

Solid Tumour Section

Mini Review

Lymphangiomyomatosis

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Identity

Alias: LAM

Note: Lymphangiomyomatosis (LAM) is a multi-system disease, affecting predominantly premenopausal women, that is characterized by proliferation of abnormal smooth muscle-like cells (LAM cells) leading to the formation of lung cysts, fluid-filled cystic tumors in the axial lymphatics (e.g., lymphangiomyomas), and abdominal tumors, primarily in the kidneys, comprising adipose cells, vascular structures and LAM cells (e.g., angiomyolipomas).

Clinics and pathology

Disease

Pulmonary disease

Note

Pulmonary disease is the main cause of morbidity and mortality. LAM usually presents with progressive breathlessness or with spontaneous recurrent pneumothorax, chylous effusions (chylothorax and ascites), or hemorrhage within an angiomyolipoma. Computed tomography scans show numerous thin-walled cysts throughout the lungs (Figure 1A and 1B), renal angiomyolipomas (Figure 2), and lymphangiomyomas (Figure 3). Pulmonary function abnormalities include airflow obstruction and gas exchange abnormalities.

Lung lesions in LAM are characterized by nodular infiltrates and clusters of LAM cells near cystic lesions and along pulmonary blood vessels, lymphatics, and bronchioles (Figure 4A and 4B). Two types of LAM cells have been described: small spindle-shaped cells and larger, epithelioid-like

cells with abundant cytoplasm. Both types of cells react with antibodies against smooth muscle cell-specific antigens (e.g., smooth muscle α -actin, vimentin, desmin) (Figure 5). The epithelioid LAM cells react with HMB-45, a monoclonal antibody that recognizes gp100, a premelanosomal protein (Figures 5, 6 and 7). The spindle-shaped cells are more likely to react with anti-proliferation cell nuclear antigen (PCNA) antibodies, suggesting a more proliferative state. Receptors for estrogen, progesterone, and growth factors have been identified in LAM cells. LAM cells appear to have neoplastic properties and may be capable of metastasis. In addition to their presence in lungs, lymphatics and kidneys, LAM cells have been isolated from blood, chyle, and urine.

Etiology

The tumor suppressor genes TSC1 and TSC2 have been implicated in the etiology of LAM, as mutations and loss of heterozygosity in the TSC genes have been detected in LAM cells (Figure 7). TSC1 encodes hamartin, a protein that plays a role in the reorganization of the actin cytoskeleton, and TSC2 encodes tuberin, a protein with roles in cell growth and proliferation. TSC1 and TSC2 may function both individually and as a cytosolic complex.

Epidemiology

LAM occurs in about one third of women with tuberous sclerosis complex (TSC), an autosomal dominant syndrome characterized by hamartoma-like tumor growths in various organs, cerebral calcifications, seizures, and mental retardation, that occurs in one of 5800 live births. Sporadic LAM is a relatively uncommon disease with a prevalence that has been estimated at 1-2.6/million women.

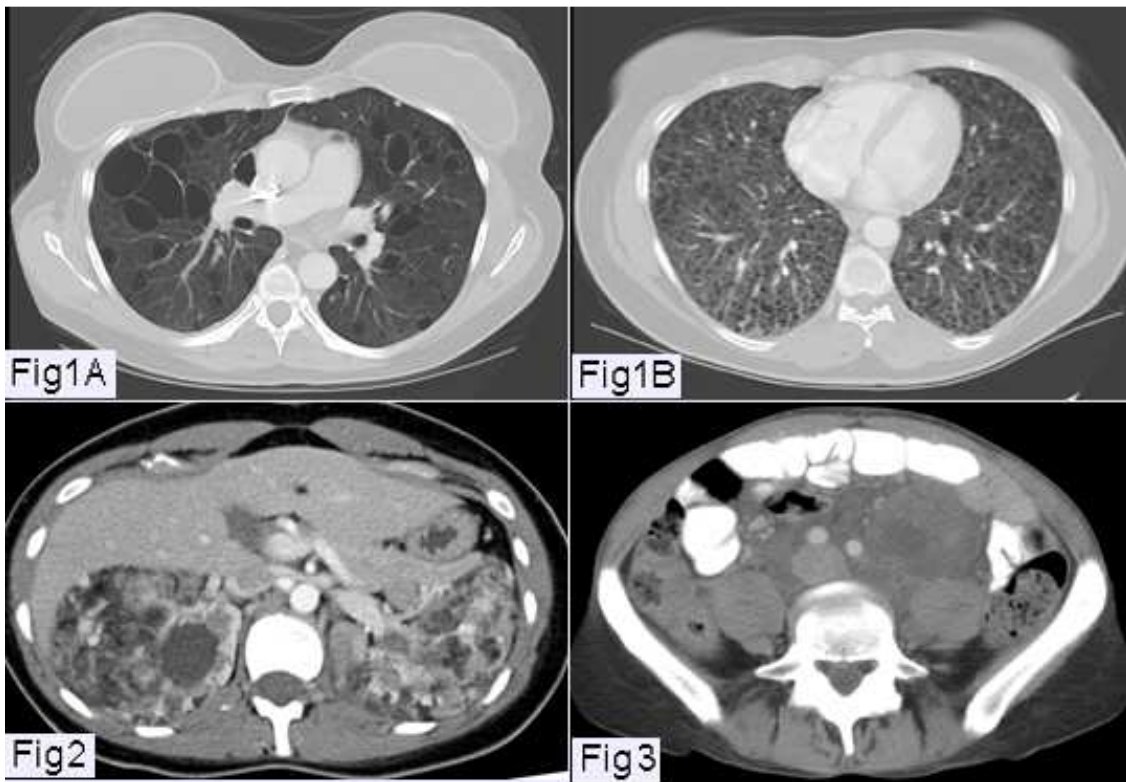


Figure 1. Chest CT scan of a patient with LAM (A) showing numerous thin-walled cysts distributed throughout the lungs. (B) The lung parenchyma is almost completely replaced by very small cysts.
 Figure 2. Abdominal CT scan of a patient with LAM showing angiomyolipomas involving both kidneys.
 Figure 3. Abdominal CT scan of a patient with LAM showing a large lymphangiomyoma located in the retroperitoneal area and surrounding the aorta and inferior vena cava.

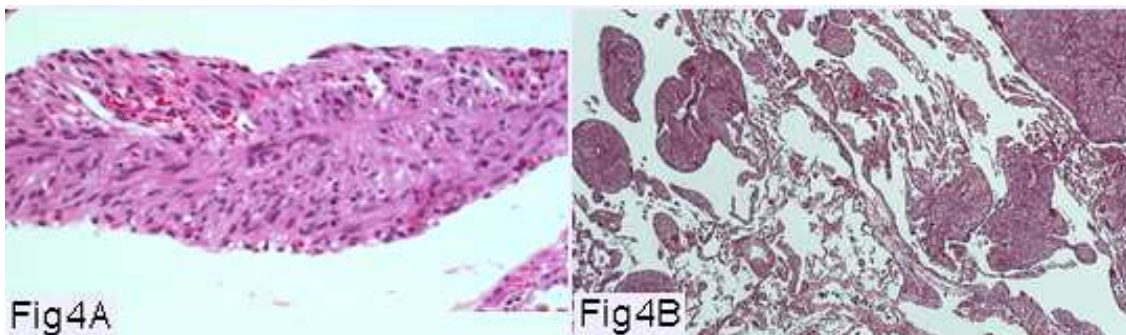


Figure 4 A and B. LAM nodule comprising spindle-shaped cells and larger epithelioid cells (A). Nodules of various sizes (B) are seen in involved lung (hematoxylin-eosin; original magnification x50).

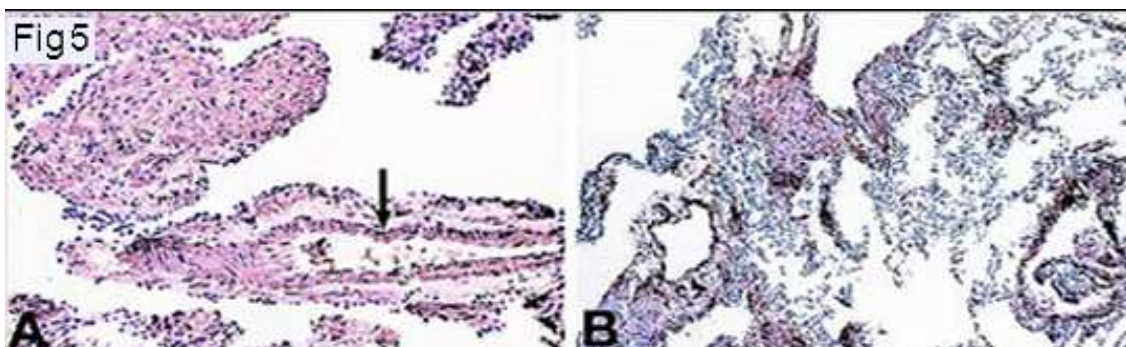
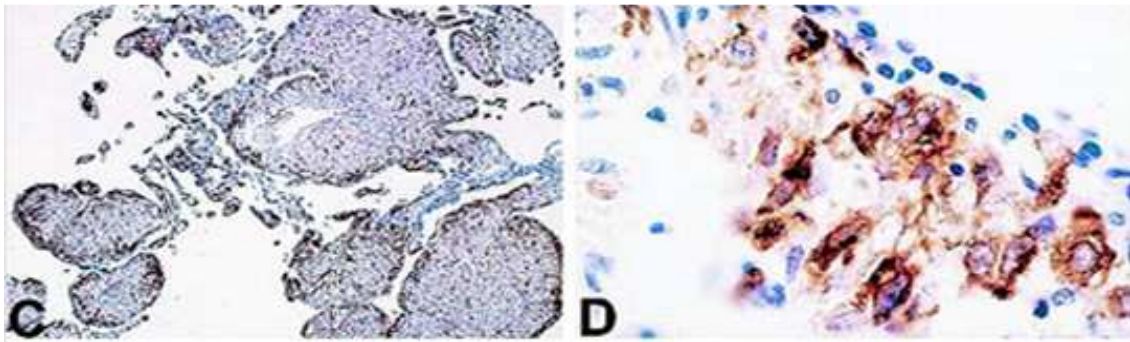


Figure 5. Immunohistochemistry of LAM cells. Immunoperoxidase method and counterstaining with hematoxylin. A and B: Immunoreactivity with α -smooth muscle actin antibodies. LAM cells show strong reactivity (A). Pulmonary vascular smooth muscle cells are also strongly positive (arrow). LAM cells in the walls of the lung cysts are also strongly reactive (arrow) (B) (original magnification x250 for each).



C: Immunoreactivity with monoclonal antibody HMB-45. Immunoreactive cells are distributed in the periphery of LAM lung nodules (arrow) (original magnification x250). D: Immunoreactivity with monoclonal antibody HMB-45. Higher-magnification view of tissue shown in C. A strong granular reaction is present in large epithelioid LAM cells adjacent to epithelial cells covering LAM lung nodules (arrow) (D) (original magnification x1000).

Fig 6

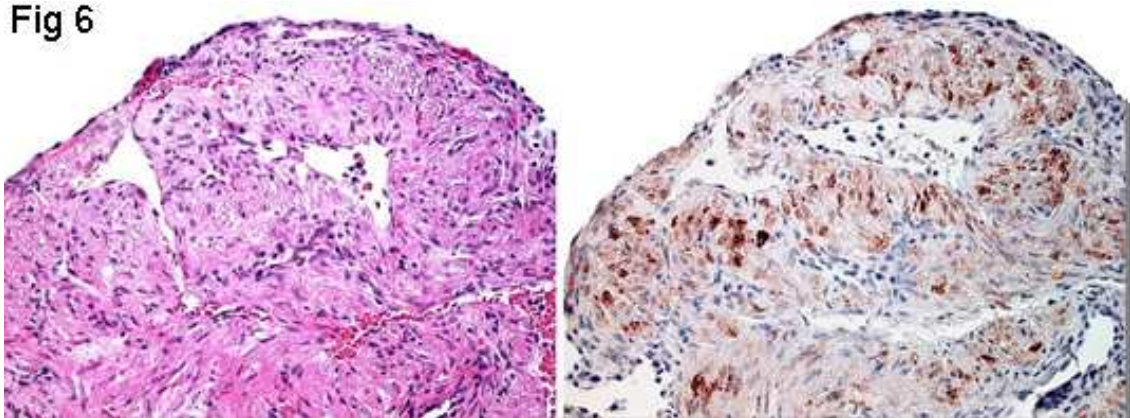


Figure 6. Left panel: close-up of LAM nodule (hematoxylin-eosin). Right panel: same nodule showing positive immunocytochemistry stain for HMB 45 (original magnification x200).

Fig 7

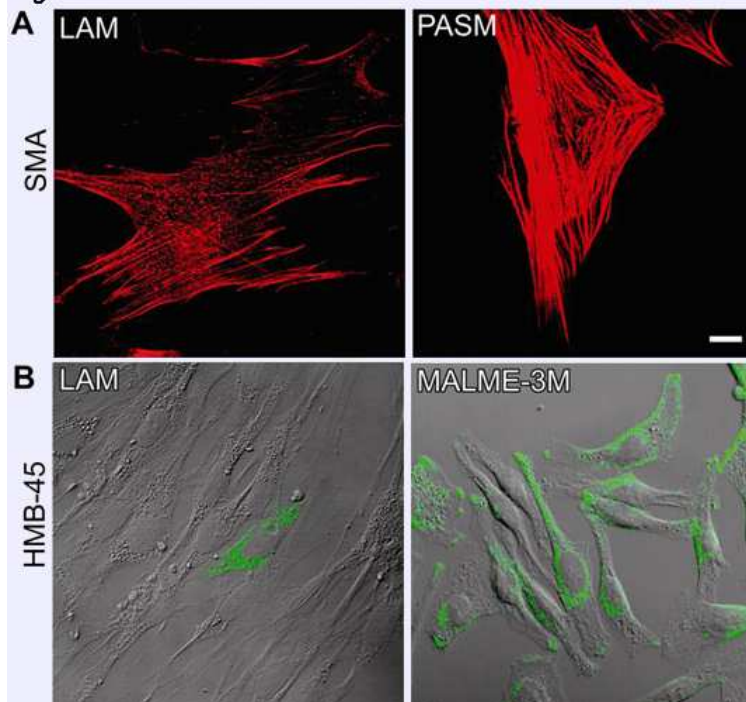
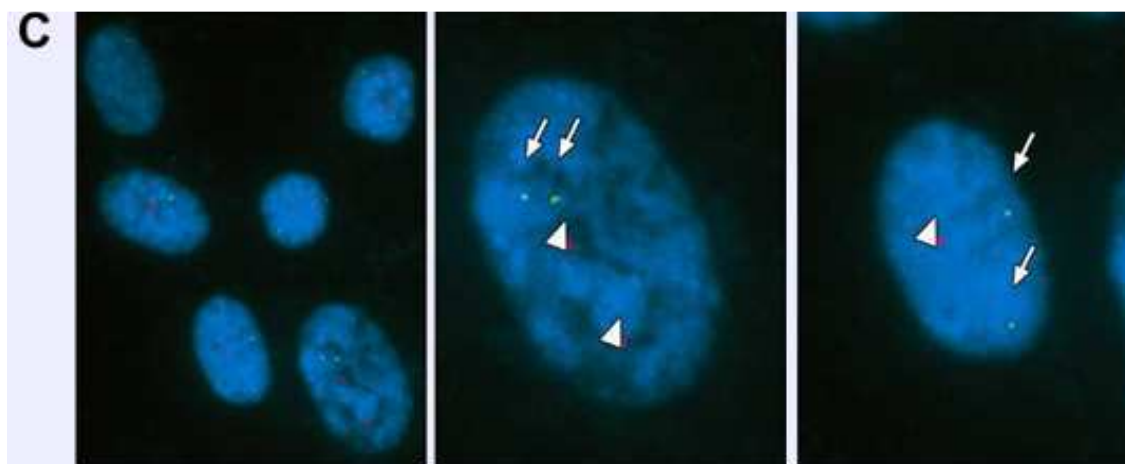


Figure 7. Characteristics of LAM cells (A-C). Reaction of LAM cells cultured from lung and pulmonary artery smooth muscle cells (PASM) with monoclonal antibody against SMA (A). Reaction of cultured LAM cells and melanoma cells (MALME-3M) with monoclonal antibody HMB-45 (B). Fluorescence in situ hybridization (FISH) for TSC1 (green) and TSC2 (red) in LAM cells showing normal presence of two of each alleles as well as abnormal presence of TSC2 alleles (left)



(C). FISH for TSC1 (green, arrow) and TSC2 (red, arrowhead) in LAM cell with one (right) or two TSC2 (left) alleles (C). Bar, 20 μ m.

Treatment

Because LAM is predominantly a disease of premenopausal women and may worsen during pregnancy, or following the administration of exogenous estrogens, hormonal manipulations have been employed. However, no controlled studies have been undertaken to determine their efficacy. A retrospective study of 275 patients found no difference in disease progression between patients treated with progesterone and patients who received no progesterone. There is also no evidence that suppression of ovarian function, either by oophorectomy or use of gonadotrophin-releasing hormone analogs, benefit patients with LAM.

Progress about the mechanisms regulating cell proliferation and migration, and angiogenesis and lymphangiogenesis, have provided a foundation for the development of new therapies.

The mammalian target of rapamycin (mTOR) appears to play a role in regulating the growth and multiplication of LAM cells (Figure 8). An inhibitor of mTOR, sirolimus (rapamycin), an antifungal macrolide antibiotic approved for immunosuppression after solid organ transplantation, has been studied as a possible treatment for LAM. In a rat model of TSC (Eker rat) with a functionally null germline mutation of *tsc2*, which spontaneously develops renal cell carcinomas, treatment with sirolimus resulted in a decrease in size of kidney tumors by both a reduction in the percentage of proliferating cells, and extensive tumor cell death. An open label study with sirolimus undertaken in twenty patients with angiomyolipomas showed a reduction in tumor size to 53.2 \pm 26.6 % of baseline at one year. An improvement in some lung function parameters was also observed. A clinical trial [MILES trial (Multicenter International Lymphangioliomyomatosis Efficacy of Sirolimus Trial)], to examine the effect of rapamycin on pulmonary function, is underway.

Patients with severe LAM or those who show an accelerated rate of decline in lung function may be referred to a lung transplantation center.

Evolution

LAM is a chronic disease, which may span decades. A retrospective analysis of 402 patients seen at NIH from 1995 to 2006 showed that 22 had died, eight of whom had undergone lung transplantation. The mortality in this large cohort was 5.5%. Of the surviving 380 patients, 38 (10%) had lung transplantation. A recent study reported a ten year survival greater than 90%.

Genes involved and proteins

TSC genes

Note

TSC1 and TSC2 are tumor suppressor genes. TSC1 (9q34) encodes the 130kDa protein hamartin, while TSC2 (16p13.3) encodes the 200kDa protein tuberin. Hamartin and tuberin may have individual functions, but they also interact to form a cytosolic complex. Loss of heterozygosity of TSC2 has been detected in LAM lesions from lung and kidney, and mutations in TSC2 occur more frequently than those in TSC1 in patients with sporadic LAM. Hamartin may play a role in the reorganization of the actin cytoskeleton, with a lack of hamartin leading to a loss of focal adhesions and detachment from substrate, resulting in cell rounding. Hamartin induces an increase in the levels of RhoGTP, an activator of ERM (ezrin-radixin-moesin) proteins, and binds to activated ERM proteins. ERM proteins bind both filamentous actin and integral membrane proteins, thereby bridging the plasma membrane and cortical actin filaments.

Tuberin has roles in pathways controlling cell growth and proliferation (Figure 8). It is a negative regulator of cell cycle progression, and loss of tuberin function shortens the G1 phase of the cell cycle. Tuberin binds p27KIP1, a cyclin-dependent kinase inhibitor, thereby preventing its degradation and leading to inhibition of the cell cycle. Upon mutation of tuberin, p27 becomes mislocalized in the cytoplasm, allowing the cell cycle to progress.

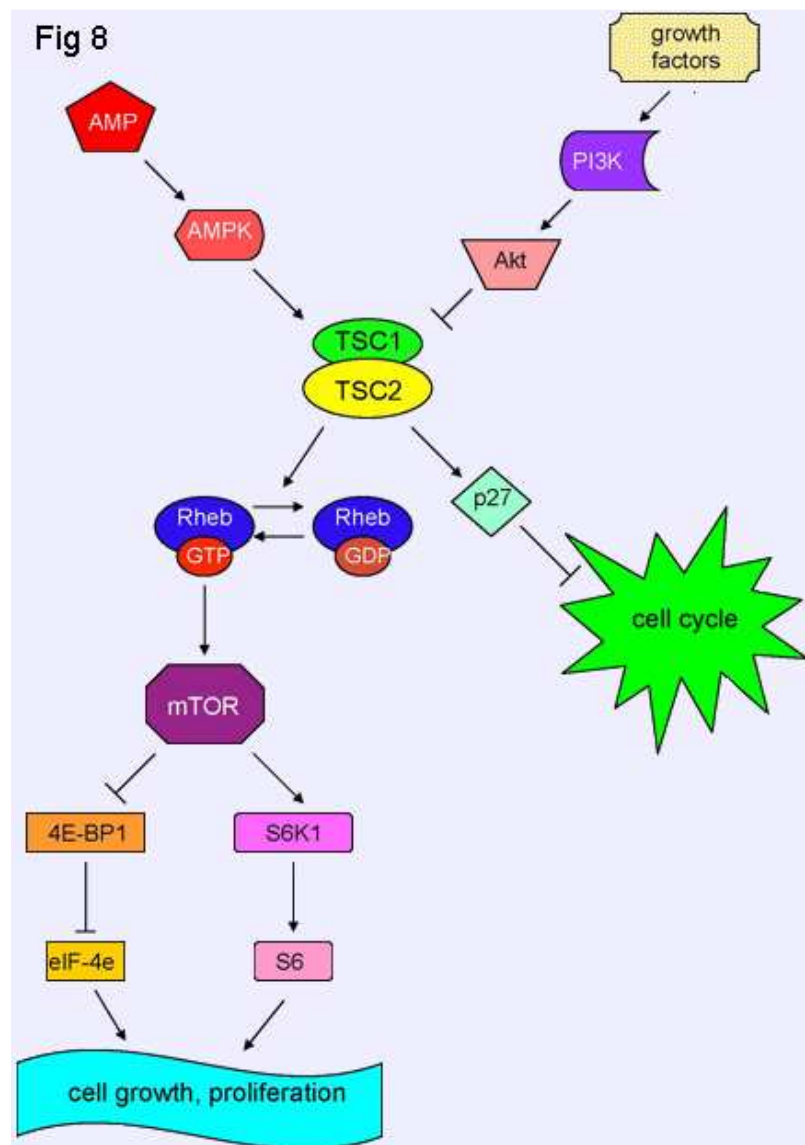


Figure 8. Schematic model of TSC1 and TSC2 pathways. The TSC1/TSC2 complex has roles in cell cycle progression and in cell growth and proliferation. Tuberlin binds p27KIP1, a cyclin-dependent kinase inhibitor, stabilizing it and resulting in inhibition of cell cycle progression. Tuberlin also has Rheb GAP activity, which converts Rheb-GTP to Rheb-GDP, resulting in inactive Rheb. Rheb controls mTOR, which is a kinase that controls translation through the phosphorylation of 4E-BP1 and S6K1. Akt, when activated by growth factors, phosphorylates tuberlin, leading to an inhibition of tuberlin and resulting in cell growth and proliferation. However, when a state of low cellular energy exists, AMPK phosphorylates tuberlin, activating it, and thereby inhibiting cell growth.

Tuberlin also integrates signals from growth factors and energy stores through its interaction with mTOR (mammalian target of rapamycin) (Figure 8). Tuberlin has Rheb GAP (Ras homolog enriched in brain GTPase-activating protein) activity, which converts Rheb-GTP to Rheb-GDP, thereby inactivating Rheb. Rheb controls mTOR, a serine/threonine kinase that phosphorylates at least two substrates: 4E-BP1, allowing cap-dependent translation, and S6K1, leading to translation of 5' TOP (terminal oligopyrimidine tract)-containing RNAs. Phosphorylation of tuberlin by Akt, which is known to be activated by growth factors, leads to inhibition of tuberlin and results in cell growth and proliferation. Phosphorylation of tuberlin by AMPK

(AMP-activated kinase) activates tuberlin and further promotes inhibition of cell growth in states of energy deprivation.

Tuberlin may also have roles in endocytosis through its interaction with rabaptin-5 and in transcriptional activation through interaction with members of the retinoid X receptor (RXR) family.

References

Berger U, Khaghani A, Pomerance A, Yacoub MH, Coombes RC. Pulmonary lymphangioliomyomatosis and steroid receptors. An immunocytochemical study. *Am J Clin Pathol.* 1990 May;93(5):609-14

Identification and characterization of the tuberous sclerosis gene on chromosome 16. *Cell.* 1993 Dec 31;75(7):1305-15

- Soucek T, Pusch O, Wienecke R, DeClue JE, Hengstschläger M. Role of the tuberous sclerosis gene-2 product in cell cycle control. Loss of the tuberous sclerosis gene-2 induces quiescent cells to enter S phase. *J Biol Chem.* 1997 Nov 14;272(46):29301-8
- Xiao GH, Shoarinejad F, Jin F, Golemis EA, Yeung RS. The tuberous sclerosis 2 gene product, tuberin, functions as a Rab5 GTPase activating protein (GAP) in modulating endocytosis. *J Biol Chem.* 1997 Mar 7;272(10):6097-100
- Henry KW, Yuan X, Koszewski NJ, Onda H, Kwiatkowski DJ, Noonan DJ. Tuberous sclerosis gene 2 product modulates transcription mediated by steroid hormone receptor family members. *J Biol Chem.* 1998 Aug 7;273(32):20535-9
- Plank TL, Yeung RS, Henske EP. Hamartin, the product of the tuberous sclerosis 1 (TSC1) gene, interacts with tuberin and appears to be localized to cytoplasmic vesicles. *Cancer Res.* 1998 Nov 1;58(21):4766-70
- Smolarek TA, Wessner LL, McCormack FX, Mylet JC, Menon AG, Henske EP. Evidence that lymphangiomyomatosis is caused by TSC2 mutations: chromosome 16p13 loss of heterozygosity in angiomyolipomas and lymph nodes from women with lymphangiomyomatosis. *Am J Hum Genet.* 1998 Apr;62(4):810-5
- Soucek T, Yeung RS, Hengstschläger M. Inactivation of the cyclin-dependent kinase inhibitor p27 upon loss of the tuberous sclerosis complex gene-2. *Proc Natl Acad Sci U S A.* 1998 Dec 22;95(26):15653-8
- Usuki J, Horiba K, Chu SC, Moss J, Ferrans VJ. Immunohistochemical analysis of proteins of the Bcl-2 family in pulmonary lymphangiomyomatosis: association of Bcl-2 expression with hormone receptor status. *Arch Pathol Lab Med.* 1998 Oct;122(10):895-902
- van Slegtenhorst M, Nellist M, Nagelkerken B, Cheadle J, Snell R, van den Ouweland A, Reuser A, Sampson J, Halley D, van der Sluijs P. Interaction between hamartin and tuberin, the TSC1 and TSC2 gene products. *Hum Mol Genet.* 1998 Jun;7(6):1053-7
- Chu SC, Horiba K, Usuki J, Avila NA, Chen CC, Travis WD, Ferrans VJ, Moss J. Comprehensive evaluation of 35 patients with lymphangiomyomatosis. *Chest.* 1999 Apr;115(4):1041-52
- Matsumoto Y, Horiba K, Usuki J, Chu SC, Ferrans VJ, Moss J. Markers of cell proliferation and expression of melanosomal antigen in lymphangiomyomatosis. *Am J Respir Cell Mol Biol.* 1999 Sep;21(3):327-36
- Nellist M, van Slegtenhorst MA, Goedbloed M, van den Ouweland AM, Halley DJ, van der Sluijs P. Characterization of the cytosolic tuberin-hamartin complex. Tuberin is a cytosolic chaperone for hamartin. *J Biol Chem.* 1999 Dec 10;274(50):35647-52
- Urban T, Lazor R, Lacronique J, Murriss M, Labrune S, Valeyre D, Cordier JF. Pulmonary lymphangiomyomatosis. A study of 69 patients. Groupe d'Etudes et de Recherche sur les Maladies "Orphelines" Pulmonaires (GERM"O" P). *Medicine (Baltimore).* 1999 Sep;78(5):321-37
- Carsillo T, Astrinidis A, Henske EP. Mutations in the tuberous sclerosis complex gene TSC2 are a cause of sporadic pulmonary lymphangiomyomatosis. *Proc Natl Acad Sci U S A.* 2000 May 23;97(11):6085-90
- Costello LC, Hartman TE, Ryu JH. High frequency of pulmonary lymphangiomyomatosis in women with tuberous sclerosis complex. *Mayo Clin Proc.* 2000 Jun;75(6):591-4
- Lamb RF, Roy C, Diefenbach TJ, Vinters HV, Johnson MW, Jay DG, Hall A. The TSC1 tumour suppressor hamartin regulates cell adhesion through ERM proteins and the GTPase Rho. *Nat Cell Biol.* 2000 May;2(5):281-7
- Franz DN, Brody A, Meyer C, Leonard J, Chuck G, Dabora S, Sethuraman G, Colby TV, Kwiatkowski DJ, McCormack FX. Mutational and radiographic analysis of pulmonary disease consistent with lymphangiomyomatosis and micronodular pneumocyte hyperplasia in women with tuberous sclerosis. *Am J Respir Crit Care Med.* 2001 Aug 15;164(4):661-8
- Maruyama H, Ohbayashi C, Hino O, Tsutsumi M, Konishi Y. Pathogenesis of multifocal micronodular pneumocyte hyperplasia and lymphangiomyomatosis in tuberous sclerosis and association with tuberous sclerosis genes TSC1 and TSC2. *Pathol Int.* 2001 Aug;51(8):585-94
- Moss J, Avila NA, Barnes PM, Litzemberger RA, Bechtle J, Brooks PG, Hedin CJ, Hunsberger S, Kristof AS. Prevalence and clinical characteristics of lymphangiomyomatosis (LAM) in patients with tuberous sclerosis complex. *Am J Respir Crit Care Med.* 2001 Aug 15;164(4):669-71
- Soucek T, Rosner M, Milozola A, Kubista M, Cheadle JP, Sampson JR, Hengstschläger M. Tuberous sclerosis causing mutants of the TSC2 gene product affect proliferation and p27 expression. *Oncogene.* 2001 Aug 9;20(35):4904-9
- Strizheva GD, Carsillo T, Kruger WD, Sullivan EJ, Ryu JH, Henske EP. The spectrum of mutations in TSC1 and TSC2 in women with tuberous sclerosis and lymphangiomyomatosis. *Am J Respir Crit Care Med.* 2001 Jan;163(1):253-8
- Gao X, Zhang Y, Arrazola P, Hino O, Kobayashi T, Yeung RS, Ru B, Pan D. Tsc tumour suppressor proteins antagonize amino-acid-TOR signalling. *Nat Cell Biol.* 2002 Sep;4(9):699-704
- Goncharova EA, Goncharov DA, Eszterhas A, Hunter DS, Glassberg MK, Yeung RS, Walker CL, Noonan D, Kwiatkowski DJ, Chou MM, Panettieri RA Jr, Krymskaya VP. Tuberin regulates p70 S6 kinase activation and ribosomal protein S6 phosphorylation. A role for the TSC2 tumor suppressor gene in pulmonary lymphangiomyomatosis (LAM). *J Biol Chem.* 2002 Aug 23;277(34):30958-67
- Inoki K, Li Y, Zhu T, Wu J, Guan KL. TSC2 is phosphorylated and inhibited by Akt and suppresses mTOR signalling. *Nat Cell Biol.* 2002 Sep;4(9):648-57
- Manning BD, Tee AR, Logsdon MN, Blenis J, Cantley LC. Identification of the tuberous sclerosis complex-2 tumor suppressor gene product tuberin as a target of the phosphoinositide 3-kinase/akt pathway. *Mol Cell.* 2002 Jul;10(1):151-62
- Tee AR, Fingar DC, Manning BD, Kwiatkowski DJ, Cantley LC, Blenis J. Tuberous sclerosis complex-1 and -2 gene products function together to inhibit mammalian target of rapamycin (mTOR)-mediated downstream signaling. *Proc Natl Acad Sci U S A.* 2002 Oct 15;99(21):13571-6
- Bittmann I, Rolf B, Amann G, Löhns U. Recurrence of lymphangiomyomatosis after single lung transplantation: new insights into pathogenesis. *Hum Pathol.* 2003 Jan;34(1):95-8
- Castro AF, Rebhun JF, Clark GJ, Quilliam LA. Rheb binds tuberous sclerosis complex 2 (TSC2) and promotes S6 kinase activation in a rapamycin- and farnesylation-dependent manner. *J Biol Chem.* 2003 Aug 29;278(35):32493-6
- Inoki K, Zhu T, Guan KL. TSC2 mediates cellular energy response to control cell growth and survival. *Cell.* 2003 Nov 26;115(5):577-90
- Karbowiczek M, Astrinidis A, Balsara BR, Testa JR, Lium JH, Colby TV, McCormack FX, Henske EP. Recurrent lymphangiomyomatosis after transplantation: genetic analyses

reveal a metastatic mechanism. *Am J Respir Crit Care Med*. 2003 Apr 1;167(7):976-82

Karbowiczek M, Yu J, Henske EP. Renal angiomyolipomas from patients with sporadic lymphangiomyomatosis contain both neoplastic and non-neoplastic vascular structures. *Am J Pathol*. 2003 Feb;162(2):491-500

Krymskaya VP. Tumour suppressors hamartin and tuberlin: intracellular signalling. *Cell Signal*. 2003 Aug;15(8):729-39

Tee AR, Manning BD, Roux PP, Cantley LC, Blenis J. Tuberous sclerosis complex gene products, Tuberlin and Hamartin, control mTOR signaling by acting as a GTPase-activating protein complex toward Rheb. *Curr Biol*. 2003 Aug 5;13(15):1259-68

Zhang Y, Gao X, Saucedo LJ, Ru B, Edgar BA, Pan D. Rheb is a direct target of the tuberous sclerosis tumour suppressor proteins. *Nat Cell Biol*. 2003 Jun;5(6):578-81

Crooks DM, Pacheco-Rodriguez G, DeCastro RM, McCoy JP Jr, Wang JA, Kumaki F, Darling T, Moss J. Molecular and genetic analysis of disseminated neoplastic cells in lymphangioliomyomatosis. *Proc Natl Acad Sci U S A*. 2004 Dec 14;101(50):17462-7

Li Y, Corradetti MN, Inoki K, Guan KL. TSC2: filling the GAP in the mTOR signaling pathway. *Trends Biochem Sci*. 2004 Jan;29(1):32-8

Rosner M, Freilinger A, Hengstschlager M. Proteins interacting with the tuberous sclerosis gene products. *Amino Acids*. 2004 Oct;27(2):119-28

Rosner M, Hengstschlager M. Tuberlin binds p27 and negatively regulates its interaction with the SCF component Skp2. *J Biol Chem*. 2004 Nov 19;279(47):48707-15

Inoki K, Corradetti MN, Guan KL. Dysregulation of the TSC-mTOR pathway in human disease. *Nat Genet*. 2005 Jan;37(1):19-24

Steagall WK, Taveira-DaSilva AM, Moss J. Clinical and molecular insights into lymphangioliomyomatosis. *Sarcoidosis Vasc Diffuse Lung Dis*. 2005 Dec;22 Suppl 1:S49-66

Johnson SR. Lymphangioliomyomatosis. *Eur Respir J*. 2006 May;27(5):1056-65

Ryu JH, Moss J, Beck GJ, Lee JC, Brown KK, Chapman

JT, Finlay GA, Olson EJ, Ruoss SJ, Maurer JR, Raffin TA, Peavy HH, McCarthy K, Taveira-Dasilva A, McCormack FX, Avila NA, Decastro RM, Jacobs SS, Stylianou M, Fanburg BL. The NHLBI lymphangioliomyomatosis registry: characteristics of 230 patients at enrollment. *Am J Respir Crit Care Med*. 2006 Jan 1;173(1):105-11

Taveira-DaSilva AM, Steagall WK, Moss J. Lymphangioliomyomatosis. *Cancer Control*. 2006 Oct;13(4):276-85

Pacheco-Rodriguez G, Steagall WK, Crooks DM, Stevens LA, Hashimoto H, Li S, Wang JA, Darling TN, Moss J. TSC2 loss in lymphangioliomyomatosis cells correlated with expression of CD44v6, a molecular determinant of metastasis. *Cancer Res*. 2007 Nov 1;67(21):10573-81

Steagall WK, Glasgow CG, Hathaway OM, Avila NA, Taveira-Dasilva AM, Rabel A, Stylianou MP, Lin JP, Chen X, Moss J. Genetic and morphologic determinants of pneumothorax in lymphangioliomyomatosis. *Am J Physiol Lung Cell Mol Physiol*. 2007 Sep;293(3):L800-8

Glasgow CG, Taveira-Dasilva AM, Darling TN, Moss J. Lymphatic involvement in lymphangioliomyomatosis. *Ann N Y Acad Sci*. 2008;1131:206-14

Harari S, Cassandro R, Chiodini I, Taveira-DaSilva AM, Moss J. Effect of a gonadotrophin-releasing hormone analogue on lung function in lymphangioliomyomatosis. *Chest*. 2008 Feb;133(2):448-54

McCormack FX. Lymphangioliomyomatosis: a clinical update. *Chest*. 2008 Feb;133(2):507-16

Rosner M, Hanneder M, Siegel N, Valli A, Hengstschlager M. The tuberous sclerosis gene products hamartin and tuberlin are multifunctional proteins with a wide spectrum of interacting partners. *Mutat Res*. 2008 Mar-Apr;658(3):234-46

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