

OPEN ACCESS JOURNAL AT INIST-CNRS

Gene Section

Review

EIF3A (eukaryotic translation initiation factor 3, subunit A)

Ji-Ye Yin, Zizheng Dong, Jian-Ting Zhang

Department of Pharmacology and Toxicology and IU Simon Cancer Center, Indiana University School of Medicine, Indianapolis, IN, USA (JYY, ZD, JTZ)

Published in Atlas Database: November 2010

Online updated version : http://AtlasGeneticsOncology.org/Genes/EIF3AID40425ch10q26.html DOI: 10.4267/2042/45982

This work is licensed under a Creative Commons Attribution-Noncommercial-No Derivative Works 2.0 France Licence. © 2011 Atlas of Genetics and Cytogenetics in Oncology and Haematology

Identity

Other names: EIF3; EIF3S10; KIAA0139; P167; TIF32; eIF3-p170; eIF3-theta; p180; p185

HGNC (Hugo): EIF3A

Location: 10q26.11

DNA/RNA

Description

The eIF3a gene spans over a region of 46 kbp DNA including 22 coding exons and 2 non-coding exons (exon 2 and exon 10).

Transcription

The eIF3a mRNA consists of about 5256 nucleotides with an open reading frame (ORF) of 4149 bases.

Pseudogene

No pseudogene has been identified.

Protein

Description

Structure: The eIF3a protein consists of 1382 amino acid residues with an apparent molecular weight of ~170 kDa as determined using SDS-PAGE (Pincheira et al., 2001b). Its primary sequence contains a PCI (Proteasome, COP9, Initiation factor 3) domain, a

spectrin domain, and a 10-amino acid repeat domain (Pincheira et al., 2001b). It has phosphorylation sites at Ser-881, Ser-1198, Ser-1336 and Ser-1364 (Damoc et al., 2007). The PCI domain spans from amino acid 405 to 495, which contains purely alpha-helix (Pincheira et al., 2001b). Since most of the proteins containing this domain are part of a multi-protein complex, it is tempting to speculate that this domain may be involved in the interaction of eIF3a with other molecules in eIF3 (Hofmann and Bucher, 1998). The spectrin domain, which consists of 112 amino acids, is a sequence almost identical to spectrin, an actin-binding protein (Pascual et al., 1997). Although the exact function of this domain remains unknown, it may be responsible for the binding of eIF3a to actin filaments (Pincheira et al., 2001a). The 10-amino acid repeat domain spanning 925-1172 amino acids is the largest domain of eIF3a. It can be divided into about 25 repeats of DDDRGPRRGA (Johnson et al., 1997; Pincheira et al., 2001b). This domain has been suggested to contribute to interaction of eIF4B and eIF3a (Methot et al., 1996). Regulatory role in gene expression: eIF3a not only functions as a regular translation initiation factor and participates in translation initiation of global mRNAs, it also regulates the translation of a subset of mRNAs which are involved in cell cycle, tumorigenesis and DNA repair (Yin et al., 2010). It has been observed that overexpression of ectopic eIF3a increases the expression of ribonucleotide reductase

0	200	400	600	800	1000	1200	1400
		PCI		Spectrin	10 AA Repeat		

Schematic presentation of eIF3a domain structure. Human eIF3a consists of 1382 amino acid residues with three putative domains of PCI, spectrin, and 10-amino acid repeat.

M2 (RRM2) and alpha-tubulin, but decreases that of p27kip without affect their mRNA levels (Dong and Zhang, 2003; Dong et al., 2004). Recently, it has also been found that eIF3a suppresses the synthesis of DNA repair proteins including: XPA, XPC, RPA 14, RPA 32 and RPA 70 KDa (Yin et al., unpublished data). Although the detailed mechanism of eIF3a regulation in translational control is yet to be determined, it is thought that eIF3a may regulate these genes at their 5'-and 3'-UTRs (Dong and Zhang, 2003; Dong et al., 2004).

Binding with other molecule: Since eIF3a is the largest subunit of the eIF3 complex, the interaction between eIF3a and other subunits of eIF3 were intensively studied. It can bind with eIF3b (Methot et al., 1997), eIF3c (Valasek et al., 2002), eIF3f (Asano et al., 1997), eIF3h (Asano et al., 1997), eIF3h (Asano et al., 1997), eIF3h (Asano et al., 2003). During the translation initiation, the amino terminal domain of eIF3a can bind with 40S protein RPS0A, while the C terminal domain binds with the 18S rRNA (Valasek et al., 2003). Apart from above molecule, eIF3a has also been shown to interact with eIF4B (Methot et al., 1996), actin (Pincheira et al., 2001a), and cytokeratin 7 (Lin et al., 2001).

Expression

eIF3a is ubiquitously expressed in all human tissues (Nagase et al., 1995; Scholler and Kanner, 1997; Pincheira et al., 2001b). However, its expression is higher in proliferating tissues such as bone marrow, thymus and fetal tissues (Pincheira et al., 2001b).

Localisation

eIF3a has been found in both cytoplasmic and membrane fractions and the cytoplasmic eIF3a appears to be phosphorylated at its serine residues (Pincheira et al., 2001a). However, 70-80% of eIF3a is cytoplasmic.

Function

eIF3a has been shown to play important roles in the biological processes: translational initiation (including generation of ribosomal subunit from 80S ribosomes, 43S pre-initiation complex formation and 48S preinitiation complex formation) (Dong and Zhang, 2006), regulation of mRNA translation (Dong and Zhang, 2003; Dong et al., 2004), differentiation and development (Liu et al., 2007), apoptosis (Nakai et al., 2005), cell cycle regulation (Dong et al., 2009), oncogenesis (Dong and Zhang, 2006; Zhang et al., 2007), and drug response (unpublished observations).

Homology

Centrosomin A and B have strong homology to eIF3a. The spectrin domain is essentially identical to spectrin.

Mutations

Note

Two SNPs (rs10787899 and rs3824830) were found to

be associated with higher risk of breast cancers (Olson et al., 2009).

Implicated in

Breast cancer

Note

eIF3a was overexpressed in breast cancer tissues.

Oncogenesis

The eIF3a was highly expressed in all tested tissues from breast cancer patients compared with normal control tissues, which indicated that it may contribute to the oncogenesis of breast cancer (Bachmann et al., 1997).

Cervical carcinoma

Note

eIF3a was found to be a molecular parameter of predicting cervical carcinoma progression and prognoses.

Prognosis

Patients with high eIF3a expression have better prognosis than those with lower ones, thus it will be useful in predicting cervical cancer prognosis (Dellas et al., 1998).

Gastric carcinoma

Note

eIF3a is an early tumor maker of gastric carcinoma.

Oncogenesis

eIF3a was highly expressed in well differentiated, early invasive stage and no-metastases gastric carcinoma (Chen and Burger, 2004).

Lung cancer

Note

eIF3a is highly expressed in lung cancer compared with normal tissues.

Prognosis

eIF3a expression in human lung cancers negatively correlates with patient response to platinum-based chemotherapy, suggesting that lung cancer patients with higher eIF3a expression level respond better to platinum-based chemotherapy (Yin et al., unpublished findings).

Oncogenesis

eIF3a was over-expressed in all types of human lung cancer. Furthermore, it is ubiquitously highly

expressed in proliferating and developing tissues. This suggested eIF3a may be involved in oncogenesis of lung cancer (Pincheira et al., 2001b).

Esophagus squamous-cell carcinoma

Note

eIF3a may be a biomaker of esophagus squamous-cell carcinoma.

Prognosis

Patients with higher eIF3a expression have better

overall survival and fewer tumor metastases than those with lower ones (Chen and Burger, 1999).

References

Nagase T, Seki N, Tanaka A, Ishikawa K, Nomura N. Prediction of the coding sequences of unidentified human genes. IV. The coding sequences of 40 new genes (KIAA0121-KIAA0160) deduced by analysis of cDNA clones from human cell line KG-1. DNA Res. 1995 Aug 31;2(4):167-74, 199-210

Méthot N, Song MS, Sonenberg N. A region rich in aspartic acid, arginine, tyrosine, and glycine (DRYG) mediates eukaryotic initiation factor 4B (eIF4B) self-association and interaction with eIF3. Mol Cell Biol. 1996 Oct;16(10):5328-34

Asano K, Vornlocher HP, Richter-Cook NJ, Merrick WC, Hinnebusch AG, Hershey JW. Structure of cDNAs encoding human eukaryotic initiation factor 3 subunits. Possible roles in RNA binding and macromolecular assembly. J Biol Chem. 1997 Oct 24;272(43):27042-52

Bachmann F, Bänziger R, Burger MM. Cloning of a novel protein overexpressed in human mammary carcinoma. Cancer Res. 1997 Mar 1;57(5):988-94

Johnson KR, Merrick WC, Zoll WL, Zhu Y. Identification of cDNA clones for the large subunit of eukaryotic translation initiation factor 3. Comparison of homologues from human, Nicotiana tabacum, Caenorhabditis elegans, and Saccharomyces cerevisiae. J Biol Chem. 1997 Mar 14;272(11):7106-13

Méthot N, Rom E, Olsen H, Sonenberg N. The human homologue of the yeast Prt1 protein is an integral part of the eukaryotic initiation factor 3 complex and interacts with p170. J Biol Chem. 1997 Jan 10;272(2):1110-6

Pascual J, Castresana J, Saraste M. Evolution of the spectrin repeat. Bioessays. 1997 Sep;19(9):811-7

Scholler JK, Kanner SB. The human p167 gene encodes a unique structural protein that contains centrosomin A homology and associates with a multicomponent complex. DNA Cell Biol. 1997 Apr;16(4):515-31

Dellas A, Torhorst J, Bachmann F, Bänziger R, Schultheiss E, Burger MM. Expression of p150 in cervical neoplasia and its potential value in predicting survival. Cancer. 1998 Oct 1;83(7):1376-83

Hofmann K, Bucher P. The PCI domain: a common theme in three multiprotein complexes. Trends Biochem Sci. 1998 Jun;23(6):204-5

Chen G, Burger MM. p150 expression and its prognostic value in squamous-cell carcinoma of the esophagus. Int J Cancer. 1999 Apr 20;84(2):95-100

Valásek L, Hasek J, Trachsel H, Imre EM, Ruis H. The Saccharomyces cerevisiae HCR1 gene encoding a homologue of the p35 subunit of human translation initiation factor 3 (eIF3) is a high copy suppressor of a temperature-sensitive mutation in the Rpg1p subunit of yeast eIF3. J Biol Chem. 1999 Sep 24;274(39):27567-72

Lin L, Holbro T, Alonso G, Gerosa D, Burger MM. Molecular interaction between human tumor marker protein p150, the largest subunit of eIF3, and intermediate filament protein K7. J Cell Biochem. 2001;80(4):483-90

Pincheira R, Chen Q, Huang Z, Zhang JT. Two subcellular localizations of eIF3 p170 and its interaction with membranebound microfilaments: implications for alternative functions of p170. Eur J Cell Biol. 2001a Jun;80(6):410-8 Pincheira R, Chen Q, Zhang JT. Identification of a 170-kDa protein over-expressed in lung cancers. Br J Cancer. 2001b Jun 1;84(11):1520-7

Valásek L, Nielsen KH, Hinnebusch AG. Direct eIF2-eIF3 contact in the multifactor complex is important for translation initiation in vivo. EMBO J. 2002 Nov 1;21(21):5886-98

Dong Z, Zhang JT. EIF3 p170, a mediator of mimosine effect on protein synthesis and cell cycle progression. Mol Biol Cell. 2003 Sep;14(9):3942-51

Dong Z, Zhang JT. Initiation factor eIF3 and regulation of mRNA translation, cell growth, and cancer. Crit Rev Oncol Hematol. 2006 Sep;59(3):169-80

Valásek L, Mathew AA, Shin BS, Nielsen KH, Szamecz B, Hinnebusch AG. The yeast eIF3 subunits TIF32/a, NIP1/c, and eIF5 make critical connections with the 40S ribosome in vivo. Genes Dev. 2003 Mar 15;17(6):786-99

Chen G, Burger MM. p150 overexpression in gastric carcinoma: the association with p53, apoptosis and cell proliferation. Int J Cancer. 2004 Nov 10;112(3):393-8

Dong Z, Liu LH, Han B, Pincheira R, Zhang JT. Role of eIF3 p170 in controlling synthesis of ribonucleotide reductase M2 and cell growth. Oncogene. 2004 May 6;23(21):3790-801

Nakai Y, Shiratsuchi A, Manaka J, Nakayama H, Takio K, Zhang JT, Suganuma T, Nakanishi Y. Externalization and recognition by macrophages of large subunit of eukaryotic translation initiation factor 3 in apoptotic cells. Exp Cell Res. 2005 Sep 10;309(1):137-48

Mayeur GL, Fraser CS, Peiretti F, Block KL, Hershey JW. Characterization of eIF3k: a newly discovered subunit of mammalian translation initiation factor eIF3. Eur J Biochem. 2003 Oct;270(20):4133-9

Damoc E, Fraser CS, Zhou M, Videler H, Mayeur GL, Hershey JW, Doudna JA, Robinson CV, Leary JA. Structural characterization of the human eukaryotic initiation factor 3 protein complex by mass spectrometry. Mol Cell Proteomics. 2007 Jul;6(7):1135-46

Liu Z, Dong Z, Yang Z, Chen Q, Pan Y, Yang Y, Cui P, Zhang X, Zhang JT. Role of eIF3a (eIF3 p170) in intestinal cell differentiation and its association with early development. Differentiation. 2007 Sep;75(7):652-61

Zhang L, Pan X, Hershey JW. Individual overexpression of five subunits of human translation initiation factor eIF3 promotes malignant transformation of immortal fibroblast cells. J Biol Chem. 2007 Feb 23;282(8):5790-800

Dong Z, Liu Z, Cui P, Pincheira R, Yang Y, Liu J, Zhang JT. Role of eIF3a in regulating cell cycle progression. Exp Cell Res. 2009 Jul 1;315(11):1889-94

Olson JE, Wang X, Goode EL, Pankratz VS, Fredericksen ZS, Vierkant RA, Pharoah PD, Cerhan JR, Couch FJ. Variation in genes required for normal mitosis and risk of breast cancer. Breast Cancer Res Treat. 2010 Jan;119(2):423-30

Yin JY, Dong Z, Liu ZQ, Zhang JT. Translational control gone awry: a new mechanism of tumorigenesis and novel targets of cancer treatments. Biosci Rep. 2010 Oct;31(1):1-15

This article should be referenced as such:

Yin JY, Dong Z, Zhang JT. EIF3A (eukaryotic translation initiation factor 3, subunit A). Atlas Genet Cytogenet Oncol Haematol. 2011; 15(7):544-546.