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1 Contrasting effects of pine plantations on two skinks: results from a large scale 'natural

- 2 experiment' in Australia
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12 Abstract

The aim of our study was to quantify the long-term response of two lizard species to the
transformation of the matrix surrounding remnant habitat patches (from agricultural land to
pine plantations).

We used a large scale (115 sites), long-term (16 years) fully replicated and controlled 16 landscape scale 'natural experiment' (the 'Nanangroe experiment', Australia) to compare 17 18 the response of lizard populations to plantation establishment. The study entailed detailed surveys of reptiles in 50 eucalypt patches surrounded by maturing Pine (*Pinus radiata*) 19 20 plantations (treatments) and populations inhabiting 55 eucalypt patches surrounded by 21 grazing land (controls). 22 We found that the Eastern Three-toed Earless Skink (Hemiergis talbingoensis) was advantaged by the establishment of the pine plantations (increasing colonization of eucalypt 23 24 patches embedded within plantations). In contrast, we found that the Southern Rainbow 25 Skink (Carlia tetradactyla) was negatively affected by surrounding areas of maturing 26 plantations. Thus our results show that plantations acted as a barrier for one species and increased connectivity for the other. We suggest that leaving areas of land devoid of pines 27 28 between remnant eucalypt patches may enhance the connectivity for species that respond negatively to plantation establishment, while maintaining the beneficial increase in 29 30 connectivity for others.

31

33 Introduction

The conversion of agricultural land to forest plantations is a major driver of global land use 34 change (Foley et al., 2005). To date, planted forest (including production-oriented 35 36 plantations and other types of planted forest (FAO, 2010) are present in most vegetated countries. Planted forests cover approximately 264 billion hectares, 7% of global forest 37 38 cover, roughly equivalent to half the Amazonian rainforest. Recent decades have witnessed 39 substantial increases in planted forest (e.g. a global increase of 5 million hectares/year was recorded by FAO between 2000-2010) due to an increased demand for wood and carbon 40 41 storage (Jackson et al., 2005; Paquette & Messier, 2010). Considering that large portions of 42 the planet are covered by forest plantations, understanding their impact on biodiversity is a 43 research priority (Brockerhoff et al., 2008; Felton et al., 2010). 44 Previous research on biodiversity on plantations focused mainly on birds (Renjifo, 2001; 45 Mortelliti, Westgate & Lindenmayer, 2014) whereas reptiles have attracted considerably 46 less attention (Gardner et al., 2007). Previous studies on reptiles have found that 47 plantations tend to decrease species richness and diversity (Glor et al., 2001; Loehle et al., 2005; Amo, López & Martín, 2006; Gardner et al., 2007; Lindenmayer et al., 2008) but see 48 49 (Vonesh, 2001). The magnitude of effects is context-specific and varies with the type of 50 plantations (e.g. eucalypt vs pine), the habitat of target species, and microhabitat 51 availability within the plantation (Glor et al., 2001; Vonesh, 2001; Loehle et al., 2005; Kanowski et al., 2006). Previous studies have been short-term and observational and have 52 53 focused mainly on species occurrence within the plantations (e.g. by contrasting assemblages in natural vs planted forests). Few studies have focused on the effects of 54 plantations as 'landscape context' (i.e. on reptiles inhabiting patches of remnant native 55 vegetation embedded within a plantation). An additional knowledge gap is that, 56 57 to date, no study within plantations has explicitly focused on colonization/extinction parameters, which are key ecological variables in fragmentation studies (Hanski & 58 59 Gaggiotti, 2004). Focusing on population turnover allows a more in-depth understanding of population dynamics than the "static" occupancy studies which are based on 'snapshot' 60 presence/absence data and also allows for a separate evaluation of what is affecting 61 62 population establishment (e.g. colonization) and what is affecting persistence (e.g. the 63 extinction risk).

64

To address key knowledge gaps about reptile responses to plantation establishment we 65 used a large-scale (115 sites over 30000 ha), long-term (16 year) fully replicated and 66 67 controlled landscape experiment, conducted in south-eastern Australia (the 'Nanangroe 68 experiment'). The aim of the 'Nanangroe experiment ' (Lindenmayer, 2009) is to compare responses of target species inhabiting 50 treatment eucalypt patches surrounded by 69 maturing Pine (Pinus radiata) plantations, with populations inhabiting 55 control eucalypt 70 71 patches surrounded by grazing land (Fig. 1). A unique feature of the Nanangroe study is that both treatments and controls have remained unmodified throughout the study period, 72 73 being the first study of its kind to focus on the effects of the 'outside' matrix on target 74 populations within eucalypt patches. 75 We chose two skinks (Scincidae) as target species: the Southern Rainbow Skink Carlia 76 tetradactyla and the Eastern Three-toed Earless Skink Hemiergis talbingoensis. The aim of 77 our study was to quantify the long-term response of these two target species to the

transformation of the matrix surrounding remnant habitat patches (from agricultural land topine plantations).

80

81 Materials and methods

82 Study area

Our research was conducted in the Nanangroe area (New South Wales, South-eastern Australia; Fig. 1). The Nanangroe area lies approximately 70 km north-west of Canberra (coordinates 34°54' - 35°4' and 148°32' - 148°18' E, altitudinal range: 250-750 m a.s.l.), covers approximately 30 km² and is characterised by hot summers and cool winters (temperate climate). The native vegetation (more than 80% of the original temperate eucalypt woodland has been cleared for grazing) is characterised by open woodlands dominated by red box (*Eucalyptus polyanthemos*), white box (*E. albens*), yellow box (*E. melliodora*),

90 Blakely's red gum (*E. blakleyi*) and red stringybark (*E. macrorhyncha*).

91

92 Experimental design of the Nanangroe project

93 Pine plantations (Pinus radiata) began being established in Nanangroe in late 1998 by Forest

- NSW for the production of timber and pulp (paper) and for carbon sequestration purposes.
- 95 Approximately 70 eucalypt patches of variable sizes were retained during the establishment

96 of the plantation, which were progressively surrounded by the emerging artificial landscape97 (Fig 1).

The selection of sites followed a replicated, random stratified procedure. The stratification
was based on three criteria : a) size of the eucalypt habitat patches; b) age-cohort of the
surrounding pine plantation; c) number of boundaries between patches and surrounding
pine stands.

The study design (Fig 1, Table S2) included: a) 50 woodland *treatment* sites (sites within eucalypt patches surrounded by *Pinus radiata* plantation); b) 55 woodland *control* sites (sites within eucalypt patches surrounded by grazing land); c) 10 pine control sites (sites within the plantation).

106 For each of the treatment and control sites, we selected a series of replicates based on the 107 size of the habitat patch. Patches were grouped according to patch size intervals: 0.5-0.9 ha 108 (13 replicates), 1.0-2.4 ha (19 replicates), 2.5-4.9 ha (15 replicates), >5.0 ha (3 replicates). 109 Eucalypt patches were surrounded by pines belonging to two age-cohorts (cohort 1= pines 110 planted in 1998; cohort 2= pines planted in 2000). Of the 50 woodland treatments, nine had 111 1-2 open boundaries with grazed land, whereas the remaining patches were completely surrounded by pines. A summary of the experimental design is provided in Table S2. The 112 113 woodland treatment sites were matched with 55 control sites surrounded by grazing land (Table S2). Domestic livestock grazing (sheep and cows) continued in all sites throughout 114 115 the whole study period, thereby eliminating potential confounding effects between 116 treatments.

117

118 *Reptile surveys*

119 We used arrays of artificial refuges (Michael *et al.*, 2012) to survey the distribution

120 (presence/absence) of the two target lizard species in each of the 115 sites. Artificial refuges

121 (ARs) were composed of: (1) one double-layered stack of corrugated galvanised steel, (2)

four 1-m long *E. dalrympleana* fence posts covered in mesh, and (3) four concrete roof tiles

123 (32 cm*42 cm). Over time, the fence posts decayed, so in 2010 we installed four 1.2 m long

124 railway sleepers at each site (Fig. 2).

125 At each site, we established two reptile monitoring stations (AR arrays) located 100 m apart.

126 Surveys were conducted in 1999, 2000, 2003, 2006, 2011 and 2013. Surveys were

127 completed during spring (August) each survey year and were conducted on clear sunny days

- between 0900 hrs and 1400 hrs by the same group of experienced herpetologists from TheAustralian National University.
- 130 Thirteen reptile species were captured during the study, of which C. tetradactyla and H.
- 131 *talbingoensis* were the two most abundant species. Capture rates for other species was low
- and did not allow us to fit occupancy models (see below).
- 133

134 Data analysis

- We selected a restricted set of predictor variables to be included in the data analyses (see 'occupancy models' below) in addition to "treatment" and "time" effect. These included: vegetation type and eucalypt tree cover surrounding the site. Vegetation type categories were measured in the field and were based on dominant or co-dominant tree species (Table S3). Eucalypt tree cover was measured in a circular buffer (250 m radius) centred in each site. Tree cover included habitat patches and isolated trees and was measured by using digitised aerial photography in ArcGIS 10.1.
- 142 We chose to use the tree cover in a buffer rather than patch size because scattered trees 143 play a crucial role in determining distribution patterns of Australian native fauna (Fischer,
- 144 Stott & Law, 2010). Furthermore, buffer-based measurements of habitat amount have been
- recommended as single predictor variables in fragmentation studies (instead of patch size
- 146 and isolation) (Fahrig, 2013).
- 147

148 Occupancy models

149 False absences (a species was present in a site but not detected) are a key source of bias in

distribution studies, particularly for studies focusing on terrestrial vertebrate species

- 151 (MacKenzie, 2005). To control for possible false absences, we fitted multiple season
- 152 occupancy models to presence/absence data from our reptile surveys thus allowing the

estimation of colonisation/extinction probabilities (MacKenzie, Nichols & Hines, 2003). We

- 154 focused our occupancy analyses on the 105 woodland eucalypt patches (50 woodland
- treatments and 55 woodland controls). We retained 10 pine control sites for separate
- analyses (detailed below).

157 We defined each eucalypt patch as a *site* (*sensu* (MacKenzie *et al.*, 2003)). We considered a 158 *visit* to a site the inspection of one of the two AR arrays. Therefore, each site was surveyed

twice on the same day during the same year. Following (MacKenzie *et al.*, 2003),

160 populations were assumed to be closed within each survey year (i.e. between visits) and

161 open to colonization/extinction between survey years.

162 We adopted the following protocol for fitting models:

163 1) The detection probability (p) parameter was either held constant across years or 164 modelled as a function of year. The variable "year" was included in two different 165 ways: a) we included it as a categorical variable to account for possible unmeasured 166 year-specific factors affecting the detectability of our two target species; b) we 167 included it as a continuous variable to check for possible trends (i.e. an increase or 168 decrease) in populations. The relative best model was selected through the Akaike 169 Information Criterion (AIC (Burnham & Anderson, 2002). The variable included in the 170 top ranking detection model was retained in all the following steps:

1712) In the second step of our protocol, we selected the most important variables172influencing Ψ_1 (probability of a site being occupied in the first study year). Ψ_1 was173modelled as a function of vegetation type and tree cover in the 250 m buffer). The174variable included in the top ranking occupancy model was retained in the following175step.

1763) We modelled the probability of colonisation (Υ) and the probability of extinction (ε)177as functions of the following variables: treatment (a site surrounded by pines vs178control), year since the beginning of the study (continuous variable) and tree cover179in the 250 m buffer. We also included two-way interactions for these variables (e.g.180treatment* year) and included models with different effects for colonisation and181extinctions: e.g. Υ (treatment*time), ε (tree cover).

182

The variable "year" was measured as time since the beginning of the study, which in the 183 184 case of treatments also acted as a proxy for the time since plantation establishment. We were unable to identify a method to include a time covariate for treatments only (the value 185 186 zero in the control sites would not be meaningful). Similarly, distinguishing between the two cohorts of plantation establishment and simultaneously modelling control sites was not 187 possible. To avoid conducting separate analyses for treatments and controls, we opted to 188 use the variable time since start of the study, and acknowledge that this was only a proxy 189 190 for time since the establishment of the plantation. Occupancy models were fitted using the 191 unmarked package for R (Fiske & Chandler, 2011).

We based our inference on model averaged estimates (including models within 2 ΔAIC
(Burnham & Anderson, 2002). Goodness of fit of each model was measured using
Nagelkerke's R².

195

Our experiment was designed to limit the risk of spatial dependence between sites (e.g. 196 197 spatial autocorrelation and pseudo-replication): a) four independent plantations were 198 included as treatments; b) woodland control sites were distributed amongst six different farms; c) average distance between sites was 9.3 km (range 0.11-21.7 km). Notwithstanding, 199 200 we checked for spatial autocorrelation in the residuals of the most parameterised model by 201 using a spline correlogram (Zuur et al., 2009). We used the R package ncf to produce 202 correlograms (Bjørnstad, 2009). We calculated correlograms for each of the six time periods 203 and could not find evidence of spatial autocorrelation for either of the two target species.

204

205 *Reptile use of the matrix sites.*

206 To evaluate the relative capabilities of each species to use/cross the pine matrix, we calculated an index of the relative abundance of each skink in the pine matrix (i.e. the 10 207 208 sampling sites located within the pine matrix) by summing the number of detections for 209 each species in these sites. We tested whether the relative abundance of the two target 210 species in the matrix were significantly different through a Kruskal-Wallis test. Effect size was calculated through a post-hoc Mann-Whitney test (i.e. it is not possible to estimate 211 212 effect size for the Kruskal-Wallis test). Both these analyses we conducted using the mass package in R (R Core Team, 2013). 213

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217

215 Results

216 Over the 16-year duration of our study, we detected *H. talbingoensis* 253 times in the

218 *C. tetradactyla* on 117 occasions in eucalypt patches (39 detections in treatments and 78 in 219 controls).

eucalypt patches (151 detections in treatments and 102 in controls), whereas we detected

220 *Occupancy models.* The probability of detecting *C. tetradactyla* varied each year (i.e. year as

categorical variable, Fig S3) whereas the probability of detecting *H. talbingoensis* increased

over time (i.e. year as continuous variable, Fig S3). The probability of a site being occupied in

223 the first sampling season was dependent on vegetation type for both species (Table 1). The variable 'treatment' was included as predictor of colonization probability in the top ranked 224 225 models for both species. However, the two lizard species showed strongly contrasting 226 responses to the establishment of the pine plantations surrounding the eucalypt patches. 227 For C. tetradactyla, colonization probability was lower in treatments, whereas in H. 228 talbingoensis the probability of colonization was higher in treatments (Fig 3; Table 1). In the 229 top ranked model for both species, the extinction probability was constant (Table 1). Predictors for probability of extinction included in the top model set (e.g. within ΔAIC<2 but 230 231 with less support compared to the first model) included year (positive beta = increase with 232 time) and habitat (negative beta = more habitat within the buffer decreased extinction risk) 233 for C. tetradactyla and treatment (negative beta = treatment sites had lower extinction risk), 234 year (positive beta) and habitat (negative beta) for *H. talbingoensis*. 235 Captures in the matrix. Throughout the study period, we captured a higher number of H. 236 talbingoensis (seven captures) than C. tetradactyla (one capture) in the pine matrix.

However, this difference was not statistically significant (Kruskall-Wallis χ^2 = 2.53, df = 1, *p* = 0.1; medium effect size: r = 0.35).

239

240 Discussion

Plantation forestry is widely recognised as major driver of global change (Foley et al., 2005). 241 242 We used a large-scale and long-term landscape transformation experiment to identify the 243 contrasting response of two lizard species to the broad-scale establishment of pine plantations. A unique aspect of our study was that we focused on the effects of changes 244 occurring in the matrix on lizard populations inhabiting eucalypt patches. We found that 245 pine plantations increased connectivity (i.e. colonization probability increased in the 246 247 eucalypt patches surrounded by plantation) for one species (*H. talbingoensis*), but acted as a barrier (i.e. the probability of colonization in eucalypt patches was lower in treatment sites) 248 249 for the other species (C. tetradactyla).

250

251 Biological interpretation of the models

252 The small taxonomic scope of our research (2 species) limits our inference on the possible

253 mechanisms that may have driven the observed patterns (Garland & Adolph, 1994).

- Below we list three (non mutually exclusive) key hypotheses that may explain the
- 255 contrasting pattern that we have observed (but see also Table S1 for additional differences
- between the target species). These hypotheses should be field-tested by future studies witha broader taxonomic scope.

258 Hypothesis 1). Thermoregulation. The Southern Rainbow Skink (C. tetradactyla) is a

- 259 heliothermic species (i.e. it gains heat by short wavelength solar radiation) whereas the
- 260 Eastern Three-toed Earless Skink (*Hemiergis talbingoensis*) is a thigmotherm (it gains heat by

direct contact with a warm substratum) (Garrick, 2008) (Table S1).

- 262 (Fischer, Lindenmayer & Cowling, 2003a) found that *C. tetradactyla* prefers open woodlands
- to woodland with more closed canopy, perhaps due to its heliothermic requirements, which
- 264 may suggest that high levels of shade associated with pine plantations (Porté, Huard &
- Dreyfus, 2004; Mott, Alford & Schwarzkopf, 2010) act as a barrier for this species and cause
 decreased colonization rates in the treatments compared with controls.
- 267 Although *H. talbingoensis* also inhabits eucalypt woodland (Fischer *et al.*, 2005a; Kay *et al.*,
- 268 2013), compared to *C. tetradactyla*, it is better adapted to living in cold and humid
- 269 environments. The field active body temperature of *H. talbingoensis* is 20.3 C° (range =
- 270 17.3 C° 23 C°), which is considerably lower than heliothermic skink species (Greer, 1989).
- 271 Importantly, the critical thermal minimum and maximum temperature range (CTmin = 6.8
- 272 C°, CTmax = 38.6 C°) is particularly low (data from (Bennett & John-Alder, 1986). In
- accordance with these findings, (Michael, Cunningham & Lindenmayer, 2011) found the
- 274 predicted response of *H. talbingoensis* to eucalypt regrowth (i.e. dense vegetation cover) to
- be positive, presumably because increased shade levels associated with regenerating
- vegetation created conditions consistent with the species autecology. Thus, according to
- this hypothesis, mode of thermoregulation (heliothermy vs thigmothermy) may be a key
- driver of their response to pine plantations (Mott *et al.*, 2010). We emphasise that to test
- this hypothesis detailed measurements of the thermal environments should be conducted(Mott *et al.*, 2010).
- 281 *Hypothesis 2) Reproductive mode.* Another key difference between the two target species
- that may help to explain the strongly contrasting responses observed is the mode of
- reproduction (Table S1). *Carlia tetradactyla* is oviparous (thus the pine matrix may be too
- cold for incubation) whereas *H. talbingoensis* is viviparous (i.e. gives birth to live young, thus
- 285 may potentially breed in the pine matrix; Table S1). Previous studies have shown that

availability of warm sites is critical for oviparous skinks (Shine, Elphick & Harlow, 1997) thus
reproductive mode may have played an important role in determining the observed
patterns (but see later discussion on matrix use).

Hypothesis 3) Prey availability and predation. Differential prey availability or predatory risk
caused by the plantations may also explain the differences observed. Pine plantation may
host different invertebrates (Barbaro *et al.*, 2012), which in turn may favour matrix-crossing
for one species compared to the other. Similarly, the low understory of pine plantations
may increase predation risk for *C. tetradactyla* but not for the burrowing skink (*H. talbingoensis*) and thus create a barrier effect for one species only.

A critical aspect that needs further clarification is the reptile use of the matrix sites.

296 Clarifying this issue will help identifying the mechanisms determining the contrasting 297 responses observed. We found captures of *H. talbingoensis* within the plantations were 298 higher than captures of *C. tetradactyla*, suggesting that this species has a higher capability 299 of moving through or persisting in pine plantations. The low p value (p = 0.1) but medium 300 effect size (0.35) suggest that lack of statistical significance may be due to poor statistical 301 power caused by low capture rates. Further analyses are therefore warranted to reach 302 definitive conclusions. The extremely low number of captures suggests that H. talbingoensis 303 occasionally moves through the plantation matrix (rather than living within them at low 304 population density; see also (Fischer et al., 2005b) for similar conclusions). Several factors could determine different matrix crossing capabilities, such as: a) thermoregulation, b) 305 306 tolerance to dry conditions (i.e. higher soil moisture in plantations may facilitate movement of the fossorial species c) different and species-specific perceptual ranges in the two types 307 308 of matrix (Sozio, Mortelliti & Boitani, 2013). Nevertheless, we emphasise that further 309 empirical evidence is required to reach more definitive conclusions on this issue. 310 Our occupancy analyses revealed that plantations affected colonization rather than extinction processes. However, we found some evidence for lower extinction risk of H. 311 talbingoensis in treatment sites (Table 1). It is likely that a longer study (e.g. >16 years) may 312 reveal an important influence of plantation establishment on local extinction risk, as a 313 decrease in colonization will likely lead to lack of immigration in already occupied patches 314 (and thus to an increase in local extinction risk)(Hanski & Gaggiotti, 2004). The patterns we 315

have discovered were not detected in previous studies in this area (Lindenmayer *et al.*,

2008) possibly because the present study was substantially longer (16 years vs 8 years).

Furthermore, our study was focused on different ecological variables (population turnoverrather than occupancy).

320

327

321 Implications for conservation

Our study is unique in focusing on the effects that plantations have on populations inhabiting eucalypt patches surrounded by plantations. Consequently, our results are applicable to plantations containing remnant patches. We emphasise that pines are the most widespread species in plantations worldwide (20% of plantations) (Brockerhoff *et al.*, 2008) thus making our study system highly representative.

We foresee a major implication of our study for reptile conservation within transformed

328 landscapes: pine plantations may act as a barrier (decreasing colonisation probability) for 329 some species (C. tetradactyla in our study) and increase connectivity for others (H. 330 talbingoensis in our study). We suggest the establishment of habitat corridors – in this case 331 open woodland or open canopy areas - may help mitigate the negative impact of 332 plantations on these species (see also (Pryke & Samways, 2001; Bertoncelj & Dolman, 2012, 2013) for similar conclusions on other taxa). The colonization rate of control patches 333 334 (surrounded by grazing) was higher than treatment sites, confirming that C. tetradactyla is 335 relatively good at crossing a matrix of open fields (Fischer et al., 2005a) especially when not heavily grazed (Fischer, Lindenmayer & Cowling, 2003b). We therefore suggest that leaving 336 areas without pines between remnant patches may enhance connectivity for this species. 337 338 Such a conservation action may be substantially cheaper than creating revegetated corridors, only resulting in foregone profit, rather than both missed profit and costs of 339 habitat restoration. Improving the quality of microhabitats (e.g. providing debris, rocks etc) 340 also may improve the suitability of plantations for reptile species (Amo et al., 2006; 341 342 Kanowski et al., 2006).

As our study was conducted on only two species we cannot provide clear explanations on which mechanisms (e.g. which life history traits) determined the contrasting responses observed. We hypothesise that mode of thermoregulation, reproductive mode and prey availability or predation risk may be potential drivers. The possibility of predicting the response of species to land use change based on life-history traits is a major challenge in conservation biology (Henle *et al.*, 2004; Koh, Sodhi & Brook, 2004; Ockinger *et al.*, 2010).

- 349 Nevertheless further multi-species studies are needed to test our hypotheses and thus to350 generalize our findings.
- 351 The results of our broad-scale landscape transformation experiment show how landscape
- 352 context surrounding remnant habitat patches affects the long-term population dynamics of
- 353 lizard populations. Converting agricultural land to pine plantations may favour some species
- and increases the colonization rate of previously unoccupied eucalypt patches but may also
- 355 create a barrier for other species. We suggest that leaving land devoid of pines between
- 356 remnant patches may enhance the connectivity for some species while maintaining the
- 357 beneficial increase in connectivity for others.
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359 References

- Amo, L., López, P. & Martín, J. (2006). Natural oak forest vs. ancient pine plantations: lizard
 microhabitat use may explain the effects of ancient reforestations on distribution and
 conservation of Iberian lizards. *Biodivers. Conserv.* 16, 3409–3422.
- Barbaro, L., Brockerhoff, E.G., Giffard, B. & van Halder, I. (2012). Edge and area effects on
 avian assemblages and insectivory in fragmented native forests. *Landsc. Ecol.* 27, 1451–
 1463.
- Bennett, A.F. & John-Alder, H. (1986). Thermal Relations of Some Australian Skinks (Sauria:
 Scincidae) Thermal Relations of Some Australian Skinks (Sauria: Scincidae). *Copeia* 1,
 57–64.
- Bertoncelj, I. & Dolman, P.M. (2012). The matrix affects trackway corridor suitability for an
 arenicolous specialist beetle. *J. Insect Conserv.* 17, 503–510.
- Bertoncelj, I. & Dolman, P.M. (2013). Conservation potential for heathland carabid beetle
 fauna of linear trackways within a plantation forest. *Insect Conserv. Divers.* 6, 300–308.
- Bjørnstad, O.N. (2009). ncf: spatial nonparametric covariance functions.
- Brockerhoff, E.G., Jactel, H., Parrotta, J.A., Quine, C.P. & Sayer, J. (2008). Plantation forests
 and biodiversity: oxymoron or opportunity? *Biodivers. Conserv.* 17, 925–951.
- Brown, G. (1991). Ecological Feeding Analysis of South-eastern Australian Scincids (Reptilia :
 Lacertilia). Aust. J. Zool. 39, 9.
- Burnham, K.P. & Anderson, D.R. (2002). *Model Selection and Multimodel Inference: A Practical Information-Theoretic Approach*. 2nd edn. Springer-Verlag.
- Fahrig, L. (2013). Rethinking patch size and isolation effects: the habitat amount hypothesis.
 J. Biogeogr. 40, 1649–1663.
- 382 FAO. (2010). *Global forest resources assessment 2010 Main report*. Rome.

Felton, A., Knight, E., Wood, J., Zammit, C. & Lindenmayer, D. (2010). A meta-analysis of
 fauna and flora species richness and abundance in plantations and pasture lands. *Biol. Conserv.* 143, 545–554.

Fischer, J., Fazey, I., Briese, R. & Lindenmayer, D.B. (2005a). Making the matrix matter:
 challenges in Australian grazing landscapes. *Biodivers. Conserv.* 14, 561–578.

<sup>Fischer, J., Lindenmayer, D. & Cowling, A. (2003a). Habitat models for the four-fingered
skink (Carlia tetradactyla) at the microhabitat and landscape scale.</sup> *Wildl. Res.* 30, 495–
504.

- Fischer, J., Lindenmayer, D. & Cowling, A. (2003b). Habitat models for the four-fingered
 skink (Carlia tetradactyla) at the microhabitat and landscape scale. *Wildl. Res.* **30**, 495.
- Fischer, J., Lindenmayer, D.B., Barry, S. & Flowers, E. (2005b). Lizard distribution patterns in
 the Tumut fragmentation "Natural Experiment" in south-eastern Australia. *Biol. Conserv.* 123, 301–315.
- Fischer, J., Stott, J. & Law, B.S. (2010). The disproportionate value of scattered trees. *Biol. Conserv.* 143, 1564–1567.
- Fiske, I. & Chandler, R. (2011). unmarked: An R package for fitting hierarchical models of
 wildlife occurrence and abundance. J. Stat. Softw. 43, 1–23.
- Foley, J.A., DeFries, R., Asner, G.P., Barford, C., Bonan, G., Carpenter, S.R., Chapin, F.S., Coe,
 M.T., Daily, G.C., Gibbs, H.K., Helkowski, J.H., Holloway, T., Howard, E.A., Kucharik, C.J.,
 Monfreda, C., Patz, J.A., Prentice, I.C., Ramankutty, N. & Snyder, P.K. (2005). Global
 consequences of land use. *Science (80-.).* **309**, 570–574.
- Gardner, T. a, Ribeiro-Júnior, M.A., Barlow, J., Avila-Pires, T.C.S., Hoogmoed, M.S. & Peres, C.
 a. (2007). The value of primary, secondary, and plantation forests for a neotropical
 herpetofauna. *Conserv. Biol.* 21, 775–87.
- Garland, T. & Adolph, S. (1994). Why not to do two-species comparative studies: limitations
 on inferring adaptation. *Physiol. Zool.* 67, 797–828.
- Garrick, D. (2008). Body surface temperature and length in relation to the thermal biology of
 lizards. *Biosci. Horizons* 1, 136–142.
- Glor, R.E., Flecker, A.S., Benard, M.F. & Power, A.G. (2001). Lizard diversity and agricultural
 disturbance in a Caribbean forest landscape 711–723.
- Greer, A. (1989). *The Biology and Evolution of Australian Lizards.* Chipping Norton, NSW:
 Surrey Beatty and Sons.
- Hanski, I. & Gaggiotti, O.E. (2004). *Ecology, Genetics, and Evolution of Metapopulations*.
 Boston: Elsevier Academic Press.
- Henle, K., Davies, K.F., Kleyer, M., Margules, C. & Settele, J. (2004). Predictors of Species
 Sensitivity to Fragmentation. *Biodivers. Conserv.* 13, 207–251.
- Jackson, R.B., Jobbágy, E.G., Avissar, R., Roy, S.B., Barrett, D.J., Cook, C.W., Farley, K. a, le
 Maitre, D.C., McCarl, B. a & Murray, B.C. (2005). Trading water for carbon with
 biological carbon sequestration. *Science* **310**, 1944–7.
- Kanowski, J.J., Reis, T.M., Catterall, C.P. & Piper, S.D. (2006). Factors Affecting the Use of
 Reforested Sites by Reptiles in Cleared Rainforest Landscapes in Tropical and
 Subtropical Australia. *Restor. Ecol.* 14, 67–76.

- Kay, G., Michael, D., Crane, M. & Okada, S. (2013). A list of reptiles and amphibians from Box
 Gum Grassy Woodlands in south-eastern Australia. *Check List* 9, 476–481.
- Koh, L.P., Sodhi, N.S. & Brook, B.W. (2004). Ecological Correlates of Extinction Proneness in
 Tropical Butterflies. *Conserv. Biol.* 18, 1571–1578.
- Lindenmayer, D., Cunningham, R.B., Macgregor, C., Crane, M., Michael, D., Fischer, J.,
 Montague-Drake, R., Felton, A. & Manning, A. (2008). Temporal changes in vertebrates
 during landscape transformation: a large-scale "natural experiment." *Ecol. Monogr.* 78,
 567–590.
- Lindenmayer, D.B. (2009). *Large-scale landscape experiments: lessons from Tumut*.
 Cambridge, UK: Cambridge University Press.
- Loehle, C., Wigley, T.B., Shipman, P. a., Fox, S.F., Rutzmoser, S., Thill, R.E. & Melchiors, M.A.
 (2005). Herpetofaunal species richness responses to forest landscape structure in
 Arkansas. *For. Ecol. Manage.* 209, 293–308.
- MacKenzie, D. (2005). What are the issues with presence-absence data for wildlife
 managers? *J. Wildl. Manage.* 69, 849–860.
- MacKenzie, D., Nichols, J. & Hines, J. (2003). Estimating site occupancy, colonization, and
 local extinction when a species is detected imperfectly. *Ecology* 84, 2200–2207.
- Michael, D.R., Cunningham, R.B., Donnelly, C.F. & Lindenmayer, D.B. (2012). Comparative
 use of active searches and artificial refuges to survey reptiles in temperate eucalypt
 woodlands. *Wildl. Res.* **39**, 149–162.
- Michael, D.R., Cunningham, R.B. & Lindenmayer, D.B. (2011). Regrowth and revegetation in
 temperate Australia presents a conservation challenge for reptile fauna in agricultural
 landscapes. *Biol. Conserv.* 144, 407–415.
- 448 Mortelliti, A., Westgate, M. & Lindenmayer, D.B. (2014). Experimental evaluation shows
 449 limited influence of pine plantations on the connectivity of highly fragmented bird
 450 populations. J. Appl. Ecol.
- Mott, B., Alford, R. a. & Schwarzkopf, L. (2010). Tropical reptiles in pine forests: Assemblage
 responses to plantations and plantation management by burning. *For. Ecol. Manage.* **259**, 916–925.
- Ockinger, E., Schweiger, O., Crist, T.O., Debinski, D.M., Krauss, J., Kuussaari, M., Petersen,
 J.D., Pöyry, J., Settele, J., Summerville, K.S. & Bommarco, R. (2010). Life-history traits
 predict species responses to habitat area and isolation: a cross-continental synthesis. *Ecol. Lett.* 13, 969–79.
- Paquette, A. & Messier, C. (2010). The role of plantations in managing the world's forests in
 the Anthropocene. *Front. Ecol. Environ.* 8, 27–34.

- 460 Porté, A., Huard, F. & Dreyfus, P. (2004). Microclimate beneath pine plantation, semi461 mature pine plantation and mixed broadleaved-pine forest. *Agric. For. Meteorol.* 126,
 462 175–182.
- 463 Pryke, S. & Samways, M. (2001). Width of grassland linkages for the conservation of
 464 butterflies in South African afforested areas. *Biol. Conserv.* 101, 85–96.
- 465 R Core Team. (2013). R: A language and environment for statistical computing.
- 466 Renjifo, L. (2001). Effect of natural and anthropogenic landscape matrices on the abundance
 467 of subandean bird species. *Ecol. Appl.* 11, 14–31.
- Shine, R., Elphick, M. & Harlow, P. (1997). The influence of natural incubation environments
 on the phenotypic traits of hatchling lizards. *Ecology* 78, 2559–2568.
- Sozio, G., Mortelliti, A. & Boitani, L. (2013). Mice on the move: Wheat rows as a means to
 increase permeability in agricultural landscapes. *Biol. Conserv.* 165, 198–202.
- Vickers, M., Manicom, C. & Schwarzkopf, L. (2011). Extending the cost-benefit model of
 thermoregulation: high-temperature environments. *Am. Nat.* 177, 452–61.
- Vonesh, J.R. (2001). Patterns of Richness and Abundance in a Tropical African Leaf-litter
 Herpetofauna ' **33**, 502–510.
- Wilson, S. & Swan, G. (2010). A Complete Guide to Reptiles of Australia. Sydney: Reed New
 Holland.
- Zuur, A.F., Ieno, E.N., Walker, N.J., Saveliev, A.A. & Smith, G.M. (2009). *Mixed effects models and extensions in ecology with R*. New York: Springer.





Eastern three-toed earless skink Hemiergis talbingoensis



Southern Rainbow Skink Carlia tetradactyla



Fig. 1. Features of the 'Nanangroe experiment' (New South Wales, Australia). A) Treatment
sites are eucalypt habitat patches surrounded by pine plantations. B) Control sites are
eucalypt habitat patches surrounded by grazing land; C-D) Target species; E) Eucalypt
habitat patch surrounded by grazing land (small growing pines visible); F-G) Later stages of *Pinus radiata* plantation development; treatments progressively surrounded by forested
landscape.



- 490 Fig. 2. Map of the study area (Nanangroe, New South Wales, Australia) with sampling sites
- 491 (grey circles = controls; black circles = treatments, black triangles= pine). The dotted area
- 492 corresponds to pine plantations, whereas the white area corresponds to grazed field.





495 Fig. 3. Model predictions (including 95% confidence intervals) based on model averaged

- 496 estimates of top ranking models (ΔAIC<2) for the Southern Rainbow Skink Carlia
- 497 tetradactyla and the Eastern Three-toed Earless Skink Hemiergis talbingoensis . Carlia
- 498 tetradactyla responded negatively to pine plantations (decrease in colonisation probability
- 499 in sites surrounded by pine plantations), whereas *H. talbingoensis* responded positively
- 500 (increase in colonisation probability in sites surrounded by pine plantations).

503	Table 1. Model ranking according to Δ AIC (delta Akaike Information Criterion); only models
504	<4 are shown. Ψ = probability of a site being occupied during the first survey, Υ = probability
505	of colonization; ϵ = probability of extinction, p = detection probability; Vegtype = vegetation
506	type (categories listed in methods); Y = year (categorical covariate); Yn = year (numeric
507	covariate); H = tree cover within 250 m buffer; nPars = number of estimated parameters; R^2
508	= Nagelkerke's coefficient of determination; (.) = constant model (no covariate).

Species	model	nPars	ΔΑΙΟ	R ²
Carlia tetradactyla	Ψ(Vegtype)Ƴ(T)ε(.)p(Y)	12	0.00	0.46
	Ψ(Vegtype)Υ(T)ε(Yn)p(Y)	13	1.16	0.47
	Ψ(Vegtype)Ƴ(T)ε(H)p(Y)	13	1.96	0.46
	Ψ(Vegtype)Υ(T)ε(T)p(Y)	13	2.00	0.46
	Ψ(Vegtype)Υ(T)ε(T*Yn)p(Y)	15	2.65	0.48
	Ψ(Vegtype)Υ(Η)ε(.)p(Y)	12	3.59	0.44
	Ψ(Vegtype)Υ(T*Yn)ε(.)p(Y)	14	3.64	0.46
	Ψ(Vegtype)Υ(.)ε(.)p(Υ)	11	3.79	0.43
Hemiergis				
talbingoensis	Ψ(Vegtype)ϒ(T)ε(.)p(Yn)	8	0.00	0.19
	Ψ(Vegtype)Υ(T)ε(T)p(Yn)	9	1.18	0.20
	Ψ(Vegtype)ϒ(T)ε(Yn)p(Yn)	9	1.80	0.19
	Ψ(Vegtype)Υ(T)ε(H)p(Yn)	9	1.89	0.19
	Ψ(Vegtype)Υ(T*Yn)ε(.)p(Yn)	10	3.15	0.20
	Ψ(Vegtype)Υ(.)ε(T)p(Yn)	8	3.50	0.16

- 513 Table S1. Life-history and ecological traits of the Southern Rainbow Skink *Carlia tetradactyla*
- and the Eastern Three-toed Earless Skink *Hemiergis talbingoensis*. Data was collated from
- the literature (Greer, 1989; Brown, 1991; Wilson & Swan, 2010).

	Southern Rainbow Skink (Carlia	Three-toed Earless Skink
	tetradactyla)	(Hemiergis talbingoensis)
Snout-Vent Length (mm)	63	60
Zoogeographical distribution	Bassian	Bassian
Life-form	Terrestrial	Fossorial
Common shelter site	Logs, rocks and leaf Litter	Logs, rocks and debris
Activity patterns	Diurnal	Diurnal but possibly
		crepuscular and nocturnal (D
		Michael pers. obs)
Mode of thermoregulation	Heliotherm	Thigmotherm
Mode of reproduction	Oviparous	Viviparous
Clutch/offspring size	2	3
Mating season	Spring	Summer
Ovideposition/Neonatal		
emergence	December/January	February/March
Reproduction frequency/year	1	1
Foraging mode	Active	Active
Dietary preference	Arthropods	Ant/Termites
Operative body temperature	25-32 ¹	20
Critical thermal min	*	6.8
Critical thermal max	42	39.3
Not available, based on co-gene	ric species (Greer, 1989; Vickers, Ma	nicom & Schwarzkopf,

517 2011)

518 ^{*}Not available,

519

Table S2. Number of sites of the Nanangroe study highlighting the factorial study design(Table modified from Lindenmayer *et al.* 2008).

Site	Context	Cohort	No. edges	No. of replicates
	Pinus radiata			
Woodland	plantation	1998	1-2	3
	Pinus radiata			
Woodland	plantation	1998	3-4	16
	Pinus radiata			
Woodland	plantation	2000	1-2	6
	Pinus radiata			
Woodland	plantation	2000	3-4	25
Woodland	Grazing land			55

527 Table S3. Main characteristics of 115 sampled sites. Context: treatment = *Eucalypt* patches surrounded by pine plantation; control = *Eucalypt* patches surrounded by grazing land; pine 528 control = sites within pine plantations. Tree cover (standardized) measured in a 250 m 529 circular buffer surrounding the site. Vegetation type: 1 = red box and red stringybark 530 (codominant) with apple box (E. bridgesiana), long-leaf box (E. goniocalyx), and broad-531 532 leaved peppermint (E. dives); (2) mountain swamp gum (E. camphora) and other kinds of vegetation (e.g., river oak Allocausarina cunninghamiana); (3) yellow box, white box, red 533 stringybark (codominant), and Blakely's red gum. 534

535

Site code	Experimental design	Tree cover	Vegetation type
AWA-1	Control	-0.162	3
AWA-10	Control	1.412	3
AWA-11	Control	0.709	3
AWA-2	Control	-0.157	3
AWA-3	Control	-0.808	3
AWA-4	Control	0.817	3
AWA-5	Control	0.086	3
AWA-6	Control	-0.014	3
AWA-7	Control	-0.110	3
AWA-8	Control	-0.436	3
AWA-9	Control	0.723	3
GRE-1	Control	-0.007	3
GRE-2	Control	0.133	3
GRE-3	Control	-0.407	3
GRE-4	Control	0.705	3
JWA-1	Control	0.416	3
JWA-10	Control	2.372	3
JWA-11	Control	2.835	3
JWA-2	Control	0.120	3
JWA-3	Control	-0.454	3
JWA-4	Control	0.747	3
JWA-5	Control	1.760	3
JWA-6	Control	-0.256	3
JWA-7	Control	-0.514	3
JWA-8	Control	0.498	3
JWA-9	Control	1.113	3
KEA-1	Control	0.547	3
KEA-2	Control	-0.134	2
KEA-3	Control	0.278	3
KEA-4	Control	1.281	3
KEA-5	Control	0.031	3
KEA-6	Control	-0.367	3

LUF-1 Control -0.571 3 LUF-10 Control -0.681 3 LUF-11 Control -0.846 3 LUF-12 Control -0.467 3 LUF-13 Control 1.029 3 LUF-14 Control 0.603 3 LUF-2 Control 1.209 3 LUF-3 Control -0.194 3 LUF-5 Control -0.646 3 LUF-6 Control -1.251 2 LUF-7 Control -0.896 3 LUF-9 Control -0.958 3 LUF-9 Control 1.642 3 SKI-1 Control 1.642 3 SKI-10 Control 1.642 3 SKI-10 Control 1.262 3 SKI-1 Control 1.262 3 SKI-3 Control -0.282 3 SKI-5 Control				
LUF-10 Control -0.681 3 LUF-11 Control -0.846 3 LUF-12 Control 1.029 3 LUF-13 Control 1.029 3 LUF-14 Control 0.603 3 LUF-2 Control -0.194 3 LUF-3 Control -0.646 3 LUF-6 Control -1.251 2 LUF-7 Control -0.896 3 LUF-7 Control -0.896 3 LUF-9 Control -0.958 3 SKI-1 Control 1.642 3 SKI-1 Control 1.642 3 SKI-2 Control 1.498 3 SKI-3 Control 1.262 3 SKI-5 Control -0.123 3 SKI-6 Control -0.123 3 SKI-7 Control 1.510 6 PIN-1 Pine control	LUF-1	Control	-0.571	3
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SKI-6 Control -0.123 3 SKI-7 Control 0.809 3 SKI-8 Control 1.755 1 SKI-9 Control 2.022 3 PIN-1 Pine control 0.610 6 PIN-10 Pine control -1.510 6 PIN-2 Pine control -1.264 6 PIN-3 Pine control -1.537 6 PIN-4 Pine control -1.568 6 PIN-5 Pine control -1.577 6 PIN-6 Pine control -1.577 6 PIN-7 Pine control -1.577 6 PIN-8 Pine control -1.577 6 PIN-9 Pine control -1.577 6 BUN-1 Treatment -1.309 3 BUN-2 Treatment -1.266 3 COT-10 Treatment 0.725 3	SKI-5	Control	-0.282	3
SKI-7 Control 0.809 3 SKI-8 Control 1.755 1 SKI-9 Control 2.022 3 PIN-1 Pine control 0.610 6 PIN-10 Pine control -1.510 6 PIN-2 Pine control -1.264 6 PIN-3 Pine control -1.537 6 PIN-4 Pine control -1.568 6 PIN-5 Pine control -1.577 6 PIN-6 Pine control -1.577 6 PIN-7 Pine control -1.577 6 PIN-8 Pine control -1.577 6 PIN-9 Pine control -1.577 6 PIN-9 Pine control -1.577 6 PUN-9 Pine control -1.577 6 BUN-1 Treatment -1.309 3 BUN-2 Treatment -1.266 3 COT-10 Treatment 0.725 3	SKI-6	Control	-0.123	3
SKI-8 Control 1.755 1 SKI-9 Control 2.022 3 PIN-1 Pine control 0.610 6 PIN-10 Pine control -1.510 6 PIN-2 Pine control -1.264 6 PIN-3 Pine control -1.537 6 PIN-4 Pine control -1.568 6 PIN-5 Pine control -1.577 6 PIN-6 Pine control -1.577 6 PIN-7 Pine control -1.577 6 PIN-8 Pine control -1.577 6 PIN-9 Pine control -1.577 6 PIN-9 Pine control -1.577 6 PIN-9 Pine control -1.577 6 BUN-1 Treatment -1.309 3 BUN-2 Treatment -1.266 3 COT-10 Treatment 0.725 3	SKI-7	Control	0.809	3
SKI-9 Control 2.022 3 PIN-1 Pine control 0.610 6 PIN-10 Pine control -1.510 6 PIN-2 Pine control -1.264 6 PIN-3 Pine control -1.537 6 PIN-4 Pine control -1.568 6 PIN-5 Pine control -1.541 6 PIN-6 Pine control -1.577 6 PIN-7 Pine control -1.577 6 PIN-8 Pine control -1.577 6 PIN-9 Pine control -1.577 6 PIN-9 Pine control -1.577 6 PIN-9 Pine control -1.577 6 PUN-1 Treatment -1.309 3 BUN-1 Treatment -1.266 3 COT-1 Treatment 0.996 1 COT-10 Treatment 0.725 3	SKI-8	Control	1.755	1
PIN-1 Pine control 0.610 6 PIN-10 Pine control -1.510 6 PIN-2 Pine control -1.264 6 PIN-3 Pine control -1.537 6 PIN-4 Pine control -1.568 6 PIN-5 Pine control -1.541 6 PIN-6 Pine control -1.577 6 PIN-7 Pine control -1.577 6 PIN-8 Pine control -1.577 6 PIN-9 Pine control -1.577 6 PIN-9 Pine control -1.577 6 BUN-1 Treatment -1.309 3 BUN-2 Treatment -1.266 3 COT-1 Treatment 0.996 1 COT-10 Treatment 0.725 3	SKI-9	Control	2.022	3
PIN-10 Pine control -1.510 6 PIN-2 Pine control -1.264 6 PIN-3 Pine control -1.537 6 PIN-4 Pine control -1.568 6 PIN-5 Pine control -1.541 6 PIN-6 Pine control -1.577 6 PIN-7 Pine control -1.577 6 PIN-8 Pine control -1.577 6 PIN-9 Pine control -1.577 6 PIN-1 Treatment -1.309 3 BUN-1 Treatment -1.266 3 COT-1 Treatment 0.996 1 COT-10 Treatment 0.725 3	PIN-1	Pine control	0.610	6
PIN-2 Pine control -1.264 6 PIN-3 Pine control -1.537 6 PIN-4 Pine control -1.568 6 PIN-5 Pine control -1.541 6 PIN-6 Pine control -1.577 6 PIN-7 Pine control -1.577 6 PIN-8 Pine control -1.227 6 PIN-9 Pine control -1.577 6 BUN-1 Treatment -1.309 3 BUN-2 Treatment -1.266 3 COT-1 Treatment 0.996 1 COT-10 Treatment 0.725 3	PIN-10	Pine control	-1.510	6
PIN-3 Pine control -1.537 6 PIN-4 Pine control -1.568 6 PIN-5 Pine control -1.541 6 PIN-6 Pine control -1.577 6 PIN-7 Pine control -1.577 6 PIN-8 Pine control -1.227 6 PIN-9 Pine control -1.577 6 BUN-1 Treatment -1.309 3 BUN-2 Treatment -1.266 3 COT-1 Treatment 0.996 1 COT-10 Treatment 0.725 3	PIN-2	Pine control	-1.264	6
PIN-4 Pine control -1.568 6 PIN-5 Pine control -1.541 6 PIN-6 Pine control -1.577 6 PIN-7 Pine control -1.577 6 PIN-8 Pine control -1.227 6 PIN-9 Pine control -1.577 6 BUN-1 Treatment -1.309 3 BUN-2 Treatment -1.266 3 COT-1 Treatment 0.996 1 COT-10 Treatment 0.725 3	PIN-3	Pine control	-1.537	6
PIN-5 Pine control -1.541 6 PIN-6 Pine control -1.577 6 PIN-7 Pine control -1.577 6 PIN-8 Pine control -1.227 6 PIN-9 Pine control -1.577 6 BUN-1 Treatment -1.309 3 BUN-2 Treatment -1.266 3 COT-1 Treatment 0.996 1 COT-10 Treatment 0.725 3	PIN-4	Pine control	-1.568	6
PIN-6 Pine control -1.577 6 PIN-7 Pine control -1.577 6 PIN-8 Pine control -1.227 6 PIN-9 Pine control -1.577 6 BUN-1 Treatment -1.309 3 BUN-2 Treatment -1.266 3 COT-1 Treatment 0.996 1 COT-10 Treatment 0.725 3	PIN-5	Pine control	-1.541	6
PIN-7 Pine control -1.577 6 PIN-8 Pine control -1.227 6 PIN-9 Pine control -1.577 6 BUN-1 Treatment -1.309 3 BUN-2 Treatment -1.266 3 COT-1 Treatment 0.996 1 COT-10 Treatment 0.725 3	PIN-6	Pine control	-1.577	6
PIN-8 Pine control -1.227 6 PIN-9 Pine control -1.577 6 BUN-1 Treatment -1.309 3 BUN-2 Treatment -1.266 3 COT-1 Treatment 0.996 1 COT-10 Treatment 0.725 3	PIN-7	Pine control	-1.577	6
PIN-9 Pine control -1.577 6 BUN-1 Treatment -1.309 3 BUN-2 Treatment -1.266 3 COT-1 Treatment 0.996 1 COT-10 Treatment 0.725 3	PIN-8	Pine control	-1.227	6
BUN-1Treatment-1.3093BUN-2Treatment-1.2663COT-1Treatment0.9961COT-10Treatment0.7253	PIN-9	Pine control	-1.577	6
BUN-2 Treatment -1.266 3 COT-1 Treatment 0.996 1 COT-10 Treatment 0.725 3	BUN-1	Treatment	-1.309	3
COT-1 Treatment 0.996 1 COT-10 Treatment 0.725 3	BUN-2	Treatment	-1.266	3
COT-10 Treatment 0.725 3	COT-1	Treatment	0.996	1
	COT-10	Treatment	0.725	3
COT-2 Treatment 0.414 1	COT-2	Treatment	0.414	1
COT-4 Treatment -0.833 1	COT-4	Treatment	-0.833	1
COT-5 Treatment -0.042 1	COT-5	Treatment	-0.042	1
COT-6 Treatment 0.074 3	COT-6	Treatment	0.074	3
COT-7 Treatment 0.123 3	COT-7	Treatment	0.123	3
COT-8 Treatment 0.710 3	COT-8	Treatment	0.710	3
COT-9 Treatment 2.004 3	COT-9	Treatment	2.004	3
EAB-1 Treatment 1.247 2	EAB-1	Treatment	1.247	2
EAB-2 Treatment -0.113 2	EAB-2	Treatment	-0.113	2

EAB-3	Treatment	-1.258	2
EAB-4	Treatment	0.771	2
EAB-5	Treatment	0.009	2
EAB-6	Treatment	1.161	1
EAB-7	Treatment	-0.408	2
EAB-8	Treatment	-0.422	2
NAN-1	Treatment	-1.206	3
NAN-10	Treatment	-0.515	3
NAN-11	Treatment	-0.526	3
NAN-12	Treatment	0.406	3
NAN-14	Treatment	0.520	3
NAN-15	Treatment	-1.089	2
NAN-16	Treatment	-0.819	3
NAN-18	Treatment	1.517	3
NAN-19	Treatment	-0.922	3
NAN-20	Treatment	0.500	3
NAN-23	Treatment	-0.457	1
NAN-24	Treatment	-0.479	1
NAN-25	Treatment	1.255	3
NAN-26	Treatment	-0.135	3
NAN-27	Treatment	0.945	3
NAN-28	Treatment	0.123	3
NAN-29	Treatment	-0.094	2
NAN-30	Treatment	0.363	2
NAN-31	Treatment	-1.041	1
NAN-34	Treatment	0.325	3
NAN-35	Treatment	-1.149	3
NAN-36	Treatment	0.333	3
NAN-37	Treatment	0.362	3
NAN-38	Treatment	0.809	2
NAN-39	Treatment	0.155	3
NAN-4	Treatment	-1.106	3
NAN-40	Treatment	1.554	3
NAN-5	Treatment	-0.618	3
NAN-6	Treatment	-0.682	3
NAN-7	Treatment	1.397	3
NAN-8	Treatment	-0.390	3



539

- 540 Fig. S1. Artificial refuges (ARs) used to monitor the presence/absence of target species. ARs
- 541 composed of: (1) one double-layered stack of corrugated galvanised steel, (2) four 1-m
- 542 mountain gum *E. dalrympleana*) fence posts covered in mesh, (3) four concrete roof tiles (32
- 543 cm*42 cm).



546 Fig. S2. Open eucalypt woodland patch (above) and pine plantation (below).







549 Fig. S3. Estimates of detection probability for the Southern Rainbow Skink Carlia

550 tetradactyla and the Eastern Three-toed Earless Skink Hemiergis talbingoensis. In Carlia

551 *tetradactyla* the probability of detection is year-specific (categorical covariate year),

s52 whereas in *H. talbingoensis* the probability of detection increases each year (numerical

553 covariate year). Bars (left) and grey areas (right) are 95% confidence intervals.