

1 **Unexpectedly rapid evolution of mandibular shape in hominins**

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3 Raia P.¹, Boggioni M.², Carotenuto F.¹, Castiglione S.¹, Di Febraro M.³, Di Vincenzo F.^{2,4},
4 Melchionna M.¹, Mondanaro A.^{1,5}, Papini A.², Profico A.², Serio C.¹, Veneziano A.², Vero V.A.¹, Rook
5 L.⁵, Meloro C.⁶, Manzi G.²

6

7 ¹*Università degli Studi di Napoli Federico II, Department of Earth Sciences, Environment and*
8 *Resources, L.go San Marcellino 10, 80138, Naples, Italy*

9 ²*Università degli Studi di Roma La Sapienza, Department of Environmental Biology, Piazzale Aldo*
10 *Moro, 5, 00185, Roma, Italy*

11 ³*Università degli Studi del Molise, Department of Biosciences and The Territory, Contrada Fonte*
12 *Lappone, 86090, Pesche, Isernia, Italy*

13 ⁴*Istituto Italiano di Paleontologia Umana, Via Ulisse Aldrovandi, 18, 00197, Roma, Italy*

14 ⁵*Università degli Studi di Firenze, Department of Earth Sciences, Via Giorgio La Pira, 4, 50121,*
15 *Florence, Italy*

16 ⁶*Liverpool John Moores University, School of Natural Science and Psychology, Byrom Street, L3 3AF,*
17 *Liverpool*

18

19 **Author for Correspondence:** Prof. Pasquale Raia, pasquale.raia@unina.it, [http://orcid.org/0000-](http://orcid.org/0000-0002-4593-8006)
20 [0002-4593-8006](http://orcid.org/0000-0002-4593-8006)

21

22 **Abstract**

23 Members of the hominins – namely the so-called ‘australopiths’ and the species of the genus
24 *Homo* – are known to possess short and deep mandibles and relatively small incisors and canines.
25 It is commonly assumed that this suite of traits evolved in early members of the clade in response
26 to changing environmental conditions and increased consumption of tough food items. With the
27 emergence of *Homo*, the functional meaning of mandible shape variation is thought to have been
28 weakened by technological advancements and (later) by the control over fire. In contrast to this
29 expectation, we found that mandible shape evolution in hominins is exceptionally rapid as
30 compared to any other primate clade, and that the direction and rate of shape change (from the
31 ape ancestor) are no different between the australopiths and *Homo*. We deem several factors
32 including the loss of honing complex, canine reduction, and the acquisition of different diets may
33 have concurred in producing such surprisingly high evolutionary rates. This study reveals the
34 evolution of mandibular shape in hominins has strong morpho-functional and ecological
35 significance attached.

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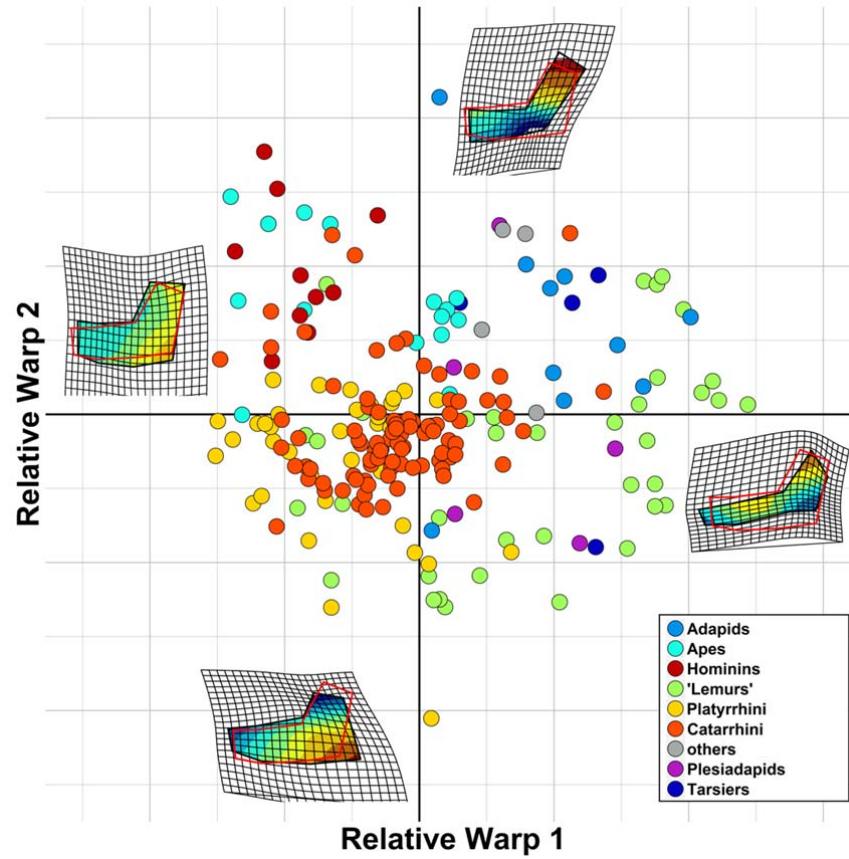
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44 **Main Text**

45 Primates are a large group of mainly arboreal, mostly tropical mammals, ranging in body size from
46 30g in Berthe's mouse lemur (*Microcebus berthae*) to 200kg in male gorilla. In terms of diet,
47 primates are nearly equally variable, being adapted to feed on insects, honey, fruits, leaves, seeds,
48 nuts, and even vertebrate meat. Such wide dietary ambit reflects in the primate mandible and
49 teeth variation. The extent to which diet actually influences the masticatory apparatus in Primates
50 is the subject of intense investigation. It is now well recognised that variation in both mandibular
51 shape and body size were the primary pathways for ecological diversification in fossil, as well as in
52 living primates¹, with diet acting primarily at high taxonomic level, while size has stronger effects
53 between closely related species². Hominins (which include the species belonging to either *Homo*
54 or to the so-called 'australopiths') make no exception to this pattern. Members of the hominin
55 clade have been long noted for their peculiar mandible shape, with short and deep corpus (the
56 horizontal part that bears the tooth-row), low-cusped molars, and reduced incisors and canines.
57 This suite of features is said to allow for a diet including tough food items such as roots and
58 seeds^{3,4}, and is linked to the reduced importance of food processing by the anterior dentition, as
59 compared to fellow apes. This habitus is common to many, but by no means to all of the
60 australopiths^{4,5}, and reached its extreme in the Early Pleistocene hominin *Paranthropus boisei*⁶,
61 consistently with the lifestyle in the grasslands the late australopiths adapted to⁷. While living in
62 open-habitats was common to *Homo* as well⁸, species in our own genus have smaller, thinner-
63 enamel cheek teeth, less robust mandible and zygomatic arches⁹, reduced masticatory muscles
64 and bite force¹⁰, and decreased protrusion of the dental arcade (i.e. prognathism). Most of the
65 differences between *Homo* and the australopiths are believed to relate to the evolution of an
66 extremely large brain in *Homo*, which is responsible for ever increasing technological abilities and,
67 later, for the control over fire. This would have eventually released adaptive pressures on the

68 mandible and teeth, by endowing efficient mechanical food processing before chewing¹¹⁻¹⁴. As
69 such, while the evolution of a mandible shape responsive to a new lifestyle and diet in
70 australopiths should make them no different from the other primates, the robust relationship
71 between mandible shape and diet presumably faded out in *Homo*, with the expected consequence
72 of low evolutionary rate of change in *Homo* mandibles.

73 To verify this hypothesis, we analysed mandibular shape variation in a large sample of
74 primates, ranging from Paleogene 'plesiadapids' to living species, by applying geometric
75 morphometrics (GMM) to the primate mandible under a new phylogenetic comparative method
76 (PCM) approach¹⁵. We assembled a dataset of 731 primate mandible images belonging to 211
77 different species and built a phylogenetic tree for those. We implemented and applied the
78 RRphylo PCM¹⁵, to the shape data ordinated via GMM (Fig. 1). Such method allows retrieving the
79 rate of shape evolution for all the branches in the tree and verifies the existence of shifts in the
80 rate of evolutionary change among clades.



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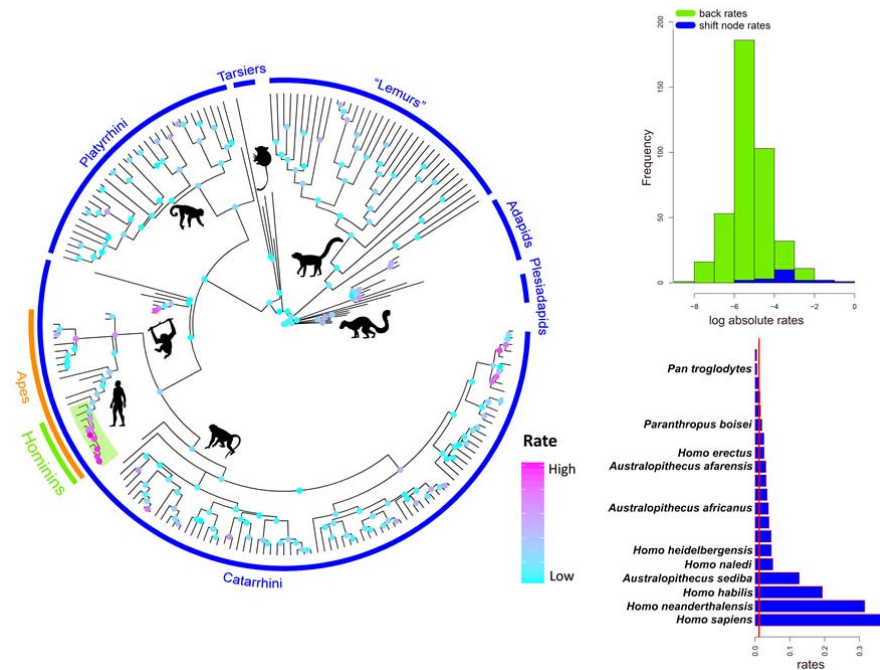
82 **Figure 1.**

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84 **Results**

85 We found the entire hominin clade to stand out among primates, accounting for a

86 disproportionately large share of the clade mandibular shape variation (Fig. 2).



87 **Figure 2.**

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89 More importantly, hominins represent the only instance of (multivariate) rate shift in mandibular
 90 shape evolution in primates, either according to RRphylo, or by using the more traditional,
 91 multivariate Brownian rate variation approach (Fig. 2). This result does not depend on the tree
 92 topology and branch lengths we adopted. We produced 100 random trees where half of the node
 93 ages were allowed to vary in between the ages of their parent and descending nodes.
 94 Contemporarily, in each random tree 50% of the tips were allowed to swap position, up to three
 95 nodes from their actual position (e.g., a *Homo erectus* - *Homo sapiens* sister species relationship,
 96 albeit *Homo neanderthalensis* and *Homo heidelbergensis* are present in the tree, is theoretically
 97 permitted in the random trees). Despite such strong rearrangement of the topology and branch
 98 lengths, the average rate of evolution calculated for the branches of the hominin clade remains
 99 statistically higher than for the remaining part of the tree (see figure S3). Since body size variation
 100 accounts for a large share of ecological diversification within primates¹, and is significantly related
 101 to shape variation (see supplementary material, and figures S6 and S7) we also repeated the

102 analyses after factoring out the effect of size on shape, by using the centroid size of the landmark
103 configuration as a proxy for size. Again, only hominins stand out for having exceptionally large
104 rates (supplementary figure S6).

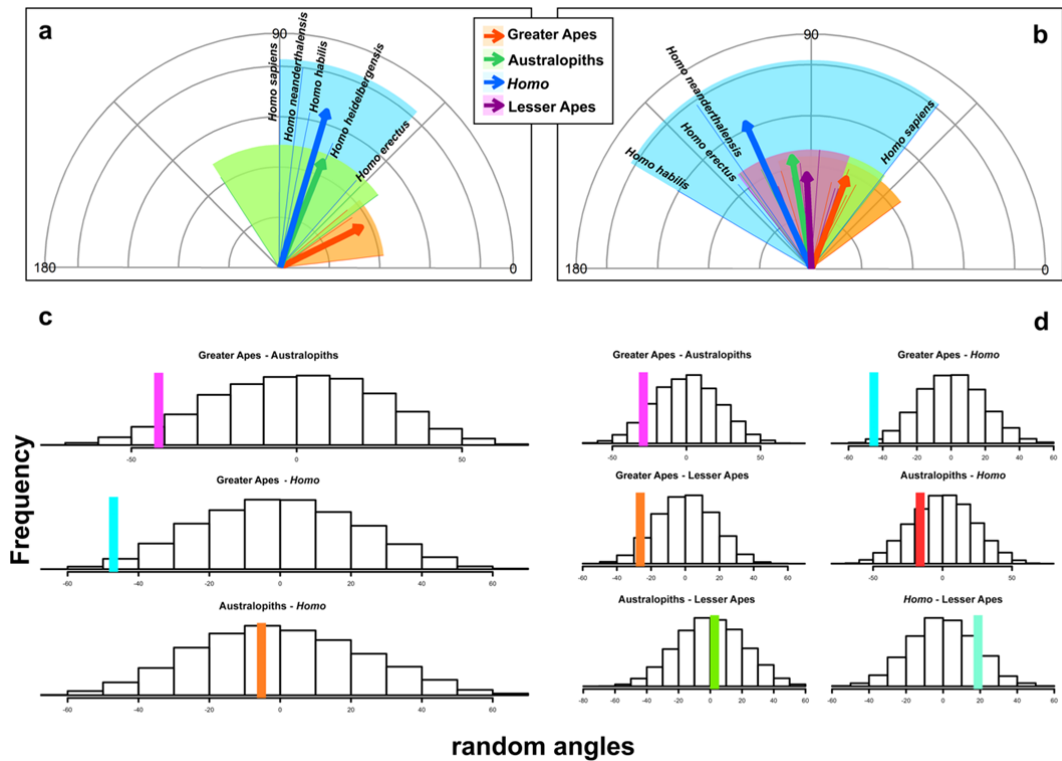
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106 *The direction of shape change, Homo and the australopiths evolved along parallel trajectories of*
107 *shape change*

108 The evolutionary rate represents the magnitude of shape change to the unit time. However, it is
109 silent as per the direction of change. RRphylo produces vectors of regression coefficients
110 (associated to the RW scores) describing the mandible shape change from one node in the tree to
111 the next. Such vectors, besides their size (magnitude) have specific directions, that can be
112 expressed in terms of the angle they form to each other, or to a specific reference. Given the
113 indication of a rate shift in mandible shape evolution accruing to all hominins, we took the most
114 recent common ancestor to the great apes in the tree as the reference and computed the angles
115 between each ape species and such ancestor. Then, we partitioned the great apes in non-hominin
116 apes (here to fore just 'ape' for simplicity), *Homo* species, and australopiths.

117 We found the mean angle of apes to the most recent common ancestor of all great apes was 26.5
118 degrees. For australopiths, the angle was 68.2 degrees, some 42 degrees more. For *Homo* species,
119 the mean angle was 73.5 degrees, 47 degrees wider than apes, but only 5.3 degrees wider than
120 the mean angle for the australopiths (Fig. 3). According to a randomization test, the difference in
121 angles between apes and australopiths, and apes and *Homo* are both significant ($p = 0.032$ and $p =$
122 0.01 , respectively). In contrast, the angle between australopiths and *Homo* is not significant ($p =$
123 0.43). This implies the trajectories of *Homo* and the australopiths are parallel, whereas both
124 diverge significantly from the other greater apes' trajectory (table 1). The same procedure
125 repeated with the inclusion into the analysis of the Hylobatidae (lesser apes) shows similar results,

126 but also indicates there is no significant difference in angles between the trajectories of lesser
 127 apes and the hominins (Fig.3 b, d).



128

129 **Figure 3.**

130

131 **Mandibular shape evolution, dental occlusion, and canine size**

132 Our results show that mandibular shape in hominins evolved faster than in any other primate
 133 clade. Contrary to our expectations, the rate of evolution in *Homo* is not smaller than in the
 134 australopiths, and the direction of the shape change velocity is one and the same for the two
 135 hominin clades. This means that the reason for the unexpected pattern of rapid mandible shape
 136 evolution observed across hominins has to be found among the characteristics shared by the
 137 australopiths and *Homo*. According to a large corpus of available data, the australopiths and *Homo*
 138 differ from each other in terms of habitat preferences, body size, patterns of sexual dimorphism,

139 diet and food processing behaviour^{16,17}. However, tool use has been hypothesized to occur in all
140 early hominids, including australopiths¹⁸⁻²⁰. Such emphasis on mechanical food processing might
141 have caused parallel evolutionary changes in the mandible of hominins. Relevant dental features
142 shared by all hominins are the reduction of maxillary canines crown height, reduced sexual
143 dimorphism²¹, and loss of the honing capacity of the C/P₃ complex²², which by contrast represents
144 a nearly ubiquitous and stable adaptation in nonhuman anthropoids. As compared to the greater
145 apes, all hominins evolved after *A. anamensis* also share a derived temporomandibular joint²³,
146 that allows for a peculiar forward translation and rotation of the mandible during mouth opening
147 in increase gape^{24,25}, and show strongly reduced anterior dentition (incisors and canines), shorter
148 mandibular corpus with more divergent rami and an increase in the absolute and relative size and
149 complexity of the post-canine dentition. The evolutionary emergence of these features has been
150 related to dietary shifts, sexual selection, or a combination of both^{26,27}. Stelzer et al.²⁸ suggest that
151 the reduction in incisors size, and the assumption of the parabolic dental arcade in *Homo* was due
152 to canine and diastema reduction, rather than being selected per se. In turn, whereas usually
153 interpreted as evolving under sexual selection, canine size in male hominins is functionally linked
154 to an increase in mechanical efficiency of the jaws, in order to preserve gape and bite force^{21,29-31}.
155 Hylander^{21,30} argued that in hominins feeding on tough foods items bite force is increased by a
156 forward shift in the position of the jaw muscles. Yet, this comes at the cost of decreasing gape. The
157 reduced gape thus becomes incompatible with vertically elongated canines, hence with a working
158 C/P₃ honing complex^{21,30}, because the P₃ has to slide forward towards the canine tip, rather than
159 producing sliding friction against the upper canine rear margin. However, there is no evidence that
160 the earliest hominins such as *Sahelanthropus*, *Ardipithecus* and *A. anamensis*, which all show a
161 non-honing C/ P₃ complex, were tough food consumers^{4,26,32,33}. Hylander³⁰ found that among the
162 living catarrhines intersexual differences in the degree of canine overlap and gape are not

163 significant only in *Homo sapiens* and the hylobatids. Inspired by these reports, we repeated the
164 multivariate angle calculation taking lesser apes in consideration. Intriguingly, whereas the
165 trajectories of the two hominin groups remain parallel, and both are significantly or marginally
166 different from the trajectory of the other great apes, hylobatids are not smaller (in multivariate
167 angle) than either hominins or great apes (table 2, table S3). Delezene³¹ showed that since the
168 inception of our own clade (i.e. with the appearance of *Sahelanthropus*, *Orrorin*, and *Ardipithecus*)
169 there was no longer any integration or covariation either between the canines and third lower
170 premolars, which is necessary for efficient honing. While this might have served to increase bite
171 force in early hominins^{34,35}, its most important evolutionary consequence could have been the
172 increased evolvability of premolars and increased pattern of reduction of the anterior dentition,
173 including incisors. Such rapid evolution in the dentition (hence in mandible shape) has profound
174 adaptive significance³⁶. It might have permitted the acquisition, in the later species, of deep
175 mandibular corpus and strong ramus^{25,37} in relationship to tough food consumption^{7,38}.
176 Differences in absolute size and relative position of the cheek teeth link to major changes in the
177 trophic niches of our ancestors during the Plio-Pleistocene^{4,9}, and to the ever more extensive use
178 of stone tools.

179 Even if many aspects of mandibular and dental morphology, as for example the high rami
180 in the mandible of the lineage *A. afarensis* – *P. boisei* and the development of megadontia in the
181 *Paranthropus* are functionally related with some major shift in diet, it is unlikely that food
182 adaptations per se may account for the high rates of mandible shape evolution along the entire
183 hominin lineage. Taking in consideration the differences in both dietary and food processing habits
184 between the australopiths and *Homo*, the vectors of the rates should be divergent, which we
185 found was not the case. Intriguingly, sexual selection cannot explain the very high rates we
186 observed in *Homo sapiens* and *Homo neanderthalensis* that are the species showing the lowest

187 level of sexual dimorphism among primates, and the ostensibly divergent shape in *Homo sapiens*
188 mandible is not shared by the Neanderthals^{36,39}.

189 We propose the reshaping of the mandible, shared by the australopiths and *Homo*, was
190 startled by both biomechanical and “structural” events such as the loss of a functioning of the C/P₃
191 honing complex²². This exaptive condition occurred early in hominin evolution and generated
192 “cascading effects” that were recruited for a number of different adaptations along and across the
193 history of the human clade, in response to the rapid environmental changes recorded in Africa
194 from the Upper Miocene through the Plio-Pleistocene.

195

196 **METHODS**

197 **Geometric Morphometrics of Primate mandibles**

198 We used Geometric Morphometrics (Gmm^{40,41}) to extract morphological data. This method permits to retrieve
199 shape information of anatomical objects after removing non-shape variation (i.e. as related to size, position
200 and orientation of the objects) by applying Generalized Procrustes Superimposition (GPA⁴²). By using the
201 TpsRelw software ver. 1.53 we performed Relative Warps Analysis on aligned coordinates (RWA⁴³) to
202 decompose shape variation into orthogonal axes of maximum variance.

203 For this study we collected (either by taking pictures directly, from digital sources, or from published
204 pictures) 731 digital images of primate hemimandibles, belonging to 211 species (148 extant, 63 extinct).

205 The number of mandibles per species ranges from 1 to 13 (median = 3, mean = 3.48). The requirements for
206 picture inclusion in the dataset were the presence of anatomical regions where landmarks had to be placed,
207 absence of distortions and breakages on the bone, and orientation perpendicular to the picture plane.

208 Fortunately, being the hemimandible a flat bone, these features were easily recognizable, even on samples
209 taken from published resources. The pictures we took directly derive from ref.². We used tpsDig2 software
210 to digitize 9 landmarks as to adequately describe the lower jaw profile (fig. S4). Gmm also returns the
211 Centroid Size (the square root of the sum of squared distances between each landmark and the centroid of

212 each configuration), a metric that permits to get back the information related to size that are removed by
 213 GPA. We regressed the natural logarithm of centroid size (Incs) and ln body mass estimates taken from the
 214 literature, to assess whether Incs works good as a proxy for body size. The regression is highly significant
 215 and positive (slope = 0.300, $R^2 = 0.844$, $p < 0.001$, fig. S5). Shape variance was decomposed into 14 axes
 216 (Relative Warps). We performed the Gmm analyses twice: on the full dataset, and on a dataset deprived from
 217 pictures we obtained from literature. The former dataset (FULL) consists of 211 species, the reduced dataset
 218 (SMALL) includes pictures for 158 species (145 extant, 13 extinct). For both dataset, we used for the rate
 219 analyses only the four first largest RW axes, as they capture some 90% of the shape variance.

220

221 **RRphylo**

222 The Phylogenetic Ridge Race Regression version we present here ('RRphylo') develops on phylogenetic
 223 ridge regression as described in¹⁵. It applies penalized ridge regression to the tree and species data. The
 224 difference between the phenotype at each tip and the phenotype at the tree root is the sum of a vector of
 225 phenotypic transformations along the root to tip path, given by equation (1)

226

$$227 \quad \Delta P = \beta_1 l_1 + \beta_2 l_2 + \dots + \beta_n l_n \quad (1)$$

228

229 where the β_{ih} and l_{ih} elements represent the regression coefficient and branch length, respectively, for each i_{th}
 230 branch along the path. As regression slopes, the β coefficients represent the actual rate of phenotypic
 231 transformation along each branch. The matrix solution to find the vector of β coefficients for all the branches
 232 is given by equation (2) ref.⁴⁴;

233

$$\hat{\beta} = (\mathbf{L}^T \mathbf{L} + \lambda \mathbf{I})^{-1} \mathbf{L}^T \mathbf{y} \quad (2)$$

234 where \mathbf{L} is the matrix of tip to root distances of the tree (the branch lengths), having tips as rows. For each
 235 row of \mathbf{L} , entries are zeroes for branches outside the tip to root path, and actual branch lengths for those
 236 branches along the path. The vector $\hat{\mathbf{y}}$ is the vector of phenotypes (tip values), $\hat{\beta}$ is the vector of regression
 237 coefficients, and λ is a penalization factor that avoids perfect predictions of $\hat{\mathbf{y}}$, therefore allowing for the
 238 estimation of the vector of ancestral states, computed as in equation (3):

239

240

$$\hat{a} = \mathbf{L}'\hat{\beta} \quad (3)$$

241

242 where \mathbf{L}' is the node to root path matrix, calculated in analogy to \mathbf{L} , but with nodes as rows.

243

After computing the rates for the tree branches, we searched for shifts in the rates across the tree.

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This rate by clade (RBC) analysis within RRphylo scans the tree to find shifts in the rate of phenotypic

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evolution. There are a number of methods available in literature to apply model-free computations of the

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evolutionary rates, yet some of them do not work with fossil phylogenies (e.g. ref ⁴⁵) or are computationally

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very intensive. With RRphylo, the Brownian rate (σ^2) is calculated for all clades as large as the user specifies

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(in terms of number of tips). Individual nodes (i.e. the clade they subtend to) are arranged according to their

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rates (i.e. in descending σ^2 value). Then, the user is left with two different options to locate a number of

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potential shifts. First, it is possible to specify the number n of shifts to be searched for all combinations of the

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n clades with the n largest σ^2 value, with size 1 to n . For instance, with $n = 3$ RRphylo will search through all

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the eight possible combinations of the 3 nodes with the largest σ^2 values (three combinations with one shift

253

only, one for each node; three combinations of two shifts at two different nodes; and a single combination

254

including all the three shifts for all $n=3$ nodes, plus Brownian motion, which means no shift applied).

255

Alternatively, all selected nodes are partitioned in groups according to their patristic distance, and the

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number of distinct groups with potential shifts is established via bootstrapped cluster analysis of the

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internodes distances. This way the number of potential shifts are located in topologically distinct parts of the

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tree. The resulting number of groups k is thus taken to be equivalent to the number of shift to be searched, by

259

examining all possible combinations of the k nodes with the largest σ^2 values. Of course, it is still possible

260

(and in fact tested) that more than one shift fall in the same region of the tree.

261

Once potential shifts are located, their combinations represent different rate variation models, which are

262

compared to each other (and to a single rate, pure Brownian motion model) by means of restricted maximum

263

likelihood fitted with the function brownieREML in phytools⁴⁶, in the case of a single variable, or mvBM in

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mvMORPH⁴⁷ in the multivariate case. The likelihoods of individual models are contrasted to each other to

265

find the best model by means of likelihood ratio test. It is important to note that whereas RRphylo assigns

266 each branch its own rate of evolution, shifts are located by assessing the likelihood of multi-rate Brownian
267 motion models.

268

269

270 *Accounting for phylogenetic uncertainty in node age and topology*

271 The distribution of evolutionary rates depends on the distribution of branch lengths and on the tree
272 topology⁴⁸. Every phylogenetic tree represents at best a phylogenetic hypothesis, which should be evaluated
273 against alternative topologies, and branch lengths. To account for phylogenetic uncertainty, we wrote an
274 Rcode that changes the tree topology and branch lengths. For every given species, the function swaps the
275 phylogenetic position up to two nodes distance. For instance, the topology ((A,(B,C)),D) could be swapped
276 to the forms ((C,D),(A,B)); (((B,D),A),C) and so on. In addition, each node age is randomly set at any age
277 between the age of its parental node, and the age of its oldest daughter node. We applied the tree swapping
278 function 100 times, computed RRphylo rates at each time, and draw the difference in mean absolute rates
279 between the human clade and the rest of the tree each time.

280

281

282 **Multivariate angle computation of evolutionary rates**

283 Our goal was to verify whether the shape trajectory in *Homo* and australopiths were parallel, and whether
284 they differed from that of non-hominin apes. One limitation with traditional trajectory analysis (e.g. ref. ⁴⁹) is
285 that it ignores phylogenetic relationships. To overcome this problem, we analysed shape trajectories by using
286 phylogenetic ridge regression results.

287 In the context of RRphylo, each branch of the tree has its own rate vector computed. With our data,
288 such rate is composed by the β coefficients of individual RW scores. The magnitude of the rate vector (i.e.
289 the evolutionary rate) is equivalent to the square root of the sum of squared β coefficients. Direction is
290 defined in reference to another vector, computing the angle between the two. Assuming **A** and **B** are two rate
291 vectors the angle between them θ is defined by equation (4):

292
$$\theta = \arccos \frac{A \cdot B}{|A||B|} \quad (4)$$

293 Thus, the path between any node in the tree and a given tip is given by the trigonometric addition of
294 successive vectors, aligned along the node to tip path, which could be summarized as a resultant vector
295 having its own magnitude and angle to the node. For instance, given a species and two successive parental
296 nodes above it, so that the node-to-species path sequence is Node1/Node2/species, the resultant vector \vec{R} is
297 given by equation (5):

298
$$\vec{R} = \vec{A}_{Node1} + \vec{B}_{Node2} + \vec{C}_{species} \quad (5)$$

299 \vec{R} is centered on Node1, so that \vec{R} will be at a certain angle to it. Here, we computed the angle between each
300 ape species and the most recent common ancestor common to all of them (the species to apes most recent
301 common ancestor angles) and contrasted the angles between species partitioned into non-hominin great apes
302 (just ‘apes’ for simplicity), species belonging to *Homo*, and the australopiths. We measured the difference in
303 mean angles between groups and generated a family of 10,000 random differences by shuffling angles
304 between individual species. If the actual mean angle difference between two groups is larger than expected
305 by chance, it means that the between groups trajectories are divergent, otherwise they are parallel.

306

307

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416

417 **Acknowledgements** We are especially in debt with Paolo Piras for the kind and profound
418 assistance with the R scripts for the geometric morphometrics analyses. We are grateful to Luca
419 Russo e Silvia Soncin for kindly providing us with the images used to compose figure 3.

420

421 **Author contributions** MM, AM, SC, VV, AP, AV, and CM collected and contributed to analyse the
422 data. CS and AP performed much of the geometric morphometrics analyses. PR, FDV and GM
423 interpreted the evolutionary meaning of the results. PR and SC wrote the phylogenetic ridge
424 regression codes. PR, SC, AM and CS wrote the multivariate angle function. All of the authors have
425 significantly contributed in producing and analysing the data, and in preparing the manuscript and
426 related material.

427

428 **The authors declare no competing financial and non-financial interests**

429

Figure legends

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431

432 **Figure 1. The major axes of mandibular shape variation in primates, retrieved from GMM. *Homo***
433 **and the australopiths almost exclusively occupy the upper left quadrant of the plot (purple**
434 **circle). At the two extremes of both axes we reported the shape deformation associated to**
435 **these axes, overlaid on the primate consensus shape (in red) and a continuous colour scale**
436 **representing the mandibular areas or more intense deformation, from areas where the**
437 **mandible widens compared to the consensus (in red) to areas where it compresses (in blue). The**
438 **image was generated by using the R package ggplot (<http://ggplot2.org/>) and our own R codes.**

439

440 **Figure 2. The evolutionary rates of mandible shape on the primate tree. The tree on the left**
441 **reports rates computed according to phylogenetic Ridge Regression (coloured dots, scaled**
442 **according to the rate value, from low= cyan, to high rates= magenta). The human clade,**
443 **highlighted with a green semitransparent box, represents the only rate shift as indicated by the**
444 **variable Brownian rate approach. On top right, the phylogenetic Ridge Regression rates (in**
445 **absolute values) computed for the branches of the tree not belonging to the human clade**
446 **(green) are contrasted to rates for the human clade (blue). On bottom right, phylogenetic Ridge**
447 **Regression rates of individual branches of the human clade (in absolute value) plus the human**
448 **clade sister species, the common chimpanzee, are collated in increasing rate value (blue bars),**
449 **and contrasted to the average rate computed over the entire tree (the vertical red line). Bars**
450 **without names correspond to internal nodes of the human clade. The image was generated by**
451 **using the R package ggplot (<http://ggplot2.org/>) and our own R codes. Animal silhouettes were**
452 **available under Public Domain license at phylopic (<http://phylopic.org/>), unless otherwise**
453 **indicated. Specifically, clockwise starting from the bottom, *Macaca***

454 (<http://phylopic.org/image/eedde61f-3402-4f7c-9350-49b74f5e1dba/>); *Homo sapiens*
455 (<http://phylopic.org/image/c089caae-43ef-4e4e-bf26-973dd4cb65c5/>); *Hylobates*
456 (<http://phylopic.org/image/0174801d-15a6-4668-bfe0-4c421fbe51e8/>); *Cebus*
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464 (<https://creativecommons.org/licenses/by-sa/3.0/>) image by Nobu Tamura (modified by
465 Michael Keesey).

467

468 **Figure 3.** Multivariate angle comparisons among non-hominin apes, *Homo* species and the australopiths,
469 assessed through multivariate angles between rate vectors. In (a) angles of *Homo*, australopiths, and
470 non-hominin greater apes (Great Apes) are depicted starting from the common origin (the ancestor of all
471 these species). The range of angles for each group is highlighted: *Homo*, transparent blue; Australopiths,
472 transparent green; Great Apes, transparent orange. Vector length is proportional to actual vector size
473 (i.e. the evolutionary rate). In (b) the same as with (a) but including lesser apes (Hylobatidae) highlighted
474 in transparent purple. In (c) the angles in (a) are tested for significance by shuffling the rates among
475 groups 10,000 times, real differences are indicated by the color bars. In (d) the angles in (b) are tested for
476 significance by shuffling the rates among groups 10,000 times, real differences are indicated by color
477 bars.

478

479 **Tables**

480 **Table 1. Multivariate angle of evolutionary rates. The row names correspond to individual comparisons**
481 **of one group to another. APE = great apes exclusive of hominins, AUS = australopiths, HOM = *Homo***
482 **species.**

comparisons	Difference in angle	p.value
APE_AUS	-41.74	0.06
APE_HOM	-47.025	0.047
AUS_HOM	-5.285	0.603

	APE	AUS	HOM
angle from the origin	26.5	68.24	73.53

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488 **Table 2. Multivariate angle of evolutionary rates. The row names correspond to individual comparisons**
489 **of one group to another. APE = great apes exclusive of hominins, AUS = australopiths, HOM = *Homo***
490 **species, HYLO = lesser apes.**

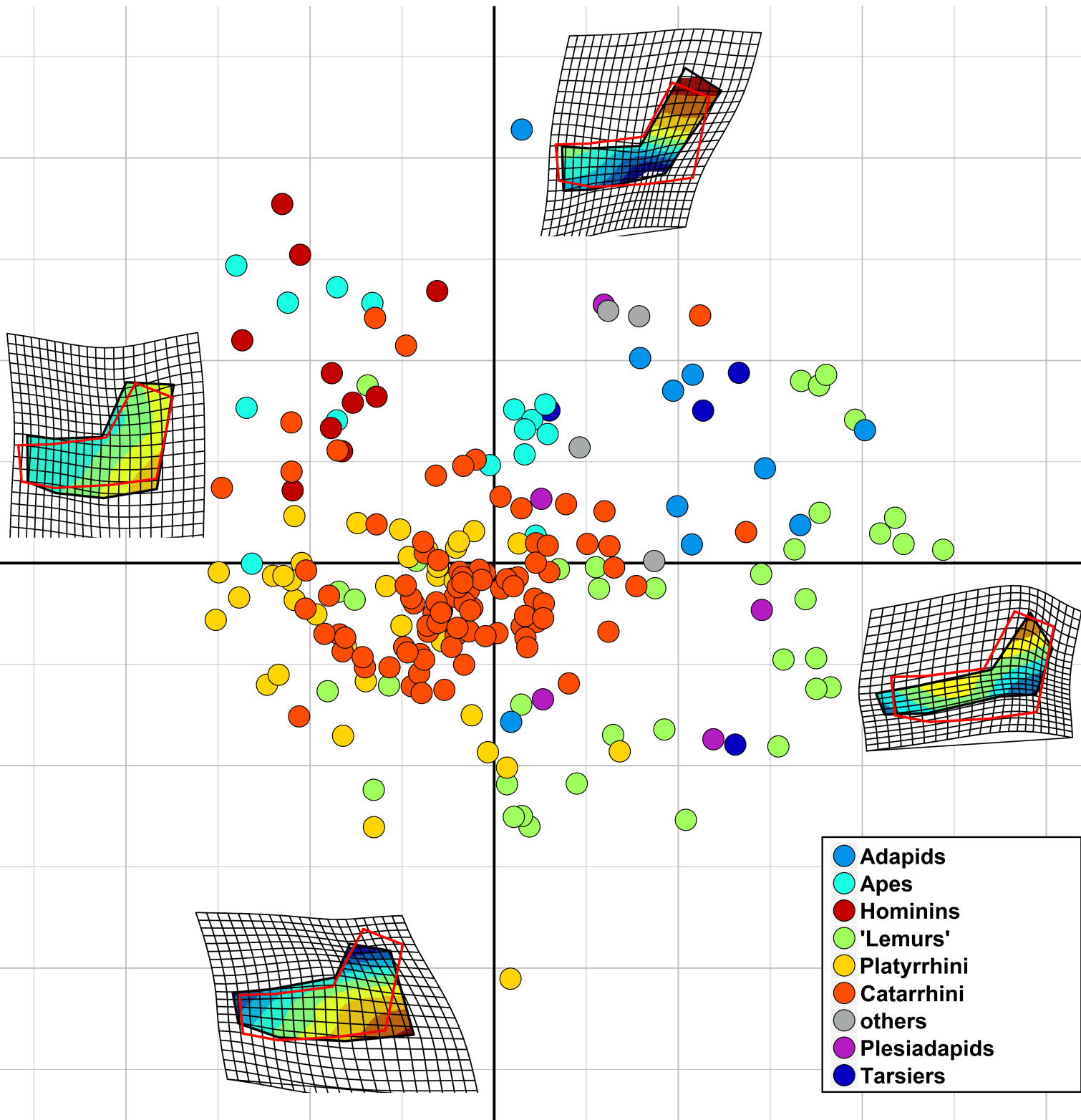
comparisons	Difference in angle	p.value
APE_AUS	-29.057	0.073
APE_HOM	-45.174	0.006
APE_HYLO	-26.33	0.057
AUS_HOM	-16.116	0.242
AUS_HYLO	2.727	0.544
HOM_HYLO	18.844	0.85

	APE	AUS	HOM	HYLO
angle from the origin	69.26	98.31	114.43	95.59

491

Relative Warp 2

Relative Warp 1



- Adapids
- Apes
- Hominins
- 'Lemurs'
- Platyrrhini
- Catarrhini
- others
- Plesiadapids
- Tarsiers

