

Unicentre CH-1015 Lausanne http://serval.unil.ch

Year : 2017

Exploring the Role of White Matter Connectivity in Cortex Maturation

Friedrichs-Maeder Cecilia

Friedrichs-Maeder Cecilia, 2017, Exploring the Role of White Matter Connectivity in Cortex Maturation

Originally published at : Thesis, University of Lausanne

Posted at the University of Lausanne Open Archive <u>http://serval.unil.ch</u> Document URN : urn:nbn:ch:serval-BIB_3B8D547CB8972

Droits d'auteur

L'Université de Lausanne attire expressément l'attention des utilisateurs sur le fait que tous les documents publiés dans l'Archive SERVAL sont protégés par le droit d'auteur, conformément à la loi fédérale sur le droit d'auteur et les droits voisins (LDA). A ce titre, il est indispensable d'obtenir le consentement préalable de l'auteur et/ou de l'éditeur avant toute utilisation d'une oeuvre ou d'une partie d'une oeuvre ne relevant pas d'une utilisation à des fins personnelles au sens de la LDA (art. 19, al. 1 lettre a). A défaut, tout contrevenant s'expose aux sanctions prévues par cette loi. Nous déclinons toute responsabilité en la matière.

Copyright

The University of Lausanne expressly draws the attention of users to the fact that all documents published in the SERVAL Archive are protected by copyright in accordance with federal law on copyright and similar rights (LDA). Accordingly it is indispensable to obtain prior consent from the author and/or publisher before any use of a work or part of a work for purposes other than personal use within the meaning of LDA (art. 19, para. 1 letter a). Failure to do so will expose offenders to the sanctions laid down by this law. We accept no liability in this respect.

UNIVERSITE DE LAUSANNE - FACULTE DE BIOLOGIE ET DE MEDECINE

Département de radiologie médicale Service de radiodiagnostic et radiologie interventionnelle

Exploring the Role of White Matter Connectivity in Cortex Maturation

THESE

préparée sous la direction du Professeur Patric Hagmann

et présentée à la Faculté de biologie et de médecine de l'Université de Lausanne pour l'obtention du grade de

DOCTEUR EN MEDECINE

par

Cecilia FRIEDRICHS-MAEDER

Médecin diplômée de la Confédération Suisse Originaire de Büchslen (FR), Lurtigen (FR)

Lausanne

2017

UNIL | Université de Lausanne Faculté de biologie et de médecine

Ecole Doctorale Doctorat en médecine

Imprimatur

Vu le rapport présenté par le jury d'examen, composé de

Directeur de thèse	Monsieur le Professeur Patric Hagmann		
Co-Directeur de thèse			
Expert	Madame la	a Professeure	Eliane Roulet Perez
<i>Vice-Directeur de l'Ecole doctorale</i>	Monsieur	le Professeur	John Prior

la Commission MD de l'Ecole doctorale autorise l'impression de la thèse de

Madame Cecilia FRIEDRICHS-MAEDER

intitulée

Exploring the Role of White Matter Connectivity in Cortex Maturation

Lausanne, le 19 septembre 2017

pour Le Doyen de la Faculté de Biologie et de Médecine

Monsieur le Professeur John Prior Vice-Directeur de l'Ecole doctorale

Résumé

De nombreuses études ont mis en évidence un processus de maturation séquentiel pour différentes régions cérébrales, notamment la matière grise corticale et ses connections de matière blanche (WM). Cependant, les mécanismes à l'origine du processus de maturation cérébrale, ainsi que la relation entre la maturation des ces deux structures demeurent mal compris. Dans cette étude, nous utilisons l'imagerie par résonance magnétique (IRM) pour obtenir simultanément des informations reflétant la maturation microstructurelle des différentes structures cérébrales, ainsi que la mise en place des connections de matière blanche durant le développement précoce. Pour ce faire, nous employons deux mesures d'IRM bien établies reflétant les changements microstructurels liés à la maturation, soit le coefficient de diffusion apparent (ADC) et le temps de relaxation T (T1), ainsi que des algorithmes mathématiques permettant de reconstruire les groupes d'axones reliant différentes régions de matière grise (connectome).

Ce travail démontre une interdépendance entre la maturation des structures de matière grise corticale et le développement de l'arbre connectionel de matière blanche sous-jacent. En effet, nous mettons en évidence un degré de maturation analogue entre les régions corticales, ainsi que les groupes de fibres axonales associées durant les stades précoces de la maturation. Un tel lien est également observé entre deux régions corticales reliées par un groupe de fibres.

Sur la base de ces observations, cette étude propose un modèle mathématique et informatique simple parlant pour un rôle clé des structures de matière blanche dans le relai des signaux maturationnels depuis un stimuli externe via les structures primaires sensorielles et atteignant finalement les structures cérébrales corticales d'ordre plus complexes.

© 2017 Cecilia Friedrichs-Maeder / PLOS ONE

The following article appeared in PLOS ONE 12(5) and was published under the <u>Creative</u> <u>Commons Attribution (CC BY) license</u>. The article may be downloaded, reused, reprinted, modified, distributed and/or its content may be copied as long as the original authors and source are cited.

Friedrichs-Maeder CL, Griffa A, Schneider J, Hüppi PS, Truttmann A, Hagmann P (2017) Exploring the role of white matter connectivity in cortex maturation. PLOS ONE 12(5): e0177466. <u>https://doi.org/10.1371/journal.pone.0177466</u>

Disclaimer of warranties

Unless otherwise separately undertaken by the Licensor, to the extent possible, the Licensor offers the Licensed Material as-is and as-available, and makes no representations or warranties of any kind concerning the Licensed Material, whether express, implied, statutory, or other. This includes, without limitation, warranties of title, merchantability, fitness for a particular purpose, non-infringement, absence of latent or other defects, accuracy, or the presence or absence of errors, whether or not known or discoverable.



Citation: Friedrichs-Maeder CL, Griffa A, Schneider J, Hüppi PS, Truttmann A, Hagmann P (2017) Exploring the role of white matter connectivity in cortex maturation. PLoS ONE 12(5): e0177466. https://doi.org/10.1371/journal.pone.0177466

Editor: Xi-Nian Zuo, Institute of Psychology, Chinese Academy of Sciences, CHINA

Received: October 13, 2016

Accepted: April 27, 2017

Published: May 17, 2017

Copyright: © 2017 Friedrichs-Maeder et al. This is an open access article distributed under the terms of the <u>Creative Commons Attribution License</u>, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: The research and authors were supported by the Swiss National Science Foundation (#33CM30D124101, #51AU40_125759).

Competing interests: The authors have declared that no competing interests exist.

RESEARCH ARTICLE

Exploring the role of white matter connectivity in cortex maturation

Cecilia L. Friedrichs-Maeder¹*, Alessandra Griffa^{1,2}, Juliane Schneider^{3,4}, Petra Susan Hüppi⁵, Anita Truttmann³, Patric Hagmann^{1,2}

 Department of Radiology, Centre Hospitalier Universitaire Vaudoise (CHUV), Lausanne, Switzerland,
Signal Processing Laboratory (LTSS), Ecole Polytechnique Fédérale de Lausanne (EPFL), Lausanne, Switzerland,
Clinic of Neonatology and Follow-up, Department of Pediatrics, Centre Hospitalier Universitaire Vaudois (CHUV), Lausanne, Switzerland,
Division of Neurology, The Hospital for Sick Children, University of Toronto, Toronto, Canada,
Division of Development and Growth, Department of Pediatrics, University of Geneva, Geneva, Switzerland

* ceciliamaeder@gmail.com

Abstract

The maturation of the cortical gray matter (GM) and white matter (WM) are described as sequential processes following multiple, but distinct rules. However, neither the mechanisms driving brain maturation processes, nor the relationship between GM and WM maturation are well understood. Here we use connectomics and two MRI measures reflecting maturation related changes in cerebral microstructure, namely the Apparent Diffusion Coefficient (ADC) and the T1 relaxation time (T1), to study brain development. We report that the advancement of GM and WM maturation are inter-related and depend on the underlying brain connectivity architecture. Particularly, GM regions and their incident WM connections show corresponding maturation levels, which is also observed for GM regions connected through a WM tract. Based on these observations, we propose a simple computational model supporting a key role for the connectome in propagating maturation signals sequentially from external stimuli, through primary sensory structures to higher order functional cortices.

Introduction

Although most of its cellular components are formed by mid-gestation, the human brain continues to mature into postnatal life [1,2]. This reflects the development of the organized structural connectivity network, which is at the basis of neural processing [3], for a review). These maturation processes consist of myelination of both white (WM) and gray matter (GM), but also of dendritic arborization, growth of terminal axon arborization and synaptic pruning in the cortex. Adequate completion of this developmental phase is crucial as insults during this period result in impaired neurodevelopmental outcome [4].

Distinct sequential patterns of GM and WM maturation are described. Cortical GM maturation is related to the development of brain functions. Evidence shows that primary sensory and motor areas differentiate [5,6] and reach mature dendritic structure [7,8], adequate intra-

cortical myelination [9,10] and mature macroscopic architecture [11] before higher-order processing areas such as fronto-parietal cortices. WM axonal maturation mainly follows a spatial gradient, with myelination beginning in central and caudal areas and progressing towards polar and rostral locations [12–15].

Evidence from histology and in vivo imaging studies suggests that a relationship exists between the maturation of axonal WM connections and local GM structural features (e.g., emergence of gyri and sulci) [16–18]. Moreover, the WM maturation level of distinct functional circuits was related to corresponding functional scores, such as performance in working memory [19] and language-related tasks [20]. Nonetheless, so far, the large-scale mechanisms driving the brain maturation patterns, and the relationship between the development of GM and WM structures, remain unclear.

While early patterns of neural cell differentiation and axonal guidance are mainly driven by genetic molecular cues [21], refinement and maturation of the produced coarse neural circuits including synaptic, axonal and dendritic outgrowth and pruning also depend on neural activity [22]. This early electrical activity is spontaneous during early development and progressively becomes experience-driven during the postnatal period [23]. Starting from the last trimester of pregnancy [24], it acts in synergy with molecular cues [25] and is fundamentally important as it provides trophic support for neuronal survival and endogenous guidance for activity dependent wiring [26–28]. Recent studies outlined a key role of action potentials for promoting myelination [29,30]. Animal studies provide evidence that early feed-forward sensory experience plays a key role in shaping local GM structure [31,32] and excitatory/inhibitory network architecture [33–35]. Altogether, these findings suggest that WM connections relay essential signals involved in cerebral plasticity and development. Accordingly, the propagation of nervous signals from one cortical area to the next through WM tracts could promote maturation of both the GM and the outgoing WM connections, and ultimately contribute to define the overall spatio-temporal patterns of brain maturation.

From these considerations, we make a simplifying hypothesis that: (i) WM axonal pathways play a key role in shaping maturation of cortical structures, and (ii) maturation propagates in a feed-forward fashion originating from primary cortical sensory areas, whose activations are initially spontaneous and later on driven by external sensory inputs, and spreading through the WM network toward higher-order processing areas over time.

Recently developed MRI sequences sensitive to microstructural changes in the cerebral cortex and the WM (diffusion tensor, T1-mapping) have emerged as sensitive assays to study developmental related changes in human brain [36–40]. Our study is designed in two parts. First, we use two of these measures, namely the Apparent Diffusion Coefficient (ADC) and the T1 relaxation time (T1). We then extensively compare these parameters within GM regions and within their incident WM axonal tracts. In the second part, we propose a computational model of brain maturation based on a random walk process, which reproduces empirical measures of cortical and WM maturation, and supports the hypothesis that the WM connectivity network plays a key role in conveying biological signals essential for brain maturation.

Materials and methods

Subjects

Neonatal scans from 9 very prematurely born infants (4 males, mean gestational age: 27.3 weeks \pm 1.0 day) acquired at term equivalent age (TEA, mean age at scan 41.0 weeks \pm 2.0 days) were selected from a prospective cohort of neonates born before 30 weeks of gestation conducted between February 2011 and May 2013 in the level III neonatology unit at the University Hospital of Lausanne, Switzerland [40]. This cohort included solely children considered

as 'cerebral low-risk', meaning that they showed no evidence for severe intraventricular haemorrhage grade III-IV and/or parenchymal haemorrhagic infarction, no high grade periventricular leucomalacia, no congenital malformations or genetic abnormalities and had normal neurological assessment at TEA according to the Hammersmith Neonatal Neurologic Examination from [41]. The local ethical committee of the Canton de Vaud approved the current study and parental written informed consent was obtained. Inclusion of the subjects in our study was based on (i) data quality, (ii) no substantial brain lesions on TEA MRI and (iii) absence of significant cognitive and psychomotor delays at 18 months of corrected age. Single subjects' characteristics are reported in <u>S1 Table</u>.

MRI data acquisition

Structural and diffusion-weighted MR images were obtained with a 3T scanner (MAGNE-TOM TrioTim, Siemens Healthcare, Germany) and a dedicated 8-channel baby head coil (NOMAG, LMT Lammers) integrated in an MR-compatible incubator. Structural scans were acquired using a double-echo MP2RAGE sequence with the following parameters: flip angle 4 degrees, TI1/TI2/TR = 900/2200/4000 ms, TE = 3.17 ms, GRAPPA R = 2, TA = 4.58 min. Two image volumes per echo time were obtained (at the 1st and 2nd inversion time INV1b and INV2b) and two morphological high resolution T1-weighted images were reconstructed: a bias-free T1-weighted image (*flatImg*) and a T1 relaxation map (*T1map*) [42]. The acquisition was sagittal with a FOV of 190 mm (feet-head) x 179 mm (ant-post) and a matrix of 256 x 241 voxels yielding an in-plane resolution of 0.7 x 0.7 mm² and 96 slices of 1.2 mm.

The diffusion-weighted acquisitions were performed using Diffusion Tensor Imaging (DTI) with a twice-refocused spin echo EPI sequence with TR/TE = 5200/84 ms and a spatial resolution of 8 mm³ (2x2x2 mm³) isotropic with an in-plane matrix of 96x96 voxels and 43 slices. Five b0-images and 30 diffusion-weighted images with b-value of 1000 s/mm² were acquired with varying directions. Acquisition time was about 3 minutes.

Anatomical brain tissue segmentation and parcellation

MP2RAGE *flatImg* scans were segmented into brain tissue classes (GM including cortical and subcortical structures, WM and cerebrospinal fluid) based on single voxel signal intensity and probabilistic tissue location priors from the publicly available neonate brain atlas by Shi et al. [43] using *SPM* software [44]. The obtained probabilistic maps were thresholded empirically to obtain binary tissue masks. The GM volume was subdivided into 90 anatomical ROIs, i.e. 45 per hemisphere, according to the Shi atlas. Each ROI was dilated in order to reach the GM-WM border, and possible holes and discontinuities in the ROI masks were corrected using mathematical morphology operations. An expert neuroradiologist reviewed the intermediary mask images at each processing step. S1 Fig shows the resulting anatomical brain masks for the 9 subjects. The MP2RAGE *flatImg* volumes were linearly registered to the first B0 volume in order to bring the brain tissue masks into the diffusion space.

Diffusion MRI processing and brain connectivity estimation

Diffusion-weighted MRI volumes were registered to the first B0 scan (rigid-body transformation, 6 degree of freedom) to correct for head motion. Next, these images were corrected for eddy current distortions using the FSL eddy correction software [45]. Tensor based reconstruction of diffusion information was performed using the *dtirecon* tool from Diffusion Toolkit [46], with the gradient table updated for the rigid-body registration performed earlier. Whole brain tractography was performed using the deterministic streamline algorithm implemented in the *Connectome Mapping Toolkit* [47,48] and using standard parameters. 32 streamlines were initiated in each WM voxel and propagated along the two opposite directions given by the local tensor orientation. Streamlines propagation was stopped if one of the following criteria was met: (i) touching the WM-GM boundary, or (ii) angle between diffusion directions in neighbouring voxels larger than 60 degrees. Fibers shorter than 6 mm and fibers not reaching the WM-GM border were discarded. Note that due to variability in head size, no stopping criterion was used for the maximal fiber length. Each reconstructed fiber trajectory reaching the WM-GM interface represented an anatomical connection between the corresponding pair of GM regions. Finally, structural connectivity matrices representing subjectwise brain networks were constructed by counting the number of streamlines connecting each pair of cortical and subcortical regions. For all analyses, we used a group-representative connectivity matrix computed by considering only the connections present for at least 50% of the subjects (excluding intra-ROI connections) and averaging the corresponding connection weights. This arbitrary group threshold was chosen based on previous work showing that a threshold between 50 and 60% yielded the best approximation of the number of existing connections and an acceptable balance between elimination of false positives and prevention of false negatives [49].

WM and GM maturation parameters

In this study, we used two MRI measures reflecting maturation related changes in cerebral microstructure, namely the Apparent Diffusion Coefficient (ADC) and the T1 relaxation time (T1). ADC corresponds to the mean diffusivity, which reflects the level of diffusion restriction. In the brain ADC is a biomarker of maturation as it decreases with age [50,51] and reflects changes in axonal diameters and packing as well as myelination [52]. T1 relaxation changes with the concentration of macromolecules and lipids and therefore is also a marker of brain maturation and myelination [40,53,54]. The MP2RAGE sequence enables rapid, robust and high resolution of maps of T1 relaxation as described in [42,55]. ADC maps were computed from reconstructed DTI data using Diffusion Toolkit dtirecon function. Next, average ADC and T1 values were computed for each GM ROI, and for each WM connection between pairs of GM regions by averaging the voxel-wise scalars along the connecting streamline trajectories. Mean ADC and T1 values were calculated over (i) voxels belonging to each GM ROI, and (ii) WM connections incident to this ROI. In order to account for partial volume effects, GM voxels whose value differed by more than one standard deviation from the ROI mean were discarded from the analysis. From the 4005 possible ROI pairs, we restricted our analysis to the group wise connectivity matrix described in the previous section yielding a total of 450 connected and 3555 unconnected ROI pairs. Pearson correlation analyses were used to assess the relationship between ADC and T1 values in ROIs and over their incident WM connections. ROIs were sorted into four different groups according to the function-related GM maturation sequence described in the literature: group SUB included the subcortical structures (Basal Ganglia, the Thalami, the Amygdala and the Hippocampus), Group PRIM comprised primary sensory and motor cortices, Group SEC secondary processing areas and Group TER higher order tertiary areas (S2 Table).

Maturation random walk model

A computational random walk model [56] was used to simulate the evolution of the brain tissue's maturation as a particle walking along the structural brain pathways. Starting in one of the primary sensory cortex regions (*rolandic operculum*, *calcarine cortex*, *postcentral gyrus*, *paracentral lobule*, *heschl gyrus*), a particle travels from one ROI (network vertex) to another along anatomical connections (edges) of the brain network. The weight *w_{ij}* of an edge connecting two vertices *i*, *j* of the brain network is equal to the normalized number of streamlines connecting the two vertices (i.e., the number of streamlines connecting vertices *i* and *j* divided by the overall number of connecting streamlines resulting from the tractography algorithm). In our computational model, the probability of moving from a vertex *i* at time point *t* to a vertex *j* at time point *t*+1 is proportional to the weight w_{ij} of the edge (*i*,*j*). Furthermore, particles are not allowed to travel back to the previous region (no-back walk). Formally, if the walk is at vertex *i* at time *t*, the probability of taking a step along one of the edges attached to *i* is $p_{ij} = \frac{w_{ij}}{\sum_{k \in N_i^t} w_{ik}}$, with N_i^t set of neighbours of vertex *i*, excluding the vertex situated at

the previous step *t*-1. We considered the number of particle transits through each GM region or WM connection, at each step of the random walk, as a score of local maturation. Random walks were initiated in each one of the ten primary cortex, and propagated for ten randomwalk steps (RWSs). This process was repeated 1000 times for each seeding area, and mean GM and WM maturation scores were computed. Finally, spatiotemporal maturation patterns yielded by each seeding area were summed, illustrating whole brain maturation scores. For each RWS, the relationship between experimental ADC and T1 values on the one side, and total brain maturation scores on the other side, was assessed with a Spearman's correlation analysis.

Results

Structural connectivity properties

Fig 1A represents the weighted group connectivity matrix for the 9 subjects, which has a density (percentage of non-zero entries) of 11.2% and no disconnected nodes. The colorbar indicates the number of streamlines for each connection. Fig 1B shows the connection length distribution for the connectivity network.

Maturation biomarkers in GM and WM

In the first part of our study, we separate the 90 GM ROIs into 4 function-related groups: group *SUB* (n = 12, *n* number of ROIs) corresponds to subcortical nuclei, group *PRIM* (n = 12) represents the primary sensory and motor cortices, group *SEC* (n = 34) secondary processing areas and group *TER* (n = 32) higher order tertiary areas (S2 Table). Connection distribution among these groups is represented in Fig 1C. Subgroups SUB and SEC display connections to all other subgroups, while subgroup PRIM mainly connects to SEC and subgroup TER to all other groups but PRIM, which is anatomically meaningful.

The distribution of mean ADC (top row) and T1 (bottom row) values in the 9 subjects is depicted for each group of ROIs in Fig 2A (left). We observe that ADC and T1 in the *SUB* group are lower than in the *PRIM* group, which in turn are lower than in the *SEC* and *TER* group and the latter have the highest values. A non parametric Jonckheere-Terpstra permutation (JT) analysis [57] confirms a significant ordered difference in these ADC (*JT* = 9.1, $p \approx 0.0$) and T1 (*JT* = 7.5, p < 10⁻¹⁰) values. We also perform this analysis for the WM tracts incident to each ROI by computing the average over the incident connections (ADC: Fig 2A right, top, T1: Fig 2A right, bottom) and observe a comparable ordered increase in ADC (*JT* = 7.4, p < 10⁻¹⁰) and T1 (*JT* = 5.7, p < 10⁻⁵) across groups. Furthermore, significant correlation analyses demonstrate that ADC (r = 0.67, p < 10⁻¹⁰, Fig 2B top) and T1 (r = 0.84, p < 10⁻¹⁰, Fig 2B bottom) values in GM and WM are highly interdependent. Yet, this correlation was not present after randomization [58] of the connectivity matrix (ADC: r = -0.06, p = 0.56; T1: r = -0.006, p = 0.95).





A Pearson's correlation analysis further shows that mean ADC of pairs of GM ROIs are correlated to the average values along the connecting WM tract (r = 0.53, $p < 10^{-10}$, Fig 2C top). This is also the case for the T1 relaxation time (r = 0.77, $p < 10^{-10}$, Fig 2C bottom). To account for possible proximity effects, analyses are reproduced on two subgroups of connections: *long* (> mean connection length, c = 179, c = number of connections) and *short* (< mean connection length, c = 271), yielding comparable results for ADC (*long*: r = 0.58, $p < 10^{-10}$; *short*: r = 0.49, $p < 10^{-10}$, S2 Fig) and T1 relaxation time (*long*: r = 0.79, $p < 10^{-10}$; *short*: r = 0.75, $p < 10^{-10}$, S3 Fig).

Taking a step further, we compare two groups of pairs of GM ROIs, those that are connected through a WM tract (450 pairs) and those that are not (3555 pairs). A Pearson's correlation analysis outlines a significant correlation in ADC between GM ROI pairs if a structural link is present between them (r = 0.34, $p < 10^{-10}$), while no association can be found if no connection is present (r = 0.03, p = 0.06). T1 relaxation time yields comparable results (connected:

PLOS ONE



Fig 2. ROIs and incident connections. A: Distribution of ADC [10^{-6} mm/s²] (top) and T1 [ms] (bottom) values for each ROI group and the incident connections. B: Scatterplot for ADC (top) and T1 (bottom) in ROIs and their incident connections, for all ROIs. Pearson's correlation: ADC r = 0.67, p < 10^{-10} ; T1 r = 0.84, p < 10^{-10} . Groups: *SUB* (n = 12) in black (diamonds), *PRIM* (n = 12) in dark gray (circles), *SEC* (n = 34) in gray (stars), *TER* (n = 32) in light gray (squares). C: Correlation between mean ADC [10^{-6} mm/s²] (top) and T1 [ms] (bottom) of GM ROI pairs and average ADC value along the connecting WM tracts (ADC: Pearson r = 0.54, p < 10^{-10} , T1: Pearson r = 0.77, p < 10^{-10} ; n = 450).

PLOS ONE

r = 0.25, p $< 10^{-5}$; not connected: r = 0.009, p = 0.7). These results are reproduced at the single-subject level (S3 and S4 Tables).

Modeling maturation

In the second part of our study, we investigate possible mechanisms driving brain tissue maturation and the role of WM connectivity by means of a random walk model [56]. We consider the number of particle transits through each GM ROI or WM connection at each RWS as a score of local maturation. We observe that simulated maturation for ROIs (Fig 3: blue) and their incident connections (Fig 3: red) correlate negatively with the ADC values from our experimental data (Fig 3: box). Highest Spearman correlations between number of particle transits and ADC values are reached at early RWSs (ROIs: r = -0.75, $p < 10^{-10}$ at RWS 2; incident connections: r = -0.68, $p < 10^{-10}$ at RWS 1) and decrease progressively through the simulation. Correlation with T1 values shows a similar pattern (ROIs: r = -0.43, $p < 10^{-3}$ at RWS 1; incident connections: r = -0.46, $p < 10^{-4}$ at RWS 1).

As a validation, we perform the same analysis (i) using the same structural WM network, starting with a random sample of 10 GM regions as seeds and (ii) using a randomized structural network with preserved node degree distribution, starting from the same sensory areas. Both these randomization were repeated 50 times (50 random seedings and 50 network randomizations). In the first case, simulated maturation values showed no (ROIs, S4 Fig left) or weak (incident connections, S4 Fig right) correlation with the experimental data. In the second





Fig 3. Modelled maturation scores projected on a standard brain surface for four representative simulated time points (RWSs): **ROI in blue (top), connections in red (bottom).** Box: Experimental data (ADC [10⁻⁶ mm/s²]) on brain surface and Pearson's correlations with modelled maturation scores for RWS 1–10. The RWS with the best correlation is represented by a star.

case, simulated maturation values showed weaker (ROIs, <u>S5 Fig</u> left) and no (incident connections, <u>S5 Fig</u> right) correlation with the experimental data.

Simulated maturation values for ROIs and connections increase over progressive RWSs of the random walk, following an expected ordered pattern: areas from group *PRIM* have higher values than group *SEC* and further than group *TER*, which show the lowest values (Fig 4). This trend is confirmed by a JT analysis for each RWS (<u>S5 Table</u>). As expected, these differences lessen as the simulation reach later RWSs (decreasing JT statistics).

Note that subcortical areas were excluded from the correlation analysis, since *on one side* our random walk model exclusively explores the role of sensory input in network mediated maturation, and on the other side subcortical areas do not mature all at once [12,59,60] and their role in the maturation process is uncertain. This was also valid for the thalami, despite their known key role in the propagation and gating of sensory signals to primary sensory regions [61], because of their particular connection network. The various thalamic nuclei have heterogeneous connections to different cortical areas (including primary sensory, but also tertiary cortices), involved in information processing [62] and poor intra-thalamic connections.



Fig 4. Histograms and brain surface plots for experimental ADC [10⁻⁶ mm/s²] values (left) and modelled maturation scores (right, four key RWSs) in ROI groups. Top: ROIs; bottom: incident connections. Highest Pearson's correlations are indicated with a star for the corresponding RWS (and colour).

Hence, considering all thalamus nuclei as one ROI would imply creating an artificial hub. Although this is often done in adult structural and functional connectivity studies, it does not properly reflect functional anatomy, especially at early developmental stages, as all thalamocortical connections do no mature at the same time.

Discussion

Relation between GM and WM development

Our findings support a strong relationship between maturation of cortical areas and their underlying WM axonal circuits. The analysis of two MRI-based indicators of brain tissue maturation, namely ADC and T1 [36,39], reveals hierarchical increasing maturation levels from subcortical structures, to primary areas, to cortices involved in secondary and tertiary brain

PLOS ONE

functions. A similar pattern is observed in the WM connections incident to these areas, suggesting a link between cortical maturation, long-range brain connectivity and WM myelination.

Our results are in agreement with the function-related maturation sequence previously described for GM structures in dissection studies where primary areas develop earlier than association and higher order processing areas [5,9,10]. Early studies examining WM myelination reported comparable findings [12,13]. However later research describes myelination as following a spatial rather than a functional and connectional gradient [14,15,39].

Early connectivity studies investigate the co-variation between cortical morphology measures (e.g., cortical thickness, surface area) in different brain regions and report highly organized correlation patterns, coinciding with the presence of underlying gross anatomical tracts [63–65]. These findings were reproduced in a developmental approach by Raznahan and colleagues who showed that cortical thickness measures for different brain regions belonging to the default mode network and one task oriented network can be predicted from well-described patterns of cortical functional and WM interconnectivity [66]. The existence of a strong relationship between GM and WM maturation is also supported by evolutionary studies. For example, Barton and Harvey highlight that the coordination of brain anatomical changes follows known patterns of structural connectivity [67].

Despite these encouraging results, explicit reports on the relationship between maturation parameters in GM and WM remain few and their results inconsistent. Cortical ADC is related to ADC in the underlying WM regions in one primary, two secondary and one tertiary brain region [68]. But a whole-brain investigation of cortical thickness and myelin content did not highlight such a relationship [69].

Compared to these findings, the strength of our study lies in the joint assessment of early stages of GM and WM maturation over the whole cerebrum at the level of (i) single structures and (ii) subsystems consisting of pairs of GM regions and their anatomical link. We show that ADC and T1 values in GM areas are consistently related to values along the axonal fiber bundles connecting them. Others, using a T2-based MRI sequence and DTI, noted a comparable relation for functional subsystems in temporal [70] and pre-frontal [71] regions. Additionally, a recent study showed a correlation between decrease in cortical thickness (interpreted as maturation) in frontal cortical regions and increase in fractional anisotropy in the WM tracts initiated from these regions, in school-age children [72]. These results point out a strong interdependence between the development of GM structures and the underlying WM connectivity network. Correlation analyses indicate that GM areas are not developing independently. Indeed we observe that ADC and T1 in GM regions connected through an anatomical link are highly related. On the same line, correlations of T2-measures [70] and cortical thickness [63,66] were reported in connected cortical regions involved in language processing and in two functional networks. Furthermore, shared expression signatures for genes involved in neuronal development and axon guidance also seem to depend on structural connectivity in different regions of the rat brain [73]. Altogether, this points to an important role of structural WM connections during the cortical maturation process.

A connectivity-based model for brain maturation

A large body of evidence suggests that sensory-driven neural activity plays a key role in shaping the maturation of GM and WM cerebral structures at the cellular level ([74], for a review). Neural signals, be they initially endogenous and later on exogenous, are relayed hierarchically through thalamo-cortical pathways from primary receiving areas to higher order processing regions [75,76]. Together with our findings on the interplay between GM and

WM maturation, this suggests that the connectivity network conveys signals implicated in maturation processes.

This work proposes a simple random-walk model to explore the role of the WM network on the aspects of the maturation process that are mechanistically related to sensory inputs. In our model, maturation particles (i.e., random walkers) are initiated in cortical sensory areas and left to diffuse across the WM network. A similar computational approach was recently used to mimic viral-like spreading of neurodegenerative diseases across the structural WM network [77,78]. Indeed we observe that simulated GM and WM maturation scores reproduce experimental early maturation patterns, and they increase hierarchically in time following function-related developmental patterns reported in literature [5,9,10].

The idea that tissues' maturation could progress in a hierarchical fashion from lower-order sensory-receiving areas to the yet immature higher-order areas in their vicinity was already proposed by Guillery [79]. Our findings suggest that the WM connectivity backbone could play a key role in relaying maturation signals. It is realistic to think that during early development, the callow brain network mainly conveys coherent sensory information to higher processing areas, which would promote the maturation of related axonal pathways and connected GM regions. In line with this, connections between lower level unimodal regions strengthen before connections towards association areas, and further on to higher order processing regions [80]. Altogether, the progression of maturation from lower order areas to higher processing regions simulated with our model resembles the evolution of GM density measures between 5 and 20 years reported by Gogtay [81]. Finally, numerous studies on prematurity support the concept that WM connectivity is critical for the development of connected cortices and its lesion leads to histological disorganization of connected GM areas [82–84].

Technical limitations

The main limits of this study are related to the characteristics of the investigated population, particularly the small sample size and the prematurity condition. On one side, premature birth is associated with delayed and altered microstructural development of cortical GM [85] and WM circuitry ([86], for a review). On the other side, these changes vary with gestational age and diffuse aggressions (e.g., nutrition, infection), which are common in this population [87]. We argue that the effects pursued in this study are coarse macroscopic maturation changes outweighing subtle inter-population differences and should also be observable in prematurely born children with 'cerebral low-risk', scanned at term equivalent age as in normal term born kids. Consistently, subject-level analyses reproduce group-level findings, excluding the presence of outliers in our cohort.

Imaging neonates is both technically and ethically challenging. First, scanning normal term newborns is ethically debatable, reason why we focused on prematurely born babies who deserve through their condition an MRI examination. Second, scans had to be done without sedation, increasing the chance of motion artefacts. Additionally, tissue contrasts in neonatal brain differ notably from those of adult brains [88]. This difference can have an impact on image processing steps. In this work, we performed careful quality checks and post-processing motion correction of our data. However, we did not quantify mean relative frame-wise displacement between interspersed non-weighted diffusion volumes as has been done by others [89], but outlier rejection was performed by a senior pediatric neuroradiologist who checked all sequences visually during acquisition and had the ones with movement artifacts repeated. Six subjects were excluded because of poor data quality due to excessive motion during scans.

Another limitation of this study concerns the estimation of tissue maturation levels using MRI. Variations in MRI signals result from multiple and inter-dependent cellular processes

[90], making the mapping between MRI parameters and cellular maturation processes challenging. ADC and T1 measures relate to multiple neurobiological aspects of the maturation process. ADC measures intra- and extra-cellular water mobility, which is influenced by dendritic arborization, axonal packing and myelin sheath thickness, while T1 provides information about lipid and macromolecules concentration associated with myelination [36,39,60,91,92]. Accordingly, a combination of these MRI parameters is well suited for the study of brain maturation in vivo [93]. Furthermore, partial volume effects and cortical ingrowing axons may contribute to the cortical ADC and T1 values in areas located at the interface between cortical GM and subcortical WM. We limited these effects by excluding GM voxels with ADC and T1 values differing by more than one standard deviation from the ROI mean from the analysis. For a more accurate estimation of the tissues maturation level, advanced MRI sequences tailored to WM microstructural investigation [94] are needed. However, these methodological approaches are still in their early stage and the applicability to newborns remains difficult due to the long acquisition time.

In order to limit the scanning time, a DTI sequence was chosen for this study. It is well known that the combination of DTI and deterministic tractography could under-estimate the degree of inter-regional connectivity between ROIs linked by complex and long WM trajectories [95,96]. However, this rather conservative approach limits the number of retrieved false positive connections [96] and seems well-suited for the correlation and random-walk analyses performed in the present study [97]. Indeed, this study concentrates on uncovering coarse macroscopic maturation mechanisms and does not attempt to provide a precise description of these.

Finally, we compared our random walk model with experimental MRI biomarkers from a single age only. The relatively coarse developmental steps of the model, along with the early developmental stage of the studied population could explain higher correlations at early RWSs. Still, although qualitative observations demonstrate a clear link with developmental patterns described in the literature, extension of this study to control populations across life-span could yield further biological insights into neurodevelopmental mechanisms.

Supporting information

S1 Fig. Anatomical ROI masks for all 9 subjects. The same transverse (first column), sagittal (second column) and coronal (third column) are displayed. Color-coding reflects single ROIs and corresponding ROIs for each hemisphere are displayed in the same colour. (EPS)

S2 Fig. Correlation between mean ADC $[10^{-6} \text{ mm/s}^2]$ of GM ROI pairs and average ADC value along the connecting WM tracts (Fig 2C top) for *short* connections (< mean connection length, c = 271, c = number of connections): Pearson r = 0.49, p < 10^{-10} ; and long connections (> mean connection length, c = 179): Pearson r = 0.58, p < 10^{-10} . (EPS)

S3 Fig. Correlation between mean T1 [ms] of GM ROI pairs and average ADC value along the connecting WM tracts (Fig 2C bottom) for *short* connections (< mean connection length, c = 271, c = number of connections): Pearson r = 0.75, p < 10^{-10} ; and long connections (> mean connection length, c = 179): Pearson r = 0.79, p < 10^{-10} . (EPS)

S4 Fig. Maturation simulation null model (1): Correlation between experimental and simulated data for 50 random seedings (random sampling of 10 regions from all possible

ROIs): ROIs (left), incident connections (right). (EPS)

S5 Fig. Maturation simulation null model (2): Correlation between experimental and simulated data for 50 randomizations of the structural network with preserved node degree distribution: ROIs (left), incident connections (right). Note that the simulations starts from the same sensory areas as the original maturation simulation. (EPS)

S1 Table. Subjects' characteristics: sbj indicates subject number; GA, gestational age; BW: birthweight; M, months; IVH, intraventricular hemorrhage grading according to Papile [98] on cerebral ultrasound (3 infants presented with IVH: 2 with grade I and 1 with bilateral grade II). Not shown here, only one infant (Sbj 3) presented with a small unilateral cerebellar hemorrage. PVL indicates periventricular leucomalacia grading according to L. de Vries [99] on cerebral ultrasound. MDI and PDI are mental and psychomotor developmental indices from Bayley Scales of Infant Development II edition, norm mean (SD) is 100 (15). The values obtained are consistent with expected findings of good evolving preterm babies. All these babies had a normal neurological exam at term equivalent age (TEA). According to the Kidokoro score [100,101] only 2 infants had a mildly abnormal score (5 and 7) wheareas all others presented with a normal score (1–3). Values are presented in mean ± SD, except for the Kidokoro score* (median and range).

(DOCX)

S2 Table. ROI allocation for each group (for one hemisphere). Note that the ROI distribution is symmetric between both hemispheres. (DOCX)

S3 Table. ADC $[10^{-6} \text{ mm/s}^2]$ single subject's values. (DOCX)

S4 Table. T1 [ms] single subject's values. (DOCX)

S5 Table. Non parametric Jonckheere-Terpstra permutation analysis (JT) for four key random walk steps (RWSs). (DOCX)

Acknowledgments

We thank Tobias Kober, Andreina Pauli and Frantisek Vasa for their help with the data processing.

Author Contributions

Conceptualization: PH AT PSH. **Data curation:** JS CLFM.

.

Formal analysis: CLFM AG PH.

Funding acquisition: PH AT PSH.

Investigation: JS AT.

Methodology: PH AT PSH.

Project administration: PH AT PSH.

Resources: PH AT PSH.

Software: CLFM AG.

Supervision: PH.

Validation: PH CLFM AG.

Visualization: CLFM.

Writing - original draft: CLFM AG PH.

Writing - review & editing: CLFM PH AT JS PSH AG.

References

- Giedd JN, Blumenthal J, Jeffries NO, Castellanos FX, Liu H, Zijdenbos A, et al. Brain development during childhood and adolescence: a longitudinal MRI study. Nat Neurosci. 1999; 2: 861–3. https://doi. org/10.1038/13158 PMID: 10491603
- Gilmore JH, Shi F, Woolson SL, Knickmeyer RC, Short SJ, Lin W, et al. Longitudinal Development of Cortical and Subcortical Gray Matter from Birth to 2 Years. Cereb Cortex. 2012; 22: 2478–2485. https://doi.org/10.1093/cercor/bhr327 PMID: 22109543
- Tau GZ, Peterson BS. Normal development of brain circuits. Neuropsychopharmacology. Nature Publishing Group; 2010; 35: 147–168.
- Marret S, Marchand-Martin L, Picaud JC, Hascoët JM, Arnaud C, Rozé JC, et al. Brain Injury in Very Preterm Children and Neurosensory and Cognitive Disabilities during Childhood: The EPIPAGE Cohort Study. PLoS One. 2013; 8: 1–10.
- 5. Brodmann K. Vergleichende Lokalisationslehre der Grosshirnrinde in ihren Prinzipien dargestellt auf Grund des Zellenbaues. Barth JA, editor. Leipzig; 1909.
- 6. Conel JL. Vol. 1. The cortex of the newborn. The postnatal development of the human cerebral cortex-Title. Oxford, England: Harvard Univ. Press; 1939.
- Travis K, Ford K, Jacobs B. Regional dendritic variation in neonatal human cortex: A quantitative Golgi study. Dev Neurosci. 2005; 27: 277–287. https://doi.org/10.1159/000086707 PMID: 16137985
- Petanjek Z, Judas M, Simic G, Rasin MR, Uylings HBM, Rakic P, et al. Extraordinary neoteny of synaptic spines in the human prefrontal cortex. Proc Natl Acad Sci U S A. 2011; 108: 13281–13286. https://doi.org/10.1073/pnas.1105108108 PMID: 21788513
- 9. Flechsig Of Leipsic P. DEVELOPMENTAL (MYELOGENETIC) LOCALISATION OF THE CEREBRAL CORTEX IN THE HUMAN SUBJECT. Lancet. 1901; 158: 1027–1030.
- Miller DJ, Duka T, Stimpson CD, Schapiro SJ, Baze WB, McArthur MJ, et al. Prolonged myelination in human neocortical evolution. Proc Natl Acad Sci. 2012; 109: 16480–16485. <u>https://doi.org/10.1073/</u> pnas.1117943109 PMID: 23012402
- Levine D, Barnes PD. Cortical maturation in normal and abnormal fetuses as assessed with prenatal MR imaging. Radiology. 1999; 210: 751–8. <u>https://doi.org/10.1148/radiology.210.3.r99mr47751</u> PMID: 10207478
- 12. Flechsig PE. Anatomie des menschlichen Gehirns und Rückenmarks auf myelogenetischer Grundlage (Vol. 1). Thieme G, editor. 1920.
- Yakovlev PI, Lecours A-R. The myelogenetic cycles of regional maturation of the brain. Regional development of the brain in early life. 1967. pp. 3–70.
- Kinney HC, Brody BA, Kloman AS, Gilles FH. Sequence of central nervous system myelination in human infancy. II. Patterns of myelination in autopsied infants. J Neuropathol Exp Neurol. 1988; 47: 217–234. PMID: 3367155
- Deoni SCL, Mercure E, Blasi A, Gasston D, Thomson A, Johnson M, et al. Mapping infant brain myelination with magnetic resonance imaging. J Neurosci. 2011; 31: 784–91. https://doi.org/10.1523/ JNEUROSCI.2106-10.2011 PMID: 21228187
- Goldman-Rakic PS, Rakic P. Experimental modification of gyral patterns. In: Geschwind N, Galaburda A, editors. Cerebral dominance: the biological foundations. Harvard University Press; 1984. pp. 179– 192.

- 17. Van Essen DC. A tension-based theory of morphogenesis and compact wiring in the central nervous system. Nature. 1997; 385: 313–318. https://doi.org/10.1038/385313a0 PMID: 9002514
- Melbourne A, Kendall GS, Cardoso MJ, Gunny R, Robertson NJ, Marlow N, et al. Preterm birth affects the developmental synergy between cortical folding and cortical connectivity observed on multimodal MRI. Neuroimage. The Authors; 2014; 89: 23–34.
- Nagy Z, Westerberg H, Klingberg T. Maturation of white matter is associated with the development of cognitive functions during childhood. J Cogn Neurosci. 2004; 16: 1227–33. <u>https://doi.org/10.1162/ 0898929041920441</u> PMID: 15453975
- Pujol J, Soriano-Mas C, Ortiz H, Sebastián-Gallés N, Losilla JM, Deus J. Myelination of languagerelated areas in the developing brain. Neurology. 2006; 66: 339–343. https://doi.org/10.1212/01.wnl. 0000201049.66073.8d PMID: 16476931
- Goodman CS, Shatz CJ. Developmental mechanisms that generate precise patterns of neuronal connectivity. Cell. 1993; 72: 77–98. PMID: 8428376
- Wong ROL, Ghosh A. Activity-dependent regulation of dendritic growth and patterning. Nat Rev Neurosci. 2002; 3: 803–812. https://doi.org/10.1038/nrn941 PMID: 12360324
- Penn AA, Shatz CJ. Brain Waves and Brain Wiring: The Role of Endogenous and Sensory-Driven Neural Activity in Development. Pediatr Res. 1999; 45: 447–458. <u>https://doi.org/10.1203/00006450-199904010-00001</u> PMID: 10203134
- 24. Kostović I, Judaš M. The development of the subplate and thalamocortical connections in the human foetal brain. Acta Paediatr Int J Paediatr. 2010; 99: 1119–1127.
- Hanson MG, Landmesser LT. Characterization of the circuits that generate spontaneous episodes of activity in the early embryonic mouse spinal cord. J Neurosci. 2003; 23: 587–600. PMID: 12533619
- Khazipov R, Luhmann HJ. Early patterns of electrical activity in the developing cerebral cortex of humans and rodents. Trends Neurosci. 2006; 29: 414–418. https://doi.org/10.1016/j.tins.2006.05.007 PMID: 16713634
- Hanganu-Opatz IL. Between molecules and experience: Role of early patterns of coordinated activity for the development of cortical maps and sensory abilities. Brain Res Rev. Elsevier B.V.; 2010; 64: 160–176.
- Kilb W, Kirischuk S, Luhmann HJ. Electrical activity patterns and the functional maturation of the neocortex. Eur J Neurosci. 2011; 34: 1677–1686. <u>https://doi.org/10.1111/j.1460-9568.2011.07878.x</u> PMID: 22103424
- 29. Wake H, Lee PR, Fields RD. Control of Local Protein Synthesis and Initial Events in Myelination by Action Potentials. Science (80-). 2011; 333: 1647–1651.
- Gibson EM, Purger D, Mount CW, Goldstein AK, Lin GL, Wood LS, et al. Neuronal Activity Promotes Oligodendrogenesis and Adaptive Myelination in the Mammalian Brain. Science (80-). 2014; 344: 1252304–1252304.
- Hubel DH, Wiesel TN. The period of susceptibility to the physiological effects of unilateral eye closure in kittens. J Physiol. 1970; 206: 419–36. PMID: 5498493
- von Melchner L, Pallas SL, Sur M. Visual behaviour mediated by retinal projections directed to the auditory pathway. Nature. 2000; 404: 871–876. https://doi.org/10.1038/35009102 PMID: 10786793
- Jiao Y. Major Effects of Sensory Experiences on the Neocortical Inhibitory Circuits. J Neurosci. 2006; 26: 8691–8701. https://doi.org/10.1523/JNEUROSCI.2478-06.2006 PMID: 16928857
- Chittajallu R, Isaac JTR. Emergence of cortical inhibition by coordinated sensory-driven plasticity at distinct synaptic loci. Nat Neurosci. 2010; 13: 1240–1248. https://doi.org/10.1038/nn.2639 PMID: 20871602
- Ko H, Cossell L, Baragli C, Antolik J, Clopath C, Hofer SB, et al. The emergence of functional microcircuits in visual cortex. Nature. 2013; 496: 96–100. <u>https://doi.org/10.1038/nature12015</u> PMID: 23552948
- Hüppi PS, Maier SE, Peled S, Zientara GP, Barnes PD, Jolesz FA, et al. Microstructural development of human newborn cerebral white matter assessed in vivo by diffusion tensor magnetic resonance imaging. Pediatr Res. 1998; 44: 584–90. https://doi.org/10.1203/00006450-199810000-00019 PMID: 9773850
- McKinstry RC, Mathur A, Miller JH, Ozcan A, Snyder AZ, Schefft GL, et al. Radial organization of developing preterm human cerebral cortex revealed by non-invasive water diffusion anisotropy MRI. Cereb Cortex. 2002; 12: 1237–43. PMID: 12427675
- Delpolyi AR, Mukherjee P, Gill K, Henry RG, Partridge SC, Veeraraghavan S, et al. Comparing microstructural and macrostructural development of the cerebral cortex in premature newborns: Diffusion tensor imaging versus cortical gyration. Neuroimage. 2005; 27: 579–586. <u>https://doi.org/10.1016/j.neuroimage.2005.04.027</u> PMID: 15921934

- Deoni SCL, Dean DC, O'Muircheartaigh J, Dirks H, Jerskey B a. Investigating white matter development in infancy and early childhood using myelin water faction and relaxation time mapping. Neuroimage. Elsevier Inc.; 2012; 63: 1038–53.
- 40. Schneider J, Kober T, Bickle Graz M, Meuli R, Hu ppi PS, Hagmann P, et al. Evolution of T1 Relaxation, ADC, and Fractional Anisotropy during Early Brain Maturation: A Serial Imaging Study on Preterm Infants. Am J Neuroradiol. 2016; 37: 155–162. https://doi.org/10.3174/ajnr.A4510 PMID: 26494693
- Dubowitz LM, Dubowitz V, Palmer P, Verghote M. A new approach to the neurological assessment of the preterm and full-term newborn infant. Brain Dev. 1980; 2: 3–14. PMID: 7416439
- Marques JP, Kober T, Krueger G, van der Zwaag W, Van de Moortele P-F, Gruetter R. MP2RAGE, a self bias-field corrected sequence for improved segmentation and T1-mapping at high field. Neuroimage. Elsevier Inc.; 2010; 49: 1271–81.
- Shi F, Yap P-T, Wu G, Jia H, Gilmore JH, Lin W, et al. Infant brain atlases from neonates to 1- and 2year-olds. PLoS One. 2011; 6: e18746. <u>https://doi.org/10.1371/journal.pone.0018746</u> PMID: 21533194
- Ashburner J, Friston KJ. Voxel-based morphometry—the methods. Neuroimage. 2000; 11: 805–21. https://doi.org/10.1006/nimg.2000.0582 PMID: 10860804
- Andersson JLR, Sotiropoulos SN. An integrated approach to correction for off-resonance effects and subject movement in diffusion MR imaging. Neuroimage. 2016; 125: 1063–1078. <u>https://doi.org/10.1016/j.neuroimage.2015.10.019 PMID: 26481672</u>
- Wang R, Benner T, Sorensen A, Wedeen VJ. Diffusion toolkit: a software package for diffusion imaging data processing and tractography. Proc Intl Soc Mag Reson Med. 2007; 15: 3720.
- Hagmann P, Kurant M, Gigandet X, Thiran P, Wedeen VJ, Meuli R, et al. Mapping human whole-brain structural networks with diffusion MRI. PLoS One. 2007; 2: e597. https://doi.org/10.1371/journal.pone. 0000597 PMID: 17611629
- **48.** Daducci A, Gerhard S, Griffa A, Lemkaddem A, Cammoun L, Gigandet X, et al. The connectome mapper: an open-source processing pipeline to map connectomes with MRI. PLoS One. 2012; 7: e48121. https://doi.org/10.1371/journal.pone.0048121 PMID: 23272041
- de Reus MA, van den Heuvel MP. Estimating false positives and negatives in brain networks. Neuroimage. Elsevier Inc.; 2013; 70: 402–409.
- Löbel U, Sedlacik J, Güllmar D, Kaiser W a, Reichenbach JR, Mentzel H-J. Diffusion tensor imaging: the normal evolution of ADC, RA, FA, and eigenvalues studied in multiple anatomical regions of the brain. Neuroradiology. 2009; 51: 253–63. https://doi.org/10.1007/s00234-008-0488-1 PMID: 19132355
- Hagmann P, Sporns O, Madan N, Cammoun L, Pienaar R, Wedeen VJ, et al. White matter maturation reshapes structural connectivity in the late developing human brain. Proc Natl Acad Sci U S A. 2010; 107: 19067–72. https://doi.org/10.1073/pnas.1009073107 PMID: 20956328
- Beaulieu C. The basis of anisotropic water diffusion in the nervous system—a technical review. NMR Biomed. 2002; 15: 435–55. https://doi.org/10.1002/nbm.782 PMID: 12489094
- Barkovich AJ, Kjos BO, Jackson DE, Norman D. Normal maturation of the neonatal and infant brain: MR imaging at 1.5 T. Radiology. 1988; 166: 173–180. https://doi.org/10.1148/radiology.166.1. 3336675 PMID: 3336675
- 54. Deoni SCL, Dean DC, Remer J, Dirks H, O'Muircheartaigh J. Cortical maturation and myelination in healthy toddlers and young children. Neuroimage. Elsevier B.V.; 2015; 115: 147–161.
- Marques JP, Gruetter R. New Developments and Applications of the MP2RAGE Sequence—Focusing the Contrast and High Spatial Resolution R1 Mapping. PLoS One. 2013; 8.
- 56. Newman MEJ. Networks: An Introduction. Oxford University Press; 2010.
- 57. Bewick V, Cheek L, Ball J. Statistics review 10: further nonparametric methods. Crit Care. 2004; 8: 196–199. https://doi.org/10.1186/cc2857 PMID: 15153238
- Rubinov M, Sporns O. Complex network measures of brain connectivity: uses and interpretations. Neuroimage. Elsevier Inc.; 2010; 52: 1059–69.
- Barkovich a. J. MR of the normal neonatal brain: assessment of deep structures. AJNR Am J Neuroradiol. 1998; 19: 1397–403. PMID: 9763367
- Nossin-Manor R, Chung a. D, Whyte HE a., Shroff MM, Taylor MJ, Sled JG. Deep Gray Matter Maturation in Very Preterm Neonates: Regional Variations and Pathology-related Age-dependent Changes in Magnetization Transfer Ratio. Radiology. 2012; 263: 510–517. <u>https://doi.org/10.1148/radiol.</u> 12110367 PMID: 22416249
- Steriade M, Jones E, McCormick D. Thalamus: v. 1. Organisation and function. Amsterdam: Amsterdam; New York: Elsevier, 1997.; 1997.

- Hirata A, Castro-Alamancos M a. Neocortex Network Activation and Deactivation States Controlled by the Thalamus. J Neurophysiol. 2010; 103: 1147–1157. https://doi.org/10.1152/jn.00955.2009 PMID: 20053845
- Lerch JP, Worsley K, Shaw WP, Greenstein DK, Lenroot RK, Giedd J, et al. Mapping anatomical correlations across cerebral cortex (MACACC) using cortical thickness from MRI. Neuroimage. 2006; 31: 993–1003. https://doi.org/10.1016/j.neuroimage.2006.01.042 PMID: 16624590
- He Y, Chen ZJ, Evans AC. Small-world anatomical networks in the human brain revealed by cortical thickness from MRI. Cereb Cortex. 2007; 17: 2407–19. https://doi.org/10.1093/cercor/bhl149 PMID: 17204824
- 65. Sanabria-Diaz G, Melie-García L, Iturria-Medina Y, Alemán-Gómez Y, Hernández-González G, Valdés-Urrutia L, et al. Surface area and cortical thickness descriptors reveal different attributes of the structural human brain networks. Neuroimage. Elsevier Inc.; 2010; 50: 1497–510.
- Raznahan A, Lerch JP, Lee N, Greenstein D, Wallace GL, Stockman M, et al. Patterns of Coordinated Anatomical Change in Human Cortical Development: A Longitudinal Neuroimaging Study of Maturational Coupling. Neuron. Elsevier Inc.; 2011; 72: 873–884.
- Barton RA, Harvey PH. Mosaic evolution of brain structure in mammals. Nature. 2000; 405: 1055– 1058. https://doi.org/10.1038/35016580 PMID: 10890446
- Smyser TA, Smyser CD, Rogers CE, Gillespie SK, Inder TE, Neil JJ. Cortical Gray and Adjacent White Matter Demonstrate Synchronous Maturation in Very Preterm Infants. Cereb Cortex. 2015; bhv164.
- Croteau-Chonka EC, Dean DC, Remer J, Dirks H, O'Muircheartaigh J, Deoni SCL. Examining the relationships between cortical maturation and white matter myelination throughout early childhood. Neuroimage. The Authors; 2016; 125: 413–421.
- 70. Leroy F, Glasel H, Dubois J, Hertz-Pannier L, Thirion B, Mangin J-F, et al. Early Maturation of the Linguistic Dorsal Pathway in Human Infants. J Neurosci. 2011; 31: 1500–1506. <u>https://doi.org/10.1523/ JNEUROSCI.4141-10.2011 PMID: 21273434</u>
- Giorgio a., Watkins KE, Douaud G, James a. C, James S, De Stefano N, et al. Changes in white matter microstructure during adolescence. Neuroimage. 2008; 39: 52–61. <u>https://doi.org/10.1016/j.</u> neuroimage.2007.07.043 PMID: 17919933
- 72. Jeon T, Mishra V, Ouyang M, Chen M, Huang H. Synchronous Changes of Cortical Thickness and Corresponding White Matter Microstructure During Brain Development Accessed by Diffusion MRI Tractography from Parcellated Cortex. Front Neuroanat. 2015; 9: 158. <u>https://doi.org/10.3389/fnana.</u> 2015.00158 PMID: 26696839
- 73. French L, Pavlidis P. Relationships between Gene Expression and Brain Wiring in the Adult Rodent Brain. Sporns O, editor. PLoS Comput Biol. 2011; 7: e1001049. <u>https://doi.org/10.1371/journal.pcbi.</u> 1001049 PMID: 21253556
- 74. White LE, Fitzpatrick D. Vision and Cortical Map Development. Neuron. 2007; 56: 327–338. <u>https://doi.org/10.1016/j.neuron.2007.10.011</u> PMID: 17964249
- 75. Kaas JH, Collins CE. The organization of sensory cortex. Curr Opin Neurobiol. 2001; 11: 498–504. PMID: 11502398
- 76. Van Essen DC. Corticocortical and thalamocortical information flow in the primate visual system. Prog Brain Res. 2005; 149: 173–85. https://doi.org/10.1016/S0079-6123(05)49013-5 PMID: 16226584
- 77. Raj A, Kuceyeski A, Weiner M. A Network Diffusion Model of Disease Progression in Dementia. Neuron. 2012; 73: 1204–1215. https://doi.org/10.1016/j.neuron.2011.12.040 PMID: 22445347
- **78.** Schmidt R, de Reus MA, Scholtens LH, van den Berg LH, van den Heuvel MP. Simulating disease propagation across white matter connectome reveals anatomical substrate for neuropathology staging in amyotrophic lateral sclerosis. Neuroimage. The Authors; 2015; 124: 762–769.
- 79. Guillery RW. Is postnatal neocortical maturation hierarchical? Trends Neurosci. 2005; 28: 512–517. https://doi.org/10.1016/j.tins.2005.08.006 PMID: 16126285
- Wierenga LM, van den Heuvel MP, van Dijk S, Rijks Y, de Reus MA, Durston S. The development of brain network architecture. Hum Brain Mapp. 2016; 37: 717–729. <u>https://doi.org/10.1002/hbm.23062</u> PMID: 26595445
- Gogtay N, Giedd JN, Lusk L, Hayashi KM, Greenstein D, Vaituzis AC, et al. Dynamic mapping of human cortical development during childhood through early adulthood. Proc Natl Acad Sci. 2004; 101: 8174–8179. https://doi.org/10.1073/pnas.0402680101 PMID: 15148381
- Pierson CR, Folkerth RD, Billiards SS, Trachtenberg FL, Drinkwater ME, Volpe JJ, et al. Gray matter injury associated with periventricular leukomalacia in the premature infant. Acta Neuropathol. 2007; 114: 619–631. https://doi.org/10.1007/s00401-007-0295-5 PMID: 17912538

- Andiman SE, Haynes RL, Trachtenberg FL, Billiards SS, Folkerth RD, Volpe JJ, et al. The Cerebral Cortex Overlying Periventricular Leukomalacia: Analysis of Pyramidal Neurons. Brain Pathol. 2010; 20: 803–814. https://doi.org/10.1111/j.1750-3639.2010.00380.x PMID: 20331617
- Dean JM, McClendon E, Hansen K, Azimi-Zonooz A, Chen K, Riddle A, et al. Prenatal Cerebral Ischemia Disrupts MRI-Defined Cortical Microstructure Through Disturbances in Neuronal Arborization. Sci Transl Med. 2013; 5: 168ra7–168ra7. https://doi.org/10.1126/scitranslmed.3004669 PMID: 23325800
- Ball G, Srinivasan L, Aljabar P, Counsell SJ, Durighel G, Hajnal J V, et al. Development of cortical microstructure in the preterm human brain. Proc Natl Acad Sci. 2013; 110: 9541–9546. <u>https://doi.org/ 10.1073/pnas.1301652110</u> PMID: 23696665
- Kiss JZ, Vasung L, Petrenko V. Process of cortical network formation and impact of early brain damage. Curr Opin Neurol. 2014; 27: 133–41. https://doi.org/10.1097/WCO.00000000000068 PMID: 24561871
- Volpe JJ. Brain injury in premature infants: a complex amalgam of destructive and developmental disturbances. Lancet Neurol. 2009; 8: 110–24. https://doi.org/10.1016/S1474-4422(08)70294-1 PMID: 19081519
- **88.** Paus T, Collins D., Evans A., Leonard G, Pike B, Zijdenbos A. Maturation of white matter in the human brain: a review of magnetic resonance studies. Brain Res Bull. 2001; 54: 255–266. PMID: <u>11287130</u>
- Roalf DR, Quarmley M, Elliott MA, Satterthwaite TD, Vandekar SN, Ruparel K, et al. The impact of quality assurance assessment on diffusion tensor imaging outcomes in a large-scale populationbased cohort. Neuroimage. 2016;
- Zatorre RJ, Fields R., Johansen-Berg H. Plasticity in Gray and White: Neuroimaging changes in brain structure during learning. Nat Neurosci. 2013; 15: 528–536.
- Dubois J, Dehaene-Lambertz G, Kulikova S, Poupon C, Hüppi PS, Hertz-Pannier L. The early development of brain white matter: a review of imaging studies in fetuses, newborns and infants. Neuroscience. 2014; 276: 48–71. https://doi.org/10.1016/j.neuroscience.2013.12.044 PMID: 24378955
- Stüber C, Morawski M, Schäfer A, Labadie C, Wähnert M, Leuze C, et al. Myelin and iron concentration in the human brain: a quantitative study of MRI contrast. Neuroimage. Elsevier Inc.; 2014; 93 Pt 1: 95–106.
- Kulikova S, Hertz-Pannier L, Dehaene-Lambertz G, Buzmakov A, Poupon C, Dubois J. Multi-parametric evaluation of the white matter maturation. Brain Struct Funct. Springer Berlin Heidelberg; 2015; 220: 3657–72.
- Kunz N, Zhang H, Vasung L, O'Brien KR, Assaf Y, Lazeyras F, et al. Assessing white matter microstructure of the newborn with multi-shell diffusion MRI and biophysical compartment models. Neuroimage. Elsevier B.V.; 2014; 96: 288–299.
- Reveley C, Seth AK, Pierpaoli C, Silva AC, Yu D, Saunders RC, et al. Superficial white matter fiber systems impede detection of long-range cortical connections in diffusion MR tractography. Proc Natl Acad Sci. 2015; 201418198.
- 96. Thomas C, Ye FQ, Irfanoglu MO, Modi P, Saleem KS, Leopold DA, et al. Anatomical accuracy of brain connections derived from diffusion MRI tractography is inherently limited. Proc Natl Acad Sci U S A. 2014; 111: 16574–16579. https://doi.org/10.1073/pnas.1405672111 PMID: 25368179
- Zalesky A, Fornito A, Cocchi L, Gollo LL, van den Heuvel MP, Breakspear M. Connectome sensitivity or specificity: which is more important? Neuroimage. Elsevier Inc.; 2016;
- 98. Papile LA, Burstein J, Burstein R, Koffler H. Incidence and evolution of subependymal and intraventricular hemorrhage: a study of infants with birth weights less than 1,500 gm. J Pediatr. 1978; 92: 529–34. PMID: 305471
- de Vries LS, Eken P, Dubowitz LM. The spectrum of leukomalacia using cranial ultrasound. Behav Brain Res. 1992; 49: 1–6.
- Kidokoro H, Neil JJ, Inder TE. New MR imaging assessment tool to define brain abnormalities in very preterm infants at term. AJNR Am J Neuroradiol. 2013; 34: 2208–14. <u>https://doi.org/10.3174/ajnr.</u> A3521 PMID: 23620070
- Kidokoro H, Anderson PJ, Doyle LW, Woodward LJ, Neil JJ, Inder TE. Brain Injury and Altered Brain Growth in Preterm Infants: Predictors and Prognosis. Pediatrics. 2014; 134: e444–e453. https://doi. org/10.1542/peds.2013-2336 PMID: 25070300