

1                   **Missing pieces of the puzzle to effectively control Digital Dermatitis**

2

3           K Orsel<sup>1</sup>, P Plummer<sup>2,3</sup>, J Shearer<sup>3</sup>, J De Buck<sup>1</sup>, SD Carter<sup>4</sup>, R Guatteo<sup>5</sup>, and HW Barkema<sup>1</sup>

4

5   <sup>1</sup> Department of Production Animal Health, Faculty of Veterinary Medicine, University of  
6   Calgary, AB, Canada

7   <sup>2</sup> Department of Veterinary Diagnostic and Production Animal Medicine, Iowa State University  
8   College of Veterinary Medicine, Ames, IA, United States of America

9   <sup>3</sup> Department of Veterinary Microbiology and Preventative Medicine, Iowa State University  
10   College of Veterinary Medicine, Ames, IA, United States of America

11   <sup>4</sup> Department of Infection Biology, Institute of Veterinary Science, University of Liverpool,  
12   United Kingdom

13   <sup>5</sup> BIOEPAR, INRA, Oniris, La Chantrerie, 44307, Nantes, France

14

15 Summary

16 Since the first report of bovine digital dermatitis (DD) in 1974, there is a large body of literature  
17 published; however, effective prevention and control of the disease remain elusive. Although  
18 many aspects of the pathogenesis of DD have been investigated, even some of the most basic  
19 questions such as the etiology of this disease remain under debate. *Treponema* spp. have been  
20 strongly associated with DD lesions and occur in abundance in advanced lesions; however,  
21 efforts to induce disease with pure cultures of these organisms have been largely underwhelming  
22 and inconsistent. Furthermore, although the disease has been present for several decades, there is  
23 limited scientific evidence regarding effective treatment of DD. Apparent discrepancies between  
24 effectiveness *in vitro* and *in vivo* has challenged the scientific community to identify new  
25 potential treatment options. With no treatment resulting in a 100% cure rate, the current  
26 expectation is manageable control, but prospects for the eradication of the disease are unlikely  
27 using current approaches. In order to develop more effective approaches to control DD on-farm,  
28 there is a critical need for a deeper understanding regarding the causation, ecology, transmission  
29 and treatment of this disease. In this article, we attempt to provide insights into specific research  
30 needs related to DD in order to assist the industry, researchers, pharmaceutical companies and  
31 research sponsors with decision-making and identified research gaps.

32 Introduction to the disease

33 Digital dermatitis (DD), a skin disorder of the feet that mainly affects cattle, was first  
34 described in 1974 in Italy (Cheli and Mortellaro, 1974). It is characterized by an inflammatory  
35 dermatitis of the skin most commonly located at the plantar aspect of the interdigital cleft,  
36 although alternative locations have been reported (Holzhauer et al., 2008). A typical lesion is a  
37 circumscribed, moist ulcerative erosive area that is painful to the touch. The raw-red granular  
38 appearance of the lesion resulted in one of its alternative names (i.e. Strawberry foot rot),  
39 although the disease is also known as footwart, hairy heel warts, raspberry heel, verrucose  
40 dermatitis, Mortellaro's disease, and papillomatous DD. Notwithstanding, DD is likely the most  
41 accurate and commonly used term.

42 The most important clinical presentation of DD is lameness (Blowey and Sharp, 1988;  
43 Bassett et al., 1990; Read and Walker, 1998), although a significant number of affected cattle  
44 lack obvious clinical signs. Lesions are painful upon palpation and prone to bleeding after their  
45 surfaces are touched. Clinically, DD presents itself as a dynamic process with morphologically  
46 distinct stages. A variety of classification systems used to describe the stages of DD development  
47 have been described (Vink, 2006; Laven, 1999; Manske et al., 2002; Krull et al., 2014a), with the  
48 most widely adopted being the M-stage scoring system developed by Döpfer et al. (1997) and  
49 amended by Berry et al. (2012). This score identifies 5 categories where M0 is defined as  
50 normal digital skin with no evidence of dermatitis; M1 if a small (< 2 cm in diameter)  
51 circumscribed red to grey epithelial defect is present; M2 if an ulcerative active  $\geq 2$  cm in  
52 diameter with a red-grey surface; M3 (healing stage) after M2 lesion surface becomes firm and  
53 scar-like; M4 (chronic stage) if the lesion surface is raised with brown or black tissue,  
54 hyperkeratotic, scaly or proliferative; and M4.1 defined as small red circumscribed lesions

55 occurring within the boundaries of an existing M4 lesion (Berry et al., 2012; Döpfer et al., 1997).  
56 Consistency in scoring methodology would be much needed for scientific comparison of study  
57 results. A number of recent review articles have summarized the current understanding of the  
58 bacterial agents, epidemiology, therapy and treatment of digital dermatitis in detail in the last 2  
59 years (Evans et al., 2016; Palmer and O'Connell, 2015; Plummer and Krull, 2017; Wilson-  
60 Welder et al., 2015a). The goal of this manuscript as part of the DISCONTTOOLS collection, is to  
61 identify and discuss significant knowledge gaps that should be addressed by the research  
62 community in order to propel the field and to drive the development of novel and effective  
63 intervention strategies for controlling this disease.

64

#### 65 Significance

66 DD is a significant concern for cattle producers and veterinarians for several reasons. The  
67 clinical manifestation of lameness associated with DD poses a significant welfare concern for  
68 cattle and represents a leading cause of culling in the dairy cattle industry throughout the world  
69 (Cramer et al., 2009; Booth et al., 2004; Charfeddine and Perez-Cabal, 2017). However, the  
70 impact of DD is not restricted to clinical disease, but includes financial losses associated with the  
71 cost of treatment, decreases in both milk production and fertility, and losses due to increased  
72 culling even in the absence of clinical symptoms (Argaez-Rodriguez et al., 1997; Gomez et al.,  
73 2015b; Bruijnis et al., 2010; Cha et al., 2010; Relun et al., 2013).

74

#### 75 Geographical distribution

76 Digital dermatitis has been described as an endemic disease of dairy cattle in most parts  
77 of the world (van Amstel et al., 1995; Holzhauer et al., 2006; Rodriguez-Lainz et al., 1998; Wells

78 et al., 1999; Solano et al., 2016). In France, the PARABOV project aiming at describing the  
79 different lesions in cattle herds, reported that 16% of the feet and 70% of the herds were affected  
80 by DD lesions (Bleriot et al., 2013).

81           Given the differences in herd size, housing and management across these different  
82 geographic areas, it is safe to say that the disease is able to adapt and persist in a wide range of  
83 ecologic and management settings. In New Zealand, where the dairy industry has been  
84 historically pasture based, DD was reported only as sporadic cases until recent years when it has  
85 been implicated as a growing concern for non-healing lesions of the sole (Vermunt and Hill,  
86 2004; Van Andel M, 2012). The situation in New Zealand, as well as some other similar  
87 observations in other countries has led to the hypothesis that DD becomes an increasingly  
88 important issue when dairy cattle management changes from a more extensive pasture based  
89 system to confinement freestall housing (Sogstad et al., 2005). In countries like the UK, where  
90 cattle have housed and pasture seasons, the disease is almost restricted to the housing season  
91 (Evans et al., 2016). There is a need to further test this hypothesis in well-designed studies along  
92 with an effort to better understand the potential drivers of this disease progression. Herd stocking  
93 density, moisture content and hydration of the foot and skin, increased herd introductions and  
94 increased time on concrete have all been discussed and considered but there is at present little  
95 definitive evidence to support any sort of relative prioritization of these based on evidence based  
96 outcomes. It is important to acknowledge and recognize that emergence of the disease in  
97 countries and production systems, like the North American pasture-based ranching system, that  
98 have previously had little to no DD provide a rich research site for these critical studies to occur.  
99 We have to, however, realize that underreporting and the disease going unnoticed might be the

100 real reason for apparent freedom of disease. Once the disease becomes endemic, these studies  
101 become much more difficult, if not impossible, to test in anything other than a simulated system.

102

### 103 Pathogens involved

104         Despite a significant number of studies focused on elucidating the etiology of DD, debate  
105 remains regarding the exact etiology. Although fungal and viral etiologies have been considered,  
106 the scientific community has largely agreed that these organisms are less likely to drive the  
107 disease process, and the field has focused its attention on bacterial organisms (Rebhun et al.,  
108 1980; Krull et al., 2014b; Zinicola et al., 2015; Brandt et al., 2011). For a detailed overview of  
109 the findings of this body of knowledge, readers are directed to the review articles referenced at  
110 the start of this manuscript; however, two consistent themes have emerged from these  
111 studies. First, DD lesions are consistently associated with an abundant and diverse population of  
112 multiple species of Treponemes (Zinicola et al., 2015; Krull et al., 2014b; Evans et al.,  
113 2016). Second, these diverse treponeme populations exist as a portion of a much more diverse  
114 and complex bacterial community that comprises the total microbiota of the DD lesions.  
115 Furthermore, the non-treponemal constituents of the microbiota are not random and instead show  
116 association with the stage of lesion development (Krull et al., 2014b, Zinicola et al., 2015). As  
117 described in more detail by Krull et al. (2014b), non-affected animals showed an abundance of  
118 *Staphylococcaceae*, *Streptococcaceae*, *Bacteroidaceae*, *Corynebacteriaceae* and  
119 *Pasteurellaceae*, replaced by other bacterial families as lesions progressed. Whereas  
120 *Spirochaetaceae* increased systematically from 0 to over 90% in chronic stages of the disease  
121 (Krull et al., 2014c). With lesions classified as active and inactive, Zinicola et al. (2015)  
122 identified *Firmicutes* and *Actinobacteria* as the predominant bacterial phyla of control animals,

123 and *Spirochetes*, *Bacteroidetes* and *Proteobacteria* as highly abundant in DD-affected animals.

124         These themes are consistent with the vast majority of the published literature on the topic  
125 and can be agreed upon by most researchers in the field. Herein, however, lies a remaining  
126 uncertainty regarding the etiologic role that each of these organisms plays in the molecular  
127 mechanisms responsible for the development of DD. We will address the research needs related  
128 to etiology in three broad areas related to 1) the role of the treponemes, 2) the role of other  
129 bacterial members in the community, and 3) the role of the interaction between the community  
130 members.

131         First, while it is clear that *Treponema* spp. are consistently present in DD lesions and  
132 make up the majority of the bacterial community in advanced lesions, it is also clear that these  
133 populations represent a diversity of species instead of a single species (Klitgaard et al., 2013;  
134 Marcatili et al., 2016; Krull et al., 2014c; Yano et al., 2009; Evans et al., 2008). This in itself  
135 poses a problem with fulfilling Koch's postulates for this disease process. At a very minimum,  
136 one must acknowledge that if treponemes are the primary etiologic agents associated with DD, it  
137 is a polytreponemal process, and this hypothesis has been argued for in the literature (Evans et  
138 al., 2008). If this hypothesis is true, it still leaves the significant question of why does the disease  
139 require the presence of multiple treponemal species instead of one? Furthermore, how do these  
140 different treponemal species interact with each other, and what is the minimum treponema  
141 consortium required for inducing clinical disease? How does the polytreponemal community  
142 change during progression of the disease? An alternate hypothesis that emerges is that the  
143 diversity of *Treponema* species present in the lesions is more suggestive of an overgrowth of  
144 opportunists that find a unique niche for expansion during the induction of DD lesions (Edwards  
145 et al., 2003; Krull et al., 2014b; Wilson-Welder et al., 2015a). Indeed, there is now much

146 evidence that the DD-associated treponemes are promiscuous opportunistic invaders of  
147 established skin lesions, particularly on feet (Evans et al., 2011), other limb skin tissues (Clegg et  
148 al., 2016a) and have been identified in a particularly virulent udder disease, ischaemic teat  
149 necrosis (Clegg et al., 2016b). This opportunistic nature of treponeme tissue invasion may also  
150 account for their strong associations with DD lesions in UK sheep (Dhawi et al., 2005) and goats  
151 (Sullivan et al., 2015b), skin lesions in UK pigs (Clegg et al., 2016d), and foot lesions in US wild  
152 elk (Clegg et al., 2015). While the morphologic appearance of DD lesions is essentially identical  
153 in beef cattle compared to dairy cattle, we have very limited information regarding the bacterial  
154 communities present in beef cattle DD and how it compares to that of dairy lesions. When beef  
155 cattle DD lesions were analyzed by PCR for the DD-associated *Treponema* spp., and also for  
156 *Dichelobacter nodosus* and *Fusobacterium necrophorum*, Sullivan et al. reported that at least 1  
157 of the known *Treponema* phylogroups associated with DD was present in all beef cattle DD  
158 lesions (Sullivan et al., 2015a). This sudden emergence of new clinical phenotypes associated  
159 with these specific bacteria is suggestive of genomic changes affecting treponeme physiology  
160 and ability to transmit between tissues, animals and even species. As such, there is a need for  
161 vigilance in case of further spread leading to new clinical phenotypes. Whether these are primary  
162 or secondary infections, the treponemes represent an important bacterial community for which  
163 there is need to better understand their physiology and ecology in lesions. In the current era of  
164 bacterial genomics there is a significant need for the identification of “type strains” for each of  
165 the species and for full genome sequencing of isolates from each of these strains. These  
166 resources would allow for the continued development and refinement of research methodologies  
167 focused on better evaluating the role that these organisms play in each stage of lesion  
168 development and any significant interactions with other bacterial species. Genome sequences



169 also allow for more informed generation of hypotheses related to the virulence and ecologic  
170 adaptation abilities that each strain possesses and how these functions interact in a central disease  
171 process. Currently, large scale genomic analyses are hampered by culture techniques struggling  
172 to isolate pure single species cultures with consistency and representing all *Treponema* species  
173 that have been demonstrated in DD lesions by metagenomic studies (Krull et al., 2014c; Zinicola  
174 et al., 2015).

175         Second, as alluded to above, constituents of the non-treponemal bacterial communities  
176 that are present in the DD lesions vary by lesion stage, but are amazingly consistent within a  
177 given stage of lesion development (Krull et al., 2014c; Zinicola et al., 2015). This finding  
178 suggests that their presence is not merely coincidental or due to background from the dairy  
179 environment, but instead suggests that there is a driving force behind the development and  
180 transition of this complex microbiota shift. There is a clear need to better understand what is  
181 driving this transition and how this transition is involved in the development, maintenance and  
182 response to therapy of digital dermatitis. Given that several of these organisms are known  
183 pathogens in other disease processes of the foot of ruminants (for example, *Dichelobacter*  
184 *nodosus*, *Fusobacterium necrophorum* and others) it is important that hypotheses are developed  
185 and tested regarding their specific role in DD. Interestingly, many of these “known” pathogens  
186 are present in low relative abundance and this fact has been used to argue that they may not be  
187 relevant to the disease process (Moe et al., 2010; Collighan and Woodward, 1997). However,  
188 recent evidence from other disease processes has demonstrated that relative abundance in  
189 phylogenomic studies needs to be interpreted with caution. This is particularly important because  
190 abundance is not necessarily commensurate with pathogenicity. Neither does it controvert or  
191 confirm etiology. For example, recent metagenomic data derived from ovine footrot, a disease

192 process with a well-known and Koch's postulates confirmed etiology of *Dichelobacter nodosus*,  
193 demonstrated that the relative abundance of that organism was between 0.5-1.9% in active  
194 lesions (Maboni et al., 2017). In contrast and as a reference point, the relative abundance of  
195 *Treponema* spp. in those same samples of ovine footrot averaged 14%. In order to address these  
196 issues and research needs, there is a need for additional genomic information and the  
197 identification of type strains for these non-treponemal species associated with DD lesions. In  
198 addition, the sensitivity to detect low abundant species involved in the pathogenicity of DD  
199 lesions needs to be increased.

200 Not surprisingly, the third area of research needs related to the etiology of DD, focuses  
201 on the interface of the two issues discussed above. The literature suggests that in other  
202 treponeme-associated diseases, such as periodontal disease in humans, the association of  
203 treponemes and other organisms extends beyond simply co-isolation and is associated with direct  
204 molecular interaction or nutritional symbiosis of the organisms (Grenier, 1992b; Grenier, 1992a;  
205 Hashimoto et al., 2003; Ito et al., 2010; Nilius et al., 1993; Simonson et al., 1992; Yao et al.,  
206 1996). Despite the fact that these organisms are very closely genetically related to the species  
207 found in DD, these types of interactions have not yet been addressed in DD research. Likewise,  
208 we must also consider the possibility that regardless of potential interaction between the bacterial  
209 species themselves, the presence of these multiple species could impact the immune response of  
210 the host, particularly by polyclonal activation of the lymphoid system and induction of  
211 immunological dysregulation (Montes et al., 2007). Alternatively, expression of virulence factors  
212 such as proteases or leukotoxins by some organisms may alter the ecological adaptation and  
213 virulence potential of other organisms in the same niche (Smalley and Olczak, 2017; Lohinai et  
214 al., 2015; Castro et al., 2017). Although these interactions have the potential to be extremely

215 complex and time consuming to study, it is likely that this broader systems approach to the  
216 complex pathobiology of DD holds potential for more fully understanding the mechanisms and  
217 roles that each of these organisms may play in the disease process. Without a clear understanding  
218 of DD etiology, development of effective vaccines for disease control as well as targeted  
219 treatments could be hampered.

220

### 221 The hosts

222 In contrast to an almost 40-year history of recognition of the importance of DD in dairy  
223 cattle, DD in beef cattle has been emerging as an increasingly recognized disease in recent years.  
224 After an initial case report from the UK (Sullivan et al., 2013), there have been several reports of  
225 DD in the North American feedlot industry (Campbell, 2014; Orsel and Schwartzkopf-  
226 Genswein, 2015). Deeper exploration of the literature suggests that DD-like lesions have been  
227 recognized in the US in beef cattle even prior to their description in dairy cattle, which may point  
228 to the potential for the disease being unrecognized (Lindley, 1974; Barthold et al., 1974). A  
229 number of questions still remain and deserve attention with regards to the growing importance of  
230 DD in beef cattle worldwide. Additional questions remain regarding what epidemiologic,  
231 environmental and management factors and changes are driving the recent emergence of DD as a  
232 recognized disease of feedlot cattle. Further efforts to understand how the disease differs from  
233 that of dairy cattle, and what knowledge can be gained from comparison of this disease across  
234 these very divergent management systems may prove fruitful in improving our understanding of  
235 the disease in both systems.

236 It has become increasingly apparent that other mammalian species, including small  
237 ruminants (sheep and goat) and wildlife (e.g. elk) can be affected with lesions of the hoof and

238 skin that have significant similarities to DD (Duncan et al., 2014; Clegg et al., 2015; Han and  
239 Mansfield, 2014; Crosby-Durrani et al., 2016). Interestingly, despite the presence of very similar  
240 organisms being isolated from these various hosts, the clinical manifestations of these diseases  
241 vary across the hosts as was eluded to before. For instance, classic bovine DD lesions are  
242 confined to the skin (hence the term dermatitis), although in cattle with DD, severe horn heel  
243 erosion are 46% more commonly reported (Gomez et al., 2015a). When treponemes are  
244 associated with non-healing sole lesions in cattle, it is primarily believed to be the result of  
245 secondary infection of pre-existing sole lesions such as sole ulcers, white line disease, toe  
246 necrosis and puncture wounds (Clegg et al., 2016a; Clegg et al., 2016c; Clegg et al., 2016d). In  
247 contrast, contagious ovine digital dermatitis, treponeme associated hoof lesions in dairy goats  
248 (Crosby-Durrani et al., 2016; Sullivan et al., 2015b) and treponeme associated hoof lesions in elk  
249 (Clegg et al., 2015; Han and Mansfield, 2014) typically present with dermatitis along with under  
250 running of the sole, and in severe cases complete avulsion of the hoof capsule. The propensity  
251 for development of these primary sole lesions in these host species raises questions regarding the  
252 difference in disease manifestation based on the host. Potential hypotheses include: 1) intrinsic  
253 differences in the host anatomy or genetics allows for differences in disease manifestation, 2)  
254 despite similarities in the treponemal species isolated, the clones involved in these diseases differ  
255 in their genetics or virulence attributes, and 3) the presence of the treponemes in these cases is  
256 more of an opportunistic infection with other organisms in the bacterial consortium driving the  
257 lesion pathogenesis. These differences in host response to the organisms along with the  
258 development of disease induction models in both cattle (Gomez et al., 2012; Krull et al., 2016a)  
259 and sheep (Wilson-Welder et al., 2015b) provide a good foundation for experimental approaches  
260 designed to address and test these hypotheses. By utilizing similar inoculums in both species and

261 observing the differences in clinical disease combined with multi-omic approaches, we can start  
262 to dissect the importance of host differences in the disease process.

263 The role of host genetics in DD lesion susceptibility has also been evaluated and has  
264 clearly demonstrated a genetic role for disease susceptibility or resistance (Scholey et al., 2012;  
265 Schopke et al., 2015). In addition, genetic parameters and breeding values have been identified  
266 for most hoof lesions and their relationships with feet and leg traits (Chapinal et al., 2013). With  
267 large variations in sire estimated breeding value for resistance to hoof lesions, the authors  
268 concluded there were long-term opportunities for genetic selection. Further research is required  
269 to determine the influence of susceptibility factors, identify the genetic basis of variation, clarify  
270 heritability of DD susceptibility and determine how host-related factors are correlated with  
271 production and health traits currently used in breeding programs (Palmer and O'Connell, 2015).

272

### 273 Immune responses to infection

274 Local dermal tissue and inflammatory response to DD infection has been evaluated using  
275 several approaches. There is a general dermal thickening in lesion development that is  
276 accompanied by varying degrees of infiltration of inflammatory cells (neutrophils and  
277 eosinophils) and changes in local cytokine concentrations (Refaai et al., 2013). Similarly, gene  
278 expression in skin biopsies from 5 bovine DD lesions and 5 healthy bovine feet were compared  
279 using RNA-Seq technology (Scholey et al., 2013). They demonstrated changes in cytokine  
280 expression (especially interleukin 1 $\beta$  being upregulated in DD lesions) and changes in expression  
281 of several other keratin or keratin associated genes. Interestingly, they detected evidence of poor  
282 local immune and inflammatory reactions to the bacterial infection present in lesions, possibly  
283 indicating a suppressed host response to DD. It has been speculated that local innate immune

284 responses may contribute to the proliferative, inflammatory conditions that perpetuate DD  
285 lesions (Wilson-Welder et al., 2015a).

286         In general, there is a limited body of knowledge in the literature regarding host innate or  
287 adaptive immune responses to DD infection. Several studies have evaluated the systemic  
288 humoral immune response of cattle and have consistently demonstrated that, despite the  
289 restricted presentations of clinical signs, systemic immune responses to treponemal antigens and  
290 some other DD-associated organisms can be identified using serology (Demirkan et al., 1999;  
291 Gomez et al., 2014a; Vink et al., 2009). However, use of these assays has not been widely  
292 implemented in diagnostic or prognostic studies, in large part due to uncertainty regarding how  
293 to utilize the outputs to effectively monitor disease in the farm. In large part, this lack of clear  
294 diagnostic serology is considered to be due to the endemic nature of disease and persistence of  
295 the DD-associated treponemes in farm environments, rendering most animals seropositive to one  
296 degree or another. Even less is known about the cell-mediated immune responses to DD and their  
297 role, if any, in disease. Future studies that evaluate both arms (humoral and cell mediated) of the  
298 immune response are warranted and have the potential to provide insights important for disease  
299 control and lesion healing. Field experience demonstrates that the majority of cattle do not  
300 develop a protective immune response that results in spontaneous lesion healing, although  
301 spontaneous healing of M1 and M2 lesions has been described (Relun et al., 2012). Efforts to  
302 compare the “typical” immune response of cattle with active DD lesions, to those of cattle that  
303 are able to clear the lesions (either spontaneously or following treatment) may provide insights  
304 into specific immune responses that are beneficial. Furthermore, these efforts need to extend  
305 across a diversity of DD-associated organisms (including multiple species of treponemes). It is  
306 likely that the greatest return on investment related to continued efforts to understand DD

307 immune responses focuses on improving our understanding of the antigenic targets, whether a  
308 TH1 or TH2 immune response predominates and which is most likely to be protective. All of the  
309 above will be essential information to boost immunity, possibly by enabling development of an  
310 effective vaccine.

311

### 312 Transmission

313 Although the exact route of transmission for DD is not fully elucidated, DD presents itself as a  
314 highly infectious disease, consistent with the experimental model of Krull et al. (2016a), in  
315 which the negative controls could be infected by being comingled with experimentally infected  
316 animals despite the feet of both animals being completely wrapped in bandages for the duration  
317 of the study. Another experimental model was used by the Liverpool research team, using sheep  
318 affected with DD lesions to induce DD in healthy animals by just mixing and intermingling in a  
319 normal farm environment with standard herd management and then chronic lesion development  
320 over time (SD Carter, personal communication). This attempt at an infection model resulted in  
321 over 50% of the naïve sheep developing contagious ovine digital dermatitis lesions, with the full  
322 range of severity, from small lesions to complete hoof evulsion requiring euthanasia (SD Carter,  
323 personal communication). The outcomes of these studies clearly demonstrate that transmission  
324 can occur when susceptible animals are housed in the same environment as those with active DD  
325 lesions. However, the fact that transmission occurred in the presence of foot wraps could suggest  
326 that direct physical contact with lesions is not required (Krull et al., 2016a). The literature has  
327 also evaluated the role that early or active host-associated DD lesions play as a primary reservoir  
328 of infectious organisms. Multiple studies have demonstrated that the quantitative levels of DD-  
329 associated treponemes are higher in host-associated tissues (including rectum, gingiva, rumen,

330 DD lesions) than in environmental samples collected from dairy environments (Evans et al.,  
331 2012b; Klitgaard et al., 2017; Rock et al., 2015). However, low numbers of DD-associated  
332 *Treponema* spp can be identified in dairy farm slurry on farms with endemic DD when using  
333 deep sequencing based phylogenomic approaches (Rock et al., 2015; Klitgaard et al., 2017).  
334 Likewise, there is evidence from multiple groups that foot trimming equipment can be  
335 contaminated with treponemes and may act as a source of infection between animals and farms  
336 (Sullivan et al., 2014; Rock et al., 2015). While there is a growing body of evidence that  
337 treponemes can be identified in samples beyond active DD lesions, the relative role of these  
338 sources as primary reservoirs of infection remains unclear. It is possible that these organisms are  
339 simply transient members of the bacterial community that are continuously shed in the  
340 environment from lesions but survive for very short periods; a hypothesis that may be more  
341 likely given the apparent affinity of treponemes for host environments. Alternatively, it is  
342 possible that the organisms are able to survive off the host for sufficient periods of time to allow  
343 disease transmission. Consequently, there is a need to better understand how these organisms  
344 adapt to the non-host environment and how long they are able to persist in the absence of host  
345 tissue and nutrients. Further complicating the issue of reservoirs of infection is the complex  
346 etiology (either polytreponemal or polybacterial) of the disease process, which results in a  
347 situation where one must potentially consider reservoirs for each of the species and the fact that  
348 there is potential that those could be different. The work thus far has focused on reservoirs of  
349 treponemes due to their known association with the disease process; however, this may be an  
350 over simplification.

351 Other routes of fomite-associated transmission should be considered, including contact  
352 with contaminated equipment, as *Treponema* spp. has been identified on hoof knives and other



353 trimming equipment (Sullivan et al., 2014; Rock et al., 2015). Transmission through insect  
354 vectors is not likely, as no vectors tested for presence of *Treponema* spp. DNA were positive  
355 (Evans et al., 2012b). However, it is reported that in a portion of dairy farms, non-lactating  
356 heifers are also affected by DD (Jacobs et al., 2017; Holzhauser et al., 2012). If undetected and  
357 untreated these animals are a continuous source of DD-affected animals for the lactating herd. It  
358 is not clear though what portion of the prevalence of DD in adult cows can be attributed to young  
359 stock entering the lactating herd after calving. There is a need for significant effort related to  
360 better understanding the relative importance of all of these potential routes of transmission on the  
361 overall epidemiology of this disease on dairy farms. Efforts in this area should consider the  
362 potential for a multi-species etiology and need to evaluate the ecologic fitness and survivability  
363 of these organisms in non-host environments. With limited knowledge regarding the key  
364 reservoir of the *Treponema* phylogroups and the role of other bacteria in pathogenesis as well as  
365 uncertainty about route of transmission, control of DD could well be hampered.

366

### 367 Experimental models

368 Robust and efficient experimental models of infection are critical to research efforts  
369 focused on better understanding the pathogenesis and etiology of DD. Several induction models  
370 have been described for use in the induction of DD lesions in both cattle and sheep (Gomez et  
371 al., 2012; Krull et al., 2016a; Wilson-Welder et al., 2015b). The most obvious benefit of an  
372 experimental model would be to evaluate the etiology of the disease; however, efforts to use the  
373 models in this manner have thus far been underwhelming. Both bovine models have attempted to  
374 induce lesions using pure culture of DD-associated *Treponema phagedenis*-like bacteria (Gomez  
375 et al., 2012; Krull et al., 2016a). While both studies observed some degree of lesion formation,

376 the size and severity of the lesions was considerably less than observed when macerated lesion  
377 material was used as the inoculum (Gomez et al., 2012). Additionally, in both studies,  
378 inoculations of pure growth treponeme isolates were performed on one foot of animals that had  
379 macerate used to induce lesions on another foot, meaning that while the one foot was only  
380 exposed to a single organism there were other organisms used in the pen and even on the same  
381 animal. This design is particularly problematic to the interpretation of the data with regards to  
382 etiology because one of the studies showed that negative control animals (i.e. animals that had  
383 their feet wrapped and inoculated with media alone) housed in the pens with animals that were  
384 induced with macerate had an induction rate and lesion severity essentially identical to those  
385 induced with pure growth organisms, whereas negative control animals that were housed in  
386 isolation remained uninfected (Krull et al., 2016a). Knowing this information, along with the  
387 experience gained in these studies, allows for the development of more robust study designs that  
388 can be effectively used to further probe the question of etiology. Considerations that need to be  
389 included in that approach include animal housing with regards to cross contamination, use of  
390 pure cultures of single organisms versus consortia of multiple pure growth organisms, the role of  
391 individual animal immunity, and the potential confounders of pre-existing immunity in animals  
392 sourced from an industry that has high endemic rates of disease and consequently a high risk of  
393 previous exposure to the disease.

394 Experimental induction models also represent a useful tool for evaluating a variety of  
395 other important issues. These include but are not limited to, experimental approaches focused on  
396 adaptive immune responses (both humoral and cell mediated), therapeutic interventions, and  
397 vaccine evaluation and development. The availability of multiple induction models allows  
398 researchers to determine which models best test their hypothesis while providing the needed

399 controls. A significant downside of current bovine models is that they tend to be quite expensive  
400 and labor intensive, so the development of a small ruminant model provides some potential cost  
401 benefits while allowing for comparison across species as described in the host portion of this  
402 manuscript.

403

#### 404 Lesion detection

405 Key to any DD control program is the efficient and consistent identification of lesions.  
406 Given a relatively distinct clinical presentation of the disease, diagnosis of DD is usually based  
407 on visual inspection of the foot. This process can be labor-intensive, and since the location of the  
408 lesion is not always easily accessed, small lesions can be easily missed (Solano et al., 2017a).  
409 Most commonly, animals are inspected in a chute that allows for safe lifting of the foot and  
410 thorough cleaning before inspection and this method of evaluation is considered the gold  
411 standard for diagnosis. To facilitate a more efficient and less labor-intensive inspection  
412 alternative means of observation in the parlor, headlocks and alleyways have been systematically  
413 compared to chute observations (Stokes et al., 2012; Winders et al., 2015; Solano et al., 2017a;  
414 Relun et al., 2011), also in young stock using pen walks (Jacobs et al., 2017). The consensus of  
415 these studies is that the highest agreement between chute and alternate observation methods  
416 occurs when the lesion status is condensed to a dichotomous presence or absence. In this  
417 situation sensitivity of lesion detection ranged from 65-100% while specificity ranged from 80-  
418 99% (Stokes et al., 2012; Winders et al., 2015; Solano et al., 2017a). When efforts are made to  
419 evaluate more precise lesion characteristics (color, erosiveness, proliferation) or score the lesions  
420 on a standardized severity scoring system the sensitivity and specificities consistently decrease to  
421 a slight to moderate level of agreement with chute evaluation (Relun et al., 2011; Winders et al.,

422 2015; Solano et al., 2017a). The presence of DD lesions at sites in the interdigital space or dorsal  
423 aspect of the foot further drops sensitivity. As might be expected, parlor observation of washed  
424 feet performed better than headlocks and pen, with pen observation showing the lowest  
425 sensitivity and specificity (Winders et al., 2015). Therefore, although DD scoring in the milking  
426 parlor as a routine practice should facilitate early detection, prompt treatment interventions, and  
427 herd monitoring, it was not sufficiently reliable to replace definitive identification of lesions  
428 done in the trimming chute. In addition, it is noteworthy that milking parlor scoring has not been  
429 implemented as a routine method of DD diagnostics and alternatives should be developed for  
430 early disease detection in automated milking systems.

431         Alternatively, detection of cows affected with DD could focus on detection of lameness.  
432 However, not all stages of DD result in visible lameness, and conversely, not all lameness results  
433 from DD. The use of locomotion score was very inconsistent in its ability to accurately identify  
434 cows with DD (Krull et al., 2016b). In fact, cows with the most severe changes in locomotion  
435 score were more likely to have other claw-horn lesions than DD, and the majority of cattle with  
436 DD failed to show high locomotion scores. These findings are consistent with the findings of  
437 Frankena et al. (2009) in which only 39% of the cows with severe DD lesions showed lameness .  
438 Therefore, DD detection is still either labor intensive as feet need to be lifted or only low to  
439 moderately sensitive based on simplified assessment methodologies. Notwithstanding, an overall  
440 lameness control program would facilitate identification of cows that need individual attention.  
441 Given that the primary welfare concern associated with DD involves induction of lameness, the  
442 field would benefit from a better understanding of the drivers of lameness as it relates to DD  
443 lesions. Clearly, the presence of a lesion alone is probably not sufficient to induce lameness,  
444 despite the fact that the lesions are universally sensitive to pressure. Likewise, the fact that

445 lameness typically improves markedly within several days following topical treatment suggest  
446 that the underlying mechanisms of pain can be minimized even in the presence of unhealed skin.

447

#### 448 Treatment

449         Given the endemic nature of DD, many field studies have been performed to identify  
450 effective treatments. With the most commonly accepted pathogenesis being based on a bacterial  
451 origin, treatments have focused on this aspect of the disease. Treatment with systemic penicillin  
452 has been shown to be efficacious but is not widely used due to the necessity of withholding milk  
453 and costs (Laven and Logue, 2006). Systemic antibiotic therapy with other antibiotics routinely  
454 used in US dairy cattle milking herds did not increase or decrease DD lesion scores (Krull et al.,  
455 2016b), and due to cost, is rarely used (Laven and Logue, 2006). Conversely, topical treatment,  
456 usually with antibiotic preparations, is the most common method employed by veterinarians and  
457 foot trimmers for the treatment of advanced lesions (Apley, 2015). There is still uncertainty and  
458 disagreement regarding the actual efficacy of treatment outcomes with topical therapy. Success  
459 rates as low as 9% and as high as 73% have been reported (Krull et al., 2016b; Cutler et al.,  
460 2013; Berry et al., 2010; Nishikawa and Taguchi, 2008; Shearer and Hernandez, 2000; Laven  
461 and Hunt, 2001). There is a pressing need for good comparative field studies using robust study  
462 designs (ideally prospective randomized controlled trials) to determine the most efficacious  
463 treatment approach. Design of these studies needs to consider and normalize the stage of lesions  
464 development, as the treatment response may vary by lesion severity. Likewise, prolonged  
465 durations of post treatment observation (upwards of 120 days) are required to confirm that  
466 lesions fully heal and do not recrudescence (Krull et al., 2016b), while shorter observation periods  
467 may allow for observation of improvement of lameness.

468           In order to evaluate a larger diversity of antibiotics and to address the issue of potential  
469 antibiotic resistance, several DD treponeme studies have used *in vitro* minimum inhibitory  
470 concentration (MIC) based approaches (Hartshorn et al., 2013; Evans et al., 2009; Evans et al.,  
471 2012a). However, it is important to recognize that the Clinical and Laboratory Standards Institute  
472 (CLSI) does not have a validated methodology or bacterial MIC cut-off points established for  
473 DD-associated bacteria. This consequently complicates clinical interpretation and utility of *in*  
474 *vitro* derived MIC data and represents an area where additional research and the development of  
475 validated cut-off points could benefit the field. Caution should be exercised when interpreting the  
476 outcomes of *in vitro* MIC data, since the pharmacokinetic and pharmacodynamic differences  
477 between drugs can greatly influence the dosage of the drug delivered to the lesion. As a result,  
478 simply comparing which drug has the lowest MIC fails to address the clinical complexity of  
479 treatment efficacy and pharmacology. For instance, topical administration of several grams of  
480 oxytetracycline directly to a lesion may result in local drug concentrations far above an MIC that  
481 could not be achieved in the same location using systemic administration. Continued efforts to  
482 better understand the potential presence of antibiotic resistance should focus on identification of  
483 genetic resistance determinants to important classes of antibiotics used in DD control. Likewise,  
484 evaluation of genetic mechanisms of resistance to heavy metals (such as copper commonly used  
485 in footbaths) is warranted.

486           The potential for various morphotypes of *Treponema* spp. has been raised as an  
487 explanation for the discrepancy of *in vitro* susceptibilities and limited effectiveness *in vivo*.  
488 During *in vitro* growth of *Treponema* spp. isolated from DD, morphological variability was  
489 observed (Döpfer et al., 2012), indicating the presence of a spiral form and a round body form.  
490 The round body forms are morphologically similar to those observed in *Borrelia burgdorferi* (a

491 related spirochete), and have been hypothesized to play a role in persistent infection as has been  
492 hypothesized for *Borrelia* (Murgia and Cinco, 2004). Additional work to fully demonstrate the  
493 roll of these morphologically variable cells in *in vivo* infections is needed, as the role of these  
494 forms in chronic Lyme disease is hotly debated (Merilainen et al., 2016; Murgia and Cinco,  
495 2004; Merilainen et al., 2015; Lantos et al., 2014). To date, very little information is available in  
496 the peer-reviewed literature that definitively identifies and details their presence in the tissue of  
497 DD lesions. Efforts to understand the biochemical and genetic drivers of cellular morphology  
498 change along with improving our understanding of the metabolic activity of these cells would aid  
499 in understanding their importance. Likewise, efforts to definitively demonstrate their significance  
500 in active lesions and the underlying molecular mechanisms related to the potential for their role  
501 in persistence of disease may allow for the identification of novel control targets for this endemic  
502 disease.

503         Due to global concerns regarding prudent antibiotic use, and the inconsistent response of  
504 DD lesions to antibiotic treatment, alternative approaches to the use of antimicrobials for control  
505 of DD are desired and have been considered. For example, the impact of altered trace mineral  
506 nutrition was evaluated in a randomized efficacy study to evaluate the effect of a premix  
507 containing concentrations of organic trace minerals and iodine (HOTMI). This study showed a  
508 reduction in the incidence of active DD lesions acquired naturally or induced by an experimental  
509 infection challenge model (Gomez et al., 2014b). The mineral premix tended to reduce the total  
510 DD infection rate and the average size of the experimentally induced lesions, although the results  
511 failed to reach the level of statistical significance. Additional work utilizing larger sample sizes  
512 are warranted to determine if the effect is real. Likewise, the mechanistic reasons for the  
513 improvement should be thoroughly evaluated in order to provide insights into the cellular

514 pathways that benefit lesion prevention. There is also a need for an improved understanding of  
515 the broader role of nutrition in DD prevention.

516

#### 517 Prevention and control

518 As reported by Potterton et al. (2012), between 2000-2011, 62 scientific papers could be  
519 identified focusing on prevention of digital dermatitis, with the seven distinct areas of interest  
520 being, standing time on concrete, claw trauma, diets and feeding, detection and treatment, heifer  
521 breeding, environmental hygiene and biosecurity. In more detail Holzhauser et al. (2012) reported  
522 the importance of prevention of transmission of disease to young stock as housed on the same  
523 farm. With DD having high within-herd prevalence, herd-level interventions are warranted to try  
524 to decrease the prevalence.

#### 525 Footbaths

526 The most commonly used herd-level intervention is a footbath, primarily used to prevent  
527 new cases through increased hygiene, but sometimes perceived important for treatment of  
528 clinical cases. Proper footbath design has been evaluated and is based on dimensions (Logue et  
529 al., 2012; Cook et al., 2012), frequency of use, product used and appropriate concentration of  
530 solution (Speijers et al., 2010; Speijers et al., 2012; Teixeira et al., 2010; Relun et al., 2012).  
531 When used, the footbath must be managed to ensure sufficient solution is consistently available  
532 to achieve full immersions of hooves of all 4 feet (Cook et al., 2012). Furthermore, fecal  
533 contamination is known to interfere with effectiveness of most footbath solutions. With copper  
534 sulphate, a common choice in North America, the pH of the concentration is critical to keep  
535 copper soluble and efficacious (Laven and Hunt, 2002; Speijers et al., 2010; Speijers et al., 2012;  
536 Teixeira et al., 2010). Optimizing footbath management according to scientific knowledge



537 reduces the prevalence of active DD lesions. On farms where footbathing practices do not meet  
538 recommendations, an automatic footbath may provide benefit (Solano et al., 2017b). With most  
539 footbath products having adverse legislative, health and safety and environmental effects, *in vitro*  
540 models have been developed to screen new footbath products. The assays designed allow for  
541 determination of minimum inhibitory concentration and minimum bactericidal concentration of  
542 disinfectants for *Treponema* spp. Additionally, manure contamination, potentially resulting in  
543 inhibition of the solution, was also mimicked. This assay was useful to categorize disinfectants,  
544 based on effects of exposure and manure concentration regarding their ability to inhibit  
545 *Treponema* spp. growth (Hartshorn et al., 2013). Despite the large body of literature, no footbath  
546 studies had acceptable efficacy in control of DD.

547         Questions have been asked about the safety for human and environmental health as  
548 related to large quantities of chemicals and minerals being used for footbaths (Laven and Logue,  
549 2006). In Canada, there is a wide variety of products in numerous combinations as well as  
550 concentrations (Solano et al., 2015). Although risks to human health due to formaldehyde have  
551 been explored (Doane and Sarenbo, 2014), it was concluded to not exceed public health  
552 guidelines. Based on frequent questions regarding antimicrobial use, environmental and health  
553 impacts, future directions should focus on early interventions and potential use of  
554 environmentally friendly products.

#### 555 Control

556         Monitoring herds with endemic disease for changes in lesion prevalence or severity and  
557 classifying cattle based on lesion monitoring has been described as one means to provide insights  
558 into on-farm management decisions making. These approaches allow producers to potentially  
559 identify higher risk animals that might need intervention or culling. The goal of this approach is

560 to achieve a manageable state of disease, but no strategy was identified to eradicate DD (Dopfer,  
561 2009). While DD eradication at herd or even country level would be the ideal objective, the  
562 literature suggests that in most cases this is extremely difficult if not impossible given the current  
563 tools available and the global nature of this disease. The combination of biosecurity, various  
564 footbaths and antimicrobials has patently not been effective in preventing disease spread or  
565 reduce severity. Consequently, we need an approach that takes a different line and preferably has  
566 more potential for prevention and control. Efforts to develop vaccines that were effective in  
567 limiting disease prevalence or severity would have significant economic and welfare benefit for  
568 the industry. The development of effective vaccines for the control of similar disease processes,  
569 such as ovine footrot, gives hope that one day these might be an option. The current research  
570 gaps identified in this manuscript, including an uncertain and complex etiology, minimal  
571 understanding of the disease transmission dynamics the significant lack of knowledge regarding  
572 the nature of protective immunity of this disease will provide challenges for vaccine  
573 development efforts in the short-term. However, we are rapidly developing a better  
574 understanding of the infective nature of DD and post-genomic technologies, such as reverse  
575 vaccinology offer hope that vaccine candidates, based on treponeme genomes, may be developed  
576 in the near future.

577

#### 578 Role of the dairy producer in control of digital dermatitis

579         There is considerable variation in producers' mindsets towards an issue like DD on their  
580 farm, leading to variation in behaviors to address DD (Garforth, 2012). The perception of risk in  
581 general for example, can vary greatly based on information source (Lam, 2007). When a  
582 preferred source, e.g. a veterinarian, addresses or informs the producer of a potential issue or

583 risk, it is important that they are also aware of the individual beliefs of that producer. If  
584 recommendations to improve a risk factor leading to DD on farm coincide with what the  
585 producer believes, the producer will be more motivated to change and improve that issue. To  
586 motivate producers to implement changes on farm, it is also important that they believe that the  
587 issue at hand is, in fact, truly a significant matter (Ritter et al., 2017). Therefore, DD diagnostics  
588 are important to keep the producer informed about within-herd prevalence of DD. Increasing  
589 knowledge in the area of interest will likely inspire farmers to want to make changes and  
590 improvements (Bruijnis et al., 2013). For example, in the UK, DairyCo launched the DairyCo  
591 Healthy Feet Programme in 2011, with a goal to reduce lameness on farms. The program  
592 increased producer' understanding and knowledge of lameness lesions. The more accurate  
593 perceptions of lameness levels on farms increased, the greater was producers enthusiasm to  
594 reduce lameness and motivation to make essential changes (Atkinson and Fisher, 2012). As seen  
595 in the UK, veterinarians and farmers attitudes towards DD have been considerably influenced by  
596 the knowledge that the DD-associated treponemes are implicated in the etiopathogenesis of many  
597 lesions outside of cattle feet. Consequently, any effective treatments or control measures for  
598 bovine DD are likely to have additive beneficial effects (Evans et al., 2016).

599 Another part of producer' motivation is driven by real or perceived economic impacts of  
600 DD control. If a published economic impact is presented as decreased milk production or  
601 increased risk of culling, there might be limited external validity of the study, and difficult to  
602 compare to local situations or had limited validity in the country of farm origin (Gomez et al.,  
603 2015b; Bruijnis et al., 2010). Therefore, locally applicable impact measures should be available  
604 for decisions making. Unfortunately, with many gaps in our knowledge of treatment and control

605 of DD, producers' motivation might be limited and the problem not adequately and consequently  
606 addressed.

607

## 608 Conclusions

609         With the identified gaps in knowledge, it has become clear that effective prevention and  
610 control of the disease is still hampered. Although several aspects of the pathogenesis of the  
611 disease have been identified, the causal agent is still under debate. Indeed, the role of *Treponema*  
612 spp. in the development of lesions is still to be clarified. Efforts to definitively determine the  
613 consortium of organisms (either polytreponemal or polybacterial) necessary for disease induction  
614 should be a top priority, but will be costly and challenging. Without knowing what specific  
615 bacterial organisms are necessary and sufficient for disease induction, all other efforts focused on  
616 better understanding organism ecology, immunity and treatment have the potential to focus on  
617 the wrong bacteria. Additional priorities for research efforts should include an improved  
618 understanding of the ecology and reservoirs of the causal agents as well as a better understanding  
619 of the immune response to those organisms and how it improves or exacerbates lesion formation.  
620 Through filling these gaps in knowledge, the most effective intervention strategy can be  
621 developed.

622

## 623 Acknowledgments

624         The authors are hugely grateful for the scientific editing by Dr. John Kastelic which has  
625 resulted in significant improvement of our manuscript. Also, the many graduate students and  
626 research fellows that have created and published much of the scientific literature are much  
627 appreciated: Stuart Ainsworth, Joe Angell, Juan-Manuel Ariza, Mare-Madeleine Auzanneau,

628 Marleen Bruggink, Caroline Beninger, Anne Chesnin, Simon Clegg, John Coatney, Hayley  
629 Crosby-Durrani, Ibrahim Demirkan, Nick Evans, Pat Gorden, Casey Jacobs, Adam Krull, Kerry  
630 Newbrook, Anne Relun, Rachel Scholey, Laura Solano, Gareth Staton, Leigh Sullivan, and Daan  
631 Vink.

632

633 References

634

- 635 Apley, M. D., 2015: Clinical evidence for individual animal therapy for papillomatous digital  
636 dermatitis (hairy heel wart) and infectious bovine pododermatitis (foot rot). *Veterinary*  
637 *Clinics North America Food Animal Practice*, 31, 81-95, vi.
- 638 Argaez-Rodriguez, F. J., D. W. Hird, J. Hernandez de Anda, D. H. Read and A. Rodriguez-  
639 Lainz, 1997: Papillomatous digital dermatitis on a commercial dairy farm in Mexicali,  
640 Mexico: incidence and effect on reproduction and milk production. *Preventive Veterinary*  
641 *Medicine*, 32, 275-286.
- 642 Atkinson, O. C. and G. Fisher, 2012: Uptake and delivery of a lameness reduction programme in  
643 North West England; preliminary findings. *17th International symposium and 9th*  
644 *international conference on lameness in ruminants*. Bristol.
- 645 Barthold, S. W., L. D. Koller, C. Olson, E. Studer and A. Holtan, 1974: Atypical warts in cattle.  
646 *Journal American Veterinary Medical Association*, 165, 276-280.
- 647 Bassett, H. F., M. L. Monaghan, P. Lenhan, M. L. Doherty and M. E. Carter, 1990: Bovine  
648 digital dermatitis. *Veterinary Record*, 126, 164-165.
- 649 Berry, S. L., D. H. Read, T. R. Famula, A. Mongini and D. Dopfer, 2012: Long-term  
650 observations on the dynamics of bovine digital dermatitis lesions on a California dairy  
651 after topical treatment with lincomycin HCl. *Veterinary Journal*, 193, 654-658.
- 652 Berry, S. L., D. H. Read, R. L. Walker and T. R. Famula, 2010: Clinical, histologic, and  
653 bacteriologic findings in dairy cows with digital dermatitis (footwarts) one month after  
654 topical treatment with lincomycin hydrochloride or oxytetracycline hydrochloride.  
655 *Journal American Veterinary Medical Association*, 237, 555-560.
- 656 Bleriot, P., G. Thomas and P. Rousel, 2013: PARABOV : Guidelines for data recording of  
657 bovine foot lesions. *Rencontre et Recherche sur les Ruminants*.
- 658 Blowey, R. W. and M. W. Sharp, 1988: Digital dermatitis in dairy cattle. *Veterinary Record*,  
659 122, 505-508.
- 660 Booth, C. J., L. D. Warnick, Y. T. Grohn, D. O. Maizon, C. L. Guard and D. Janssen, 2004:  
661 Effect of lameness on culling in dairy cows. *Journal of Dairy Science*, 87, 4115-4122.
- 662 Brandt, S., V. Apprich, V. Hackl, R. Tober, M. Danzer, C. Kainzbauer, C. Gabriel, C. Stanek and  
663 J. Kofler, 2011: Prevalence of bovine papillomavirus and Treponema DNA in bovine  
664 digital dermatitis lesions. *Veterinary Microbiology*, 148, 161-167.
- 665 Bruijnis, M. R., H. Hogeveen, G. C and E. N. Stassen, 2013: Dairy farmers' attitudes and  
666 intentions towards improving dairy cow foot health. *Livestock Science* 155, 103-113.

- 667 Bruijnis, M. R., H. Hogeveen and E. N. Stassen, 2010: Assessing economic consequences of foot  
668 disorders in dairy cattle using a dynamic stochastic simulation model. *Journal of Dairy*  
669 *Science*, 93, 2419-2432.
- 670 Campbell, J., 2014: Digital dermatitis emerges in beef cattle. *The Western Producer*.
- 671 Castro, S. A., R. Collighan, P. A. Lambert, I. H. Dias, P. Chauhan, C. E. Bland, I. Milic, M. R.  
672 Milward, P. R. Cooper and A. Devitt, 2017: Porphyromonas gingivalis gingipains cause  
673 defective macrophage migration towards apoptotic cells and inhibit phagocytosis of  
674 primary apoptotic neutrophils. *Cell Death Disease*, 8, e2644.
- 675 Cha, E., J. A. Hertl, D. Bar and Y. T. Grohn, 2010: The cost of different types of lameness in  
676 dairy cows calculated by dynamic programming. *Preventive Veterinary Medicine*, 97, 1-  
677 8.
- 678 Chapinal, N., A. Koeck, A. Sewalem, D. F. Kelton, S. Mason, G. Cramer and F. Miglior, 2013:  
679 Genetic parameters for hoof lesions and their relationship with feet and leg traits in  
680 Canadian Holstein cows. *Journal of Dairy Science*, 96, 2596-2604.
- 681 Charfeddine, N. and M. A. Perez-Cabal, 2017: Effect of claw disorders on milk production,  
682 fertility, and longevity, and their economic impact in Spanish Holstein cows. *Journal of*  
683 *Dairy Science*, 100, 653-665.
- 684 Cheli, R. and C. Mortellaro, 1974: La dermatite digitale del bovino. *Proceedings of the 8th*  
685 *International Conference on Diseases of Cattle*, pp. 208-213. Milan.
- 686 Clegg, S. R., J. Bell, S. Ainsworth, R. W. Blowey, N. J. Bell, S. D. Carter and N. J. Evans,  
687 2016a: Isolation of digital dermatitis treponemes from cattle hock skin lesions. *Veterinary*  
688 *Dermatology*, 27, 106-112e129.
- 689 Clegg, S. R., S. D. Carter, J. P. Stewart, D. M. Amin, R. W. Blowey and N. J. Evans, 2016b:  
690 Bovine ischaemic teat necrosis: a further potential role for digital dermatitis treponemes.  
691 *Veterinary Record*, 178, 71.
- 692 Clegg, S. R., H. E. Crosby-Durrani, J. Bell, R. Blundell, R. W. Blowey, S. D. Carter and N. J.  
693 Evans, 2016c: Detection and Isolation of Digital Dermatitis Treponemes from Bovine  
694 Pressure Sores. *Journal of Comparative pathology*, 154, 273-282.
- 695 Clegg, S. R., K. G. Mansfield, K. Newbrook, L. E. Sullivan, R. W. Blowey, S. D. Carter and N.  
696 J. Evans, 2015: Isolation of digital dermatitis treponemes from hoof lesions in Wild  
697 North American Elk (*Cervus elaphus*) in Washington State, USA. *Journal of Clinical*  
698 *Microbiology*, 53, 88-94.
- 699 Clegg, S. R., L. E. Sullivan, J. Bell, R. W. Blowey, S. D. Carter and N. J. Evans, 2016d:  
700 Detection and isolation of digital dermatitis treponemes from skin and tail lesions in pigs.  
701 *Research in Veterinary Science*, 104, 64-70.
- 702 Collighan, R. J. and M. J. Woodward, 1997: Spirochaetes and other bacterial species associated  
703 with bovine digital dermatitis. *FEMS Microbiology Letters*, 156, 37-41.
- 704 Cook, N. B., J. Rieman, A. Gomez and K. Burgi, 2012: Observations on the design and use of  
705 footbaths for the control of infectious hoof disease in dairy cattle. *The Veterinary*  
706 *Journal*, 193, 669-673.
- 707 Cramer, G., K. D. Lissemore, C. L. Guard, K. E. Leslie and D. F. Kelton, 2009: The association  
708 between foot lesions and culling risk in Ontario Holstein cows. *Journal of Dairy Science*,  
709 92, 2572-2579.
- 710 Crosby-Durrani, H. E., S. R. Clegg, E. Singer, J. W. Angell, N. J. Evans, S. D. Carter, R. J.  
711 Blundell and J. S. Duncan, 2016: Severe Foot Lesions in Dairy Goats Associated with  
712 Digital Dermatitis Treponemes. *Journal of Comparative Pathology*, 154, 283-296.

713 Cutler, J. H. H., G. Cramer, J. J. Walter, S. T. Millman and D. F. Kelton, 2013: Randomized  
714 clinical trial of tetracycline hydrochloride bandage and paste treatments for resolution of  
715 lesions and pain associated with digital dermatitis in dairy cattle. *Journal of Dairy*  
716 *Science*, 96, 7550-7557.

717 Demirkan, I., R. L. Walker, R. D. Murray, R. W. Blowey and S. D. Carter, 1999: Serological  
718 evidence of spirochaetal infections associated with digital dermatitis in dairy cattle. *The*  
719 *Veterinary journal*, 157, 69-77.

720 Dhawi, A., C. A. Hart, I. Demirkan, I. H. Davies and S. D. Carter, 2005: Bovine digital  
721 dermatitis and severe virulent ovine foot rot: a common spirochaetal pathogenesis. *The*  
722 *Veterinary Journal*, 169, 232-241.

723 Doane, M. and S. Sarenbo, 2014: Exposure of farm laborers and dairy cattle to formaldehyde  
724 from footbath use at a dairy farm in New York State. *Science Total Environment*, 487,  
725 65-71.

726 Dopfer, D., 2009: The dynamics of digital dermatitis in dairy cattle and the manageable state of  
727 disease. *CanWest Veterinary Conference*. Banff, AB.

728 Dopfer, D., K. Anklam, D. Mikheil and P. Ladell, 2012: Growth curves and morphology of three  
729 *Treponema* subtypes isolated from digital dermatitis in cattle. *The Veterinary Journal*,  
730 193, 685-693.

731 Dopfer, D., A. Koopmans, F. A. Meijer, I. Szakall, Y. H. Schukken, W. Klee, R. B. Bosma, J. L.  
732 Cornelisse, A. J. van Asten and A. A. ter Huurne, 1997: Histological and bacteriological  
733 evaluation of digital dermatitis in cattle, with special reference to spirochaetes and  
734 *Campylobacter faecalis*. *The Veterinary Record*, 140, 620-623.

735 Duncan, J. S., J. W. Angell, S. D. Carter, N. J. Evans, L. E. Sullivan and D. H. Grove-White,  
736 2014: Contagious ovine digital dermatitis: an emerging disease. *The Veterinary Journal*,  
737 201, 265-268.

738 Edwards, A. M., D. Dymock and H. F. Jenkinson, 2003: From tooth to hoof: treponemes in  
739 tissue-destructive diseases. *Journal of Applied Microbiology*, 94, 767-780.

740 Evans, N. J., R. W. Blowey, D. Timofte, D. R. Isherwood, J. M. Brown, R. Murray, R. J. Paton  
741 and S. D. Carter, 2011: Association between bovine digital dermatitis treponemes and a  
742 range of 'non-healing' bovine hoof disorders. *The Veterinary Record*, 168, 214.

743 Evans, N. J., J. M. Brown, I. Demirkan, R. Birtles, C. A. Hart and S. D. Carter, 2009: In vitro  
744 susceptibility of bovine digital dermatitis associated spirochaetes to antimicrobial agents.  
745 *Veterinary Microbiology*, 136, 115-120.

746 Evans, N. J., J. M. Brown, I. Demirkan, R. D. Murray, W. D. Vink, R. W. Blowey, C. A. Hart  
747 and S. D. Carter, 2008: Three unique groups of spirochetes isolated from digital  
748 dermatitis lesions in UK cattle. *Veterinary Microbiology*, 130, 141-150.

749 Evans, N. J., J. M. Brown, C. Hartley, R. F. Smith and S. D. Carter, 2012a: Antimicrobial  
750 susceptibility testing of bovine digital dermatitis treponemes identifies macrolides for in  
751 vivo efficacy testing. *Veterinary Microbiology*, 160, 496-500.

752 Evans, N. J., R. D. Murray and S. D. Carter, 2016: Bovine digital dermatitis: Current concepts  
753 from laboratory to farm. *The Veterinary Journal*, 211, 3-13.

754 Evans, N. J., D. Timofte, D. R. Isherwood, J. M. Brown, J. M. Williams, K. Sherlock, M. J.  
755 Lehane, R. D. Murray, R. J. Birtles, C. A. Hart and S. D. Carter, 2012b: Host and  
756 environmental reservoirs of infection for bovine digital dermatitis treponemes. *Veterinary*  
757 *Microbiology*, 156, 102-109.

758 Frankena, K., J. G. Somers, W. G. Schouten, J. V. van Stek, J. H. Metz, E. N. Stassen and E. A.  
759 Graat, 2009: The effect of digital lesions and floor type on locomotion score in Dutch  
760 dairy cows. *Preventive Veterinary Medicine*, 88, 150-157.

761 Garforth, C. J., 2012: Effective communication to improve udder health: can social science help?  
762 Wageningen Academic Publishers.

763 Gomez, A., K. S. Anklam, N. B. Cook, J. Rieman, K. A. Dunbar, K. E. Cooley, M. T. Socha and  
764 D. Dopfer, 2014a: Immune response against *Treponema* spp. and ELISA detection of  
765 digital dermatitis. *Journal of Dairy Science*, 97, 4864-4875.

766 Gomez, A., N. Bernardoni, J. Rieman, A. Dusick, R. Hartshorn, D. H. Read, M. T. Socha, N. B.  
767 Cook and D. Dopfer, 2014b: A randomized trial to evaluate the effect of a trace mineral  
768 premix on the incidence of active digital dermatitis lesions in cattle. *Journal of Dairy  
769 Science*, 97, 6211-6222.

770 Gomez, A., N. B. Cook, N. D. Bernardoni, J. Rieman, A. F. Dusick, R. Hartshorn, M. T. Socha,  
771 D. H. Read and D. Dopfer, 2012: An experimental infection model to induce digital  
772 dermatitis infection in cattle. *Journal of Dairy Science*, 95, 1821-1830.

773 Gomez, A., N. B. Cook, J. Rieman, K. A. Dunbar, K. E. Cooley, M. T. Socha and D. Dopfer,  
774 2015a: The effect of digital dermatitis on hoof conformation. *Journal of Dairy Science*,  
775 98, 927-936.

776 Gomez, A., N. B. Cook, M. T. Socha and D. Dopfer, 2015b: First-lactation performance in cows  
777 affected by digital dermatitis during the rearing period. *Journal of Dairy Science*, 98,  
778 4487-4498.

779 Grenier, D., 1992a: Demonstration of a bimodal coaggregation reaction between *Porphyromonas*  
780 *gingivalis* and *Treponema denticola*. *Oral Microbiology and Immunology*, 7, 280-284.

781 Grenier, D., 1992b: Nutritional interactions between two suspected periodontopathogens,  
782 *Treponema denticola* and *Porphyromonas gingivalis*. *Infection Immunology*, 60, 5298-  
783 5301.

784 Han, S. and K. G. Mansfield, 2014: Severe hoof disease in free-ranging Roosevelt elk (*Cervus*  
785 *elaphus roosevelti*) in southwestern Washington, USA. *Journal Wildlife Diseases*, 50,  
786 259-270.

787 Hartshorn, R. E., E. C. Thomas, K. Anklam, M. G. Lopez-Benavides, M. Buchalova, T. C.  
788 Hemling and D. Dopfer, 2013: Short communication: minimum bactericidal  
789 concentration of disinfectants evaluated for bovine digital dermatitis-associated  
790 *Treponema phagedenis*-like spirochetes. *Journal dairy science*, 96, 3034-3038.

791 Hashimoto, M., S. Ogawa, Y. Asai, Y. Takai and T. Ogawa, 2003: Binding of *Porphyromonas*  
792 *gingivalis* fimbriae to *Treponema denticola* dentilisin. *FEMS Microbiology Letters*, 226,  
793 267-271.

794 Holzhauser, M., C. J. Bartels, D. Dopfer and G. van Schaik, 2008: Clinical course of digital  
795 dermatitis lesions in an endemically infected herd without preventive herd strategies. *The  
796 Veterinary Journal*, 177, 222-230.

797 Holzhauser, M., B. Brummelman, K. Frankena and T. J. Lam, 2012: A longitudinal study into the  
798 effect of grazing on claw disorders in female calves and young dairy cows. *The  
799 Veterinary Journal*, 193, 633-638.

800 Holzhauser, M., C. Hardenberg, C. J. Bartels and K. Frankena, 2006: Herd- and cow-level  
801 prevalence of digital dermatitis in the Netherlands and associated risk factors. *Journal of  
802 Dairy Science*, 89, 580-588.



803 Ito, R., K. Ishihara, M. Shoji, K. Nakayama and K. Okuda, 2010: Hemagglutinin/Adhesin  
804 domains of *Porphyromonas gingivalis* play key roles in coaggregation with *Treponema*  
805 *denticola*. *FEMS Immunology & Medical Microbiology*, 60, 251-260.

806 Jacobs, C., K. Orsel and H. W. Barkema, 2017: Prevalence of digital dermatitis in young stock in  
807 Alberta, Canada, using pen walks. *Journal of Dairy Science*, Accepted for publication.

808 Klitgaard, K., A. Foix Breto, M. Boye and T. K. Jensen, 2013: Targeting the treponemal  
809 microbiome of digital dermatitis infections by high-resolution phylogenetic analyses and  
810 comparison with fluorescent in situ hybridization. *Journal of Clinical Microbiology*, 51,  
811 2212-2219.

812 Klitgaard, K., M. L. Strube, A. Isbrand, T. K. Jensen and M. W. Nielsen, 2017: Microbiota  
813 analysis of environmental slurry and its potential role as a reservoir of bovine digital  
814 dermatitis pathogens. *Applied and Environmental Microbiology*.

815 Krull, A. C., V. L. Cooper, J. W. Coatney, J. K. Shearer, P. J. Gorden and P. J. Plummer, 2016a:  
816 A Highly Effective Protocol for the Rapid and Consistent Induction of Digital Dermatitis  
817 in Holstein Calves. *PloS one*, 11, e0154481.

818 Krull, A. C., J. K. Shearer, P. J. Gorden, V. L. Cooper, G. J. Phillips and P. J. Plummer, 2014a:  
819 Deep sequencing analysis reveals temporal microbiota changes associated with  
820 development of bovine digital dermatitis. *Infection Immunology*, 82, 3359-3373.

821 Krull, A. C., J. K. Shearer, P. J. Gorden, V. L. Cooper, G. J. Phillips and P. J. Plummer, 2014b:  
822 Deep sequencing analysis reveals temporal microbiota changes associated with  
823 development of bovine digital dermatitis. *Infection Immunology*, 82, 3359-3373.

824 Krull, A. C., J. K. Shearer, P. J. Gorden, V. L. Cooper, G. J. Phillips and P. J. Plummer, 2014c:  
825 Deep sequencing analysis reveals temporal microbiota changes associated with  
826 development of bovine digital dermatitis. *Infection Immunology*, 82, 3359-3373.

827 Krull, A. C., J. K. Shearer, P. J. Gorden, H. M. Scott and P. J. Plummer, 2016b: Digital  
828 dermatitis: Natural lesion progression and regression in Holstein dairy cattle over 3 years.  
829 *Journal of Dairy Science*, 99, 3718-3731.

830 Lam, T. J. G. M., J. Jansen, B. Van den Borne, and J. Van Veersen, 2007: A structural approach  
831 of udder health improvement via private practitioners: ups and downs., *Proceedings 46th*  
832 *Annual Meeting National Mastitis Council*, pp. 142-151. San Antonia, TX.

833 Lantos, P. M., P. G. Auwaerter and G. P. Wormser, 2014: A systematic review of *Borrelia*  
834 *burgdorferi* morphologic variants does not support a role in chronic Lyme disease.  
835 *Clinical Infectious Disease*, 58, 663-671.

836 Laven, R. A., 1999: The environment and digital dermatitis. *Cattle Pract.*, 7, 349-354.

837 Laven, R. A. and H. Hunt, 2001: Comparison of valnemulin and lincomycin in the treatment of  
838 digital dermatitis by individually applied topical spray. *Veterinary Record*, 149, 302-303.

839 Laven, R. A. and H. Hunt, 2002: Evaluation of copper sulphate, formalin and peracetic acid in  
840 footbaths for the treatment of digital dermatitis in cattle. *Veterinary Record*, 151, 144-  
841 146.

842 Laven, R. A. and D. N. Logue, 2006: Treatment strategies for digital dermatitis for the UK. *The*  
843 *Veterinary Journal*, 171, 79-88.

844 Lindley, W. H., 1974: Malignant verrucae of bulls. *Veterinary Medicine Small Animal Clinics*,  
845 69, 1547-1550.

846 Logue, D. N., T. Gibert, T. Parkin, S. Thomson and D. J. Taylor, 2012: A field evaluation of a  
847 footbathing solution for the control of digital dermatitis in cattle. *The Veterinary Journal*,  
848 193, 664-668.

849 Lohinai, Z., B. Keremi, E. Szoko, T. Tabi, C. Szabo, Z. Tulassay, J. C. DiCesare, C. A. Davis, L.  
850 M. Collins and M. Levine, 2015: Biofilm Lysine Decarboxylase, a New Therapeutic  
851 Target for Periodontal Inflammation. *Journal of Periodontology*, 86, 1176-1184.

852 Maboni, G., A. Blanchard, S. Frosth, C. Stewart, R. Emes and S. Totemeyer, 2017: A distinct  
853 bacterial dysbiosis associated skin inflammation in ovine footrot. *Scientific Reports*, 7,  
854 45220.

855 Manske, T., J. Hultgren and C. Bergsten, 2002: Topical treatment of digital dermatitis associated  
856 with severe heel-horn erosion in a Swedish dairy herd. *Preventive Veterinary Medicine*,  
857 53, 215-231.

858 Marcatili, P., M. W. Nielsen, T. Sicheritz-Ponten, T. K. Jensen, C. Schafer-Nielsen, M. Boye, M.  
859 Nielsen and K. Klitgaard, 2016: A novel approach to probe host-pathogen interactions of  
860 bovine digital dermatitis, a model of a complex polymicrobial infection. *BMC Genomics*,  
861 17, 987.

862 Merilainen, L., H. Brander, A. Herranen, A. Schwarzbach and L. Gilbert, 2016: Pleomorphic  
863 forms of *Borrelia burgdorferi* induce distinct immune responses. *Microbes Infection*, 18,  
864 484-495.

865 Merilainen, L., A. Herranen, A. Schwarzbach and L. Gilbert, 2015: Morphological and  
866 biochemical features of *Borrelia burgdorferi* pleomorphic forms. *Microbiology*, 161, 516-  
867 527.

868 Moe, K. K., T. Yano, K. Misumi, C. Kubota, K. Nibe, W. Yamazaki, M. Muguruma and N.  
869 Misawa, 2010: Detection of antibodies against *Fusobacterium necrophorum* and  
870 *Porphyromonas levii*-like species in dairy cattle with papillomatous digital dermatitis.  
871 *Microbiology Immunology*, 54, 338-346.

872 Montes, C. L., E. V. Acosta-Rodriguez, M. C. Merino, D. A. Bermejo and A. Gruppi, 2007:  
873 Polyclonal B cell activation in infections: infectious agents' devilry or defense  
874 mechanism of the host? *Journal of Leukocyte Biology*, 82, 1027-1032.

875 Murgia, R. and M. Cinco, 2004: Induction of cystic forms by different stress conditions in  
876 *Borrelia burgdorferi*. *APMIS : acta pathologica, microbiologica, et immunologica*  
877 *Scandinavica*, 112, 57-62.

878 Nilius, A. M., S. C. Spencer and L. G. Simonson, 1993: Stimulation of in vitro growth of  
879 *Treponema denticola* by extracellular growth factors produced by *Porphyromonas*  
880 *gingivalis*. *Journal of Dental Research*, 72, 1027-1031.

881 Nishikawa, A. and K. Taguchi, 2008: Healing of digital dermatitis after a single treatment with  
882 topical oxytetracycline in 89 dairy cows. *Veterinary Record*, 163, 574-576.

883 Orsel and Schwartzkopf-Genswein, 2015: Lameness and claw lesions in dairy and beef cattle.  
884 *CenCan*. Winnipeg.

885 Palmer, M. A. and N. E. O'Connell, 2015: Digital Dermatitis in Dairy Cows: A Review of Risk  
886 Factors and Potential Sources of Between-Animal Variation in Susceptibility. *Animals*  
887 *(Basel)*, 5, 512-535.

888 Plummer, P. and A. Krull, 2017: Clinical Perspectives of Digital Dermatitis in Dairy and Beef  
889 Cattle. *Veterinary Clinics of North America: Food Animal Practice*.

890 Potterton, S. L., N. J. Bell, H. R. Whay, E. A. Berry, O. C. Atkinson, R. S. Dean, D. C. Main and  
891 J. N. Huxley, 2012: A descriptive review of the peer and non-peer reviewed literature on  
892 the treatment and prevention of foot lameness in cattle published between 2000 and 2011.  
893 *Veterinary Journal*, 193, 612-616.

- 894 Read, D. H. and R. L. Walker, 1998: Papillomatous digital dermatitis (footwarts) in California  
895 dairy cattle: clinical and gross pathologic findings. *Journal Veterinary Diagnostic*  
896 *Investigation*, 10, 67-76.
- 897 Rebhun, W. C., R. M. Payne, J. M. King, M. Wolfe and S. N. Begg, 1980: Interdigital  
898 papillomatosis in dairy cattle. *Journal of American Veterinary Medical Association*, 177,  
899 437-440.
- 900 Refaai, W., R. Ducatelle, P. Geldhof, B. Mihi, M. El-shair and G. Opsomer, 2013: Digital  
901 dermatitis in cattle is associated with an excessive innate immune response triggered by  
902 the keratinocytes. *BMC Veterinary Research*, 9, 193.
- 903 Relun, A., R. Guatteo, P. Roussel and N. Bareille, 2011: A simple method to score digital  
904 dermatitis in dairy cows in the milking parlor. *Journal of Dairy Science*, 94, 5424-5434.
- 905 Relun, A., A. Lehebel, N. Bareille and R. Guatteo, 2012: Effectiveness of different regimens of a  
906 collective topical treatment using a solution of copper and zinc chelates in the cure of  
907 digital dermatitis in dairy farms under field conditions. *Journal of Dairy Science*, 95,  
908 3722-3735.
- 909 Relun, A., A. Lehebel, A. Chesnin, R. Guatteo and N. Bareille, 2013: Association between  
910 digital dermatitis lesions and test-day milk yield of Holstein cows from 41 French dairy  
911 farms. *Journal of Dairy Science*, 96, 2190-2200.
- 912 Ritter, C., J. Jansen, S. Roche, D. F. Kelton, C. L. Adams, K. Orsel, R. J. Erskine, G. Benedictus,  
913 T. J. Lam and H. W. Barkema, 2017: Invited review: Determinants of farmers' adoption  
914 of management-based strategies for infectious disease prevention and control. *Journal of*  
915 *Dairy Science*.
- 916 Rock, C., A. Krull, P. Gorden, J. Shearer and P. Plummer, 2015: Metagenomic evaluation of the  
917 dairy farm environment and facilities for evidence of digital dermatitis associated  
918 bacteria. *International Ruminant Lameness Conference*. Valdivia, Chile.
- 919 Rodriguez-Lainz, A., P. Melendez-Retamal, D. W. Hird and D. H. Read, 1998: Papillomatous  
920 digital dermatitis in Chilean dairies and evaluation of a screening method. *Preventive*  
921 *Veterinary Medicine*, 37, 197-207.
- 922 Scholey, R. A., R. W. Blowey, R. D. Murray, R. F. Smith, J. Cameron, J. P. Massey, W. E.  
923 Ollier and S. D. Carter, 2012: Investigating host genetic factors in bovine digital  
924 dermatitis. *Veterinary Record*, 171, 624.
- 925 Scholey, R. A., N. J. Evans, R. W. Blowey, J. P. Massey, R. D. Murray, R. F. Smith, W. E.  
926 Ollier and S. D. Carter, 2013: Identifying host pathogenic pathways in bovine digital  
927 dermatitis by RNA-Seq analysis. *The Veterinary Journal*, 197, 699-706.
- 928 Schopke, K., A. Gomez, K. A. Dunbar, H. H. Swalve and D. Dopfer, 2015: Investigating the  
929 genetic background of bovine digital dermatitis using improved definitions of clinical  
930 status. *Journal of Dairy Science*, 98, 8164-8174.
- 931 Shearer, J. K. and J. Hernandez, 2000: Efficacy of two modified nonantibiotic formulations  
932 (Victory) for treatment of papillomatous digital dermatitis in dairy cows. *Journal of*  
933 *Dairy Science*, 83, 741-745.
- 934 Simonson, L. G., K. T. McMahon, D. W. Childers and H. E. Morton, 1992: Bacterial synergy of  
935 *Treponema denticola* and *Porphyromonas gingivalis* in a multinational population. *Oral*  
936 *Microbiology and Immunology*, 7, 111-112.
- 937 Smalley, J. W. and T. Olczak, 2017: Heme acquisition mechanisms of *Porphyromonas gingivalis*  
938 - strategies used in a polymicrobial community in a heme-limited host environment.  
939 *Molecular Oral Microbiology*, 32, 1-23.

- 940 Sogstad, A. M., T. Fjeldaas, O. Osteras and K. P. Forshell, 2005: Prevalence of claw lesions in  
941 Norwegian dairy cattle housed in tie stalls and free stalls. *Preventive Veterinary*  
942 *Medicine*, 70, 191-209.
- 943 Solano, L., H. W. Barkema, C. Jacobs and K. Orsel, 2017a: Validation of the M-stage scoring  
944 system for digital dermatitis on dairy cows in the milking parlor. *Journal of Dairy*  
945 *Science*, 100, 1592-1603.
- 946 Solano, L., H. W. Barkema, S. Mason, E. A. Pajor, S. J. LeBlanc and K. Orsel, 2016: Prevalence  
947 and distribution of foot lesions in dairy cattle in Alberta, Canada. *Journal of Dairy*  
948 *Science*, 99, 6828-6841.
- 949 Solano, L., H. W. Barkema, E. A. Pajor, S. Mason, S. J. LeBlanc, J. C. Zaffino Heyerhoff, C. G.  
950 Nash, D. B. Haley, E. Vasseur, D. Pellerin, J. Rushen, A. M. de Passille and K. Orsel,  
951 2015: Prevalence of lameness and associated risk factors in Canadian Holstein-Friesian  
952 cows housed in freestall barns. *Journal of Dairy Science*, 98, 6978-6991.
- 953 Solano, L., H. W. Barkema, C. Pickel and K. Orsel, 2017b: Effectiveness of a standardized  
954 footbath protocol for prevention of digital dermatitis. *Journal of Dairy Science*, 100,  
955 1295-1307.
- 956 Speijers, M. H., L. G. Baird, G. A. Finney, J. McBride, D. J. Kilpatrick, D. N. Logue and N. E.  
957 O'Connell, 2010: Effectiveness of different footbath solutions in the treatment of digital  
958 dermatitis in dairy cows. *Journal of Dairy Science*, 93, 5782-5791.
- 959 Speijers, M. H., G. A. Finney, J. McBride, S. Watson, D. N. Logue and N. E. O'Connell, 2012:  
960 Effectiveness of different footbathing frequencies using copper sulfate in the control of  
961 digital dermatitis in dairy cows. *Journal of Dairy Science*, 95, 2955-2964.
- 962 Stokes, J. E., K. A. Leach, D. C. J. Main and H. R. Whay, 2012: The reliability of detecting  
963 digital dermatitis in the milking parlour. *The Veterinary Journal*, 193, 679-684.
- 964 Sullivan, L. E., R. W. Blowey, S. D. Carter, J. S. Duncan, D. H. Grove-White, P. Page, T.  
965 Iveson, J. W. Angell and N. J. Evans, 2014: Presence of digital dermatitis treponemes on  
966 cattle and sheep hoof trimming equipment. *Veterinary Record*, 175, 201-201.
- 967 Sullivan, L. E., S. D. Carter, R. Blowey, J. S. Duncan, D. Grove-White and N. J. Evans, 2013:  
968 Digital dermatitis in beef cattle. *Veterinary Record*, 173, 582.
- 969 Sullivan, L. E., N. J. Evans, R. W. Blowey, D. H. Grove-White, S. R. Clegg, J. S. Duncan and S.  
970 D. Carter, 2015a: A molecular epidemiology of treponemes in beef cattle digital  
971 dermatitis lesions and comparative analyses with sheep contagious ovine digital  
972 dermatitis and dairy cattle digital dermatitis lesions. *Veterinary Microbiology*, 178, 77-  
973 87.
- 974 Sullivan, L. E., N. J. Evans, S. R. Clegg, S. D. Carter, J. E. Horsfield, D. Grove-White and J. S.  
975 Duncan, 2015b: Digital dermatitis treponemes associated with a severe foot disease in  
976 dairy goats. *Veterinary Record*, 176, 283.
- 977 Teixeira, A. G., V. S. Machado, L. S. Caixeta, R. V. Pereira and R. C. Bicalho, 2010: Efficacy of  
978 formalin, copper sulfate, and a commercial footbath product in the control of digital  
979 dermatitis. *Journal of Dairy Science*, 93, 3628-3634.
- 980 van Amstel, S. R., S. van Vuuren and C. L. Tutt, 1995: Digital dermatitis: report of an outbreak.  
981 *Journal of South African Veterinary Association*, 66, 177-181.
- 982 Van Andel M, R. T., Thompson K, Vink WD, 2012: Review of recent bovine digital dermatitis-  
983 like lesions in cattle. *Surveillance*, 39.
- 984 Vermunt, J. J. and F. I. Hill, 2004: Papillomatous digital dermatitis in a Holstein-Friesian bull.  
985 *New Zealand Veterinary Journal*, 52, 99-101.

- 986 Vink, W. D., 2006: Investigating the epidemiology of bovine digital dermatitis: causality,  
987 transmission and infection dynamics. University of Liverpool, UK.
- 988 Vink, W. D., G. Jones, W. O. Johnson, J. Brown, I. Demirhan, S. D. Carter and N. P. French,  
989 2009: Diagnostic assessment without cut-offs: application of serology for the modelling  
990 of bovine digital dermatitis infection. *Preventive Veterinary Medicine*, 92, 235-248.
- 991 Wells, S. J., L. P. Garber and B. A. Wagner, 1999: Papillomatous digital dermatitis and  
992 associated risk factors in US dairy herds. *Preventive veterinary medicine*, 38, 11-24.
- 993 Wilson-Welder, J. H., D. P. Alt and J. E. Nally, 2015a: Digital Dermatitis in Cattle: Current  
994 Bacterial and Immunological Findings. *Animals (Basel)*, 5, 1114-1135.
- 995 Wilson-Welder, J. H., J. Nally, D. Alt and P. Plummer, 2015b: Development of a digital  
996 dermatitis model in sheep. *International Ruminant Lameness Conference*. Valdivia,  
997 Chile.
- 998 Winders, T., M. Socha and G. Cramer, 2015: An evaluation of the agreement between digital  
999 dermatitis scoring methods in the parlor, pen and hoof-trimming chute. *International*  
1000 *Lameness in Ruminant Conference*. Valdivia, Chile.
- 1001 Yano, T., R. Yamagami, K. Misumi, C. Kubota, K. K. Moe, T. Hayashi, K. Yoshitani, O. Ohtake  
1002 and N. Misawa, 2009: Genetic heterogeneity among strains of *Treponema phagedenis*-  
1003 like spirochetes isolated from dairy cattle with papillomatous digital dermatitis in Japan.  
1004 *Journal of Clinical Microbiology*, 47, 727-733.
- 1005 Yao, E. S., R. J. Lamont, S. P. Leu and A. Weinberg, 1996: Interbacterial binding among strains  
1006 of pathogenic and commensal oral bacterial species. *Oral Microbiology and Immunology*,  
1007 11, 35-41.
- 1008 Zinicola, M., H. Higgins, S. Lima, V. Machado, C. Guard and R. Bicalho, 2015: Shotgun  
1009 Metagenomic Sequencing Reveals Functional Genes and Microbiome Associated with  
1010 Bovine Digital Dermatitis. *PloS one*, 10, e0133674.
- 1011
- 1012