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A Thieno[2,3-*d*]pyrimidine Scaffold is a Novel Negative Allosteric Modulator of the Dopamine D₂ Receptor

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Abstract: Recently, a novel negative allosteric modulator (NAM) of the D₂-like dopamine receptors **1** was identified through virtual ligand screening. This ligand comprises a thieno[2,3-*d*]pyrimidine scaffold that does not feature in known dopaminergic ligands. Herein, we provide pharmacological validation of an allosteric mode of action for **1**, revealing that it is a NAM of dopamine efficacy and identify the structural determinants of this allostery. We find that key structural moieties are important for functional affinity and negative cooperativity, whilst functionalization of the thienopyrimidine at the 5- and 6-positions results in analogues with divergent cooperativity profiles. Successive compound iterations have yielded analogues exhibiting a 10-fold improvement in functional affinity, as well as enhanced negative cooperativity with dopamine affinity and efficacy. Furthermore, our study reveals a fragment-like core that maintains low μM affinity and robust negative cooperativity with markedly improved ligand efficiency.

▪ INTRODUCTION

The dopamine D₂ receptor (D₂R), a class A G protein-coupled receptor (GPCR), is a therapeutic target for central nervous system disorders such as schizophrenia and Parkinson's disease. Accordingly, it has been the subject of intensive drug discovery efforts for the last 60 years. Such efforts have focused on targeting the

1 orthosteric binding site of the D₂R where dopamine (DA) binds.^{1,2} Clinically marketed drugs indicated for the
2 treatment of schizophrenia display efficacy with respect to treating the positive symptoms of the disease.
3 These drugs all act either as competitive antagonists or partial agonists at the D₂R. However, these drugs can
4 cause deleterious side-effects. First-generation antipsychotics (FGAs) are associated with a high frequency of
5 extrapyramidal motor symptoms (EPS) caused by antagonism of D₂R signaling in the nigrostriatal system.³
6 Although, second generation antipsychotic (SGAs) are thought to be associated with a lower risk of EPS such
7 drugs are associated with other limiting side effects such as metabolic side effects.⁴
8
9 In order to adequately address the unmet medical need associated with antipsychotic drug discovery, novel
10 approaches are required. NAMs that display limited negative cooperativity with dopamine have been proposed
11 as a safer therapeutic option to treat the positive symptoms of schizophrenia, in particular for the onset of EPS,
12 because they would cause only partial blockade of the D₂R.⁵ Indeed, targeting allosteric sites within GPCRs
13 may be associated with a number of advantages over classical orthosteric drug discovery such as improved
14 subtype-selectivity and thus reduced off-target side-effects, as well as maintenance of spatiotemporal patterns
15 associated with endogenous neurohumoral signalling because they still allow the endogenous neurotransmitter
16 to bind the receptor.^{6,7}

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35 Virtual screening of orthosteric and putative allosteric sites within the human dopamine D₃ receptor
36 (D₃R) recently identified an active thienopyrimidine compound (**1**, Figure 1).⁸ In this study, screening was
37 undertaken using two optimised crystal-structure-based models (PDB ID: 3PBL)⁹: the receptor with an empty
38 (unliganded) binding pocket (D₃R^{APO}), and the receptor complex with DA (D₃R^{Dopa}). In particular, D₃R^{Dopa}
39 was used to screen for allosteric compounds that can bind along with the endogenous ligand. Interestingly, **1**,
40 although identified using D₃R^{APO}, was predicted to bind in an allosteric pose and showed the highest potency
41 of DA inhibition in vitro.⁸ Furthermore, this compound also displayed activity at the D₂R, and as such, may
42 represent a novel allosteric scaffold for this receptor.

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53 To date, very few ligands have been characterised as allosteric modulators of D₂-like dopamine receptors
54 including the drug-like ligand SB269652 (Figure 1); which acts to negatively modulate dopamine affinity at
55 the D₂R and D₃R.¹⁰⁻¹² Structure-based virtual ligand screening (VLS) provides an attractive path for efficient
56 discovery of allosteric ligands. However, it is important to validate this approach through optimisation of
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initial VLS hits.^{13,14} Currently, there is no structure-activity relationship (SAR) data associated with **1**, but there is much scope for further structural interrogation. We therefore undertook a study to identify novel allosteric modulators for the D₂R, based on the structure of **1**. Through modifications to the scaffold of **1**, we aimed to identify key molecular features responsible for its allosteric action at the D₂R (Figure 2). We applied analytical pharmacological methods to allow us to supplement this SAR study with measures of allosteric ligand action to include not only allosteric ligand affinity (K_B), but also the magnitude of modulatory effects upon dopamine affinity (α) and efficacy (β). Such an approach may be useful as a starting point to understand the relationship between allosteric ligand behavior and the in vivo efficacy of such modulators. Indeed, to explore negative allosteric modulation as an approach to treat the positive symptoms of schizophrenia it would be advantageous to have a set of D₂R NAMs that display a range of different allosteric behaviors from modulation of dopamine affinity through to modulation of dopamine efficacy. Herein, we report the design, synthesis, and detailed pharmacological characterisation of **1**, together with a library of structural analogues. We provide evidence that **1** does indeed act via an allosteric site at the D₂R and reveal that **1** exerts distinct pharmacology to that of SB269652, whereby it acts to modulate dopamine efficacy rather than dopamine affinity. Promising low molecular weight candidates with marketedly improved lipophilic ligand efficiencies (LLE) have been identified, and thus represent an attractive starting point for the development of novel D₂R NAMs that modulate dopamine efficacy, affinity or both.

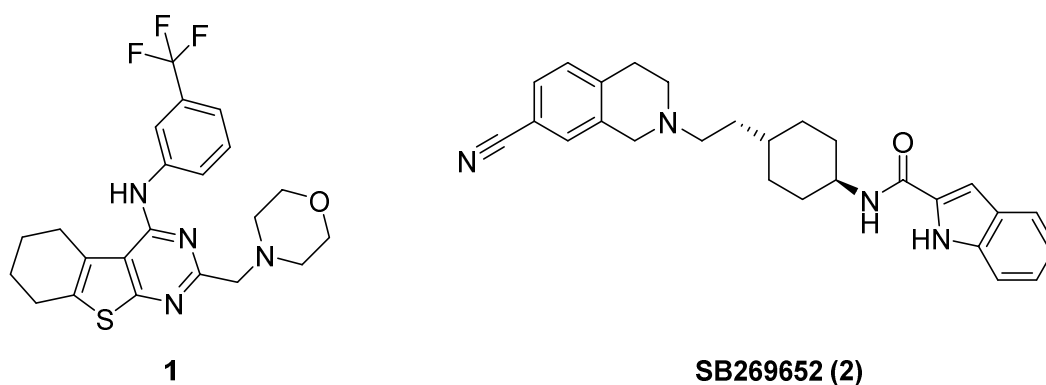
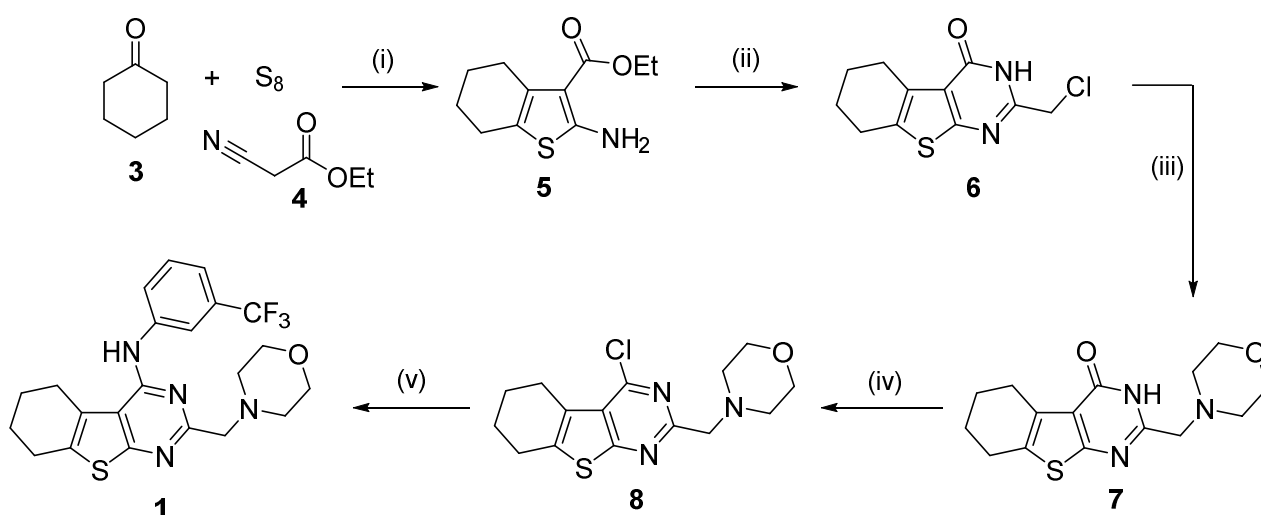


Figure 1. Compound **1** (above left) (2-(morpholinomethyl)-*N*-(3-(trifluoromethyl)phenyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4-amine) (above left) was identified from a virtual ligand screen targeting orthosteric and allosteric pockets of the dopamine D₂-like receptors. SB269652 (**2**) (above right), is the first drug-like NAM of the dopamine D₂ receptor.¹⁰

RESULTS & DISCUSSION

Chemistry. In order to validate the pharmacological activity of lead compound **1**, it was resynthesised using a five step chemical synthesis outlined in Scheme 1.¹⁵ Gewald chemistry permitted the one-pot assembly of ethyl 2-amino-4,5,6,7-tetrahydrobenzo[*b*]thiophene-3-carboxylate **5** via commercially available cyclohexanone **2**, ethyl cyanoacetate **3** and sulfur, catalysed by diethylamine in absolute ethanol at 60 °C to afford the fused thiophene **5** as brilliant orange needles.¹⁵ Next, **5** was subjected to an acid catalysed intermolecular cyclisation in 1,4-dioxane at room temperature using chloroacetonitrile and hydrogen chloride gas generated *in situ* (Kipp's apparatus) to yield the 2-chloro substituted thieno[2,3-*d*]pyrimidin-4(3*H*)-one core scaffold **6** as a precipitated white solid.¹⁶ Subsequent *N*-alkylation with morpholine was achieved in DMF at 100 °C followed by cooling to room temperature to precipitate the desired product **7**. Activation of the pyrimidinone was accomplished with an adapted procedure using phosphorus oxychloride (POCl₃) and catalytic DMF in toluene to give the corresponding chloropyrimidine **8** as a brown oil in good yield. Finally, **8** was subjected to microwave assisted nucleophilic aromatic substitution with 3-(trifluoromethyl)aniline in *i*-PrOH to afford the target compound **1**, following chromatography and recrystallisation from chloroform.

Scheme 1. Chemical Synthesis of 1^a



^aReagents and conditions: (i) morpholine, 0-45 °C, 80%; (ii) chloroacetonitrile, 1,4-dioxane, HCl_(g) (Kipp's apparatus), 50 °C, 2 h, 85%; (iii) DMF, morpholine, Et₃N, 100 °C, 2 h, 80%; (iv) POCl₃, toluene, 100 °C, 4 h, 75%; (v) *i*-PrOH, *m*-trifluoromethylaniline, MWI, 2 h, 70%.

1 *Single Modifications to Fused Cyclohexane Moiety, 2-, and 4-Positions of 1*. In addition to **1**, we generated a
2 focused library of thieno[2,3-*d*]pyrimidine analogues bearing single structural modifications to examine the
3 effect on functional binding affinity and allosteric pharmacology of the scaffold (Figure 2). This included
4 modifying the fused ring system – initially observing the effect of differing ring size (fused cyclopentane **14a**,
5 cycloheptane **14b** and cyclooctane **14c**) (Schemes 2 and 3), as well as the effects of removing substitution at
6 the 5- and 6-position of the thienopyrimidine **14j**. Moreover, we explored functionalisation of both the 5- and
7 6-positions with various substituents (6-phenyl, **14d**; 5-phenyl, **14e**; 6-benzyl, **14f**; 5-cyclohexyl, **14g**; 5,6-
8 dimethyl, **14h**; 6-cyclohexyl, **14i**) (Scheme 3) to provide an understanding of the chemical space surrounding
9 this moiety for further future elaboration. We investigated the effect of removing the CF₃ substituent on the
10 aryl ring **16a**, and the influence of incorporating additional substituents with different electronic effects (3-F,
11 **16b**; 3-OCH₃, **16c**; 3-OH, **17**) (Scheme 4). In addition to this, we explored the role of the secondary aliphatic
12 amine bridging the substituted aryl moiety in **1** through the synthesis of two analogues bearing *N*-methylamino
13 (**18**) and *N,N*-diethylamino (**19**) substituents (Scheme 4). The role of this secondary amine was further probed
14 through the removal of its hydrogen-bond donor property via isosteric replacement with an ether linkage (**20**)
15 (Scheme 4). This would permit the assessment and influence of hydrogen bond donor removal and any
16 subsequent functional effects on the allosteric pharmacology of **1**. The *N*-methylamino derivative (**18**) would
17 test the importance of aromaticity whilst still maintaining hydrogen-bond characteristics of **1**, while the *N,N*-
18 diethylamino analogue (**19**) investigates the impact of removing aromaticity and hydrogen-bond donor
19 properties at the 4-position. Additionally, the ether derivative (**20**) assesses the importance of removing
20 hydrogen-bond donor properties while maintaining aromaticity. We also focused on the 2-position of **1**, where
21 we sought to evaluate the impact of removing the 2-morpholinomethyl moiety entirely (**23a**, Scheme 5),
22 subsequently conferring no substitution at the 2-position of the thienopyrimidine. In addition to this, we
23 investigated isosteric replacement of the morpholino O for CH₂ to afford the piperidine analog **26** (Scheme 6),
24 as well as a similar replacement of N for CH to furnish the tetrahydropyran analog **33a** (Scheme 7). Moreover,
25 we investigated the effects of increasing the methylene linker length bearing morpholine (ethylene, **39a**;
26 propylene, **39b**; butylene, **39c**) (Scheme 8).

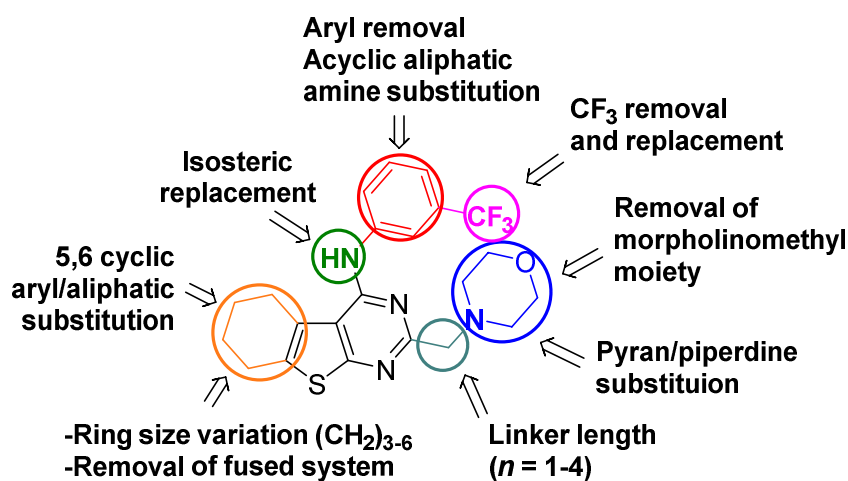


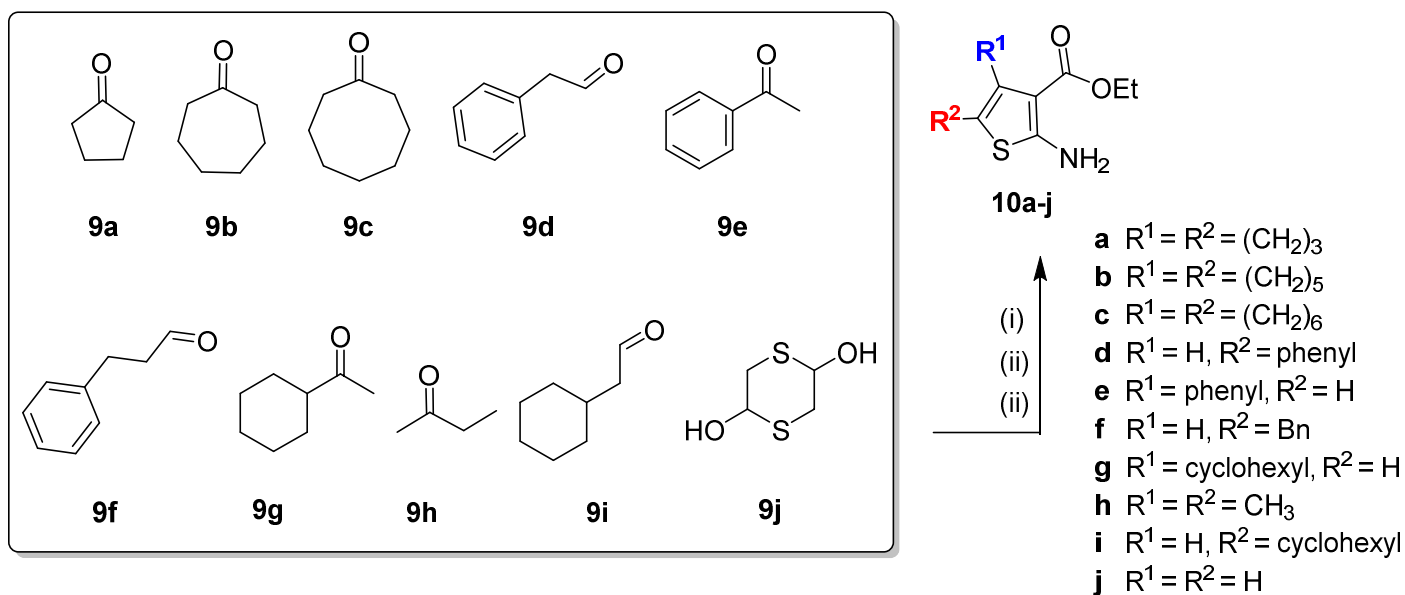
Figure 2. Areas of structural modification of **1** conducted during initial SAR investigation.

Hybrid Modification of 1. Several additional compounds were synthesised to further investigate the effects of the aforementioned structural modifications. These modifications included deletion of the fused cyclohexane system of the tricyclic scaffold, removal of the 2-morpholinomethyl pendant, isosteric replacement through substitution of the morpholine ring for tetrahydropyran, and the incorporation of a 1,4-butylene linker; all bearing the *N,N*-diethylamino functionality in place of the aryl *m*-CF₃ moiety. These subsequent analogues (**23b**, Scheme 5), (**33b**, Scheme 7), (**40**, Scheme 9), (**44**, Scheme 11) were synthesised as outlined previously by varying the nature of the nucleophile used in the final nucleophilic aromatic substitution. Finally, a further four analogues (**43a-b**, Scheme 10; **47a-b**, Scheme 11), were synthesised to examine the effects of simultaneous deletion/installment of functionality with respect to maintaining one of either *m*-CF₃-aryl or *N,N*-diethylamino substituents at the 4-position of **1**.

The thieno[2,3-*d*]pyrimidine moiety (Figure 2) is prevalent in a variety of reported compounds, thus its synthesis is well established.¹⁷⁻²² Rapid access to a series of analogues substituted at the 5- and 6-positions was achieved using the appropriate aldehyde or ketone as a starting material, by means of Gewald-type chemistry (Scheme 2). An alternative method to traditional Gewald chemistry using solvent-free conditions for the preparation of substituted 2-aminothiophene derivatives from respective ketones/aldehydes in the presence of morpholine as catalyst was used to furnish a parallel set of analogues (Scheme 2).²³ The commercial availability of aldehydes (**9d**, **9f**, **9i**) or ketones (**9a-c**, **9e**, **9g**, **9h**) and their usefulness in Gewald-type chemistries or variations thereof permitted easy access to a variety of substituted thienocarboxylate

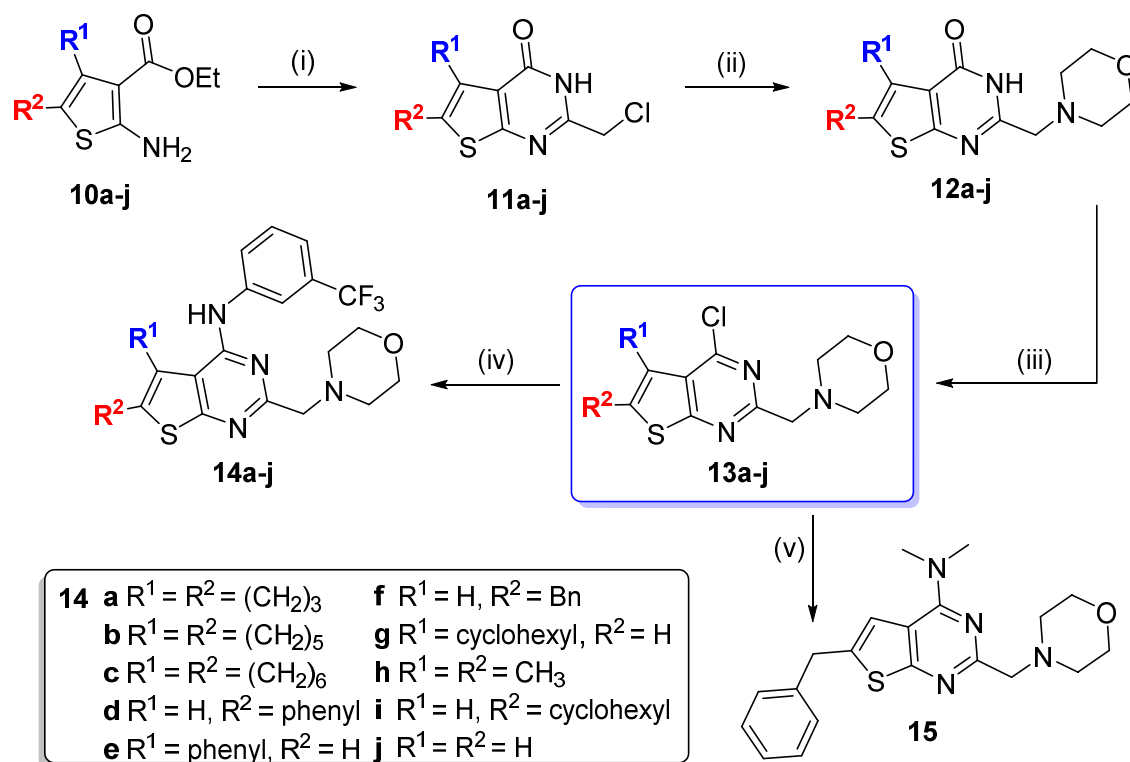
precursors **10a-i** bearing a primary amine and ethyl ester as functional handles. Ethyl 2-aminothiophene-3-carboxylate **10j**, was obtained in accordance with an adapted procedure²⁴ (Scheme 2) using the commercially available mercaptoacetaldehyde dimer, 1,4-dithiane-2,5-diol (**9j**). This reagent was successfully used to afford **10j** as a yellow crystalline solid in good yield.

Scheme 2. Chemical Synthesis of 2-Amino-3-Carboethoxythiophenes^a



^aReagents and conditions: (i) sulfur, abs. EtOH, ethyl cyanoacetate, 60 °C, 3-4 h, 50-80% (**10a-c**); (ii) sulfur, morpholine (neat), ethyl cyanoacetate, r.t., ~18 h, 45-80% (**10d-i**); (iii) Et₃N, ethyl cyanoacetate, DMF, 45 °C, 0.5 h, 55% (**10j**).

To allow construction of the 2-chloromethyl-substituted pyrimidine bearing a functional handle for further elaboration, the appropriate thiophenes **10a-j** were subjected to an intermolecular cyclisation reaction under acidic conditions in the presence of chloroacetonitrile to afford the desired thieno[2,3-*d*]pyrimidine core which could be utilised for further derivatisation (Scheme 3). Cyclisation required dry HCl gas to be generated in situ (Kipps' apparatus) and to be passed through a solution containing the respective nitrile and thiophene, furnishing the corresponding 2-haloalkyl pyrimidinones **11a-j** in high yields.¹⁶

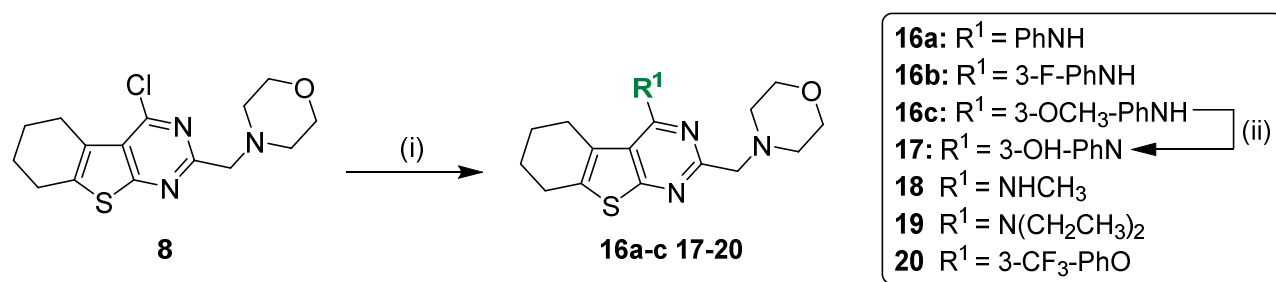
Scheme 3. Chemical Synthesis of Analogues of 1 with Modifications to Fused Cyclohexane System^a

^aReagents and conditions: (i) thiophene, chloroacetonitrile, $HCl_{(g)}$, 50 °C – reflux, 4-24 h, 20-95%; (ii) DMF, Et_3N , 100°C, 4-8 h, 50-90 %; (iii) $POCl_3$, $DMF_{(cat.)}$, toluene, 3-5 h, 70-90%; (iv) 3-(trifluoromethyl)-aniline, Et_3N , *i*-PrOH, MWI, 1-3 h, 60-90%; (v) **15**, DMF, 100°C, 33%.

N-Alkylation of morpholine with **11a-j** was conducted in DMF and typically proceeded smoothly to provide the corresponding morpholinomethyl analogues **12a-j**. Products could often be precipitated with H_2O , washed with an appropriate solvent and/or recrystallised. Carbonyl activation was achieved through the use of $POCl_3$ along with catalytic DMF in toluene to attain clean conversion to the corresponding chloropyrimidines (**13a-j**). Finally, the appropriate chloropyrimidine (**13a-j**) could be taken up in *i*-PrOH and subjected to microwave-assisted nucleophilic aromatic substitution in the presence of the appropriate nucleophile (primary aromatic or secondary acyclic aliphatic amine) and an excess of Hünigs base to furnish target analogues **14a-j** in good yields. Initially, DMF was trialed as a potential reaction solvent for nucleophilic aromatic substitution, however produced sub-optimal yields and by-products, and thus was not used further. Compound **15** was isolated as a reaction by-product during the attempted synthesis of **14f**, due to the presence of *N,N*-dimethylamine existing as an impurity in DMF, thus it was characterised and tested for its *in vitro* activity.

Substituents at the thienopyrimidine 4-position. Compounds **16a-c** were similarly obtained as previously outlined, through the use of various substituted anilines via nucleophilic aromatic substitution with key chloropyrimidine intermediate **8** (Scheme 4). Compound **16c** was used to access the phenolic analogue via *O*-demethylation employing boron tribromide in DCM at 0 °C to afford the corresponding phenol **17** in good yield. To assess the impact of removing aromaticity from the 4-position, several additional analogues of **1** were synthesised (Scheme 4). To do this, key intermediate **8** was again subjected to nucleophilic aromatic substitution using an excess of *N*-methyl or *N,N*-diethylamine under basic conditions in *i*-PrOH which furnished the corresponding *N*-methylamino and *N,N*-diethylamino analogues in good yields as light brown oils (**18** & **19**, respectively). To advance our understanding of the aniline's role within **1**, we generated a compound deficient of a hydrogen-bond donor group through isosteric replacement with an ether linkage but still bearing the aryl-CF₃ substituent (**20**, Scheme 4). To achieve this compound **8** was subjected to MWI in the presence of 3-(trifluoromethyl)phenol and a 10% molar equivalent of phase-transfer catalyst tris[2-(2-methoxyethoxy)ethyl]amine (TRIDENT). Subsequent chromatographic purification afforded the target ether-derivative **20** as a transparent oil in 72% yield.

Scheme 4. Synthesis of Analogues Modified at the 4-Position^a

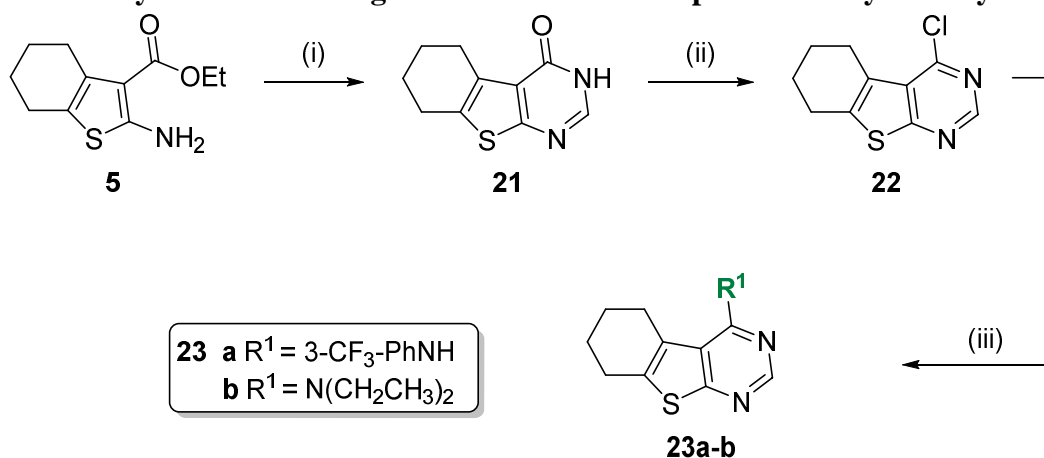


^aReagents and conditions: (i) substituted aniline, *i*PrOH, MWI, 2 h, 65-90% (**16a-c**); (ii) BBr₃ (1 M in DCM), r.t., overnight, 53% (**17**); (iii) *i*-PrOH, methylamine, MWI, 140 °C, 2.5 h, 85% (**18**); *i*-PrOH, *N,N*-diethylamine, MWI, 140 °C, 3 h, 91% (**19**); *i*-PrOH, 3-(trifluoromethyl)phenol, TRIDENT_(cat.), MWI, 6 h, 72% (**20**).

Substituents at the thienopyrimidine 2-position. To assess the effects of removing the 2-morpholinomethyl moiety (Scheme 5), 4,5,6,7-tetrahydrobenzothiophene **5** was refluxed for several hours in

neat formamide to construct the corresponding pyrimidinone **21**. This compound was similarly converted to 4-chloro analogue **22**, followed by nucleophilic aromatic substitution with 3-(trifluoromethyl)aniline to yield compound **23a** (Scheme 5). To further assess the effect of introducing an *N,N*-diethylamino substituent at the 4-position of **1**, we synthesised an additional analogue that incorporated this amine in conjunction with removal of the 2-morpholinomethyl substituent (**23b**, Scheme 5). This compound was accessed using key intermediate **22** via nucleophilic aromatic substitution thereby permitting us to further probe the effect of incorporating an aliphatic 4-substituent but still bearing similar hydrogen-bonding capabilities.

Scheme 5. Chemical Synthesis of Analogues of 1 devoid of Morpholinomethyl Moiety^a

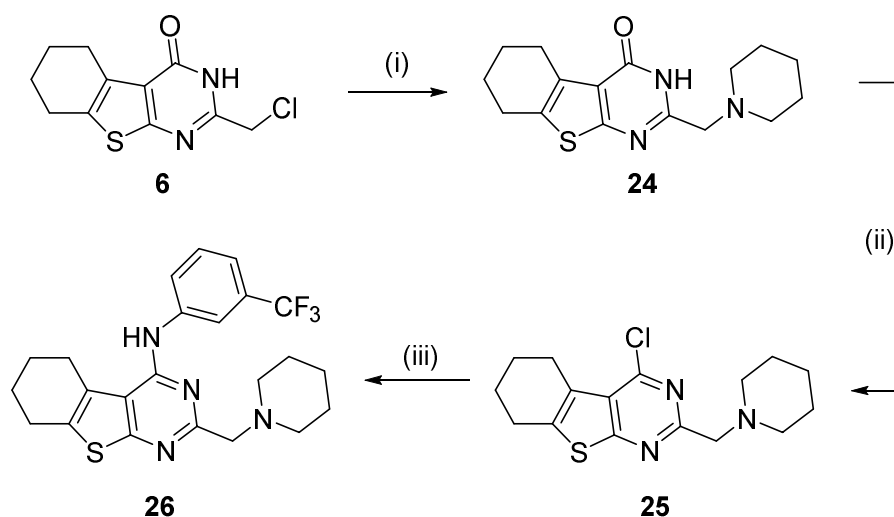


^aReagents and conditions: (i) formamide, 170 °C, 12 h, 85%; (ii) POCl₃, DMF_(cat.), toluene, 100 °C, 3-5 h, 90%; (iii) *i*-PrOH, *m*-CF₃-aniline, DIPEA, MWI, 2 h, 75% (**23a**); (iv) *i*-PrOH, Et₂NH, DIPEA, MWI, 2 h, 90% (**23b**).

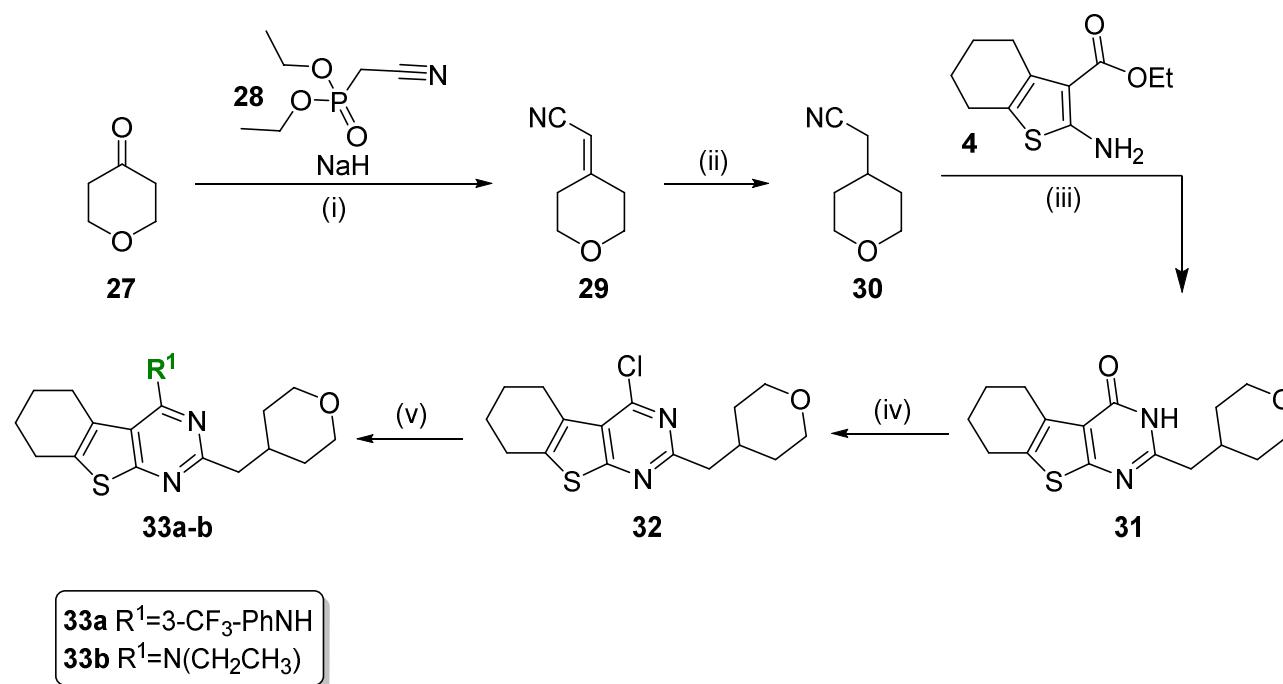
Next, we focused on the morpholine moiety to probe the effect on the allosteric pharmacology of **1** by replacing the O or N heteroatoms with a methylene or methine group to give piperidine analogue, **26** and tetrahydropyran analogue, **33a**, respectively. Compound **26** (Scheme 6) was accessed via the *N*-alkylation of piperidine with key intermediate **6**, again followed by chlorination (**25**) and substitution to afford **26** in good yield. Compound **33a** (Scheme 7) was furnished via a multi-step synthesis that initially involved activation of diethyl cyanomethylphosphonate **28** with NaH, followed by drop-wise addition of tetrahydro-4*H*-pyran-4-one **27** in diethyl ether. Vacuum distillation and recrystallisation from petroleum-ether afforded the corresponding nitrile **29** as a transparent crystalline solid in moderate yield. Catalytic hydrogenation employing Pd/C as catalyst in EtOAc and AcOH under atmospheric pressure gave the corresponding saturated nitrile **30** as a

transparent oil in high purity. Intermolecular cyclisation of thiophene **5** with **30** proceeded in 1,4-dioxane under standard acidic conditions (Kipp's apparatus) to afford the thienopyrimidinone **31**, though requiring reflux temperatures to achieve full conversion. Subsequent DMF-catalysed POCl₃ activation in toluene provided the chloropyrimidine **32**, followed by nucleophilic aromatic substitution in *i*-PrOH to afford the target tetrahydropyran **33a** as a white foam in moderate yield. To better assess the effects of *N,N*-diethylamino substitution of the 4-position in combination with the tetrahydropyran functionality, one additional analogue was synthesised (**33b**). This compound was readily accessed by reacting the appropriate starting material **32** under standard conditions to afford the target as a light gold oil in good yield (Scheme 7).

Scheme 6. Chemical Synthesis of Piperidine Analogue^a



^aReagents and conditions: (i) piperidine, Et₃N, 100 °C, 2 h, 71%; (ii) toluene, 100 °C 4 h, 46%; (iii) *i*-PrOH, *m*-trifluoromethylaniline, MWI, 2 h, 70%.

Scheme 7. Chemical Synthesis of Tetrahydro-2H-pyran-4-yl Analogues of **1**^a

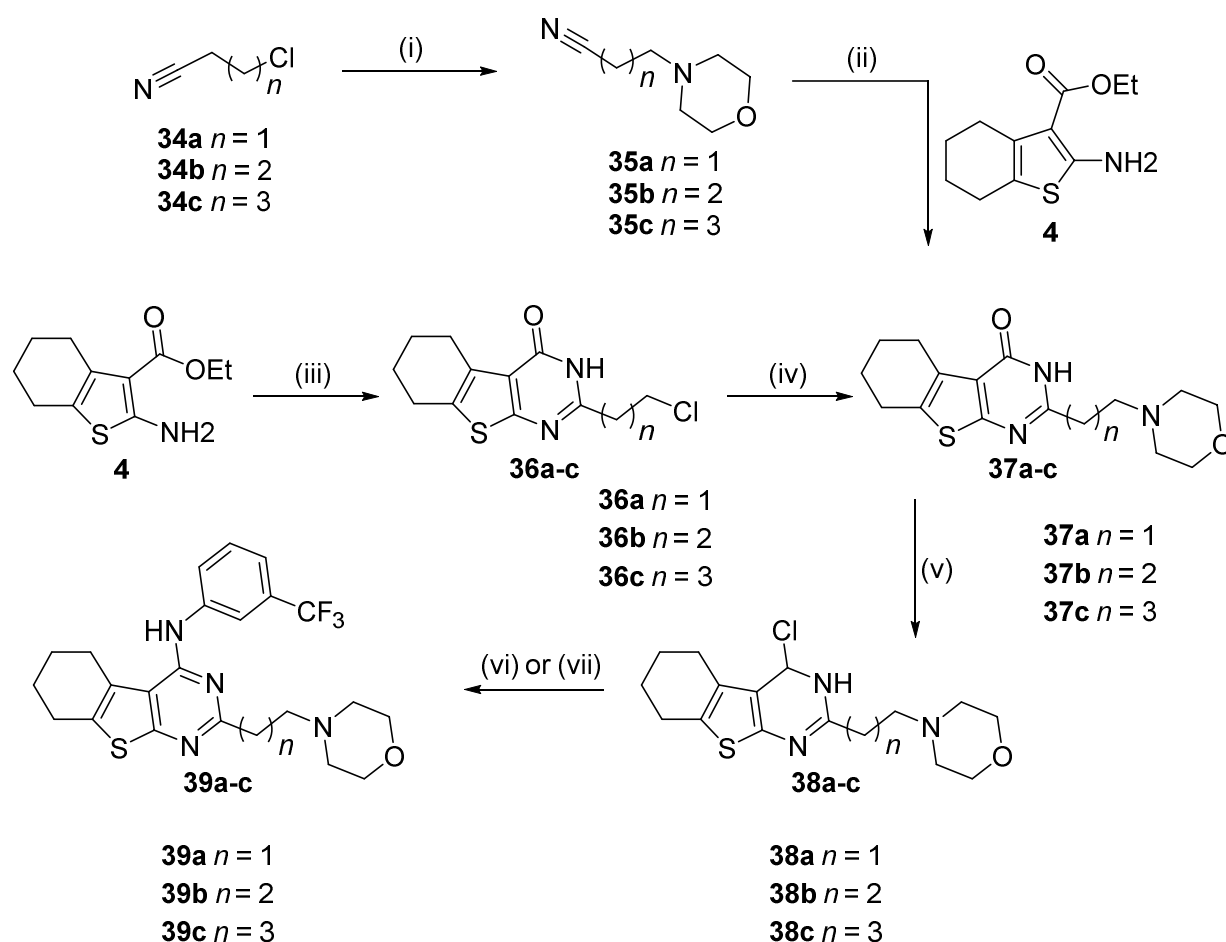
^aReagents and conditions: (i) Et₂O, 0 °C - r.t. 24 h, 39%; (ii) 10% Pd/C, EtOAc, AcOH, 1 atm, r.t. 5 h, 100%; (iii) 1,4-dioxane, HCl_(g), r.t. – reflux, 13 h, 93%; (iv) toluene, POCl₃, DMF_(cat.), 100 °C, 4 h, 24%; (v) amine, *i*-PrOH, MWI 120 °C, 3 h, 48% (**33a**); (vi) *i*-PrOH, Et₂NH, MWI, 120 °C, 2 h, 85% (**33b**).

Chain-extension of the 2-substituent linker of 1. To investigate the effects of increasing the methylene linker length on the allosteric pharmacology of **1**, we generated a series of analogues of differing methylene chain length (2-4) bridging the morpholine substituent to the pyrimidine core (Scheme 8). Commercially available primary chloroalkyl nitriles: 3-chloropropionitrile **34a**, 4-chlorobutanenitrile **34b** and 5-chloropentanenitrile **34c**, were reacted in the presence of thiophene **5** under standard acidic conditions in order to furnish the corresponding chain-extended pyrimidinones (**36a-c**). These nitriles were substantially less reactive than chloroacetonitrile, and thus required reflux temperatures to complete the conversion of the amidinium species formed *in situ* to the corresponding 2-chloro-pyrimidinone.

Using standard conditions, alkylation of morpholine with the appropriate chain extended chloro-pyrimidinone was attempted to generate the corresponding morpholinoethyl, morpholinopropyl, and morpholinobutyl substituted intermediates **37a-c**. This methodology successfully afforded the morpholinoethyl pyrimidinone **37a** in good yield as a yellow crystalline solid (Scheme 8), however pyrimidinone derivatives bearing the longer chloroalkyl chains were suspected to be eliminating under basic conditions. ¹H NMR analysis provided

confirmation of favoured intramolecular cyclisation under both ambient and reflux conditions, generating two tetracyclic regioisomers in a 1:1 ratio.²⁵ In light of this, additional efforts focused on construction of the *N*-substituted nitrile moiety first (Scheme 8). This was accomplished by stirring the appropriate nitrile (**34a-c**) with an excess of morpholine in refluxing toluene for several hours to furnish the corresponding *N*-morpholino nitriles (**35a-c**) in good yields as transparent oils. These intermediates were reacted with **5** under acidic conditions to afford the target morpholinopropyl and morpholinobutyl pyrimidinones in good yields (**37b-c**) followed by chlorination (**38a-c**) and subsequent nucleophilic aromatic substitution to furnish the final chain extended analogues **39a-c**.

Scheme 8. Synthesis of Morpholino-Substituted Nitriles and Corresponding Alkyl Chain-Extended Analogues of **1^a**

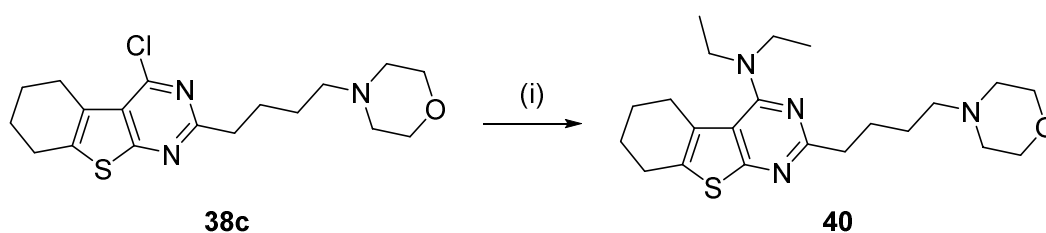


^aReagents and conditions: (i) morpholine, toluene, reflux, 7 h, 45-65% (**35a-c**); (ii) 1,4-dioxane, nitrile, HCl(g), r.t. 12 h, reflux 4 h, 57-80% (**37b-c**); (iii) **34a-c**, 1,4-dioxane, HCl(g), r.t. 12 h – reflux 3 h, 60-83% (**36a-**

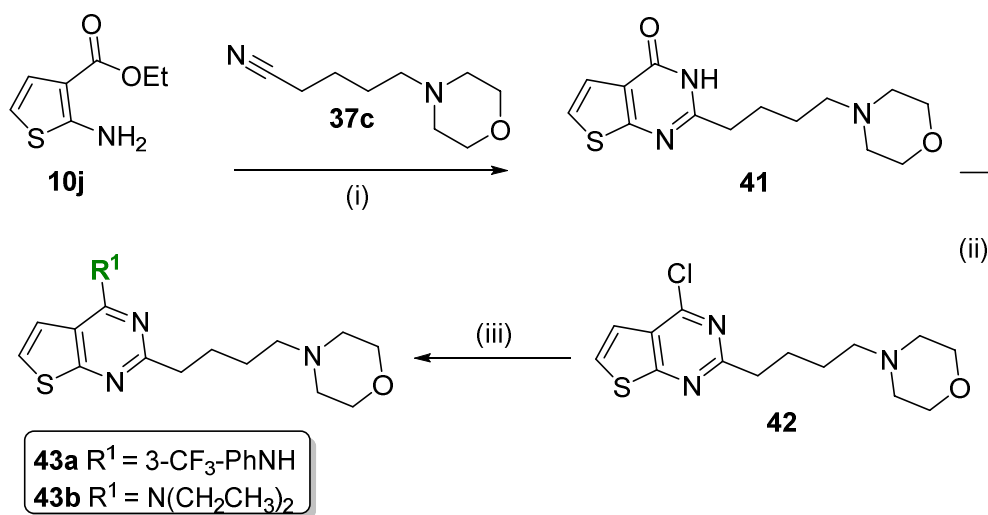
c); (iv) DMF, 50 °C, 4 h, 56% (**37a**); (v) POCl₃, toluene, DMF_(cat.), 2-5 h, 67-88% (**38a-c**); (vi) NaH, 3-(trifluoromethyl)aniline, DMF, 100 °C, 6 h, 27% (**39a**); (vii) *i*-PrOH, MWI, 2-4 h, 56-59% (**39b-c**).

Hybrid Functionalisation of 2- and 4-positions of 1. The subsequent pharmacological effects of incorporating extended morpholinoalkyl substituents into analogues of **1** (**39a-c**) conveyed interesting changes in the functional properties of this scaffold. The 1,4-butylene linker was of particular interest. Thus, compound **40** was synthesised using intermediate **38c** (Scheme 9) via nucleophilic aromatic substitution to enhance our understanding of the SAR surrounding the 4-position of **1** with respect to *N,N*-dimethylamino substitution. In addition to further investigate the 1,4-butylene linker, removal of the fused functionality within **1** (**14j**) resulted in a loss of not only binding affinity, but rendered this analogue functionally inactive. To this end, we sought to investigate the effect of removing the fused cyclohexane system whilst maintaining the morpholinobutyl functionality in conjunction with one of either 3(trifluoromethyl)anilino or *N,N*-diethylamino substituents at the 4-position (Scheme 10). These two additional analogues were accessed by reacting the appropriate unsubstituted thiophene **10j** with **35c** under acidic conditions to provide cyclised product **41**, followed by chlorination **42** and nucleophilic aromatic substitution employing conditions as outlined previously to furnish **43a-b** in good yields (Scheme 10).

Scheme 9. Synthesis of *N,N*-Diethylamino Analogue Bearing Morpholinobutyl Substituent & Fused Cyclohexane Moiety^a

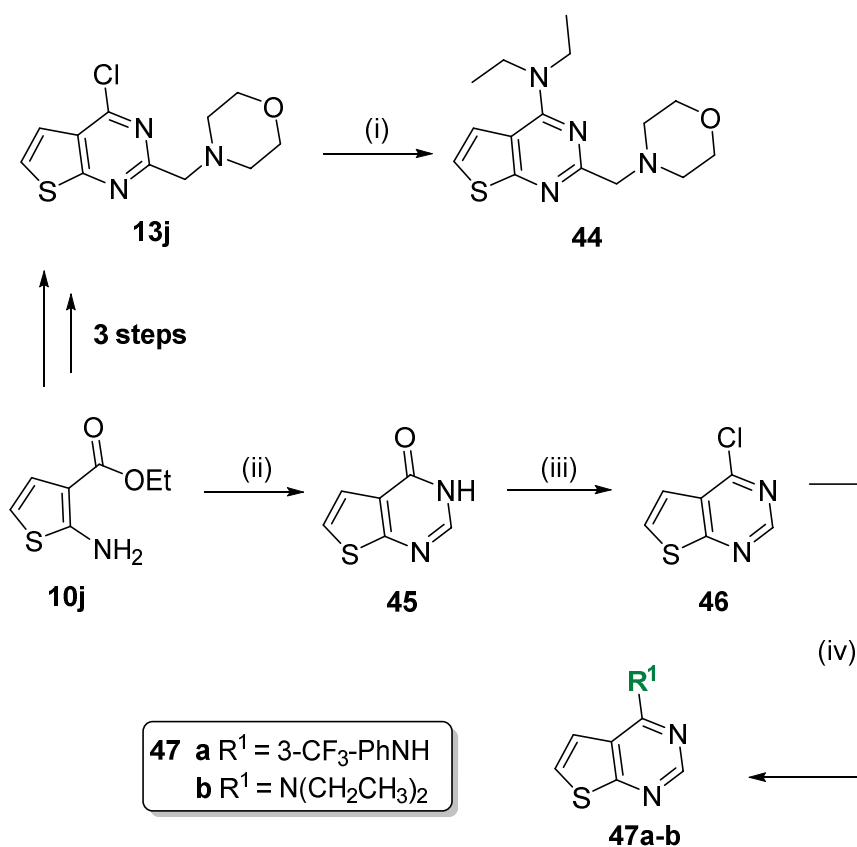


^aReagents and conditions: (i) *i*-PrOH, Et₂NH, MWI, 120 °C, 3 h, 85%

Scheme 10. Further Analogues of **1** Bearing the Morpholinobutyl Substituent^a

^aReagents and conditions: (i) 1,4-dioxane, HCl_(g), r.t overnight, reflux 3-6 h, 57-65%; (ii) toluene, POCl₃, DMF_(cat.), 3 h, 60-88% (iii) *i*-PrOH, Et₂NH, MWI, 120 °C, 1.5 h, 80% (**43a**); *i*-PrOH, Et₂NH, MWI, 120 °C, 4 h, 90% (**43b**).

In addition to this, we wanted to observe the effects of *N,N*-diethylamino substitution of **14j** as a direct comparison with 3-(trifluoromethyl)aniline. Thus, **13j**, whose synthesis is depicted in Scheme 3, was reacted with *N,N*-diethylamine under microwave irradiation to afford **44** in good yield (Scheme 11). Moreover, we wanted to observe the effect of aromatic and aliphatic substituents at the 4-position of **1** in conjunction with concurrent removal of the fused cyclohexane and morpholinomethyl moieties. Removal of said functionalities on the structure of **1** effectively results in a low molecular weight fragment-like core. These compounds were readily accessed using chemistry as outlined previously (Scheme 11) to furnish the target analogues **47a-b** in high yields.

Scheme 11. Further Thieno[2,3-*d*]pyrimidin-4-amine Analogues of **1**^a

^aReagents and conditions: (i) Et₂NH, DIPEA, *i*-PrOH, MWI, 120 °C 1.5 h, 94% (**44**); (ii) formamide, 170 °C, 6 h, 76%; (iii) toluene, POCl₃, DMF_(cat.), 3 h, 88%; (iv) *m*-CF₃-aniline, DIPEA, *i*-PrOH, MWI, 120 °C 2.5 h, 80% (**47a**); Et₂NH, DIPEA, *i*-PrOH, MWI, 120 °C 1.5 h, 90% (**47b**).

Pharmacology. Confirmation of an allosteric mode of action for **1** at the D₂R. Our previous study predicted that **1** can adopt an allosteric pose within the D₃R^{APO} model and revealed that this compound also displayed activity at the D₂R, our primary target of interest.⁸ Prior to initiating an investigation into **1**, we wanted to confirm its activity at the D₂R, and determine if it displays competitive or allosteric pharmacology. Equilibrium radioligand binding experiments were performed on **1** to monitor its effect upon orthosteric ligand ([³H]raclopride) binding in membranes prepared from FlpIn CHO cells stably expressing the long isoform of the human D₂R (hD_{2L}R).²⁶ This assay may also provide indication of an allosteric versus competitive mode of action. If the compound displays incomplete or partial displacement of the radioligand this is consistent with the saturable effect of an allosteric modulator whereby the degree of negative cooperativity with the radioligand determines the level of radioligand displacement. In cases where one observes only partial

1 displacement, data may be fit using an allosteric ternary complex model to derive a value of affinity, denoted
2 as K_B , and cooperativity with [^3H]raclopride (α).²⁷ However, in our assay, **1** was able to completely displace
3 [^3H]raclopride. Therefore, this assay did not allow us to discriminate between high negative cooperativity with
4 [^3H]raclopride. Thus, to determine evidence of an
5 the [^3H]raclopride or competition for the orthosteric binding site (Table 1). Thus, to determine evidence of an
6 allosteric mode of action for this ligand, we extended our evaluation to measurements of [^3H]raclopride
7 dissociation from the D₂R (Figure 3A). Dissociation of the radioligand was measured following addition of a
8 high concentration of cold orthosteric ligand (10 μM haloperidol) to prevent [^3H]raclopride rebinding in the
9 absence or presence of **1**. In the presence of 100 μM of **1**, the dissociation rate of [^3H]raclopride from the D₂R
10 significantly increased, (Table 2 and Figure 3A) consistent with **1** binding to an allosteric site within the D₂R.²⁸

21 Our pharmacological analysis using the above radioligand binding assay did not provide information
22 regarding the modulatory effect of **1** upon the neurotransmitter dopamine. Therefore, we tested **1** in an assay
23 measuring inhibition of forskolin-stimulated cyclic adenosine monophosphate (cAMP) accumulation through
24 activation of the hD_{2L}R stably expressed in FlpIn CHO cells using a BRET biosensor.²⁹ The concentration-
25 dependent response of dopamine was measured in the presence of increasing concentrations of **1** and 10 μM
26 forskolin. Application of an operational model of allosterism to the concentration-response data yielded an
27 estimate of affinity of **1** for the unoccupied receptor (K_B), and its cooperativity with dopamine where α is
28 cooperativity exerted upon dopamine binding, and β denotes modulation of dopamine efficacy.³⁰ These data
29 are reported in Table 4 and are presented as logarithms to base 10 to allow statistical comparison.³¹ Values of
30 α or $\beta < 1$ signify negative cooperativity with dopamine. The lead compound **1** acted as a negative allosteric
31 modulator at the D₂R, with low μM functional affinity ($K_B = 3.9 \mu\text{M}$). The depression in the maximal response
32 upon the dopamine dose-response curve caused by increasing concentrations of **1** is characteristic of the action
33 of a NAM of agonist efficacy (Figure 3B). In order to fit these data, the cooperativity of **1** with dopamine
34 affinity (α) was constrained to be neutral, allowing us to estimate a value of modulatory action upon dopamine
35 efficacy ($\beta = 0.28$) that equates to a maximal 3-fold decrease (Figure 3B, Table 4). This action is different to
36 the action of the competitive antagonist haloperidol at the D₂R that acts to cause a dextral shift in the dopamine
37 dose-response curve with no affect upon the maximal effect of dopamine (Figure 3C). Thus, by using
38 complementary pharmacological approaches including equilibrium and dissociation kinetic binding analyses,

together with experiments measuring D₂ receptor function, we have confirmed that **1** acts through a non-competitive mechanism to modulate the activity of dopamine at the D₂R i.e. it is NAM of the efficacy of dopamine. The thienopyrimidine scaffold of **1** is unique to known dopaminergic receptor ligands. Therefore, a novel allosteric scaffold for the D₂R has been identified and validated. Moreover, further validation has been provided for virtual allosteric ligand screening at the dopamine D₂-like receptors.

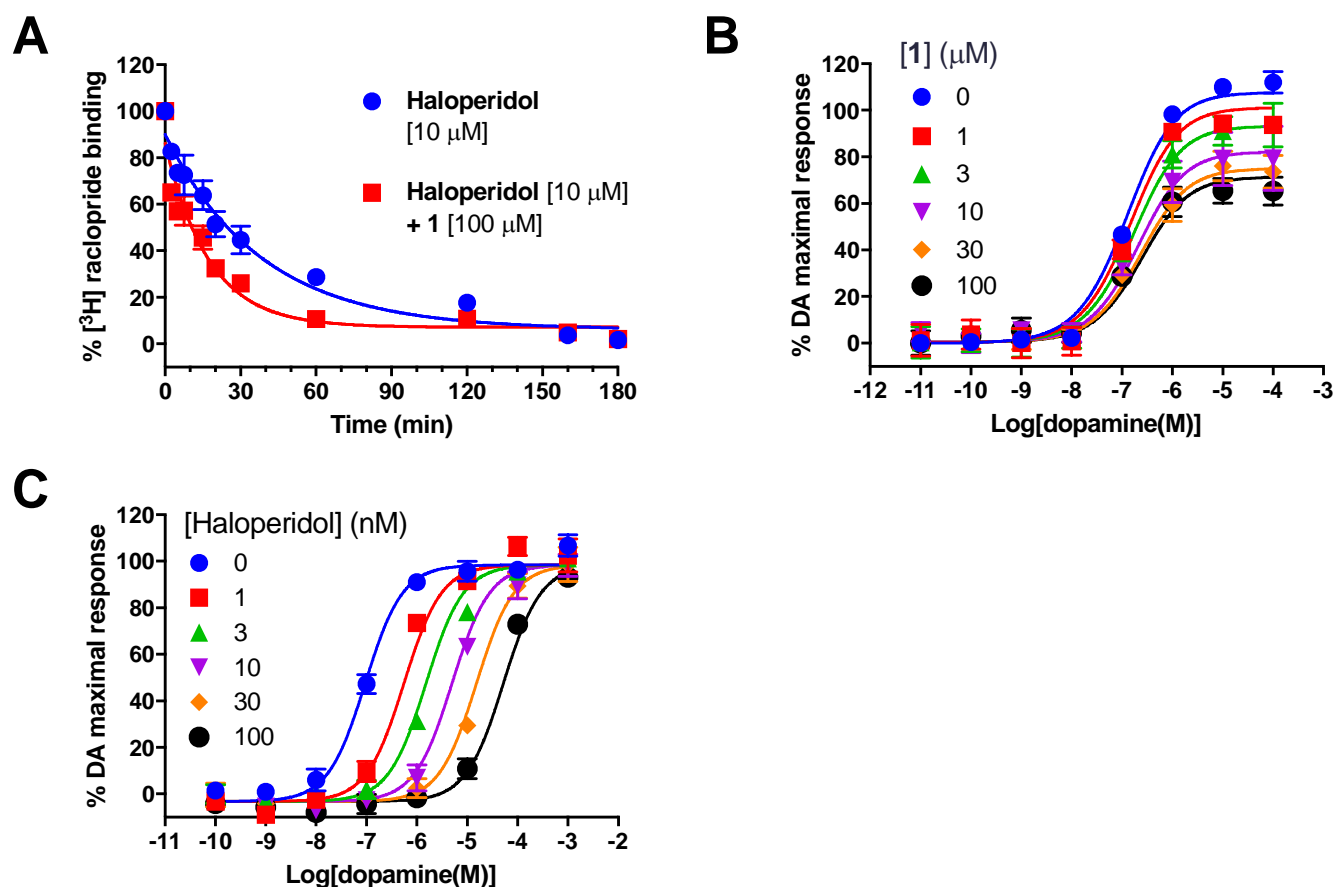


Figure 3. Compound 1 displays allosteric pharmacology at the D₂R. (A) **1** acts to increase the rate of dissociation of [³H]raclopride from the hD₂L_R expressed in FlpIN CHO membranes consistent with an allosteric mode of action (Table 2). (B) In an assay measuring inhibition of forskolin-stimulated cAMP production using a BRET biosensor, **1** acts to cause limited depression in the maximal response of dopamine with more modest effects upon dopamine potency (these data were fitted with an operational model of allosterism to derive values of affinity and cooperativity, Table 4). In contrast increasing concentrations of haloperidol (C) causes a limitless dextral shift in dopamine potency with no effect on dopamine maximal response

consistent with competitive antagonism and can be fit with a Gaddum-Schild model to derive a value of affinity $pA_2 = 9.67 \pm 0.10$. Data is the mean \pm SEM of three independent experiments performed in duplicate.

Radioligand binding analysis of analogues of 1: To begin to explore the structural features of **1** that determine its allosteric activity, we employed the equilibrium [³H]raclopride binding assay described in our characterisation of **1** above. For the majority of compounds, we observed complete displacement of the radioligand. Therefore, while this simple assay provides us with both confirmation of target activity and a value of affinity, for these compounds, we cannot distinguish between a competitive mode of interaction or an allosteric interaction that exerts high negative cooperativity upon [³H]raclopride binding. We first examined modifications to the fused cyclohexane system of **1** (Table 1). Replacement of the 6-membered saturated ring with a fused cyclopentane (**14a**) or cycloheptane (**14b**) system made no significant impact on binding affinity. However, replacement with the cyclooctane (**14c**) system caused a significant 6-fold decrease in affinity. No significant effect on affinity was seen when various other substituents were introduced to the 5- and 6-positions of the thienopyrimidinone (**14d-g, i**). Conversely, introduction of the 5-phenyl substituent (**14e**) did result in an apparent decrease in negative cooperativity manifested by an incomplete displacement of [³H]raclopride that allowed us to determine a value of cooperativity, $\alpha = 0.13$ (Table 1). 5,6-Dimethyl substitution (**14h**), as well as removal of the fused system (**14j**) both caused a significant 5-10-fold decrease in affinity ($K_B = 250$ and $120 \mu\text{M}$, respectively). To further explore the role of the *N*-(3-(trifluoromethyl)phenyl) substituent of **1**, we generated a small series of analogues of **1** that incorporated changes to this functionality, including removal of the CF₃ substituent (**16a**), as well as substitution with various groups (3-F, **16b**), (3-OCH₃, **16c**), (3-OH, **17**). Removal of the CF₃ substituent caused a 6-fold decrease in affinity whereas both the 3-OH and 3-OMe analogues displayed similar affinity to the parent compound (**1**). To investigate the effects of removing the *N*-(3-trifluoromethyl)phenyl substituent at the 4-position, we synthesised a series of analogues of **1** that included an *N,N*-dimethylamino group (**15**) that was isolated as a by-product from the synthesis of **14f** as a result of trialing DMF as a reaction solvent, as well as analogues bearing *N*-methylamino (**18**), and *N,N*-diethylamino (**19**) moieties. In addition to this, we investigated the effect of removing the secondary amine bridging the aryl moiety to the pyrimidine moiety of **1** by isosteric

1 replacement with an ether linkage (**20**). These analogues all saw no significant change in affinity as compared
2 to **1**. Interestingly, however, **19** displayed only partial displacement of [³H]raclopride at the D₂R ($K_B = 6.0$
3 μM , $\alpha = 0.23$) (Table 1, Figure 4A). These data demonstrate that the secondary amine plays no role in the
4 functional affinity of **1**, but modification of this feature can modulate the negative cooperativity with
5 [³H]raclopride. Additional analogues were synthesised to examine the effect of increasing the methylene
6 linker length bearing the morpholine substituent (**39a-c**), as well as the effect of replacing either the oxygen
7 or nitrogen atoms within morpholine with CH₂ and CH (**26** & **33a**, respectively, Table 1). Of note, the addition
8 of a morpholinobutyl substituent (**39c**) not only appeared to increase affinity 5-fold but also to cause an
9 apparent decrease in negative cooperativity such that this compound was only able to partially inhibit the
10 binding of [³H]raclopride ($\alpha = 0.09$). Interestingly, while incorporation of piperidine **26** saw no changes in
11 affinity, the tetrahydropyran analogue **33a** lost affinity and apparent negative cooperativity with
12 [³H]raclopride as the highest concentration of **33a** only partially displaced the radioligand from the receptor
13 ($K_B = 100 \mu\text{M}$, $\alpha = 0.39$) (Table 1, Figure 4A). Similarly, removal of the morpholinomethyl moiety **23a** did
14 not affect binding affinity despite having effects on negative cooperativity with only partial displacement the
15 radioligand again observed ($K_B = 31 \mu\text{M}$, $\alpha = 0.39$) (Table 1, Figure 4A). In summary, the removal of the
16 morpholinomethyl moiety, or incorporation of *N,N*-diethylamino, tetrahydropyranomethyl, and
17 morpholinobutyl functionalities yielded ligands that exhibited incomplete displacement of [³H]raclopride,
18 suggesting that these functionalities were important determinants of cooperativity (compounds **23a**, **19**, **33a**,
19 and **39c**, respectively). Accordingly, analogues were synthesised integrating these changes with additional
20 structural changes (Table 1). We synthesised four analogues combining the incorporation of the *N,N*-
21 diethylamino moiety into these novel analogues of **1**, including **23b** (morpholinomethyl deletion), (**33b**)
22 (tetrahydropyranomethyl), **40** (morpholinobutyl), and (**44**) (fused cyclohexane deletion). In addition to these
23 modifications, we wanted to examine the effects of bis-functionalisation with respect to incorporating *m*-
24 trifluoromethylaniline or *N,N*-diethylamine. We synthesised two compounds devoid of the fused cyclohexane
25 system and morpholinomethyl moieties but maintained substitution with one of either *m*-
26 trifluoromethylaniline or *N,N*-diethylamine, to reveal fragment-like low molecular weight analogues (**47a** &
27 **47b** respectively). While both maintained affinity at the D₂R, the latter compound was only able to partially

1 displace [³H]raclopride and a value of $\alpha = 0.10$ could be determined reflecting moderate negative
2 cooperativity. Excitingly, our finding that **47a** and **47b**, analogues both devoid of the fused system and
3 morpholinomethyl moieties that maintain affinity, indicate that the scaffold of **1** can be ‘pruned back’ to reveal
4 modulators of the D₂R with low molecular weights and high ligand efficiencies. A further two compounds
5 were synthesised that comprised the morpholinobutyl moiety but were devoid of the fused cyclohexane
6 system, and were substituted with either *m*-trifluoromethyl-aniline or *N,N*-diethylamine (**43a** & **43b**
7 respectively). Affinity was maintained for these analogues showing that the fused cyclohexane may be
8 removed from the scaffold without detriment. It is interesting to note that **44** displayed 100-fold improved
9 affinity ($K_B = 1 \mu\text{M}$) from its parent analogue **14j** ($K_B = 120 \mu\text{M}$), demonstrating the *N,N*-diethylamino
10 substituent is favourable over the *m*-trifluoromethyl substituted aryl system. Similarly, upon introduction of
11 this amine to **33a**, compound **33b** gained a 3-fold improvement in affinity ($K_B = 35 \mu\text{M}$), but acted to
12 completely displace [³H]raclopride suggestive of higher negative cooperativity or a competitive mode of
13 interaction. This is consistent with our observation that, while **23a** partially displaced [³H]raclopride,
14 compound **23b** was able to completely displace the radioligand. In summary, then, our exploration of the
15 thienopyrimidine scaffold of **1** revealed that a vast array of modifications are tolerated in terms of affinity for
16 the D₂R but that subtle modifications to the 4-position may modulate the degree of negative cooperativity
17 between the allosteric ligand and orthosteric probe [³H]raclopride. As the degree of negative cooperativity on
18 [³H]raclopride binding significantly changed upon *N,N*-diethylamino substitution (**19**), resulting in the
19 incomplete displacement of the radioligand, we sought to obtain additional confirmation of an allosteric mode
20 of interaction. The profile of **19** was further explored through radioligand binding dissociation kinetic
21 experiments. In the presence of **19**, the dissociation rate of the [³H]raclopride from the D₂R again significantly
22 increased (Table 2). This experiment provided further validation of the allosteric mode of action of **1** and its
23 structural analogues.
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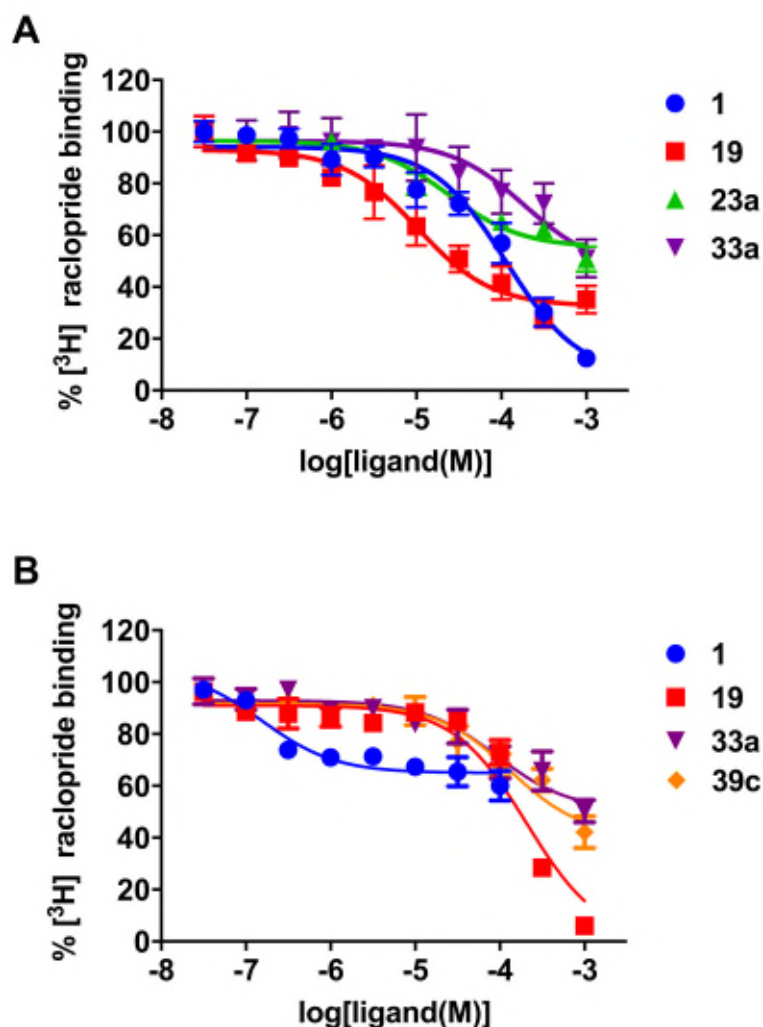
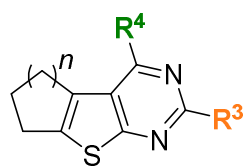


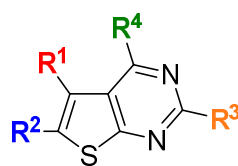
Figure 4. Analogues of 1 act to partially displace orthosteric radioligand binding at the D₂R and D₃R.

(A) In equilibrium radioligand binding experiments, **1** acts to completely displace $[^3\text{H}]$ raclopride from the hD₂L_R expressed in FlpIN CHO membranes, an effect consistent with a competitive mode of binding or an allosteric mode of binding that exerts high negative cooperativity with $[^3\text{H}]$ raclopride (**Table 1**). In contrast compounds **19**, **23a**, and **33a** act to only partially displace $[^3\text{H}]$ raclopride at the highest concentration consistent with an allosteric mode of action and moderate negative cooperativity (these data were fitted with an allosteric ternary complex model to derive estimates of affinity (K_B) and cooperativity with $[^3\text{H}]$ raclopride (α), **Table 1**). (B) **1** acts to partially displace $[^3\text{H}]$ raclopride from the hD₃R expressed in FlpIN CHO membranes at the highest concentration measured, whereas **19** completely displaces $[^3\text{H}]$ raclopride from the D₃R (**Table 3**). Data is the mean \pm SEM of three independent experiments performed in duplicate.

Table 1. Ability of 1 and derivatives of 1 to displace the radiolabelled antagonist [³H]raclopride at the D₂L_R expressed in FlpIn CHO cells.



**1, 14a-c, 16a-c, 17-20,
23a-b, 26, 33a-b, 39a-c, 40**



14d-j, 15, 43a-b, 44, 47a-b

	<i>n</i>	R ¹	R ²	R ³	R ⁴	p <i>K</i> _B (<i>K</i> _B , μM) ^a	Log α (α) ^b
1	2	-	-			4.78 ± 0.11 (16.6)	-
14a	1	-	-			4.23 ± 0.12 (59)	-
14b	3	-	-			4.43 ± 0.21 (37)	-
14c	4	-	-			3.98 ± 0.17 (106)	-
14d	-	H				4.35 ± 0.13 (45)	-
14e	-		H			4.31 ± 0.11 (48.6)	-0.88 ± 0.10 (0.13)
14f	-	H				4.30 ± 0.18 (50)	-
14g	-		H			4.42 ± 0.20 (38)	-
14h	-	CH ₃	CH ₃			3.59 ± 0.15 (250)	-
14i	-	H				4.34 ± 0.18 (45.8)	-
14j	-	H	H			3.91 ± 0.20 (120)	-
15	-	H			N(CH ₃) ₂	4.51 ± 0.14 (30)	-
16a	2	-	-			4.00 ± 0.16 (99)	-
16b	2	-	-			nd	nd
16c	2	-	-			4.32 ± 0.14 (48)	-
17	2	-	-			4.45 ± 0.10 (35)	-

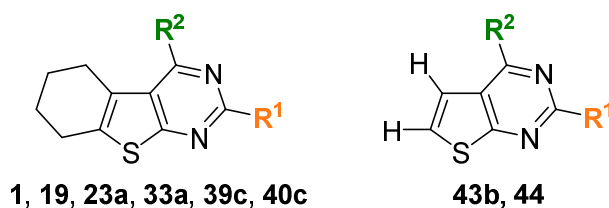
Table 2. Compounds 1 and 19 act to increase the rate of [³H]Raclopride dissociation from membrane homogenates of FlpIn CHO Cells Stably Expressing the D_{2L}R.^b

	<i>k</i> _{off} (min ⁻¹)		<i>t</i> _{1/2} (min)	
	haloperidol	haloperidol + modulator	haloperidol	haloperidol + modulator
1	^a 0.020 ± 0.004	^a 0.038 ± 0.005*	^a 34.2	^a 18.1
19	0.014 ± 0.001	0.038 ± 0.003*	49.2	18.2

Values represent the mean ± S.E.M. from at least three independent experiments performed in quadruplicate. *Analysis with a student's *t*-test revealed that [³H]raclopride displayed a significantly higher (*p* < 0.05) *k*_{off} in the presence of haloperidol plus modulator as compared to haloperidol alone.

Finally, we sought to assess receptor sub-type selectivity at the D₃R versus the D₂R; accordingly, equilibrium radioligand binding experiments were conducted on **1** and a further seven structural analogues (**19**, **23a**, **33a**, **39c**, **40c**, **43b** and **44**, Table 3). Molecules were selected based upon their profiles at the D₂R acquired from [³H]raclopride binding experiments at this receptor, where some showed increased binding affinities and divergent cooperativity profiles. Interestingly, two compounds displayed selectivity for the D₂R over the D₃R **39c** (30-fold selectivity) and **44** (10-fold selectivity). In contrast, compounds **1**, **19** and **23a** displayed selectivity towards the D₃R (57-fold, 23-fold, 140-fold, respectively). All other compounds displayed no significant subtype selectivity (**33a**, **40**, **43b**). It is interesting to note that many compounds assayed at the D₃R saw strikingly different levels of negative cooperativity with [³H]raclopride binding compared to that observed at the D₂R (Table 3, Figure 4B). For example, at the D₂R, **19** only partially displaces [³H]raclopride at the highest concentration measured, however, at the D₃R it completely displaces [³H]raclopride at the highest concentration despite having a lower functional binding affinity (*K*_B = 89.7 μM) (Table 3, Figure 4B). As **1** was identified via virtual ligand screening using a D₃R crystal structure, it is unsurprising that **1** displayed subtype selectivity for this receptor. However, our study has identified D₂R-selective compounds that may be used as a starting point for the future development of D₂R subtype-selective ligands.

Table 3. Ability of 1 and derivatives of 1 to displace the radiolabelled antagonist [³H]raclopride at the D₃R expressed in FlpIn CHO cells



	R¹	R²	^b pK_B (K _B , μM)	D₂/D₃ selectivity	^a log α (α)
1			6.54 ± 0.27 (0.3)	57	-0.40 ± 0.04 (0.39)
19		N(CH ₂ CH ₃) ₂	3.85 ± 0.16 (140)	23	-
23a	H		6.65 ± 0.21 (0.2)	140	-0.39 ± 0.03 (0.06)
33a			4.28 ± 0.23 (52)	2	-0.44 ± 0.23 (0.37)
39c			3.97 ± 0.18 (106)	0.03	-0.69 ± 0.20 (0.20)
40		N(CH ₂ CH ₃) ₂	4.99 ± 0.05 (10)	1.6	-
43b		N(CH ₂ CH ₃) ₂	6.06 ± 0.03 (0.9)	3.5	-
44		N(CH ₂ CH ₃) ₂	4.95 ± 0.06 (11)	0.1	-

^aEstimate of the negative logarithm of the equilibrium dissociation constant ± SEM determined by radioligand binding.

^bEstimate of the logarithm of the net cooperativity factor between the modulator and [³H]raclopride. Values represent the mean ± SEM from at least three independent experiments performed in duplicate.

Functional Analysis (cAMP) of analogues of 1. We extended our characterisation of these compounds to their effect upon dopamine action in the cAMP assay described above. Interestingly, all compounds above were shown to retain activity in our equilibrium radioligand binding experiments, yet many failed to have any effects upon dopamine binding and/or signalling efficacy within the same concentration range. This observation is likely an example of probe dependence whereby these ligands exert negative cooperativity on raclopride binding but display neutral cooperativity upon dopamine binding and efficacy. In agreement with this hypothesis, while our binding assay found that **1** displayed a robust effect upon [³H]raclopride binding,

1 our functional data suggests that **1** acts to modulate dopamine efficacy but has little effect upon dopamine
2 affinity.
3

4 We first examined modifications to the 5,6-fused cyclohexane system of **1** (Table 4). We found that
5 increasing the fused ring size greater than 6-carbons renders these analogues inactive in this functional assay
6 (**14b-c**), while the five-membered cyclopentane system (**14a**) maintained functional affinity ($K_B = 11.4 \mu\text{M}$)
7 but changed the nature of the cooperativity whereby we observed a modulatory effect on dopamine affinity (α
8 = 0.13) rather than modulation of efficacy ($\beta = 1$). Both 6-phenyl substitution (**14d**) and modifications to the
9 5-position with phenyl (**14e**) or cyclohexyl (**14g**) substituents resulted in a loss of activity. Activity was
10 maintained for the 6-cyclohexyl substituted analogue (**14i**, $K_B = 5.4 \mu\text{M}$, Figure 5A), but in contrast to the
11 action of **1** that modulates dopamine efficacy, this ligand acted to modulate dopamine affinity only ($\alpha = 0.13$)
12 (Table 4, Figure 5A). 5,6-Dimethyl substitution (**14h**), as well as removal of the fused functionality entirely
13 (**14j**), rendered these analogues inactive in our functional assays, thus indicating a small window in which **1**
14 can be modified surrounding the 5,6-positions and still maintain allosteric pharmacology with dopamine.
15 Removal of the CF_3 substituent on the aryl ring of **1** (**16a**, Table 5) or replacement with 3-F (**16b**), 3-OMe
16 (**16c**) or 3-OH (**17**) had no effect upon functional affinity. However, the addition of methoxy or hydroxy
17 substituents resulted in additional modulatory effects upon dopamine affinity ($\alpha = 0.19$ and $\alpha = 0.06$
18 respectively).
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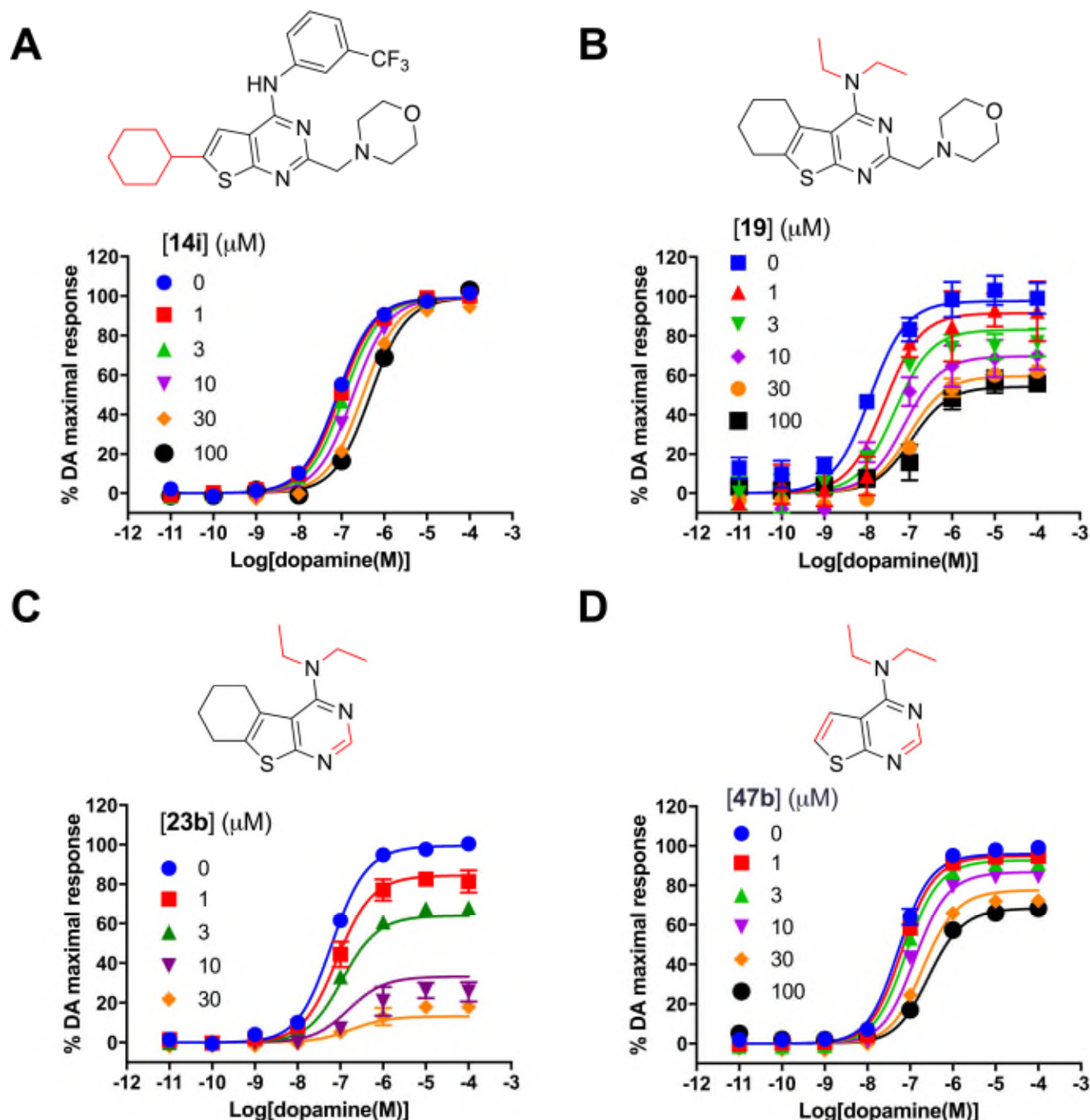
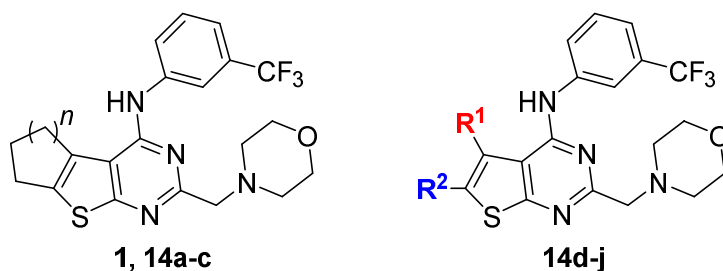


Figure 5. Exploration of the thieno[2,3-*d*]pyrimidine scaffold of 1 reveals small molecule modulators of the D_{2L}R with distinct functional effects. (A) Modification of the 5,6-fused system in the form of a 6-cyclohexane substitution (**14i**, Table 4) resulted in negative cooperativity upon dopamine affinity with no effect upon dopamine efficacy (B). Replacement of the phenyl ring of 1 with *N,N*-diethyl (**19**, Table 6) caused a 6-fold increase in affinity and 4-fold increase in negative cooperativity with dopamine. (C) Removal of the morpholinomethyl moiety along with incorporation of the *N,N*-diethylamino substitution (**23b**, Table 7) maintained affinity but resulted in a large increase in negative cooperativity with dopamine efficacy. (D) Remarkably, a fragment-like derivative devoid of both the morpholinomethyl and fused cyclohexane moieties

but incorporating the *N,N*-diethylamino substitution (**47b**, **Table 8**) maintained functional affinity and displayed robust negative cooperativity with dopamine efficacy. All data are the mean \pm SEM of three independent experiments performed in duplicate. Data were fit using the operational model of allosterism to derive estimates of affinity and cooperativity with dopamine.

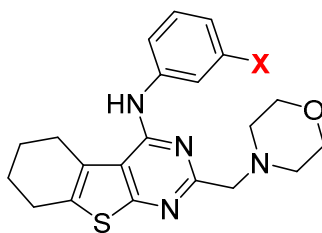
Table 4. Functional Parameters for Analogues of 1 with Chemical Modifications to the 5,6-Fused System Derived from cAMP BRET Assay



	<i>n</i>	R¹	R²	^a <i>pK_B</i> (<i>K_B</i> , μ M)	^b Log α (α)	^c Log β (β)
1	1	-	-	5.41 \pm 0.22 (3.87)	= 0	-0.55 \pm 0.08 (0.28)
14a	2	-	-	4.95 \pm 0.40 (11.4)	-0.90 \pm 0.32 (0.13)	= 0
14b	3	-	-	nd		
14c	4	-	-	nd		
14d	-	H		nd		
14e	-		H	nd		
14f	-	H		nd		
14g	-		H	nd		
14h	-	CH ₃	CH ₃	nd		
14i	-	H		5.27 \pm 0.17 (5.37)	-0.90 \pm 0.11 (0.13)	= 0
14j	-	H	H	nd		

^aEstimate of the negative logarithm of the equilibrium dissociation constant determined in an cAMP functional assay.

^bEstimate of the logarithm of the net cooperativity factor between the modulator and dopamine. ^cEstimate of the logarithm of the modulatory effect upon efficacy factor induced by the allosteric modulator. nd = inactive at concentrations up to 100 μ M. Values represent the mean \pm S.E.M. from at least three independent experiments performed in duplicate.

Table 5. Functional Parameters for Analogues of 1 with Modifications to Aryl Moiety

	X	^a $pK_B (K_B, \mu M)$	^b $\text{Log } \alpha (\alpha)$	^c $\text{Log } \beta (\beta)$
1	CF ₃	5.41 ± 0.22 (3.87)	= 0	-0.55 ± 0.08 (0.28)
16a	H	5.13 ± 0.31 (7.46)	= 0	-0.95 ± 0.24 (0.11)
16b	F	5.08 ± 0.44 (8.39)	= 0	-0.40 ± 0.11 (0.40)
16c	OMe	5.10 ± 0.13 (8.02)	-0.73 ± 0.16 (0.19)	-0.53 ± 0.14 (0.29)
17	OH	5.55 ± 0.12 (2.81)	-1.22 ± 0.16 (0.06)	-3.0

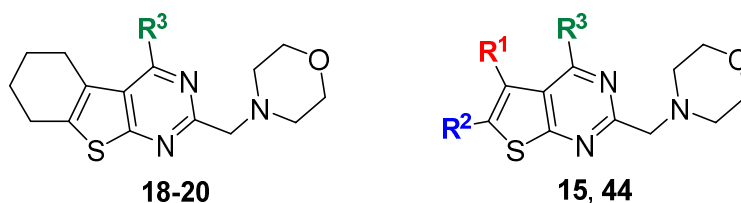
^aEstimate of the negative logarithm of the equilibrium dissociation constant determined in an cAMP functional assay.

^bEstimate of the logarithm of the net cooperativity factor between the modulator and dopamine. ^cEstimate of the logarithm of the modulatory effect upon efficacy factor induced by the allosteric modulator. nd = inactive at concentrations up to 100 μM . Values represent the mean \pm S.E.M. from at least three independent experiments performed in duplicate.

We then focused on the importance of the aryl ring within **1** by replacing the phenyl ring with acyclic aliphatic amines (Table 6). These analogues still maintained a hydrogen-bond donor-acceptor, but lacked the aromatic moiety in this region. The *N*-methylamino analogue (**18**) was inactive whereas the *N,N*-diethylamino analogue (**19**) saw a 6-fold increase in functional affinity and a 4-fold increase in negative cooperativity with dopamine efficacy ($K_B = 660$ nM, $\beta = 0.08$, Figure 5B). To further probe the effects of this substituent, we tested an additional analogue of **19** in which the fused cyclohexane system was removed entirely whilst maintaining the *N,N*-diethyl functionality (**44**). While the analogue bearing no substitution at the 5- and 6-position (**14j**) displayed no activity in this functional assay, to our surprise compound **44** maintained affinity ($K_B = 3$ μM) as well as robust negative cooperativity with dopamine affinity and efficacy ($\alpha = 0.08$, $\beta = 0.24$). Interestingly, due to the presence of the *N,N*-dimethylamino functionality, 6-benzyl substitution (**15**) no longer rendered the molecule inactive, as seen previously with **14f** (Table 4), but instead, caused affinity and negative allosteric cooperativity to be maintained (Table 6). Together, these data demonstrate that the nature of the 4-substituent (aromatic or acyclic aliphatic amine) has significant effects upon negative cooperativity with dopamine.

Isosteric replacement of the secondary amine with an ether (**20**, Table 6) did not affect affinity in our binding assay, moreover this compound showed no activity in the functional assay highlighting the importance of the secondary amine contained within **1** and the potential requirement of a hydrogen-bond donor-acceptor at the 4-position for cooperativity with dopamine.

Table 6. Functional Parameters for Analogues of 1 with Chemical Modifications to the 4-position & 5,6-Fused System Derived from cAMP BRET Assay



	R¹	R²	R³	^a pK_B (<i>K_B</i> , μM)	^b Log <i>a</i> (<i>a</i>)	^c Log <i>β</i> (<i>β</i>)
15	H		N(CH ₃) ₂	5.31 ± 0.15 (4.90)	-0.17 ± 0.14 (0.68)	-0.84 ± 0.10 (0.15)
18	-	-	NCH ₃	nd		
19	-	-	N(CH ₂ CH ₃) ₂	6.18 ± 0.16 (0.662)	-0.17 ± 0.17 (0.68)	-1.10 ± 0.10 (0.08)
20	-	-		nd		
44	H	H	N(CH ₂ CH ₃) ₂	5.50 ± 0.10 (3.13)	-1.10 ± 0.16 (0.08)	-0.62 ± 0.15 (0.24)

^aEstimate of the negative logarithm of the equilibrium dissociation constant determined in an cAMP functional assay.

^bEstimate of the logarithm of the net cooperativity factor between the modulator and dopamine. ^cEstimate of the logarithm of the modulatory effect upon efficacy factor induced by the allosteric modulator. nd = inactive at concentrations up to 100 μM. Values represent the mean ± S.E.M. from at least three independent experiments performed in duplicate.

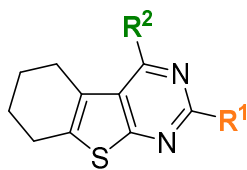
Our binding experiments revealed that modifications at the 2-position of the thienopyrimidine can influence the degree of negative cooperativity between the modulator and [³H]raclopride. Therefore, we examined the functional impact of removing the morpholinomethyl moiety with respect to varying substituents at the 4-position. Maintaining the *m*-(trifluoromethyl)anilino substituent (**23a**) did not affect functional affinity but resulted in a 3-fold increase in the modulatory effect upon dopamine efficacy ($\beta = 0.10$, Table 7). The

1 *N,N*-diethylamino substituent (**23b**) not only maintained functional affinity ($K_B = 1 \mu\text{M}$), but gave rise to
2 almost complete attenuation of dopamine signalling at a concentration of $10 \mu\text{M}$ indicative of a very high
3 negative modulatory effect upon dopamine efficacy (Figure 5C, Table 7). Interestingly, the pyrimidinone
4 analogue (**21**) was inactive and is in agreement with our finding that the secondary amine is a key structural
5 determinant of cooperativity.
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11 We next focused on the non-functionalised methylene spacer group within **1**, and the importance of
12 linker length (Table 7). Increasing linker length by one carbon atom had no effect on affinity, however the
13 nature of allostery was altered as **39a** modulated dopamine affinity with little effect upon dopamine efficacy
14 ($\alpha = 0.23$). Further extension of linker length by one carbon **39b** engendered an additional modulatory effect
15 upon dopamine efficacy ($K_B = 1.39 \mu\text{M}$, $\alpha = 0.21$, $\beta = 0.40$). Moreover, further extension of the linker **39c** was
16 observed to not only increase functional affinity ($K_B = 611 \text{ nM}$), but also increased negative cooperativity with
17 dopamine efficacy ($\beta = 0.03$). The addition of the *N,N*-diethylamino substituent (**40**) resulted in a similar
18 affinity as determined for **39c**, but displayed pharmacology best fit with a competitive model (Table 7).
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30 Interestingly, isosteric replacement of the morpholino nitrogen with a methine group (**33a**) caused a
31 10-fold increase in functional affinity ($K_B = 622 \text{ nM}$), as well as enhancing the negative allosteric modulatory
32 effect upon dopamine efficacy ($\beta = 0.13$). This finding suggests that the ionizable nitrogen within **1** is not a
33 structural requirement to maintain modulatory activity at the D_2R . However, the *N,N*-diethylamino
34 substitution (**33b**) was not tolerated in conjunction with the tetrahydropyran as the compound was inactive in
35 this functional assay. Isosteric replacement of the morpholino oxygen for a methylene group via the
36 incorporation of piperidine (**26**) maintained functional affinity ($K_B = 6.52 \mu\text{M}$), but saw a reduction in negative
37 allosteric cooperativity ($\alpha = 0.49$, $\beta = 0.68$) compared to **33a**.
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Table 7. Functional Parameters for Analogues of 1 with Chemical Modifications to the 2-Position
Derived from cAMP BRET Assay



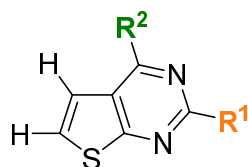
	R¹	R²	^a pK_B (K_B, μM)	^b Log α (α)	^c Log β (β)
21	H	C=O	nd		
23a	H		4.60 ± 0.17 (25.4)	= 0	-1.02 ± 0.35 (0.10)
23b	H	N(CH ₂ CH ₃) ₂	5.72 ± 0.13 (1.92)	= 0	= -3.0
26			-5.16 ± 0.13 (6.98)	-0.46 ± 0.12 (0.34)	-0.22 ± 0.08 (0.60)
33a			6.22 ± 0.26 (0.622)	-0.89 ± 0.29 (0.13)	-0.63 ± 0.19 (0.23)
33b		N(CH ₂ CH ₃) ₂	nd		
39a			6.01 ± 0.17 (0.981)	-0.63 ± 0.05 (0.23)	= 0
39b			5.79 ± 0.11 (1.63)	-0.68 ± 0.12 (0.21)	-0.40 ± 0.08 (0.40)
39c			6.21 ± 0.11 (0.611)	-0.72 ± 0.18 (0.19)	-1.50 ± 0.15 (0.03)
40		N(CH ₂ CH ₃) ₂	5.80 ± 0.20 (1.77)	^e Schild Slope: 1.07 ± 0.11	

^aEstimate of the negative logarithm of the equilibrium dissociation constant ± SEM determined in an cAMP functional assay. ^bEstimate of the logarithm of the net cooperativity factor between the modulator and dopamine ± SEM determined in an cAMP functional assay. nd = inactive at concentrations up to 100 μM. Values represent the mean ± S.E.M. from at least three independent experiments performed in duplicate.

We also investigated the impact of bis-functionalisation of **1** with respect to incorporating various amines at the 4-position of **1** in this functional assay (Table 8). Two analogues of **1** that incorporate the morpholinobutyl substituent, in conjunction with removal of the fused cyclohexane system, together with one of either *m*-CF₃-anilino (**43a**) or *N,N*-diethylamino substituents (**43b**) maintained functional affinity ($K_B = 1.3$ μM and 1.4 μM, respectively) comparable to **1**, but now displayed a higher level of negative cooperativity

with both dopamine affinity and efficacy (Table 8). These data further indicate that linker length extension of the morpholine moiety acts to increase negative cooperativity with dopamine.

Table 8. Functional Parameters for Analogues of 1 with Bi-Functionalisation in Conjunction with Aromatic or Aliphatic 4-Substituted Amine



	R¹	R²	^a pK_B (<i>K_B</i> , μM)	^b Log α (α)	^c Log β (β)
43a			5.85 ± 0.10 (1.4)	-0.88 ± 0.16 (0.13)	-1.21 ± 0.16 (0.06)
43b		N(CH ₂ CH ₃) ₂	5.89 ± 0.09 (1.3)	-1.34 ± 0.21 (0.05)	-0.82 ± 0.18 (0.15)
47a	H		6.13 ± 0.21 (0.8)	-0.43 ± 0.06 (0.37)	= 0
47b	H	N(CH ₂ CH ₃) ₂	5.31 ± 0.07 (4.6)	= 0	-0.89 ± 0.04 (0.13)

^aEstimate of the negative logarithm of the equilibrium dissociation constant determined in an cAMP functional assay.

^bEstimate of the logarithm of the net cooperativity factor between the modulator and dopamine. ^cEstimate of the logarithm of the modulatory effect upon efficacy factor induced by the allosteric modulator. Values represent the mean ± S.E.M. from at least three independent experiments performed in duplicate.

In agreement with our binding data, analogues of **1** devoid of both the morpholinomethyl and fused cyclohexane moieties but incorporating one of either *m*-(trifluoromethyl)aniline (**47a**), or *N,N*-diethylamine (**47b**, Figure 5D) maintained functional affinity and displayed robust negative cooperativity with dopamine efficacy and affinity despite the lack of a substituent at the 2-position or the presence of the fused cyclohexane system ($K_B = 1.3 \mu\text{M}$, $\alpha = 0.05$, $\beta = 0.15$ and $K_B = 1.4 \mu\text{M}$, $\alpha = 0.13$, $\beta = 0.06$, respectively, table 8). These data demonstrate the utility of the thieno[2,3-*d*]pyrimidine scaffold as a NAM of the dopamine effect as ‘pruned back’ analogues maintain negative allosteric cooperativity whilst preserving functional affinity relative to **1**. Through successive compound iterations, both the lipophilic ligand efficiency (LLE) and ligand efficiency (LE) of the scaffold have been enhanced significantly compared to that of **1**, with the most efficient

compound being **47b** (Figure 6, LLE = 2.48, LE = 0.54). This reveals the versatility of the scaffold for further structural optimisation and development of ligands that non-competitively modulate dopamine affinity and signalling efficacy at the D₂R. Collectively, these results demonstrate that analogues of **1** mediate their effects through an allosteric mode of action. While we were able to gain only relatively modest (10-fold) improvement in affinity, our SAR study revealed compounds with a range of modulatory effects upon dopamine efficacy and/or affinity.

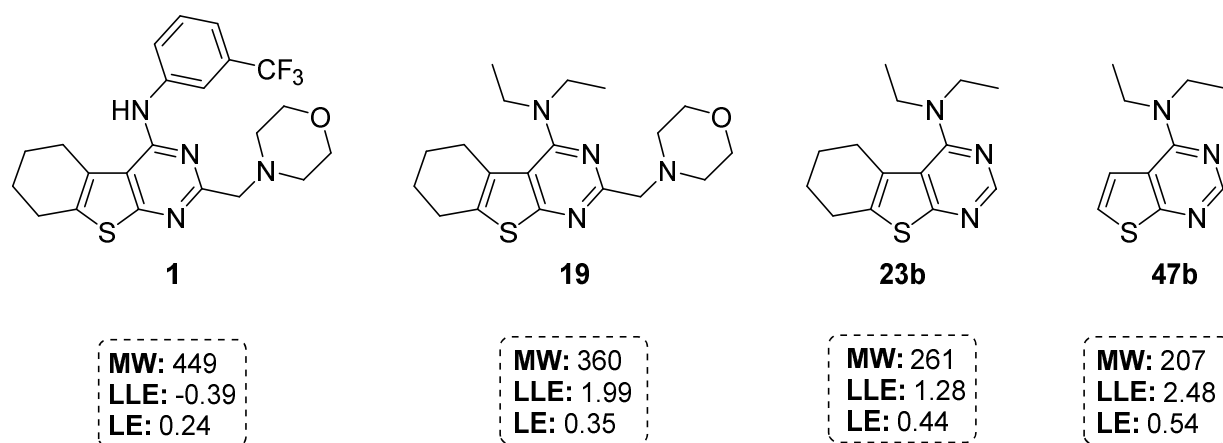
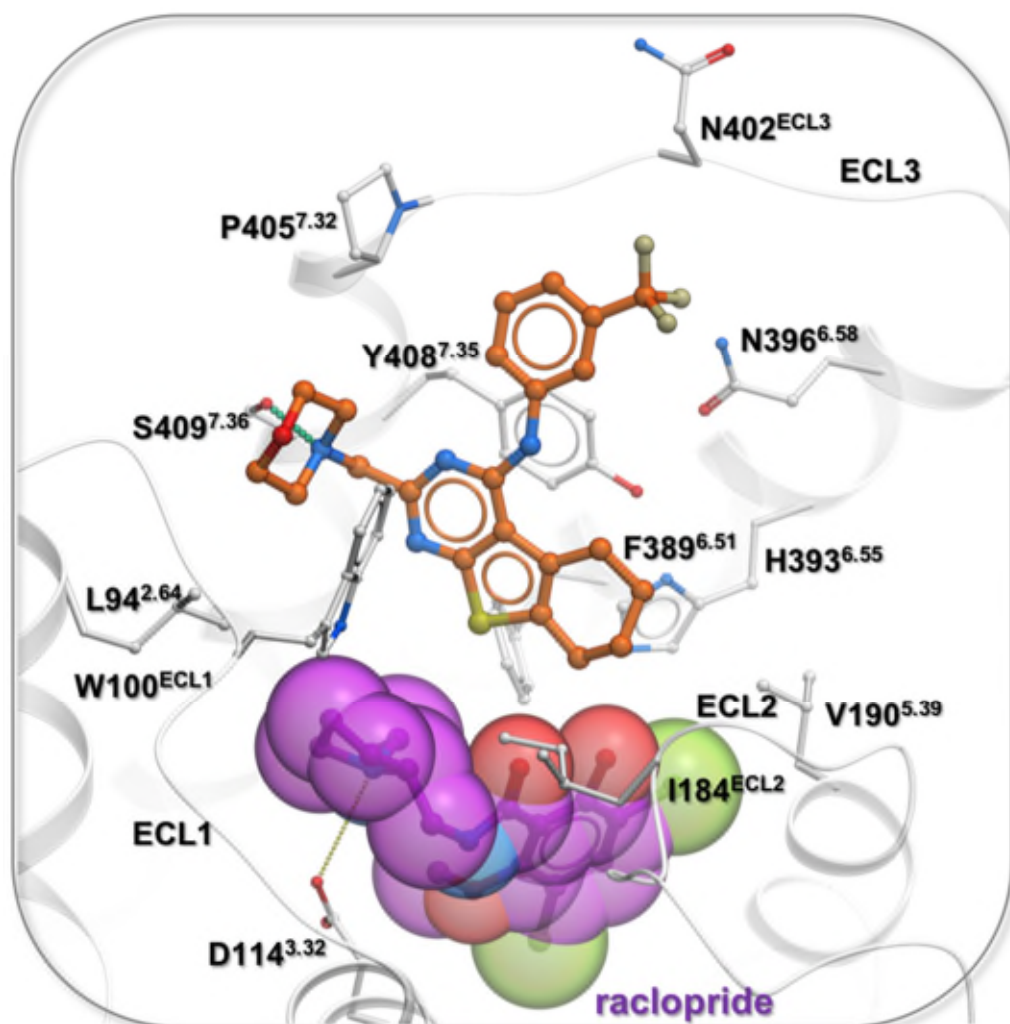


Figure 6. Lipophilic ligand efficiencies (LLE), and ligand efficiencies (LE) of **1**, **19**, **23b**, and **47b** calculated according to equilibrium dissociation constants derived from functional interaction experiments measuring cAMP accumulation. Multiple compound iterations have yielded improvements in the scaffold's efficiency through decreased molecular weight and functional binding affinity. This is particularly evident with compound **47b** as it shows a marked improvement in both LLE and LE compared to that of **1**, resulting in a promising fragment-like starting scaffold that can be elaborated through multiple vectors.

Molecular Docking Studies. The recently solved structure of D₂R receptor (PDB ID: 6C38)³² provides an opportunity to accurately model the allosteric binding of compound **1** and a library of structurally diverse analogues. Specifically, we were interested in the interactions of these allosteric compounds with the orthosteric ligand, as reflected in the [³H]raclopride equilibrium binding assays (Table 1). While the D₂R structure was solved with the atypical antipsychotic drug risperidone, we first generated a conformational model of raclopride bound to the D₂R (D₂R^{Raclo}). Flexible docking of raclopride shows a pose similar to the pose of eticlopride found in the D₃R structure. Both raclopride and eticlopride are known to bind to the

1 orthosteric pocket of the D₂R commonly occupied by dopamine, however, since it is bulkier than dopamine,
2 the allosteric pocket in the D₂R^{Raclo} complex is shallower, as compared to the D₂R^{Dopa} complex.
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4 Docking of compound **1** in the D₂R^{Raclo} complex consistently yielded the best scoring pose shown in Figure
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7 7. This pose is stabilised by a hydrogen bond between the ionised morpholine secondary amine and the oxygen
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9 atom of the side-chain of S409^{7.36} (Ballesteros-Wenstein nomenclature³³), as well π - π stacking interactions of
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11 the thienopyrimidine moiety with Y408^{7.35} and W100^{ECL1} side chains. In addition, the fused cyclohexane ring
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13 system is positioned in the hydrophobic sub-pocket located between helices V and VI formed by the residues
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15 I184^{ECL2}, V190^{5.39}, H394^{6.55}, whereas the phenyl ring bearing the *m*-CF₃ substituent is placed between
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17 helices VI and VII, and ECL3, and is lined by the residues N396^{6.58}, N402^{ECL3}, P405^{7.32}, and Y408^{7.35} (Figure
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57 **Figure 7.** Compound **1** (orange carbon atoms) predicted binding mode in complex with raclopride (magenta
58 carbon atoms) (D₂R^{Raclo} conformational model (PDB ID: 6C38)). D₂R structural model is shown in light grey
59 cartoon and stick presentation.
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2 The key SAR data support the consistent binding pose of compound **1** and its analogues as illustrated in Figure
3 8 and Supplementary Figure 2. First, we examined modifications to the fused cyclohexane system of **1**, (**14a-**
4 **j**). Most analogues bind in a similar mode, maintaining the key interactions of the scaffold (Supplementary
5 Figure 2A). At the same time, the docking illustrates a modest binding dependence on the size of this fused
6 hydrophobic ring, as replacement of the cyclohexane with either cyclopentane (**14a**) or cycloheptane (**14b**)
7 would make less favorable hydrophobic interactions, reducing allosteric binding affinity by ~2-4-fold (Figure
8 8A). Moreover, compounds that lack the fused ring system (**14h, j**) showed an even more pronounced decrease
9 in binding, consistent with loss of interactions with this hydrophobic sub-pocket (Figure 8B).

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21 Next, we examined compounds with variations to in the *m*-CF₃ substituent (**16a-c, 17**) positioned between
22 helices VI and VII, and ECL3, but exposed to solvent. As the CF₃ substituent is favorable for solvent
23 interactions, its removal in **16a** explains the observed 6-fold decrease in binding for this compound. The
24 reduction is much smaller for derivatives that have polar substituents in this position, 3-OH (**17**) and 3-OMe
25 (**16c**) (Supplementary Figure 2B).

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Next, we examined compounds with variations in *N*-(3-trifluoromethyl)phenylamino substituent (**15, 18, 19**).
The docking poses place *N,N*-dimethylamino (**15**), *N*-methylamino (**18**), and *N,N*-diethylamino moieties (**19**)
into the hydrophobic sub-pocket in the proximity of Y408^{7,35} and W100^{ECL1}, with **19** having the most
extensive hydrophobic contacts. This is in agreement with an improved binding affinity for **19** (Supplementary
Figure 2C). The secondary amine of compound **1** is not engaged in any interactions, and its isosteric
replacement to an ether (**20**) did not impact the predicted binding mode (Supplementary Figure 2D), nor its
binding affinity (Table 1).

Interestingly, removal of the morpholinomethyl moiety (**23a**) maintained the π - π stacking interactions of the
thienopyrimidine moiety with the Y408^{7,35} and W100^{ECL1}, though slightly shifting the scaffold, enabling an
additional hydrogen bond interaction with T412^{7,39}. We hypothesize that though loss of the morpholine system
abolished the interaction with the S409^{7,36} (Figure 8C), the new hydrogen bond at least partially compensated
the loss, maintaining binding affinity of this analogue (Table 1). However, replacing morpholine with

1 piperidine (**26**) leads to 2.5-fold decrease in affinity, reflecting replacement of a polar oxygen with a carbon
2 in this solvent-exposed position (Supplementary Figure 2E).
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4 Moreover, removal of the tertiary amine in the morpholine ring abolishes the hydrogen bond interaction of
5 **33a** with S409^{7,36} (Figure 8D), with the importance of this hydrogen bond corroborated by a ~7-fold decrease
6 in affinity for this compound (Table 1).
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10 Furthermore, we evaluated the effect of elongating the methylene linker to the morpholine substituent (**39a-**
11 **c**). In our model, increasing the linker length did not hinder the ability of the compounds to maintain crucial
12 interactions, and even helped to improve complementarity of **39c** with the binding pocket, in accordance with
13 its improved affinity (Figure 8E).
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20 Other docked analogues, characterised by morpholinomethyl deletion (**23b**), tetrahydropyranomethyl (**33b**),
21 morpholinobutyl (**40**), and fused cyclohexane deletion (**44**), also maintained the crucial π - π interactions with
22 Y408^{7,35} and W100^{ECL1} (Supplementary Figure 2F). Similar poses were obtained for the two compounds **43a-**
23 **b**, comprised of the morpholinobutyl moiety and four carbon linker, but devoid of the fused cyclohexane
24 system (Supplementary Figure 2G).
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32 Finally, we docked compounds devoid of the fused cyclohexane system and morpholinomethyl moieties but
33 maintained substitution with one of either *m*-trifluoromethylanilino or *N,N*-diethylamino (**47a-b**). This
34 dramatically reduced the size of the scaffold, but also maintained π - π stacking interactions with Y408^{7,35} and
35 W100^{ECL1}, while shifting to gain a hydrogen bond interaction with T412^{7,39} (Figure 8F), which is in agreement
36 with only a minor loss of binding affinity (Table 1).
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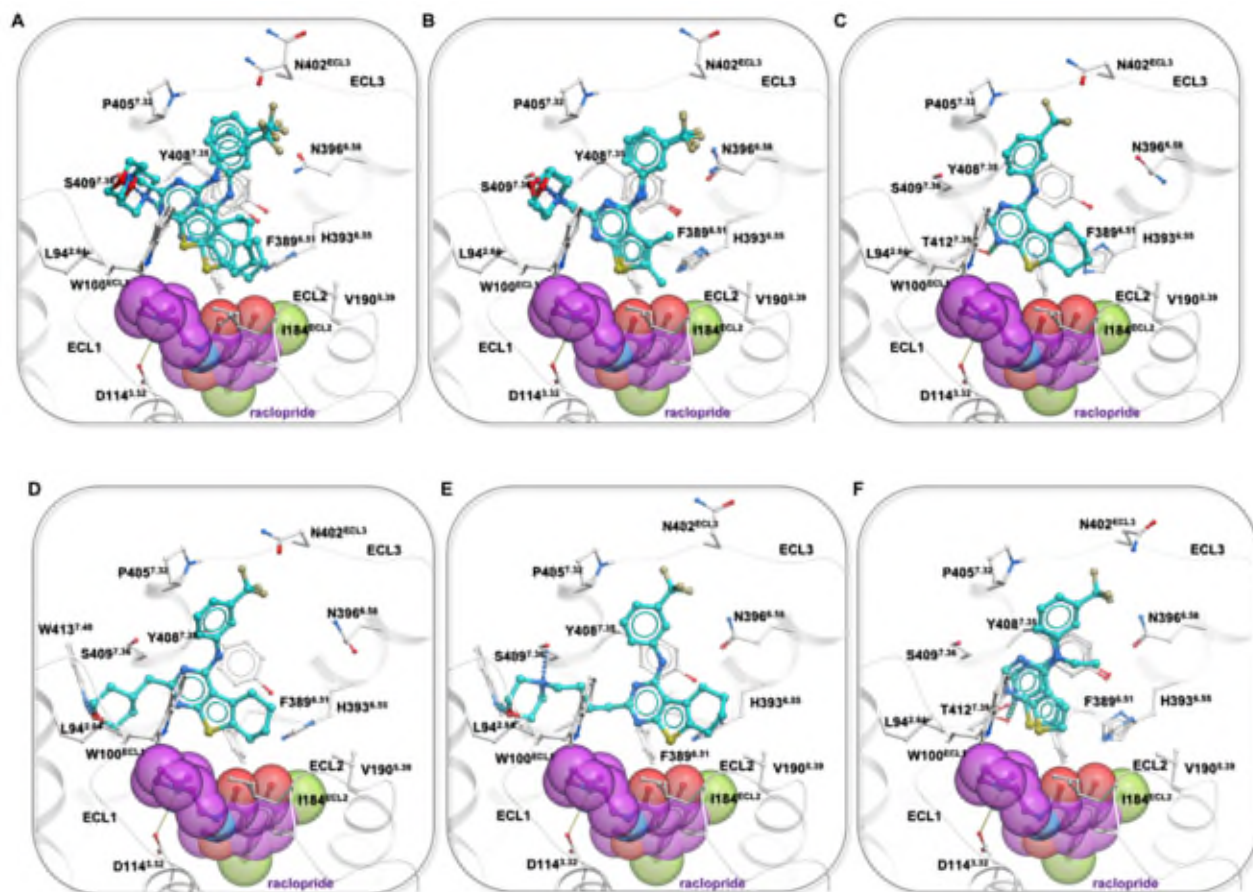


Figure 8. Compound 1 analogues in complex with D_2R^{Raclo} (PDB ID: 6C38). Compounds with variation in the fused cyclohexane system: (A) Compounds **14a, c**. (B) Compounds **14h, j**. (C) Compound devoid of morpholinomethyl moiety (**23a**). (D) Compound with tetrahydropyran moiety (**33a**). (E) Compounds with variations in alkyl linker length to the morpholine motif (**39c**). (F) Fragment-like compounds (**47a-b**).

CONCLUSIONS

In this study, we report the pharmacological validation of a VLS hit, confirming it exerts its effect via an allosteric mode of action at the D_2R , as predicted by our initial modeling studies. We subsequently describe the design and characterisation of a structurally novel series of dopamine D_2R NAMs based on this thieno[2,3-*d*]pyrimidine scaffold, demonstrating its scope for future development of D_2R NAMs. The impact of structural modification was assessed with respect to thienopyrimidine substituents (2-, 4-, 5- and 6-positions), isosteric replacement of the aniline nitrogen with respect to hydrogen-bonding, as well as the importance of bearing an

1 ionisable nitrogen at differing linker lengths on the 2-substituent and its subsequent impact on the allosteric
2 pharmacology of **1**. The majority of active compounds displayed comparable binding affinities to that of **1**.
3 Rather, the structural modifications within this study had a profound effect upon the extent of modulation of
4 dopamine binding and/or efficacy. Indeed, while **1** exerted its effect predominally via negative modulation of
5 dopamine efficacy, we identified analogues of **1** that displayed modulatory effects upon dopamine affinity
6 (e.g. **14a**, **14i**, **39a**), and those that displayed modulatory effects upon both dopamine affinity and efficacy
7 (e.g. **17**, **19**, **43a**). Maintaining a hydrogen-bond donor (secondary amine) at the 4-position was found to be a
8 requirement for cooperativity with dopamine, as thienopyrimidinone analogues were inactive in our functional
9 assay. Moreover, the ether analogue of **1** (**20**) was also inactive in our functional assay, suggesting this
10 modification is not well tolerated. Most significantly, we show that this scaffold can be effectively ‘pruned
11 back’ of morpholinomethyl and fused cyclohexane moieties, to reveal a low molecular weight fragment-like
12 core with low μM functional affinity and very high lipophilic ligand efficiency (**47a-b**). Our molecular
13 docking studies, using the recently determined crystal structure as a template, provide some insight into the
14 molecular determinants of this SAR. Thus, this minimal thieno[2,3-*d*]pyrimidine scaffold provides an
15 attractive starting point for further structural interrogation and elaboration of the core scaffold to achieve high
16 affinity, low molecular weight NAMs of the D₂R with varying effects upon dopamine binding and/or function.
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39 ■ EXPERIMENTAL SECTION:

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42 **Chemistry: General Information and Synthetic Procedures.** Chemicals and solvents were purchased
43 from standard suppliers and used without further purification. Davisil silica gel (40–63 μm) for flash column
44 chromatography was supplied by Grace Davison Discovery Sciences (Victoria, Australia), and deuterated
45 solvents were purchased from Cambridge Isotope Laboratories, Inc. (USA, distributed by Novachem PTY.
46 Ltd., Victoria, Australia). Reactions were monitored by thin layer chromatography on commercially available
47 precoated aluminum-backed plates (Merck Kieselgel 60 F₂₅₄). Visualisation was by examination under UV
48 light (254 and 366 nm). A solution of ninhydrin (in ethanol) was used to visualise primary and secondary
49 amines. All organic extracts collected after aqueous workup procedures were dried over anhydrous MgSO₄ or
50 Na₂SO₄ before gravity/vacuum filtering and evaporation to dryness. Organic solvents were evaporated *in*
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vacuo at ≤ 40 °C (water bath temperature). ^1H NMR and ^{13}C NMR spectra were recorded on a Bruker Avance Nanobay III 400 MHz Ultrashield Plus spectrometer at 400.13 and 100.62 MHz, respectively. Chemical shifts (δ) are recorded in parts per million (ppm) with reference to the chemical shift of the deuterated solvent. Coupling constants (J) are recorded in Hz, and the significant multiplicities described by singlet (s), doublet (d), triplet (t), quadruplet (q), broad (br), multiplet (m), doublet of doublets (dd), and doublet of triplets (dt). Spectra were assigned using appropriate COSY, distortionless enhanced polarization transfer (DEPT), HSQC, and HMBC sequences. LC-MS were run to verify reaction outcome and purity using an Agilent 6120 series single quad coupled to an Agilent 1260 series HPLC. The following buffers were used: buffer A, 0.1% formic acid in; buffer B, 0.1% formic acid in MeCN. The following gradient was used with a Poroshell 120 EC-C18 50 mm \times 3.0 mm 2.7 μm column, and a flow rate of 0.5 mL/min and total run time of 5 min; 0–1 min 95% buffer A and 5% buffer B, from 1 to 2.5 min up to 0% buffer A and 100% buffer B, held at this composition until 3.8 min, 3.8–4 min 95% buffer A and 5% buffer B, held until 5 min at this composition. Mass spectra were acquired in positive and negative ion mode with a scan range of 100–1000 m/z . UV detection was carried out at 214 and 254 nm. All retention times (t_R) are quoted in minutes. All screening compounds were of >95% purity unless specified in the individual monologue. All NMR experiments were performed in CDCl_3 to permit comparison of the spectra of the various analogues. Experiments were performed in acetone- d_6 , DMSO- d_6 , or MeOH- d_4 if selected analogues lacked solubility in CDCl_3 .

General Synthetic Procedures. General Procedure A for the Gewald Synthesis of 2-Amino-3-Carboethoxy Thiophenes 5, 10a-c. To a stirred solution of ethyl cyanoacetate (1 equiv.) in absolute ethanol was added sulfur powder (1 equiv.), followed by the aldehyde or ketone (1 equiv) at r.t. under N_2 and the resulting mixture cooled to 0–5 °C. To this was added diethylamine (1 equiv.) and the reaction mixture warmed to r.t. and subsequently stirred at 50–55 °C for 5 h. The reaction was concentrated under reduced pressure and the residue was chromatographed on silica eluting with petroleum ether/EtOAc (10:1) to give the desired compound as a solid or oil which can be recrystallised from an appropriate solvent as indicated.

General Synthetic Procedures. General Procedure B for the Gewald Synthesis of 2-Amino-3-Carboethoxy Thiophenes 5, 10d-i. An equimolar mixture of powdered sulfur and morpholine was stirred until total dissolution of the sulfur. After, the ethyl cyanoacetate (1 equiv.) and the aldehyde or ketone (1 equiv.) were

1 added to the reactional mixture, which was stirred at room temperature for ~18 h. After completion of the
2 reaction, as monitored by TLC, the crude product was chromatographed on silica with an appropriate eluent
3 as indicated to afford the desired compound which could be recrystallised from an appropriate solvent as
4 indicated.
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9 *General Procedure C for the Preparation of 6, 11a-j, 31, 36a-c, 37b-c, 41.* The required amino ethyl ester
10 thiophene (1 equiv) and appropriate nitrile (1.2 equiv) were taken up in 1,4-dioxane, heated to 50 °C, and
11 passed with dry HCl gas (Kipp's apparatus) until the starting material was absent in 3-24 h or a precipitate
12 formed. The reaction mixture was filtered directly and the solid washed with *n*-hexane to obtain the desired
13 compound. Depending on purity, the solid may be recrystallised from an appropriate solvent or reacted without
14 further purification. Alternatively, the reaction mixture can be concentrated under reduced pressure and the
15 residue chromatographed with an appropriate eluent as indicated.
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25 *General Procedure D for the Preparation of 7, 12a-j, 37a.* The required 2-chloro-thienopyrimidinone (1
26 equiv.) was taken up in DMF. To this solution/suspension was added morpholine (2.5 equiv) and the resulting
27 solution/suspension was stirred at 100 °C for 2-3 h. The reaction mixture was then cooled to room temperature
28 or ~4 °C to induce crystallisation of product. Any crystals were collected using vacuum filtration, washed
29 with cold *i*-PrOH and again recrystallised from *i*-PrOH. In the case of present morpholinium hydrochloride,
30 the solids were dissolved in CHCl₃, washed with 5% NaHCO₃ solution, water, brine, and the organic extracts
31 concentrated *in vacuo*. If no crystallisation from reaction mixture was evident, the residue can be
32 chromatographed on silica using 1% Et₃N/EtOAc unless otherwise indicated.
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44 *General Procedure E for the Preparation of Chloropyrimidines 8, 13a-j, 22, 25, 38a-c, 42, 46.* The required
45 thienopyrimidinone (1 equiv.) was taken up in toluene. To this solution/suspension was added phosphorous
46 oxychloride (4 equiv), *N,N*-dimethylformamide (cat.) and the solution/suspension was stirred at 100 °C. After
47 TLC has indicated complete consumption of starting material, the reaction mixture was slowly introduced into
48 a stirred suspension of ice water and the pH adjusted to ~12 with 6 M NaOH. The aqueous phase was extracted
49 with EtOAc and the combined organic extracts dried over anhydrous sodium sulfate and concentrated *in vacuo*.
50 The resulting oil/solid, depending on purity, can be reacted without further purification, or further purified
51 using column chromatography (1:99 Et₃N:EtOAc) or recrystallised as indicated.
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General Procedure F for the Synthesis of 1, 14a-j, 16a-c, 18-20, 23a-b, 33a-b, 39a-c, 40, 43a-b, 44, 47a-b.

In a suitable microwave reaction vessel the required chloropyrimidine (1equiv.) was taken up in *i*-PrOH. To this was added the required amine (1.1 equiv) and the mixture irradiated under stirring at 120 °C for 1-2 h. Upon completion of the reaction, the mixture can purified using silica gel chromatography to afford the compound. Similarly, any precipitate can be collected under vacuum and washed with cold *i*-PrOH to afford the desired compound.

2-(Morpholinomethyl)-*N*-(3-(trifluoromethyl)phenyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4-amine (1). General procedure F. The residue was chromatographed on silica (1:99 MeOH/CHCl₃) which furnished the title compound as a white foam (160 mg, 70%). LCMS (*m/z*): 449.2 [M + H]⁺. HPLC: *t*_R 7.912 min. HRMS (*m/z*): C₂₃H₂₃F₃N₃OS requires 449.1671 [M + H]⁺; found 449.1661. ¹H NMR δ 8.17 (s, 1H), 7.93 (dd, *J* = 8.1, 1.6 Hz, 1H), 7.48 (t, *J* = 8.0 Hz, 1H), 7.34 – 7.31 (d, *J* = 7.7 Hz, 1H), 7.26 (s, 1H), 3.77 (s, 2H), 3.09 (t, *J* = 6.0 Hz, 2H), 2.87 (t, *J* = 6.0 Hz, 2H), 2.58 (s, 4H), 2.04 – 1.97 (m), 1.98 – 1.94 (m, 2H), 1.66 (dt, *J* = 11.2, 5.6 Hz, 4H). ¹³C NMR (CDCl₃) δ 167.5, 161.4, 154.2, 139.7, 135.0, 131.4 (q, *J* = 32.3 Hz), 129.5, 124.2, 124.1 (q, *J* = 272.4 Hz), 123.2, 119.7 (q, *J* = 3.8 Hz), 117.1 (q, *J* = 4.0 Hz), 115.1, 65.8, 54.8, 26.6, 26.0, 24.3, 22.7, 22.5.

2-Amino-4,5,6,7-tetrahydrobenzo[*b*]thiophene-3-carboxylate³³ (4). General Procedure A. The reaction mixture was kept for 30 min at 0 °C and formed crystals were collected under vacuum filtration, washed with chilled ethanol (2 × 50 mL) and dried to give the title compound as bright yellow needles which were used as such for the next reaction without further purification (2.83 g, 80%). mp: 110-114 °C. ¹H NMR (*d*₆-acetone) δ 6.88 (s, 2H), 4.21 (q, *J* = 7.1 Hz, 2H), 2.72 – 2.60 (m, 2H), 2.45 (m, 2H), 1.81 – 1.63 (m, 4H), 1.29 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (*d*₆-acetone) δ 165.6, 162.9, 131.9, 116.2, 104.1, 58.7, 26.8, 24.1, 23.1, 22.7, 13.9. LCMS (*m/z*): 226.2 [M + H]⁺. HPLC: *t*_R 8.389 min. HRMS (*m/z*): C₁₁H₁₅NO₂S requires 225.0818 [M + H]⁺; found 225.0818.

2-(Chloromethyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4(3*H*)-one (6).¹⁶ General procedure C. The reaction mixture was filtered directly and the solid washed with *n*-hexane to obtain the desired compound as a beige solid (4.21 g, 93%). LCMS (*m/z*): 255.1 [M + H]⁺. HPLC: *t*_R 6.858 min. HRMS (*m/z*): C₁₁H₁₁ClN₂OS requires 255.0354 [M + H]⁺; found 255.0353. ¹H NMR (*d*₆-DMSO) δ 12.72 (s, 1H),

4.56 (s, 2H), 2.85 (t, $J = 5.8$ Hz, 2H), 2.73 (t, $J = 5.8$ Hz, 2H), 1.76 (ddd, $J = 7.5, 6.8, 4.4$ Hz, 4H). ^{13}C NMR (d_6 -DMSO) δ 162.4, 158.6, 152.4, 133.7, 131.3, 122.1, 43.0, 25.6, 24.9, 22.8, 22.1.

2-(Morpholinomethyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4(3*H*)-one (7). General

procedure D. Reaction cooled and crystals collected under vacuum filtration to obtain the desired compound as a white crystalline solid (2.59 g, 70%). LCMS (m/z): 306.1 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 4.334 min. ^1H NMR (CDCl_3) δ 10.14 (s, 1H), 3.80 – 3.73 (m, 4H), 3.57 (s, 2H), 3.09 – 3.02 (m, 2H), 2.94 (t, $J = 7.3$ Hz, 2H), 2.63 – 2.57 (m, 4H), 2.51 – 2.40 (m, 2H). ^{13}C NMR (CDCl_3) δ 168.6, 158.2, 152.2, 140.1, 138.7, 119.5, 66.7, 60.6, 53.6, 32.2, 29.5, 28.9, 27.

4-((4-Chloro-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-2-yl)methyl)morpholine (8).

General Procedure E. The residue was chromatographed on silica (1:99 $\text{Et}_3\text{N}:\text{EtOAc}$) to afford the compound as a brown oil (270 mg, 67%). LCMS (m/z): 324.1 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 5.112 min. ^1H NMR (CDCl_3) δ 3.80 (s, 2H), 3.06 (t, $J = 4.2$ Hz, 2H), 2.84 (t, $J = 4.2$ Hz, 2H), 2.52 – 2.49 (m, 4H), 1.93 – 1.86 (m, 2H), 1.67 – 1.57 (m, 4H), 1.49 – 1.36 (m, 2H). ^{13}C NMR (CDCl_3) δ 169.4, 161.4, 152.9, 138.9, 126.9, 126.9, 65.2, 54.7, 26.3, 26.0, 25.7, 24.1, 22.2.

Ethyl 2-amino-5,6-dihydro-4*H*-cyclopenta[*b*]thiophene-3-carboxylate (10a).¹⁵ General Procedure A. The

reaction was concentrated under reduced pressure and the residue eluted through a silica gel column (10:1 petroleum ether:EtOAc) to afford the compound as a yellow solid which later turned a dark brown (1.18 g, 37%). General procedure B. The reaction was concentrated under reduced pressure and the residue eluted through a silica gel column (10:1 petroleum ether:EtOAc) to afford the compound as a yellow solid which later turned brown (2.21 g, 44%). LCMS (m/z): 212.1 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 8.102 min. ^1H NMR (CDCl_3) δ 5.85 (s, 2H), 4.24 (q, $J = 7.1$ Hz, 2H), 2.82 (ddd, $J = 7.9, 3.9, 1.9$ Hz, 2H), 2.74 – 2.68 (m, 2H), 2.35 – 2.26 (m, 2H), 1.32 (t, $J = 7.1$ Hz, 1H). ^{13}C NMR (CDCl_3) δ 166.4, 165.9, 142.8, 121.4, 103.0, 59.5, 30.9, 29.0, 27.4, 14.6.

Ethyl 2-amino-5,6,7,8-tetrahydro-4*H*-cyclohepta[*b*]thiophene-3-carboxylate (10b). General procedure A.

The reaction was concentrated under reduced pressure and the residue chromatographed on silica (10:1 petroleum ether:EtOAc). The resulting residue was recrystallised from CHCl_3 /petroleum ether to afford the desired compound as yellow crystals (1.55 g, 48%). LCMS (m/z): 240.2 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 8.791 min.

General procedure B. The crude product was purified by silica gel column chromatography (10:1 petroleum ether:EtOAc) to afford the title compound as a yellow solid (3.87 g, 72%). ^1H NMR (CDCl_3) δ 5.76 (s, 2H), 4.28 (q, $J = 7.1$ Hz, 2H), 3.00 – 2.94 (m, 2H), 2.60 – 2.54 (m, 2H), 1.85 – 1.76 (m, 2H), 1.68 – 1.56 (m, 4H), 1.34 (t, $J = 7.1$ Hz, 3H). ^{13}C NMR (CDCl_3) δ 166.1, 159.9, 138.0, 121.4, 107.6, 59.6, 32.2, 28.7, 28.7, 27.9, 27.0, 14.5.

Ethyl 2-amino-4,5,6,7,8,9-hexahydrocycloocta[*b*]thiophene-3-carboxylate (10c). General procedure A.

The solvent was removed under reduced pressure and the residue eluted through a silica gel column (10:1 petroleum ether:EtOAc). The product was recrystallised from CHCl_3 /petroleum ether to give the desired compound as yellow crystals (1.32 g, 37%). LCMS (m/z): 254.2 [$\text{M} + \text{H}$] $^+$. HPLC: t_{R} 9.405 min. General procedure B. After completion of the reaction (24 h), as monitored by TLC, the mixture was concentrated under reduced pressure and the crude product was purified by silica gel column chromatography (10:1 petroleum ether:EtOAc) to afford the title compound as a yellow solid (2.63 g, 65%). ^1H NMR (CDCl_3) δ 5.94 (s, 1H), 4.27 (q, $J = 7.1$ Hz, 2H), 2.85 – 2.77 (m, 2H), 2.61 – 2.57 (m, 2H), 2.43 – 2.36 (m, 1H), 1.91 – 1.82 (m, 1H), 1.65 – 1.59 (m, 2H), 1.58 – 1.52 (m, 2H), 1.49 – 1.41 (m, 2H), 1.37 – 1.26 (m, 3H). ^{13}C NMR (CDCl_3) δ 166.1, 161.5, 134.9, 120.2, 106.2, 59.5, 32.1, 29.9, 26.8, 26.6, 25.9, 25.7, 14.4.

Ethyl 2-amino-5-phenylthiophene-3-carboxylate (10d). General procedure B. After completion of the

reaction, as monitored by TLC, the mixture was concentrated under reduced pressure and the crude product was purified by silica gel column chromatography (10:1 petroleum ether:EtOAc). This resulting brown solid was recrystallised from *i*-PrOH to afford the title compound as dark orange needles (6.30 g, 54%). LCMS (m/z): 248.1 [$\text{M} + \text{H}$] $^+$. HPLC: t_{R} 7.946 min. ^1H NMR (CDCl_3) δ 7.47 – 7.42 (m, 2H), 7.36 – 7.30 (m, 2H), 7.25 (s, 1H), 7.23 – 7.18 (m, 1H), 6.02 (s, 2H) 4.31 (q, $J = 7.1$ Hz, 2H), 1.37 (t, $J = 7.1$ Hz, 3H). ^{13}C NMR (CDCl_3) δ 165.5, 162.1, 134.1, 128.9, 126.7, 124.9, 124.8, 121.3, 108.1, 60.0, 14.6.

Ethyl 2-amino-4-phenylthiophene-3-carboxylate (10e). General procedure B. The reaction mixture was

concentrated under reduced pressure and the crude product was purified by silica gel column chromatography (10:1 petroleum ether:EtOAc). This resulting yellow solid was recrystallised from *i*-PrOH to afford the title compound as yellow crystals (4.88 g, 49%). LCMS (m/z): 248.1 [$\text{M} + \text{H}$] $^+$. HPLC: t_{R} 7.379 min. ^1H NMR

(CDCl₃) δ 7.33 – 7.28 (m, 5H), 6.10 (s, 2H), 6.07 (s, 2H), 4.04 (q, J = 7.1 Hz, 2H), 0.93 (t, J = 7.1 Hz, 3H).

¹³C NMR (CDCl₃) δ 165.8, 163.9, 141.7, 138.6, 129.0, 127.3, 126.9, 105.6, 59.5, 13.7.

Ethyl 2-amino-5-benzylthiophene-3-carboxylate (10f). General procedure B. After completion of the reaction, as monitored by TLC, the mixture was concentrated under reduced pressure and the crude product chromatographed on silica (10:1 petroleum ether:EtOAc) to afford the title compound as a yellow oil (2.71 g, 35%). LCMS (m/z): 262.1 [M + H]⁺. HPLC: t_R 8.633 min. ¹H NMR (CDCl₃) δ 7.30 – 7.26 (m, 2H), 7.23 – 7.18 (m, 3H), 6.69 (t, J = 1.0 Hz, 1H), 5.83 (s, 2H), 4.22 (q, J = 7.1 Hz, 2H), 3.87 (s, 2H), 1.30 (t, J = 7.1 Hz, 1H). ¹³C NMR (CDCl₃) δ 165.4, 162.3, 139.9, 128.5, 128.49, 126.5, 125.1, 122.8, 106.1, 59.6, 36.0, 14.5.

Ethyl 2-amino-4-cyclohexylthiophene-3-carboxylate (10g). General procedure B. After completion of the reaction, as monitored by TLC, the reaction mixture was concentrated under reduced pressure and the crude product purified by silica gel column chromatography (10:1 petroleum ether:EtOAc). Collected fractions were concentrated under reduced pressure and placed in the freezer for 2 h. Formed solids were collected under vacuum and washed with petroleum ether to afford the compound as clear crystals (3.56 g, 38%). LCMS (m/z): 254.2 [M + H]⁺. HPLC: t_R 8.936 min. ¹H NMR (CDCl₃) δ 6.06 (s, 2H), 5.85 (s, 1H), 4.29 (q, J = 7.1 Hz, 2H), 3.03 (tt, J = 11.4, 2.5 Hz, 1H), 1.97 (d, J = 12.0 Hz, 2H), 1.80 (d, J = 12.6 Hz, 2H), 1.73 (d, J = 12.5 Hz, 1H), 1.37 (t, J = 7.2 Hz, 5H), 1.28 – 1.15 (m, 3H). ¹³C NMR (CDCl₃) δ 166.2, 164.5, 147.4, 100.6, 59.7, 39.4, 34.1, 27.1, 26.6, 14.1.

Ethyl 2-amino-4,5-dimethylthiophene-3-carboxylate (10h). General procedure B. After completion of the reaction, as monitored by TLC, the mixture was concentrated under reduced pressure and the crude product was purified by silica gel column chromatography (10:1 petroleum ether:EtOAc) to afford the title compound as a pale yellow solid which was recrystallised from *i*-PrOH (3.41 g, 38%). LCMS (m/z): 200.1 [M + H]⁺. HPLC: t_R 7.265 min. ¹H NMR (CDCl₃) δ 5.90 (s, 2H), 4.27 (q, J = 7.1 Hz, 2H), 2.16 (d, J = 0.7 Hz, 3H), 2.15 (d, J = 0.7 Hz, 2H), 1.34 (t, J = 7.1 Hz, 3H). ¹³C NMR (CDCl₃) δ 166.2, 161.0, 130.4, 113.9, 107.0, 59.7, 14.9, 14.5, 12.4.

Ethyl 2-amino-5-cyclohexylthiophene-3-carboxylate (10i). General procedure B. After completion of the reaction, as monitored by TLC, the mixture was concentrated under reduced pressure and the crude product was chromatographed on silica (10:1 petroleum ether:EtOAc) to afford the title compound as a gold oil (3.05

g, 73%). LCMS (m/z): 254.2 $[M + H]^+$. HPLC: t_R 8.918 min. 1H NMR ($CDCl_3$) δ 6.62 (d, $J = 1.1$ Hz, 1H), 5.79 (s, 2H), 4.25 (q, $J = 7.1$ Hz, 2H), 2.57 – 2.48 (m, 1H), 2.00 – 1.91 (m, 2H), 1.82 – 1.76 (m, 2H), 1.72 – 1.65 (m, 1H), 1.36 – 1.29 (m, 7H), 1.23 – 1.13 (m, 1H). ^{13}C NMR ($CDCl_3$) δ 165.6, 160.9, 133.3, 119.1, 106.2, 59.6, 39.0, 34.9, 26.4, 26.0, 14.6.

Ethyl 2-aminothiophene-3-carboxylate (10j).³⁴ Triethylamine (7.34 mL, 52.6 mmol) was added dropwise over 10 min to a mixture of 2,5-dihydroxy-1,4-dithiane (8.09 g, 52.6 mmol), ethyl cyanoacetate (11.2 mL, 105 mmol), and dimethylformamide (40 mL). The mixture was stirred for at 45 °C for 30 min, diluted with 0.4 M acetic acid and extracted with ether. The ethereal layer was dried over sodium sulphate. The residue was eluted through a silica gel column (10:1 petroleum ether:EtOAc) and the resulting fractions collected, concentrated under reduced pressure and cooled at 4 °C to afford the compound as a light yellow crystalline solid (5.00 g, 55%). LCMS (m/z): 172.1 $[M + H]^+$. HPLC: t_R 6.123 min. 1H NMR ($CDCl_3$) δ 6.98 (d, $J = 5.8$ Hz, 1H), 6.18 (d, $J = 5.8$ Hz, 1H), 4.27 (q, $J = 7.1$ Hz, 2H), 1.34 (t, $J = 7.1$ Hz, 3H). ^{13}C NMR ($CDCl_3$) δ 165.5, 162.7, 126.6, 107.2, 106.9, 59.8, 14.6.

2-(Chloromethyl)-3,5,6,7-tetrahydro-4H-cyclopenta[4,5]thieno[2,3-d]pyrimidin-4-one (11a). General procedure C. The precipitate was collected and washed with cold 1,4-dioxane to afford the compound as a beige solid (2.56 g, 70%). LCMS (m/z): 292.1 $[M + H]^+$. HPLC: t_R 4.216 min. 1H NMR (d_6 -DMSO) δ 12.72 (s, 1H), 4.54 (s, 2H), 2.97 – 2.85 (m, 4H), 2.37 (p, $J = 7.2$ Hz, 2H). ^{13}C NMR (d_6 -DMSO) δ 167.2, 157.9, 151.7, 139.7, 138.7, 119.3, 42.7, 29.1, 28.6, 27.5.

2-(Chloromethyl)-3,5,6,7,8,9-hexahydro-4H-cyclohepta[4,5]thieno[2,3-d]pyrimidin-4-one (11b).

General procedure C. The precipitate was collected and washed with cold 1,4-dioxane to afford the compound as a white powder (5.00 g, 86%). LCMS (m/z): 269.1 $[M + H]^+$. HPLC: t_R 7.039 min. 1H NMR (d_6 -DMSO) δ 12.71 (s, 1H), 4.57 (s, 2H), 3.31 – 3.21 (m, 2H), 2.86 – 2.79 (m, 2H), 1.90 – 1.78 (m, 2H), 1.69 – 1.49 (m, 4H). ^{13}C NMR (d_6 -DMSO) δ 160.4, 158.7, 151.7, 137.6, 136.6, 122.2, 42.6, 32.0, 29.1, 27.3, 27.1, 26.9.

2-(Chloromethyl)-5,6,7,8,9,10-hexahydrocycloocta[4,5]thieno[2,3-d]pyrimidin-4(3H)-one (11c).

General procedure C. The precipitate was collected and washed with cold 1,4-dioxane to afford the compound as a white powder (3.01 g, 78%). LCMS (m/z): 283.0 $[M + H]^+$. HPLC: t_R 7.426 min. 1H NMR (d_6 -DMSO) δ 12.68 (s, 1H), 4.55 (s, 2H), 3.06 – 2.99 (m, 2H), 2.90 – 2.82 (m, 2H), 1.59 (s, 4H), 1.41 – 1.36 (m, 2H), 1.29

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– 1.19 (m, 2H). ^{13}C NMR (d_6 -DMSO) δ 161.6, 158.2, 151.7, 136.1, 133.8, 121.6, 42.6, 31.5, 29.8, 26.1, 25.4, 25.2, 24.3.

2-(Chloromethyl)-5-phenylthieno[2,3-*d*]pyrimidin-4(3*H*)-one (11d). General procedure C. The bright orange suspension was filtered and the resulting solids were washed with cold 1,4-dioxane which afforded the desired compound as a yellow solid (4.33 g, 82%). LCMS (m/z): 277.1 [$\text{M} + \text{H}$] $^+$. HPLC: t_{R} 6.476 min. ^1H NMR (d_6 -DMSO) δ 12.71 (s, 1H), 7.55 (s, 1H), 7.53 (dd, $J = 7.9, 1.3$ Hz, 2H), 7.41 – 7.37 (m, 2H), 7.35 (dd, $J = 6.6, 2.7$ Hz, 1H). ^{13}C NMR (d_6 -DMSO) δ 165.3, 157.8, 153.1, 138.5, 135.1, 129.4, 127.6, 127.5, 121.8, 119.9, 42.5.

2-(Chloromethyl)-6-phenylthieno[2,3-*d*]pyrimidin-4(3*H*)-one (11e). General procedure C. The precipitate was collected from the crude reaction mixture and washed with hot *i*-PrOH to afford the compound as a white solid which was used without the need for further purification (4.57 g, 64%). LCMS (m/z): 277.1 [$\text{M} + \text{H}$] $^+$. HPLC: t_{R} 6.752 min. ^1H NMR (d_6 -DMSO) δ 12.72 (1H), 7.80 (s, 1H), 7.78 – 7.73 (m, 2H), 7.45 (dd, $J = 8.1, 6.7$ Hz, 2H), 7.38 – 7.34 (m, 1H), 4.62 (s, 2H). ^{13}C NMR (d_6 -DMSO) δ 162.9, 157.6, 153.2, 140.4, 132.6, 129.3, 128.8, 125.8, 125.1, 117.3, 42.6.

6-Benzyl-2-(chloromethyl)thieno[2,3-*d*]pyrimidin-4(3*H*)-one (11f). General procedure C. The solvents were removed under reduced pressure and the residue was chromatographed on silica (40% petroleum ether in EtOAc). The collected fractions were concentrated under reduced pressure and the resulting solid was recrystallised from CHCl_3 affording the compound as white-yellow crystals (2.74 g, quantitative yield). LCMS (m/z): 291.1 [$\text{M} + \text{H}$] $^+$. HPLC: t_{R} 6.680 min. ^1H NMR (d_6 -DMSO) δ 7.62 (s, 1H), 7.35 – 7.28 (m, 4H), 7.25 (dd, $J = 11.1, 4.3$ Hz, 1H), 7.16 (s, 1H), 4.54 (s, 2H), 4.20 (s, 2H). ^{13}C NMR (d_6 -DMSO) δ 163.1, 157.6, 152.5, 142.9, 139.5, 128.7, 128.6, 123.7, 119.1, 42.7, 35.6.

2-(Chloromethyl)-5-cyclohexylthieno[2,3-*d*]pyrimidin-4(3*H*)-one (11g). General procedure C. The precipitate was collected from the crude reaction mixture and washed with 1,4-dioxane. This afforded the desired compound as a yellow solid which was used without the need for further purification (2.75 g, 70%). LCMS (m/z): 283.1 [$\text{M} + \text{H}$] $^+$. HPLC: t_{R} 7.595 min. ^1H NMR (d_6 -DMSO) δ 12.73 (s, 1H), 7.21 (s, 1H), 4.57 (s, 2H), 3.26 (t, $J = 9.6$ Hz, 1H), 1.95 (d, $J = 9.8$ Hz, 2H), 1.77 (d, $J = 11.6$ Hz, 2H), 1.70 (d, $J = 12.9$ Hz, 1H),

1.41 – 1.28 (m, 4H), 1.22 (dt, $J = 24.7, 9.6$ Hz, 1H). ^{13}C NMR (d_6 -DMSO) δ 164.6, 158.3, 152.6, 144.8, 120.9, 116.6, 42.5, 37.9, 33.1, 26.3, 25.8.

2-(Chloromethyl)-5,6-dimethylthieno[2,3-*d*]pyrimidin-4(3*H*)-one (11h). General procedure C. The precipitate was collected from the crude reaction mixture, and washed with hot CHCl_3 to afford the compound as a yellow solid (3.38 g, 84%). LCMS (m/z): 229.0 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 5.717 min. ^1H NMR (d_6 -DMSO) δ 12.69 (s, 1H), 4.55 (s, 2H), 2.36 (d, $J = 0.8$ Hz, 3H), 2.34 (d, $J = 0.7$ Hz, 1H). ^{13}C NMR (d_6 -DMSO) δ 161.2, 158.4, 151.9, 130.4, 128.8, 122.4, 42.6, 12.7, 12.7.

2-(Chloromethyl)-6-cyclohexylthieno[2,3-*d*]pyrimidin-4(3*H*)-one (11i). General procedure C. The precipitate was collected from the crude reaction under vacuum filtration and washed with cold 1,4-dioxane. The remaining filtrate was concentrated reduced under pressure and the resulting residue chromatographed on silica (40:60 EtOAc:petroleum ether) to afford the compound as a yellow solid (2.56 g, 64%). LCMS (m/z): 283.0 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 7.614 min. ^1H NMR (d_6 -DMSO) δ 12.67 (s, 1H), 7.09 (d, $J = 0.5$ Hz, 1H), 4.57 (s, 2H), 2.89 – 2.78 (m, 1H), 1.97 (d, $J = 8.5$ Hz, 2H), 1.76 (d, $J = 2.3$ Hz, 2H), 1.65 (d, $J = 12.5$ Hz, 1H), 1.46 – 1.28 (m, 4H), 1.24 – 1.15 (m, 1H). ^{13}C NMR (d_6 -DMSO) δ 161.9, 157.7, 152.3, 149.8, 123.6, 116.1, 42.7, 38.9, 34.5, 25.7, 25.3.

2-(Chloromethyl)thieno[2,3-*d*]pyrimidin-4(3*H*)-one (11j). General procedure C. The reaction mixture was concentrated under reduced pressure and the residue chromatographed on silica (50:50 petroleum ether:EtOAc) which afforded the compound as a pale white solid (1.58 g, 21%). LCMS (m/z): 201.0 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 4.394 min. ^1H NMR (d_6 -DMSO) δ 7.62 (d, $J = 5.8$ Hz, 1H), 7.40 (d, $J = 5.8$ Hz, 1H), 4.58 (s, 2H). ^{13}C NMR (d_6 -DMSO) δ 163.8, 157.9, 152.8, 124.8, 123.6, 121.6, 42.7.

2-(Morpholinomethyl)-3,5,6,7-tetrahydro-4*H*-cyclopenta[4,5]thieno[2,3-*d*]pyrimidin-4-one (12a). General Procedure D. The residue was chromatographed on silica (1:99 $\text{Et}_3\text{N}:\text{EtOAc}$) affording the compound as a beige solid (2.52 g, 86%). LCMS (m/z): 292.1 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 4.216 min. HRMS (m/z): $\text{C}_{14}\text{H}_{17}\text{N}_3\text{O}_2\text{S}$ requires 292.112 $[\text{M} + \text{H}]^+$; found 292.114. ^1H NMR (CDCl_3) δ 10.14 (s, 1H), 3.80 – 3.73 (m, 4H), 3.57 (s, 2H), 3.09 – 3.02 (m, 2H), 2.94 (t, $J = 7.3$ Hz, 2H), 2.63 – 2.57 (m, 4H), 2.51 – 2.40 (m, 2H). ^{13}C NMR (CDCl_3) δ 168.6, 158.2, 152.2, 140.1, 138.7, 119.5, 66.7, 60.6, 53.6, 29.5, 28.9, 27.

2-(Morpholinomethyl)-3,5,6,7,8,9-hexahydro-4H-cyclohepta[4,5]thieno[2,3-d]pyrimidin-4-one (12b).

General Procedure D. A precipitate was collected collected and recrystallised from $\text{CHCl}_3/i\text{-PrOH}$ affording the compound as a white fluffy solid (3.29 g, 83%). LCMS (m/z): 320.1 $[\text{M} + \text{H}]^+$. HPLC: t_R 5.186 min. HRMS (m/z): $\text{C}_{16}\text{H}_{21}\text{N}_3\text{O}_2\text{S}$ requires 320.1433 $[\text{M} + \text{H}]^+$; found 320.1427. ^1H NMR (CDCl_3) δ 10.09 (s, 1H), 3.83 – 3.70 (m, 4H), 3.55 (s, 2H), 3.37 – 3.25 (m, 2H), 2.88 – 2.78 (m, 2H), 2.60 (s, 4H), 1.89 (ddd, $J = 8.3, 7.5, 4.3$ Hz, 2H), 1.74 – 1.63 (m, 4H). ^{13}C NMR (CDCl_3) δ 161.8, 158.9, 137.8, 137.2, 130.1, 122.8, 66.8, 60.7, 53.6, 32.6, 30.1, 28.1, 27.8, 27.4.

2-(Morpholinomethyl)-5,6,7,8,9,10-hexahydrocycloocta[4,5]thieno[2,3-d]pyrimidin-4(3H)-one (12c).

General Procedure D. A precipitate was collected and recrystallised from $\text{CHCl}_3/i\text{-PrOH}$ to afford the compound as a white fluffy solid (1.75 g, 63%). LCMS (m/z): 334.1 $[\text{M} + \text{H}]^+$. HPLC: t_R 5.489 min. HRMS (m/z): $\text{C}_{17}\text{H}_{23}\text{N}_3\text{O}_2\text{S}$ requires 334.1592 $[\text{M} + \text{H}]^+$; found 334.1584. ^1H NMR (CDCl_3) δ 10.17 (s, 1H), 3.81 – 3.71 (m, 4H), 3.54 (s, 2H), 3.13 – 3.05 (m, 2H), 2.90 – 2.81 (m, 2H), 2.67 – 2.51 (m, 4H), 1.78 – 1.61 (m, 4H), 1.45 (dd, $J = 9.9, 4.7$ Hz, 2H), 1.37 – 1.25 (m, 2H). ^{13}C NMR (CDCl_3) δ 163.1, 158.6, 152.4, 136.3, 134.4, 122.2, 66.9, 60.8, 53.7, 31.9, 30.3, 27.1, 26.1, 25.8, 25.2.

2-(Morpholinomethyl)-6-phenylthieno[2,3-d]pyrimidin-4(3H)-one (12d). General Procedure D. Upon cooling of the reaction mixture, a precipitate formed which was collected under vacuum and washed with cold $i\text{-PrOH}$ affording the compound as a white solid (2.71 g, 91%). LCMS (m/z): 328.1 $[\text{M} + \text{H}]^+$. HPLC: t_R 5.092 min. HRMS (m/z): $\text{C}_{17}\text{H}_{17}\text{N}_3\text{O}_2\text{S}$ requires 328.1118 $[\text{M} + \text{H}]^+$; found 328.1114. ^1H NMR ($d_6\text{-DMSO}$) δ 10.85 (s, 1H), 7.78 (s, 1H), 7.78 – 7.73 (m, 2H), 7.46 (dd, $J = 8.2, 6.8$ Hz, 2H), 7.41 – 7.34 (m, 2H), 3.64 – 3.55 (m, 4H), 3.48 (s, 2H), 2.50 (dd, $J = 6.1, 3.0$ Hz, 4H). ^{13}C NMR ($d_6\text{-DMSO}$) δ 163.3, 157.8, 155.1, 139.3, 132.8, 129.3, 128.6, 125.8, 124.7, 117.1, 66.1, 60.4, 53.0.

2-(Morpholinomethyl)-5-phenylthieno[2,3-d]pyrimidin-4(3H)-one (12e). General Procedure D. A precipitate was collected under vacuum and recrystallised from $\text{CHCl}_3/i\text{-PrOH}$ to afford the desired compound as bright yellow crystals (2.67 g, 83%). LCMS (m/z): 328.1 $[\text{M} + \text{H}]^+$. HPLC: t_R 4.853 min. HRMS (m/z): $\text{C}_{17}\text{H}_{17}\text{N}_3\text{O}_2\text{S}$ requires 328.1123 $[\text{M} + \text{H}]^+$; found 328.1114. ^1H NMR (CDCl_3) δ 10.61 (s, 1H), 7.58 – 7.54 (m, 2H), 7.44 – 7.35 (m, 3H), 7.10 (s, 1H), 3.71 – 3.66 (m, 4H), 3.53 (s, 2H), 2.56 – 2.50 (m, 4H). ^{13}C NMR (CDCl_3) δ 166.7, 158.4, 154.2, 139.6, 135.4, 129.5, 127.9, 127.9, 120.3, 66.8, 60.7, 53.6.

6-Benzyl-2-(morpholinomethyl)thieno[2,3-*d*]pyrimidin-4(3*H*)-one (12f). General Procedure D. The residue was recrystallised from CHCl₃/*i*-PrOH to afford the compound as beige crystals (2.47 g, 78%). LCMS (*m/z*): 342.1 [M + H]⁺. HPLC: *t*_R 5.208 min. HRMS (*m/z*): C₁₈H₁₉N₃O₂S requires 342.1278 [M + H]⁺; found 342.1271. ¹H NMR (CDCl₃) δ 10.09 (s, 1H), 7.35 – 7.30 (m, 4H), 7.26 (dt, *J* = 5.0, 1.4 Hz, 3H), 7.13 (t, *J* = 1.1 Hz, 1H), 4.15 (s, 2H), 3.78 – 3.74 (m, 4H), 3.55 (s, 2H), 2.61 – 2.54 (m, 4H). ¹³C NMR (CDCl₃) δ 164.8, 157.8, 153.2, 142.4, 138.8, 128.9, 128.8, 127.1, 123.9, 119.2, 66.8, 60.7, 53.7, 36.9.

5-Cyclohexyl-2-(morpholinomethyl)thieno[2,3-*d*]pyrimidin-4(3*H*)-one (12g). General Procedure D. *i*-PrOH was introduced to a cooled reaction mixture and the resulting precipitate collected to afford the compound as bright yellow crystals (1.56 g, 63% yield). LCMS (*m/z*): 334.1 [M + H]⁺. HPLC: *t*_R 5.715 min. HRMS (*m/z*): C₁₇H₂₃N₃O₂S requires 334.1587 [M + H]⁺; found 334.1584. ¹H NMR (CDCl₃) δ 10.15 (s, 1H), 6.82 (s, 1H), 3.79 – 3.72 (m, 4H), 3.56 (s, 2H), 3.35 (tt, *J* = 11.6, 2.8 Hz, 1H), 2.62 – 2.56 (m, 4H), 2.05 (d, *J* = 11.7 Hz, 2H), 1.86 – 1.71 (m, 3H), 1.54 – 1.41 (m, 2H), 1.33 (tt, *J* = 7.3, 3.5 Hz, 2H), 1.29 – 1.18 (m, 1H). ¹³C NMR (CDCl₃) δ 166.1, 158.5, 153.3, 146.0, 121.3, 115.2, 66.9, 60.6, 53.7, 38.7, 33.8, 26.8, 26.4.

5,6-Dimethyl-2-(morpholinomethyl)thieno[2,3-*d*]pyrimidin-4(3*H*)-one (12h). General Procedure D. Upon cooling of the reaction mixture, a formed precipitate was collected and washed with petroleum ether to afford the desired compound as a bright yellow solid (2.74 g, 88%). LCMS (*m/z*): 280.2 [M + H]⁺. HPLC: *t*_R 4.198 min. HRMS (*m/z*): C₁₃H₁₇N₃O₂S requires 280.1119 [M + H]⁺; found 280.1114. ¹H NMR (CDCl₃) δ 10.13 (s, 1H), 3.79 – 3.71 (m, 4H), 3.53 (s, 2H), 2.63 – 2.56 (m, 4H), 2.46 (d, *J* = 0.6 Hz, 3H), 2.37 (s, 3H). ¹³C NMR (CDCl₃) δ 162.7, 158.7, 152.6, 130.3, 129.5, 122.9, 66.9, 60.7, 53.7, 13.2, 13.1.

6-Cyclohexyl-2-(morpholinomethyl)thieno[2,3-*d*]pyrimidin-4(3*H*)-one (12i). General Procedure D. The solids were filtered from the cooled reaction mixture and recrystallised from *i*-PrOH to afford the compound as white fluffy crystals (2.57 g, 60%). LCMS (*m/z*): 334.2 [M + H]⁺. HPLC: *t*_R 5.603 min. HRMS (*m/z*): C₁₇H₂₃N₃O₂S requires 334.1593 [M + H]⁺; found 334.1584. ¹H NMR (CDCl₃) δ 10.17 (s, 1H), 7.13 (d, *J* = 1.0 Hz, 1H), 3.79 – 3.71 (m, 4H), 3.55 (s, 2H), 2.83 – 2.73 (m, 1H), 2.61 – 2.53 (m, 4H), 2.10 – 2.02 (m, 2H), 1.83 (dd, *J* = 9.9, 2.9 Hz, 2H), 1.73 (ddd, *J* = 9.6, 5.6, 1.5 Hz, 1H), 1.50 – 1.32 (m, 4H), 1.30 – 1.18 (m, 1H). ¹³C NMR (CDCl₃) δ 163.5, 158.0, 152.8, 150.2, 123.7, 115.8, 66.9, 60.8, 53.7, 39.9, 34.9, 26.3, 25.9.

1 **2-((Morpholinomethyl)thieno[2,3-*d*]pyrimidin-4(3*H*)-one (12j).** General Procedure D. The residue was
2 chromatographed on silica (1:99 Et₃N:EtOAc) which afforded the compound as a gold solid (1.10 g, 58%).
3
4 LCMS (*m/z*): 252.1 [M + H]⁺. HPLC: *t*_R 3.009 min. HRMS (*m/z*): C₁₁H₁₃N₃O₂S requires 252.0803 [M + H]⁺;
5
6 found 252.0801. ¹H NMR (CDCl₃) δ 10.28 (s, 1H), 7.44 (d, *J* = 5.8 Hz, 1H), 7.21 (d, *J* = 5.8 Hz, 1H), 3.78 –
7
8 3.73 (m, 4H), 3.57 (s, 2H), 2.63 – 2.55 (m, 4H). ¹³C NMR (CDCl₃) δ 165.3, 158.2, 153.8, 123.7, 123.1, 121.9,
9
10 66.8, 60.7, 53.7.
11

12
13 **4-((4-Chloro-6,7-dihydro-5*H*-cyclopenta[4,5]thieno[2,3-*d*]pyrimidin-2-yl)methyl)morpholine (13a).**

14
15 General Procedure E. The residue was chromatographed on silica (1:99 Et₃N:EtOAc) to afford the compound
16
17 as a brown oil (270 mg, 67%). LCMS (*m/z*): 310.1 [M + H]⁺. HPLC: *t*_R 5.230 min. ¹H NMR (CDCl₃) δ 3.85
18
19 (s, 1H), 3.78 – 3.75 (m, 2H), 3.17 – 3.11 (m, 2H), 3.07 – 3.02 (m, 2H), 2.63 – 2.56 (m, 4H), 2.56 – 2.47 (m,
20
21 1H). ¹³C NMR (CDCl₃) δ 174.3, 160.5, 152.9, 144.6, 136.2, 124.6, 66.9, 64.9, 53.8, 30.3, 29.5, 27.5.
22
23

24
25 **4-((4-Chloro-6,7,8,9-tetrahydro-5*H*-cyclohepta[4,5]thieno[2,3-*d*]pyrimidin-2-yl)methyl)morpholine**

26
27 **(13b).** General Procedure E. The residue was chromatographed on silica (1:99 Et₃N:EtOAc) to afford the
28
29 compound as a gold oil (2.62 g, 70%). LCMS (*m/z*): 338.1 [M + H]⁺. HPLC: *t*_R 5.949 min. ¹H NMR (CDCl₃)
30
31 δ 3.61 (s, 2H), 3.56 – 3.49 (m, 2H), 3.14 – 3.03 (m, 2H), 2.77 – 2.66 (m, 2H), 2.42 – 2.34 (m, 4H), 1.69 (dd,
32
33 *J* = 11.3, 5.9 Hz, 2H), 1.51 (ddd, *J* = 10.7, 9.4, 5.4 Hz, 4H). ¹³C NMR (CDCl₃) δ 167.8, 159.4, 152.5, 143.1,
34
35 131.7, 126.7, 66.3, 64.1, 53.3, 31.4, 29.6, 27.6, 26.5, 26.0.
36
37

38
39 **4-((4-Chloro-5,6,7,8,9,10-hexahydrocycloocta[4,5]thieno[2,3-*d*]pyrimidin-2-yl)methyl)morpholine**

40
41 **(13c).** General Procedure E. The residue was chromatographed on silica (1:99 Et₃N:EtOAc) to afford the
42
43 compound as a gold oil (1.91 g, 67%). LCMS (*m/z*): 352.1 [M + H]⁺. HPLC: *t*_R 6.259 min. ¹H NMR (CDCl₃)
44
45 δ 3.44 (s, 2H), 3.40 – 3.25 (m, 4H), 2.75 – 2.65 (m, 2H), 2.61 – 2.48 (m, 2H), 2.26 – 2.15 (m, 4H), 1.45 – 1.24
46
47 (m, 4H), 1.08 (dd, *J* = 9.5, 4.8 Hz, 2H), 0.91 – 0.84 (m, 2H). ¹³C NMR (CDCl₃) δ 168.2, 159.4, 152.0, 141.3,
48
49 128.4, 125.8, 65.9, 63.8, 53.0, 30.8, 29.4, 27.2, 25.5, 24.5, 24.1.
50
51

52
53 **4-((4-Chloro-6-phenylthieno[2,3-*d*]pyrimidin-2-yl)methyl)morpholine (13d).** General Procedure E. The
54
55 residue was recrystallised from CHCl₃/petroleum ether to afford the compound as pale white crystals (2.45 g,
56
57 90%). LCMS (*m/z*): 346.1 [M + H]⁺. HPLC: *t*_R 6.418 min. ¹H NMR (*d*₆-DMSO) δ 8.08 (s, 1H), 7.96 (d, *J* =
58
59
60

6.9 Hz, 2H), 7.58 – 7.48 (m, 3H), 4.67 (s, 2H), 3.89 (s, 4H), 3.60 – 3.26 (m, 4H). ^{13}C NMR (d_6 -DMSO) δ 167.7, 153.5, 145.9, 131.7, 130.2, 130.1, 129.5, 126.8, 114.9, 63.4, 59.9, 51.8.

4-((4-Chloro-5-phenylthieno[2,3-*d*]pyrimidin-2-yl)methyl)morpholine (13e). General Procedure E. The residue was chromatographed on silica (1:99 Et₃N:EtOAc) which afforded the compound as a gold solid 1.02 g, 96%). LCMS (m/z): 346.1 [M + H]⁺. HPLC: t_R 5.595 min. ^1H NMR (d_6 -DMSO) δ 7.95 (s, 1H), 7.68 – 7.29 (m, 5H), 3.80 (s, 2H), 3.63 – 3.53 (m, 4H), 2.55 – 2.51 (m, 4H). ^{13}C NMR (d_6 -DMSO) δ 169.72, 161.43, 153.8, 134.9, 134.7, 130.0, 128.2, 127.8, 127.5, 124.7, 66.2, 63.7, 53.1.

4-((6-Benzyl-4-chlorothieno[2,3-*d*]pyrimidin-2-yl)methyl)morpholine (13f). General Procedure E. The residue was recrystallised from CHCl₃/*i*-PrOH to afford the compound as pale yellow crystals (1.55 g, 70%). LCMS (m/z): 360.1 [M + H]⁺. HPLC: t_R 6.261 min. ^1H NMR (d_6 -DMSO) δ 7.34 (d, J = 4.4 Hz, 1H), 4.35 (s, 2H), 3.76 (s, 2H), 3.60 – 3.50 (m, 4H), 2.56 – 2.44 (m, 4H). ^{13}C NMR (d_6 -DMSO) δ 168.5, 152.4, 148.2, 138.7, 128.8, 128.7, 127.9, 126.9, 116.8, 114.9, 67.1, 65.9, 52.9, 36.1.

4-((4-Chloro-5-cyclohexylthieno[2,3-*d*]pyrimidin-2-yl)methyl)morpholine (13g). General Procedure E. The residue was chromatographed on silica (1:99 Et₃N:EtOAc) which afforded the compound as a gold oil (1.14 g, 69%). LCMS (m/z): 352.1 [M + H]⁺. HPLC: t_R 6.418 min. ^1H NMR (CDCl₃) δ 7.21 (d, J = 0.7 Hz, 1H), 3.88 (s, 2H), 3.82 – 3.73 (m, 4H), 3.45 – 3.32 (m, 1H), 2.68 – 2.57 (m, 4H), 2.11 (d, J = 12.8 Hz, 2H), 1.87 (d, J = 12.9 Hz, 2H), 1.80 (d, J = 12.9 Hz, 1H), 1.53 – 1.35 (m, 4H), 1.33 – 1.19 (m, 1H). ^{13}C NMR (CDCl₃) δ 171.4, 161.1, 154.5, 141.3, 125.7, 120.9, 66.9, 64.7, 53.9, 38.8, 34.4, 26.8, 26.2.

4-((4-Chloro-5,6-dimethylthieno[2,3-*d*]pyrimidin-2-yl)methyl)morpholine (13h). General Procedure E. The residue was recrystallised from CHCl₃/*i*-PrOH to afford the compound as pale white crystals (1.55 g, 70%). LCMS (m/z): 298.1 [M + H]⁺. HPLC: t_R 4.925 min. ^1H NMR (CDCl₃) δ 3.96 (s, 2H), 3.87 – 3.79 (m, 4H), 2.76 (s, 4H), 2.53 (d, J = 0.5 Hz, 3H), 2.50 (s, 3H). ^{13}C NMR (CDCl₃) δ 168.9, 153.5, 136.4, 128.0, 125.1, 118.4, 66.3, 63.9, 53.4, 14.3, 14.2.

4-((4-Chloro-6-cyclohexylthieno[2,3-*d*]pyrimidin-2-yl)methyl)morpholine (13i). The residue was chromatographed on silica (1:99 Et₃N:EtOAc) affording the compound as a gold oil which later solidified under vacuum to produce a light yellow/white solid (1.64 g, 64%). LCMS (m/z): 352.1 [M + H]⁺. HPLC: t_R 6.663 min. ^1H NMR (CDCl₃) δ 7.05 (d, J = 1.1 Hz, 1H), 3.85 (s, 2H), 3.78 – 3.73 (m, 4H), 2.95 – 2.83 (m,

1H), 2.64 – 2.55 (m, 4H), 2.10 (dd, $J = 13.4, 1.3$ Hz, 2H), 1.91 – 1.83 (m, 2H), 1.80 – 1.71 (m, 1H), 1.57 – 1.35 (m, 4H), 1.32 – 1.21 (m, 1H). ^{13}C NMR (CDCl_3) δ 168.6, 160.9, 155.3, 153.3, 128.7, 113.7, 66.9, 65.0, 53.8, 40.5, 34.8, 26.3, 25.8.

4-((4-Chlorothieno[2,3-*d*]pyrimidin-2-yl)methyl)morpholine (13j). General Procedure E. The residue was chromatographed on silica (1:99 $\text{Et}_3\text{N}:\text{EtOAc}$) which afforded the compound as a gold oil later solidifying under vacuum (1.12 g, 73%). LCMS (m/z): 270.0 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 1.169 min. ^1H NMR (CDCl_3) δ 7.58 (d, $J = 6.0$ Hz, 1H), 7.41 (d, $J = 6.0$ Hz, 1H), 3.91 (s, 2H), 3.81 – 3.74 (m, 4H), 2.70 – 2.60 (m, 4H). ^{13}C NMR (CDCl_3) δ 169.5, 162.1, 155.1, 128.2, 127.9, 119.8, 66.9, 64.9, 53.9.

2-(Morpholinomethyl)-*N*-(3-(trifluoromethyl)phenyl)-6,7-dihydro-5H-

cyclopenta[4,5]thieno[2,3-*d*]pyrimidin-4-amine (14a). General Procedure F. The residue was chromatographed on silica (1:99 $\text{Et}_3\text{N}:\text{EtOAc}$) and the resulting residue recrystallised from CHCl_3 -petroleum ether to afford the title compound as a white powder (127 mg, 52%). LCMS (m/z): 435.1 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 6.750 min. HRMS (m/z): $\text{C}_{21}\text{H}_{21}\text{F}_3\text{N}_4\text{OS}$ requires 435.1467 $[\text{M} + \text{H}]^+$; found 435.1464. ^1H NMR (d_6 -acetone) δ 8.49 (s, 1H), 8.13 (d, $J = 8.2$ Hz, 1H), 7.86 (s, 1H), 7.58 (t, $J = 8.0$ Hz, 1H), 7.40 (d, $J = 7.7$ Hz, 1H), 3.68 (s, 2H), 3.66 – 3.59 (m, 4H), 3.31 (ddd, $J = 5.7, 3.5, 1.7$ Hz, 2H), 3.09 – 2.99 (m, 2H), 2.63 – 2.58 (m, 4H), 2.58 – 2.49 (m, 2H). ^{13}C NMR (d_6 -acetone) δ 173.4, 161.6, 154.7, 141.3, 137.8, 131.1 (q, $J = 32.0$ Hz), 130.2, 125.3 (q, $J = 272.1$ Hz), 125.1, 119.9 (d, $J = 3.9$ Hz), 118.2 (d, $J = 4.1$ Hz), 113.4, 67.5, 65.8, 54.5, 30.1, 29.6, 28.5.

2-(Morpholinomethyl)-*N*-(3-(trifluoromethyl)phenyl)-6,7,8,9-tetrahydro-5Hcyclohepta[4,5]thieno[2,3-*d*]pyrimidin-4-amine (14b). General Procedure F. The residue was chromatographed on silica (1:99 $\text{Et}_3\text{N}:\text{EtOAc}$) to afford the title compound as a gold oil (90.3 mg, 12%). LCMS (m/z): 463.1 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 7.226 min. HRMS (m/z): $\text{C}_{21}\text{H}_{21}\text{F}_3\text{N}_4\text{OS}$ requires 463.1782 $[\text{M} + \text{H}]^+$; found 463.1774. ^1H NMR (CDCl_3) δ 8.10 (s, 1H), 7.75 (dd, $J = 8.1, 1.7$ Hz, 1H), 7.45 (t, $J = 7.9$ Hz, 1H), 7.33 (d, $J = 7.8$ Hz, 1H), 7.22 (s, 1H), 3.76 – 3.73 (m, 4H), 3.74 (s, 2H), 3.16 – 3.08 (m, 2H), 2.93 – 2.89 (m, 2H), 2.67 – 2.59 (m, 4H), 1.98 – 1.78 (m, 4H), 1.88 – 1.79 (m, 2H). ^{13}C NMR (CDCl_3) δ 166.4, 160.1, 154.1, 139.6, 138.9, 131.3 (q, $J = 32.4$ Hz), 129.4, 129.3, 124.1 (q, $J = 272.5$ Hz), 123.6, 119.9 (q, $J = 3.8$ Hz), 117.5 (q, $J = 3.9$ Hz), 116.4, 67.0, 65.1, 53.8, 30.4, 30.3, 29.1, 26.9, 26.4.

2-(Morpholinomethyl)-N-(3-(trifluoromethyl)phenyl)-5,6,7,8,9,10hexahydrocycloocta[4,5]thieno[2,3-d]pyrimidin-4-amine (14c). General Procedure F. The residue was chromatographed on silica (1:99 Et₃N:EtOAc) to afford the compound as gold oil (60.2 mg, 11%). LCMS (*m/z*): 477.2 [M + H]⁺. HPLC: *t_R* 7.442 min. HRMS (*m/z*): C₂₄H₂₇F₃N₄O requires 477.194 [M + H]⁺; found 477.193. ¹H NMR (CDCl₃) δ 8.23 (s, 1H), 7.77 (dd, *J* = 8.1, 1.7 Hz, 1H), 7.46 (t, *J* = 7.9 Hz, 1H), 7.35 (s, 1H), 7.32 (d, *J* = 5.8 Hz, 1H), 3.77 (s, 2H), 3.78 – 3.74 (m, 4H), 3.07 (dd, *J* = 11.8, 5.5 Hz, 2H), 2.95 – 2.88 (m, 2H), 2.69 – 2.62 (m, 4H), 1.92 – 1.81 (m, 2H), 1.78 – 1.69 (m, 2H), 1.60 – 1.51 (m, 2H), 1.39 (dd, *J* = 10.5, 5.3 Hz, 2H). ¹³C NMR (CDCl₃) δ 167.0, 160.3, 153.9, 139.6, 138.3, 131.4 (q, *J* = 32.5 Hz), 129.4, 126.5, 124.1 (d, *J* = 272.5 Hz), 123.45, 119.9 (q, *J* = 3.8 Hz), 117.4 (q, *J* = 3.9 Hz), 115.2, 67.1, 65.3, 53.9, 31.7, 30.1, 28.0, 26.3, 26.2, 25.2.

2-(Morpholinomethyl)-6-phenyl-N-(3-(trifluoromethyl)phenyl)thieno[2,3-d]pyrimidin-4-amine (14d). General Procedure F. The residue was chromatographed on silica (1:99 Et₃N:EtOAc) to afford the title compound as a gold solid (244 mg, 51%). LCMS (*m/z*): 471.1 [M + H]⁺. HPLC: *t_R* 7.124 min. HRMS (*m/z*): C₂₄H₂₁F₃N₄OS requires 471.1467 [M + H]⁺; found 471.1461. ¹H NMR (*d*₆-DMSO) δ 9.88 (s, 1H), 8.58 (s, 1H), 8.25 (s, 1H), 8.18 (d, *J* = 8.18 Hz, 1H), 7.73 (d, *J* = 7.5 Hz, 2H), 7.62 (t, *J* = 8.0 Hz, 1H), 7.53 (t, *J* = 7.6 Hz, 2H), 7.43 (t, *J* = 7.8 Hz, 2H), 3.66 (s, 2H), 3.62 – 3.57 (m, 4H), 2.58 – 2.52 (m, 4H). ¹³C NMR (*d*₆-DMSO) δ 166.6, 161.5, 153.8, 140.5, 139.2, 133.0, 129.7, 129.4, 129.4 (d, *J* = 31.4 Hz), 128.8, 125.8, 124.3 (q, *J* = 272.4 Hz), 123.7, 118.9 (d, *J* = 3.8 Hz), 116.8, 116.6 (d, *J* = 4.1 Hz), 114.9, 66.2, 64.8, 53.3.

2-(Morpholinomethyl)-5-phenyl-N-(3-(trifluoromethyl)phenyl)thieno[2,3-d]pyrimidin-4-amine (14e). General Procedure F. The residue was chromatographed on silica (1:29:70 Et₃N:petroleum ether:EtOAc) to afford the compound as a gold oil (307 mg, 75%). LCMS (*m/z*): 471.1 [M + H]⁺. HPLC: *t_R* 7.195 min. HRMS (*m/z*): C₂₄H₂₁F₃N₄OS requires 471.1465 [M + H]⁺; found 471.1461. ¹H NMR (CDCl₃) δ 8.08 (s, 1H), 7.60 (dd, *J* = 6.4, 2.5 Hz, 3H), 7.55 (d, *J* = 2.2 Hz, 1H), 7.54 (d, *J* = 3.8 Hz, 1H), 7.34 (t, *J* = 7.9 Hz, 1H), 7.25 (dd, *J* = 13.4, 5.5 Hz, 2H), 7.21 (s, 1H), 7.01 (s, 1H), 3.85 (s, 2H), 3.82 – 3.77 (m, 4H), 2.76 – 2.66 (m, 4H). ¹³C NMR (CDCl₃) δ 168.5, 161.9, 154.6, 139.3, 135.9, 133.77, 131.33 (q, *J* = 32.2 Hz), 129.6, 129.4, 129.3 (d, *J* = 2.8 Hz), 123.3 (q, *J* = 272.2 Hz), 122.3, 121.5, 119.6 (q, *J* = 3.8 Hz), 116.5 (q, *J* = 4.0 Hz), 113.3, 67.0, 65.3, 53.8.

6-Benzyl-2-(morpholinomethyl)-N-(3-(trifluoromethyl)phenyl)thieno[2,3-*d*]pyrimidin-4-amine (14f).

General Procedure F. The reaction precipitate was collected under vacuum and washed with cold *i*-PrOH to afford the compound as a white solid (37.5 mg, 55%). LCMS (m/z): 485.2 [M + H]⁺. HPLC: t_R 7.162 min. HRMS (m/z): C₂₅H₂₃F₃N₄OS requires 485.1624 [M + H]⁺; found 485.1617. ¹H NMR (*d*₆-DMSO) δ 10.39 (s, 1H), 8.36 (d, *J* = 8.5 Hz, 1H), 8.19 (s, 1H), 7.92 (s, 1H), 7.61 (t, *J* = 8.0 Hz, 1H), 7.45 (d, *J* = 7.7 Hz, 1H), 7.39 – 7.33 (m, 4H), 7.30 – 7.25 (m, 1H), 4.52 (s, 2H), 4.30 (s, 2H), 3.89 (s, 4H), 3.40 (s, 4H). ¹³C NMR (*d*₆-DMSO) δ 166.2, 153.9, 142.6, 139.9, 139.2, 129.8, 129.4, 129.1, 128.7, 128.7, 126.8, 125.3, 124.2 (q, *J* = 272.3 Hz), 119.6, 117.6 (d, *J* = 3.9 Hz), 117.4, 116.8, 63.2, 59.8, 51.8, 36.2.

5-Cyclohexyl-2-(morpholinomethyl)-N-(3-(trifluoromethyl)phenyl)thieno[2,3-*d*]pyrimidin-4-amine

(14g). General Procedure F. The residue was chromatographed on silica (1:29:70 Et₃N:petroleum ether:EtOAc) to afford the title compound as a yellow oil which later solidified under vacuum (260 mg, 66%). LCMS (m/z): 477.2 [M + H]⁺. HPLC: t_R 7.618 min. HRMS (m/z): C₂₄H₂₇F₃N₄OS requires 477.1937[M + H]⁺; found 477.193. ¹H NMR (CDCl₃) δ 8.33 (s, 1H), 7.75 (dd, *J* = 8.0, 0.9 Hz, 1H), 7.49 (t, *J* = 7.9 Hz, 1H), 7.37 (d, *J* = 6.6 Hz, 2H), 7.01 (s, 1H), 3.80 (s, 2H), 3.79 – 3.75 (m, 4H), 2.86 (t, *J* = 9.6 Hz, 1H), 2.74 – 2.62 (m, 4H), 2.23 (d, *J* = 10.4 Hz, 2H), 2.04 – 1.98 (m, 2H), 1.89 (d, *J* = 12.9 Hz, 1H), 1.64 – 1.47 (m, 4H), 1.45 – 1.31 (m, 1H). ¹³C NMR (CDCl₃) δ 169.7, 161.2, 154.7, 139.6, 138.8, 131.6 (q, *J* = 32.3 Hz), 129.6, 124.2 (q, *J* = 272.4 Hz), 123.2, 120.1 (q, *J* = 3.8 Hz), 117.3 (q, *J* = 4.0 Hz), 117.2, 114.1, 67.1, 65.2, 53.9, 41.3, 34.2, 26.9, 26.1.

5,6-Dimethyl-2-(morpholinomethyl)-N-(3-(trifluoromethyl)phenyl)thieno[2,3-*d*]pyrimidin-4-amine

(14h). General Procedure F. The residue was chromatographed on silica (1:29:70 Et₃N:petroleum ether:EtOAc) to afford the title compound as a gold oil (214 mg, 40%). LCMS (m/z): 423.2 [M + H]⁺. HPLC: t_R 6.577 min. HRMS (m/z): C₂₀H₂₁F₃N₄OS requires 423.1464 [M + H]⁺; found 423.1461. ¹H NMR (CDCl₃) δ 8.20 (s, 1H), 7.81 (dd, *J* = 8.1, 1.2 Hz, 1H), 7.46 (t, *J* = 8.0 Hz, 1H), 7.38 (s, 1H), 7.34 (d, *J* = 7.8 Hz, 1H), 3.78 – 3.75 (m, 6H), 2.68 – 2.63 (m, 4H), 2.60 (s, 3H), 2.47 (s, 3H). ¹³C NMR (CDCl₃) δ 166.8, 160.5, 154.3, 139.5, 131.9, 131.4 (q, *J* = 32.3 Hz), 129.5, 123.8 (q, *J* = 272.2 Hz), 123.5 (d, *J* = 1.2 Hz), 120.0 (q, *J* = 3.8 Hz), 117.4 (q, *J* = 4.0 Hz), 116.1, 67.1, 65.3, 53.9, 14.6, 13.7.

6-Cyclohexyl-2-(morpholinomethyl)-N-(3-(trifluoromethyl)phenyl)thieno[2,3-*d*]pyrimidin-4-amine

(14i). General Procedure F. The residue was chromatographed on silica (1:99 Et₃N:EtOAc) to afford the compound as a white solid (258 mg, 54 %). LCMS (*m/z*): 477.2 [M + H]⁺. HPLC: *t_R* 7.666 min. HRMS (*m/z*): C₂₄H₂₇F₃N₄OS requires 477.1942 [M + H]⁺; found 477.193. ¹H NMR (400 MHz, *d*₆-DMSO) δ 9.72 (s, 1H), 8.57 (s, 1H), 8.17 (d, *J* = 8.3 Hz, 1H), 7.62 (d, *J* = 0.9 Hz, 1H), 7.59 (d, *J* = 8.1 Hz, 1H), 7.39 (d, *J* = 7.7 Hz, 1H), 3.64 (s, 2H), 3.61 – 3.56 (m, 4H), 2.90 (tt, *J* = 10.8, 5.5 Hz, 1H,A), 2.56 – 2.51 (m, 4H), 2.07 (d, *J* = 9.7 Hz, 2H), 1.81 (dd, *J* = 9.3, 3.0 Hz, 2H), 1.71 (d, *J* = 12.7 Hz, 1H), 1.51 – 1.35 (m, 4H), 1.26 (ddd, *J* = 17.7, 10.6, 6.0 Hz, 1H). ¹³C NMR (*d*₆-DMSO) δ 165.9, 160.5, 153.4, 148.5, 140.7, 129.7, 129.3 (d, *J* = 31.4 Hz), 124.3 (q, *J* = 272.1 Hz), 123.6, 118.7 (d, *J* = 3.8 Hz), 116.5 (d, *J* = 4.0 Hz), 115.6, 113.5, 66.2, 64.7, 53.2, 39.1, 34.4, 25.6, 25.4.

2-(Morpholinomethyl)-N-(3-(trifluoromethyl)phenyl)thieno[2,3-*d*]pyrimidin-4-amine (14j). General

Procedure F. The precipitate was filtered and washed with cold *i*-PrOH affording the compound as a brown solid (199 mg, 45%). LCMS (*m/z*): 395.1 [M + H]⁺. HPLC: *t_R* 5.894 min. HRMS (*m/z*): C₁₈H₁₇F₃N₄OS requires 395.1106 [M + H]⁺; found 395.1148. ¹H NMR (*d*₆-DMSO) δ 10.68 (s, 1H), 8.43 (d, *J* = 8.5 Hz, 1H), 8.29 (d, *J* = 5.9 Hz, 2H), 7.82 (d, *J* = 6.0 Hz, 1H), 7.62 (t, *J* = 8.0 Hz, 1H), 7.46 (d, *J* = 7.7 Hz, 1H), 4.56 (s, 2H), 3.92 (s, 4H), 3.40 (s, 4H). ¹³C NMR (*d*₆-DMSO) δ 166.6, 154.7, 139.9, 129.7, 129.2 (d, *J* = 31.6 Hz), 128.8, 125.4, 124.7, 124.2 (q, *J* = 272.4 Hz), 120.3, 119.7 (d, *J* = 3.9 Hz), 117.7 (d, *J* = 3.9 Hz), 116.4, 63.3, 59.8, 51.9

6-Benzyl-N,N-dimethyl-2-(morpholinomethyl)thieno[2,3-*d*]pyrimidin-4-amine (15). General Procedure

F. The residue was chromatographed on silica (1:99 Et₃N:EtOAc) to afford the compound as a gold oil (91 mg, 33%). LCMS (*m/z*): 369.2 [M + H]⁺. HPLC: *t_R* 6.012 min, 93% purity (214 nm). HRMS (*m/z*): C₂₀H₂₄N₄OS requires 369.1747 [M + H]⁺; found 369.1744. ¹H NMR (CDCl₃) δ 7.30 (dd, *J* = 9.7, 4.8 Hz, 1H), 7.26 – 7.20 (m, 3H), 7.11 (s, 1H), 4.14 (s, 2H), 3.80 – 3.72 (m, 4H), 3.69 (s, 2H), 3.32 (s, 6H), 2.71 – 2.60 (m, 1H). ¹³C NMR (CDCl₃) δ 169.6, 160.7, 160.0, 157.6, 139.1, 138.2, 128.6, 128.5, 126.7, 118.7, 114.5, 66.8, 65.0, 53.6, 39.8, 36.9.

2-(Morpholinomethyl)-N-phenyl-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4-amine (16a).

General Procedure F. The residue was eluted through a silica gel column (99:1 EtOAc:Et₃N) and proceeded

to afford the title compound as a light pink oil which later solidified at room temperature (48.8 mg, 20%). mp: 129-135 °C. LCMS (m/z): 381.2 [M + H]⁺. HPLC: t_R 6.294 min. HRMS (m/z): C₂₁H₂₄N₄OS requires 381.1748 [M + H]⁺; found 381.1744. ¹H NMR (CDCl₃) δ 7.76 – 7.72 (m, 2H), 7.39 – 7.33 (m, 2H), 7.17 (s, 1H), 7.13 – 7.07 (m, 1H), 3.80 – 3.77 (m, 4H), 3.76 (s, 2H), 3.10 – 3.02 (m, J = 6.0 Hz, 2H), 2.88 – 2.78 (m, 2H), 2.69 – 2.62 (m, 4H), 2.02 – 1.96 (m, 2H), 1.96 – 1.89 (m, 2H). ¹³C NMR (CDCl₃) δ 167.0, 160.7, 154.7, 138.9, 134.4, 128.9, 124.4, 123.5, 120.5, 114.9, 67.0, 65.1, 53.9, 26.5, 25.5, 22.6, 22.5.

***N*-(3-Fluorophenyl)-2-(morpholinomethyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4-**

amine (16b). General Procedure F. Chromatographed on silica (1:99 EtOAc:Et₃N) to give the final product as a yellow/orange solid (90.2 mg, 17%). mp: 141-150 °C. LCMS (m/z): 399.2 [M + H]⁺. HPLC: t_R 6.569 min. HRMS (m/z): C₂₁H₂₃FN₄OS requires 399.1654 [M + H]⁺; found 399.1649. ¹H NMR (CDCl₃) δ 7.97 (dt, J = 11.5, 2.1 Hz, 1H), 7.32 – 7.26 (m, 1H), 7.24 – 7.19 (m, 2H), 6.80 (td, J = 8.2, 1.7 Hz, 1H), 3.83 – 3.80 (m, 4H), 3.80 (s, 2H), 3.06 (t, J = 5.7 Hz, 2H), 2.86 (t, J = 5.7 Hz, 2H), 2.72 – 2.65 (m, 4H), 2.02 – 1.97 (m, 2H), 1.97 – 1.91 (m, 2H). ¹³C NMR (CDCl₃) δ 165.8 (d, J = 247.2 Hz), 161.9, 160.7, 154.4, 140.7 (d, J = 11.2 Hz), 134.9, 129.9 (d, J = 9.7 Hz), 124.3, 115.2 (d, J = 2.7 Hz), 115.1, 109.9 (d, J = 21.6 Hz), 107.78 (d, J = 27.1 Hz), 67.1, 65.1, 53.9, 26.5, 25.6, 22.6, 22.5.

***N*-(3-Methoxyphenyl)-2-(morpholinomethyl)-5,6,7,8-tetrahydrobenzo [4,5]thieno[2,3-*d*]pyrimidin-4-**

amine (16c). General Procedure F. This residue was purified using column chromatography (50:50 petroleum ether/EtOAc) to afford the final product as a yellow oil (60.5 mg, 18%). ¹H NMR (CDCl₃) δ 7.65 (s, 1H, H), 7.25 (t, J = 8.1 Hz, 1H), 7.18 (s, 1H), 7.12 (ddd, J = 8.2, 2.1, 0.8 Hz, 1H), 6.65 (ddd, J = 8.2, 2.5, 0.8 Hz, 1H), 3.86 (s, 3H), 3.81 – 3.76 (m, J = 8.3, 3.5 Hz, 6H), 3.06 (t, J = 5.9 Hz, 2H), 2.84 (t, J = 5.9 Hz, 2H), 2.67 (s, 4H), 2.03 – 1.96 (m, 2H), 1.96 – 1.89 (m, 2H). LCMS (m/z): 411.3 [M + H]⁺. HPLC: t_R 6.521 min. HRMS (m/z): C₂₂H₂₆N₄O₂S requires 411.1853 [M + H]⁺; found 411.1849. ¹³C NMR (CDCl₃) δ 167.0, 160.2, 154.6, 140.2, 134.6, 129.6, 124.4, 115.1, 112.5, 108.7, 106.6, 66.9, 65.2, 60.4, 55.3, 53.8, 26.4, 25.5, 22.6, 22.4.

3-((2-(Morpholinomethyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4-yl)amino)phenol (17).

To a solution of *N*-(3-methoxyphenyl)-2-(morpholinomethyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4-amine (195 mg, 475 μmol) in DCM (4.00 mL) at 0 °C was added BBr₃ (1 M in DCM) (2.37 mL, 2.37 mmol). The mixture was stirred at r.t. under a N₂ environment overnight, and then poured into ice-

1 water. The solution was adjusted to pH 6 by addition of sat. NaHCO₃. The DCM was removed under reduced
2 pressure and the aqueous residue extracted with EtOAc (3 x 20 mL). The combined organic extracts were
3 washed with brine, dried (Na₂SO₄), filtered and concentrated *in vacuo*. The resulting residue was eluted
4 through a silica gel column (5:95 MeOH:DCM) to give the title compound as a white/grey solid (100 mg,
5 53%). mp: 98-105°C. LCMS (*m/z*): 397.2 [M + H]⁺. HPLC: *t_R* 5.850 min. HRMS (*m/z*): C₂₁H₂₄N₄O₂S requires
6 397.1699 [M + H]⁺; found 397.1693. ¹H NMR (CDCl₃) δ 8.37 (s, 1H), 7.73 (t, *J* = 2.0 Hz, 1H), 7.14 (s, 1H),
7 7.11 (t, *J* = 8.1 Hz, 1H), 6.87 (ddd, *J* = 8.0, 2.0, 0.7 Hz, 1H), 6.56 (ddd, *J* = 8.1, 2.3, 0.7 Hz, 1H), 3.79 (s, 2H),
8 3.78 – 3.73 (m, 4H), 2.98 (t, *J* = 5.4 Hz, 2H), 2.80 – 2.72 (m, 6H), 1.98 – 1.83 (m, 4H). ¹³C NMR (CDCl₃) δ
9 166.4, 160.3, 157.4, 154.5, 139.8, 134.4, 129.7, 124.6, 115.1, 111.5, 111.1, 108.3, 66.8, 64.0, 52.8, 26.4, 25.5,
10 22.5, 22.4.

23 ***N*-Methyl-2-(morpholinomethyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4-amine (18).**

24 General Procedure F. Upon completion of reaction the reaction vessel was stored at -4 °C overnight.
25 Precipitates were collected and washed with cold *i*-PrOH to afford the compound as light brown crystals (300
26 mg, 76%). LCMS (*m/z*): 319.1[M + H]⁺. HPLC: *t_R* 5.162 min. HRMS (*m/z*): C₁₆H₂₂N₄OS requires 319.1593
27 [M + H]⁺; found 319.1587. ¹H NMR (CDCl₃) δ 5.23 (dd, *J* = 9.4, 5.2 Hz, 1H), 3.82 – 3.74 (m, 4H), 3.71 (s,
28 2H), 3.10 (d, *J* = 4.8 Hz, 3H), 2.92 – 2.85 (m, 2H), 2.82 – 2.75 (m, 2H), 2.70 – 2.65 (m, 4H), 1.98 – 1.81 (m,
29 4H). ¹³C NMR (CDCl₃) δ 165.9, 161.0, 158.0, 132.8, 125.2, 114.6, 67.1, 65.4, 53.9, 28.1, 26.5, 25.5, 22.7.

39 ***N,N*-Diethyl-2-(morpholinomethyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4-amine (19).**

40 General Procedure F. The reaction mixture was concentrated under reduced pressure and the residue
41 chromatographed on silica (5% MeOH-DCM) to afford the compound as a gold oil (299 mg, 90%). LCMS
42 (*m/z*): 361.2 [M + H]⁺. HPLC: *t_R* 5.162 min. HRMS (*m/z*): C₁₉H₂₈N₄OS requires 361.2055 [M + H]⁺; found
43 361.2057. ¹H NMR (CDCl₃) δ 3.71 (t, *J* = 4.7 Hz, 6H), 3.41 (q, *J* = 7.0 Hz, 4H), 2.90 – 2.76 (m, 4H), 2.65 –
44 2.55 (m, 4H), 1.92 – 1.82 (m, 2H), 1.79 – 1.68 (m, 2H), 1.08 (t, *J* = 7.0 Hz, 6H). ¹³C NMR (CDCl₃) δ 168.9,
45 161.7, 159.1, 133.7, 127.4, 120.0, 66.9, 65.0, 53.6, 44.7, 26.6, 25.8, 23.1, 22.9, 12.5.

55 **4-((4-(3-(Trifluoromethyl)phenoxy)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-2-**

56 **yl)methyl)morpholine¹⁶ (20).** A mixture of 4-((4-chloro-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-
57 *d*]pyrimidin-2-yl)methyl)morpholine (350 mg, 1.08 mmol), 3-(trifluoromethyl)phenol (135 μL, 1.11 mmol),
58
59
60

1 K_2CO_3 (299 mg, 2.16 mmol), and tris[2-(2-methoxyethoxy)ethyl]amine (TRIDENT) (34.0 μL , 108 μmol)
2 were taken up in *i*-PrOH and irradiated at 40 W for 6 h in an 8 mL pressure tube. The reaction mixture was
3 diluted with water, and extracted with DCM. The combined organic extracts were dried and concentrated *in*
4 *vacuo*. The residue was chromatographed on silica (1:99 $\text{Et}_3\text{N}:\text{EtOAc}$) to afford the title compound as pale
5 yellow oil (351 mg, 72%). LCMS (m/z): 450.1 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 7.432 min. HRMS (m/z): $\text{C}_{22}\text{H}_{22}\text{F}_3\text{N}_3\text{O}_2\text{S}$
6 requires 450.1468 $[\text{M} + \text{H}]^+$; found 450.1458. ^1H NMR (CDCl_3) δ 7.56 – 7.48 (m, 3H), 7.41 (dt, $J = 7.4, 1.7$
7 Hz, 1H), 3.72 – 3.60 (m, 6H), 3.04 (t, $J = 4.8$ Hz, 2H), 2.85 (t, $J = 4.8$ Hz, 2H), 2.54 – 2.50 (m, 4H), 1.96 –
8 1.85 (m, 4H). ^{13}C NMR (CDCl_3) δ 169.6, 162.2, 160.5, 152.7, 136.2, 131.9 (q, $J = 32.9$ Hz), 130.0, 126.8,
9 125.3 (d, $J = 0.8$ Hz), 123.7 (q, $J = 272.4$ Hz), 122.0 (q, $J = 3.8$ Hz), 119.1 (q, $J = 3.9$ Hz), 117.5, 66.8, 64.3,
10 53.6, 25.8, 25.6, 22.9, 22.4.

11 **5,6,7,8-Tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4(3*H*)-one³³ (21).** Ethyl-2-amino-4,5,6,7-
12 tetrahydrobenzothiophene-3-carboxylate (4.05 g, 17.8 mmol) was taken up in formamide (20 mL) and heated
13 to 170 °C under a nitrogen atmosphere. The heating was continued for 12 h and the progress of the reaction
14 was monitored by TLC. The reaction mixture was allowed to cool to r.t., and the precipitated solids were
15 collected and washed with petroleum spirits to afford the compound as brown crystals (3.12 g, 85%). LCMS
16 (m/z): 207.1 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 5.537 min. HRMS (m/z): $\text{C}_{10}\text{H}_{10}\text{N}_2\text{OS}$ requires 207.0585 $[\text{M} + \text{H}]^+$; found
17 207.0587. ^1H NMR (d_6 -DMSO) δ 7.99 (s, 1H), 2.86 (t, $J = 5.9$ Hz, 2H), 2.73 (t, $J = 5.8$ Hz, 2H), 1.88 – 1.64
18 (m, 4H). ^{13}C NMR (d_6 -DMSO) δ 162.4, 157.6, 144.8, 132.1, 130.8, 122.6, 25.3, 24.4, 22.4, 21.7.

19 **4-Chloro-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidine (22).** General Procedure E. The residue
20 was chromatographed on silica (99:1 $\text{EtOAc}:\text{Et}_3\text{N}$) which afforded the compound as a gold solid (1.85 g,
21 68%). LCMS (m/z): 225.1 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 8.139 min. ^1H NMR (CDCl_3) δ 8.70 (s, 1H), 3.09 (t, $J = 4.3$
22 Hz, 2H), 2.88 (dd, $J = 5.0, 3.5$ Hz, 2H), 1.92 (dt, $J = 6.1, 2.9$ Hz, 4H). ^{13}C NMR (CDCl_3) δ 169.02, 153.3,
23 151.7, 139.7, 128.9, 127.3, 26.4, 26.2, 22.5, 22.3.

24 ***N*-(3-(Trifluoromethyl)phenyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4-amine (23a).**

25 General Procedure F. The crude reaction was concentrated under reduced pressure and the residue
26 chromatographed on silica eluting with (1:29:70 $\text{Et}_3\text{N}:\text{petroleum ether}:\text{EtOAc}$) to afford the compound as an
27 oil which later solidified under vacuum to give a beige solid (265 mg, 68%). LCMS (m/z): 350.1 $[\text{M} + \text{H}]^+$.

HPLC: t_R 8.367 min. HRMS (m/z): $C_{17}H_{14}F_3N_3S$ requires 350.0936 $[M + H]^+$; found 350.0933. 1H NMR ($CDCl_3$) δ 8.52 (s, 1H), 7.96 (s, 1H), 7.91 (d, $J = 8.2$ Hz, 1H), 7.48 (t, $J = 8.0$ Hz, 1H), 7.36 (d, $J = 7.8$ Hz, 1H), 7.22 (s, 1H), 3.08 (t, $J = 6.0$ Hz, 1H), 2.86 (t, $J = 6.0$ Hz, 1H), 2.05 – 1.91 (m, 4H). ^{13}C NMR ($CDCl_3$) δ 166.7, 15.6, 152.4, 139.3, 135.5, 131.5 (d, $J = 32.4$ Hz), 129.64, 126.1 (d, $J = 1.0$ Hz), 124.1 (q, $J = 272.5$ Hz), 120.3 (q, $J = 3.8$ Hz), 117.5 (q, $J = 3.9$ Hz), 116.8, 26.6, 25.6, 22.6, 22.4.

***N,N*-Diethyl-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4-amine (23b).** General procedure F.

The solution was concentrated *in vacuo* and the residue chromatographed on silica (4:1 EtOAc:petroleum ether) which afforded the desired compound as a light gold oil that solidified under high vacuum to give a pale yellow solid (233 mg, 80%). mp^{exp} : 60-62 °C. LCMS (m/z): 262.1 $[M + H]^+$. HPLC: t_R 6.281 min. HRMS (m/z): $C_{14}H_{19}N_3S$ requires 262.1376 $[M + H]^+$; found 262.1372. 1H NMR ($CDCl_3$) δ 8.50 (s, 1H), 3.46 (q, $J = 7.1$ Hz, 4H), 2.96 – 2.82 (m, 4H), 1.97 – 1.90 (m, 2H), 1.84 – 1.76 (m, 2H), 1.13 (t, $J = 7.1$ Hz, 6H). ^{13}C NMR ($CDCl_3$) δ 168.3, 162.1, 151.3, 134.2, 127.7, 122.1, 44.9, 26.6, 25.8, 23.0, 22.9, 12.4.

2-(Piperidin-1-ylmethyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4(3*H*)-one (24).

General procedure D. The reaction mixture was poured into water. The formed precipitates were collected under vacuum filtration and recrystallised from DMF. The mother liquor was extracted with EtOAc, washed with water, brine and the combined organic extracts concentrated *in vacuo*. The residue was chromatographed on silica (1:99 MeOH:EtOAc) to afford the target compound as cream white solid (350 mg, 73%), mp : 182-184 °C. LCMS (m/z): 304.2 $[M + H]^+$. HPLC: t_R 5.228 min. HRMS (m/z): $C_{16}H_{21}N_3OS$ requires 303.1356 $[M + H]^+$; found 303.14. 1H NMR δ 9.95 (s, 1H), 3.46 (s, 2H), 3.04 – 2.97 (m, 2H), 2.77 – 2.73 (m, 2H), 2.54 – 2.44 (m, 4H), 1.92 – 1.78 (m, 4H), 1.62 (m, 4H), 1.48 (d, $J = 5.2$ Hz, 2H). ^{13}C NMR ($CDCl_3$) δ 163.7, 158.3, 153.8, 132.8, 131.5, 121.9, 60.8, 54.8, 25.9, 25.6, 25.1, 23.7, 23.0, 22.3.

4-Chloro-2-(piperidin-1-ylmethyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidine (25).

General Procedure E. The residue was eluted through a silica gel column (99:1 EtOAc:Et₃N) which proceeded to give the compound as a golden oil (154 mg, 41%). LCMS (m/z): 324.1 $[M + H]^+$. HPLC: t_R 6.231 min. HRMS (m/z): $C_{16}H_{20}ClN_3S$ requires 322.114 $[M + H]^+$; found 322.1139. 1H NMR ($CDCl_3$) δ 3.80 (s, 2H), 3.06 (t, $J = 4.2$ Hz, 2H), 2.84 (t, $J = 4.2$ Hz, 2H), 2.52 – 2.49 (m, 4H), 1.93 – 1.86 (m, 4H), 1.67 – 1.57 (m,

4H), 1.49 – 1.36 (m, 2H). ¹³C NMR (CDCl₃) δ 169.4, 161.4, 152.9, 138.9, 126.9, 126.9, 65.2, 54.7, 26.3, 26.0, 25.7, 24.1, 22.5, 22.2.

2-(Piperidin-1-ylmethyl)-N-(3-(trifluoromethyl)phenyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-d]pyrimidin-4-amine (26). General Procedure F. This residue was purified on silica gel (99:1 EtOAc:Et₃N) to give the final product as a yellow oil which later solidified at room temperature (100 mg, 24%). mp: 121–126 °C. LCMS (*m/z*): 447.3 [M + H]⁺. HPLC: *t*_R 7.602 min. HRMS (*m/z*): C₂₃H₂₅F₃N₄S requires 447.1828 [M + H]⁺; found 447.1825. ¹H NMR δ 8.17 (s, 1H), 7.93 (dd, *J* = 8.1, 1.6 Hz, 1H), 7.48 (t, *J* = 8.0 Hz, 1H), 7.34 – 7.31 (d, *J* = 7.7 Hz, 1H), 7.26 (s, 1H), 3.77 (s, 2H), 3.09 (t, *J* = 6.0 Hz, 2H), 2.87 (t, *J* = 6.0 Hz, 2H), 2.58 (s, 4H), 2.04 – 1.97 (m), 1.98 – 1.94 (m, 2H), 1.66 (dt, *J* = 11.2, 5.6 Hz, 4H), 1.48 – 1.39 (m, 2H). ¹³C NMR (CDCl₃) δ 167.5, 161.4, 154.2, 139.7, 135.0, 131.4 (q, *J* = 32.3 Hz), 129.5, 124.2, 124.1 (q, *J* = 272.4 Hz), 123.2, 119.7 (q, *J* = 3.8 Hz), 117.1 (q, *J* = 4.0 Hz), 115.1, 65.8, 54.8, 26.6, 26.0, 25.7, 24.3, 22.7, 22.5.

2-(Tetrahydro-4H-pyran-4-ylidene)acetonitrile³⁵ (29). To a suspension of sodium hydride (60% mineral dispersion oil) (2.37 g, 59.3 mmol) in diethyl ether (150 mL) cooled to 0 °C was added diethyl cyanomethylphosphonate (9.55 mL, 59.3 mmol) dropwise followed by a solution of tetrahydro-4H-pyran-4-one (5.00 mL, 53.9 mmol) in diethyl ether (150 mL). After the addition was completed the reaction was warmed to room temperature and stirred overnight. H₂O and EtOAc were added, the organic layer was separated, washed with brine, dried over Na₂SO₄, filtered and concentrated *in vacuo* to provide the compound as a brown oil. The crude oil was then subjected to vacuum distillation (20 mbar) to reveal the title compound as a transparent oil which crystallised upon standing. This distillate was dissolved in hot petroleum ether and left in the fridge overnight to recrystallise. The crystals were collected and dried to afford the desired compound as a white crystalline solid (2.56 g, 39%). LCMS (*m/z*): 124.2 [M + H]⁺. HPLC: *t*_R 3.390 min. ¹H NMR (CDCl₃) δ 5.19 – 5.16 (m, 1H), 3.81 – 3.71 (m, 4H), 2.67 – 2.60 (m, 2H), 2.43 – 2.37 (m, 2H). ¹³C NMR (CDCl₃) δ 162.7, 116.2, 93.6, 68.3, 68.1, 35.9, 33.7.

2-(Tetrahydro-2H-pyran-4-yl)acetonitrile³⁵ (30). To a solution of 2-(tetrahydro-4H-pyran-4-ylidene)acetonitrile (2.65 g, 20.8 mmol), in ethyl acetate (25 mL) & acetic acid (1 mL), stirred under N₂ was added 10% Pd/C (84.0 mg, 4.17 mmol). The reaction mixture was evacuated under reduced pressure, purged with hydrogen and stirred for 5 h at rt under 1 atm of pressure. TLC showed consumption of the starting

material after 5 hours. The reaction was filtered (Celite[®]) and the filtrate was concentrated *in vacuo* to provide the compound as pale yellow oil which was used for the next reaction without any further purification (2.61 g, quantitative yield). ¹H NMR (CDCl₃) δ 4.04 – 3.98 (m, 2H), 3.40 (td, *J* = 11.9, 2.1 Hz, 2H), 2.32 (d, *J* = 6.8 Hz, 2H), 1.93 (qdd, *J* = 10.8, 7.0, 3.9 Hz, 2H), 1.75 (ddd, *J* = 12.6, 3.5, 1.6 Hz, 2H), 1.51 – 1.38 (m, 2H). ¹³C NMR (CDCl₃) δ 118.1, 67.3, 32.3, 32.1, 24.4.

2-((Tetrahydro-2H-pyran-4-yl)methyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4(3H)-one

(31). General Procedure C. The reaction mixture was filtered directly and the solid washed with petroleum ether to obtain the desired compound as a beige solid (4.23 g, 93%). mp: 250-252 °C. LCMS (*m/z*): 255.1 [M + H]⁺. HPLC: *t_R* 6.858 min. HRMS (*m/z*): C₁₁H₁₁ClN₂OS requires 255.0354 [M + H]⁺; found 255.0353. ¹H NMR (*d*₆-DMSO) δ 7.44 (s, 1H), 4.56 (s, 2H), 2.85 (t, *J* = 5.8 Hz, 2H), 2.73 (t, *J* = 5.8 Hz, 2H), 1.76 (ddd, *J* = 7.5, 6.8, 4.4 Hz, 4H). ¹³C NMR (*d*₆-DMSO) δ 163.1, 158.5, 156.0, 131.0, 130.5, 120.4, 66.8, 40.8, 33.8, 32.2, 25.3, 24.4, 22.5, 21.8.

4-Chloro-2-((tetrahydro-2H-pyran-4-yl)methyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidine

(32). General Procedure E. The residue was chromatographed on silica (1:99 Et₃N:EtOAc) to afford the compound as a gold oil (350 mg, 24%). LCMS (*m/z*): 323.2 [M + H]⁺. HPLC: *t_R* 9.066 min, >95% purity (254 nm). ¹H NMR (CDCl₃) δ 3.93 (dd, *J* = 11.5, 2.4 Hz, 2H), 3.38 (td, *J* = 11.7, 2.3 Hz, 2H), 3.06 (d, *J* = 2.1 Hz, 2H), 2.91 (d, *J* = 7.2 Hz, 2H), 2.85 (d, *J* = 2.1 Hz, 2H), 2.28 – 2.17 (m, 1H), 1.90 (dt, *J* = 6.0, 2.9 Hz, 4H), 1.60 – 1.52 (m, 2H), 1.51 – 1.39 (m, 2H). ¹³C NMR (CDCl₃) δ 169.3, 163.4, 152.9, 138.2, 126.9, 126.3, 67.8, 45.7, 35.1, 32.8, 26.3, 25.9, 22.5, 22.3.

2-((Tetrahydro-2H-pyran-4-yl)methyl)-*N*-(3-(trifluoromethyl)phenyl)-5,6,7,8-

tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4-amine (33a). General Procedure F. The crude reaction mixture was evaporated under reduced pressure and the residue chromatographed on silica (1% MeOH-CHCl₃) which furnished the title compound as a white foam (160 mg, 48%). LCMS (*m/z*): 448.2 [M + H]⁺. HPLC: *t_R* 7.945 min. HRMS (*m/z*): C₂₃H₂₄F₃N₃OS requires 448.1673 [M + H]⁺; found 448.1665. ¹H NMR (CDCl₃) δ 8.31 (s, 1H), 7.72 (dd, *J* = 8.1, 1.7 Hz, 1H), 7.47 (t, *J* = 7.9 Hz, 1H), 7.36 – 7.33 (m, 1H), 7.24 (s, 1H), 3.95 (dd, *J* = 11.5, 2.4 Hz, 2H), 3.41 (td, *J* = 11.8, 2.1 Hz, 2H), 3.07 (t, *J* = 6.0 Hz, 2H), 2.84 (dd, *J* = 13.1, 6.7 Hz, 4H), 2.26 (dq, *J* = 15.1, 7.5, 3.7 Hz, 1H), 2.03 – 1.90 (m, 4H), 1.63 (dd, *J* = 13.0, 1.9 Hz, 2H),

1.52 – 1.43 (m, 2H). ^{13}C NMR (CDCl_3) δ 167.3, 163.4, 154.2, 139.5, 134.3, 131.3, (q, $J = 32.2$ Hz), 124.3, 124.1, (q, $J = 272.5$ Hz) 123.0, 119.7, (q, $J = 3.8$ Hz), 117.2, (q, $J = 4.0$ Hz), 114.5, 67.9, 46.2, 34.9, 32.9, 26.5, 25.5, 22.6, 22.5.

***N,N*-Diethyl-2-((tetrahydro-2*H*-pyran-4-yl)methyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-**

***d*]pyrimidin-4-amine (33b).** General procedure F. Chromatographed on silica (50:50 petroleum ether:EtOAc) furnishing the target compound as a gold oil (150 mg, quantitative yield). LCMS (m/z): 360.2 [$\text{M} + \text{H}$] $^+$. HPLC: t_{R} 6.664 min. HRMS (m/z): $\text{C}_{20}\text{H}_{29}\text{N}_3\text{OS}$ requires 360.2107 [$\text{M} + \text{H}$] $^+$; found 360.2104. ^1H NMR (CDCl_3) δ 3.94 (dd, $J = 11.4, 2.4$ Hz, 2H), 3.44 (dd, $J = 14.1, 7.1$ Hz, (m, 4H), 3.42 – 3.34 (m, 2H), 2.89 (t, $J = 5.9$ Hz, 2H), 2.84 (t, $J = 6.3$ Hz, 2H), 2.79 (d, $J = 7.2$ Hz, 2H), 2.27 – 2.15 (m, 1H), 1.96 – 1.88 (m, 2H), 1.81 – 1.74 (m, 2H), 1.59 (dd, $J = 13.0, 1.7$ Hz, 2H), 1.44 (ddd, $J = 11.9, 10.3, 6.0$ Hz, m, 2H), 1.12 (t, $J = 7.0$ Hz, 6H). ^{13}C NMR (CDCl_3) δ 168.9, 162.0, 161.9, 132.9, 127.6, 119.6, 68.1, 45.9, 44.9, 34.9, 33.0, 26.7, 25.8, 23.1, 23.0, 12.5.

4-Morpholinobutanenitrile³⁶ (35b). Morpholine (3.66 mL, 42.3 mmol) and 4-chlorobutanenitrile (2.00 mL, 21.6 mmol) were taken up in toluene (30 mL) and stirred at reflux temperature for 7 h. The reaction mixture was filtered directly and the solids washed with CH_2Cl_2 . The filtrate was acidified with 1M KHSO_4 solution and extracted with further CH_2Cl_2 , and the organic extracts were discarded. The aqueous phase was then adjusted to pH 12 with 1M NaOH and extracted with CH_2Cl_2 (3 x 25 mL). The organic extracts were dried over anhydrous Na_2SO_4 , and concentrated *in vacuo* to yield the compound as a transparent oil which was utilised without further purification (2.12 g, 65%). LCMS (m/z): 155.1 [$\text{M} + \text{H}$] $^+$. ^1H NMR (CDCl_3) δ 3.77 – 3.66 (m, 4H), 2.53 – 2.37 (m, 8H), 1.83 (p, $J = 6.9$ Hz, 2H). ^{13}C NMR (CDCl_3) δ 119.8, 66.9, 56.7, 53.5, 22.5, 14.9.

5-Morpholinopentanenitrile³⁶ (35c). Morpholine (6.93 mL, 80.4 mmol) and 5-chlorovaleronitrile (4.50 mL, 40.2 mmol) were taken up in toluene (50 mL) and stirred at reflux temperature for 8 h. The reaction mixture was filtered directly and the solids washed with CH_2Cl_2 . The filtrate was acidified with 1M KHSO_4 solution and extracted with further CH_2Cl_2 , and the organic extracts were discarded. The aqueous phase was then adjusted to pH 12 with 1M NaOH and extracted with CH_2Cl_2 (3 x 30 mL). The organic extracts were dried over anhydrous Na_2SO_4 , and concentrated *in vacuo* to yield the compound as a transparent oil which was

utilised without further purification (2.93 g, 43%). LCMS (m/z): 169.2 [M + H]⁺. ¹H NMR (CDCl₃) δ 3.77 – 3.61 (m, 4H), 2.52 – 2.29 (m, 8H), 1.80 – 1.57 (m, 4H). ¹³C NMR (CDCl₃) δ 119.7, 67.0, 57.8, 53.7, 25.4, 23.4, 17.1.

2-(2-Chloroethyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4(3*H*)-one (36a). General

procedure C. The reaction was allowed to cool and the precipitate was collected, washed with cold 1,4-dioxane, triturated with petroleum ether, filtered and dried to afford the compound as a beige powder (4.95 g, 83%). LCMS (m/z): 269.1 [M + H]⁺. HPLC: t_R 6.677 min. ¹H NMR (*d*₆-DMSO) δ 12.38 (s, 1H), 3.98 (t, *J* = 6.6 Hz, 2H), 3.09 (t, *J* = 6.6 Hz, 2H), 2.84 (t, *J* = 5.8 Hz, 2H), 2.71 (t, *J* = 5.8 Hz, 2H), 1.83 – 1.67 (m, 4H). ¹³C NMR (*d*₆-DMSO) δ 162.6, 158.4, 154.2, 131.6, 130.7, 120.8, 41.5, 36.6, 25.3, 24.4, 22.5, 21.8.

2-(3-Chloropropyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4(3*H*)-one (36b). General

procedure C. The reaction mixture was allowed to cool and the formed precipitate was collected and washed with cold 1,4-dioxane to yield a light brown solid (3.16 g, 60%). LCMS (m/z): 283.1 [M + H]⁺. HPLC: t_R 7.018 min. ¹H NMR (*d*₆-DMSO) δ 12.29 (s, 1H), 3.70 (t, *J* = 6.5 Hz, 1H), 2.85 (t, *J* = 5.8 Hz, 2H), 2.78 – 2.73 (m, 2H), 2.73 – 2.68 (m, 2H), 2.15 (dq, *J* = 13.6, 6.6 Hz, 2H), 1.77 (ddt, *J* = 8.6, 5.2, 2.6 Hz, 4H). ¹³C NMR (*d*₆-DMSO) δ 162.6, 158.5, 156.4, 131.2, 130.6, 120.6, 44.6, 30.9, 29.3, 25.3, 24.4, 22.5, 21.8.

2-(4-Chlorobutyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4(3*H*)-one (36c). The reaction

mixture was allowed to cool and water (150 mL) was added. The precipitate was collected, washed with MeOH, and recrystallised from DMF to afford the compound as light brown crystals (6.56 g, 83% yield). LCMS (m/z): 297.1 [M + H]⁺. HPLC: t_R 7.276 min. ¹H NMR (*d*₆-DMSO) δ 12.22 (s, 1H), 3.66 (t, *J* = 6.3 Hz, 2H), 2.85 (t, *J* = 6.0 Hz, 2H), 2.71 (t, *J* = 5.8 Hz, 2H), 2.61 (t, *J* = 7.2 Hz, 2H), 1.86 – 1.68 (m, 8H). ¹³C NMR (*d*₆-DMSO) δ 163.1, 158.5, 157.0, 131.2, 130.5, 120.4, 45.0, 32.9, 31.3, 25.2, 24.4, 24.1, 22.5, 21.8.

2-(2-Morpholinoethyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4(3*H*)-one (37a). General

Procedure D. Residue chromatographed on silica (1:99 Et₃N:EtOAc) to yield the compound as a white solid (800 mg, 56%). LCMS (m/z): 320.1 [M + H]⁺. HPLC: t_R 4.909 min. HRMS (m/z): C₁₆H₂₁N₃O₂S requires 320.1433 [M + H]⁺; found 320.1427. ¹H NMR (CDCl₃) δ 12.51 (s, 1H), 3.82 – 3.71 (m, 4H), 2.96 (t, *J* = 5.9 Hz, 2H), 2.89 – 2.83 (m, 2H), 2.84 – 2.78 (m, 2H), 2.74 (t, *J* = 5.9 Hz, 2H), 2.62 – 2.56 (q, *J* = 3.8 Hz, 4H),

1.90 – 1.73 (m, 4H). ¹³C NMR (CDCl₃) δ 163.9, 159.7, 155.6, 132.6, 131.3, 121.4, 66.9, 55.7, 53.2, 30.1, 25.6, 25.2, 23.1, 22.4.

2-(3-Morpholinopropyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4(3*H*)-one³⁷ (37b).

General Procedure C. The reaction mixture was filtered directly and the solids washed with petroleum ether to obtain the desired compound as a fluffy beige solid (3.51 g, 80%). LCMS (*m/z*): 334.1 [M + H]⁺. HPLC: *t_R* 4.925 min. HRMS (*m/z*): C₁₇H₂₃N₃O₂S requires 334.1586 [M + H]⁺; found 334.1584. ¹H NMR (*d*₆-DMSO) δ 11.54 (s, 1H), 3.99 – 3.81 (m, 4H), 3.42 (d, *J* = 12.3 Hz, 2H), 3.15 – 3.07 (m, 2H), 3.06 – 3.01 (m, 2H), 2.85 (t, *J* = 5.5 Hz, 2H), 2.70 (t, *J* = 7.2 Hz, 4H), 2.17 (dq, *J* = 15.4, 7.6 Hz, 2H), 1.86 – 1.69 (m, 4H). ¹³C NMR (*d*₆-DMSO) δ 162.9, 158.9, 156.5, 131.7, 131.0, 121.1, 63.5, 55.5, 51.3, 31.2, 25.7, 24.9, 22.9, 22.2, 20.8.

2-(4-Morpholinobutyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4(3*H*)-one (37c). General

Procedure C. The reaction mixture was filtered directly and the solids collected and washed with hot *i*-PrOH to afford the desired compound as a white solid (2.34 g, 57%). LCMS (*m/z*): 348.2 [M + H]⁺. HPLC: *t_R* 5.034 min. HRMS (*m/z*): C₁₈H₂₅N₃O₂S requires 348.1749 [M + H]⁺; found 348.1740. ¹H NMR (*d*₆-DMSO) δ 11.36 (s, 1H), 3.99 – 3.77 (m, 4H), 3.36 (d, *J* = 12.2 Hz, 2H), 3.13 – 2.93 (m, 2H), 2.83 (t, *J* = 5.6 Hz, 2H), 2.70 (t, *J* = 5.5 Hz, 2H), 2.64 (t, *J* = 6.8 Hz, 2H), 1.86 – 1.63 (m, 8H). ¹³C NMR (*d*₆-DMSO) δ 162.6, 158.5, 157.1, 131.3, 130.7, 120.6, 63.2, 55.5, 50.9, 33.1, 25.4, 24.5, 24.0, 22.6, 22.1, 21.9.

4-(2-(4-Chloro-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-2-yl)ethyl)morpholine (38a).

General Procedure E. The residue was chromatographed on silica (1:99 Et₃N:EtOAc) to afford the compound as a gold oil (270 mg, 78%). LCMS (*m/z*): 338.1 [M + H]⁺. HPLC: *t_R* 5.824 min. ¹H NMR (CDCl₃) δ 3.87 – 3.52 (m, 4H), 3.29 – 3.12 (m, 2H), 3.05 (s, 2H), 2.98 – 2.89 (m, 2H), 2.85 (s, 2H), 2.59 – 2.50 (m, 4H), 1.94 – 1.84 (m, 4H). ¹³C NMR (CDCl₃) δ 169.4, 163.3, 152.9, 138.1, 126.9, 126.4, 67.0, 57.2, 53.4, 35.9, 26.3, 25.9, 22.5, 22.3.

4-(3-(4-Chloro-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-2-yl)propyl)morpholine (38b).

General Procedure E. The residue was chromatographed on silica (1:99 Et₃N:EtOAc) to afford the compound as a gold oil (1.06 g, 67%). LCMS (*m/z*): 352.1 [M + H]⁺. HPLC: *t_R* 5.938 min. ¹H NMR (CDCl₃) δ 3.66 – 3.61 (m, 4H), 3.08 – 2.95 (m, 4H), 2.88 – 2.78 (m, 2H), 2.47 – 2.37 (m, 4H), 2.11 – 2.00 (m, 2H), 1.91 (dt, *J*

1 = 6.0, 2.9 Hz, 4H). ^{13}C NMR (CDCl_3) δ 169.4, 164.8, 152.8, 137.9, 126.9, 126.2, 66.9, 58.4, 53.6, 26.3, 25.9,
2 25.4, 22.5, 22.2.
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4 **4-(4-(4-Chloro-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-2-yl)butyl)morpholine (38c).**

5 General Procedure E. The residue was chromatographed on silica (1:99 $\text{Et}_3\text{N}:\text{EtOAc}$) to afford the compound
6 as a gold oil (1.42 g, 88%). LCMS (m/z): 366.1 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 6.255 min. ^1H NMR (CDCl_3) δ 3.75 –
7 3.63 (m, 4H), 3.07 – 3.02 (m, 2H), 3.02 – 2.97 (m, 2H), 2.87 – 2.82 (m, 2H), 2.44 – 2.34 (m, 6H), 1.98 – 1.79
8 (m, 6H), 1.62 – 1.53 (m, 2H). ^{13}C NMR (CDCl_3) δ 169.4, 164.9, 152.9, 138.0, 126.9, 126.2, 66.9, 58.7, 53.7,
9 26.5, 26.2, 26.2, 25.9, 22.5, 22.2.
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18 **2-(2-Morpholinoethyl)-*N*-(3-(trifluoromethyl)phenyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]**
19 **pyrimidin-4-amine (39a).** General Procedure F. The crude reaction mixture was concentrated under
20 reduced pressure and the residue chromatographed on silica (1:99 $\text{Et}_3\text{N}:\text{EtOAc}$). The product was isolated and
21 found to have co-eluted with a reaction by-product (m/z : 347.2 $[\text{M} + \text{H}]^+$), this mixture was concentrated under
22 reduced pressure, and the resulting bright yellow solid was suspended in Et_2O . The solids were collected under
23 vacuum filtration & washed with Et_2O to afford the desired compound as pale yellow crystalline solid (233
24 mg, 27%). LCMS (m/z): 463.2 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 7.025 min. HRMS (m/z): $\text{C}_{23}\text{H}_{25}\text{F}_3\text{N}_4\text{O}$ requires 463.1783
25 $[\text{M} + \text{H}]^+$; found 463.1774. ^1H NMR (CDCl_3) δ 8.28 (s, 1H), 7.71 (dd, $J = 7.9, 1.5$ Hz, 1H), 7.44 (t, $J = 7.9$
26 Hz, 1H), 7.35 – 7.31 (m, 1H), 7.23 (s, 1H), 3.87 – 3.60 (m, 2H), 3.12 – 3.04 (m, 4H), 2.97 – 2.91 (m, 2H),
27 2.84 (t, $J = 6.0$ Hz, 2H), 2.59 – 2.49 (m, 4H), 2.07 – 1.89 (m, 4H). ^{13}C NMR (CDCl_3) δ 167.4, 163.3, 154.4,
28 139.6, 134.4, 131.4 (q, $J = 32.3$ Hz), 129.4, 124.4, 124.1 (q, $J = 272.4$ Hz), 123.2, 119.8 (q, $J = 3.8$ Hz), 117.4
29 (q, $J = 4.0$ Hz), 114.7, 67.2, 57.4, 53.6, 36.3, 26.6, 25.6, 22.6, 22.5.
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46 **2-(3-Morpholinopropyl)-*N*-(3-(trifluoromethyl)phenyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]**
47 **pyrimidin-4-amine (39b).** General Procedure F. The residue was chromatographed on silica (1:99
48 $\text{Et}_3\text{N}:\text{EtOAc}$) to afford the title compound as a light brown oil which later solidified under high vacuum to
49 give a dark white solid (383 mg, 56%). LCMS (m/z): 477.2 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 6.671 min. HRMS (m/z):
50 $\text{C}_{24}\text{H}_{27}\text{F}_3\text{N}_4\text{OS}$ requires 477.1937 $[\text{M} + \text{H}]^+$; found 477.1930. ^1H NMR (CDCl_3) δ 8.24 (s, 1H), 7.78 (dd, $J =$
51 8.2, 1.8 Hz, 1H), 7.45 (t, $J = 7.9$ Hz, 1H), 7.36 – 7.31 (m, 1H), 7.22 (s, 1H), 3.70 – 3.66 (m, 4H), 3.07 (t, $J =$
52 6.0 Hz, 2H), 2.96 – 2.88 (m, 2H), 2.84 (t, $J = 6.0$ Hz, 2H), 2.44 (dd, $J = 8.4, 6.6$ Hz, 6H), 2.13 – 1.89 (m, 6H).
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¹³C NMR (CDCl₃) δ 167.3, 164.7, 154.3, 139.5, 134.1, 131.3, (q, *J* = 32.1 Hz), 129.3, 124.3, 124.1, (q, *J* = 272.4 Hz), 123.1, 119.7, (q, *J* = 3.9 Hz), 117.2, (q, *J* = 4.0 Hz), 114.5, 67.1, 58.5, 53.6, 37.1, 26.5, 25.5, 25.2, 22.5, 22.4.

2-(4-Morpholinobutyl)-*N*-(3-(trifluoromethyl)phenyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-

***d*]pyrimidin-4-amine (39c).** General Procedure F. The residue was chromatographed on silica (1:99 Et₃N:EtOAc) to afford the title compound as a pale yellow crystalline solid (400 mg, 59%). LCMS (*m/z*): 491.2 [M + H]⁺. HPLC: *t*_R 6.671 min. HRMS (*m/z*): C₂₅H₂₉F₃N₄OS requires 491.2102 [M + H]⁺; found 491.2087. ¹H NMR (CDCl₃) δ 8.30 (s, 1H), 7.74 (dd, *J* = 8.2, 1.8 Hz, 1H), 7.46 (t, *J* = 7.9 Hz, 1H), 7.35 – 7.31 (m, 1H), 7.22 (s, 1H), 3.76 – 3.62 (m, 4H), 3.06 (t, *J* = 6.0 Hz, 2H), 2.94 – 2.88 (m, 2H), 2.84 (t, *J* = 6.0 Hz, 2H), 2.47 – 2.33 (m, 6H), 1.97 (dddd, *J* = 23.5, 21.1, 13.6, 8.1 Hz, 6H), 1.59 (tt, *J* = 9.8, 6.6 Hz, 2H). ¹³C NMR (CDCl₃) δ 167.3, 164.8, 154.2, 139.5, 134.1, 131.26, (q, *J* = 32.2 Hz), 129.3, 124.07, (q, *J* = 272.6 Hz), 123.0, 119.6, (q, *J* = 3.8 Hz), 117.19, (q, *J* = 4.0 Hz), 114.4, 67.0, 58.9, 53.7, 38.9, 26.5, 26.2, 25.5, 22.5, 22.4.

***N,N*-Diethyl-2-(4-morpholinobutyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4-amine (40).**

General procedure F. Pale yellow solid (145 mg, 94%). mp: 60-62 °C. LCMS (*m/z*): 403.3 [M + H]⁺. HPLC: *t*_R 5.370 min. HRMS (*m/z*): C₂₂H₃₄N₄OS requires 403.2532 [M + H]⁺; found 405.2326. ¹H NMR (CDCl₃) δ 3.76 – 3.65 (m, 4H) 3.44 (q, *J* = 7.0 Hz, 4H) 2.94 – 2.79 (m, 6H) 2.45 (d, *J* = 3.8 Hz, 4H), 2.39 (dd, *J* = 8.6, 7.0 Hz, 2H), 1.96 – 1.88 (m, 2H), 1.87 – 1.81 (m, 2H), 1.78 (dtd, *J* = 8.9, 5.9, 2.9 Hz, 2H), 1.63 – 1.52 (m, 2H), 1.12 (t, *J* = 7.0 Hz, 6H). ¹³C NMR (CDCl₃) δ 168.9, 163.5, 162.1, 132.8, 127.6, 119.6, 66.9, 58.9, 53.7, 44.8, 38.8, 26.8, 26.5, 26.1, 25.9, 23.2, 23.1, 12.6.

2-(4-Morpholinobutyl)thieno[2,3-*d*]pyrimidin-4(3*H*)-one (41). General Procedure C. The reaction vessel

was decanted leaving a dark solid mass. The vessel was washed with 1,4-dioxane and the solids were collected and stirred in hot 1,4-dioxane to wash away any 1,4-dioxane soluble impurities. The crude solid was chromatographed on silica (5% MeOH-DCM) affording the desired compound as a fluffy beige solid (1.22 g, 57%). LCMS (*m/z*): 294.2 [M + H]⁺. HPLC: *t*_R 3.664 min. HRMS (*m/z*): C₁₄H₁₉N₃O₂S requires 294.1277 [M + H]⁺; found 294.1271. ¹H NMR (CDCl₃) δ 7.44 (d, *J* = 5.8 Hz, 1H), 7.19 (d, *J* = 5.8 Hz, 1H), 3.78 – 3.73 (m, 4H), 2.87 (t, *J* = 7.5 Hz, 2H), 2.51 – 2.48 (m, 2H), 2.47 – 2.43 (m, 2H), 1.90 (dt, *J* = 15.1, 7.5 Hz, 2H), 1.70 –

1.61 (m, 2H). ^{13}C NMR (CDCl_3) δ 166.0, 160.1, 157.8, 122.3, 122.2, 121.4, 66.4, 58.1, 53.4, 45.8, 34.4, 25.3, 25.2.

4-(4-(4-Chlorothieno[2,3-*d*]pyrimidin-2-yl)butyl)morpholine (42). General Procedure D. The residue was chromatographed on silica (1:99 Et_3N - EtOAc) which afforded the desired compound as a golden oil (485 mg, 51%). LCMS (m/z): 312.2 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 4.734 min. ^1H NMR (CDCl_3) δ 7.52 (d, $J = 6.0$ Hz, 1H), 7.39 (d, $J = 6.0$ Hz, 1H), 3.73 – 3.69 (m, 4H), 3.10 – 3.02 (m, 2H), 2.46 – 2.42 (m, 4H), 2.42 – 2.37 (m, 2H), 1.92 (dt, $J = 15.5, 7.7$ Hz, 2H), 1.60 (ddd, $J = 17.9, 8.6, 6.7$ Hz, 2H). ^{13}C NMR (CDCl_3) δ 169.6, 166.5, 154.9, 127.2, 127.1, 119.9, 67.1, 58.9, 53.9, 38.9, 26.7, 26.3.

2-(4-Morpholinobutyl)-*N*-(3-(trifluoromethyl)phenyl)thieno[2,3-*d*]pyrimidin-4-amine (43a). General procedure F. The solvent was concentrated *in vacuo* and the brown residue was chromatographed on silica (98:1:1 $\text{EtOAc}/\text{MeOH}/\text{Et}_3\text{N}$) furnishing the target compound as a transparent oil, later turning into a white powder under high vacuum (201 mg, 80%). LCMS (m/z): 437.2 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 5.670 min. HRMS (m/z): $\text{C}_{21}\text{H}_{23}\text{F}_3\text{N}_4\text{OS}$: requires 437.1621 $[\text{M} + \text{H}]^+$; found 437.1617. ^1H NMR (CDCl_3) δ 8.33 (t, $J = 1.8$ Hz, 1H), 7.80 (dd, $J = 8.1, 0.8$ Hz, 1H), 7.47 (t, $J = 7.9$ Hz, 1H), 7.40 (s, 1H), 7.36 (d, $J = 7.8$ Hz, 1H), 7.31 (d, $J = 6.0$ Hz, 1H), 7.25 (d, $J = 6.0$ Hz, 1H), 3.73 – 3.67 (m, 4H), 3.00 – 2.94 (m, 2H), 2.46 – 2.34 (m, $J = 13.7, 6.1$ Hz, 6H), 1.98 – 1.88 (m, 2H), 1.64 – 1.55 (m, 2H). ^{13}C NMR (CDCl_3) δ 168.4, 166.2, 154.4, 139.5, 131.31 (q, $J = 32.3$ Hz), 129.4, 123.9 (q, $J = 272.2$ Hz), 123.4, 123.3, 120.0 (q, $J = 3.7$ Hz), 117.6 (q, $J = 4.0$ Hz), 116.6, 114.8, 67.0, 58.8, 53.7, 39.1, 26.2.

***N,N*-Diethyl-2-(4-morpholinobutyl)thieno[2,3-*d*]pyrimidin-4-amine (43b).** General procedure F. The solvents were evaporated under reduced pressure and the residue was chromatographed on silica (3:2 EtOAc :petroleum ether) furnishing the desired compound as a gold oil (178 mg, 89%). LCMS (m/z): 349.2 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 4.108 min. HRMS (m/z): $\text{C}_{18}\text{H}_{28}\text{N}_4\text{OS}$: requires 349.2063 $[\text{M} + \text{H}]^+$; found 349.2057. ^1H NMR (CDCl_3) δ 7.26 (d, $J = 6.2$ Hz, 1H), 7.09 (d, $J = 6.2$ Hz, 1H), 3.74 (q, $J = 7.1$ Hz, 4H), 3.70 (dd, $J = 7.8, 3.1$ Hz, 4H), 2.81 (t, $J = 7.5$ Hz, 2H), 2.44 – 2.41 (m, 4H), 2.40 – 2.35 (m, 2H), 1.86 (dt, $J = 15.3, 7.6$ Hz, 2H), 1.58 (tt, $J = 9.8, 6.6$ Hz, 2H), 1.31 (t, $J = 7.0$ Hz, 6H). ^{13}C NMR (CDCl_3) δ 169.9, 165.2, 156.9, 121.2, 119.3, 112.5, 67.1, 59.1, 53.8, 43.9, 38.9, 26.7, 26.3, 13.4.

1 ***N,N*-diethyl-2-(morpholinomethyl)thieno[2,3-*d*]pyrimidin-4-amine (44)**. General procedure F. The
2 solution was concentrated under reduced pressure and the residue chromatographed on silica (4:1 EtOAc:
3 petroleum ether) affording the desired compound as a light brown oil (125 mg, 94%). mp: 60-62 °C. LCMS
4 (*m/z*): 262.1 [M + H]⁺. HPLC: *t*_R 6.281 min. HRMS (*m/z*): C₁₅H₂₂N₄OS requires 307.1589 [M + H]⁺; found
5 307.1587. ¹H NMR (CDCl₃) δ 7.29 (d, *J* = 6.2 Hz, 1H), 7.15 (d, *J* = 6.2 Hz, 1H), 3.80 – 3.77 (m, 4H), 3.76
6 (dd, *J* = 12.4, 5.5 Hz, 4H), 3.71 (s, 2H), 2.75 – 2.59 (m, 4H), 1.31 (t, *J* = 7.0 Hz, 3H). ¹³C NMR (CDCl₃) δ
7 169.9, 160.9, 156.8, 121.1, 120.2, 113.1, 66.9, 65.2, 53.8, 44.0, 13.4.

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16 **Thieno[2,3-*d*]pyrimidin-4(3*H*)-one (45)**. Ethyl 2-aminothiophene-3-carboxylate (1.25 g, 7.30 mmol) was
17 taken up in formamide (15 mL) and heated to 170 °C under a N₂ atmosphere. The heating was continued for
18 5 h, in which the progress of the reaction was monitored by LC/MS. The crude reaction mixture was diluted
19 with EtOAc and extracted with aqueous 1M HCl solution (3 x 30 mL). The organic extracts were discarded,
20 the aqueous phase adjusted to ~pH 10 and further extracted with EtOAc (3 x 30 mL). The organic extracts
21 were collected and dried over anhydrous sodium sulphate and the solvent was concentrated under reduced
22 pressure affording the compound as a yellow solid (845 mg, 76%). LCMS (*m/z*): 153.1 [M + H]⁺. HPLC: *t*_R
23 3.424 min. ¹H NMR (*d*₆-DMSO) δ 12.50 (s, 1H), 8.14 (s, 1H), 7.59 (d, *J* = 5.8 Hz, 1H), 7.41 (d, *J* = 5.8 Hz,
24 1H). ¹³C NMR (*d*₆-DMSO) δ 164.6, 157.9, 146.1, 125.1, 124.3, 122.1.

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37 **4-Chlorothieno[2,3-*d*]pyrimidine (46)**. General Procedure E. The black crude residue was chromatographed
38 on silica (60:40 EtOAc:petroleum ether) to afford the compound as a fluffy beige solid (695 mg, 77%). LCMS
39 (*m/z*): 170.9 [M + H]⁺. HPLC: *t*_R 5.394 min. ¹H NMR (CDCl₃) δ 8.88 (s, 1H), 7.64 (d, *J* = 6.0 Hz, 1H), 7.46
40 (d, *J* = 6.0 Hz, 1H). ¹³C NMR (CDCl₃) δ 168.8, 155.0, 152.8, 129.6, 128.4, 119.2.

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46 ***N*-(3-(trifluoromethyl)phenyl)thieno[2,3-*d*]pyrimidin-4-amine (47a)**. General procedure F. The solvent
47 was evaporated under reduced pressure and the residue was chromatographed on silica (70:29:1 petroleum
48 ether:EtOAc:Et₃N). This afforded the target compound as a white/pale yellow solid (225 mg, 65%). mp: 60-
49 62 °C. LCMS (*m/z*): 296.0 [M + H]⁺. HPLC: *t*_R 6.842 min. HRMS (*m/z*): C₁₃H₈F₃N₃S requires 296.0466 [M
50 + H]⁺; found 296.0464. ¹H NMR (400 MHz, *d*₆-DMSO) δ 9.92 (s, 1H), 8.60 (s, 1H), 8.33 (t, *J* = 2.0 Hz, 1H),
51 8.23 (dd, *J* = 8.1, 2.0 Hz, 1H), 7.93 (d, *J* = 6.0 Hz, 1H), 7.80 (d, *J* = 6.0 Hz, 1H), 7.63 (t, *J* = 8.0 Hz, 1H), 7.44
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(ddt, $J = 7.8, 1.7, 0.9$ Hz, 1H). ^{13}C NMR (d_6 -DMSO) δ 166.8, 154.5, 152.9, 140.3, 129.7, 129.3 (d, $J = 31.5$ Hz), 124.5, 124.3 (q, $J = 272.6$ Hz), 119.4, 119.2 (q, $J = 3.7$ Hz), 117.2, 116.9 (q, $J = 4.0$ Hz).

***N,N*-diethylthieno[2,3-*d*]pyrimidin-4-amine (47b)**. General procedure F. The bright blue crude reaction was evaporated of any volatile solvents and the resulting residue chromatographed on silica (50:50 petroleum ether:EtOAc) affording the compound as a light green powder (223 mg, 91%). LCMS (m/z): 208.1 $[\text{M} + \text{H}]^+$. HPLC: t_{R} 4.098 min. HRMS (m/z): $\text{C}_{10}\text{H}_{13}\text{N}_3\text{S}$: requires 208.0984 $[\text{M} + \text{H}]^+$; found 208.0903. ^1H NMR (CDCl_3) δ 8.43 (s, 1H), 7.32 (d, $J = 6.2$ Hz, 1H), 7.19 (d, $J = 6.2$ Hz, 1H), 3.76 (q, $J = 7.1$ Hz, 4H), 1.33 (t, $J = 7.1$ Hz, 6H). ^{13}C NMR (CDCl_3) δ 169.1, 157.1, 153.1, 121.4, 120.5, 114.8, 44.1, 13.3.

Pharmacological Characterisation. *Materials.* Dulbecco's modified Eagle's medium, Flp-In CHO cells, and hygromycin B were purchased from Invitrogen (Carlsbad, CA). Fetal bovine serum (FBS) was purchased from ThermoTrace (Melbourne, VIC, Australia). [^3H]raclopride, GF/C 96-well filter plates, and MicroScint-0 were from PerkinElmer (Boston, MA). All other reagents were purchased from Sigma Aldrich (St. Louis, MO).

Cell culture. FlpIn Chinese Hamster Ovary (CHO) cells (Invitrogen, Carlsbad, CA, USA) stably expressing the hD_{2L}R were grown and maintained in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal bovine serum (FBS), and 200 $\mu\text{g}/\text{mL}$ of Hygromycin-B, and maintained at 37 $^\circ\text{C}$ in a humidified incubator containing 5% CO_2 .

[^3H]Raclopride Binding Assay: The membranes of FlpIn CHO cells expressing the hD_{2L}R were prepared as previously described.³⁸ Radioligand inhibition assays were performed by co-incubating membrane homogenates (15 μg protein) with varying concentrations of test compound in binding buffer (20 mM HEPES, 100 mM NaCl, 6 mM MgCl_2 , 1 mM EGTA, 1 mM EDTA; pH 7.4) containing 0.5 nM of [^3H]raclopride to a final volume of 300 μL in the absence or presence of the competing compounds, and incubated at 25 $^\circ\text{C}$ for 2-3 h. Each test compound was dissolved in dimethyl sulfoxide and diluted in the binding buffer into 10 half- $^{10}\log$ serial dilutions: final test concentrations ranged from 10^{-3} M to 10^{-7} M. All compound concentrations were tested in duplicate. Nonspecific binding was determined using 10 μM haloperidol (Sigma-Aldrich, St. Louis, MO). In the kinetic binding assay, the dissociation of [^3H]raclopride was started after being incubated

for 60 minutes. Binding was terminated by fast flow filtration over GF/C membrane unifier plates (PerkinElmer) using a Uniplate 96-well harvester (PerkinElmer) followed by five washes with ice-cold 0.9% NaCl. MicroScint-0 (50 μL /well) was added, plates were sealed (TopSealTM), and radioactivity was measured in a MicroBeta2 LumiJET MicroBeta counter (PerkinElmer). The affinity (K_d) of [³H]raclopride at the D_{2L}R determined in saturation binding assays is 1.07 ± 0.12 nM and the B_{max} value is 1.51 ± 0.26 pmol/mg protein.

Cell culture and transfection for cAMP assay: FlpIn CHO cells stably expressing the human dopamine D_{2L} receptor were maintained in DMEM supplemented with 5% foetal calf serum (FBS) and 0.2 mg/mL hygromycin at 37 °C in a humidified incubator supplied with 5% CO₂. For transfection, the cells were grown in 10 cm culture dishes until sub-confluent. A mixture of 4 μg plasmid DNA containing BRET-based cAMP (CAMYEL) biosensor construct²⁹ and 25 μg 20 kDa linear polyethylene imine (PEI) in 500 μL 150 mM NaCl was added into a dish of cells.

cAMP measurement: The cellular cAMP levels were measured with the CAMYEL BRET-based biosensor for cAMP.²⁹ One day after transfection, cells were trypsinised and seeded in white 96-well microplates. The cells were then cultured for an additional day, rinsed twice with Hank's Balanced Salt Solution (HBSS) and were then incubated in fresh HBSS. The *Renilla* luciferase (*RLuc*) substrate coelenterazine-h was added to reach a final concentration of 5 μM . The cells were stimulated with dopamine in the presence of 10 μM forskolin (final concentration). For the antagonism assay, the antagonists were added 30 min prior to stimulation. The BRET signals were measured using a BMG Lumistar counter 30 min after stimulation. The BRET signal (BRET ratio) was determined by calculating the ratio of the light emitted at 535 \pm 30 nm (YFP) to the light emitted at 475 \pm 30 nm (RLuc).

Data analysis. Computerized nonlinear regression, statistical analyses and simulations were performed using Prism 6.0 (GraphPad Prism 6.0b Software, San Diego, CA).

Analysis of radioligand binding experiments:

Competition-binding curves between [³H]raclopride and **1** and analogues of **1** were fitted to the allosteric ternary complex model using the following equation.²⁷

$$Y = \frac{\frac{[A]}{K_A}}{\frac{[A]}{K_A} + \left(\frac{1 + \frac{[B]}{K_B}}{1 + \alpha \frac{[B]}{K_B}} \right)} \quad (1)$$

where Y is percentage (vehicle control) binding; [A] and [B] are the concentrations of [³H]raclopride and modulator, respectively; K_A and K_B are the equilibrium dissociation constants of [³H]raclopride and modulator, respectively; and α is the cooperativity between [³H]raclopride and modulator. Values of $\alpha > 1$ denote positive cooperativity, values < 1 (but > 0) denote negative cooperativity and values = 1 denote neutral cooperativity. For analogues of **1** that displayed incomplete displacement of [³H]raclopride, a value of α could be determined experimentally. However, for analogues that acted to completely displace the radioligand then we could not distinguish an allosteric mode of action (with high negative cooperativity) from a competitive mode of interaction. In these cases, we fit the above model fixing the value of α to assume high negative cooperativity ($\alpha = 0.0001$). Values of K_B determined using this fit did not deviate significantly from values of affinity derived using a competitive model.

Radioligand binding dissociation kinetic data was analysed and fit to an exponential decay equation using non-linear regression to determine the rate constant.

$$\text{Total Binding} = \text{NS} + (\text{Total} - \text{NS}) \times e^{-k_{\text{off}}t} \quad (2)$$

Where NS is non-specific binding and t is time expressed in minutes. The dissociation rate constant (k_{off}) is expressed in units of inverse, min^{-1} . The half-life for dissociation is also calculated as $\ln(2)/k_{\text{off}}$ or $0.6931/k_{\text{off}}$.

Analysis of functional data. All concentration-response data were fitted to the following modified four-parameter Hill equation to derive potency estimates.³⁹

$$E = \text{Basal} + \frac{(E_{\text{max}} - \text{Basal}) \cdot [A]^{nH}}{[A]^{nH} + EC_{50}^{nH}} \quad (3)$$

where E is the effect of the system, nH is the Hill slope and EC₅₀ is the concentration of agonist [A] that gives the midpoint response between basal and maximal effect of dopamine or other agonists (E_{max}), which are the lower and upper asymptotes of the response, respectively.

To determine the mode of interaction of **1** and derivatives of **1** at the D₂R in relation to the agonist dopamine, data were fit to both a competitive and allosteric model and the best fit compared statistically. A logistic equation of competitive agonist-antagonist interaction was globally fitted to data from functional experiments measuring the interaction between dopamine and all analogues of **1**:

$$Response = Bottom + \frac{(E_{max} - Bottom)}{1 + \left(\frac{10^{-pEC_{50}} \left[1 + \left(\frac{[B]}{10^{-K_B}} \right)^s \right]}{[A]} \right)^{nH}} \quad (4)$$

Where *s* represents the Schild slope for the antagonist and pA₂ represents the negative logarithm of the molar concentration of antagonist that makes it necessary to double the concentration of agonist needed to elicit the original submaximal response obtained in the absence of antagonist. The same data describing the interaction between all analogues of **1** and dopamine were also analyzed using a complete operational model of allosterism and agonism according to equation 5:²⁷

$$E = \frac{E_m (\tau_A [A] (K_B + \alpha \beta [B]) + \tau_B [B] K_A)^{nH}}{([A] K_B + K_A K_B + K_A [B] + \alpha [A] [B])^{nH} + (\tau_A [A] (K_B + \alpha \beta [B]) + \tau_B [B] K_A)^{nH}} \quad (5)$$

Where E_m is the maximum possible cellular response, [A] and [B] are the concentrations of orthosteric and allosteric ligands, respectively, K_A and K_B are the equilibrium dissociation constant of the orthosteric and allosteric ligands, respectively, τ_A and τ_B (constrained to -100) are operational measures of orthosteric and allosteric ligand efficacy (which incorporate both signal efficiency and receptor density), respectively, α is the binding cooperativity parameter between the orthosteric and allosteric ligand, and β denotes the magnitude of

1 the allosteric effect of the modulator on the efficacy of the orthosteric agonist. K_A was constrained to 617 nM,
2 and represents a value of functional affinity determined by an operational model of agonism applied to dose-
3 response data of dopamine in the presence of increasing concentrations of the alkylating agent
4 phenoxybenzamine. For compounds that caused a limited rightward shift of the dopamine dose-response curve
5 but no decrease in E_{max} , data were fit using an operational model of allosterism where $\text{Log}\beta$ was constrained
6 to 0 to represent neutral cooperativity with dopamine efficacy. For compounds that produced an unlimited
7 decrease in the maximal response of dopamine $\text{Log}\beta$ was constrained to -100.
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16 For each of the compounds the two equations (models) were then compared for their fit using an extra-
17 sum-of-squares F test. All of the data points and values shown in the figures and tables are the mean \pm s.e.m.
18 of at least three separate experiments performed in duplicate unless otherwise stated.
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25 **Molecular Docking Simulations.** The D_2R crystal structure (PDB ID 6C38)³² was prepared by the
26 regularisation algorithm implemented in ICM-Pro molecular modeling software.⁴⁰ Raclopride was docked
27 into the orthosteric pocket of the D_2R using an energy-based docking protocol implemented in the ICM-Pro
28 software.⁴⁰ The best energy binding pose predicted for raclopride in the D_2R was found to be similar to the
29 binding pose of closely related eticlopride in the co-crystallised complex with the D_3R .⁴² The D_2R -raclopride
30 complex was further optimised using Biased Probability Monte Carlo (BPMC) optimisation of the surrounding
31 residues in the proximity of 5 Å of ligand. In the final D_2R -raclopride complex, the RMSD between common
32 atoms of raclopride and the crystallographic pose of eticlopride did not exceed 0.5 Å.
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44 Compound **1** and its thirty-three analogues were docked into the D_2R -raclopride complex using an energy-
45 based docking protocol implemented in the ICM-Pro software.^{40, 41} Compounds were generated from 2D
46 representations and their 3D geometry was optimised using MMFF-94 force field. The docking was performed
47 in the rectangular box that comprised the whole extracellular part of the receptor above raclopride, including
48 the extracellular loops. Docking simulations used BPMC optimisation of the compounds internal coordinates
49 in the pre-calculated grid energy potentials of the receptor. The exhaustive sampling of the compound
50 conformational space was done with the thoroughness parameter set to 10, and at least three independent
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docking runs performed for each compound. The final docking poses were selected based on optimal ligand receptor interactions and ICM binding scores.

For D₃R docking, we used the previously published D₃R-eticlopride crystal structure (PDB ID: 3PBL)⁹, where raclopride was docked in place of eticlopride. Compound **1** and its eight analogues were then docked into D₃R-raclopride complex. Due to the hypothesised high flexibility of the E90^{2,65} residue side chain, it was set up as “explicitly flexible group” during the docking. Further docking method was used as abovementioned.

▪ ASSOCIATED CONTENT

Supporting Information

Coordinates for computational models (D₂R PDB ID: 6C38) and molecular formula strings (CSV) have been included in Supporting Information. Authors will release the atomic coordinates and experimental data upon article publication.

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Notes The authors declare no competing financial interest.

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■ ABBREVIATION LIST

cAMP, cyclic adenosine monophosphate; D₂R, dopamine D₂ receptor; D₃R, dopamine D₃ receptor; dopamine, DA; GPCR, G protein-coupled receptor; FGA, first-generation antipsychotic; SGA, second-generation antipsychotic; EPS, extrapyramidal symptoms; NAM, negative allosteric modulator; VLS, virtual ligand screening; SAR, structure-activity relationship; CHO, chinese hamster ovary; cAMP, cyclic adenosine monophosphate; BRET, bioluminescence resonance energy transfer; FBS, foetal bovine serum; HBSS, hank's balanced salt solution; SEM, standard error of mean; DMF, *N,N*-dimethylformamide, POCl₃, phosphorus oxychloride; *i*-PrOH, isopropyl alcohol; EtOAc, ethyl acetate; DCM, dichloromethane; MWI, microwave irradiation; TRIDENT, tris[2-(2-methoxyethoxy)ethyl]amine.

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