amongst its homologues	55
3.2.3 AtCPR5 is predicted to have 4 or 5 transmembrane regions	56
3.2.4 AtCPR5 carries two casein kinase phosphorylation sites	62
3.2.5 AtCPR5 carries multiple transcription or translation initiation sites	62
3.2.6 AtCPR5 is annotated to contain intrinsically disordered regions	65
3.2.7 AtCPR5 is also predicted to contain coiled-coil domains	65
3.2.8 CPR5 is predicted to have glycine zipper	67
3.3 DISCUSSION	71
3.3.1 AtCPR5 appears to be an important membrane protein	71
Chapter 4 AtCPR5 may be one of the Intrinsically Disordered Proteins	72
4.1 INTRODUCTION	72
4.2 RESULTS	75
4.2.1 Like typical IDRs, CPR5 IDR is highly polymorphic in amino acid composition	75
4.2.2 Amino acid composition of AtCPR5 IDRs is consistent with typical IDRs	75
4.2.3 IDRs of CPR5 and CPR5-like proteins appear to have INDELS	79

4.2.4 AtCPR5 IDRs are annotated to contain MoRFs and disordered binding regions.....	79
4.2.5 AtCPR5 disordered regions annotate to be unfolding and flexible regions	80
4.3 DISCUSSION	81
4.3.1 AtCPR5 appears to be an intrinsically disordered protein	81
4.3.2 Having IDP characteristics, AtCPR5 could exert pleiotropic effects.....	81
Chapter 5 <i>In silico</i> characterisation and purification of AtCPR5 protein for structural studies	84
5.1 INTRODUCTION.....	84
5.2 RESULTS.....	86
5.2.1 Eliminations of predicted transmembrane domains	86
5.2.2 Elimination of predicted AtCPR5 IDRs.....	86
5.2.3 Eliminations of TM and IDR show no improvement in crystallizability of CPR5 truncated proteins	87
5.2.4 <i>CPR5</i> constructs were cloned in pET32 expression vector	87
5.2.5 Integration and orientation of <i>CPR5</i> transgenes were confirmed via PCR and restriction analyses.....	88
5.2.6 AtCPR5 recombinant proteins were soluble in Rosetta-gami cells.....	93
5.2.7 Purification of His-tagged AtCPR5 recombinant proteins	93
5.2.8 Purification of untagged versions using ion exchange chromatography.....	98
5.2.9 Identification of CPR5 protein by Mass Spectrometry	98
5.2.10 CPR5 protein was failed to be purified using size exclusion chromatography	99
5.3 DISCUSSION	102
5.3.1 Potential consequences of transmembrane domains eliminations.....	102
5.3.2 Putative impacts of IDRs eliminations on CPR5 structure	103
5.3.3 Purification of AtCPR5 protein is not trivial.....	104
Chapter 6 Development of CPR5 mutants based on <i>in silico</i> information.....	106
6.1 INTRODUCTION.....	106
6.2 RESULTS.....	107
6.2.1 Putative IDRs were deleted to study their roles in CPR5 functions	107
6.2.2 Putative start codons were mutated to study CPR5 isoforms.....	108

6.2.3 NLS clusters were mutated individually as well as collectively.....	108
6.2.4 Casein kinase phosphorylation site 1 and 2 were deleted and mutated.....	109
6.2.5 Putative leucine residues of coiled-coil domains were mutated into asparagine.....	112
6.2.6 Putative glycine residues of glycine zipper were mutated alanine.....	112
6.2.7 CPR5 transgenes were complemented into <i>cpr5-2</i> plants.....	113
6.3 DISCUSSION	118
6.3.1 Mutagenesis studies are expected to identify roles of various putative motifs of <i>CPR5</i>	118
6.3.2 <i>CPR5</i> transgenes were introduced into <i>cpr5-2</i> plants using floral dipping.....	120
Chapter 7 AtCPR5 regulates balance between plant growth and disease	
resistance.....	122
7.1 INTRODUCTION.....	124
7.2 RESULTS.....	126
7.2.1 <i>CPR5</i> native promoter is able to drive <i>SynCPR5</i> expression.....	126
7.2.2 <i>SynCPR5</i> complements aberrant trichomes, lesions and early leaf senescence	127
7.2.3 Leaf size and number of trichomes on <i>SynCPR5</i> plants is consistent with their <i>SynCPR5</i> expression levels.....	128
7.2.4 <i>SynCPR5</i> fails to complement upregulated levels of <i>PR1</i> in <i>SynCPR5</i> lines	130
7.2.5 <i>SynCPR5</i> and <i>cpr5-2</i> plants both show reduced <i>PDF1.2</i> levels.....	130
7.2.6 <i>SynCPR5</i> regulate resistance independent of <i>SynCPR5</i> and <i>PR1</i> levels.....	131
7.2.7 <i>SynCPR5</i> plants show wildtype-like ploidy levels	133
7.2.8 <i>metCPR5</i> complement aberrant trichomes, HR-like lesions and leaf yellowing shown by <i>cpr5-2</i>	136
7.2.9 <i>metCPR5b</i> lines show higher transcript abundance than wildtype.....	137
7.2.10 <i>metCPR5b</i> and <i>metCPR5bc</i> have bigger leaves than wildtype.....	137
7.2.11 <i>metCPR5b</i> and <i>metCPR5bc</i> have wildtype-like epidermal pavement cells	138
7.2.12 <i>metCPR5b</i> and <i>metCPR5bc</i> leaves have higher ploidy levels	144
7.2.13 <i>metCPR5b</i> and <i>metCPR5bc</i> have lower or wildtype-like <i>PR1</i> levels.....	145
7.2.14 <i>metCPR5</i> plants conferred enhanced susceptibility to <i>P syringae</i>	145
7.3 DISCUSSION	148
7.3.1 <i>SynCPR5</i> transgenic plants have variable <i>SynCPR5</i> expression levels	148

7.3.2 <i>SynCPR5</i> complements majority of <i>cpr5-2</i> compromised phenotypes	149
7.3.3 Phenotypes suggest quantitative as well as qualitative roles for CPR5	150
7.3.4 <i>PDF1.2</i> is expressed at lower levels in <i>SynCPR5</i> and <i>cpr5-2</i> than <i>CPR5</i>	151
7.3.5 Putative roles for nucleotide residues associated with alternative start codons	153
7.3.6 Putative roles of CPR5 in regulation of balance between growth and resistance	157

Chapter 8 AtCPR5 putative NLS clusters appear to have limited function in CPR5 nuclear localisation and may be part of a structural component of the Intrinsically Disordered Region162

8.1 INTRODUCTION.....	164
8.2 RESULTS.....	167
8.2.1 AtCPR5 is predicted to contain 3 NLS-encoding clusters	167
8.2.2 Putative NLS clusters are highly conserved among AtCPR5 homologues	168
8.2.3 CPR5 putative NLS-encoding clusters are predicted to be flanked by CKI and CKII phosphorylation sites.....	173
8.2.4 <i>nlsCPR5</i> complemented lines show variation in growth.....	173
8.2.5 <i>nlsCPR5</i> plants show reduced <i>nlsCPR5</i> expression levels.....	174
8.2.6 <i>nlsCPR5</i> transgenic lines display smaller leaves	178
8.2.7 Epidermal pavement cells are smaller on <i>nlsCPR5</i> transgenic leaves.....	178
8.2.8 Cell expansion appears to be affected in leaves of <i>nlsCPR5</i> transgenic plants.....	179
8.2.9 Degree of trichome aberration varies among <i>nlsCPR5</i> lines.....	183
8.2.10 <i>nlsCPR5</i> transgenic plants confer resistance to <i>Pseudomonas syringae</i>	191
8.2.11 <i>nlsCPR5</i> mutants show elevated levels for <i>PR1</i> and <i>PR5</i> genes	191
8.2.12 <i>nlsCPR5</i> transgenic plants display downregulation of <i>PDF1.2</i>	191
8.2.13 <i>nlsCPR5</i> mutant plants show reduced plant height	192
8.2.14 Collective mutations in NLS-encoding clusters results in early bolting.....	195
8.2.15 Putative CPR5 NLS clusters and an intrinsically disordered region belong to the same region of CPR5 protein.....	195
8.2.16 <i>Del37CPR5</i> and <i>Del63CPR5</i> complement HR-like lesions and early leaf yellowing.....	196
8.2.17 <i>Del63CPR5</i> transgenic lines display reduced transcript abundance	197

8.2.18 Typical pattern of trichomes is restored by <i>Del37CPR5</i> but not by <i>Del63CPR5</i> and <i>Del98CPR5</i>	197
8.2.19 <i>Del37CPR5</i> and <i>Del63CPR5</i> both produce leaves of intermediate sizes	201
8.2.20 Sizes of epidermal cells are consistent with leaf sizes of <i>DelCPR5</i> plants	201
8.2.21 Nuclei from <i>DelCPR5</i> leaf cells show reduced ploidy levels	201
8.2.22 <i>Del63CPR5</i> confers resistance but <i>Del37CPR5</i> is susceptible to <i>PstDC3000</i>	202
8.2.23 <i>PR1</i> levels are not restored in <i>Del63CPR5</i> transgenic plants.....	202
8.3 DISCUSSION	207
8.3.1 <i>In silico</i> studies suggest CPR5 to be a nuclear protein	207
8.3.2 Some CPR5 phenotypes/functions appear to be dependent on <i>CPR5</i> expression.....	208
8.3.3 Putative NLS clusters appear to have limited function	209
8.3.4 First 37 amino acids of CPR5 are crucial for wildtype-like leaf sizes	210
8.3.5 <i>nlsCPR5</i> -like phenotypes on <i>DelCPR5</i> plants suggest NLS clusters to be part of IDRs ...	211
Chapter 9 Characterization of putative CPR5 leucine and glycine motifs	215
9.1 INTRODUCTION.....	215
9.2 RESULTS.....	217
9.2.1 AtCPR5 protein is predicted to contain 4 coiled-coil domains.....	217
9.2.2 AtCPR5 protein is predicted to contain glycine motifs	217
9.2.3 <i>ccdCPR5</i> plants display <i>cpr5</i> -like phenotypes.....	220
9.2.4 Glycine residues of the glycine motif show intolerance to mutations	220
9.3 DISCUSSION	223
9.3.1 Leucine residues are crucial for CPR5 functioning	223
9.3.2 Glycine 452 and 459 are also crucial for CPR5 functions.....	223
Chapter 10 FINAL DISCUSSION AND FUTURE PROSPECTS	226
10.1 AtCPR5 is a nuclear membrane protein.....	226
10.2 CPR5 may exert pleiotropic effects by modifying NPC-mediated selectivity and entry of nuclear proteins	228
10.3 CPR5 IDRs and Coiled-coil-domains may be involved in CPR5-mediated NPC gating.....	230
Chapter 11 APPENDICES.....	234

11.1 List of primers.....	234
11.1.1 Primers for the amplification of untagged <i>CPR5</i> constructs.....	234
11.1.2 List of primers for the amplification of <i>CPR5</i> constructs with N-ter His-tag.....	234
11.1.3 List of primers for the amplification of <i>CPR5</i> constructs with C-ter His-tag.....	235
11.1.4 List of primers used for real-time quantifications.....	235
11.1.5 List of genotyping primers.....	236
11.2 List of accession number of CPR5-like proteins.....	237
11.3 Students' t-test results.....	238
11.3.1 Expression levels: Statistical significance at $p < 0.05$ (Students' t-test).....	238
11.3.2 Trichomes with three appendages: Statistical significance at $p < 0.05$ (Students' t-test).....	238
11.3.3 Area of third leaf pair: Statistical significance at $p < 0.05$ (Students' t-test).....	239
11.3.4 Area of fourth leaf pair: Statistical significance at $p < 0.05$ (Students' t-test).....	239
11.3.5 <i>PR1</i> transcript abundance: Statistical significance at $p < 0.05$ (Students' t-test).....	240
11.3.6 Expression levels: Level of statistical significance at $p < 0.05$ (Students' t-test).....	241
REFERENCES.....	242

LIST OF FIGURES

Figure 1.1 Annotated architecture of <i>CPR5</i> Gene.....	7
Figure 2.1 Positions of primers used for real-time qRT-PCR quantifications.....	34
Figure 2.2 Schematic representation of transgene cassette of pET32.....	35
Figure 2.3 pET32 sequence composition and landmark information.....	36
Figure 2.4 Schematic diagram for serial dilutions.....	52
Figure 3.1 Multiple sequence alignment of CPR5 putative homologues.....	61
Figure 3.2 Annotation, position and number of TMs in CPR5 protein.....	64
Figure 3.3 Positions of in-frame start codons and RNA stem-loop structures.....	66

Figure 3.4 Position and number of Intrinsically Disordered Regions	69
Figure 3.5 Position and number of annotated coiled-coil regions	70
Figure 3.6 Position of glycine residues in CPR5 protein.....	70
Figure 4.1 Position of disordered and binding regions in AtCPR5	77
Figure 4.2 Presence of polymorphism in the N-terminus of CPR5 and CPR5-like proteins sequences	78
Figure 4.3 Folding and unfolding propensities of regions of CPR5.....	80
Figure 5.1 Positions of various selected truncations in CPR5 protein.....	90
Figure 5.2 Migration of <i>CPR5</i> cDNA amplified fragment	91
Figure 5.3 Amplification of <i>CPR5</i> constructs	91
Figure 5.4 Confirmation of integration of constructs in pET32 by PCR.....	92
Figure 5.5 Confirmation of integration of constructs in pET32 by restriction analyses	92
Figure 5.6 Confirmation of expression and solubility of CPR5 tagged recombinant proteins.....	96
Figure 5.7 Confirmation of binding affinity of CPR5 recombinant protein B	97
Figure 5.8 Confirmation of expression and purification of untagged version	100
Figure 5.9 CPR5 identification by Mass-spectrometry	101
Figure 6.1 Construct designing of <i>Del37CPR5</i> , <i>Del63 CPR5</i> and <i>Del98CPR5</i> transgenes	110
Figure 6.2 Designing of <i>metCPR5</i> transgenes.....	111
Figure 6.3 Similarities and differences between <i>CPR5</i> wildtype and synthetic gene and positions of structural sites	116
Figure 7.1 <i>SynCPR5</i> complementation, transcript and trichomes quantification.....	129
Figure 7.2 Mean leaf area of third and fourth rosette leaf pairs.....	132
Figure 7.3 Relative expression levels of <i>PR1</i> in <i>SynCPR5</i> plants	132
Figure 7.4 Relative expression levels of <i>PDF1.2</i> in <i>SynCPR5</i> plants	134

Figure 7.5 <i>PstDC3000</i> infection growth assay	134
Figure 7.6 Ploidy levels in leaves of <i>SynCPR5</i> transgenic plants.....	135
Figure 7.7 Overexpression, <i>metCPR5</i> , Col-0 and <i>cpr5-2</i> plants at 17 DAS.....	139
Figure 7.8 <i>metCPR5</i> transgene and trichome quantification in <i>metCPR5</i> plants.....	140
Figure 7.9 Mean area of leaves and epidermal pavement cells of <i>metCPR5</i> plants.....	143
Figure 7.10 Ploidy levels in leaves of <i>SynCPR5</i> transgenic plants.....	146
Figure 7.11 Relative expression levels of <i>PR1</i> in <i>SynCPR5</i> plants.....	147
Figure 7.12 <i>PstDC3000</i> infection growth assay	147
Figure 8.1 Positions of potential nuclear localisation signal (NLS) facilitating basic residues in AtCPR5 and its homologues	170
Figure 8.2 Position of annotated NLS clusters and casein kinase phosphorylations sites among CPR5-like proteins	172
Figure 8.3 <i>nlsCPR5</i> and <i>DelCPR5</i> plants at 17 DAS.....	175
Figure 8.4 Transcript abundance of <i>CPR5</i> in <i>nlsCPR5</i> and <i>DelCPR5</i> transgenic lines	176
Figure 8.5 Mean leaf areas of <i>nlsCPR5</i> lines.....	177
Figure 8.6 Epidermal pavement cell size	182
Figure 8.7 Measurements of ploidy levels in leaves of <i>nlsCPR5</i> mutants at 24 DAS	185
Figure 8.8 Type and number of trichome appendages at 16 DAS.....	190
Figure 8.9 <i>Pseudomonas syringae</i> pv <i>DC3000</i> growth assay.....	193
Figure 8.10 Quantification of transcript abundance of <i>PR1</i> , <i>PR5</i> , and <i>PDF1.2</i>	194
Figure 8.11 Plant height measurement	198
Figure 8.12 Plant growth and bolting of <i>nlsCPR5</i> transgenic lines at 27 DAS.....	200
Figure 8.13 Type and average number of trichomes on <i>DelCPR5</i> leaves	203
Figure 8.14 Mean leaf area of <i>DelCPR5</i>	203

Figure 8.15 Mean area of epidermal pavement cells	204
Figure 8.16 Ploidy levels measurement in leaves of <i>DelCPR5</i> mutants	205
Figure 8.17 <i>Pseudomonas syringae</i> infection assay	206
Figure 8.18 Position of structural motifs present in CPR5 first 98 amino acids	214
Figure 9.1 Position of putative leucine and glycine residues of leucine and glycine motifs	219
Figure 9.2 Plant growth and early leaf senescence on <i>ccdCPR5</i> and <i>GlnCPR5</i> leaves.....	222
Figure 10.1 Proposed CPR5 schematic diagram in nuclear lemma and its working	233

LIST OF TABLES

Table 4.1 Positions of predicted MoRFs and binding regions in CPR5	77
Table 5.1 Predicted position of TM domains in CPR5 protein	89
Table 5.2 XtalPred probabilities for CPR5 recombinant proteins to be crystallized ...	89
Table 6.1 Mutations in putative NLS-encoding residues	111
Table 6.2 Designing of CK phosphorylation site mutants.....	114
Table 6.3 Designing of leucine and glycine zipper mutants	114
Table 6.4 List of <i>SynCPR5</i> transgenes and their transformation efficiency	117
Table 7.1 Average number of cells per image.....	143

LIST OF ABBREVIATIONS

aa	Amino acid
ABA	Abscisic acid
ABI5	ABA-INSENSITIVE 5
<i>acd6</i>	<i>ACCELERATED CELL DEATH 6</i>
APC/C	anaphase-promoting complex/cyclosome
BRI	BRASSINOSTEROID-INSENSITIVE
BRs	Brassinosteroids
Ca	Calcium
CaM 35S promoter	Cauli Mosaic Virus 35S promoter
CC-domains	Coiled-coil domains
<i>CDC20</i>	<i>CELL DIVISION CYCLE 20</i>
CDH	<i>CDH HOMOLOG 1</i>
CDK	Cyclin-Dependant-Kinase
CK	Casein kinase
CKI	CDK inhibitor
<i>CNGC</i>	<i>Cyclic Nucleotide Gated Channels</i>
<i>COI1</i>	<i>CORONATINE-INSENSITIVE 1</i>
Col-0	<i>Arabidopsis</i> ecotype Columbia
<i>CPR5</i>	<i>CONSTITUTIVE EXPRESSER of PATHOGENESIS-RELATED GENES 5</i>
DAS	Days after sowing
<i>EDR1</i>	<i>ENHANCED DISEASE RESISTANCE 1</i>
<i>EDS5</i>	<i>ENHANCED DISEASE SUSCEPTIBILITY 5</i>
<i>EIN2</i>	<i>ETHYLENE INSENSITIVE 2</i>
ET	Ethylene
ETI	Effector-Triggered Immunity
<i>FZR</i>	<i>FIZZY-RELATED</i>
GA	Gibberellin
<i>GeBP</i>	<i>GL1 ENHANCER BINDING PROTEIN</i>
GFP	Green Fluorescent Protein
<i>GIG1</i>	<i>GIGAS cell 1</i>
<i>GPLs</i>	<i>GeBP</i> -like proteins
GST	Glutathione-S-transferases
H ₂ O ₂	Hydrogen Peroxide
HR	Hypersensitive Response
<i>HXK</i>	<i>Hexokinase</i>
<i>HYS1</i>	<i>HYPERSENESCENCE1</i>
IDP	Intrinsically Disordered Protein
IDR	Intrinsically Disordered Region
JA	Jasmonic Acid
<i>JAR1</i>	<i>JASMONATE RESISTANT 1</i>
<i>JAZ1</i>	<i>JASMONATE-ZIM-DOMAIN PROTEIN 1</i>

K	Potassium
KRPs	<i>KIP-RELATED PROTEINS</i>
<i>Ler-0</i>	<i>Landsberg erecta</i>
LOX	Lipoxygenases
<i>LSD</i>	<i>LESIONS SIMULATING DISEASE</i>
MeJA	Methyl jasmonate
MoRFs	Molecular Recognition Features
mRNA	Messenger RNA
NDGA	Nordihydroguaiaretic acid
NE	Nuclear Envelope
NLS	Nuclear Localization Signal
NO	Nitric oxide
NPC	Nuclear Pore Complex
<i>NPR1</i>	<i>NON-EXRESSER of PATHOGENESIS-RELATED GENES 1</i>
OE lines	Overexpression lines
<i>OLD1</i>	<i>ONSET OF LEAF DEATH1</i>
<i>OSD1</i>	<i>OMISSION of SECOND DIVISION 1</i>
<i>PAD4</i>	<i>PHYTOALEXIN DEFICIENT 4</i>
PAMPs/ MAPMs	Pathogen- or microbe-associated molecular patterns
PCD	Programmed Cell Death
PCR	Poly Chain Reaction
<i>PDF1.2</i>	<i>PLANT DEFENSIN 1.2</i>
<i>PIF</i>	<i>PHYTOCHROME INTERACTING FACTOR</i>
<i>PR1</i>	<i>PATHOGENESIS-RELATED 1</i>
PRRs	Pathogen Recognition Receptors
<i>PstDC3000</i>	<i>Pseudomonas syringae pv DC3000</i>
PTI	PAMP-Triggered Immunity
qRT-PCR	Quantitative Reverse-Transcriptase Poly Chain Reaction
R proteins	Resistance proteins
RB	Retinoblastoma
ROS	Reactive Oxygen Species
<i>RPM1</i>	<i>RESISTANCE TO PSEUDOMONAS SYRINGAE PV MACULICOLA1</i>
<i>RPS2</i>	<i>RESISTANCE TO PSEUDOMONAS SYRINGAE</i>
SA	Salicylic Acid
<i>SAGs</i>	<i>SENESCENCE-ASSOCIATED GENES</i>
SAR	Systemic Acquired Resistance
<i>SIM</i>	<i>SIAMESE</i>
<i>SMR</i>	<i>SIAMESE-RELATED</i>
SOD	Superoxide Dismutase
TFs	Transcription Factors
TM	Transmembrane
tRNA	Transfer RNA
<i>UVI4</i>	<i>UV INSENSITIVE 4</i>

