Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

# PHYLOGENETICS, DIVERGENCE AND MORPHOLOGY OF NEW ZEALAND ELEOTRIDAE (*GOBIOMORPHUS* GILL)

A thesis

presented in partial fulfilment

of the requirements for the Degree

of

## Master of Science in Ecology

at

Massey University, Palmerston North

New Zealand

by

**Emily Atkinson** 



#### ABSTRACT

The genus *Gobiomorphus* Gill (1863) is the only representative freshwater Eleotridae in New Zealand and is comprised of seven species, of which four are diadromous. The species are endemic to New Zealand and are widespread around lowland streams and coasts (with non-diadromous species penetrating further inland). The only other *Gobiomorphus* species are *G. coxii* and *G. australis*, which are endemic to Australia. Eleotridae are stocky fishes of small size (up to 150 mm in length) (McDowall, 1990) and are characterised by two dorsal fins, large pectoral fins, separate thoracic pelvic fins (fused in gobies) and the absence of a lateral line (McDowall, 1990; Allen *et al.*, 2002).

Gobiomorphus has had a particularly turbulent taxonomic history in the literature, spanning approximately 150 years from Gill (1863) to the present, where many species have been synonymised with one another (particularly, most Gobiomorphus species were synonymised with G. gobioides) due the plasticity of many morphological characters. Additionally, similar morphologies have led to identification difficulties. Phenotypic plasticity can also make cladistic approaches difficult (e.g. Vrijenhoek, 1998; Orti et al., 1994), for example there has been a lot of controversy surrounding G. alpinus and its species status. Furthermore, studies of evolution using morphological characters often lack an accurate perspective on relationships and origins of fish species, in particular, little information exists on the evolutionary origins of the Gobiomorphus genus. Genetic studies have contributed to resolving problems with taxonomically difficult groups by detecting diversity between morphologically similar species (where DNA variation is often not expressed phenotypically), and examining geographical divergence within species (e.g. Vrijenhoek, 1998; Kocher et al., 1989). Thus, this thesis employed two regions of mitochondrial DNA (cytochrome b and control region) to resolve issues surrounding species identification, morphological variation, phylogenetic relationships (including divergence), origins and the evolution of diadromy within the Gobiomorphus group.

Mitochondrial DNA sequences were obtained from all seven *Gobiomorphus* species in New Zealand, as well as from both Australian *Gobiomorphus*. The morphology of both *G. basalis* and *G. breviceps* in the lower North Island was also examined. The results suggested that the Australasian *Gobiomorphus* are a polyphyletic group, although with the exclusion of *G. australis* the rest of the species formed a

monophyletic group. The Australian group formed a polyphyletic group basal to the New Zealand monophyletic group. *Gobiomorphus hubbsi*, a diadromous species was found to be a sister group to the New Zealand *Gobiomorphus*. Clock calibrations indicated that the New Zealand and Australian groups have been isolated for about 6-37 Myr, suggesting that the New Zealand species dispersed here (in a single event) from Australia post-Gondwana break-up. These results are discussed in terms of New Zealand's geological history. Once in New Zealand there was a series of radiations; the most recent radiation produced the non-diadromous species (*G. breviceps*, and a *G. basalis*, *G. cotidianus* (although not all populations are diadromous) and *G. alpinus* species complex). Furthermore, *G. huttoni* and *G. gobioides* (both diadromous) formed a monophyletic group that is part of the first radiation, indicating that diadromy is a primitive feature of *Gobiomorphus*.

Mitochondrial DNA accurately distinguished between *G. breviceps* and *G. basalis* (suggesting a genetic basis to morphological variation), and coupled with morphological data, identified pectoral fin ray counts as the best quantitative character for differentiating the species. However, within species high morphological variation was observed that did not fit expected patterns of geographical divergence. Limited time periods may have obscured subtle morphological divergence between catchments. Mitochondrial DNA revealed some unique haplotypes within both catchments, whereas some catchments shared identical haplotypes. The lack of divergence between catchments may have been due in part to connections during the Pleistocene, whereas populations with unique haplotypes may have been isolated for a greater length of time.

Collectively, these studies highlighted the usefulness of mitochondrial DNA for exploring; phylogenetic relationships (including divergence) and solving problems with taxonomically difficult groups, and origins of fish species. Furthermore, the use of molecular data coupled with morphological data can be used to aid in the improvement of identification of morphologically similar species.

#### PREFACE and ACKNOWLEDGEMENTS

Each chapter has its own acknowledgements section. Here I thank those people whose assistance was not specific to any one chapter of this thesis.

First I would like to thank my supervisor, Dr Michael Joy for providing excellent guidance throughout the long and often difficult journey that a thesis can often be. I would also like to thank Mike for his humour (although often very dry) and for the 'counselling' sessions that myself and other students made full use of! Furthermore, I would like to thank my co-supervisors Dr Mark Stevens and Dr Russell Death. Thankyou Mark for your excellent tutoring and crash course in genetic analysis; from the laboratory work to the write-up. Without you I do not think my thesis would have been so interesting! Russell you have my gratitude for the help that you gave me early on during my undergraduate study. You are the one that facilitated my interests in freshwater ecology and helped me plan my future career, which is turning out to be rather fun! Without you I may not have realised my aims in life.

I would also like to acknowledge the support given to me by the Julie Alley bursary, which gave me some much needed financial aid.

I am also very grateful to the other post-graduate students around the ecology department (as well as a few in the Allan Wilson Centre) who provided many hours of entertainment, especially during lunch breaks that may have at times gone on for a little too long! Having people in the same boat as you also helps you stay sane and focussed. Angela, thank-you for your guidance and providing for me with an outlet when things got a little frustrating! I also really enjoyed those coffee breaks (which often went a little longer than planned) even when we didn't actually get coffee, and I hope we will always find the time to sit down together every once in awhile. I also must thank Richard Seaton for sharing his office with me, especially as I gave him so much cheek! One day Richard you'll be able to get me back (how does the score stand? 100 to me and 1 to you I think!).

The support provided by the rest of the staff in Ecology has been phenomenal. A big thank-you goes to all the technicians for helping with field work and anything else I needed, and also to the office staff who provided so much help on so many levels. I would also like to thank the rest of the lecturers in the department for many helpful comments on my project and for giving me the encouragement I often needed. During

my time in Ecology I also really enjoyed the morning tea shouts (at which all postgraduate students could be found!), which provided me with much sustenance. Long live the morning tea shouts!

To my Mum and Dad; thank-you so much for everything you have done for me, without you I could not have achieved what I have. Your love, support and confidence in my ability has meant the world to me. My achievements are yours to be proud of too. Thank-you also to my siblings (James, Melissa, Claire and Benjamin) for just being there and for fun chats on the phone. Nanna, Grandad, and my many Aunties and Uncles have also contributed; they have always provided me with support and shown a strong interest in what I am up to, so for that I thank-you. My cat Mia has also helped in her own way. Her constant need for my attention and loud purrs have reminded me to think beyond my own problems and needs.

Last, but certainly not least a big thanks goes to my partner Mike Ushaw, who has been infinitely patient with me and provided me with much love and support, which is essential when completing a task such as a thesis. Cheers for sticking around, especially as I may not have always been in the best mood! I would also like to thank Mike's family who have treated me as one of their own and taken me into the fold. This has been especially wonderful, as my own family is in the South Island, so it is nice to know I have support in the North!

If I have forgotten anyone, my apologies; many people contribute in positive ways to your life, but it is very difficult to thank everyone, but I think I have covered the main ones.

# TABLE of CONTENTS

ABSTRACTi
PREFACE and ACKNOWLEDGEMENTSiii
LIST of TABLESvii
LIST of FIGURESvii
LIST of APPENDICESix
THESIS INTRODUCTION1
Literature Cited
CHAPTER I - PHYLOGENY OF NEW ZEALAND AND AUSTRALIAN
ELEOTRIDAE WITH A FOCUS ON GOBIOMORPHUS
Abstract9
Introduction10
Materials and Methods
Specimen collection13
DNA analyses17
Data analyses18
Results
Cytochrome b19
Control region
Discussion
Conclusions
Acknowledgements
Literature Cited
CHAPTER II - GENES AND MORPHOLOGY: A COMPARATIVE EXAMINATION
OF C. PREVICERS AND C. RASALIS IN THE LOWER NORTH ISLAND. NEW

OF G. BREVICEFS AND G. BASALIS IN THE LOWER NORTH ISLAND, I	NE W
ZEALAND	45
Abstract	46
Introduction	47
Materials and Methods	49

Specimen collection49
DNA analyses
Morphological analyses
Data analyses
Results
Molecular analysis
Morphological analysis
Discussion
Conclusions
Acknowledgements
Literature Cited
THESIS CONCLUSION
Future Research
Literature Cited
Appendicesfacing page 82

#### LIST of TABLES

**TABLE I.1** Sampling locations for all specimens throughout New Zealand and
 Australia (see also Smith et al., 2005; Akihito et al., 2000 and Thacker & Hardman, 2005). [] indicate sequences vet to be placed on GenBank, numbers indicate number of sequences for each mtDNA region, and (-) no sequences for the region. \*- sequenced by M. Stevens and B. Hicks (unpublished data), all others collected for the purpose of this study. Sequences with GenBank accession numbers: AY644 - Smith et al. (2005); AY722 – Thacker & Hardman (2005); ABO212 – Akihito et al. (2000).....15 **TABLE I.2** The 375 variable sites for cytochrome *b* region for *Gobiomorphus*. Locations are indicated using location codes from Table 1. Identical character states are indicated by dots. A – Gobiomorphus gobioides ......facing page 20 TABLE I.3 Sequence divergence (both regions of mtDNA) between G. coxii and the extant New Zealand Gobiomorphus taxa......31 
**TABLE I.4** The 368 variable sites for control region sequences for Gobiomorphus.
 Locations are indicated using codes from Table 1. Identical character states are indicated by dots. A = G. gobioides.....facing page 24 **TABLE II.1** Variable sites. The first set of variable sites is the cytochrome b sequences and the second is the control region sequences. Locations where each haplotype was found are indicated using location codes from fig. 3 and haplotypes are given a single letter code (i.e. A,B etc.). Identical character states are indicated by dots.

The (-) indicates a deletion..... facing page 55

#### LIST of FIGURES

the mitochondrial DNA (cytochrome b) gene using only unique sequences. Bootstrap confidence limits (500 replicates) shown above nodes. Codes are those used in Table 1. Number of identical haplotypes present at any site is given in parentheses. Symbols indicate life history patterns; open circle = non-diadromous, closed circle = diadromous, half circle = presence of both life history patterns. G. huttoni species in bold is Chatham Island haplotype. A – N/KR(2), W/WR(1), BOP/LT(2), BOP/RR(3), BOP/KR(1), NE/M(3), EC/G(3), M/N(2), WC/CR(2). B – N/SS(1), W/M(3), M/N(5), WC/LB(2). C - N/KR(1), N/LN(3), N/N(3), N/LW(3), N/WR(2), W/M(1). D - M/T(8), M/W(1), M/TS(9), M/K(12), M/M(1), Wa/K(7), Wa/WR(1), Wa/H(2), Wa/M(1), We/K(2), We/ES(1). E – N/KR(2), N/WR(2), N/Mi(2), N/M(1), N/US(2), W/M(2).....23 **FIGURE I.3** Maximum likelihood phylogram based on the substitution model HKY+I+G (-InL = 2123.9351; base frequencies set to A = 0.3922, C = 0.1775, G = 0.1356, T = 0.2947) derived from Modeltest vers. 3.7 (see methods), using a 384-bp fragment of the mitochondrial DNA (control region) gene using only unique sequences. Bootstrap confidence limits (500 replicates) shown above nodes. Codes are those used in Table 1. Number of identical haplotypes present at any site is given in parentheses. Symbols indicate life history; open circle = non-diadromous species, closed circle = diadromous species, half circle = presence of both life history patterns. G. huttoni in bold is the Chatham Island haplotype......27 FIGURE II.1 The location of the sites sampled for bullies in the Wellington, Manawatu and Wairarapa regions, North Island, New Zealand......50 FIGURE II.3 Haplotype network (neighbour joining with 100 bootstraps) showing mtDNA haplotypes and their relative connections, with bootstrap support. Data from: **FIGURE II.4** Discriminant analysis of all three species based on morphological data. The species form three distinct clusters with some overlap. Up= G. breviceps, Cr = G. *basalis* and Co = G. 

### LIST of APPENDICES

#### THESIS INTRODUCTION

The freshwater bullies of New Zealand form part of the genus Gobiomorphus Gill, 1863 (Eleotridae), which consists of nine species to date. The name Gobiomorphus can be broken down into two parts, with Gobio recognising the similarity between them and the European gudgeon Gobio gobio and morphus taken from the Latin word representing form or shape (McDowall, 1990). They are stocky fishes of small size (up to 150 mm in length) and are characterised by two dorsal fins, large pectoral fins, separate thoracic pelvic fins (fused in gobies) and the absence of a lateral line (McDowall, 1990; Allen et al., 2002). The bullies are widely distributed throughout New Zealand and two species (G. coxii (Cox's gudgeon) and G. australis (striped gudgeon)) are endemic to Australia and found on the Victorian south-eastern coast. The species occupy both brackish- and inland fresh-water habitats and are typically benthic, feeding on small aquatic invertebrates. While some members of the genus spend their entire life cycle in freshwater, four of the seven New Zealand species are diadromous, where part of their life cycle is spent in salt water. In particular, New Zealand Gobiomorphus are amphidromous (a form of diadromy) where the larvae migrate out to sea for a feeding phase before returning as juveniles to freshwater (McDowall, 1998).

Morphological characters have been used historically in systematics, however, several characteristics (e.g. fin ray numbers) that have been used to describe fish are often phenotypically plastic, where morphology tends to reflect the environmental conditions. This has made cladistic approaches difficult (Vrijenhoek, 1998; Orti *et al.*, 1994). Thus, in the early literature many species were not recognised (most species were originally described as the one species; *G. gobioides*) due to their similar morphology. Additionally, little information exists on the evolutionary origins of this genus, including explanations regarding how *Gobiomorphus* reached New Zealand, where the Eleotridae family is at southern-most limit (McDowall, 1975). However, many genetic studies have contributed to resolving problems with taxonomically difficult groups by detecting diversity between morphologically similar species (where DNA variation is often not expressed phenotypically), and examining geographical divergence within species (e.g. Vrijenhoek, 1998; Kocher *et al.*, 1989). In particular, mitochondrial DNA (mtDNA) has allowed for high resolution analyses of population-level questions in many species of fishes (Orti *et al.*, 1994), and has provided insights

into the origins, evolution and phylogenetics of many New Zealand fishes including Galaxiidae and *Gobiomorphus* (e.g. Waters *et al.*, 2000; Smith *et al.*, 2005). Thus, this thesis consists of two chapters dealing with genetic species identification, morphological variation, phylogenetic relationships (including divergence), origins and the evolution of diadromy within the *Gobiomorphus* group.

Chapter I utilised two regions of mitochondrial DNA (control and cytochrome *b* regions) to examine and resolve phylogenetic relationships within the *Gobiomorphus* genus and examine the relationship between *Gobiomorphus*, *Philypnodon* and *Eleotris*. Additionally, this chapter aimed to provide an understanding of the somewhat contentious view of species origins for this Australasian group, with a special focus on the New Zealand species and the evolution of diadromy.

Although five out of the seven currently recognised species were recognised in the early literature, there were some difficulties with identification. For example, *Gobiomorphus* was once considered part of the *Eleotris* and *Philypnodon* genera—*G. gobioides* (giant bully), *G. huttoni* (redfin bully), and *G. basalis* (Cran's bully) (*G. cotidianus* (common bully) was not described until 1975 by McDowall) were all part of the genus *Eleotris* before being moved to *Gobiomorphus* in the late 1800's; *G. hubbsi* (bluegill bully) and *G. breviceps* (upland bully) were part of the genus *Philypnodon* before being placed in *Gobiomorphus* in the 1960's. Chapter I aimed to address the confusion surrounding genetic species identification (and thus bully systematics) in New Zealand by applying phylogenetic analyses to this species group.

Additionally, while the origin of typically migratory New Zealand Galaxiidae has been well studied, with species distributions assigned as being either dispersal- or vicariance-driven (McDowall, 2002; Waters & Burridge, 1999), little is known of the origins of *Gobiomorphus* in New Zealand. However, some authors have suggested a role for dispersal in creating current *Gobiomorphus* distributions. For example, McDowall (1975) suggested that *Gobiomorphus* may have reached New Zealand via dispersal in prevailing ocean currents, and believes that *G. coxii* is ancestral to or has common ancestry with New Zealand *Gobiomorphus*. Meanwhile, Thacker and Unmack (2005) examined the origins of the Australian *Hypseleotris* species whose widespread distribution encompasses South Africa, Japan, Southeast Asia and Australia, and suggested that this species dispersed from Southeast Asia via marine environments before invading freshwaters in Australia. These findings highlight the dispersal potential of fish species within the Eleotridae family. However, *Gobiomorphus* origins may alternatively fit the vicariance theory such that current distributions may be a result of Gondwanan plate tectonics and speciation. Therefore, Chapter I also aims to examine this 'dispersal vs. vicariance' issue by examining which of these methods resulted in Gobiomorphus colonising New Zealand, and determining whether this involved single, or multiple dispersal events (if *Gobiomorphus* distribution is dispersal-driven). Finally, Chapter I examines the evolution of diadromy in Gobiomorphus. The presence of a saltwater-tolerant phase in diadromous fish species that allows dispersal has led to the belief that ancestry for these species must be marine. This has been argued against by McDowall (2004) who believes that diadromy simply implies an ancestry to other diadromous species elsewhere rather than a purely marine ancestry. To further complicate the issue, the genus *Rhyacichthys* is exclusively freshwater and is basal to the Eleotridae (Akihito et al., 2000; Thacker & Hardman, 2005), possibly indicating a freshwater ancestry for Gobiomorphus, and suggesting that non-diadromous bullies should have a more basal position within species groups than diadromous bullies. Alternatively, diadromy may be an ancestral character, which appears to be the case for galaxiid fishes, where the presence of a marine phase is found in the basal members of clades, but is absent in more derived species (Waters et al., 2000). Potentially diadromous species in the New Zealand Gobiomorphus could belong in basal position within the Gobiomorphus group, with non-diadromous species being of more recent descent. Therefore, placement of species on phylogenetic trees will help to infer ancestry or derivation of diadromy in Gobiomorphus.

Chapter II focuses on the identification and divergence of two non-migratory, morphologically similar species (*G. breviceps* and *G. basalis*) in the lower North Island of New Zealand. *Gobiomorphus breviceps* has a widespread distribution in New Zealand, encompassing the South Island as well as lower North Island's Manawatu, Wairarapa and Wellington regions (McDowall, 1990). Conversely, *G. basalis* has a widespread yet intermittent distribution in the North Island that reaches its southern limit at Wellington (McDowall, 2000). Both species occupy a variety of habitats at low altitudes (McDowall, 1975; 1990; 2000). The two species are sexually dimorphic; the adult males often have bright and distinct colouration that the females and juveniles lack. Colouration is often used as a diagnostic tool and is useful for identifying male bullies, but can cause complications when females and juveniles are being compared *in situ.* The overlap in distribution of the two species has led to confusion in the identification of females and juveniles in the lower North Island, which is further confounded by the presence of *G. cotidianus* (common bully) a morphologically similar species.

Morphological characters including body ratios and serially repeated structures such as fin rays (Christiansen *et al.*, 1988) allow species to be further distinguished. However, they may be strongly influenced by environmental factors, causing variation in populations (Smith *et al.*, 2003; King *et al.*, 2003). For example, *G. alpinus* (Tarndale bully) occupies an extreme high-altitude environment and has the lowest number of first dorsal spines for *Gobiomorphus*, which Smith *et al.* (2003) suggest may be the result of the cold water that the species live in. The large variation in meristic counts (e.g. fin ray counts) observed between populations in *G. basalis* and *G. breviceps* may also be a result of environmental influence. For example, *G. breviceps* have 4-7 first dorsal spines and *G. basalis* have 6-8 first dorsal spines. This overlap in fin ray counts between *G. breviceps* and *G. basalis* can make identification of species difficult.

Therefore, this chapter aimed to utilise the control and cytochrome *b* regions of mtDNA to accurately distinguish between *G. breviceps* and *G. basalis*. Upon species identification, meristic information is applied to individuals in order to identify diagnostic morphological characters. Additionally, the genetic and morphological information for both species is used to examine divergence (both genetic and morphological) to ascertain geographical patterns. It is thought that there would be genetic and morphological divergence between catchments, as there will be no dispersal (non-diadromous species), and there would be no genetic or morphological divergence within catchments as dispersal would be possible.

Collectively, the two chapters of this thesis address the phylogenetic relationships, origins and evolution of diadromy in *Gobiomorphus*, as well as resolving issues surrounding identification and morphological variation. This work demonstrates how genetic data can be used to resolve difficult taxonomic questions, and clarify evolutionary patterns and origins in fish species. The thesis concludes with an overall summary, and then addresses priorities for future research.

#### LITERATURE CITED

- Akihito, Iwata, A., Kobayashi, T., Ikeo, K., Imanishi, T., Ono, H., Umehara, Y.,
  Hamamatsu, C., Sugiyama, K., Ikeda, Y., Sakamoto, K., Fumihito, A., Ohno, S.
  & Gojobori, T. (2000). Evolutionary aspects of gobioid fishes based upon a phylogenetic analysis of mitochondrial cytochrome *b* genes. *Gene* 259: 5-15.
- Allen, G. R., Midgley, S.H., Allen, M. (2002). Field guide to the Freshwater Fishes of Australia. Perth, Western Australian Museum.
- Christiansen, F. B., Nielsen, V.H., Simonsen, V. (1988). Genetics of *Zoarces* populations XV. Genetic and morphological variation in Mariager Fjord. *Hereditas* 109: 99-112.
- Gill, T. (1863). On the gobioids of the eastern coast of the United States. *Proceedings of the Academy of Natural Sciences, Philadelphia* **5**: 267-271.
- King, K. J., Young, K.D., Waters, J.M. & Wallis, G.P. (2003). Preliminary genetic analysis of koaro (*Galaxias brevipinnis*) in New Zealand lakes: Evidence for allopatric differentiation among lakes but little population subdivision within lakes. *Journal of the Royal Society of New Zealand* 33(3): 591-600.
- Kocher, T.D., Thomas, W. K., Meyer, A., Edwards S.V., Pääbo, S. Villablanca F.X.,
   Wilson A.C. (1989). Dynamics of Mitochondrial DNA evolution in animals:
   Amplification and sequencing with conserved primers. *Proceedings of the Academy of Natural Sciences.* 86: 6196-6200.
- McDowall, R. M. (1975). A Revision of the New Zealand species of *Gobiomorphus* (Pisces: Eleotridae). *National Museum of New Zealand Records* 1(1): 1-32.

- McDowall, R. M. (1990). New Zealand freshwater fishes: A natural history and guide. Auckland, Heinemann Reed MAF Publishing group.
- McDowall, R.M. (1998). Driven by diadromy: its role in the historical and ecological biogeography of the New Zealand freshwater fish fauna. *Italian Journal of Zoology* 65, suppl: 73-85.
- McDowall, R. M. (2000). The Reed Field Guide to New Zealand Freshwater Fishes. Auckland, Reed Publishing.
- McDowall, R. M. (2002). Accumulating evidence for a dispersal biogeography of southern cool temperate freshwater fishes. *Journal of Biogeography* 29: 207-219.
- McDowall, R. M. (2004). Ancestry and amphidromy in island freshwater fish faunas. *Fish and Fisheries* **5**: 75-85.
- Orti G., Bell, M. A., Reimchen, T.E., Meyer, A. (1994). Global Survey of Mitochondrial DNA Sequences in the Threespine Stickleback: Evidence for Recent Migrations. *Evolution* 48(3): 608-622.
- Smith, P. J., McVeagh, S.M., Allibone, R. (2003). The Tarndale bully revisited with molecular markers: an ecophenotype of the common bully *Gobiomorphus cotidianus* (Pisces: Gobiidae). *Journal of the Royal Society of New Zealand* 33(3): 663-673.
- Smith, P. J. McVeagh, S.M., Allibone, R. (2005). Extensive genetic differentiation in Gobiomorphus breviceps from New Zealand. Journal of Fish Biology 67: 627-639.
- Thacker, C., Hardman, M.A. (2005). Molecular phylogeny of basal gobioid fishes:
   Rhyacichthyidae, Odontobutidae, Xenisthmidae, Eleotridae (Teleostei:
   Perciformes: Gobioidei). *Molecular Phylogenetics and Evolution* 37: 858-871.

- Thacker, C., Unmack, P.J. (2005). Phylogeny and Biogeography of the Eleotrid Genus Hypseleotris (Teleostei: Gobioidei: Eleotridae), with Redescription of H. cyprinoides. Records of the Australian Museum 57(1): 1-13.
- Vrijenhoek, R. C. (1998). Conservation genetics of freshwater fish. Journal of Fish Biology 53(Supplement A): 394-412.
- Waters, J. M., Lopez, J.A., Wallis, G.P. (2000). Molecular Phylogenetics and Biogeography of Galaxiid Fishes (Osteichthyes: Galaxiidae): Dispersal, Vicariance, and the Position of *Lepidogalaxias salamandroides*. Systematic Biology 49(4): 777-795.
- Waters, J. M., Burridge, C.P. (1999). Extreme intraspecific Mitochondrial DNA Sequence Divergence in *Galaxias maculatus* (Osteichthys: Galaxiidae). One of the World's Most Widespread Freshwater Fish. *Molecular Phylogenetics and Evolution* 11(1): 1-12.