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1 **Title**

2 Classification of sheep urination events using accelerometers to aid improved measurements of
3 livestock contributions to nitrous oxide emissions

4

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22 Conflict of interest: None

Abbreviations

VeDBA: Vectorial Dynamic Body Acceleration; VeDBAs: Smoothed Vectorial Dynamic Body Acceleration; PSD: Power spectrum Density; StX, StY, StZ: Static acceleration on the X, Y, and Z axes; DyX, DyY, DyZ: Dynamic acceleration on the X, Y and Z axes; TP: True Positives; TN: True Negatives; FP: False Postives; FN: False Negatives

23 **Abstract**

24 Livestock emissions account for 74 % of nitrous oxide contributions to greenhouse gases in the UK.
25 However, it remains uncertain how much is directly attributable to localised sheep urination events,
26 which could generate nitrous oxide emission ‘hot spots’. Currently, IPCC emission factors are mainly
27 extrapolated from lowland grazing systems and do not incorporate temporal or spatial factors related
28 to sheep behaviour and movement. Being able to gather data that reliably measures when, where, and
29 how much sheep urinate is necessary for accurate calculations and, to inform best management
30 practices for reducing greenhouse gas emissions and minimizing emission-based climate change.

31 Animal-attached movement sensors have been shown to be effective in classifying different
32 behaviours, albeit with varying classification accuracy depending on behaviour types. Previous
33 studies have used accelerometers on cattle and sheep to assess active and non-active behaviours to
34 help with grazing management, although no study has yet attempted to identify sheep urination events
35 using this method.

36 We attached tri-axial accelerometer sensor tags to thirty Welsh Mountain ewes for thirty days to
37 assess if we could identify urination events. We used random forest models using different sliding
38 mean windows to classify behaviours. Urination had a distinctive pattern and could be identified from
39 accelerometer data, with a 5 s window providing the best recall and a 10 s window giving the best
40 precision. ‘State’ behaviours considered (foraging, walking, running, standing and lying down) were
41 also identified with high recall and precision. This demonstrates the extent to which the identification
42 of discrete ‘event’ behaviours may be sensitive to the window size used to calculate the summary
43 statistics. The method shows promise for identifying urination in sheep and other livestock, being
44 minimally invasive compared to other methods, and has clear potential to inform agricultural
45 management practices and policies.

46

47 **Keywords**

48 Biologging, Climate change, Discrete behaviour, Greenhouse gas emissions, Sheep, Urination

49 **1. Introduction**

50 Agriculture contributes to 10 % of the total greenhouse gas emissions in the UK, with 74 % arising
51 from nitrous oxide (N₂O) and 51 % from methane emissions (DEFRA, 2016). The latter is largely due
52 to enteric fermentation by cattle and sheep (DEFRA, 2016), but N₂O is principally generated in the
53 soil via nitrification and subsequent denitrification. Urine from livestock contains high concentrations
54 of urea which can be hydrolysed in the soil to ammonium and subsequently nitrified. This means that
55 urine patches can act as ‘hot spots’ for N₂O emissions (Hoogendoorn *et al.*, 2016; Marsden, Jones &
56 Chadwick, 2016). There are uncertainties regarding the estimates of direct N₂O emission levels from
57 urine and dung deposited by livestock, particularly from sheep and extensively grazed systems.
58 Emission factors are currently extrapolated from cattle studies conducted in intensively managed
59 systems (UNFCCC, 2016). The uncertainties surrounding N₂O emissions are also higher because
60 precise measurements that incorporate spatial and temporal factors, along with animal behaviour and
61 movement, are lacking (DEFRA, 2016). Being able to monitor when livestock urinate and understand
62 any behavioural patterns that elucidate where and how often they urinate would help to reduce this
63 uncertainty. Combining such data with other experimental studies to measure direct N₂O emissions
64 released from soil due to urination in relation to edaphic factors, would enable more accurate
65 calculations and better understanding of its contribution to climate change.

66

67 Previous studies have utilised thermistors in conjunction with GPS to determine the spatial
68 distribution of urination events (Betteridge *et al.*, 2010). These have been modified to include a
69 measure of urine volume and nitrogen content via refractive index (Betteridge *et al.*, 2013;
70 Misselbrook *et al.*, 2016; Shepherd *et al.*, 2016). Flow meters in combination with data loggers have
71 also been used to record cattle urine frequency and volume (Ravera *et al.*, 2015), but all these
72 methods are quite invasive. The use of tri-axial accelerometers attached to a range of animals has
73 proven to be a powerful method for determining animal behaviour (Shepard *et al.*, 2008; Nathan *et*
74 *al.*, 2012; McClune *et al.*, 2014), although they have not yet been used to specifically detect urination
75 events.

76 Methods used for analysing accelerometer data vary in terms of variables used to classify behaviours
77 and the precise way the data are processed. Approaches used include template-matching (Walker *et*
78 *al.*, 2015) and various clustering approaches (Sakamoto *et al.*, 2009; Nathan *et al.*, 2012), with
79 accuracy depending on circumstance. In many clustering methods, the size of window used to
80 summarise the data plays an important role in the accuracy with which the data can be classified
81 (Gjoreski, Gams & Chorbev, 2010; McClune *et al.*, 2014). For example, Lush *et al.* (2015) used a 5 s
82 window to classify brown hare (*Lepus europaeus*) behaviour resulting in high levels of classification
83 accuracy for running, feeding and vigilance behaviours (> 90 %), but less than 50 % accuracy for
84 resting, scratching and grooming. Similarly, McClune *et al.* (2014) used a 2 s window to analyse
85 badger (*Meles meles*) behaviour and classified resting with nearly 100 % accuracy, but trotting,
86 walking and snuffling was between 75 – 80 % accuracy, while Wang *et al.* (2015) also used a 2 s
87 window to classify puma (*Puma concolor*) behaviour and achieved greater than 90 % classification
88 accuracy for resting, walking, running and trotting, whilst feeding was 64 % and grooming was 0 %.

89

90 The variation in classification accuracies stem, in part, from the length of time over which a behaviour
91 is expected to occur (Robert *et al.*, 2009). Behaviours, such as running, walking, feeding and resting
92 that tend to occur over extended periods of minutes or longer and regarded as ‘state’ behaviours
93 (Martin & Bateson, 1993), which facilitates their classification. In contrast, the short duration of many
94 ‘event’ behaviours (Martin & Bateson, 1993), such as urination, makes them particularly sensitive to
95 the window length used in the analysis (Robert *et al.*, 2009; Alvarenga *et al.*, 2015).

96

97 In this study, we used tri-axial accelerometers on Welsh Mountain ewes and then employed random
98 forest models on the data using different sliding mean windows to assess if we could identify
99 urination events. Accelerometers have been used previously on cattle and sheep to define active and
100 non-active behaviours such as standing, lying down, feeding, walking and running using 3, 5, and 10 s
101 windows (Martiskainen *et al.*, 2009; Robert *et al.*, 2009; Marais *et al.*, 2014; Alvarenga *et al.*, 2015).

102 However, this is the first study to attempt to use this approach to determine sheep urination events.
103 Ewes exhibit a characteristic squat when they urinate, hence we hypothesised that a rear-mounted tri-
104 axial accelerometer could reliably identify this behaviour. If successful it would provide a
105 methodology that could improve the accuracy of N₂O emission estimates and help to define how
106 much sheep contribute to greenhouse gas emissions.

107

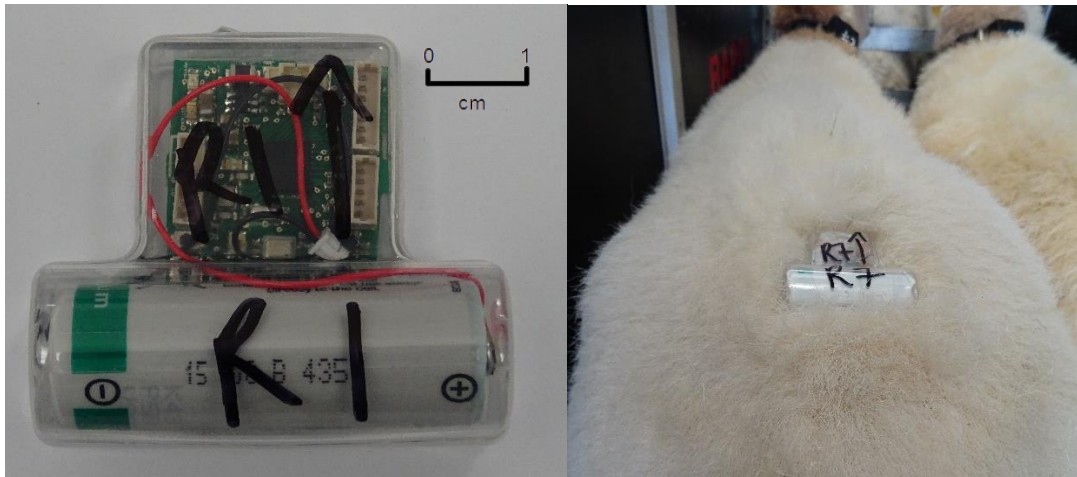
108 **2. Material and methods**

109 The study was carried out in a semi-improved enclosed 11.5 ha upland pasture at Bangor University's
110 Henfaes Research Centre, Abergwyngregyn, North Wales (53°13'13.75" N, 4°0'34.88" W). We
111 attached a 'Daily Diary' tag (Wildbyte Technologies Ltd, UK) to each of 30 barren Welsh Mountain
112 ewes for 30 d from 12th May – 16th June, 2016. Rear-mounted accelerometers were used since
113 accelerometers mounted on a collar were not able to detect urination events. Average sheep weight
114 was 36.8 kg (SD = 6.87 kg) and average age was 4.2 y (SD = 1.2). The work and methods used were
115 approved by Swansea University's Animal Welfare and Ethical Review Group (Reference IP-1516-5)
116 and by Bangor University's College of Natural Sciences Ethics Committee (Ethics approval code
117 CNS2016DC01).

118

119 *2.1 Daily Diary tags*

120 The Daily Diaries' recorded accelerometer data at 40 Hz on each of the three orthogonal axes; X
121 (surge), Y (sway), and Z (heave). The tags were powered by an A cell battery that was enclosed in a
122 vacuform plastic housing and sealed using Poly Cement (Humbrol, Hornby Hobbies, UK) (Fig. 1). A
123 small patch of wool was sheared from the rump of the sheep above their hips and the tags attached to
124 the remaining shorter wool using a solvent free epoxy adhesive (Fig. 1). Positioning the tag at the rear
125 of the sheep maximised the possibility of detecting the change in posture that occurs when sheep
126 urinate. The tags weighed 50 g which was less than 0.002 % of their body weight, and therefore was
127 likely to have minimal or no impact on sheep behaviour (Hobbs-Chell *et al.*, 2012).



128

129 **Fig. 1:** Rear tag consisting of a Daily Diary and an A cell battery and a tag in position on the rear of
130 the sheep.

131

132 *2.2 Behavioural measurements*

133 Twenty of the tagged sheep were filmed using a Panasonic HC-W570 full HD camcorder (Panasonic
134 UK & Ireland) over four separate filming sessions to record the different types of sheep behaviour.

135 Not all thirty sheep were filmed due to difficulties of observing all of them within the field. Sheep (n
136 = 20) were filmed for 5 min at a time unless they moved out of view. A total of 335 min of behaviour
137 from the video footage was logged, representing 15.9 ± 11.7 mins per sheep. Using the timestamp, the
138 logged behaviours were synchronised to the accelerometer data to create a labelled behaviour file. An
139 ethogram was produced of the main behaviours (Table 1). Six main behaviours were used to label the
140 accelerometer data and in subsequent analysis. Infrequently observed behaviours were omitted.

141 Urination events created a distinctive pattern within the acceleration trace that was identified using the
142 observed dataset (Fig. 2). Filmed urination events had an average duration of 7 s (SD = 4.9 seconds).

143

Behaviour	Description	Sample (seconds)
Foraging	Feeding with head down, small movements of head side to side and small steps forward	7595
Walking	Moving at slow pace	2170
Running	Moving at fast pace	126
Standing	Stationary with head raised	1653
Lying	Lying down with head raised or lowered	8345
Urinating	Rear of sheep lowers in a squatting position	127
Scratching	Using the back leg to scratch body or head	64
Grooming	Bending head to lick leg	8
Interaction	Physical interaction between two sheep such as head butting	8

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146 **Table 1:** Ethogram of sheep behaviour and number of seconds of observed behaviour logged (335
 147 min) from video footage of 20 sheep. Behaviours in bold are those used for further analysis.

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156 **Fig. 2:** Example time series of raw acceleration of the X, Y and Z axes from 40 Hz sampling rate
157 showing a single urination event of (a) 11 s duration, and (b) 5 s identified from the observed
158 behaviour (bounded in black box). The shaded rectangle represents a 3 s window. Urination is
159 associated with a sharp increase in the acceleration of the X axis combined with a decrease in
160 acceleration along the Z axis, and the Y axis generally remaining low, unless the sheep turns its head.

161

162 *2.3 Random Forest model*

163 Random Forests are machine learning models that test large numbers of regression or classification
164 trees on a training dataset to identify the best ensemble model. R (version 3.2.5), RandomForest
165 package (Liaw & Wiener, 2002) and RATTLE (R Analytical Tool To Learn Easily, Williams 2007)
166 were used for analysis. Previous studies have shown the merits of using random forest as a robust
167 method to classify behaviour from accelerometer data that also allows classification accuracy to be
168 measured for individual behaviours (Nathan *et al.*, 2012; Lush *et al.*, 2015; Fehlmann *et al.*, 2017).

169

170 A series of descriptive statistics were calculated using a 3, 5 and 10 s sliding windows on the
171 accelerometer data for the labelled behaviour dataset. These window sizes were chosen to allow
172 comparison with other behaviours and other studies that used the same window sizes. The variables
173 calculated were the static and dynamic acceleration (for each axis), the pitch, sway, Vectorial
174 Dynamic Body Acceleration (VeDBA), smoothed VeDBA with the mean, standard deviation,
175 minimum and maximum for all variables calculated. In addition, the maximum Power spectrum
176 Density (PSD) and associated frequency and second maximum PSD and frequency for each axis
177 (Wang *et al.*, 2015; Pagano *et al.*, 2017) were also calculated (Table 2, see Fehlmann *et al.*, (2017) for
178 example R code). This gave 52 variables to be used in the initial model. 75 % of the labelled dataset
179 was used as the training data to create the random forest model, with the remaining 25 % used to
180 validate the model's accuracy (how well the model classified the behaviours). 500 trees were grown
181 with 5 splits at each node. The mean decrease in accuracy was used to improve the model (Cutler *et*

182 *al.*, 2007) and resulted in VeDBA, dynamic acceleration, and frequency variables being removed,
 183 reducing the number of variables used in the final models to 30 (Table 2). A random forest model was
 184 created for each of the time windows to assess how window size affected the accuracy with which
 185 each of the main behaviours could be classified. We were particularly interested in how well the
 186 model could classify urination events.

187

Variable	Label	Definition
Raw acceleration	Raw X, Y, Z	Raw output of each acceleration channel
Static acceleration*	StX, StY, StZ	$StX = \frac{1}{n} \sum_{i=0}^{n-1} RawX - i$
Dynamic acceleration	DyX, DyY, DyZ	$DyX = StX - RawX$
Vectorial Dynamic Body Acceleration	VeDBA	$\sqrt{DyX^2 + DyY^2 + DyZ^2}$
Smoothed VeDBA*	VeDBAs	VeDBA calculated over sliding mean of 3, 5 or 10 s
Pitch*	Pitch	$Asin(StZ)$
Sway*	Sway	$Asin(StY)$
Power Spectrum Density* (PSD) and Frequency	PSD1X, PSD1Y, PSD1Z, PSD2X, PSD2Y, PSD2Z,	Fast Fourier analysis to calculate dominant frequencies, and respective strengths for windows of 3, 5 or 10 s for DyX , DyY and DyZ . Values used were the maximum and second maximum PSD and associated frequency calculated for each axis.

188

189 **Table 2:** Calculated variables from the raw X, Y, and Z acceleration axes used in the models. *
 190 indicates those variables used in the final models.

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194 2.4 Comparisons between models

195 To assess model performance for classifying the six behaviours, a confusion matrix was created based
196 on the number of true positives (TP), which was the number of events correctly classified, the true
197 negative (TN), which was those events correctly identified as being a different behaviour, the false
198 positive (FP), where behaviours were incorrectly classified as the behaviour, and false negative (FN),
199 where the behaviour was incorrectly classified as another behaviour (Martiskainen *et al.*, 2009;
200 Alvarenga *et al.*, 2015). This allowed us to calculate the precision ($TP / (TP+FP)$) and recall/sensitivity
201 ($TP / (TP+FN)$) for each time window generated from the validation data.

202 The Kappa statistic ($Kappa = (observed\ accuracy - expected\ accuracy) / (1 - expected\ accuracy)$), was
203 also calculated to compare models and evaluate the classifiers by comparing the observed accuracy with
204 the expected accuracy against random chance (Cutler *et al.*, 2007; Martiskainen *et al.*, 2009; Alvarenga
205 *et al.*, 2015).

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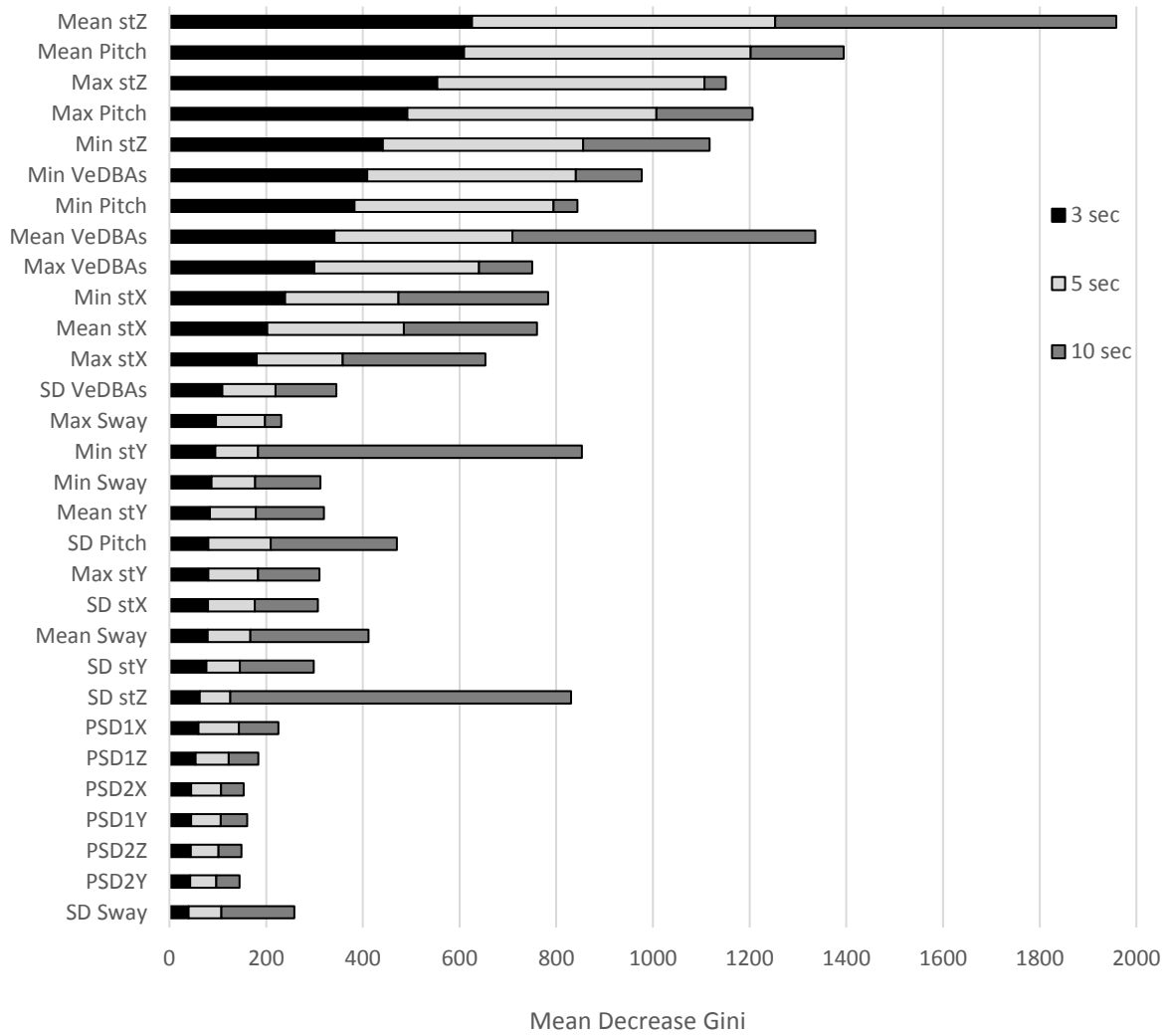
207 **3 Results**

208 3.1 Model fitting

209 The mean static acceleration of the Z axis was the most useful variable for classifying behaviours from
210 our acceleration data across all three different time windows (3, 5 and 10 s models; Fig. 3). Static
211 acceleration (Z and Y axis), pitch and smoothed VeDBA were also important for distinguishing among
212 behaviours performed by the sheep for each of our models, but the mean smoothed VeDBA, minimum
213 static acceleration of the Y axis (Min stY) and standard deviation of the static acceleration of the X axis
214 (SD stX) had higher importance in the 10 s model compared to both the 3 and 5 s models.

215

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217

218 **Fig. 3:** Variable importance for the 3, 5 and 10 s window models. For terms see Table 2.

219

220 The 3 s window model classified most behaviours with the lowest error rate, for both the training
 221 (Table 3) and the validation data (Table 4). Foraging was an exception to this, being classified with
 222 lower error on the 10 s window for the training data (2.6 %), as was urination, which was classified
 223 with lower error on the 5 s window validation data (28.0 %), although the training data error was
 224 much higher (54.3 %).

225

226

227

Behaviour	Class Error (%)		
	3 s window	5 s window	10 s window
Foraging	3.1	3.1	2.6
Walking	9.9	13.4	19.0
Running	16.6	18.8	27.9
Standing	21.5	23.5	23.0
Lying	0.2	0.3	0.4
Urinating	31.5	54.3	67.4
OOB estimate of error rate (%)	4.38	5.22	5.88

228

229 **Table 3:** Class errors (amount of classification error) for each behaviour using the training data to
230 create the Random Forest model for each time window.

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Observed behaviour (%)	Predicted behaviour (%)						Class Error
	Foraging	Walking	Running	Standing	Lying	Urinating	
3 s window	Overall error = 4 %, average class error = 13 %						
Foraging	97.7	2.2	0.1	0.06	0.0	0.0	2.0
Walking	7.7	91.3	0.2	0.2	0.6	0.0	9.0
Running	10	0.0	90.0	0.0	0.0	0.0	10.0
Standing	17.5	1.3	0.0	80.5	0.3	0.5	20.0
Lying	0.0	0.0	0.0	0.0	100	0.0	0.0
Urinating	16.0	8.0	0.0	4.0	8.0	64	36.0
Performance	Kappa = 0.945						Mean %
Precision	93.9	90.3	90.0	99.1	99.7	88.9	93.7
Recall/Sensitivity	97.7	91.3	90.0	80.5	99.9	64.0	87.2
5 s window	Overall error = 5 %, average class error = 15 %						Class Error
Foraging	97.0	2.8	0.1	0.1	0.0	0.1	3.0
Walking	12.3	86.5	0.4	0.5	0.4	0.0	14.0
Running	10.0	8.3	81.7	0.0	0.0	0.0	18.0
Standing	19.0	4.9	0.0	73.6	0.0	0.3	24.0
Lying	0.0	0.0	0.1	0.0	99.9	0.0	0.0
Urinating	16.7	0.0	0.0	0.0	11.1	72.2	28.0
Performance	Kappa = 0.927						Mean %
Precision	92.6	86.6	92.5	98.2	99.8	81.3	91.8
Recall/Sensitivity	97.0	86.5	81.7	75.7	99.9	72.2	85.5
10 s window	Overall error = 5 %, average class error = 20 %						Class Error
Foraging	97.4	1.9	0.1	0.7	0.0	0.0	3.0
Walking	15.2	84.4	0.0	0.2	0.2	0.0	16.0
Running	13.2	13.2	71.7	0.0	1.9	0.0	28.0
Standing	19.5	1.9	0.0	78.1	0.3	0.3	22.0
Lying	0.1	0.0	0.0	0.0	99.9	0.0	0.0
Urinating	5.6	0.0	0.0	27.8	16.7	50.0	50.0
Performance	Kappa = 0.926						Mean %
Precision	92.1	90.0	97.4	93.8	99.7	90.0	93.8
Recall/Sensitivity	97.4	84.4	71.7	78.1	99.9	50.0	80.3
Mean precision	92.9	89.0	93.3	97.0	99.7	86.7	
Mean recall	97.3	87.4	81.1	78.1	99.9	62.1	

242

243 **Table 4:** Confusion matrix of the validation datasets and the performance of the Random Forest model
244 in classifying six sheep behaviours using three different mean sliding time windows (3, 5 and 10 s). The
245 numbers in bold are the correct classifications. (Values are percentages)

246 *3.2 Model accuracy and performance*

247 Overall, the 3 s window model performed the best for most of the behaviours, with the highest kappa
248 statistic (Table 4). In fact, the kappa statistic was very high across all three models and, according to
249 Landis and Koch's (1977) criteria, was almost perfect (0.81 – 1.00). Running was predicted with the
250 highest precision in the 10 s window model, whereas, urination had the highest precision in the 10 s
251 window and the highest recall in the 5 s window.

252 The 3 s model had the highest mean recall across all six behaviours (Table 4). All behaviours except
253 urination had high mean precision and recall (> 75 %) across all models. Urination had high mean
254 precision (86.7 %) but the mean recall was lower at 62.1 %.

255

256 **4. Discussion**

257 *4.1 Behaviour identification in sheep*

258 Overall, the random forest approach identified the behaviours well, with the 3 s window model
259 performing the best for classifying 'state' behaviour (e.g. foraging, walking and lying) and relatively
260 well for the 'event' behaviour we were interested in; that is, urination, for both precision and recall.
261 Unsurprisingly, our ability to detect state behaviours were little affected by the size of window used,
262 because the duration of the window was great enough to incorporate multiples of any repetitive
263 frequency within the behaviour, while only being a small fraction of the likely length of any bout of
264 the behaviour. However, longer time windows have been found to perform less well, as found in a
265 study on cattle behaviour (Robert et al. 2009).

266

267 Conversely, urination, a discrete event behaviour, was the least well classified out of all the
268 behaviours, with the degree of success depending greatly on window size. In fact, although the 5 s
269 window model classified urination with the highest classification accuracy on the validation data the
270 classification accuracy for the training model was only 54 %. High training data error and low

271 validation error is indicative of a poorly fitting model (Sujatha, Prabhakar & Devi, 2013). Ideally, the
272 validation error should be low, and the training error marginally higher. Therefore, the 3 s model, with
273 a training error of 31.5 % and validation error of 36 %, indicates a better model fit. Model precision
274 for urination was relatively high across all models. However, it was the recall, critical for showing
275 how good a classification model is at correctly identifying the behaviour, which varied greatly. This
276 could be because the window may miss either the start and/or the end of urination events, which are
277 defined by the change in pitch (and the value of smoothed acceleration X and Z) as the sheep squats
278 and returns to standing (Fig. 2), interspaced with lower VeDBA, because sheep remain stationary
279 whilst urinating. Therefore, the interplay between window size and the duration of the urination event
280 may modulate the classification error overall. In addition, the sample size of urination events was one
281 of the lowest of our selected behaviours, as it was difficult to film, resulting in a reduced training
282 dataset to inform the model.

283

284 Urination had a visually very distinctive pattern within the raw acceleration data (Fig. 2), which arises
285 from the time-separated ‘squat’, ‘hold’ and ‘return-to-standing’ sequence. Such readily identifiable
286 patterns in the accelerometer trace may be better dealt with by an algorithm that accurately defines the
287 time-based order of important variables in sequence, as done by template matching (Walker *et al.*,
288 2015), for example. The immediate difficulty here, is coping with variable durations within such event
289 behaviours. It may also be more difficult for identifying behaviours that occur simultaneously within
290 state behaviours.

291

292 Despite the issues associated with identifying infrequent and transient behaviours like urination, this
293 study has nonetheless identified urination events from accelerometer data. This approach, therefore,
294 provides valuable information about urination frequency and duration. When combined with high-
295 resolution GPS data (e.g. Haddadi *et al.*, 2011) it can provide spatial and temporal information on
296 urine emissions (Fig. A1). This method of using rear-mounted tags to identify urination events would

297 not be suitable to detect urination events of rams, as they do not exhibit the characteristic squat
298 movement that is used for ewes. However, the number of rams grazing compared to breeding ewes
299 would be negligible and therefore would not have as much impact on greenhouse gas emissions.
300 Given that sheep movement is not random (Harris & O'Connor, 1980) their patterns of urination are
301 not expected to be either. In fact previous work over a six-day trial estimated that sheep deposit about
302 30 % of their urine over only 7.5 % of the pasture area used for grazing (Betteridge et al. 2010). This
303 heterogeneity of urine deposition to pasture soils could create highly concentrated 'hot spot' areas that
304 potentially release N₂O through nitrification and subsequent denitrification. By combining
305 information on where and when sheep urinate with data on N₂O emissions from urine patches on
306 different soil types and under different environmental conditions, could improve greenhouse gas
307 estimates from grazed pastures.

308

309 *4.2 Conclusions*

310 We suggest that our method of using a rear-mounted tri-axial accelerometer may provide a non-
311 invasive method to record urination events in sheep and other livestock to estimate urination patterns
312 (frequency and duration). This would provide important information to measure livestock urination
313 contributions to greenhouse gas emissions and to inform better agricultural management practices and
314 policies.

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328 **References**

- 329 Alvarenga, F.A.P., Borges, I., Palkovič, L., Rodina, J., Oddy, V.H. & Dobos, R.C. (2015). Using a
330 three-axis accelerometer to identify and classify sheep behaviour at pasture. *Appl. Anim. Behav. Sci.* **181**, 91–99.
- 331
- 332 Betteridge, K., Costall, D.A., Li, F.Y., Luo, D. & Ganesh, S. (2013). Why we need to know what and
333 where cows are urinating – a urine sensor to improve nitrogen models. *Proc. New Zeal. Grassl. Assoc.* **75**, 33–38.
- 334
- 335 Betteridge, K., Hoogendoorn, C., Costall, D., Carter, M. & Griffiths, W. (2010). Sensors for detecting
336 and logging spatial distribution of urine patches of grazing female sheep and cattle. *Comput. Electron. Agric.* **73**, 66–73.
- 337
- 338 Cutler, D.R., Edwards, T.C., Beard, K.H., Cutler, A., Hess, K.T., Gibson, J. & Lawler, J.J. (2007).
339 Random forests for classification in ecology. *Ecology* **88**, 2783–92.
- 340 DEFRA. (2016). Agricultural Statistics and Climate Change. *Dep. Environ. Food Rural Aff.* 1–109.
- 341 Fehlmann, G., O’Riain, M.J., Hopkins, P.W., O’Sullivan, J., Holton, M.D., Shepard, E.L.C. & King,
342 A.J. (2017). Identification of behaviours from accelerometer data in a wild social primate. *Anim. Biotelemetry* **5**, 6.
- 343
- 344 Gjoreski, H., Gams, M. & Chorbev, I. (2010). 3-Axial Accelerometers Activity Recognition. *ICT Innov.* 51–58.
- 345
- 346 Haddadi, H., King, A.J., Wills, A.P., Fay, D., Lowe, J., Morton, A.J., Hailes, S. & Wilson, A.M.
347 (2011). Determining association networks in social animals: Choosing spatial-temporal criteria
348 and sampling rates. *Behav. Ecol. Sociobiol.* **65**, 1659–1668.
- 349 Harris, P.S. & O’Connor, K.F. (1980). The Grazing Behaviour of Sheep (*Ovis aries*) on a High-
350 Country Summer Range in Canterbury, New Zealand. *N. Z. J. Ecol.* **3**, 85–96.
- 351 Hobbs-Chell, H., King, A.J., Sharratt, H., Haddadi, H., Rudiger, S.R., Hailes, S., Morton, A.J. &

352 Wilson, A.M. (2012). Data-loggers carried on a harness do not adversely affect sheep
353 locomotion. *Res. Vet. Sci.* **93**, 549–552.

354 Hoogendoorn, C.J., Luo, J., Lloyd-west, C.M., Devantier, B.P., Lindsey, S.B., Sun, S., Pacheco, D.,
355 Li, Y., Theobald, P.W. & Judge, A. (2016). Agriculture , Ecosystems and Environment Nitrous
356 oxide emission factors for urine from sheep and cattle fed forage rape (*Brassica napus* L .) or
357 perennial ryegrass / white clover pasture (*Lolium perenne* L ./ *Trifolium repens*). *Agric.*
358 *Ecosyst. Environ.* **227**, 11–23.

359 Landis, J.R. & Koch, G.G. (1977). The Measurement of Observer Agreement for Categorical Data.
360 *Int. Biometric Soc.* **33**, 159–174.

361 Liaw, A. & Wiener, M. (2002). Classification and Regression by randomForest. *R News* **2**, 18–22.

362 Lush, L., Ellwood, S., Markham, A., Ward, A.I. & Wheeler, P. (2015). Use of tri-axial accelerometers
363 to assess terrestrial mammal behaviour in the wild. *J. Zool.* **298**, 257–265.

364 Marais, J., Petrus, S., Roux, L., Wolhuter, R. & Niesler, T. (2014). Automatic classification of sheep
365 behaviour using 3-axis accelerometer data. In *Pattern Recognition Association of South Africa:*
366 1–6. Puttkammer, M. & Eiselen, R. (Eds). . Cape Town, South Africa.

367 Marsden, K.A., Jones, D.L. & Chadwick, D.R. (2016). The urine patch diffusional area: An important
368 N₂O source? *Soil Biol. Biochem.* **92**, 161–170.

369 Martin, P. & Bateson, P. (1993). *Measuring behavior: An introductory guide*. Second edi. Cambridge,
370 England: Cambridge University Press.

371 Martiskainen, P., Järvinen, M., Skön, J.P., Tiirikainen, J., Kolehmainen, M. & Mononen, J. (2009).
372 Cow behaviour pattern recognition using a three-dimensional accelerometer and support vector
373 machines. *Appl. Anim. Behav. Sci.* **119**, 32–38.

374 McClune, D.W., Marks, N.J., Wilson, R.P., Houghton, J., Montgomery, I.W., McGowan, N.E.,
375 Gormley, E. & Scantlebury, M. (2014). Tri-axial accelerometers quantify behaviour in the
376 Eurasian badger (*Meles meles*): towards an automated interpretation of field data. *Anim.*

377 *Biotelemetry* **2**, 1–6.

378 Misselbrook, T., Fleming, H., Camp, V., Umstatter, C., Duthie, C.A., Nicoll, L. & Waterhouse, T.
379 (2016). Automated monitoring of urination events from grazing cattle. *Agric. Ecosyst. Environ.*
380 **230**, 191–198.

381 Nathan, R., Spiegel, O., Fortmann-Roe, S., Harel, R., Wikelski, M. & Getz, W.M. (2012). Using tri-
382 axial acceleration data to identify behavioral modes of free-ranging animals: general concepts
383 and tools illustrated for griffon vultures. *J. Exp. Biol.* **215**, 986–96.

384 Pagano, A.M., Rode, K.D., Cutting, A., Owen, M.A., Jensen, S., Ware, J.V., Robbins, C.T., Durner,
385 G.M., Atwood, T.C., Obbard, M.E., Middel, K.R., Thiemann, G.W. & Williams, T.M. (2017).
386 Tri-axial accelerometers remotely identify wild polar bear behaviors. *Endanger. Species Res.* **32**,
387 19–33.

388 Ravera, B.L., Bryant, R.H., Cameron, K.C., Di, H.J., Edwards, G.R. & Smith, N. (2015). Use of a
389 urine meter to detect variation in urination behaviour of dairy cows on winter crops. *Proc. New
390 Zeal. Soc. Anim. Prod.* **75**, 84–88.

391 Robert, B., White, B.J., Renter, D.G. & Larson, R.L. (2009). Evaluation of three-dimensional
392 accelerometers to monitor and classify behavior patterns in cattle. *Comput. Electron. Agric.* **67**,
393 80–84.

394 Sakamoto, K.Q., Sato, K., Ishizuka, M., Watanuki, Y., Takahashi, A., Daunt, F. & Wanless, S.
395 (2009). Can ethograms be automatically generated using body acceleration data from free-
396 ranging birds? *PLoS One* **4**, e5379.

397 Shepard, E., Wilson, R., Quintana, F., Gómez Laich, a, Liebsch, N., Albareda, D., Halsey, L., Gleiss,
398 a, Morgan, D., Myers, A., Newman, C. & McDonald, D. (2008). Identification of animal
399 movement patterns using tri-axial accelerometry. *Endanger. Species Res.* **10**, 47–60.

400 Shepherd, M.A., Welten, B.G., Costall, D., Cosgrove, G.P., Pirie, M. & Betteridge, K. (2016).
401 Evaluation of refractive index for measuring urinary nitrogen concentration in a sensor worn by

402 grazing female cattle. *New Zeal. J. Agric. Res.* **60**, 23–31.

403 Sujatha, M., Prabhakar, S. & Devi, G. (2013). A Survey of Classification Techniques in Data Mining.
404 *Int. J. Innov. Eng. Technol.* **2**, 86–92.

405 UNFCCC. (2016). UK Greenhouse Gas Inventory, 1990 to 2014: Annual report for submission under
406 the framework convention on climate change. *Dep. Energy Clim. Chang.* 1–569.

407 Walker, J.S., Jones, M.W., Laramée, R.S., Holton, M.D., Shepard, E.L., Williams, H.J., Scantlebury,
408 D.M., Marks, N.J., Magowan, E. a, Maguire, I.E., Bidder, O.R., Di Virgilio, A. & Wilson, R.P.
409 (2015). Prying into the intimate secrets of animal lives; software beyond hardware for
410 comprehensive annotation in “Daily Diary” tags. *Mov. Ecol.* **3**, 1–16.

411 Wang, Y., Nickel, B., Rutishauser, M., Bryce, C., Williams, T., Elkaim, G. & Wilmers, C. (2015).
412 Movement, resting, and attack behaviors of wild pumas are revealed by tri-axial accelerometer
413 measurements. *Mov. Ecol.* **3**, 2.

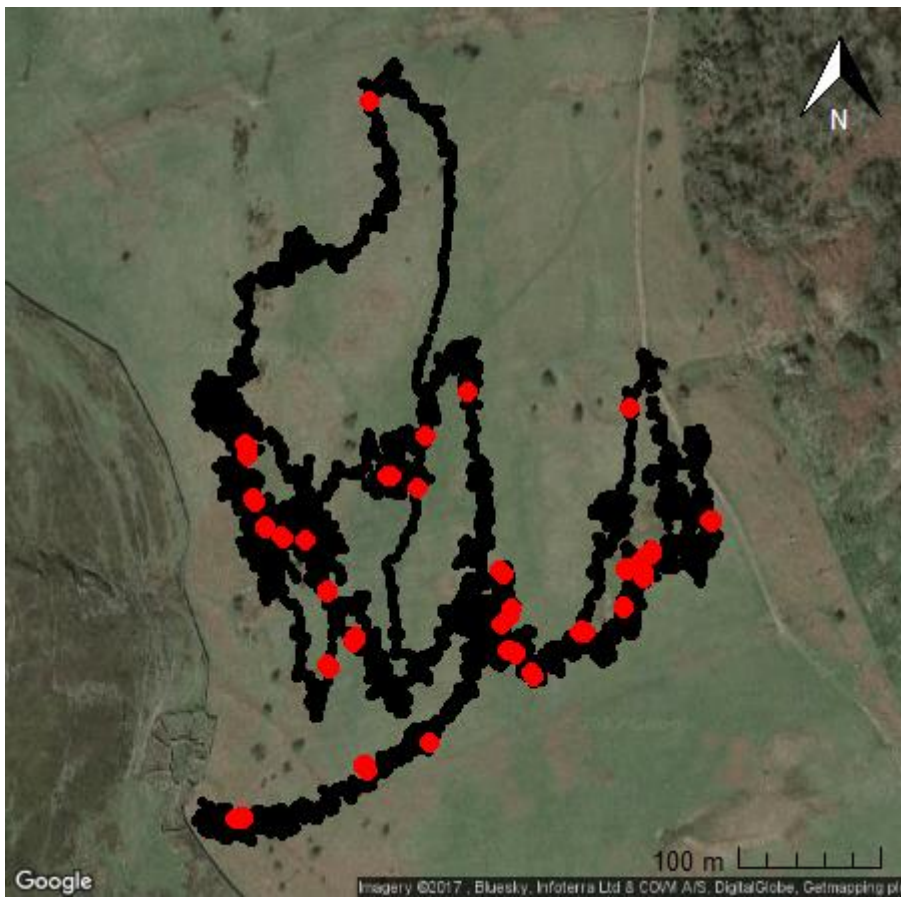
414 Williams, G. (2007). Rattle: A graphical user interface for data mining in R using GTK. *R J.* **1**, 45–55.

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416

417 **Appendix**

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419

420 **Fig. A1:** Movement of 1 sheep over the duration of a day plotted on the study site. Red dots are

421 urination events.