



DIGITAL ACCESS TO
SCHOLARSHIP AT HARVARD
DASH.HARVARD.EDU



HARVARD LIBRARY
Office for Scholarly Communication

Cutaneous melanoma in women###

The Harvard community has made this article openly available. [Please share](#) how this access benefits you. Your story matters

Citation	Roh, Mi Ryung, Philip Eliades, Sameer Gupta, Jane M. Grant-Kels, and Hensin Tsao. 2015. "Cutaneous melanoma in women###." International Journal of Women's Dermatology 3 (1 Suppl): S11-S15. doi:10.1016/j.ijwd.2017.02.003. http://dx.doi.org/10.1016/j.ijwd.2017.02.003 .
Published Version	doi:10.1016/j.ijwd.2017.02.003
Citable link	http://nrs.harvard.edu/urn-3:HUL.InstRepos:33029908
Terms of Use	This article was downloaded from Harvard University's DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA



Cutaneous melanoma in women☆☆☆



Mi Ryung Roh, MD^{a,b}, Philip Eliades, BS^{a,c}, Sameer Gupta, BA^a,
Jane M. Grant-Kels, MD^d, Hensin Tsao, MD, PhD^{a,*}

^a Wellman Center for Photomedicine, Department of Dermatology, Massachusetts General Hospital, Harvard Medical School, Boston, MA

^b Department of Dermatology, Cutaneous Biology Research Institute, Yonsei University College of Medicine, Seoul, Korea

^c Tufts University School of Medicine, Boston, MA

^d Dermatology Department, University of CT Health Center, Farmington, CT

ARTICLE INFO

Article history:

Received 13 December 2014

Received in revised form 12 January 2015

Accepted 13 January 2015

ABSTRACT

Background: Gender disparity in melanoma outcome is consistently observed, suggesting that gender is as an important prognostic factor. However, the source of this gender disparity in melanoma remains unclear.

Objective: This article reviews advances in our understanding of gender differences in melanoma and how such differences may contribute to outcomes.

Methods: A broad literature search was conducted using the PubMed database, with search terms such as 'gender differences in melanoma' and 'sex differences in melanoma.' Additional articles were identified from cited references.

Results: Herein, we address the gender-linked physiologic differences in skin and melanoma. We discuss the influence of estrogen on a woman's risk for melanoma and melanoma outcomes with regard to pregnancy, oral contraceptives, hormone replacement therapy, and UV tanning.

Conclusions: The published findings on gender disparities in melanoma have yielded many advances in our understanding of this disease. Biological, environmental, and behavioral factors may explain the observed gender difference in melanoma incidence and outcome. Further research will enable us to learn more about melanoma pathogenesis, with the goal of offering better treatments and preventative advice to our patients.

© 2015 The Authors. Published by Elsevier Inc. on behalf of Women's Dermatologic Society. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Introduction

The incidence of cutaneous melanoma (CM) continues to increase in the Caucasian population in the United States. In 2014, women only accounted for 42% of the 76,100 new melanoma cases and only 33% of the 9,710 deaths associated with CM in the United States [1]. These trends are consistently observed in populations around the world. Indeed, gender disparity in melanoma outcome is so consistently observed that gender has been suggested as an important prognostic factor, despite not being formerly incorporated in staging algorithms [2]. The source of this gender disparity in melanoma remains unclear, but likely represents both biological and behavioral etiologies. Here, we review the current knowledge of how the disease of melanoma differs between men and women.

Gender-linked physiologic differences in skin

Skin is a dynamic, complex, integrated arrangement of cells, tissues, and matrix elements that mediates a diverse array of functions, includ-

ing physical permeability barrier, protection from infectious agents, thermoregulation, sensation, ultraviolet (UV) protection, wound repair and regeneration, and outward physical appearance. These various functions of skin are mediated by its major layers: the epidermis, dermis, and subcutaneous fat. The dermis contains water, ground substance, and elastic fibers that, in combination with the layer of subcutaneous fat, account for the majority of the skin's thickness. The thickness of the skin is greater in men than in women at all ages [3,4]. Skin thickness decreases in men and women starting at the age of 45, and women's skin gets 10% thinner after menopause [5,6]. Skin color is modulated by melanin, hemoglobin, and other chromophores. Melanin is synthesized in melanocytes, dendritic cells located in the basal layer of the epidermis. Melanocytes are known to be differently distributed according to anatomic sites. No gender-related difference has been reported; however, within individual ethnic groups, men have been reported to have darker skin color [7], which may be related to a more vascularized upper dermis [8] and more melanin synthesis [9]. These differences are likely modulated, in part, by hormones, given that they manifest during puberty and increase with age [10].

Men and women differ in the metabolism of and response to androgens and estrogens [11]. Estrogens are known to accelerate wound healing, improve inflammatory disorders, increase epidermal thickness, and protect against photoaging of the skin [12]. The cellular effects of estrogens are mediated by estrogen receptors (ERs), ER α and ER β , which

☆ Conflicts of interest: The authors state no conflicts of interest.

☆☆ This article is a reprint of a previously published article. For citation purposes, please use the original publication details; International Journal of Women's Dermatology 1 (2015) 21–25. DOI of original item: [10.1016/j.ijwd.2015.01.001](https://doi.org/10.1016/j.ijwd.2015.01.001).

* Corresponding author.

E-mail address: htsao@partners.org (H. Tsao).

belong to the nuclear steroid hormone receptor superfamily. ER α and ER β are widely expressed in human tissue, but have differential distributions in various tissues, including the skin. ER α is primarily expressed in the uterus, liver, kidneys, breasts, and heart, whereas ER β is primarily detected in what are known as nonclassical estrogen-responsive tissues: the ovaries, colon, lungs, adipose tissue, prostate, bladder, and skin [13]. Notably, melanomas do express ER β , but its expression does not differ between male and female tumors [14]. Cutaneous ER levels are generally known to be higher in women as compared with men. However, relative levels of ER α and ER β in men and women are not well understood. In women, the amount of ERs declines after menopause with declining levels of estradiol, which, through positive feedback, has a pro-synthetic effect on ERs [13]. Expression declines more rapidly in ER β than in ER α , resulting in an increased ER α :ER β ratio in the skin. In contrast to estrogens, androgens—such as testosterone and 5 α -dihydroxytestosterone—may be able to promote melanoma tumorigenesis. Androgen receptors, which have a similar mechanism of action as estrogen receptors, have been described in human melanoma cells [15,16].

There are also several baseline differences in the immune systems of men and women. On average, women have higher measured IgG and IgM levels, as well as a greater percentage of CD3 + T lymphocytes, as compared with men, suggesting that men have a relative attenuation of the adaptive immune response compared with women [17]. This is further evidenced by observations that men are more susceptible to bacterial and viral infections [18], while women are more prone to autoimmune and inflammatory diseases [19]. Men are also more prone to skin cancer; this increased risk may be partly explained by their heightened susceptibility to ultraviolet-induced immunosuppression compared with women [20]. Ultraviolet (UV) irradiation is known to inhibit contact hypersensitivity or delayed-type hypersensitivity. Also, the sex hormones may have an additional differential effect on immune cells, as both estrogen and androgen receptors are expressed in immune cells [21].

The mechanisms and mechanics of oxidative stress also differ between sexes. By-products of oxygen metabolism lead to the production of reactive oxygen species (ROS), which can damage a wide range of molecules, leading to oxidative stress [22]. Male cells of various tissue types have an elevated ROS cellular environment, due to the expression of lower levels of antioxidant enzymes as compared with females [23]. Additionally, estrogens appear to have direct antioxidant and protective effects, while testosterone may potentiate oxidative stress. This difference between testosterone and estrogen may contribute to disparities in melanoma outcome [24].

Gender-specific differences in melanoma

Gender-specific differences in melanoma epidemiology are well established. The probability of developing melanoma during one's lifetime is 1.72% in males and 1.22% in females [25]. In the Netherlands, a large population-based cohort study including 10,538 melanoma patients from 1993 to 2004 analyzed the gender difference in melanoma survival after adjusting for tumor-related variables (Breslow thickness, histology, tumor site, and metastatic and nodal status). They found that the relative excess risk of mortality was 2.70 (95% CI [2.38, 3.06]) in males versus females. The female survival advantage remained after adjusting for multiple confounding variables, including tumor thickness [26]. Gamba et al. [27] analyzed data from 26,107 individuals, aged 15–39 years, from the U.S. National Cancer Institute's Surveillance, Epidemiology, and End Results registry. They reported that young men had a 55% lower rate of melanoma survival compared with age-matched young women, and concluded that male sex, within all specific age groups and across all tumor thickness categories, histologic subtypes, and anatomic sites, is associated with a disproportionate burden of melanoma deaths. A population-based cohort study including 11,774 melanoma patients from the Munich Cancer Registry (Germany) analyzed gender differences in survival and disease progression across all stages of CM. Localized melanomas in women showed a lower risk of metastasis, resulting in better survival when compared with men, even after first disease progression [28]. In

localized melanoma, men generally had worse characteristics at diagnosis, such as older age; increased likelihood of having an ulcerated or thicker primary tumor; and melanomas more commonly located on the head, neck, and trunk instead of the extremities. However, even after diagnosis, men continue to have disadvantages compared with women [28]. In various studies, women showed a longer delay before relapse and a higher cure rate compared to men [26,28]. Table 1 summarizes the sex risk estimates of melanoma survival in various studies.

The natural history of melanoma in women parallels the physiologic hormonal changes they experience. The incidence of melanoma is rare before puberty, rises abruptly through the reproductive ages until approximately 50 years of age, and then diminishes after menopause [33]. In agreement with the aforementioned survival advantages of young women and women with localized melanoma, women who present with stage IV disease also show higher survival rates compared to men, who have a two-fold greater death rate from melanoma [34]. Kemeny et al. [35] also reported that premenopausal women show a higher survival rate compared to postmenopausal women, which was more pronounced in women with advanced disease. Although the survival advantage decreases with age, postmenopausal women still have better rates of survival compared with men.

Gender disparity in melanoma may have genetic underpinnings. Kocarnik et al. [36] confirmed eight single nucleotide polymorphisms that modulate melanoma risk. Interestingly, one single nucleotide polymorphism, rs16891982, in the *SLC45A2* gene influenced melanoma risk differently by sex, with higher risk association observed in males (*OR* 5.5 in males vs. 2.37 in females). The *SLC45A2* gene codes for an ion transporter protein in the melanosome and likely shapes melanoma risk by modulating melanosome activity by changing the internal pH [37]. Notably, estrogen increases expression of tyrosinase [38], the rate-limiting enzyme in melanin production located in the melanosome, and rs16891982 has been associated in this enzyme's activity and expression [39]. Synthesizing these observations, Kocarnik et al. [36] hypothesize that the gender-modified risk associated with rs16891982 may be mediated through hormonal influence on a critical enzyme in melanogenesis. While this study explored inherited germline polymorphisms, inherent differences in genetic burden or immunogenicity of the tumors themselves are an intriguing, yet poorly explored, area of research.

Estrogen and melanoma

The most widely studied mechanism regarding gender disparity in melanoma is the influence of sex hormone levels and estrogen receptor expression. Various laboratory studies with cultured cells and animal models support a role for estrogens in contributing to the better survival rates among women. Kanda and Watanabe [40] showed that incubation of 17- β -estradiol inhibited the growth of human metastatic melanoma cells *in vitro*, and concomitantly reduced constitutive interleukin-8 (IL-8) mRNA and secretion. The melanoma cell growth inhibited by 17- β -estradiol was counteracted by exogenously added IL-8, which suggests that estrogen mediates growth suppression in melanoma cells by inhibiting IL-8 expression. Of note, this effect was only observed in ER-positive cells, indicating that estrogen may mediate an inhibitory effect on melanomas via ER and IL-8 *in vitro*. Richardson et al. [41] and Roy et al. [42] reported that 17- β -estradiol inhibited the invasive activity of melanoma cells, but did not have antitumor activity in C57BL/6 mice inoculated with syngeneic B16 tumors. Cho et al. [43] reported that implanted B16 murine melanoma cells grew more rapidly in ER β knockout mice compared to congenic C56BL/6 mice with intact ER β , therefore suggesting that the presence of ER β was protective against tumor growth. It has been demonstrated that melanocytic lesions express detectable levels of ER α and ER β mRNA and protein [44,45]. However, ER β was the predominant ER type in melanocytic lesions, indicating that estrogen might play a role in melanocyte physiology via ER β [44]. ER β immunoreactivity of melanoma cells was dependent on the microenvironment: melanoma cells in close contact with keratinocytes were more reactive than invasive melanoma, and the

Table 1
Summary of Selected Studies Demonstrating Female Survival Advantage.

Reference	Year	End Point	Country	No. of Patients	Adjusted Risk Estimates ¹	95% CI
Balch et al. [29]	2001	DSS	United States	13,581	0.84	0.76 to 0.92
de Vries et al. [26]	2008	RS	The Netherlands	10,538	0.53 ^{2,3}	0.48 to 0.61
Xing et al. [30]	2010	DSS	United States (SEER)	37,519	0.67 ³	0.60 to 0.75
Joose et al. [28]	2011	DSS	Germany	11,774	0.62	0.56 to 0.70
Collins et al. [31]	2011	DSS ⁴	United States (SEER)	142,653	0.65 ³	0.62 to 0.68
Thompson et al. [32]	2011	DSS	International AJCC Consortium	10,233	0.69	0.61 to 0.79

Abbreviations: AJCC, American Joint Committee on Cancer; DSS, disease-specific survival; RS, relative survival (estimate of DSS).

¹ Relative risk of women compared with men; presented as hazard ratio unless otherwise specified.

² Presented as relative excess risk.

³ Value reported here is the inverse of the original risk estimate, because men were compared with women in the cited publication.

⁴ For patients who underwent surgery.

ER β protein expression decreased progressively with increased Breslow thickness and resulted in more invasive melanoma [46]. Another study demonstrated that ER β expression in CMs without and with lymph node metastases were higher and lower, respectively, than in the healthy skin surrounding them, suggesting a role for ER β in the metastatic process of melanoma [44]. Together, these studies suggest that the loss of ER β expression may be an important step in melanoma progression and/or the stimulation of melanoma proliferation.

Pregnancy, oral contraceptives, and hormone replacement therapy

Approximately one third of all women diagnosed with CM are of childbearing age [47]. The observation of physiological hyperpigmentation on multiple cutaneous sites during pregnancy yielded the hypothesis that pregnancy-related hormones might influence the course of CM in women. Pregnancy is also considered to be a state of immunosuppression, one that helps prevent the rejection of the fetus expressing paternal alloantigens [48]. However, recent studies found no significant dermatoscopic or abnormal histologic changes in nevi during pregnancy [49]. Also, clinical, epidemiologic, and laboratory studies have found that pregnancy does not significantly change the characteristics or prognosis of CM [50]. Multiple controlled studies in the 1980s consistently showed no significant impact on the survival of women diagnosed with localized melanoma (AJCC stage I or II) during pregnancy [51–56]. Since pregnancy does not seem to influence the prognosis or evolution of localized malignant melanoma, the recommendations set forth by the European Society of Medical Oncology for evaluating a patient diagnosed with melanoma remain the same regardless of pregnancy status [57]. Counseling patients diagnosed with melanoma regarding future pregnancies and hormone replacement therapy should be based on established prognostic factors. If poor prognostic factors exist, clinicians will often advise patients to wait 2 to 3 years to become pregnant, thereby avoiding pregnancy during the window of time with the highest likelihood of recurrence [50]. However, there are no standardized guidelines for the postponement of the further pregnancies after the diagnosis of melanoma; counseling must be individualized for each patient, taking into consideration her age, fertility status, availability of oocyte cryopreservation, disease prognosis, and family support.

Multiple studies have investigated the relationship between the use of oral contraceptives (OCs) and the risk of CM, including two recently published meta-analyses. The first of these, an analysis of 18 studies, reported an overall OR of 0.95 (95% CI [0.82, 1.15]) [58]. The second, an analysis of 10 studies encompassing 3,796 cases and 9,442 controls, also reported a summation OR of 0.95 (95% CI [0.87, 1.04]) [59]. The authors of a pooled analysis of 10 case-control studies also failed to find an association between risk of CM and ever-use of OCs (pooled OR, 0.86; 95% CI [0.74, 1.01]) [60]. There were also no relationships between risk of melanoma and OC length of use, age at first use, or current use. These studies all make the consensus conclusion that there is no evidence of a relationship between risk of melanoma and use of OCs. Based on a limited number of studies, a similar conclusion has been made regarding the relationship, or lack thereof, between hormone replacement therapy (HRT) and risk of malignant

melanoma. Gupta and Driscoll [61] analyzed 12 studies, 10 of which showed no association between use of HRT and melanoma. The two studies that showed increased risk of melanoma with HRT use did not control for potential confounding variables. Consequently, a history of localized melanoma is not considered a contraindication for the use of HRT.

UVA tanning: a major melanoma risk factor for young women

Melanoma is the most prevalent cancer among 25- to 29-year-old females, and under the age of 40, women have a higher incidence of melanoma than men [62]. In several studies, risky tanning behavior was more common among women, younger people, and people of low socioeconomic status [63]. These emerging data on the incidence rates of skin cancers in young women younger than age 40 years compared with men of that age group suggest that the etiology is partly due to excessive, repeated exposures to unnaturally large amounts of ultraviolet A (UVA) light from UVA-rich lamps. Indoor tanning has been widely practiced in northern Europe and the United States since the 1980s [64]. Indoor tanning equipment mainly emits light in the UVA range, but a fraction (<5%) of this spectrum is in the ultraviolet B range [65]. In frequent users of UVA-rich lamps, the amount of annual UVA exposure is roughly doubled. It was also reported that high-pressure UVA tanning beds, with dose rates up to 13 times that of the summer sun, have the potential to quadruple the annual UVA exposure of frequent tanners [66]. Two recent meta-analyses [65,67] reported that the risk of CM is increased by 16% and 20%, respectively, for those who have ever used indoor tanning devices. Even more concerning, the risk of melanoma doubled when use started before the age of 35 years. These findings—that early exposure and more frequent use of indoor tanning beds significantly increases the risk of melanoma—represent a global trend [65]. Therefore, in 2009, the International Agency for Research on Cancer classified the entire ultraviolet spectrum and indoor tanning devices as carcinogenic to humans (group 1) [68]. This classification was based on evidence from basic and epidemiologic studies demonstrating that UVA has an important role in skin carcinogenesis [69] and that UVA-producing tanning lamps are capable of inducing the types of DNA damage associated with photocarcinogenesis [70].

As melanomas arising in young women are commonly associated with tanning bed use, these diseases can be minimized by educating patients to avoid artificial tanning habits and strongly regulating their use. The World Health Organization, the International Commission on Non-ionizing Radiation Protection, and the European Society of Skin Cancer Prevention have all maintained that the highest regulatory priorities should be the restriction of sunbed use by people younger than 18 years of age and the banning of unsupervised indoor tanning facilities. Such restrictions have been implemented in Australia and several European countries, including Austria, France, Germany, Portugal, Belgium, Scotland, and Spain.

In May 2014, the U.S. Food and Drug Administration formally reclassified sunlamp products and UV lamps intended for use in sunlamp products from low-risk (class I) to moderate-risk (class II) devices. Along with this reclassification came the mandate that all sunlamp

products carry a visible black-box warning on the device that explicitly states that the sunlamp product should not be used on persons younger than 18 years of age. Furthermore, certain marketing information associated with these devices must provide additional and specific warning statements and contraindications (<http://www.fda.gov/NewsEvents/Newsroom/PressAnnouncements/ucm399222.htm>). On legislative fronts, only nine states (CA, DE, IL, LA, MN, NV, OR, TX, and VT) have banned access to indoor tanning for adolescents younger than age 18 years.

Conclusion

Mounting evidence suggests that both biological and environmental factors explain observed gender differences in melanoma incidence and outcome. Innate gender differences in hormones, immune homeostasis, and oxidative stress likely contribute to the sexual disparity in melanoma survival. Beyond these innately biological differences, behaviors, such as tanning bed use, also clearly shape melanoma risk. Understanding gender differences allows for targeted interventions. For instance, increased awareness of the modifiable risks of tanning behavior has yielded bans in nine states, with pending legislation in additional states that promises to explicitly ban under-18 indoor tanning. Similarly, better understanding of innate biological differences in melanoma between the genders may yield improved targeted therapies.

References

- [1] Siegel R, Ma J, Zou Z, Jemal A. Cancer statistics, 2014. *CA Cancer J Clin* 2014;64:9–29.
- [2] Homsji J, Kashani-Sabet M, Messina JL, Daud A. Cutaneous melanoma: prognostic factors. *Cancer Control* 2005;12:223–9.
- [3] Sandby-Moller J, Poulsen T, Wulf HC. Epidermal thickness at different body sites: relationship to age, gender, pigmentation, blood content, skin type and smoking habits. *Acta Derm Venereol* 2003;83:410–3.
- [4] Shuster S, Black MM, McVitie E. The influence of age and sex on skin thickness, skin collagen and density. *Br J Dermatol* 1975;93:639–43.
- [5] Leveque JL, Corcuff P, de Rigo J, Agache P. In vivo studies of the evolution of physical properties of the human skin with age. *Int J Dermatol* 1984;23:322–9.
- [6] Panyakhamlerd K, Chotnopparattapattara P, Taechakraichana N, Kukulprasong A, Chaikittisilpa S, Limpaphayom K. Skin thickness in different menopausal status. *J Med Assoc Thai* 1999;82:352–6.
- [7] Kalla A, Tiwari SC. Sex differences in skin colour in man. *Acta Genet Med Gemellol (Roma)* 1970;19:472–6.
- [8] Tur E. Physiology of the skin—differences between women and men. *Clin Dermatol* 1997;15:5–16.
- [9] Mehrai H, Sunderland E. Skin colour data from Nowshahr City, northern Iran. *Ann Hum Biol* 1990;17:115–20.
- [10] Kelly RI, Pearce R, Bull RH, Leveque JL, Mortimer PS. The effects of aging on the cutaneous microvasculature. *J Am Acad Dermatol* 1995;33:749–56.
- [11] Chen W, Thiboutot D, Zouboulis CC. Cutaneous androgen metabolism: basic research and clinical perspectives. *J Invest Dermatol* 2002;119:992–1007.
- [12] Thornton MJ. The biological actions of estrogens on skin. *Exp Dermatol* 2002;11:487–502.
- [13] Hall G, Phillips TJ. Estrogen and skin: the effects of estrogen, menopause, and hormone replacement therapy on the skin. *J Am Acad Dermatol* 2005;53:555–68 [quiz 69–72].
- [14] Ohata C, Tadokoro T, Itami S. Expression of estrogen receptor beta in normal skin, melanocytic nevi and malignant melanomas. *J Dermatol* 2008;35:215–21.
- [15] Allil PA, Visconti MA, Castrucci AM, Isoldi MC. Photoperiod and testosterone modulate growth and melanogenesis of s91 murine melanoma. *Med Chem* 2008;4:100–5.
- [16] Morvillo V, Luthy IA, Bravo AI, Capurro MI, Donaldson M, Quintans C, et al. Atypical androgen receptor in the human melanoma cell line IIB-MEL-J. *Pigment Cell Res* 1995;8:135–41.
- [17] Bouman A, Heineman MJ, Faas MM. Sex hormones and the immune response in humans. *Hum Reprod Update* 2005;11:411–23.
- [18] Brown Jr LA. Pathogenesis and treatment of pseudofolliculitis barbae. *Cutis* 1983;32:373–5.
- [19] Da Silva JA. Sex hormones and glucocorticoids: interactions with the immune system. *Ann N Y Acad Sci* 1999;876:102–17 [discussion 17–8].
- [20] Dao Jr H, Kazin RA. Gender differences in skin: a review of the literature. *Gend Med* 2007;4:308–28.
- [21] Damian DL, Patterson CR, Stapelberg M, Park J, Barnetson RS, Halliday GM. UV radiation-induced immunosuppression is greater in men and prevented by topical nicotinamide. *J Invest Dermatol* 2008;128:447–54.
- [22] Malorni W, Campesi I, Straface E, Vella S, Franconi F. Redox features of the cell: a gender perspective. *Antioxid Redox Signal* 2007;9:1779–801.
- [23] Vina J, Borrás C, Gambini J, Sastre J, Pallardo FV. Why females live longer than males? Importance of the upregulation of longevity-associated genes by oestrogenic compounds. *FEBS Lett* 2005;579:2541–5.
- [24] Jousse A, de Vries E, van Eijck CH, Eggermont AM, Nijsten T, Coebergh JW. Reactive oxygen species and melanoma: an explanation for gender differences in survival? *Pigment Cell Melanoma Res* 2010;23:352–64.
- [25] Tremblay GB, Tremblay A, Copeland NG, Gilbert DJ, Jenkins NA, Labrie F, et al. Cloning, chromosomal localization, and functional analysis of the murine estrogen receptor beta. *Mol Endocrinol* 1997;11:353–65.
- [26] de Vries E, Nijsten TE, Visser O, Bastiaannet E, van Hattem S, Janssen-Heijnen ML, et al. Superior survival of females among 10,538 Dutch melanoma patients is independent of Breslow thickness, histologic type and tumor site. *Ann Oncol* 2008;19:583–9.
- [27] Gamba CS, Clarke CA, Keegan TH, Tao L, Swetter SM. Melanoma survival disadvantage in young, non-Hispanic white males compared with females. *JAMA Dermatol* 2013;149:912–20.
- [28] Jousse A, de Vries E, Eckel R, Nijsten T, Eggermont AM, Holzel D, et al. Gender differences in melanoma survival: female patients have a decreased risk of metastasis. *J Invest Dermatol* 2011;131:719–26.
- [29] Balch CM, Soong SJ, Gershenwald JE, Thompson JF, Reintgen DS, Cascinelli N, et al. Prognostic factors analysis of 17,600 melanoma patients: validation of the American Joint Committee on Cancer melanoma staging system. *J Clin Oncol* 2001;19:3622–34.
- [30] Xing Y, Chang GJ, Hu CY, Askew RL, Ross MI, Gershenwald JE, et al. Conditional survival estimates improve over time for patients with advanced melanoma: results from a population-based analysis. *Cancer* 2010;116:2234–41.
- [31] Collins KK, Fields RC, Baptiste D, Liu Y, Moley J, Jeffe DB. Racial differences in survival after surgical treatment for melanoma. *Ann Surg Oncol* 2011;18:2925–36.
- [32] Thompson JF, Soong SJ, Balch CM, Gershenwald JE, Ding S, Coit DG, et al. Prognostic significance of mitotic rate in localized primary cutaneous melanoma: an analysis of patients in the multi-institutional American Joint Committee on Cancer melanoma staging database. *J Clin Oncol* 2011;29:2199–205.
- [33] Strouse JJ, Fears TR, Tucker MA, Wayne AS. Pediatric melanoma: risk factor and survival analysis of the surveillance, epidemiology and end results database. *J Clin Oncol* 2005;23:4735–41.
- [34] Korn EL, Liu PY, Lee SJ, Chapman JA, Niedzwiecki D, Suman VJ, et al. Meta-analysis of phase II cooperative group trials in metastatic stage IV melanoma to determine progression-free and overall survival benchmarks for future phase II trials. *J Clin Oncol* 2008;26:527–34.
- [35] Kemeny MM, Busch E, Stewart AK, Menck HR. Superior survival of young women with malignant melanoma. *Am J Surg* 1998;175:437–44 [discussion 44–5].
- [36] Kocarnik JM, Park SL, Han J, Dumitrescu L, Cheng I, Wilkens LR, et al. Replication of associations between GWAS SNPs and melanoma risk in the Population Architecture Using Genomics and Epidemiology (PAGE) study. *J Invest Dermatol* 2014;134:2049–52.
- [37] Kondo T, Hearing VJ. Update on the regulation of mammalian melanocyte function and skin pigmentation. *Expert Rev Dermatol* 2011;6:97–108.
- [38] Kim NH, Cheong KA, Lee TR, Lee AY. PDZK1 upregulation in estrogen-related hyperpigmentation in melasma. *J Invest Dermatol* 2012;132:2622–31.
- [39] Cook AL, Chen W, Thurber AE, Smit DJ, Smith AG, Bladen TG, et al. Analysis of cultured human melanocytes based on polymorphisms within the SLC45A2/MATP, SLC24A5/NCKX5, and OCA2/P loci. *J Invest Dermatol* 2009;129:392–405.
- [40] Kanda N, Watanabe S. 17beta-estradiol, progesterone, and dihydrotestosterone suppress the growth of human melanoma by inhibiting interleukin-8 production. *J Invest Dermatol* 2001;117:274–83.
- [41] Richardson B, Price A, Wagner M, Williams V, Lorigan P, Browne S, et al. Investigation of female survival benefit in metastatic melanoma. *Br J Cancer* 1999;80:2025–33.
- [42] Roy S, Reddy BS, Sudhakar G, Kumar JM, Banerjee R. 17beta-estradiol-linked nitro-L-arginine as simultaneous inducer of apoptosis in melanoma and tumor-angiogenic vascular endothelial cells. *Mol Pharm* 2011;8:350–9.
- [43] Cho JL, Allanson M, Reeve VE. Oestrogen receptor-beta signalling protects against transplanted skin tumour growth in the mouse. *Photochem Photobiol Sci* 2010;9:608–14.
- [44] de Giorgi V, Mavilia C, Massi D, Gozzini A, Aragona P, Tanini A, et al. Estrogen receptor expression in cutaneous melanoma: a real-time reverse transcriptase-polymerase chain reaction and immunohistochemical study. *Arch Dermatol* 2009;145:30–6.
- [45] Schmidt AN, Nanney LB, Boyd AS, King Jr LE, Ellis DL. Oestrogen receptor-beta expression in melanocytic lesions. *Exp Dermatol* 2006;15:971–80.
- [46] Mor G, Sapi E, Abrahams VM, Rutherford T, Song J, Hao XY, et al. Interaction of the estrogen receptors with the Fas ligand promoter in human monocytes. *J Immunol* 2003;170:114–22.
- [47] Lens M. Melanoma during pregnancy: epidemiology, diagnosis, staging, clinical picture. *Recent Results Cancer Res* 2008;178:165–74.
- [48] Brady MS, Noce NS. Pregnancy is not detrimental to the melanoma patient with clinically localized disease. *J Clin Aesthet Dermatol* 2010;3:22–8.
- [49] Akturk AS, Bilen N, Bayramgurler D, Demirsoy EO, Erdogan S, Kiran R. Dermoscopy is a suitable method for the observation of the pregnancy-related changes in melanocytic nevi. *J Eur Acad Dermatol Venereol* 2007;21:1086–90.
- [50] Driscoll MS, Grant-Kels JM. Hormones, nevi, and melanoma: an approach to the patient. *J Am Acad Dermatol* 2007;57:919–31 [quiz 32–6].
- [51] Daryanani D, Plukker JT, De Hullu JA, Kuiper H, Nap RE, Hoekstra HJ. Pregnancy and early-stage melanoma. *Cancer* 2003;97:2248–53.
- [52] Lens MB, Rosdahl I, Ahlbom A, Farahmand BY, Synnerstad I, Boeryd B, et al. Effect of pregnancy on survival in women with cutaneous malignant melanoma. *J Clin Oncol* 2004;22:4369–75.
- [53] MacKie RM, Bufalino R, Morabito A, Sutherland C, Cascinelli N. Lack of effect of pregnancy on outcome of melanoma. For The World Health Organisation Melanoma Programme. *Lancet* 1991;337:653–5.
- [54] O'Meara AT, Cress R, Xing G, Danielsen B, Smith LH. Malignant melanoma in pregnancy. A population-based evaluation. *Cancer* 2005;103:1217–26.

- [55] Reintgen DS, McCarty Jr KS, Vollmer R, Cox E, Seigler HF. Malignant melanoma and pregnancy. *Cancer* 1985;55:1340–4.
- [56] Slingluff Jr CL, Reintgen DS, Vollmer RT, Seigler HF. Malignant melanoma arising during pregnancy. A study of 100 patients. *Ann Surg* 1990;211:552–7 [discussion 8–9].
- [57] Pentheroudakis G, Orecchia R, Hoekstra HJ, Pavlidis N, Group EGW. Cancer, fertility and pregnancy: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol* 2010;21(Suppl. 5):v266–73.
- [58] Gefeller O, Hassan K, Wille L. Cutaneous malignant melanoma in women and the role of oral contraceptives. *Br J Dermatol* 1998;138:122–4.
- [59] Karagas MR, Stukel TA, Dykes J, Miglionico J, Greene MA, Carey M, et al. A pooled analysis of 10 case-control studies of melanoma and oral contraceptive use. *Br J Cancer* 2002;86:1085–92.
- [60] Persson I, Yuen J, Bergkvist L, Schairer C. Cancer incidence and mortality in women receiving estrogen and estrogen-progestin replacement therapy—long-term follow-up of a Swedish cohort. *Int J Cancer* 1996;67:327–32.
- [61] Gupta A, Driscoll MS. Do hormones influence melanoma? Facts and controversies. *Clin Dermatol* 2010;28:287–92.
- [62] Bleyer A, Viny A, Barr R. Cancer in 15- to 29-year-olds by primary site. *Oncologist* 2006;11:590–601.
- [63] Branstrom R, Ullen H, Brandberg Y. Attitudes, subjective norms and perception of behavioural control as predictors of sun-related behaviour in Swedish adults. *Prev Med* 2004;39:992–9.
- [64] Schneider S, Kramer H. Who uses sunbeds? A systematic literature review of risk groups in developed countries. *J Eur Acad Dermatol Venereol* 2010;24:639–48.
- [65] Boniol M, Autier P, Boyle P, Gandini S. Cutaneous melanoma attributable to sunbed use: systematic review and meta-analysis. *BMJ* 2012;345:e4757.
- [66] Miller SA, Hamilton SL, Wester UG, Cyr WH. An analysis of UVA emissions from sunlamps and the potential importance for melanoma. *Photochem Photobiol* 1998;68:63–70.
- [67] Colantonio S, Bracken MB, Beecker J. The association of indoor tanning and melanoma in adults: systematic review and meta-analysis. *J Am Acad Dermatol* 2014;70:847–57.
- [68] El Ghissassi F, Baan R, Straif K, Grosse Y, Secretan B, Bouvard V, et al. A review of human carcinogens—part D: radiation. *Lancet Oncol* 2009;10:751–2.
- [69] Mouret S, Forestier A, Douki T. The specificity of UVA-induced DNA damage in human melanocytes. *Photochem Photobiol Sci* 2012;11:155–62.
- [70] International Agency for Research on Cancer Working Group on artificial ultraviolet (UV) light, skin cancer. The association of use of sunbeds with cutaneous malignant melanoma and other skin cancers: A systematic review. *Int J Cancer* 2007;120:1116–22.