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Reports on Polar and Marine Research

The Expedition PS107 of the Research Vessel POLARSTERN to the Fram Strait and the AWI-HAUSGARTEN in 2017

Edited by Ingo Schewe with contributions of the participants

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Titel: Polarstern unter einem Regenbogen (Foto: Jana Bäger, Alfred-Wegener-Institut, 16. August 2017)

Cover: Polarstern under the rainbow (Photo: Jana Bäger, Alfred Wegener Institute, 16th August 2017)

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PS107

(ARK-XXXI/2)

23 July 2017 – 19 August 2017

Tromsø - Tromsø

Chief scientist Ingo Schewe

Coordinator Rainer Knust

Contents

1. ÜBERBLICK UND FAHRTVERLAUF

Ingo Schewe AWI

Die wissenschaftlichen Arbeiten während der Expedition PS107 widmeten sich im wesentlichen der Fortführung der Langzeitbeobachtungen am LTER (Long-Term Ecological Research) Observatorium HAUSGARTEN mit Untersuchungsgebieten westlich von Spitzbergen und am Kontinentalhang vor Ost-Grönland (Abbildung 1.1). Sehr erleichtert wurden alle Arbeiten während der Expedition durch die diesjährigen, sehr ruhigen Witterungsbedingungen sowie die nur lockerere Eisbedeckung, so dass alle vor der Expedition geplanten Verankerungsarbeiten und Geräteeinsätze realisiert werden konnten. Die Arbeiten stellten einen weiteren Beitrag zur Sicherstellung der Langzeitbeobachtungen am LTER Observatorium HAUSGARTEN dar, in denen der Einfluss von Umweltveränderungen auf ein arktisches Tiefseeökosystem untersucht und dokumentiert wird. Diese Arbeiten wurden auch in diesem Jahr in enger Zusammenarbeit der HGF-MPG Brückengruppe für Tiefsee-Ökologie und -Technologie, der PEBCAO-Gruppe (Phytoplankton Ecology and Biogeochemistry in the Changing Arctic Ocean) des AWI und der Helmholtz-Hochschul-Nachwuchsgruppe SEAPUMP (Seasonal and regional food web interactions with the biological pump) durchgeführt. Die Expedition wurde außerdem genutzt, um eine Reihe von Geräten, Sensoren und Verankerungen des HGF Infrastruktur-Projektes FRAM (Frontiers in Arctic marine Monitoring) auszutauschen. Das FRAM Ozean-Beobachtungs-System ermöglicht eine ganzjährige Beobachtung von Ökosystemen im Arktischen Ozean von der Meereisdecke bis zum Meeresboden. Eine Komponente ist dabei ein Netz von stationären Verankerungen in der Framstraße, welche ganzjährig Daten zur Erdsystem-Dynamik, Klimavariabilität und den Veränderungen in den Ökosystemen sammelt. Damit erfüllt es nationale und internationale Aufgaben hin zu einem besseren Verständnis, welche Auswirkung Veränderungen in den Meeresströmungen und Wassermassenverteilungen sowie der Rückgang des Meereises auf die Funktion der marinen Ökosysteme in der Arktis haben. Der Einsatz bereits existierender und neu entwickelter Sensoren und Beobachtungs-Plattformen erlaubt die synchronisierte Erfassung von wichtigen Parametern zur Untersuchung physikalischer, chemischer und biologischer Prozesse im Ozean.

Kurzzeitig unterbrochen wurden die Arbeiten zu den Langzeitstudien in diesem Jahr für die Realisierung eines kleinen komplementären Programms zur Untersuchung von mesoskaligen Prozessen, an einem der typischen Frontensysteme in der zentralen Framstrasse. Zur hochauflösenden Erfassung des Frontensystems kamen neben einer geschleppten CTD und dem AUTOFIM die bordeigenen Sensoren der *Polarstern*, sowie der neue Web-GIS basierte Ice-Viewer – um eine derartige Front überhaupt erst einmal aufzuspüren - zum Einsatz. Ergänzt wurde dieses wissenschaftliche Programm durch Untersuchungen zur Sensitivität von arktischem Zooplankton auf potentielle Temperaturveränderungen und ob sich ökophysiologische Effekte bis auf die Ebene von Zooplankton-Gemeinschaften niederschlagen.

Insgesamt leisten die Erkenntnisse und Arbeiten der Expedition PS107 Beiträge zu verschiedenen nationalen und internationalen Forschungs- und Infrastrukturprojekten (INTAROS, FRAM, SIOS, OceanSITES) sowie dem Forschungsprogramm PACES II (Polar Regions and Coasts in the changing Earth System) des AWI. Im Arbeitspaket WP4 (Arctic sea ice and its interaction with ocean and ecosystems) des PACES-II Programms werden die mit dem Rückgang des Meereises verbundenen Ökosystemverschiebungen im Pelagial und im tiefen Ozean ermittelt und quantifiziert, und Rückkopplungsprozesse auf zeitliche und räumliche Prozesse untersucht.

Abb. 1.1: Haupt-Arbeitsgebiete und HAUSGARTEN Stationen während PS107. Siehe https://doi.pangaea.de/10.1594/PANGAEA.881581 für eine Darstellung des master tracks in Verbindung mit der Stationsliste.

Fig. 1.1: Main working areas and sampling sites during PS107. See https://doi.pangaea.de/10.1594/PANGAEA.881581 to display the master track in conjunction with

the list of stations.

SUMMARY AND ITINERARY

The scientific work during expedition PS107 substantially supported the time-series studies at the LTER (Long-Term Ecological Research) observatory HAUSGARTEN, where we document Global Change induced environmental variations on a polar deep-water ecosystem. The work was mainly carried out in research areas westerly off Svalbard and at the East-Greenland continental slope (Fig. 1.1). Due to the calm weather situation as well as the scattered ice conditions this year working on board was very easy. On the whole the expedition PS107 was extremely successful. All mooring and station work was carried out as previously planned prior to the expedition. This expedition supported extensively the time-series studies at the LTER (Long-Term Ecological Research) observatory HAUSGARTEN, where we document Global Change induced environmental variations on a polar deep-water ecosystem. This work is carried out in close co-operation between the HGF-MPG Joint Research Group on Deep-Sea Ecology and Technology, the PEBCAO Group (Phytoplankton Ecology and Biogeochemistry in the Changing Arctic Ocean) at AWI and the Helmholtz Young Investigators Group SEAPUMP (Seasonal and regional food web interactions with the biological pump), representing a joint effort between the AWI, the MARUM - Center for Marine Environmental Sciences, and the University of Bremen. The expedition was also dedicated to accomplish installations for the HGF infrastructure project FRAM (Frontiers in Arctic marine Monitoring). The FRAM Ocean Observing System aims at permanent presence at sea, from ocean surface to the deep-sea, for the provision of near real-time data on Earth system dynamics, climate variability and ecosystem change. It serves national and international tasks towards a better understanding of the effects of change in ocean circulation, water mass properties and sea-ice retreat on Arctic marine ecosystems and their main functions and services. FRAM implements existing and next-generation sensors and observatory platforms, allowing synchronous observation of relevant ocean variables as well as the study of physical, chemical and biological processes in the ocean. Experimental and event-triggered platforms complement the observational platforms.

We shortly interrupted our long-tern studies for realizing a complementing programme with the aim to investigate mesoscale processes along one of those typical frontal systems in the central Fram Strait. For high resolution surveying of the frontal system we used besides the Underway-CTD and the AUTOFIM system the on-board sensors of *Polarstern* as well as the new web-GIS based Ice-Viewer, for - first of all - tracing such kind of front. The scientific programme was complemented by investigations for the sensitivity of Arctic zooplankton to potential temperature changes and its effects on ecophysiological level as well responses of the zooplankton community. In any case, also for our long-term investigations in this area it is of exceptional importance to get a holistic picture of processes within the different ecosystems. It will become most exciting when all analyses in our home laboratories will be accomplished and we will start to join all puzzle pieces to get them into the context of results from former years.

As a whole all those insights and operations of the expedition PS107 mean a contribution to various national and international research- and infrastructure-projects like INTAROS, SIOS, OceanSITES as well as to AWIs own research programme PACES-II (Polar Regions and Coasts in the changing Earth System).

2. WEATHER CONDITIONS DURING PS107

Jens Kieser, Juliane Hempelt **DWD**

23.7.2017: In the evening of the 23rd of July *Polarstern* left Tromsø heading towards the research area 'HAUSGARTEN' within the Fram Strait. The cruising area was situated under a high pressure zone extending from the Barents Sea to the sea area of Iceland. Mostly light winds were blowing from northerly or easterly directions. Due to the cold water of the fjords the air temperature was not much higher than 10°C. Visibility was good.

24./25.7.2017: On 24 July research vessel *Polarstern* passed the axis of the high pressure zone mentioned above. After a period of light motion of air in the morning winds increased to 4 or 5 Bft and shifted to southwesterly directions as the cruising area got between the high pressure zone and a trough over the Greenland Sea. Initially fog was present, later low level clouds dominated. Air temperature dropped from 12°C to 6°C.

From the trough mentioned above a low was moving eastward across Svalbard on the $25th$. Associated low level clouds influenced the cruising area. Afterwards the ridge of an Iceland high passed the Fram Strait eastward. Light to moderate winds shifted to north for a while. Fog patches were present. The temperature of air declined further on from 8°C to -1°C. Water temperatures about 7°C were measured at first but later we reached an area where water temperatures of 0°C were present.

26.-28.7.2017: During the following days the working area in the 'HAUSGARTEN' was influenced by a low that moved from the sea area north of Greenland to the Barents Sea. In front of the low southerly winds were temporarily blowing strong (6 Bft). On 27 July winds shifted from southwest to north and strengthened up again to 5 Bft. Low level clouds with light drizzle or grain dominated. Due to the presence of fog patches visibility was remarkably reduced. An exception was the $27th$ when the sky was clear for a longer time. At first air temperatures between 0°C and 4°C were measured, later -2°C to 1°C. Significant wave heights of more than 0.5 m were not observed.

29.7.-4.8.2017: During the following days till 4 August a series of weakly pronounced high pressure ridges and low pressure troughs crossed the working area 'HAUSGARTEN' eastward. Mostly light to moderate winds blew from variable directions. Air temperatures between -3°C and +2°C and water temperatures between -1°C and +3°C were measured. Foggy and sunny phases alternated as well as good and poor visibilities. Significant wave heights of more than a half meter were scarcely reached.

5.8./6.8.2017: On 5 August a weakly pronounced low pressure system established over the southern Fram Strait, while north of the working area a high pressure zone built. Due to the constellation of the pressure fields northeasterly to easterly winds result over the working area, temporarily reaching 6 Bft. Mist or fog dominated. The temperature of the ambient air was -2°C to 0°C. The significant wave height didn't exceed 0.5 m.

7.8.-9.8.2017: On 7 August the low pressure system moved slowly to the northern Fram Strait where it gradually filled. Simultaneously a trough was building southward from the Arctic Ocean along the coast of Greenland. The working area was situated at the eastern flank of the trough within an extensive southerly or southwesterly air current carrying moist air into the working area. An overcast sky with low clouds was present. Visibility was reduced by fog, drizzle and freezing drizzle. A southerly or southwesterly wind was blowing mostly at 4 Bft, interrupted by periods of light winds. The temperature of the air varied between -2°C and +1°C, the temperature of water between -1°C and +1°C. Sea state was marginal.

10.8.-13.8.: The general weather situation started to change on 10 August. At first a high pressure ridge crossed the research area, while a low pressure system formed over the Norwegian Sea and Iceland. Initially wind was blowing light to moderate from south or southeast. On 11 August wind turned to northeast and freshened up. An intense cyclone embedded within the low pressure system mentioned above was located over the central Norwegian Sea on 12 August. At this time a secondary low developed over southern Sweden. Winds of 5 Bft were observed nearly during the entire 12 August. At this time cloud conditions changed between nearly overcast periods, short sunny moments and foggy episodes. Temperatures around the freezing point were observed.

On the 13^th we cruised in a more eastern area where water temperatures of 6° C to 8° C were measured. Air temperature increased a bit and visibility improved significantly. A light to moderate wind blew from north. From the southeast a swell with significant height of 3 m propagated into the research area.

14.8.-16.8.: On 14 August an intense cyclone was still located over the northern Norwegian Sea. It was weakening, while the secondary low mentioned above moved across the Barents Sea and took over the role of the new central low. The cruising area got on the northeasterly edge of the low pressure area and wind shifted to north or northwest. It increased again up to 5 Bft. The north-northwesterly wind carried a cooler air mass into the research area close west of Spitzbergen. Temperatures dropped from 7° C to 0 $^{\circ}$ C in the afternoon of the 14th. Simultaneously the visibility became poor. Water temperatures about 7°C or 8°C were measured further on.

On 15 August the low pressure system over the Barents Sea temporarily lost its influence on the cruising area. *Polarstern* cruised close west of Svalbard and later inside of the archipelago. Wind calm down for a while, as a weak intermittent high influenced the cruising area. In the evening of the 15th a new low approached from the Fram Strait. The low passed *Polarstern* and Spitzbergen in the morning of 16 August. At the rear of the low wind increased considerably. Strong to near gale force winds (6-7 Bft) from west later from northwest blew. In the exit of the fjord west of Spitzbergen *Polarstern* steamed through rough sea with significant wave heights up to 4 m. During the 16th rain and drizzle occurred over the fjords near Longyearbyen and showery precipitation in the evening when we left Spitzbergen. Inside the fjords temperatures between 4 and 8°C were measured.

17.-19.8.: At first strong northwesterly winds blew on the edge of a low pressure system centered over the Kara Sea. The air mass was relatively dry and cool with temperatures between 3°C and 6°C. During the night to the 18th *Polarstern* crossed a high pressure zone extending between a high over Greenland and a further high over Eastern Europe. Afterwards we sailed on the northern edge of a low pressure area located over the Norwegian Sea and Scandinavia. Wind veered to east and increased to 6 Bft. Visibility was good. Northwest of the Fjords near Tromsø air temperatures about 11°C were reached corresponding with the sea surface temperatures. During the passage from Spitzbergen to Norway significant wave heights between 2 and 3 m were observed. Wind calm down as we entered the fjords near Tromsø. In the morning of 19 August *Polarstern* reached Tromsø with light winds and temperatures between 12°C and 15°C.

For further statistics see attached Figs. $2.1 - 2.5$.

Fig. 2.1: Distribution of wind force during PS107

Fig. 2.2: Distribution of wind direction during PS107

Fig. 2.3: Distribution of visibility during PS107

Fig. 2.4: Distribution of cloud coverage during PS107

Fig. 2.5: Distribution of ceiling during PS107

3. MEASUREMENTS OF ATMOSPHERIC WATER VAPOR, AEROSOL AND THIN CLOUDS USING FT SPECTROSCOPY IN THE INFRARED

Philipp Richter not on board: Mathias Palm, Christine Weinzierl, Justus Notholt, Matthias Buschmann

Universität Bremen

Grant No. AWI_PS107_03

Objectives

This project is embedded in the project AC3, which aims at understanding the enhanced warming in the Arctic. Clouds and atmospheric feedback are believed to play a major role.

We used mobile FTIR spectrometers onboard the *Polarstern* cruise PS106 (ARK XXXI/1, ARK XXXI/2) and PS107 (ARK XXXII) in the Arctic to study the geographical and temporal distribution of H₂O, HDO, thin clouds and aerosol in the Arctic. Together with the same suite of measurements at the AWIPEV research base in Ny-Ålesund, Spitsbergen the measurements will allow us to assess the representatively of the supersite in Ny-Ålesund for the Arctic.

a) Measurements of columnar H_2O and HDO

Solar absorption spectroscopy in the infrared can be used to determine the distribution of many infrared trace gases in the atmosphere. In particular it is possible to measure ${\sf H}_{\sf 2}$ O in high quality (Palm, 2008) and the isotopic composition H_2O in the troposphere (Schneider, 2006). The isotopic ratio of H_2O and HDO can be used to study the history of the sampled air parcels (Frankenberg, 2009).

b) Measurements of properties of aerosol layers and thin clouds

Emission spectroscopy in the infrared can be used to study the composition of aerosols and properties of thin clouds (Rathke, 2000a; Rathke, 2000b). Since the self-emission of the atmosphere is measured, those measurements are independent of an external light source like moon or sun.

Work at sea

Two FTIR instruments are housed in a custom build container for measurements in emission and solar absorption mode. The measurements were performed whenever weather conditions permited, i.e. clear sky for solar absorption measurements and dry conditions (no precipitation) for emission measurements. The performance of the measurements requires manual operation and oversight; therefore two participants took over to cover most of day and night.

Preliminary (expected) results

1. Low altitude resolution profiles of $H₂O$ and HDO and their ratio will be derived from the solar absorption measurements. A trajectory model will be used to track the path of the air-parcels measured.

2. The structure of radiation emitted from the atmosphere is analyzed to derive properties like the optical depth, the mean radius, and chemical composition of particles contained in clouds or aerosol layers.

Fig. 3.1: Spectra recorded on the 2nd of August 2017 at two different conditions: cirrus (blue) and altocumulus (orange)

Fig. 3.1 shows two spectra recorded in emission mode for different conditions: cirrus clouds to clear sky (08:41, blue color) and a thin cloud layer of altocumulus at an altitude of around 4,830 m (17:01, orange colour). The imminent effect of a thin cloud is the attenuation of the radiation coming from above and adding radiation, a baseline, due to its temperature. The shape of the baseline contains information about the liquid water path, the optical depth and the mean radius of the droplets in case of water clouds. If there is an effect due to ice is subject of a PhD study.

Data management

The obtained data will be made available to the project partners via the database PANGAEA and made available for the public after publication or 5 years at most.

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4. PHYSICAL OCEANOGRAPHY OF FRONTS IN FRAM STRAIT

Wilken-Jon von Appen¹, Laura Hehemann¹, Vibe Schourup-Kristensen¹, Claudia Wekerle¹ 1 AWI

Grant No. AWI_PS107_04

Objectives

Physical changes in the polar oceans caused by climate change, such as the sea-ice retreat in the Arctic Ocean or the increase in wind forcing in the Southern Ocean, will impact oceanic dynamics, particularly on scales of ~1–10 km ("submesoscale"). Those motions produce the largest vertical water velocities and are most effective at regulating the upper ocean vertical density gradients ("stratification"), thereby directly driving the light and nutrient availability to plankton in the upper 50 m.

Vertical motions in the ocean are important for primary production because they move both phytoplankton and nutrients into and out of the euphotic layer. Specifically, in post-bloom summer conditions, vertical upward nutrient transport from depth into the euphotic layer can sustain phytoplankton growth. It is thus crucial to determine what conditions are conducive to vertical motions. Large scale organized motions lead to advective transport where properties are moved along with the same water mass for significant distances. Small scale random motions (turbulent mixing), on the other hand, lead to turbulent transport where properties are consecutively transferred between adjacent water masses. Thus both vertical advective transports and vertical turbulent transports (mixing) can change the phytoplankton and nutrient distributions.

A fundamental scale in physical oceanography is the Rossby radius, which is the horizontal scale over which horizontal oceanic motions are roughly uniform. The first baroclinic Rossby radius Rd = NH/f decreases with the Coriolis parameter f, which is related to the Earth's rotation, while it scales linearly with the stratification N and the water depth H. If horizontal motions are confined to the mixed layer depth hML rather than the full water depth H, they have a horizontal scale of the mixed-layer Rossby radius RML = NhML/f. Mesoscale motions with horizontal scales comparable to Rd, which ranges from ~100km in low latitudes to as little as \sim 5 km in the polar oceans (von Appen et al., 2016), tend to be associated with weak advective vertical velocities. In contrast, submesoscale motions, with horizontal scales of RML, which ranges from \sim 10 km in low latitudes to \sim 1 km in the polar oceans, can lead to large advective vertical velocities (Thomas, 2008) such as in frontal. Thus submesoscale motions can drive large vertical advective property transports. Submesoscale motions can also strongly affect stratification and thereby either increase or decrease vertical turbulent property transports (Klein and Lapeyre, 2009).

For these reasons, the cruise attempted to sample submesoscale fronts, where such vertical motions are expected to be especially strong, with consequences for biology and the redistribution and mixing of Atlantic Water and Polar Water in Fram Strait.

Work at sea

CTD All information on CTD casts are provided in Tables 4.1 - 4.4.

Station name	Station # Latitude PS107/		Longitude	Max depth [m]	In-situ pumps	Salt		Config. Comments
S-III	002/01	78° 36.151' N	005° 01.517' E	2293	X	X	$\mathbf{1}$	
S-III	002/04	78° 36.258' N	005° 00.248' E	10			1	X
S-III	002/13	78° 36.496' N	005° 03.325' E	251			$\mathbf{1}$	
HG-IV	006/03	79° 03.864' N	004° 10.865' E	2404	X	X	1	X
HG-IV	006/08	79° 03.905' N	004° 10.087' E	100			$\mathbf{1}$	X
HG-V	007/01	79° 03.198' N	003° 44.980' E	1500		X	1	
HG-VI	008/01	79° 02.788' N	003° 36.351' E	250			$\mathbf{1}$	X
Front	010/04	78° 58.610' N	002° 29.658' E	500			1	
Front	012/03	78° 56.689' N	002° 42.102' E	500			$\mathbf{1}$	
Front	014/01	78° 55.622' N	002° 51.016' E	506			1	X
Front	016/03	79° 00.273' N	002° 17.015' E	500			$\mathbf{1}$	
Front	018/03	78° 59.153' N	002° 45.347' E	1500		X	1	X
HG-VII	019/01	79° 03.561' N	003° 28.918' E	1500		X	$\mathbf{1}$	X
HG-IX	020/01	79° 07.407' N	002° 49.132' E	5513	X	X	$\mathbf{1}$	X
HG-IX	020/08	79° 08.013' N	002° 50.686' E	400			$\overline{2}$	X
Station 0°E	021/01	78° 57.856' N	000° 00.228' E	1500		X	$\overline{2}$	
Station 0°E	021/05	78° 57.706' N	000° 02.377' E	300			$\overline{2}$	
EG-IV	022/01	78° 49.000' N	002° 48.039' W	2530	X	X	$\overline{2}$	X
EG-IV	022/06	78° 49.129' N	002° 43.015' W	251			$\overline{2}$	
EG-III	024/01	78° 51.345' N	003° 56.499' W	1944		X	$\overline{2}$	X
HG-EGC mooring	025/01	78° 49.867' N	002° 47.511' W	250			2	X
EG-II	028/01	78° 56.019' N	004° 38.088' W	1507		X	$\overline{2}$	X
EG-I	029/01	78° 59.693' N	005° 28.411' W	944	X	X	$\overline{2}$	X
Auel I	030/01	79° 18.089' N	001° 59.776' W	1500		X	$\overline{2}$	X
Auel II	031/01	79° 26.035' N	000° 00.160' E	1500		X	$\overline{2}$	X
Auel III	032/01	79° 34.928' N	002° 00.709' E	1500		X	$\overline{2}$	
N-IV	033/01	79° 44.256' N	004° 26.102' E	2560		X	$\sqrt{2}$	X
N-IV	033/06	79° 43.632' N	004° 30.175' E	250			$\overline{2}$	
N-V	034/01	79° 58.189' N	002° 54.400' E	2563	X	Χ	$\mathbf 2$	
N-V	034/05	80° 00.019' N	002° 56.414' E	250			2	
N-III	036/01	79° 35.268' N	005° 10.327' E	2720	X	X	$\sqrt{2}$	X
HG-III	037/01	79° 06.491' N	004° 35.961' E	1876		X	2	
HG-II	042/01	79° 07.845' N	004° 54.174' E	1503		X	$\overline{2}$	X
HG-II	042/05	79° 07.872' N	004° 54.353' E	200			2	X
HG-I	043/01	79° 08.385' N	006° 05.448' E	1247	X	X	$\sqrt{2}$	X
HG-I	043/05	79° 09.165' N	006° 06.915' E	250			2	
SV-IV	044/03	79° 02.044' N	007° 00.111' E	1268		X	$\sqrt{2}$	X
SV-III	045/01	79° 00.270' N	008° 21.819' E	740		X	2	X
SV-II	047/01	78° 58.974' N	009° 30.484' E	220			$\overline{2}$	
SV-I	048/01	79° 01.755' N	011° 06.101' E	272			\overline{c}	

Tab. 4.1: List of CTD casts. Comments to individual casts are given in Tab. 4.2

Station name	Comment
PS107/002/04	For Holger Auel's group.
PS107/006/03	T and S profiles have many spikes from 100 to 700 meters during downcast. Approximately 0.1 ml/L difference between the oxygen sensors. Upcast of T and S less spiky.
PS107/006/08	Beam transmission error. Value from 0 to 100 meters app. -1.3. At 100 meters, value equals 90.
PS107/008/01	Beam transmission error. Value ~-1.3.
PS107/014/01	Beam transmission error. Value ~-1.3. Last bottle closed after CTD was brought on deck and then back in the water. Extra CTD cast file for last bottle, named PS107_014_01_bottle24.hex.
PS107/018/03	Beam transmission reasonable after CTD was below 320 meters.
PS107/019/01	Beam transmission worked after 300 meters.
PS107/020/01	Conductivity2 and oxygen sensors broken during the cast.
PS107/020/08	Beam transmission calibration coefficients questionable. Jump in transmission at 100 meters depth of upcast. Both altimeter and beam transmission show spikes at 100 meters. Fluorescence -0.01 at depth.
PS107/022/01	Some difference in oxygen sensors between up- and downcast (~2nm drift of ship).
PS107/024/01	On upcast, all profiles have strange loop from 350 to 400 meters, but then return to profile from downcast. At the same depths the two sensors have large differences.
PS107/025/01	Cast for HG-EGC mooring.
PS107/028/01	Altimeter has many false readings during upcast.
PS107/029/01	Oxygen sensor disturbed after pumping. Irregular upcast profile with large difference to downcast. Does, however, have same profile shape as downcast. Sensors may be damaged, but were not changed.
PS107/030/01	Cast for Holger Auel's group. Oxygen looks reasonable again.
PS107/031/01	Cast for Holger Auel's group.
PS107/032/01	Cast for Holger Auel's group.
PS107/033/01	Beam transmission larger in upcast than downcast.
PS107/036/01	Sensor test carried out before the cast leading to two log sheets and two data files for cast; one for sensor test and one for standard CTD cast. Name of datafile for sensor test: PS107_036_01_PAR_sensor test. hex. Difference in temperature: T2-T1=0.001 for downcast and 0.000 for upcast. During upcast, CTD was taken to 50 meters, and then again lowered to 100 meters before bottle 2 and 3 were closed.
PS107/042/01	Testing of CTD-ADCP. Interference with altimeter signal leading to large errors in altimeter. Close to the bottom $(\sim]30$ m) the altimeter works as usual.
PS107/042/05	Water samples for Holger Auel's group.
PS107/043/01	Difference between sensors larger in upcats than downcast
PS107/044/03	LADCP on during cast leading to interference with altimeter. Altimeter works from ~50 meters above bottom.
PS107/045/01	Upcast salinity noisy.

Tab. 4.2: Comments to individual CTD casts

Tab. 4.3: CTD rosette configuration 1

Tab. 4.4: CTD rosette configuration 2

*: Winch EL31 was used until station PS107/021/05. Winch EL32 was used from station PS107/022/01 onwards.

UCTD

The winch for the underway CTD (UCTD) (serial number: 0204) was first mounted to the stern of the ship on the port side. A loop, created by splicing, was added to the end of the Kevlar line and was checked between UCTD casts in case of wear; it was re-spliced a few times when the loop appeared worn or burnt. Before the first cast and between each cast, the batteries were charged (3 hours) while the UCTD was sitting in a flask-container holding milliQ water and a few drops of Triton X-100. This was important, because, if the UCTD was dry, the conductivity sensor would have an offset during the first downcast.

To begin data gathering, the UCTD was connected to the computer using the command line terminal programme: UCTDterm. The data was processed following Ullman and Herbert (2014). After each transect, the data was removed from the UCTD to allow space for further data acquisition.

During the casts the ship speed exceeded no more than 7 knots and the target depth for each UCTD cast was approximately 200 m. A transect began with the removal of the start plug (to begin data capture) and was directly released from the stern of the ship into the water. Each cast had an 80-second drop time where the UCTD could descend until the target depth. The UCTD was wheeled in by the winch for approximately 4 minutes until it reached a distance of 5-10 m from the stern of the ship. The process was repeated for the length of each profile. In areas of sparse ice, the ship could maneuver around the ice floes, but when the ice became increasingly difficult to navigate, it was necessary to remove the UCTD from the water for a period of time until the ice concentration was thin enough to continue. When the ship went through and broke the ice, the ice tended to wrap around the ship leaving no clear path for the UCTD. This was extremely problematic for the thin Kevlar line and could result in easily damaging the UTCD. In situations of complete ice coverage, the ship captain would inform the deck crew in enough time to wind in the winch and pull the UCTD on deck. This occurred multiple times throughout the profiles, for a duration of anywhere from 5–20 min.

There were 6 transects in total, conducted at two target areas. Table 4.5 shows the station profile numbers, date/time, depth and location with each cast transect. The first target area was for a UCTD campaign that was carried out approximately 5 nm southwest of central Hausgarten heading towards an ocean front. The complete campaign ran from July 29, 2017 at a latitude of 78°00.91'N and a longitude 2.0351°0.91'E, until July 31, 2017 at a latitude of 78°00.97'N and a longitude of 2.0647°0.97'E. Fig. 4.1 shows the map of the casts for each transect across the front (A, B, C, and E). The second target area was for a transect running across the West Spitsbergen Current from central Hausgarten directly east to the most eastern station (SV-1). The complete transect was carried out from August 11, 2017 at a latitude of 79°00.05'N and a longitude of 3.6026°0.05'E and ended on August 14, 2017 at a latitude of 79°00.02'N and a longitude of 10.898°0.02'E. Fig. 3 shows the map of the casts for each transect $(1 – 7)$ that was carried out between Hausgarten stations.

A problematic issue was with the data storage itself. The UTCD failed to record a long transect (D) of three hours that repeated the transect A. After a series of tests conducted afterwards, the issue was determined to be only intermittent and could not be pin-pointed. Fortunately, the UCTD collected data for the remaining transects.

Tab. 4.5: UCTD casts

Vessel Mounted ADCP

Processing of the ship ADCP raw data was carried out with the Matlab package OSSI (Ocean surveyor sputum interpreter) Version 1.7. Determination of heading correction parameters was based on the time period 24 Jul – 30 Jul 2017. The following parameters were chosen: misalignment = 0.9, ampfactor = 1.02, tr_depth = 11 and pg_threshold =25. Note that water depths during this time period were deeper than 200 m. An averaging interval of 60 s was used for the data processing.

The processed ship ADCP data is saved in four matlab-files:

PS107_1_hc.mat: time period 24 Jul 2017 06:37 - 31 Jul 2017 14:33 PS107_2_hc.mat: time period 31 Jul 2017 19:36 - 14 Aug 2017 00:23 PS107_4_hc.mat: time period 14 Aug 2017 04:45 - 14 Aug 2017 06:29 PS107_6_hc.mat: time period 14 Aug 2017 06:33 - 15 Aug 2017 14:51

Problems on 14 Aug occurred due to synchronization of data from the sADCP computer to an external hard drive. If another program than VMDAS accesses the file to which VMDAS writes currently collected data, e.g. if a file synchronization tool accesses it, VMDAS produces an error and stops data acquisition. This results in lost data and ought to be avoided. Only when realized by e.g. the Labor ELO and manually restarted, does the raw data acquisition continues. VMDAS finishes writing to a file after the file size has been reached. During PS107, this was 10 MB (setting MaximumFileSize=10 in file VMDAS configuration). Then the 10 MB file can be accessed by another programme, e.g. for file synchronization. In the future, for near real time data analysis, we plan to access files much faster. Maximum file size then has to be set to e.g. 1 MB. Then it should be assured that synchronization problems don't occur. VMDAS settings used during PS107 are provided in Table 4.6.

```
;--------------------------------------------------------------------------
\lambda; ADCP Command File for use with VmDas software.
; 
; ADCP type: 150 Khz Ocean Surveyor 
; Setup name: for Polarstern in 6/2014
; Setup type: Low resolution, long range profile (Narrowband)
; 
; NOTE: Any line beginning with a semicolon in the first 
; column is treated as a comment and is ignored by 
; the VmDas software.
;
; NOTE: This file is best viewed with a fixed-point font (e.g. courier).
; Modified Last: 12Jun2014
;--------------------------------------------------------------------------
/
; Restore factory default settings in the ADCP
cr1
; set the data collection baud rate to 115200 bps, 
; no parity, one stop bit, 8 data bits
; NOTE: VmDas sends baud rate change command after all other commands in 
; this file, so that it is not made permanent by a CK command.
cb611
; Set for narrowband single-ping profile mode (NP), 100 (NN) 4 meter bins (NS), 
; 2 meter blanking distance (NF), 390 cm/s ambiguity vel (WV)
WP000
```
Tab. 4.6: VMDAS settings during PS107

NP001 NN080 NS0400 NF0400 ;WV390 ; Disable single-ping bottom track (BP), ; Set maximum bottom search depth to 1200 meters (BX) BP000 ;BX12000 ; output velocity, correlation, echo intensity, percent good ND111100000 ; Ping as fast as possible TP000000 ; Since VmDas uses manual pinging, TE is ignored by the ADCP ; and should not be set. ;TE0000000 ; Set to calculate speed-of-sound, no depth sensor, external synchro heading ; sensor, pitch or roll being used, no salinity sensor, use internal transducer ; temperature sensor EZ1011101 ; Output beam data (rotations are done in software) EX00000 ; Set transducer misalignment (hundredths of degrees). ; Ignored here but set in VmDAS options. ;EA00000 ; Set transducer depth (decimeters) ED00110 ; Set Salinity (ppt) ES35 ;set external triggering and output trigger; no trigger CX0,0 ; save this setup to non-volatile memory in the ADCP CK

WaMoS

The wave radar system on *Polarstern* uses radar reflectance from surface waves to determine the ocean surface current velocity and direction. For the first part of the cruise until August 10, 2017, the ship was in sea-ice covered regions where surface waves were dampened by the sea-ice. Only from August 10, 2017 onwards there had been sufficient surface roughness for WaMoS to be able to determine the current velocity. The data was screened according to significant wave height and otherwise saved during PS107.

Salinometer

Water samples were taken from depths >1.000 m at the stations marked "salt" in Table 4.1. These samples were then run on the Optimare Precision Salinometer SN006 to determine their salinity. The standard protocol developed by the Physical Oceanography of Polar Seas group at AWI was followed.

Preliminary (expected) results

In order to learn about the structure of a submesocale front in Fram Strait, we consulted at satellite radar imagery. An image from July 26, 2017 revealed a nearly straight line of sea-ice of almost 50 km length and only 500 m width. *Polarstern* moved to the front and then occupied 6 parallel cross-frontal sections (Fig. 4.1) with a total of 97 UCTD casts.

The UCTD measurements from the frontal survey showed that the along-front changes were much weaker than the cross-front changes. It also revealed denser Atlantic Water in the center of the front at 200 m depth than on either side of the front. A strong (exceeding 0.5 m/s) geostrophic along-front jet was also observed with the vessel mounted ADCP. Fig. 4.2 shows the temperature and density distribution in 6 parallel cross-front sections.

In order to detect possible frontal zones in the West Spitsbergen Current and in order to continue annual surveys of the WSC as they had been undertaken in previous years by AWI, we occupied a long transect from roughly 3.5°E to 11°E near 79°N (Fig. 4.3). The section, which is comprised of 136 UTCD casts, can be seen in Fig. 4.4 and agrees well with the climatological situation in August (e.g. von Appen et al., 2016), but it also reveals a strong front between 5°E and 6°E with raised isopycnals. This provides a different view to the traditional lower resolution sampling.

Fig. 4.2: Temperature in °C (color) and density in kg/m3 (contour lines) of the cross-frontal transects (transects A, B, C, E)

Fig. 4.3: Locations of the UCTD casts during the high-resolution transect across the West Spitsbergen Current

Fig. 4.4: Temperature, salinity, and density plot of the section across the West Spitsbergen Current from 3.5° E to 11° E

Data management

The raw data collected on PS107 is archived at Pangaea. When the data has been processed, the processed data will be made available on Pangaea as well.

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5. MOORING WORK OF THE FRAM PROJECT IN LTER HAUSGARTEN

1 AWI

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Objectives

Ongoing climate change will impact marine ecosystems in a variety of ways, and polar environments are believed to be particularly sensitive to these changes. Increasing atmospheric CO2 concentration will lead to a reduction of ocean pH, which is expected to exert a significant impact on marine calcifying organisms. Reductions in sea-ice cover and warming of water masses will modify ocean stratification and restrict the supply of nutrients to the euphotic zone. Taken together with anticipated changes in light conditions, net phytoplankton growth is expected to change with important ramifications for Arctic ecosystem structure and biogeochemical fluxes. In particular the quantity of photosynthetically produced organic matter exported from the surface ocean is likely to change under these conditions.

These changes have clear implications for the sequestration of atmospheric CO2 and the structure and function of benthic ecosystems that principally rely on this energy source from the pelagic. Considering the importance of these processes, reliably detecting the effects of climate change and understanding the influence of anthropogenic forcing on Arctic ecosystems is a clear priority. As part of our efforts to observe and detect long-term changes in Arctic Ocean ecosystems, we deployed a network of fixed- point moorings in the Fram Strait. On the one hand these moorings reflect a continuation of the well-established and long-term efforts of monitoring particle flux in the Fram Strait as part of the LTER Observatory HAUSGARTEN. In addition to this, new mooring arrays were designed to sample biogeochemical and physical properties in the upper water column to link with particle flux observations. These arrays include autonomous water samplers and autonomous particle samplers that are capable of collecting discrete samples with weekly resolution and preserving them for detailed analysis upon recovery. This work is part of the Frontiers in Arctic marine Monitoring (FRAM) infrastructure project.

To determine the transfer of organic matter from the upper productive layer in the water column to the bottom of the ocean, measurements of settling particles are performed by means of year-round moored sediment traps that provide information on the quantity and seasonality of vertical particle flux. The moorings are also equipped with current meters (RCMs) and selfrecording CTDs, to gain information on the hydrographic conditions in the study area. Results obtained by these instruments are of major importance for the interpretation of the results derived by the sediment traps, as the settling particles can be transported over long distances before arriving at the seabed.

To determine the seasonal changes in nutrient concentrations in the euphotic zone, water samplers were deployed in 2016 (PS99.2) and also on the current cruise (PS107) at approximately 20 m and 80 m depth. In total, 24 discrete samples will be taken with weekly to monthly resolution (depending on season) to follow the biological drawdown of nutrients. The moorings are also equipped with a physical and biogeochemical sensor package including SBE37-SMP-ODO (temperature, salinity, oxygen), SAMI pH, SAMI pCO₂, Wetlabs PAR (photosynthetically active radiation), Wetlabs Ecotriplet (Chlorophyll and CDOM fluorescence plus scattering), SUNA Deep Nitrate, current meters, and Acoustic Doppler Current Profilers. The combination of these sensors and the water samplers, in combination with the deployment of a profiling winch facilitates the assessment of seasonal stratification and nutrient concentrations above and below the pycnocline. The nutrient drawdown enables an estimate of new production. Furthermore, the samples will be used for DNA sequencing to examine seasonal changes in bacterial community structure. The particle samplers collect and preserve filters for DNA extraction and sequencing that together with the fluorescence sensors allow us to track the progression of phytoplankton biomass and community composition over different seasons. These efforts give us a novel year-round description of biological, chemical, and physical processes in the Fram Strait.

In addition, water samplers were deployed at ~80 m water depth in the anticipated cores of the West Spitsbergen and East Greenland Currents. Together with detailed measurements of physical parameters, these samples will be used to measure the nutrient composition of water masses flowing into and out of the Arctic Ocean through the Fram Strait.

Work at sea

6 moorings and one bottom lander were recovered on PS107 and redeployed in a similar configuration (Table 5.1) for planned recoveries in 2018. The mooring work was successful without major complications.

As part of the cruise several moorings were deployed in the Fram Strait, to combine physical and biological autonomous observations. The HG-EGC-4 and HG-N-FEVI-35 moorings were redeployed in the same configuration as in 2016. In addition we recovered and deployed four out of six moorings comprising two triangular cluster arrays that combine physical and biogeochemical measurements in the eastern part of the Fram Strait at the existing HG-IV and F4 stations. The HG-IV-FEVI-36 mooring was redeployed configured with three sediment traps and a McLane RAS 500 water sampler at 80 m and an upward looking ADCP at 400m. Additionally, larval samplers and recruitment surfaces were added at four depths on three moorings (Fig. 5.1). Two near-surface moorings were also recovered and deployed at HG-IV (HG-IV-S-2) and F4 (F4-S-2) that were equipped with McLane RAS 500 water samplers and McLane PPS particle samplers at 20 m (Fig. 5.2). Furthermore, biogeochemical sensor packages were attached to these frames that included the following: SBE37-SMP-ODO (temperature, salinity, oxygen), SAMI pH, SAMI pCO2, Wetlabs PAR (photosynthetically active radiation), and Wetlabs Ecotriplet (Chlorophyll and CDOM fluorescence and scattering), SUNA Deep Nitrate. In addition to the traditional and near surface moorings the third components of these triangular cluster arrays are surface profiling winches. One of these moorings, HG-IV-SWIPS-2017, is equipped with a CTD sensor package (Fig. 5.3).

Name		Longitude	Latitude		Deployment time				Recovery time					Depth	Deployment station	Recovery station	
	Degrees	Minutes	Degrees	Minutes	Year	Month	Day	Hour	Minute	Year	Month	Day	Hour	Minute	Meters		
Recoveries																	
HG-IV-FEVI-34	$\overline{4}$	19,97 E		79 0.00 N	2016			7 11 17 47		2017	7	27	6	9	2612	PS99/072-1	PS107/003-2
HG-IV-S-1	4	15,71 E		79 1,38 N	2016		7 11	7	37	2017	7	27	4	20 ₁	2542	PS99/070-1	PS107/003-1
HG-IV- SWIPS-2016	4	24.30 E		79 1,40 N	2016			7 12 16	2	2017	7	27	12	9	2480	PS99/076-1	PS107/005-1
$F4-S-1$	6	57.89 E		79 0.71 N	2016		7 10 7		31	2017	8	13	8	57	1223	PS99/067-1	PS107/044-1
HG-N-FEVI-33	4	29,42 $E $		79 44,19N	2016	75		$\overline{7}$	35	2017	8	8	5	57	2641	PS99/055-6	PS107/033-8
HG-EGC-3	2	47,63 W 78 49,86N			2016			6 30 15 10		2017	8	3	9	57 ¹	2535	PS99/048-7	PS107/023-1
Lander-2016	4	6,72 E		79 4,68 N	2016			7 12 17 50		2017	$\overline{7}$	27	9	56	2494	PS99/077-1	PS107/004-1
Deployments																	
HG-IV-FEVI-36	4	20,02 E		79 0.00 N	2017		8 11 6		4						2609	PS107/038-1	
HG-IV-S-2	4	15.71 E		79 1.36 N	2017		8 11 8		14						2599	PS107/038-2	
HG-IV- SWIPS-2017	4	24,31 E		79 1,39 N	2017			8 11 12 12							2535	PS107/038-4	
$F4-S-2$	6	57,86 E		79 0.70 N 2017				8 15 10 16							1260	PS107/049-1	
HG-N-FEVI-35	4	31,44 E		79 44,49N	2017	89		16	37						2692	PS107/035-1	
HG-EGC-4	2	47,53 W 78 49,86N			2017	84		13	3						2589	PS107/025-2	
Lander-2017	4			6,74 E 79 4,68 N 2017				8 11 14 23							2493	PS107/038-7	

Tab. 5.1: Overview of mooring recoveries and deployments on PS107

Fig. 5.1: Larval samplers and recruitment surfaces attached to 3 moorings in 4 depths

Fig. 5.2: Recovery of the 20 m biogeochemical sensor and sampler package on F4-S-1. The instruments spent one year in the euphotic zone.

Fig. 5.3: SWIPS system before deployment in HG-IV-SWIPS-2017. The yellow body is the winch and storage unit. The orange body is the profiler.

The biological and chemical sensors as well as the water and particle samplers that were recovered on PS107 had been programmed in German summer time and not UTC. All of the sensors and samplers deployed on PS107 were programmed in UTC with the exception of the water sampler on HG-EGC-4 which was programmed in German summer time.

The sampling schedules of the samplers and the sediment traps are archived together with the sensor raw data of the instruments recovered on PS107 on Pangaea.

During PS107 the two shallow moorings deployed in 2016 during the PS99 cruise, containing McLane Remote Access Sampler 500 (RAS), McLane PPS particle samplers, Microcat CTDs, SAMI pHs, SAMI pCO₂, Wetlabs PARs (photosynthetically active radiation) and Wetlabs EcoTriplets (CDOM, Clorophyll and fluorescence) were recovered.

The SAMI pH, recovered at F4-S-1 stopped measuring due to low battery after 10 months and 1 week. Since the necessary cable was missing on board, the data recovered at HG-IV-S-1 was not downloaded from the SAMI pH during the cruise.

The SAMI pCO $_{\rm 2}$ sensors, recovered at F4-S-1 and HG-IV-S-1 measured for 365 days in an interval of 1 hr.

The Wetlabs PAR, recovered at F4-S-1 did not collect any data while in water. There must have been a problem with it being turned on. When recovered, its batteries were drained. When replacing the batteries on board, it started its programmed duty cycle. The Wetlabs PAR, recovered at HG-IV-S-1 worked as programmed from deployment until recovery in an interval of 1 hr and 2 sec. This is different from a planned 1 hr interval and highlights that the instrument programming necessary for the Wetlabs sensors is highly counter-intuitive.

The Wetlabs Triplets, recovered at F4-S-1 and HG-IV-S-1 was programmed for 1,000 cycles with an interval of 1hr. The 1,000 cycles where accomplished after 41 days.

When recovered, the batteries were drained. After replacing the batteries, the sensors started running their programmed duty cycles.

All of the sensors encountered bio-fouling on their housings. However, and the copper antifouling protections and wipers worked well.

All recovered sensors were manually cleaned from bio-fouling and the SAMI sensors were flushed with de-ionized water.

As part of the cruise, the two shallow moorings F4-S-1 and HG-IV-S-1 were replaced by two new moorings F4-S-2 and HG-IV-S-2, which will be recovered in 2018. The new moorings have the same configuration as 2016 but in addition, 2 SUNA nitrate sensors are attached to the RAS samplers, equipped with external batteries. At F4-S-2 the nitrate sensor is equipped with an anti-fouling cage to protect the optical chamber. At HG-IV-S-2 the nitrate sensor is equipped with a so called "bio-wiper" which also prevents bio-fouling. Both sensors were calibrated on board and tested with a standard nitrate concentration of 8 µMol. The measuring interval of both sensor was set to 3 hr.

The SUNA pH sensors were tested in sea-water on board and the deployment interval was set to 3hr. The reason for the longer interval is the drained battery encountered during the previous deployment. Furthermore, the sensors were flushed with de-ionized water before deployment.

The SUNA pCO2 sensors were set to the same interval as 2016 of 1 hr and were flushed with de-ionized water.

The performance of the recovered and deployed RAS water samplers and the PPS particle samplers is documented in Table 5.2.

Tab. 5.2: Performance of the RAS water samplers and the PPS particle samplers

The SWIPS winch performed the first water column profiles from 140 m to 15 m depth in fall and winter. After initially performing two planned profiles 3 days apart, the system went on a hiatus from mid July 2016 to mid November 2016. Then it performed another 17 profiles in winter. Then the system fatally failed due to salt-water leakage at the slip-ring. The data was stored on the winch system, but the profiler dragged all of the cable out from the winch and then sheared the cable through and got lost at sea.

The cups of the sediment traps recovered are shown below and the trap performance is described.

FEVI-33-Oben

All cup rotated as planned. Since the last cup was planned to rotate into position after the recovery date, bottle #19 was open during recovery and bottle #20 had not recovered.

FEVI-33-Unten

The rotation system had stalled at bottle #3, which was still open during recovery. Therefore, only bottle #1 and #2 are reliable.

FEVI-34-Oben

All cup rotated as planned. Since the last cup was planned to rotate into position after the recovery date, bottle #19 was open during recovery and bottle #20 had not recovered.

FEVI-34-Mitte

All cup rotated as planned. Since the last cup was planned to rotate into position after the recovery date, bottle #19 was open during recovery and bottle #20 had not recovered.

FEVI-34-Unten

The rotation system had stalled at bottle #5, which was still open during recovery. Therefore, only bottle #1 through #4 are reliable.

EGC-3-Oben

All cup rotated as planned. Since the last cup was planned to rotate into position after the recovery date, bottle #19 was open during recovery and bottle #20 had not recovered.

EGC-3-Unten

All cup rotated as planned. Since the last cup was planned to rotate into position after the recovery date, bottle #19 was open during recovery and bottle #20 had not recovered.

F4S-1

All cups rotated as planned and the trap was recovered with the funnel position at the empty hole.

LANDER

All cups rotated as planned and the trap was recovered with the funnel position at the empty hole.

Preliminary (expected) results

The data present an unprecedented dataset characterizing the seasonal progression of physical, biological, and chemical parameters in the Arctic Ocean with the marked absence of year-round fluorescence data. This documents the onset of stratification in the water column and how this is related to salinity and temperature. It also allows to follow the development of the phytoplankton bloom and to look at the seasonal phenology of dominant eukaryotic and bacterio plankton groups. Nutrient inputs to the surface layer and the ensuing biological consumption will allow us to estimate the magnitude of new production. By comparing the fluxes measured in sediment traps, we can estimate the fraction of production exported out of the surface layer and the dominant mechanisms characterizing this export.

The preliminary physical/biogeochemical time series from the HG-IV and F4 clusters are shown in Figs 5.4-5.7. The temperature and salinity profiles taken by the SWIPS winch on HG-IV-SWIPS-2016 are shown in Figs 5.8 - 5.9.

Fig. 5.4: Pressure, temperature, salinity, water speed, and chlorophyll concentration values at the HG-IV cluster in 2016-2017. The red, blue, green, cyan pressure time series show where the respectively colored time series are measured. The black dots superimposed on the pressure time series show the time/pressure of particle samples. The gray dots above 180 m superimposed on the pressure time series show the time/pressure of water samples. The gray dots below 180 m superimposed on the pressure time series show the time/pressure when the sediment trap rotated between cups. The vertical magenta lines superimposed on the pressure time series show the time/pressure of CTD profiles taken by the SWIPS system. All sensor values are factory calibrated and non-final.

Fig. 5.5: PAR, CDOM, scattering, CO2 , and pH values at the HG-IV cluster in 2016-2017 at the depth of the red pressure time series in Fig. 5.4. All sensor values are factory calibrated and non-final.

Fig. 5.6: Same as Fig. 5.4, but for the F4 cluster, where only one mooring of the cluster was recovered in 2017

Fig. 5.7: Same as Fig. 5.5, but for the F4 cluster

Fig. 5.8: Temperature profiles between July 2016 and February 2017 at HG-IV-SWIPS-2016

Fig. 5.9: Same as Fig. 5.8, but for salinity

Data management

The raw sensor data and sample schedules collected on PS107 are archived at Pangaea. The sensor configuration of the recovered and deployed moorings is documented on sensor.awi. de. When the data has been processed, it will be made available on Pangaea as well. The unrestricted availability from Pangaea will depend on the required time and effort for acquisition of individual datasets and its status of scientific publication.

6. PELAGIC FOOD WEB: INTERACTIONS WITH THE BIOLOGICAL PUMP

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Grant No. AWI_PS170_06

Objectives and scientific program

Our main objective during the cruise was to study the controlling mechanisms for attenuation and export of organic carbon flux through the water column. This was done by detailed investigations of particle dynamics in relations to plankton community structure and aggregate composition. We did this by looking at both large and small scale, i.e. on a whole water perspective using *in-situ* optics and drifting traps on short time-scale or by using our BioOptical Platform for long-term patterns in aggregate size-distribution and abundance. The small scale studies were done on individual aggregates collected in from different depths through the water column.

Work at sea and first preliminary results

a) In-situ long-term monitoring of abundance, size-distribution, sinking velocity of settling aggregates

During the *Polarstern* cruise PS99-2 we re-deployed the prototype of the BioOptical Platform (BOP) on the FEVI-34 mooring at the HG-IV station. We developed the BOP system to be able to follow aggregate dynamics at high temporal resolution at different seasons throughout a whole year. BOP uses an *in-situ* camera system to determine daily size-distribution, abundance and size-specific sinking velocities of settling particles at one particular depth throughout one year (during the FEVI-34 deployment it was at 1,200 m). This is done by having a settling cylinder where particles sink through. At the bottom part of the settling cylinder we have attached a perpendicular camera system that records 5 minutes image sequences daily. At the bottom of the settling cylinder we attached a rotation table with 20 collection cups so each cup could be placed under the settling column for a pre-determined collection period (Fig. 6.1, Table 6.1). The cups are filled with a viscous gel which preserves the size and three dimensional structures of particles sinking into the gel. This makes it possible to identify and quantify different particle types as well as their compositions.

The BOP system was based on a modified sediment trap (KUM GmbH) where the collection funnel was replaced with a polycarbonate cylinder to avoid that the settling particles were sliding/rolling down the sides of the funnel, which may alter their physical structure. The polycarbonate cylinder has an inner diameter of 35 mm and functions as a settling cylinder that excludes ocean currents while the particles settling through it (Fig. 6.1). The camera system placed at the lower part of the settling cylinder consisted of an industrial camera (Basler), a fixed focal length lens (Edmund Optics) and a single board computer including a SSD hard disc and custom made power and time management circuitry. The images were illuminated by a custom made visible light source providing backlight. The whole camera system was powered by a Li-Ion battery (24V, 1670Wh, SubCTech GmbH) (Fig. 6.2).

Fig. 6.1: The old BOP system deployed during PS99-2 and recovered during PS107 with the polycarbonate settling column and camera illumination switched on (left image) and the rotation table with collection cups (right image). The large cups are for long collection periods and the small black cup are filled with viscous gel for short period collection.

Recovery of the BOP at FEVI-34, HG-IV (deployed during PS 99-2)

During this cruise (PS107), we recovered the BOP system at the FEVI-34 mooring at station HG-IV (PS107 3-2) on the 27 July 2017. The cups had all rotated through to the last cup and all contained material. From this we could confirm that the BOP system was able to capture settling particles at all seasons.

The camera system captured 5 min of images every second day from the 11 July 2016 and until 18 March 2017, which resulted in 122 days of image sequences. This premature end of image sequence collections was caused by earlier drainage of the Li-battery due to failure on the power module manager (PMM). We have replaced the PMM on all BOP systems and should not have these issues any longer.

Tab. 6.1: Programming of the of the BOP system deployed during PS99-2 and recovered during PS107. Periodical measurements by the camera system for 5 minutes every second day and opening times for each of the 20 gel cups.

Deployment of the BOP system at FEVI-36 (deployed during PS107)

We deployed a new version of the BOP system, which was equipped with two rotating tables and capable of collecting sinking particles in 40 gel-cups (Fig. 6.2). The camera system was similar to that described above. We deployed the BOP system on the FEVI-36 mooring at station PS107_38-2 on the 11 August 2017 at 79°00.01'N and 04°20.02'E. Please see further information about the mooring under the mooring cruise report. We timed the cup openings according to the deep ocean sediment traps on the same mooring, but ensured that we would have several gel cups with only three days of opening period at each collection period of the deep ocean sediment traps (Table 6.2). This was to ensure that we would not have particles falling on top of each other, which would prevent of from doing image analyses on the particles collected in the gel traps. We programmed the camera for measurements of particle type, sizedistribution, abundance, and sinking velocities to switch on daily at 12:00 (UTC) and capture one image every four seconds for 31 minutes.

Fig 6.2: The new BOP system with the polycarbonate settling column (left image) and the two rotation tables (right image)

b) Marine snow catcher

We deployed 26 marine snow catchers (MSC) to collected *in-situ* formed marine snow.

Fig. 6.3: The marine snow catcher was deployed via the ship's winch system and lowered to a specific depth, whereafter it was closed by a messenger weight and releaser. After recovery the MSC was positioned upright on deck to allow the particles to settle to the bottom part of the MSC.

The MSCs consisted of a 100 l cylindrical water sampler with a particle collection tray at the bottom (Fig. 6.3, Table 6.3). The MSCs were deployed with a winch to the target depth and closed via a release mechanism that was activated with a drop weight. The closed MSCs were placed on deck for a few hours to allow the collected particles to sink to the collection tray at the lower part of the MSCs. We gently drained the water from the 100 l cylinder and removed the collection tray, which now contained the settling particles.

We used the collected aggregates to determine their size-specific sinking velocities, microscopical observations of the aggregate composition, and made filtrations for DNA. The sinking velocities were determined using a video setup in a temperature controlled cooling room. The aggregates were individually placed in an aquarium with a length and depth of 10 cm and a height of 25 cm. The aquarium was closed with a lid to avoid evaporation and convection in the aquarium. The lid had a small hole that allowed us to place individual aggregates at the top part of the aquarium. We made stack images of individual aggregates from each station using a microscope. These images provide a detailed semi three dimensional composition of the aggregates (Fig. 6.4).

Tab. 6.3: Deployments of the marine snow catcher with information about station name, MSC number, date of deployment, time for deployment, latitude, longitude, and deployment depth.

SEAPUMP

FRELMHOLTZ

Fig. 6.4: Example of a stacked image of an aggregate collected at station SV-III near Svalbard. The aggregate contained several chains of diatoms but mainly consisted of small fragments of diatoms.

c) Drifting sediment traps

We used an array of free-drifting sediment traps to measure the export fluxes at 100 m, 200 m, and 400 m depth (Table 6.4, Fig. 6.5). Each collection depth had a trap station that consisted of four cylindrical collection tubes with a gyroscopical attachment (Fig. 6.5). Three of the four collection cylinders at each depth were used to collect samples for biogeochemical measurements of total dry weight, particulate organic carbon, particulate organic nitrogen, particulate inorganic carbon, and silica. The fourth trap cylinder at each depth was equipped with a viscous gel that preserved the structure, shape and size of the fragile settling particles (Fig. 6.6). After recovery of the drifting trap, the samples for bulk fluxes were frozen for later analysis in the home laboratory. The particles collected in the gel traps were photographed with a digital camera on board and frozen for further detailed investigations in the home laboratory. The image analyses of the gel traps will be used to determine the composition, abundance and size distribution of the sinking particles.

We deployed the drifting trap six times during the PS107 cruise. The drifting array consisted of a surface buoy equipped with an Iridium satellite unit the provided trap positions every 10 minutes with a resolution of two minutes, we used two benthos floats for buoyancy and 14 small buoyancy balls were placed between the surface buoy and the two benthos floats to act as wave breakers and thereby reducing the hydrodynamic effects on the sediments traps. The trap cylinders were mounted to a sediment station with gimbal mounts ensuring that they were maintaining a vertical position in the water column. Each cylinder was 1 m tall and had a diameter 10.4 cm, which resulted in a collection area of 84.95 cm².

Fig. 6.5: Images of the free-drifting sediment trap close to the ice edge at East-Greenland (left). One of the trap stations with the four sediment trap cylinders (right).

Four of the six trap deployments were equipped with gel traps. These were at HAUSGARTEN central stations (HG-IV), the East-Greenland station (EG-III), the HAUSGARTEN North station (N4), and the Svalbard station (SV-III). We deployed two further drifting trap during a survey of front system between the HAUSGARTEN central region and the East-Greenland stations. However, since we needed to deploy two drifting trap directly after each other, we only had two collection cylinders at each collection depth and decided use both collection cylinder for biogeochemical analyses in order to have enough material.

Fig. 6.6: Examples of the particles collected in the gel traps at 100 m, 200 m, and 400 m at the four stations: East-Greenland (EG-III), Central Hausgarten (HG-IV), Svalbard station (SV-III), and the northern station (N4).

The gel trap collections showed that the composition of sinking particles were different at the four stations (Fig. 6.6). At the East-Greenland station (EG-III) we mainly collected copepod fecal pellets at 100 m, while the most frequent particles were dense amphipod fecal pellets at the two deeper depths. At the central HAUSGARTEN station, we mainly observed small dense aggregates composed of degraded fecal pellets and few copepod and krill fecal pellets at the deeper depths (200 m and 400 m). The differences between the HAUSGARTEN central station (HG-IV) majority of the settling particles consisted of marine snow and fecal pellets. Both the northern station (N4) and the station close to Svalbard (SV-III) had many copepod

and krill fecal pellets sinking through 100 m and 200 m depths, while the 400 m traps mainly showed small aggregates composed of degraded fecal pellets (Fig. 6.6).

d) Vertical profiles with the In Situ Camera system

The In Situ Camera (ISC) consisted of an industrial camera with removed infrared filter (from Basler) with backend electronics for timing, image acquisition and storage of data and a fixed focal length lens (16 mm Edmund Optics). Furthermore a DSPL battery (24V, 38Ah) was used to power the system (Fig. 6.7).

Fig. 6.7: Deployment of the In Situ Camera (ISC), consisting of an industrial camera and lens with electronics, an infrared light source, the DSPL battery and a Seabird SBE19 CTD.

A single board computer was both used as the operating system for the infrared camera and to acquire the images from the camera and send them to a SSD hard drive where they were stored. The illumination was provided by a custom made light source that consisted of infrared LEDs which were placed in an array in front of the camera. The choice of the infrared illumination was done to avoid disturbing the zooplankton that potentially would feed on the settling particles. With this geometrical arrangement of the camera and the light source we obtained shadow images of particles through the water column. We captured 2 images per second and lowered the ISC with 0.3 meters per second (lowest possible speed of winch).

Sampling: we made 24 vertical profiles with the IR-Camera. Profile 01 (just to10 m) was used for calibration. We had some hardware issues during the sixth profile, which led to data loss during this profile. However, we could fix the problem quite fast and so we could repeat this profile. On the camera frame also a Seabird SBE19 CTD was mounted. With a, better correlation of depth and images can be achieved. During the whole cruise both instruments were used in standalone mode and data was then afterwards correlated with Matlab.

With that we got 22 successful profiles (9 x 500 m, 1 x 730 m, 1 x 900 m, 11 x 1,000 m,). An overview of all measured profiles can be found in Table 6.5.

Tab. 6.5: List of stations where the *in-situ* Camera (ISC) was deployed in the profiling mode (Wdepth is water depth and Pdepth is profile depth).

Data management

When the data has been finally processed, it will be made available on Pangaea. The unrestricted availability from Pangaea will depend on the required time and effort for acquisition of individual datasets and its status of scientific publication.

7. PELAGIC FREE-LIVING AND PARTICLE-ASSOCIATED BACTERIAL AND ARCHAEAL COMMUNITIES IN FRAM STRAIT

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1 AWI 2 MPI-MM 3 MARUM

Grant No. AWI_PS107_07

Outline and objectives

Long-term studies at the LTER HAUSGARTEN observatory in the past two decades have revealed changes in pelagic microbial communities, *e.g.* shifts in the phytoplankton community from diatom-dominated towards *Phaeocystis*-dominated communities (Soltwedel et al., 2015). But no baseline has been established yet for bacterial and archaeal communities, in order to assess temporal changes in their structure and function as a result of changing environmental conditions. In the framework of the FRAM project, a Molecular Microbial Observatory is currently established. Within this framework, the collected samples will be used to study the composition and function of bacterial and archaeal communities of different water masses in the Fram Strait (Soltwedel et al., 2013). The work is coordinated with and linked to the sampling of eukaryotic microbial communities (see Chapter 8). In addition, samples for bacterial and archaeal community structure were collected as part of an interdisciplinary effort to understand the extent of meso-scale water column dynamics (see Chapter 4).

Sinking particulate organic matter (POM) plays a major role in linking pelagic and benthic ecosystems (De La Rocha and Passow, 2007). Sinking particles get quickly colonized by microbial communities and degraded on their way through the water column, eventually reaching the seafloor. Characterizing colonization patterns and the composition and function of bacteria and archaea on sinking particles will thus help to better understand organic matter export in different regions of Fram Strait. Our main objective during the expedition was to better understand the effect of "seed" free-living microbial communities from different water masses, on the colonization and degradation patterns of POM. An experimental set-up was complemented by sample collection of natural particles using Marine Snow Catcher (see Chapter 6).

Work at sea

Water samples for bacterial and archaeal community structure analyses will be obtained using 12L Niskin bottles housed on a rosette equipped with SBE conductivity–temperature–depth (CTD) sensors. Triplicates from each sample will be filtered using peristaltic pumps through a 0.22 µm Sterivex© filter (see Table 7.1). During the sampling of meso-scale water structures, the sampling was conducted in fractionated filtration through 5 or 3 um nylon filters for particlesassociated fraction and then 0.22 µm for the free-living fraction (see Table 7.2). Same filtration set-up was used for the Marine Snow Catcher sampling (see Table 7.3).

The particle incubation experiment was conducted for periods of 11 days each time, using freshly sampled water from distinct North Atlantic waters (AW) that were collected at station S3 from 60 m depth, and the surface Polar waters (PSW) that were collected at station EG1 from 60 m depth. In parallel to the water collection using niskin bottles, the same water masses were sampled using Marine Snow Catcher (MSC) for natural occurring particle-associated communities on marine aggregates.

Tab. 7.1: Water column samples for DNA extractions and total prokaryotic cell counts, collected at LTER HAUSGARTEN stations. DNA samples were filtered on 0.22 µm Sterivex© filters and stored at -20°C. For fluorescence *in-situ* Hybridization (FISH), samples were fixed using 2 % formaldehyde, filtered on 0.22 µm membranes and stored at -20°C.

Tab. 7.2: Water column samples for DNA extractions and total prokaryotic cell counts, collected during the meso-scale water structure survey. DNA samples were sequentially filtered on 5 or 3 and 0.22 µm filters and stored at -20°C. For fluorescence *in-situ* Hybridization (FISH), samples were fixed using 2% formaldehyde, filtered on 0.22 µm membranes and stored at -20°C. Marine Snow Catcher samples were collected in the noted stations.

Tab. 7.3: Marine Snow Catcher samples for DNA extractions, collected at LTER HAUSGARTEN stations. The samples were sequentially filtered on 5 or 3 and 0.22 µm filters and stored at -20° C.

Data management

Post-cruise data archival will be hosted by the information system PANGAEA at the World Data Centre for Marine Environmental Sciences (WDC-MARE), which is operated on a longterm base by the Alfred Wegener Institute Helmholtz-Centre for Polar and Marine Research (AWI), Bremerhaven and the Center of Marine Environmental Sciences (MARUM), Bremen. Scientific data retrieved from observations, measurements and home-based data analyses will be submitted to PANGAEA either upon publication or with password protection as soon as the data is available and quality-assessed. This includes also biological data, for most of which parameters are already defined in PANGAEA. Molecular data will be deposited in public repositories such as NCBI and ENA. Biological samples will be stored deep frozen or fixed at the Max Planck Institute for Marine Microbiology (MPIMM) in Bremen. One of the three replicates from each sample will be used for the long term biological archive of the Molecular Observatory at the Alfred Wegener Institute Helmholtz-Centre for Polar and Marine Research (AWI), Bremerhaven.

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8. FUNCTIONAL GLYCOMICS OF ALGAL BLOOMS IN THE ARCTIC OCEAN

Grant No. AWI_PS107_08

Objectives

Marine algae are contributing to half of the global photosynthetic carbon fixation (Falkowski et al., 1998; Field et al. 1998), producing substantial quantities of reduced carbon compounds such as polysaccharides, lipids and proteins. Many algal species do not grow continuously, but in temporary blooms (Teeling et al., 2016) limited by nutrients, grazing predators and viral infections (Valiela et al., 1997). During this algal blooms, organic matter (OM) is released, which can subsequently trigger blooms of planktonic bacteria with a successions of distinct bacterial clades over time (Teeling et al., 2016, 2012). The remineralization of algal biomass by heterotrophic bacteria is an important step in the marine carbon cycle, but so far little is known about the key degraders and their enzymatic pathways.

Algal polysaccharides

Polysaccharides are composed of long chains of monosaccharides linked by glycosidic bonds. Marine algae contain large amounts of polysaccharides, ranging from $4 - 76\%$ of the dry weight depending on seasonal variations and algal species (Kraan, 2012). Polysaccharides physically support the thallus, have storage properties (Kraan, 2012) or are secreted as exudates. Algal polysaccharides serve as energy source for heterotrophic bacteria in form of dissolved organic matter (DOM) or they spontaneously aggregate into particles, which sink into deeper water layers (biological pump) and can serve as habitat and food source for bacteria (Passow, 2002; Verdugo et al. ,2004). One very abundant polysaccharide is laminarin, a linear β-1,3 glycan of brown algae consisting of ~ 20 to 30 glucose residues with β-1,6-linked side chains (Kraan, 2012).

Polysaccharide-degrading bacteria

Bacteria utilizing polysaccharides posses proteins specialized for carbohydrate degradation, such as sugar transporter, transcriptional regulators, and carbohydrate-active enzymes (CAZymes) including polysaccharide lyases (PL), glycoside hydrolyases (GH), carbohydrate esterases, and glycosyl transferases (Lombard et al. 2014). In the phylum Bacteroidetes, CAZymes are frequently organized in large operon like structures (Thomas et al., 2011), termed polysaccharide utilization loci (PUL) (Sonnenburg et al., 2010).

During *Polarstern* cruise PS107 (ARK-XXX1/2), we collected particulate organic matter (POM) from filtered seawater to obtain vertical profiles of laminarin concentration during an algal bloom in the Arctic Ocean by using carbohydrate active enzymes (CAZYmes) from marine heterotrophic bacteria (Becker et al., 2017). This novel method allows an accurate quantification of this important polysaccharide and therefore, serves further insights into the marine carbon cycle.

Work at sea

During the *Polarstern* cruise PS107 (ARK-XXX1/2), we took water samples by using Large Volume Water Transfer Systems(WTS-LV), also known as *in-situ* pumps, in order to obtain vertical profiles of laminarin concentration. The *in-situ* pumps were successfully deployed at seven stations (Table 8.1) in the Artic HAUSGARTEN area. The pumps were attached to the CTD rope (Fig. 8.1) and filtered approximately 300 L in 90 minutes onto 142 mm GF/F glass microfiber filters with a pore size of 0.7 µm in five different depths (surface, chlorophyll maximum, 300 m, 1,000 m, sea floor). The filters were frozen at -20°C, since we were not able to analyse them on board. Additionally, water samples were collected with a CTD/rosette water sampler at the corresponding depths of the *in-situ* pumps and stored at -20°C. To identify key heterotrophic bacteria and to study their enzymatic pathways for algal polysaccharide utilization, surface water obtained with a CTD/rosette was sampled for later DNA extraction and for fluorescence *in-situ* hybridization (FISH) in the laboratory in Bremen. FISH samples were fixed with formalin for 1 h at room temperature (RT), filtered onto 0.22 µm polycarbonate (PC) filters (Millipore) and stored at -20°C. Samples for DNA extraction were filtered onto 10 µm PC filters, 3 µm and 0.22 µm, flash frozen in liquid nitrogen and stored at -80°C.

Tab. 8.1: Deployment of *in-situ* pumps in the HAUSGARTEN area during cruise PS107

Preliminary (expected) results

For the quantification of laminarin in particulate organic carbon (POC), we will extract the polysaccharide from the filters and determine the concentration using CAZymes from marine heterotrophic bacteria (Becker et al., 2017). The data will be compared to the samples taken in 2016 during *Polarstern* cruise PS99.2. We expect high polysaccharide concentrations in the euphotic zone and a decrease with depth (Fig. 8.2).

Data management

The data will be available for cruise participants upon request and uploaded on the PANGAEA database after data processing.

Fig. 8.1: Deployment of the in-situ pumps at the CTD rope (A) and storage of filters after filtration (B)

Fig. 8.2: Filtered POC from surface (A), chlorophyll maximum (B), 300 m (C) and seafloor at ~5,350 m (D) at station HG-IX

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9. PLANKTON ECOLOGY AND BIOGEOCHEMISTRY IN A CHANGING ARCTIC OCEAN (PEBCAO)

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1 AWI 2 GEOMAR

Grant No. AWI_PS107_09

Background and Objectives

The project PEBCAO (Plankton Ecology and Biogeochemistry in a Changing Arctic Ocean) is focused on the plankton community and the microbial processes relevant for biogeochemical cycles of the Arctic Ocean. This research focus is acknowledging that the Arctic Ocean has gained increasing attention over the past years because of the drastic decrease in sea ice and increase in temperature, which is about twice as fast as the global mean rate. In addition, the chemical equilibrium and the elemental cycling in the surface ocean will alter due to oceanacidification. These environmental changes will have consequences for the biogeochemistry and ecology of the Arctic pelagic system. The effects of changes in the environmental conditions on the polar plankton community can only be detected through long-term observation of the species and processes. Our studies on plankton ecology have started in 1991 and sampling has been intensified by the PEBCAO-team since 2009 in the Fram Strait at ~79°N. Since then our studies are based on combining a broad set of analysed parameters. This includes e.g. classical bulk measurements and microscopy, optical measurements and satellite observations, molecular genetic approaches, or cutting edge methods for zooplankton observations to study plankton ecology in a holistic approach.

Over the past eight years we have compiled complementary information on annual variability in plankton composition, primary production, bacterial activity or zooplankton composition (Nöthig et al., 2015). Previous assessments in the observation area indicated that protist composition in the WSC changed in the summer months. A dominance of diatoms was replaced by a dominance of *Phaeocystis pouchetii* and other small pico- and nanoplankton species. Our recent regular annual observations in Fram Strait suggest that TEP concentration in the observation area could be correlated with *Phaeocystis pouchetii* abundance (Engel et al., 2017). These data were complemented by our molecular genetic investigations that provided new insights into eukaryotic microbial community composition with special emphasis on the contribution of pico- eukaryotes to plankton communities (Metfies et al., 2016).

Biogeochemistry and phytoplankton

Climate induced changes will impact the biodiversity in pelagic ecosystems. At the base of the food web, we expect small algae to gain more importance in mediating element and matter turnover as well as matter and energy fluxes in future Arctic pelagic systems. In order to examine changes, including the smallest fractions, molecular methods are applied to complement traditional microscopy. The characterization of the communities with molecular methods is independent of cell-size and distinct morphological features. The assessment of the biodiversity and biogeography of Arctic phytoplankton will be based on the analysis of ribosomal genes with next generation sequencing technology, Automated Ribosomal Intragenic Sequence Analysis (ARISA), and quantitative PCR. Besides molecular methods the set of parameters investigated includes classical bulk measurements (e.g. chlorophyll a, POC/N, biogenic silica) and microscopy.

Automated sampling for molecular analyses

Marine phytoplankton distribution displays high spatial heterogeneity or "patchiness". As a consequence, comprehensive observation of marine phytoplankton requires sampling with high spatial and temporal resolution. The latter is a labour intensive task and requires high amounts of ship time. The newly developed **auto**mated **fi**ltration system for **m**arine microbes (AUTOFIM) has high potential to reduce the described effort related to adequate sampling of marine phytoplankton. It is coupled to the ships pump system and allows filtration of a sampling volume up to 5 litres. In total 12 filters can be taken and stored in a roundel. Prior to the storage a preservative can be applied to the filters to prevent degradation of the sample material, that can be used for molecular or biochemical analyses. Filtration can be triggered after defined regular time intervals or remote controlled from a scientist. Alternatively, filtration could be event-triggered if the filtration system would be operated in connection with the FerryBox System, an *in-situ* measurement device for the monitoring of oceanographic parameter (temperature, salinity, pH etc.) installed on-board *Polarstern*. AUTOFIM (Fig. 9.1) provides the technical background for automated high resolution collection of marine samples for molecular analyses (Metfies et al., 2016). During expedition PS92 of RV *Polarstern* to the Arctic Ocean in summer 2015, AUTOFIM was used for the first time to collect samples from the upper water column at a depth of \sim 10 m, which is the depth of the inlet of the ships water pump system.

Fig. 9.1 A: AUTOFIM installed on board RV Polarstern (1: Sample reservoir; 2: Filtration; 3: Archive for preserved filters. B: Filtration-module (1: Filter stacker; 2:Filtration cap).

Optical methods for continuous data on phytoplankton community structure and colored dissolved organic matter (CDOM)

The distribution and composition of phytoplankton and CDOM in the Fram Strait are influenced by different environmental factors such as the water masses exchanges between the Arctic and Atlantic basins, the intensity of phytoplankton blooms and the riverine outflow. At this cruise we focused on collecting a high spatial and temporal resolved data set on phytoplankton biomass and composition, particulates and CDOM at the surface and for the full euphotic zone in the Fram Strait by taking continuous optical measurements which directly give information on inherent and apparent optical properties (IOPs, and AOPs, respectively). These will be validated by direct biological and chemical analysis of frequently taken water samples in order to derive information on abundance and composition of phytoplankton, other particulates and CDOM,. The whole large data set on phytoplankton and CDOM and the measurements of optical properties by radiometers at stations will also be used to validate the new Sentinel-3 sensor OLCI level-2 ocean products for high latitudes.

Bacterioplankton

The bioreactivity of particulate and dissolved organic matter is determined by its biochemical composition and diagenetic state. The loss of organic matter within and below the euphotic zone is mainly mediated by the degradation activity of heterotrophic bacteria, colonizing sinking particles and their surroundings. Hence, bacterial activity co-determines the efficiency of carbon export to the deep ocean. Furthermore, bacterioplankton plays a fundamental role at the basis of microbial foodwebs. Dissolved organic matter is almost exclusively accessible for bacterial cells that make it available for higher trophic levels by the production of bacterial biomass. Effects of increasing temperature and decreasing pH on bacterial communities and their activity are thereby of outstanding importance, but yet hardly considered. Studies conducted in the past decades revealed strong physiological responses of marine bacteria to changing temperature and pH, but their relevance for biogeochemical cycles in the future ocean is only poorly investigated.

Zooplankton

Many zooplankton species are associated with a specific water mass. For example, the boreal copepod *Calanus finmarchicus* is transported northward with the North Atlantic current whereas the sibling species *C. glacialis* inhabits Arctic water masses. Rising water temperatures and altered hydrographical conditions could therefore result in a shift in the zooplankton species composition in the Fram Strait and the Arctic ocean. Zooplankton might also be affected by a decrease in seawater pH due to uptake of anthropogenic carbon dioxide (ocean acidification). This could have severe consequences for the ecosystem functioning. To detect the impact of climate change on pelagic ecosystems of the Arctic, we studied the zooplankton community composition and depth distribution in the HAUSGARTEN area during PS99.2 and compare these with previous studies from the same area.

Work at sea

Samples for a large variety of parameters have been collected in the area of the 'deep-sea long-term observatory HAUSGARTEN' of the Alfred-Wegener-Institute located in the Fram Strait including the frontal zone separating the warm and cold water masses originating from the West Spitsbergen current and the East Greenland current. Sampling was accomplished by the PEBCAO team from CTD casts, sampling with the automated filtration device AUTOFIM and by net hauls is summarized in tables 9.1-9.3 .

Biogeochemistry (AG Engel)

We sampled seawater of 5-6 depths by a CTD/rosette sampler at the HAUSGARTEN area to determine the impact of microbial processes on the cycling of organic matter. Samples have been taken for dissolved biogeochemical parameters. Samples for dissolved organic carbon/ total dissolved nitrogen (DOC/TDN) and Total Alkalinity (TA) were filtered over 0.7 µm GMF syringe filters and stored at 4°C. Amino Acids and carbohydrates were samples for the dissolved fractions (DAA, DCHO, filtered over 0.45 µm Acrodisc filters) and particulate fraction (PAA, PCHO, filtered onto pre-combusted GFF filters) and stored frozen at -20°C. Concentrations will be determined by the use of IC and HPLC at GEOMAR in Kiel, respectively. At selected stations (HG-I, EG-III, N-V) a size fractionation resulted in PAA and PCHO samples for two phytoplankton size fractions (>10 μm and 10 - 0.47 μm) and bacterioplankton (<0.47 – 0.2 μm).

Additionally, samples for transparent exopolymer particles (TEP) and Coomassie stainable particles (CSP) were taken and stored at -20°C until analysis by photometry and microscopy back at GEOMAR. Bacteria and phytoplankton abundance will be determined using flowcytometry. Samples were taken from CTD water and stored at -20°C .

Tab. 9.1: Sample List (Biogeochemical parameters sampled from CTD casts)

DOC: dissolved organic carbon; TDN: total dissolved nitrogen; TEP: transparent exopolymer particles; CSP: *Coomassie* stainable particles; TA: total alkalinity; DCHO: dissolved carbohydrates; DAA: dissolved amino acids; Bac: bacterial cell numbers; Phyto: phytoplankton cell numbers; PCHO: particulate carbohydrates; PAA: particulate amino acids; * stations where size fractionation for PCHO/PAA occurred. Chl.*a*: chlorophyll *a*; POC/N/bSi: particulate organic carbon, nitrogen & biogenic particulate silica; TPP: total phosphorus. In addition, Seston (TPM, total particulate matter) samples were taken at stations where sediment trap moorings were deployed (HG-IV, N4, EG-IV). Here particulate parameters were taken down to close above bottom. Samples for microscopy have been taken at all Chl.*a* stations for the upper 50 – 100m.

Phytoplankton (AG Nöthig and AG Metfies)

Seawater samples were taken at 6-12 depths by a CTD/rosette sampler in the HAUSGARTEN area to determine the impact of microbial processes on organic matter cycling. The water from the rosette was filtered for analyzing biogeochemical parameters such as chlorophyll *a (unfractionated, and fractionated on 10 µm, 3 µm and 0.2 µm)*, seston, particulate organic carbon and nitrogen (POC/N), and particulate biogenic silica (PbSi) as well as total particulate phosphorus (TPP). In addition, samples were collected for microscopy to determine phytoand protozooplankton abundance. Furthermore, samples were collected from the top 100 m depth for molecular analyses in order to assess differences in the phytoplankton community composition and cellular activity by 18S meta-barcoding and meta-transcriptome analyses. Samples for 18S meta-barcoding analyses were fractionated by three filtrations on 10.0µm, 3.0µm and 0.2 µm filters. One additional archive sample was collected from every depth by filtration on 0.2 µm to provide a sample for future analyses methods. All samples were preserved, refrigerated or frozen at -20°C or -80°C for storage until analyses in the home laboratories (AWI, Bremerhaven).

Automated sampling for molecular analyses

During this cruise we used AUTOFIM on *Polarstern* in order to assess its applicability on board ships and to carry out filtrations with high spatial resolution in parallel to underway surveys of the physical oceanography. On one hand we collected water samples by AUTOFIM and a CTD/rosette sampler (5 depths) at 20 stations in Atlantic Water, polar water and on the Svalbard shelf. Samples were collected with AUTOFIM in parallel to the CTD/rosette sampler in order to evaluate to what extent the protist community composition in the AUTOFIM sample is representative for the protist community in the photic zone at the same sampling site. In addition we used AUTOFIM to collect samples regular intervals of 15-20 min in parallel to 10 transect studies of the underway CTD. All samples were filtered and preserved or frozen at 20°C for further molecular genetic analyses in the home laboratory.

Tab. 9.2: Sampling for Molecular Analyses (AUTOFIM: samples automatically collected for molecular analyses on a 0.4 µm filter; DNA Euk: DNA of eukaryotes; RNA Euk: samples collected a 3°C on an 0.2 µm filter; Archiv Filter: samples collected on an 0.2 µm Sterivex-filter.)

Optical methods for continuous data on phytoplankton community structure and colored dissolved organic matter (CDOM) (AG Bracher)

We continuously ran an *in-situ* hyperspectral transmission and absorption meter (ACs, WETLabs) during the cruise for underway surface water sampling. The instrument was mounted to the surface seawater supply with a membrane and operated in flow-through mode with a time-programmed filter to allow alternating measurements of the total and CDOM+water beam attenuation and absorption (IOPs) of seawater (Fig. 9.2). Flow-control and a debubbler system ensured water flow through the instrument with no air bubbles.

To validate the AC-s measurements, we took regularly (every 3 hours) surface water samples from the AC-S outflow for pigment analysis (further processed with High Performance Liquid Chromatography (HPLC) back at AWI) and the absorption of particulate matters, phytoplankton and CDOM. The absorption of the bulk particulate matters and phytoplankton were directly measured on board using Quantitative Filter Technique Integrating Cavity Absorption Meter (QFT-ICAM) (Fig. 9.3) and CDOM absorption with Liquid Waveguide Capillary Cell system (LWCC, WPI) (Fig. 9.4).

In addition to the underway programme, a second AC-s instrument was mounted on a steel frame together with a pressure sensor and a set of hyperspectral radiometer (RAMSES sensor, TRIOS) and operated successfully during 17 CTD stations. The frame was veered down to 140 m at a speed of 0.1 m/s with stops every 5 m and heaved with a continuous speed of 0.1 m/s to allow a better collection of radiometric data. For the validation of both devices we further took samples at 5 water depths (surface, DCM, Below DCM, 75 m and 100 m) for HPLC pigment analysis, absorption of particulate matters and phytoplankton and CDOM absorption and fluorescence at 26 CTD stations in the Fram Strait. Besides, we mounted a third RAMSES sensor on the monkey deck during the whole cruise for continuous spectrally resolved measurements of down welling irradiance in air (bottom of atmosphere irradiance). The RAMSES underwater radiance and irradiance data corrected by the in-air RAMSES incoming irradiance data will be used to calculate Remote Sensing Reflectance (R_{RS}). The *in-situ* R_{RS} data obtained under clear sky will be used to validate the ESA-Sentinel-3 OLCI imaging spectrometer R_{RS} data which are used to derive information on surface biogeochemistry (phytoplankton, other particulates and CDOM). All RAMSES R_{pS} data will be used to optimize models linking those to the inherent optical properties (measured by the AC-s) and to the geophysical quantities derived from those. In addition also the chlorophyll-a (chl-a) data obtained from the HPLC measurements will be used to validate the ESA-Sentinel-3 OLCI imaging spectrometer chl-a.

Fig. 9.2: Examples of absorption (red) and attenuation (blue) spectra acquired with the AC-S instrument

Fig. 9.3: Examples of absorption spectra of particulate matters (with peaks) and non-algal particles (without peaks) measured with the QFT-ICAM technique. The colors indicate different sample measurement.

Tab. 9.3: Bio-optical parameters sampled at PS107 stations. HPLC: High Pressure Liquid Chromatography; CDOM: Coloured Dissolved Organic Matter; PAB: Particulate absorption; RAMSES: upwelling and downwelling radiation in the water.

Zooplankton (AG Niehoff)

Mesozooplankton community composition and depth distribution were investigated at 11 HAUSGARTEN stations, 2 transit stations at the 0° meridian as well as 5 stations during a frontal zone study (see chapter 4; Fig. 4.1), using three different devices. For analyzing the largescale zooplankton distribution in the upper 1,500 m of the water column, we used a MultiNet which was equipped with 5 nets of 150 um mesh size. The MultiNet was towed vertically at 11 stations (Fig. 9.5) to sample five different depths layers (1,500-1,000-500-200-50-0 m). The samples were then immediately preserved in formalin buffered with hexamethylentetramine. To analyse the small-scale distribution of zooplankton species in the upper 1,000 m of the water column, a combination of the optical plankton recorder LOKI (Lightframe On-sight Key species Investigation) and the acoustic device AQUAscat was deployed at 18 stations (Fig. 9.5). LOKI was taking pictures of zooplankton organisms and particles at a rate of 18 frames per second while being towed vertically through the water column. Simultaneously,

depth, temperature, conductivity, oxygen content and fluorescence were recorded to relate the zooplankton abundance to the environmental conditions. The AQUAscat was mounted to the upper part of the LOKI frame, facing sideward and recording the acoustic backscatter at 0.5, 1, 2 and 4 mHz.

Fig. 9.5: Mesozooplankton sampling stations in the Hausgarten area. Blue dots: MultiNet + LOKI/ AQUAscat casts. Yellow dots: LOKI/AQUAscat casts.

Preliminary/expected results

Most samples will be analysed in home laboratories at AWI in Bremerhaven (biogeochemical parameters, phytoplankton abundance and molecular biology, zooplankton community composition and distribution), respectively GEOMAR in Kiel (bacterioplankton).

Zooplankton

Zooplankton abundances as roughly estimated during preservation of the Multinet samples were generally highest in the eastern part of the Fram Strait, where the relatively warm Atlantic water masses of the West Spitsbergen current prevail, and lowest in the western part, which is influenced by the cold polar water masses of the East Greenland current. At all stations, the zooplankton community was dominated by calanoid copepods. In the Atlantic domain, the boreal species *Calanus finmarchicus* was often found in large numbers from the surface down to 1,000 m depth, whereas its sibling species, the Arctic *C. hyperboreus*, was mainly found below 1,000 m. In the polar domain of the Fram Strait, *C. hyperboreus* was also found at the surface, however, in relatively low numbers. The polar species *C. glacialis* was only present close to the East Greenland shelf. Other mesozooplankton taxa that occurred regularly were chaetognaths, ostracods, amphipods, euphausiids and jellyfish.

During the frontal study, LOKI/Aquascat casts down to 500 m water depth were conducted at the centre of the frontal feature, at its rim and outside of it. A first evaluation of the LOKI pictures showed that total zooplankton abundances were about twice as high in the centre as compared to the outside.

Detailed analyses of the MultiNet samples will be done later on at the AWI laboratories using a ZooScan and a stereo microscope. LOKI data and AQUAscat profiles will be analysed at the AWI laboratories and at Polar Scientific Ltd, Scotland, respectively.

Data management

Analyses of i.e. net or sediment trap samples, require tedious and time-consuming processing (species identification and enumeration) and, therefore, these analyses will take longer than chemical measurements. Thus, depending on the parameter as well as on the methods used for the analyses, it will take up to three years to complete our analyses. As soon as the data sets are available, other cruise participants may request and use them. When the data are to be published, they will also be submitted to the PANGAEA Data Publisher for Earth & Environmental Science and are then open for external use.

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10. SENSITIVITY OF ARCTIC ZOOPLANKTON TO TEMPERATURE CHANGE: FROM ECOPHYSIOLOGICAL EFFECTS TO COMMUNITY RESPONSE

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Background and objectives

In the Arctic marginal seas, such as Fram Strait, closely related zooplankton sister species of polar vs. boreal-Atlantic origin occur sympatrically. It is expected that the polar representatives will be replaced by boreal-Atlantic congeners in the course of global climate change and warming. In order to better understand the dynamics and potential effects of this shift in species composition, species-specific sensitivities of polar vs. boreal-Atlantic species to temperature increase were determined experimentally and with molecular genetic methods.

The Arctic Ocean and adjacent ice-covered seas are the areas most rapidly and strongly affected by global warming over the coming decades. Climate models predict a rise in air temperature in the Arctic by 3 to 6°C over the coming 50 years. The temperature increase will be similar throughout the year, but with stronger effects during the summer season, when it will result in a longer and more intense melting period of the sea ice. This changes the extent, thickness and seasonal coverage of sea ice, strongly affecting not only ice-associated biota, but also pelagic communities beneath the ice.

Zooplankton are particularly suitable as indicators of environmental change due to their rapid response (generally short life-cycles), direct coupling to physical forcing (relatively passive drifters) and the fact that they are not subject to targeted harvesting, which could bias or obscure other environmental impacts. In the North Atlantic, a northward shift of several hundred kilometers of the distribution ranges of many zooplankton species has been observed with more southerly species replacing northerly relatives at higher latitudes.

Climate change induced impacts on species composition also occur in Arctic marine ecosystems. Repetitive analyses of zooplankton community structure demonstrate substantial changes in species composition and biodiversity between the 1990s and 2006, both in Fram Strait and in Svalbard fjord systems. Boreal-Atlantic species have shifted in distribution further north and now dominate plankton communities in Fram Strait. For instance, in the AWI Hausgarten sediment trap time series from 2000 to 2012, the boreal-Atlantic amphipod *Themisto compressa* only occurred from 2004 onwards. Until 2010 only older individuals were present, whereas in 2011 the presence of ovigerous females and recently hatched juveniles provided first evidence of reproductive success of this southern invader at high-Arctic latitudes.

Such changes in species composition will have a strong impact on secondary production of Arctic seas, pelagic-benthic coupling processes and sedimentation rates, in particular as most of the boreal-Atlantic species are smaller and have a lower lipid content than their polar relatives. This has profound consequences for marine food chains in the Arctic. Often the larger, more lipid-rich polar species are the preferred prey for fish and seabirds.

Different components of polar food webs will react differently to climate change and increasing Atlantic inflow. Based on model calculations, a mismatch in the timing of the phytoplankton bloom and the seasonal ascent of the dominant copepod *Calanus hyperboreus* from its hibernation depth could disrupt pelagic food chains and lead to a system dominated by microzooplankton that would not support higher trophic levels and fisheries. In contrast, other studies predict an increased zooplankton production and a better food supply for pelagic fish in conjunction with warmer sea surface temperatures. Thus, present predictions are still highly controversial and the effects of an increasing Atlantic inflow on pelagic biodiversity and productivity represent key questions for future ecological research in the Arctic. Further studies on the physiological and ecological response of key species to ocean warming and an increasing Atlantic inflow are needed to assess and forecast potential impacts of global change on marine pelagic ecosystems in Arctic seas.

During PS107, our research focused on the ecophysiology, gene activity and gene regulation of polar vs. boreal-Atlantic zooplankton sister species in order to establish at which temperature thresholds changes in zooplankton species composition will occur and what consequences they will have. In detail, we worked on the following research questions:

- How do polar and boreal-Atlantic zooplankton sister species differ in their temperature tolerance and thresholds?
- How do they differ in gene activity and gene regulation during incubations at increasing ambient temperatures?
- How do polar and boreal-Atlantic zooplankton sister species differ in lipid content, fatty acid composition and, hence, nutritional value for potential predators?
- To what extent are the different pelagic species dependent on ice algal primary production, which will diminish in the course of global warming?
- To what extent are novel eDNA sampling and analytical techniques applicable and suitable for studies on mesozooplankton distribution in the Arctic?
- How did mesozooplankton species composition in Fram Strait change over the past 20 years?

Work at sea

Mesozooplankton was sampled by stratified vertical hauls down to 1,500 m with opening and closing nets (Hydro-Bios MultiNet, 150 µm mesh size) at 17 stations. At most stations, standard depth intervals of 1,500-1,000-500-200-50-0 m were sampled to allow comparison with previous data and integration with samples collected by AWI colleagues. Macrozooplankton such as amphipods was collected by double oblique tows with a Bongo net (500 and 330 µm mesh size). Usually two successive Bongo net casts were conducted: first a shallow cast with 60 m maximum wire length and equipped with an under-water camera, thereafter a deep cast with 450 m maximum wire length. Sampling concentrated on two transects across Fram Strait at 79°N coinciding with the majority of the HAUSGARTEN stations and slightly north in order to cover the different hydrographic regimes.

Specimens of the target species were sorted immediately after the catch in a temperaturecontrolled lab container and used for respiration and feeding experiments onboard, incubations at increasing ambient temperatures or genetic and biochemical analyses in the home labs. The remains of the samples were preserved in formaldehyde, ethanol or RNA later for later quantitative analysis of species composition and abundance.

To establish species-specific temperature sensitivities, more than 120 respiration measurements were conducted onboard with polar and boreal-Atlantic zooplankton species at different ambient temperatures (0°C, 4°C, 8°C). Individuals were placed in gas-tight incubation bottles, filled with filtered and oxygenated sea water, and kept in a water bath in a temperature-controlled incubator usually for 24 hours. Respiration rates were recorded by high resolution optode respirometry throughout the incubation at different temperatures. In parallel, additional specimens were incubated at the same temperatures over several days in order to study differences in gene activity and gene regulation between polar and boreal-Atlantic zooplankton and potential responses in gene regulation to increasing ambient temperatures. The transcriptomic analyses will be performed in cooperation with the University of the Algarve.

In order to investigate whether the gene expression under thermal stress varies between the Atlantic *Themisto abyssorum* and the genuine Arctic *T. libellula* and within each species across its geographical range, we carried out experiments where specimens were exposed to either a gradual increase in temperature or a "heat shock". Additional molecular genetic analyses of gene expression (transcriptomics) in the home lab will show which genes are most important for temperature acclimation in these species and whether gene expression differs interspecifically, i.e. between sister species and/or intraspecifically, i.e. between different geographic populations of the same species.

In order to establish differences in lipid content, composition and nutritional value, individuals of polar and boreal-Atlantic zooplankton species were collected and deep-frozen at 80°C onboard for determination of dry mass and lipid content at Bremen University. A quantitative assessment of the different caloric and nutritional value of polar vs. boreal-Atlantic zooplankton will help to better understand the effects of shifts in zooplankton species composition on higher trophic levels such as fish and seabirds and for the structure and secondary production of Arctic marine ecosystems in general.

In cooperation with colleagues at AWI (Barbara Niehoff, Nicole Hildebrandt), long-term changes in mesozooplankton abundance, distribution and species composition over the past 20 years will be studied. For that purpose, comparative data are available from the same region of Fram Strait collected in 1997 (ARK XIII), 2006 (MSM 02/4), and 2016 (PS100).

Preliminary (expected) results

Description of Bongo net tows at stn. HG-IV

Two bongo net casts were carried out in the vicinity of a fin whale group aggregating and feeding during an entire day near the station HG-IV. The EK60 echosounder indicated the presence of big swarms of possibly euphausiids at a depth of 170-250 m at station HG-IV, at 1.5 nm southeast of that station, in the vicinity of the whales feeding, where the Bongo hauls were carried out, the echosounder showed smaller swarms forming around 200-300 m and between 200 m and the surface. The organisms recovered with the bongo tows showed a clear bathymetric segregation with *Themisto libellula* as only dominant taxon at the surface haul (0- 60 m) and *Calanus finmarchicus*, some *Thysanoessa inermis* and chaetognaths in the deeper haul (0-450 m), where *T. libellula* was much less abundant. The video recordings of the shallow haul did not show any presence of swarming euphausiids.

Mesozooplankton eDNA sampling and analysis

In order to establish whether novel environmental DNA (eDNA) sampling and analytical techniques would be applicable and suitable for studies on Arctic mesozooplankton, field sampling of *in-situ* eDNA was combined with an experimental approach to study eDNA excretion by zooplankton and eDNA degradation at different water temperatures. Water samples were taken at twelve stations from five different depths by means of a rosette water sampler attached to the CTD sonde and filtered to bind eDNA to a 0.2 µm cellulose filter. In addition, different densities of zooplankton species were incubated on board in 10 L buckets at different temperatures and their eDNA excretion monitored over several days. Thereafter, animals were removed from the buckets and the degradation of eDNA further monitored for several more days. Samples will be analysed by quantitative PCR and sequencing at the University of Bremen. Already during the cruise, a proof of principle was achieved. Zooplankton eDNA could be extracted from filters, and a PCR with specific primers showed positive results.

Data management

Data and samples to be collected during the cruise will be analysed by the cruise participants and collaborating scientists. They will also provide material for thesis projects at the Universities of Bremen and the Algarve. The results will be published within three to five years after the cruise. Geo-referenced data sets will be archived and made publicly accessible via the PANGAEA database, jointly operated by MARUM and AWI. DNA sequence data to be obtained from molecular genetic analyses will be archived and published in GenBank. Quantitative plankton samples preserved in formaldehyde or ethanol will be stored at BreMarE, Bremen University.

11. HAUSGARTEN: IMPACT OF CLIMATE CHANGE ON ARCTIC BENTHIC ECOSYSTEMS

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Grant No. AWI_PS107_11

Objectives

The Alfred-Wegener-Institut Helmholtz-Zentrum für Polar- und Meeresforschung (AWI) established the LTER (Long-Term Ecological Research) observatory HAUSGARTEN to detect and track the impact of large-scale environmental changes in the transition zone between the northern North Atlantic and the central Arctic Ocean, and to determine experimentally the factors controlling deep-sea biodiversity. Since 2014, this observatory has been successively extended within the frame of the HGF financed infrastructure project FRAM (Frontiers in Arctic marine Monitoring) and currently covers 21 permanent sampling sites on the West-Spitsbergen and East-Greenland slope at water depths between 250 and 5,500 m. Regular sampling as well as the deployment of moorings and different free-falling systems (bottom-lander) which act as local observation platforms, has taken place since the observatory was established back in 1999.

Fig. 11.1: Permanent sampling sites of the LTER Observatory HAUSGARTEN in Fram Strait (yellow markers: benthic sampling sites; green triangles: mooring sites; thick red arrows: North Atlantic Current)

During the *Polarstern* expedition PS107 investigations of the deep-sea benthos were conducted at all HAUSGARTEN stations (Fig. 11.1). The research programme covered ecosystem compartments from smallest benthic bacteria up to large epifauna within the benthic deep-sea realm. Benthic investigations comprised biochemical analyses to estimate the input of organic matter from phytodetritus sedimentation and to determine the activity and biomass of the small benthic biota. Further sediments were retrieved to study the composition of small sedimentinhabiting organisms (meiofauna). Results from these studies will help to describe the ecostatus of the benthic system.

Work at sea

Biogenic sediment compounds and meiofauna

Virtually undisturbed sediment samples were taken using a video-guided multicorer (TV- MUC; Fig. 11.2). Various biogenic compounds from these sediments were analysed to estimate activities (i.e. bacterial exoenzymatic activity) and the total biomass (i.e. particulate proteins, phospholipids) of the smallest sediment-inhabiting organisms. Results will help to describe ecosystem changes in the benthal of the Arctic Ocean. Sediments retrieved by the TV-MUC will also be analysed for the quantitative and qualitative assessment of the small benthic biota (meiofauna). The uppermost five centimetres of sediments, retrieved with the MUC, were sub-sampled to analyse parameters indicating the input of organic matter to the seafloor as well as sediment-bound biomass and benthic activity. Additional samples were taken to analyse the abundance and biomass of bacteria as well as meiofauna densities and the diversity patterns of nematodes. Sediment-bound chloroplastic pigments (chlorophyll *a* and its degradation products) represent a suitable indicator for the input of phytoplanktonic detritus to the seafloor, representing the major food source for benthic organisms. They can be analysed with high sensitivity by fluorometric methods. To estimate the potential heterotrophic activity of bacteria, we measured cleaving rates of extracellular enzymes using the model-substrate FDA (fluorescein-di-acetate) in incubation experiments. Bacterial activity and chloroplastic pigments were analysed on board. All other sub-samples were stored for later analyses of various biochemical bulk parameters at the home lab.

Fig. 11.2: Sediment sampling using a video-guided multicorer (TV-MUC)

Fig. 11.3: Giant Box Corer

Spatial and temporal variations in the structure of macrofaunal benthic communities

Samples were collected from 20 stations in the region of Fram Strait (Fig. 11.1). At each station samples were collected by a Giant Box Corer (0.25 m^2 sampling area; Fig. 11.3): two subsamples for macrozoobenthos (20 cm deep samples of 0.1 $m²$ surface area, samples sieved on 0.5 mm sieves and fixed with 4 % buffered formaldehyde), 6 subsamples for meiozoobentos (small cores of 10 cm^2 surface area, 5 cm deep): three preserved with 4% formaldehyde and three preserved in ethanol. Moreover, two samples for bacteria (surface sediments collected with a spoon, one samples frozen in -80ºC and one preserved in 10% formaldehyde) and three samples for sediment characteristics (small cores of 10 cm² surface area, 5 cm deep samples for grain size and particulate organic carbon (POC) content and 2 cm deep for photosynthetic pigments). Samples for grain size and POC were frozen in -20ºC, while samples for photosynthetic pigments were frozen in -80ºC.

Sample processing: In laboratory all metazoan organisms will be identified to the possible lowest taxonomic resolution, counted, photographed with camera connected to stereomicroscope and measured with use of digital image analysis in LAS v4.2 software. For the polychaete specimens, which were fragmented during the sample sieving, total length of the specimens will be calculated using equations of the regression between the total length and the width of a selected chaetiger (specific to the family or order; Górska et al., manuscript in preparation). Semiautomated image analysis method will be used for nematode measurements (Mazurkiewicz et al., 2016). Individual biovolumes will be converted to dry weight [µg]. Benthic biomass size spectra will be constructed by plotting the total biomass in each size class against the log2 transformed size of a class. Each individual will be classified into log2 size classes based on its dry weight [µg]. Individual biomass data will be used to calculate annual production and respiration of the studied communities. Production/biomass ratio and annual secondary production as well as mass specific respiration rate and respiration for macrofauna will be estimated using the Artificial Neural Network models as proposed by Brey (2012). Meiofaunal production and respiration will be estimated using equations published by Schwinghamer et al. (1986). Also selected sediment samples from box corer will be analysed for environmental parameters including chloroplastic pigments, organic carbon content, total nitrogen, isotopic signatures (δ13C), grain size, water content in sediments will be assessed as an indicator of sediment stability.

Preliminary (expected) results

Biogenic sediment compounds and meiofauna

Comparing the concentrations of sediment-bound Chlorophyll *a* and the potential bacterial esterase activity (Fig. 11.4) along the east-westerly transect, crossing the Fram-Strait, we detected the expected huge differences between both hemispheres. Those are superimposed by a general decrease of values with increasing water as well as sediment depth.

Upcoming analyses of additional parameters at the home laboratory will show whether the observed long-term trends at HAUSGARTEN observatory will continue and to which extend Climate Change induced processes might be responsible for the observed changes within the deep-sea ecosystem.

Spatial and temporal variations in the structure of macrofaunal benthic communities

A first inspection of the collected megafauna on board gave us the opportunity to take some nice preliminary pictures of several common species (Fig. 11.5).

Fig. 11.4: Sediment bound Chlorophyll a and esterase activity in the upper five sediment centimetres across the Fram-Strait (transect shown on map)

Fig. 11.5: Various macrofauna species from box-corer casts (from left to right, top: Amphipoda, Diastylis sp., Ophioridea; middle: Elpidia sp., Chiton, Gastropoda; down: Polychaeta, Polychaeta, Sipunculidea)

Collected material allows us to provide an extensive (in terms of spatial and temporal ranges) assessment of structure (standing stocks and size structure) and function (secondary production, respiration and carbon demand) of the benthic component of the Arctic deep sea ecosystems. It will also provide observations of the current response to the increasing advection of Atlantic water and associated changes in productivity of the subpolar seas and the basis for the future scenarios predictions. The changes in benthic productivity will have further consequences for the carbon flow through the ecosystem and the reintroduction of the detrital carbon into the food webs, as well as sustaining the benthivorous upper trophic levels (fish and sea mammals) that can have societal and economic impacts, especially in the era of the increasing fishing activity in the opening Arctic. Increasing carbon consumption by benthic communities may also impact the levels of carbon burial in deeper sediment layers (organic carbon sequestration) which is considered to be high in the Arctic sediments (Smith et al., 2015) but can vary depending on the rate of consumption by benthic biota.

Data management

Further sample processing will be carried out at home laboratories. Data acquisition from the several types of investigation will be differently time-consuming. The time periods from post processing to data provision will vary from one year maximum for sensor data, to several years for organism related datasets. Until then preliminary data will be available to the cruise participants and external users after request to the senior scientist. The finally processed data will be submitted to the PANGAEA data library. The unrestricted availability from PANGAEA will depend on the required time and effort for acquisition of individual datasets and its status of scientific publication.

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12. FRAM POLLUTION OBSERVATORY: MARINE ANTHROPOGENIC LITTER AND MICROPLASTICS IN DIFFERENT ARCTIC ECOSYSTEMS

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Objectives and scientific programme

Marine litter has long been on the political and public agenda, as it has been recognized as a rising pollution problem affecting all oceans and coastal areas of the world. There is currently a discrepancy of several orders of magnitude between estimates of global inputs of plastic litter, with figures derived from field measurements highlighting again the question: 'Where is all the plastic?'. Degradation of larger litter items into smaller particles termed 'microplastics' may be one reason for this discrepancy. Another possibility is that certain ecosystem compartments such as water column have not been considered so far with the latest research suggesting that the Arctic is an accumulation area for marine plastic. A newly added component to the open-ocean infrastructure FRAM (FRontiers in Arctic marine Monitoring) allows for the observation of marine litter and microplastics and other pollutants in different ecosystem compartments over long time scales.

The sampling campaign during PS107 has aimed to collect samples from different marine compartments to answer these open questions. Six different sampling campaigns were executed during PS107 to assess the spatial and temporal distribution of macrolitter and microplastic. Photographic surveys undertaken by a towed camera system (Ocean Floor Observation System, OFOS) were done to observe the change of the amount of macrolitter on the seafloor. Analyses of OFOS images obtained previously at the LTER HAUSGARTEN indicates a significant increase of litter on the seafloor between 2002 and 2014 (Bergmann and Klages, 2012; Tekman et al., 2017). Recent evidence suggests high concentrations of microplastics, a degradation product of larger plastic items, in Arctic sediments (Bergmann et al., 2017). The upper layer of sediment was sampled with a Video Multi Corer for further analysis of microplastic particles in deep-sea sediment. *In-situ* pumps were deployed to get filtered water samples from different target depths of the water column. Litter was also recorded floating at the sea surface in the HAUSGARTEN area (Bergmann et al., 2015). A towed neuston microplastic catamaran was used at locations where it was feasible to tow through ice sheets to assess the amount of microplastic particles in surface waters. Visual litter surveys were done to monitor the amount of floating marine litter during the transit from/ to Tromsø and around the Fram Strait. Snow samples were taken on an ice floe to assess the role of athmospheric transport in microplastic distribution at the Arctic. This work contributes to the Pollution Observatory of the infrastructure program FRAM.

Work at sea

Seafloor

Five OFOS (Ocean Floor Observation System) transects were executed at the East Greenland (EGIV) and Hausgarten (S3, HGI, HGIV, N3) stations. 15 sediment samples were taken from multiple corer deployments along the latitudinal and bathymetric transect and frozen in tinfoil for assessments of microplastic concentrations (G. Gerdts) and analysis of other pollutants. Sediment sample from HGII was splitted into layers of 1cm to assess microplastic amounts in the layers. Sea cucumbers (*Elpidia heckeri*), starfish (*Bathybiaster vexilifer*), sea spiders (*Colloscendeis proboscidea*) and some other unidentified organisms found in sediment samples were frozen in glass jars for further analysis of microplastic in their tissues.

Water Column

Five *In-situ* pumps were deployed during deep CTD casts at 8 stations at the East Greenland (EGI, EGIV) and HAUSGARTEN (S3, HGI, HGIV, HGIX, N3, N5) to 5 depths (Near surface, chlorophyll maximum, 300m, 1000m, near seafloor) of the water column. Between 200 and 400 liter of seawater was filtrated during 1 to 2 hours of deployment.

Sea Surface

49 neuston surveys of around 1h duration were conducted to determine densities of floating litter when the ship was in transit to another station or from/to Tromsø or during underway CTD casts. 17 Volunteers took turns in shifts during the transit between Longyearbean and Tromsø. Nine samples (EGI, EGIV, S3, HGI, HGIV, HGIX, N3, N5, SVI) were taken by a neuston catamaran to determine macro- and microplastic concentrations (> 300 µm) at the sea surface. For some areas, high amount of ice sheets prevented to deploy the catamaran in the vicinity of standard HAUSGARTEN station locations. At these areas, the catamaran was deployed at the first feasible location during the transect to the next station where it was possible to tow 2-3 miles without ice sheets. Snow samples were taken during a helicopter flight to an ice floe to assess airborne transport of microplastic in the Arctic.

Preliminary (expected) results

Floating litter was observed during 34 out of 49 transects. 139 floating litter items were recorded. 130 of recorded items were plastic. The distribution of floating litter items was patchy but preliminary results showed that the transects with most items are concentrated in central Fram Strait area. Several macrolitter items was observed in catamaran samples from HGIX and SVI (Fig. 12.1).

Further analysis with (μ) fourier transform infrared spectroscopy (FTIR) is needed to assess microplastic amounts in catamaran, snow (Fig. 12.2), sediment and *in-situ* pump (Fig. 12.3) samples.

Obvious litter items were observed during the OFOS transect at the central HAUSGARTEN station HGIV (Fig. 12.4). Possible litter items were recorded at all other sampling stations. However standard image analysis is needed to be certain whether those objects are litter items or not.

Fig. 12.1: Photos: (1, 2) Floating litter items. Credit: Kajetan Deja Photo: (3) Catamaran sample from SVI. Credit: Mine Tekman

Fig. 12.2: Images documenting helicopter-based snow-sampling for microplastic on an ice floe Photo: (1) Sampling area. Credit: Kajetan Deja; Photo: (2) Sampling snow. Credit: Kajetan Deja

Fig 12.3: Photo: (1) Deployment of in-situ pump for microplastic analysis. Credit: Lennard Frommhold Photo: (2) Taking blank sample onboard. Credit: Mine Tekman

Data management

All OFOS images, videos and metadata will be uploaded to PANGAEA as will be data on microplastic concentration and results from neuston litter surveys. These data will also be uploaded to the online portal 'LITTERBASE' [\(www.litterbase.org](http://www.litterbase.org)).

Fig. 12.4: Litter items found on the seafloor during the OFOS transect at HGIV (Photos: M. Tekman/OFOS/AWI).

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APPENDIX

- **A.1 PARTICIPATING INSTITUTIONS**
- **A.2 CRUISE PARTICIPANTS**
- **A.3 SHIP'S CREW**
- **A.4 STATION LIST**

A.1 TEILNEHMENDE INSTITUTE / PARTICIPATING INSTITUTIONS

A.2 FAHRTTEILNEHMER / CRUISE PARTICIPANTS

A.3 SCHIFFSBESATZUNG / SHIP'S CREW

A.4 STATIONSLISTE / STATION LIST PS107

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