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Effects of Elevated Atmospheric CO₂ on Photosynthesis, Growth and Biomass in *Shorea platycarpa* F. Heim (Meranti Paya)

(Kesan Peningkatan CO₂ dalam Atmosfera Terhadap Fotosintesis, Pertumbuhan dan Biojisim *Shorea platycarpa* F. Heim (Meranti Paya))

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ABSTRACT

Elevated atmospheric CO₂ is widely reported to stimulate the plant growth and affect plant physiological processes. However, studies on the respond of tropical plant species to elevated CO₂ are quite limited and remain largely unknown. The objective of this study was to investigate the effects of elevated atmospheric CO₂ treatments on the photosynthetic characteristics, growth and biomass in *Shorea platycarpa*. Saplings of *S. platycarpa* were grown for seven months in the open roof gas chamber supplied with elevated CO₂ (800±50 µmol mol⁻¹) and in the shade house with ambient CO₂ (400±50 µmol mol⁻¹). Measurements of *S. platycarpa* growth and photosynthetic characteristics were made at frequent intervals. Biomass characteristics were determined using destructive methods after seven months of treatment and nondestructive method was used for leaf area index (LAI) determination. Photosynthetic rate (A) of *S. platycarpa* was not significantly affected by elevated CO₂. Increased water use efficiency (WUE) of *S. platycarpa* grown in elevated CO₂ was due to the reduced stomatal conductance (g_s) and transpiration rate (E). The CO₂ elevation had no significant effect on the *S. platycarpa* relative growth rates (RGR) and biomass but significantly reduced the leaf area. A weak correlation was found between photosynthetic rate (A) and relative growth rate (RGR). The results clearly showed that photosynthesis, growth rate and biomass of *S. platycarpa* were not significantly enhanced by elevated CO₂. The findings indicated that elevated CO₂ did not affect a relatively slow growing and a late successional peat swamp tree species.

Keywords: Biomass; elevated CO₂; growth rate; photosynthesis; *Shorea platycarpa*

ABSTRAK

Peningkatan CO₂ dalam atmosfera telah dilaporkan secara meluas kerana dapat meningkatkan pertumbuhan pokok dan memberi kesan kepada proses fisiologi pokok. Namun begitu, kajian ke atas tindak balas spesies pokok tropika terhadap peningkatan CO₂ agak terhad dan sebahagian besarnya tidak diketahui. Objektif kajian ini ialah untuk mengkaji kesan rawatan peningkatan CO₂ dalam atmosfera ke atas ciri fotosintesis, pertumbuhan dan biojisim *Shorea platycarpa*. Anak pokok *S. platycarpa* telah ditanam selama tujuh bulan di dalam bilik gas dengan bumbung terbuka dan dibekalkan dengan peningkatan CO₂ (800±50 µmol mol⁻¹) dan di dalam rumah teduh dengan ambien CO₂ (400±50 µmol mol⁻¹). Pengukuran secara selang berulang ke atas pertumbuhan *S. platycarpa* dan ciri fotosintesis telah dibuat. Selepas tujuh bulan rawatan, ciri biojisim telah ditentukan dengan menggunakan kaedah musnah dan tanpa musnah untuk indeks luas daun (LAI). Kadar fotosintesis (A) *S. platycarpa* tidak dipengaruhi secara signifikan oleh peningkatan CO₂. Peningkatan kecekapan penggunaan air (WUE) *S. platycarpa* yang hidup di CO₂ berganda disebabkan oleh pengurangan stomata konduktans (g_s) dan kadar transpirasi (E). Peningkatan CO₂ tidak memberi kesan signifikan ke atas kadar pertumbuhan relatif (RGR) dan biojisim tetapi mengurangkan luas daun secara signifikan. Korelasi lemah wujud antara kadar fotosintesis (A) dengan kadar pertumbuhan relatif (RGR). Hasil jelas menunjukkan bahawa fotosintesis, kadar pertumbuhan dan biojisim *S. platycarpa* tidak meningkat secara signifikan dengan peningkatan CO₂. Penemuan tersebut memberi indikasi bahawa peningkatan CO₂ tidak memberi kesan kepada spesies pokok paya gambut dengan ciri pertumbuhan perlahan secara relatif dan lewat sesaran.

Kata kunci: Biojisim; fotosintesis; kadar pertumbuhan; peningkatan CO₂; *Shorea platycarpa*

INTRODUCTION

For the 1000 years prior to the Industrial Revolution CO₂ was stable at about 270 µmol mol⁻¹; today CO₂ is approximately 38% higher at 372 µmol mol⁻¹ and by the middle of this century it is predicted to reach 550 µmol mol⁻¹ and to surpass 700 µmol mol⁻¹ by the end of the century (Long et al. 2004). Extensive botanical

investigations have been done to comprehend the potential effect of elevated atmospheric CO₂ concentration, e.g. studies on *Populus tremuloides*, *Betula papyrifera*, *Pinus taeda* and *Quercus ilex* (Karnosky 2003). However, not all plant species respond the same way to elevated CO₂. There is a considerable variation in the direction and magnitude of growth responses to elevated CO₂, partly depending

on the duration of the exposure, plant development, species (e.g. species that differ in inherent growth rate or type of photosynthetic pathway) and the availability of primary resources (Poorter & Pérez-Soba 2001). Even though growth of *Ochroma lagopus*, a pioneer species and *Pentaclethra macroloba*, a climax species increased significantly in elevated CO₂, *O. lagopus* responded more to elevated CO₂ compared to *P. macroloba* (Oberbauer et al. 1985). In addition, Liang et al. (2001) reported that elevated CO₂ did suppress photosynthesis of *Pongamia pinnata*, a shade tolerant species in both low and high light condition. Nonetheless, more than half of the recognized angiosperm families have not had a single species evaluated for CO₂ response in a field environment and there are large gaps in knowledge for a number of keystone species and genera, including those in the Dipterocarpaceae family (Leakey et al. 2012).

Shorea platycarpa also known as Meranti paya is a timber tree, belonging to Dipterocarpaceae family and is considered as endangered species due to forest conversion (Ashton et al. 1988). This species is generally found in deep peat swamp forest and the wood is commonly used for joinery and furniture (Choo et al. 1998). Compared to temperate regions and crop plants, the studies on elevated CO₂ on peat swamp trees is relatively limited. Peatland represent a major store of soil carbon and sink for carbon dioxide. Plants play an important roles in sequestering carbon through photosynthesis process and the amount and composition of carbon forms that plants sequester can be related to plant trait (De Deyn et al. 2008). Hence, the investigation on the responses of such important species in the peat swamp ecosystem to elevated CO₂ is timely. The present study describes the growth and physiological responses of a dominant, late successional (Hamzah et al. 2009), tropical tree species of a peat swamp forest i.e. *Shorea platycarpa* grown under elevated CO₂ with main focus on photosynthetic leaf gas exchange, relative growth rate and biomass allocation.

MATERIALS AND METHODS

PLANT MATERIAL, GROWTH CONDITION AND EXPERIMENTAL DESIGN

The experiments were conducted from March to October 2015 in an open roof chamber with elevated CO₂ that of 800±50 µmol mol⁻¹ and in the shade house with ambient CO₂ that of 400±50 µmol mol⁻¹ at the Universiti Kebangsaan Malaysia (UKM). The elevated concentration of CO₂ (800 ± 50 µmol mol⁻¹) were selected to match with the predicted concentration at the end of the century from a global climate models under WGIII scenario of IPCC (2014). Eighteen months old saplings of *S. platycarpa* were collected from Forest Research Institute Malaysia (FRIM). Seeds of this species are not readily available due to the non-mast years. For several species of dipterocarps, mast years occur unpredictably at intervals of 2-10 years (Ashton et al. 1988). Saplings growth responses can be

measured easily compared to adult trees. Hence, they are more amenable to experimentation by short term research projects. Seedling growth and survival may also be more responsive to environmental than that of large trees (Zagt & Werger 1998). Microclimatic measurements were taken within and outside the chamber (shade house) at different time intervals. Humidity and temperature were monitored and controlled using ventilation system. Temperature and relative humidity were 0.1-0.2°C and 2-4% respectively, relatively higher in the chamber compared to shade house. The light intensity in the chamber was 94-95% of the shade house/ambient level. Treatment of CO₂ (800±50 µmol mol⁻¹) was done by daily automated continuous injection of pure CO₂ for 2 h starting from 9 until 11 am until the desired concentration (800±50 µmol mol⁻¹) was reached. CO₂ was supplied through CO₂ cylinder which was connected to air delivery system of the open roof chamber and air blower, and the concentration was regulated by dilution with air stream generated by the blower. CO₂ concentration in the chamber was monitored and administered using CO₂ analyzer. After two hours of CO₂ injection, the CO₂ levels inside the chamber and the ambient CO₂ was virtually the same. Growing medium used was ground parts topsoil podzolic types that were mixed with organic fertilizer with a ratio of 4:1. The saplings were planted in polybags (25 cm diameter × 30 cm depth). NPK was applied (15:15:15) with a low dose of 5 g/polybag/month. The pots were watered periodically to sustain soil moisture.

PLANT GROWTH

Plant growth ($n=18$) was assessed by regular, nondestructive measurements. Plants height, stem diameter and number of leaf were measured weekly for seven months. Plant height (cm) was measured vertically from the soil surface to the tip of youngest leaf (at the top of the plant) using a measuring tape. In addition, the main stem diameter (mm) was measured at a consistent point (5 cm above the base of the stem) using callipers. Relative height growth rate (RHGR), stem basal diameter growth rate (SDGR) and number of leaf growth rate (NLGR) were calculated every month according to Jach and Ceulemans (1999).

$$RGR = (\ln l_2 - \ln l_1) / (t_2 - t_1) \text{ where } l_1 = \text{the height / diameter at time } t_1 \text{ and } l_2 = \text{the height / diameter at } t_2.$$

BIOMASS DETERMINATION

At the end of the experiment, the plants were cut at the base and divided into above ground and below ground parts. The roots were carefully washed in water to remove all soil particles and then both components were oven dried at 65°C for seven days. For growth analysis, the dry weight (DW) of the average above ground biomass (AGB), average below ground biomass (BGB) and below ground: above ground ratio (BG: AG) were calculated. The (BG: AG) was determined as below ground dry matter divided by the sum of above ground dry matter.

LEAF AREA INDEX DETERMINATION

A simple nondestructive method was used to estimate the leaf area index (LAI) using digital image taken *in situ* and public domain software ImageJ. Samples were taken after three months of treatment (first phase) and after seven months of treatment (second phase). To record the image of a leaf, a 30 cm ruler was placed on board with white paper where the leaf was positioned. Then, a photograph of each young, and fully expanded leaf ($n=160$ leaves originating from 8 plants) was taken using digital camera. The digital images were then processed in ImageJ for leaf area index (LAI) estimation.

PHOTOSYNTHETIC LEAF GAS EXCHANGE MEASUREMENTS

The measurements of the photosynthetic rate (A), stomatal conductance (g_s), transpiration rate (E) and water use efficiency (WUE) was made at frequent intervals throughout seven months treatment with the LI-6400 Portable Photosynthesis System (Li-cor, Lincoln, NE, USA). The levels of A, g_s , E, and WUE were measured at ambient CO_2 concentration ($400 \mu mol mol^{-1}$), air temperature ($28-29^\circ C$), relative humidity (about 65%) and photosynthetic photon flux density (PPFD, $1000 \mu mol mol^{-1}$) in the leaf chamber. For each treatment; ambient ($400 \pm 50 \mu mol mol^{-1}$) and elevated CO_2 ($800 \pm 50 \mu mol mol^{-1}$) concentration, measurements were made on randomly selected fully expanded leaf from each plant ($n=18$) from 9 am to 12 pm.

STATISTICAL ANALYSIS

An analysis of variance was conducted to evaluate the effects of elevated CO_2 on several growth characteristics using one way ANOVA of MINITAB version 16.0 software package. Growth characteristics, biomass characteristics, photosynthetic characteristics and leaf area index were included in the analysis of variance (ANOVA). The data were tested for normality using Anderson-Darling's test and where necessary raw data were transformed logarithmically to meet assumptions of normality. Statistical significance of the mean data was assessed by Tukey's multiple range test ($p \leq 0.05$). Relationship between gas exchange and growth parameters were analyzed by correlations and regression analysis using MINITAB version 16.0.

RESULTS

Elevated CO_2 did not enhanced the RHGR (Figure 1(a)), SDGR (Figure 1(b)) and NLGR (Figure 1(c)) of the plants. The highest peak of RHGR for ambient CO_2 and elevated CO_2 was 0.0043 and $0.0036 cm cm^{-1} day^{-1}$, respectively. However, only at month five there was a significant difference ($p < 0.05$) in RHGR between these two treatments. This was probably being influenced by the variability of individual growth performance. The highest peak of SDGR for ambient CO_2 and elevated CO_2 was 0.0048 and $0.0054 cm cm^{-1} day^{-1}$, respectively. The SDGR in plants between ambient CO_2 and elevated CO_2 did not show significant

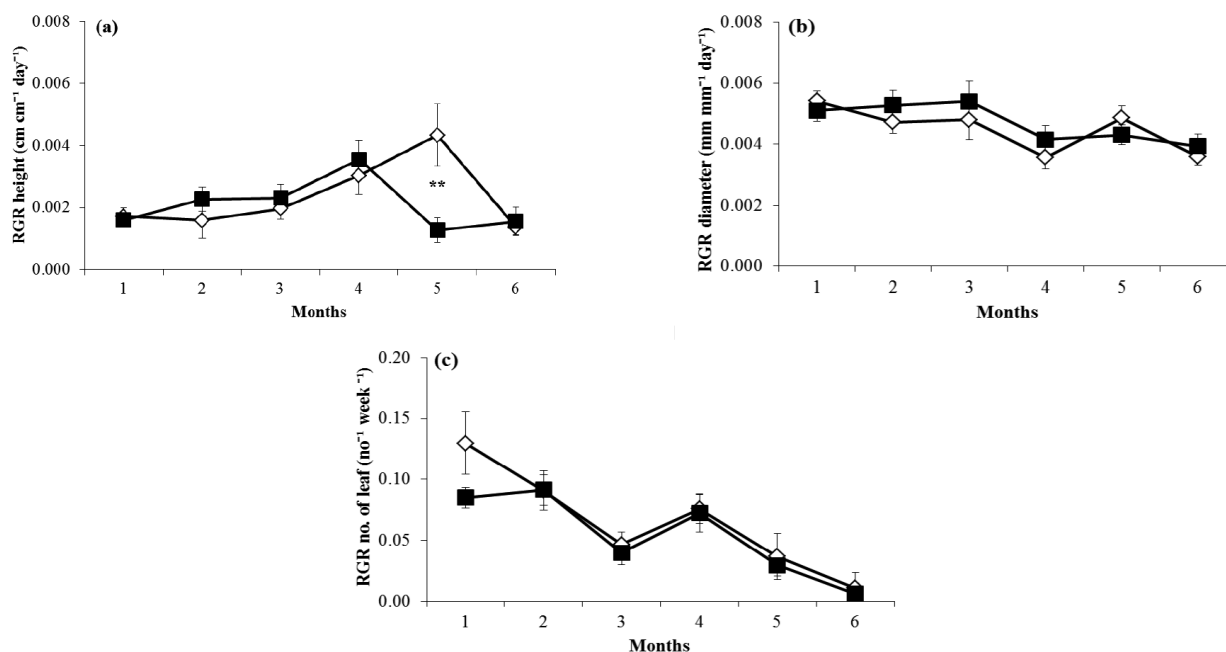


FIGURE 1. Time course of monthly relative growth rate (RGR) for height (a), diameter (b) and number of leaf (c) of *Shorea platycarpa* during seven months of treatment (Mac - October 2015) under ambient CO_2 and elevated CO_2 . An $\diamond = 400 \pm 50 \mu mol mol^{-1}$, $\blacksquare = 800 \pm 50 \mu mol mol^{-1}$. Values represent means \pm SE; $n=18$. The statistical significance is based on one-way ANOVA ($p \leq 0.05$). An asterisk (**) in (a) indicates a significant difference ($p < 0.01$) between the ambient and elevated CO_2 treatment

($p < 0.05$) variations in seven months of treatment. The NLGR from month one to month seven for both ambient CO₂ and elevated CO₂ treatment was continuously decreasing from 0.13 to 0.011 no.⁻¹ week⁻¹ and 0.085 to 0.006 no.⁻¹ week⁻¹, respectively. Overall, there were no significant differences ($p < 0.05$) in the mean relative growth rate (RGR) of height, diameter and number of leaf of *S. platycarpa* after seven months treatment in elevated CO₂ (Table 1).

There was no significant difference in the mean dry weight biomass of *S. platycarpa* grown in the two CO₂ conditions at ($p < 0.05$) (Table 1). Though, a decreased biomass yield was observed for plant grown in elevated CO₂. Total dry weight biomass was 56.26 g higher in ambient CO₂ compared to elevated CO₂, which 48.12 g of it was from the above ground biomass. The below ground biomass was 8.14 g higher in ambient CO₂ compared to elevated CO₂. There was no significant ($p < 0.05$) CO₂ effect on BG: AG, even though the ratios were slightly higher in elevated CO₂. On the other hand, leaf area index (LAI) (Table 1) shows significant difference ($p < 0.05$) after seven months of treatment compared to after three months of treatment, indicating that duration of treatment under elevated CO₂ did give significant effect on the LAI of *S. platycarpa*. The LAI of *S. platycarpa* increased by 159.7% and 3.6% by the end of seven months treatment in the ambient and elevated CO₂, respectively.

The photosynthetic rate (A) was averagely increased by 82% and 85% by the end of seven months treatment in the ambient and elevated CO₂, respectively. Photosynthetic rate (A) of plant grown in elevated CO₂ did increase in the second month of treatment, but then it was getting lower than (A) for plants grown in ambient CO₂ (Figure 2(a)). Decreased stomatal conductance (g_s) (Figure 2(b)) and transpiration rate (E) (Figure 2(c)) in *S. platycarpa* under elevated CO₂ were recorded. After seven month of treatments, the value of stomatal conductance (g_s) and transpiration rate (E) of plants kept at elevated CO₂,

compared with that of their counterparts grown in ambient CO₂, was 0.4% and 51.9% lower, respectively. As expected, transpiration rates (E) were significantly lower ($p < 0.05$) at elevated CO₂ than at ambient CO₂ from early treatment until end of treatment. Transpiration was reduced due to a lower stomatal conductance (g_s). The water loss transpiration then will correspond to water use efficiency (WUE). Therefore, because of lower transpiration rate (E), plants grown under elevated CO₂ showed an average increase in water use efficiency (WUE) by 88% than their respective controls.

The correlation analysis between photosynthetic rates (A) and relative growth rate (RGR) after seven-month treatment showed a non-significant weak correlation ($r = 0.4$, $p < 0.05$) for both RHGR (Figure 3(a)) and (SDGR) (Figure 3(b)), indicating that the photosynthetic rate (A) is not a primary influence for RHGR and SDGR of *S. platycarpa* under elevated CO₂.

DISCUSSION

EFFECTS OF ELEVATED CO₂ ON GROWTH

Growth increments of *S. platycarpa* were not profoundly influenced by elevated CO₂ despite (A) were increased in both condition. In many cases (e.g. Rasineni et al. 2011), high CO₂ usually increases the photosynthetic rate and thereby the growth rate. However, in the present study and some other experiments (Novriyanti et al. 2012), there were no positive effects of elevated CO₂ on growth of *S. platycarpa*. Nevertheless, in particular, the mean SDGR was higher under elevated CO₂ condition compared to ambient CO₂, whereas mean RHGR in elevated CO₂ was equivalent to the ambient CO₂ condition. Stem diameter was more responsive to CO₂ treatment compared to plant height. Many studies reported that elevated CO₂ resulted in the formation of wider tree rings in conifers (*Pinus radiata*

TABLE 1. Effects of atmospheric CO₂ (ambient; 400±50 µmol mol⁻¹ and elevated 800±50 µmol mol⁻¹) on relative growth rate (RGR), biomass and leaf area index (LAI) in *Shorea platycarpa* after seven months of treatment (Mac - October 2015)

Characters	Ambient CO ₂	Elevated CO ₂	P level
<i>Plant growth (RGR)</i>			
RGR height (cm cm ⁻¹ day ⁻¹)	0.002 ± 0.00	0.002 ± 0.00	0.46 ns
RGR diameter (mm mm ⁻¹ day ⁻¹)	0.004 ± 0.00	0.005 ± 0.00	0.46 ns
RGR no. of leaf (no ⁻¹ week ⁻¹)	0.07 ± 0.00	0.05 ± 0.00	0.28 ns
<i>Biomass (g DW plant⁻¹)</i>			
Below ground	41.51 ± 4.61	33.37 ± 6.23	0.32 ns
Above ground	137.71 ± 15.44	89.59 ± 21.83	0.11 ns
Total biomass	179.22 ± 18.51	122.96 ± 27.45	0.13 ns
BG:AG ratio	0.31 ± 0.02	0.40 ± 0.04	0.12 ns
<i>Leaf area index (cm² plant⁻¹)</i>			
LAI (3 months of treatment)	2.24 ± 0.04	2.18 ± 0.03	0.10 ns
LAI (7 months of treatment)	4.50 ± 0.06	3.84 ± 0.08	0.00 ***

Values represent means ± SE; n=18. The calculation of statistical significance is based on one-way ANOVA ($p = 0.05$). The symbol used *** $p < 0.001$; ** $p < 0.01$; * $p < 0.05$; not significant (ns) $p > 0.05$.

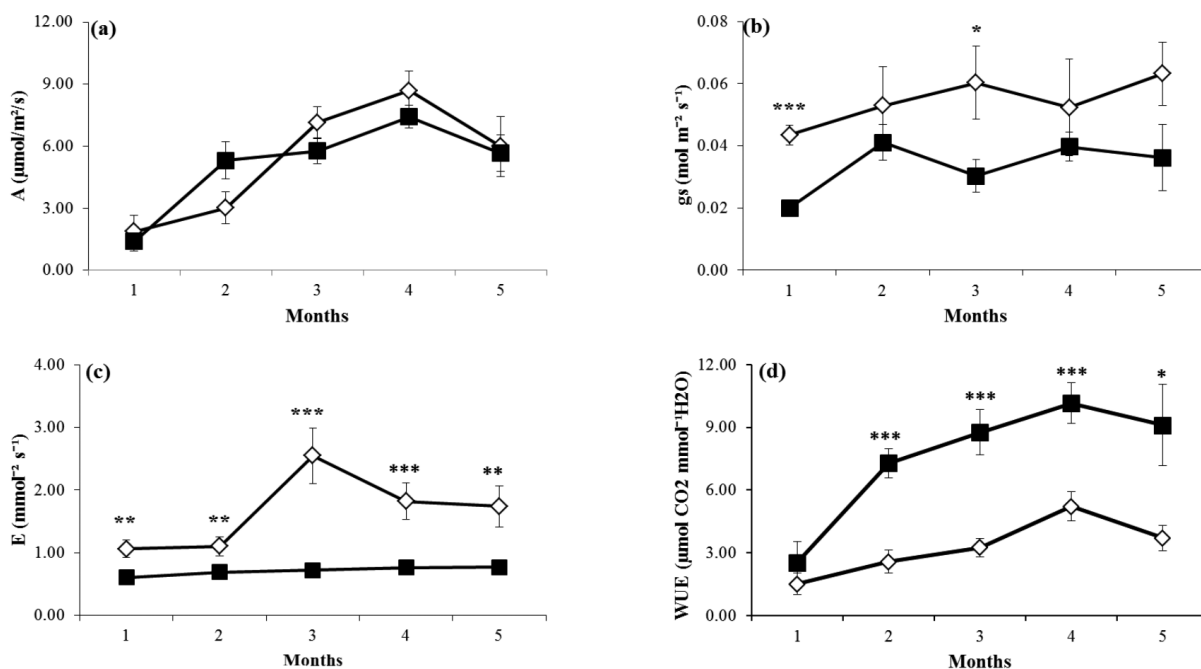


FIGURE 2. The effect of different CO₂ (400±50 μmol mol⁻¹ and 800±50 μmol mol⁻¹) on photosynthetic rate (a), stomatal conductance (b), transpiration rate (c), and water use efficiency (d) of *Shorea platycarpa* in five months of treatment. An ◇ = 400 ± 50 μmol mol⁻¹; ■ = 800 ± 50 μmol mol⁻¹. Values represent means ± SE; n=18. The statistical significance is based on one-way ANOVA (p<0.05). The symbol used indicates *** p<0.001; ** p<0.01 and * p<0.05

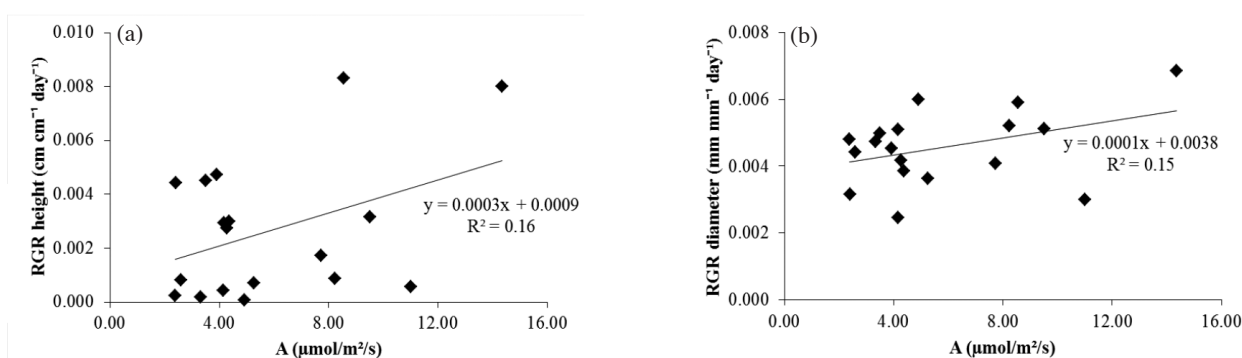


FIGURE 3. Correlation between photosynthetic rate (A) to relative growth rate (RGR) of height (a), ($r = 0.4$, $p = 0.10$) and RGR of stem diameter (b), ($r = 0.4$, $p = 0.11$) of *Shorea platycarpa* in ambient CO₂ and elevated CO₂. Relationship between A and RGR of height and diameter was not significant at ($p < 0.05$)

and *Pinus sylvestris*) and hardwoods (*Quercus ilex* and *Liquidambar styraciflua*) though some studies found no difference in growth ring width (*Picea abies* and *Larix kaempferi*) (Yazaki et al. 2005). Cambial activities, is the frequency of cell division, affects the radial growth of plants. Stem expansion could indicate that elevated CO₂ affects the apical meristem and cambium differently or has different effects on cell division and cell expansion, or both. Elevated CO₂ may affect plant development not only by altering photosynthetic activity, but also by altering apical cell development (Yazaki et al. 2001).

The total number of leaves for *S. platycarpa* in both ambient and elevated CO₂ was increased, however the

growth rate was continually decreased. Low nutrient level could generally limit the NLGR of plants to CO₂ enrichment. Some environmental limitations may slow down growth than photosynthesis. Thus, supposedly, assimilates were not allocated more to growth of those species, but shifted to other allocation, instead (defense system or food reserve) (Novriyanti et al. 2012). It also appears that response could also be related to experimental duration. This climax species was known as slow growing species. Therefore, the stimulation developed very slowly. By prolonging the experimental duration, this species will probably enter a phase during which a significant above ground effect becomes measurable.

EFFECTS OF ELEVATED CO₂ ON BIOMASS

Elevated CO₂ has the potential to yield significant increase of biomass accumulation and this have been observed in many plants. Effects of elevated CO₂ on tree seedlings or juvenile trees generally showed an increase in above ground and below ground biomass (*Gmelina arborea*) (Rasineni et al. 2011). However, in this experiment there were no detectable increase in the above ground and below ground biomass of *S. platycarpa* grown in elevated CO₂. Variation in biomass responses to elevated CO₂ has been attributed to a large number of confounding factors, such as the length of study, plant functional group and species morphological physiology. Dijkstra et al. (2002) reported that elevated CO₂ did significantly increase the above ground biomass of dominant species, *Quercus myrtifolia*, tended to increase the above ground biomass of *Q. chapmanii*, but has no effect on above ground biomass of the subdominant species, *Q. geminata*. Some environmental limitations such as water stress and nutrient deficiency (N, P and K⁺) may retard the biomass allocation. In response to environmental cues, plants re-adjust partitioning of photosynthates to maintain a functional equilibrium between shoot and root growth, since they are interdependent (Mooney & Winner 1991). Increased C supply from elevated atmospheric CO₂ can preferentially induce the distribution of photosynthate below ground because plants tend to allocate photosynthate to tissues needed to acquire the most limiting resource, whereas, when CO₂ is elevated, the most limiting resource becomes water or nutrients (Prior et al. 2011). The production of new leaf or leaf loss and the nutrient availability during treatments might give an added effect on the LAI result. Oikawa et al. (2013) reported that elevated CO₂ did stimulate the LAI in the soybeans cultivars but the LAI was also being regulated by the leaf production rate and leaf loss. The effects of (CO₂) on LAI are probably modified by N availability, which strongly controls leaf area growth (Anten et al. 2003). The LAI of plants in elevated CO₂ might be affected by the insufficiency of N uptake. The elevated CO₂ affect the ability of soil-root system to supply N. In addition, low dosage of N supplied during the experiment might also contribute to this interactive effect of CO₂ and N. Subsequently, this will decrease the acquisition of N in the plants and yet the demand for N is increasing. Kim et al. (2001) found LAI for a given N uptake to be greater

for plants under elevated CO₂, but only when N uptake itself was high, and not when it was low.

EFFECTS OF ELEVATED CO₂ ON PHOTOSYNTHESIS

The present study demonstrates the responses of *S. platycarpa* with significant variation in photosynthetic performance to increasing atmospheric CO₂. Elevated CO₂ did not significantly ($p < 0.05$) increase (A) in *S. platycarpa* leaves (Table 2), however it did induce significant ($p < 0.05$) changes in (g_s), (E) and (WUE) (Table 2). Though the leaf photosynthetic rates (A) give non-significant effects, it is believed that by prolonging the treatments it will significantly changes the outcomes. The photosynthetic rate (A) can be reduced during long term elevated CO₂ exposure, often referred as photosynthetic acclimation or down-regulation. However, tree species have a wide range of responses that are correlated with their successional status. Down-regulation of photosynthesis in *P. macroloba*, a late successional slow growing climax species was decreased after long periods of exposure to elevated CO₂ (Oberbauer et al. 1985) but this effect was not reduced over time in *Luehea seemannii*, a fast growing canopy tree (Lovelock et al. 1999).

On average, the stomatal conductance (g_s) of *S. platycarpa* only decreased by 0.1% in elevated CO₂ compared to their counterparts grown in ambient CO₂. Following the decrease in (g_s), it is expected that plants will use less water and become more efficient for water use. This data is consistent with the classic response of a decrease in stomatal conductance in plants grown in elevated CO₂. Similar result have been observed in climax species, *Pentaclethra macroloba* (Oberbauer et al. 1985), tropical canopy species, *Luehea seemanni* (Lovelock et al. 1999) and shade-tolerant species, *Pongamia pinnata* (Liang et al. 2001). Stomatal conductance (g_s) represents the abilities of stomata, which are the channels of CO₂ absorption and water loss transpiration in plants. As shown in Figure 2(d), (WUE) increased with CO₂, especially on the 2nd, 3rd and 4th months of treatment, suggesting that it is mainly due to significantly decreased of transpiration. The plant reacts to water deficit with a rapid closure of stomata to avoid further loss of water through transpiration. Under high CO₂ environment, reduced (g_s) and (E) resulting in substantial increase in (WUE) which contributed to increased water use efficiency (WUE) (Prior et al. 2011).

TABLE 2. Effects of atmospheric CO₂ (ambient; 400±50 µmol mol⁻¹ and elevated 800±50 µmol mol⁻¹) on photosynthetic rate (A), stomatal conductance (g_s), transpiration rate (E) and water use efficiency (WUE) in *Shorea platycarpa*

Parameters	Ambient CO ₂	Elevated CO ₂	P level
Photosynthetic rate (A) (µmol/m ² /s)	5.35 ± 0.58	5.11 ± 0.42	0.74 ns
Stomatal conductance (g _s) (mol m ⁻² s ⁻¹)	0.22 ± 0.01	0.17 ± 0.00	0.002 **
Transpiration rate (E) (mmol ⁻² s ⁻¹)	0.35 ± 0.09	-0.41 ± 0.05	0.000 ***
Water use efficiency (µmol CO ₂ mmol ⁻¹ H ₂ O)	0.95 ± 0.12	1.78 ± 0.11	0.000 ***

Values represent means ± SE; n=18. The calculation of statistical significant is based on one-way ANOVA ($p=0.05$). The symbol used *** $p < 0.001$; ** $p < 0.01$; * $p < 0.05$; not significant (ns) $p > 0.05$

In this study, *S. platycarpa* did not show any significant ($p < 0.05$) correlation between growth rate and photosynthetic rate. However, according to Kenzo et al. (2011), high growth rate of *Dyera costulata* and *Dipterocarpus baudii* were observed under high light conditions may correspond to the high photosynthetic rate. A variety of hypotheses have been proposed that prolonged exposure to elevated CO₂ might decrease the photosynthetic rate (*A*) of plants. These responses were attributed to secondary responses related to either excess carbohydrate accumulation or decreased N content. Down-regulation or acclimation of photosynthesis is generally accompanied by a large increase in leaf carbohydrates (Cheng et al. 1998). Photosynthetic down regulation has been observed in hybrid larch F1 (*Larix gmelinii* var. *japonica* × *Larix kaempferi*) and *Quercus suber* seedlings. This response was not initiated by stomatal closure but mainly by the reduction in the assimilation capacity which increase the total soluble sugars and starch in leaves (Faria et al. 1996; Watanabe et al. 2011). Excess of carbohydrate remained in the leaf as sugars or starch made leaves heavier per unit area, thus, contributed to an ineffective transformation of photosynthetic carbon gain into new growth (Kirschbaum 2011). Enhanced storage of carbohydrates within the tree may lead to increased growth in the year following the elevated CO₂ treatment (Lovelock et al. 1999).

CONCLUSION

This study pointed out that elevated CO₂ did not give significant effect on the relative growth rate (RGR), photosynthetic rate (*A*) and biomass characteristics of *S. platycarpa*. *S. platycarpa* is a slow growing species. Possibly, the study indicated that elevated carbon concentration does not affect a relatively slow growing tree species and a late successional tree species. However, a longer-term exposure of elevated CO₂ on *S. platycarpa* would confirm the findings of this study. The study was conducted on the sapling. Perhaps, next study should look at different growth stages of *S. platycarpa* such as at seedling and juvenile/semi adult stages.

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REFERENCES

- Anten, N.P.R., Hirose, T., Onoda, Y., Kinugasa, T., Kim, H.Y., Okada, M. & Kobayashi, K. 2003. Elevated CO₂ and nitrogen availability have interactive effects on canopy carbon gain in rice. *New Phytologist* 161: 459-471.
- Ashton, P.S., Givnish, T.J. & Appanah, S. 1988. Staggered flowering in the Dipterocarpaceae: New insights into floral induction and the evolution of mast fruiting in the aseasonal tropics. *The American Naturalist* 132(1): 44-66.
- Cheng, S., Moore, B. & Seemann, J.R. 1998. Effects of short- and long-term elevated CO₂ on the expression of ribulose-1,5-bisphosphate carboxylase/oxygenase genes and carbohydrate accumulation in leaves of *Arabidopsis thaliana* (L.) Heynh. *Plant Physiology* 116: 715-723.
- De Deyn, G.B., Cornelissen, J.H.C. & Bardgett, R.D. 2008. Plant functional traits and soil carbon sequestration in contrasting biomes. *Ecology Letters* 11(5): 516-531.
- Dijkstra, P., Hymus, G., Colavito, D., Vieglais, D.A., Cundari, C.M. & Johnson, D.P. 2002. Elevated atmospheric CO₂ stimulates aboveground biomass in a fire-regenerated scrub-oak ecosystem. *Global Change Biology* 8: 90-103.
- Faria, T., Wilkins, D., Besford, R.T., Pereira, M.V.J.S. & Chaves, M.M. 1996. Growth at elevated CO₂ leads to down-regulation of photosynthesis and altered response to high temperature in *Quercus suber* L. seedlings. *Journal of Experimental Botany* 47(304): 1755-1761.
- Gan, K.S., Lim, S.C. & Choo, K.T. 1998. Timber Notes - Light Hardwoods (I). (Meranti bakau, Dark red meranti, Light red meranti, White meranti, Yellow meranti). *Timber Technology Bulletin* No. 9.
- Hamzah, K.A., Parlan, I., Kassim, A.R., Hassan, C.H., Akeng, G. & Said, N.M. 2009. Ecological characteristics of a *Gonystylus bancanus*-rich area in Pekan Forest Reserve, Pahang, Malaysia. *Tropical Life Sciences Research* 20(2): 15-27.
- IPCC. 2014. *Climate Change 2014: Synthesis Report. Contribution of Working Groups I, II and III to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*, edited by Core Writing Team, Pachauri, R.K. & Meyer, L.A. IPCC, Geneva, Switzerland. p. 151.
- Jach, M.E. & Ceulemans, R. 1999. Effects of elevated atmospheric CO₂ on phenology, growth and crown structure of Scots pine (*Pinus sylvestris*) seedlings after two years of exposure in the field. *Tree Physiology* 19: 289-300.
- Karnosky, D.F. 2003. Impacts of elevated atmospheric CO₂ on forest trees and forest ecosystems: Knowledge gaps. *Environment International* 29: 161-169.
- Kenzo, T., Yoneda, R., Matsumoto, Y., Azani, A.M. & Majid, M.N. 2011. Growth and photosynthetic response of four Malaysian indigenous tree species under different light conditions. *Journal of Tropical Forest Science* 23(3): 271-281.
- Kim, H.Y., Lieffering, M., Miura, S., Kobayashi, K. & Okada, M. 2001. Growth and nitrogen uptake of CO₂-enriched rice under field conditions. *New Phytologist* 150: 223-229.
- Kirschbaum, M.U.F. 2011. Does enhanced photosynthesis enhance growth? Lessons learned from CO₂ enrichment studies. *Plant Physiology* 155(1): 117-124.
- Leakey, A.D.B., Bishop, K. & Ainsworth, E. 2012. A multi-biome gap in understanding of crop and ecosystem responses to elevated CO₂. *Current Opinion in Plant Biology* 15(3): 228-236.

- Liang, N., Tang, Y. & Okuda, T. 2001. Is elevation of carbon dioxide concentration beneficial to seedling photosynthesis in the understory of tropical rain forests? *Tree Physiology* 21: 1047-1055.
- Long, S.P., Ainsworth, E.A., Rogers, A. & Ort, D.R. 2004. Rising atmospheric carbon dioxide: Plants FACE the future. *Annu. Rev. Plant Biol.* 55: 591-628.
- Lovelock, C.E., Virgo, A., Popp, M., Winter, K. & Environmental, S. 1999. Effects of elevated CO₂ concentrations on photosynthesis, growth and reproduction of branches of the tropical canopy tree species, *Luehea seemannii* Tr. & Planch. *Plant, Cell and Environment* 22: 49-59.
- Mooney, H.A. & Winner, W.E. 1991. Partitioning response of plants to stress. In *Response of Plants to Multiple Stresses*, edited by Mooney, H.A., Winner, W.E. & Pell, E.J. New York: Academic Press.
- Novriyanti, E., Watanabe, M., Kitao, M., Utsugi, H. & Uemura, A. 2012. High nitrogen and elevated [CO₂] effects on the growth, defense and photosynthetic performance of two eucalypt species. *Environmental Pollution* 170: 124-130.
- Oberbauer, S.V., Strain, B.R. & Fetcher, N. 1985. Effects of CO₂ enrichment on seedling physiology and growth of two tropical trees. *Physiologia Plantarum* 65: 352-356.
- Oikawa, S., Okada, M. & Hikosaka, K. 2013. Effects of elevated CO₂ on leaf area dynamics in nodulating and non-nodulating soybean stands. *Plant Soil* 373: 627-639.
- Poorter, H. & Pérez-Soba, M. 2001. The growth response of plants to elevated CO₂ under non-optimal environmental conditions. *Oecologia* 129(1): 1-20.
- Prior, S., Runion, G., Marble, S., Rogers, H., Gilliam, C. & Allen Torbert, H. 2011. A review of elevated atmospheric CO₂ effects on plant growth and water relations: Implications for horticulture. *Horticultural Science* 46(2): 158-162.
- Rasineni, G.K., Guha, A. & Reddy, A.R. 2011. Responses of *Gmelina arborea*, a tropical deciduous tree species, to elevated atmospheric CO₂: Growth, biomass productivity and carbon sequestration efficacy. *Plant Science* 181(4): 428-438.
- Watanabe, M., Watanabe, Y., Kitaoka, S., Utsugi, H., Kita, K. & Koike, T. 2011. Growth and photosynthetic traits of hybrid larch F1 (*Larix gmelinii* var. japonica × *L. kaempferi*) under elevated CO₂ concentration with low nutrient availability. *Tree Physiology* 31: 965-975.
- Yazaki, K., Funada, R.Y.O., Mori, S., Maruyama, Y., Abaimov, A.P., Kayama, M. & Koike, T. 2001. Growth and annual ring structure of *Larix sibirica* grown at different carbon dioxide concentrations and nutrient supply rates. *Tree Physiology* 3: 1223-1229.
- Yazaki, K., Maruyama, Y., Mori, S., Koike, T. & Funada, R. 2005. Effects of elevated carbon dioxide concentration on wood structure and formation in trees. In *Plant Responses to Air Pollution and Global Changes*, edited by Omasa, K., Nouchi, I. & De Kok, L.J. Tokyo: Springer-Verlag. pp. 89-97.
- Zagt, R.J. & Werger, M.J.A. 1998. Community structure and the demography of primary species in tropical rain forest. In *Dynamics of Tropical Communities*, edited by Newbery, D.M., Prins, H.H.T. & Brown, N.D. Oxford: Blackwell Science. pp. 193-219.

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