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## Modulation of Y<sub>356</sub> Photooxidation in *E. coli* Class Ia Ribonucleotide Reductase by Y<sub>731</sub> Across the $\alpha_2$ : $\beta_2$ Interface

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### Abstract

Substrate turnover in class Ia ribonucleotide reductase (RNR) requires reversible radical transport across two subunits over 35 Å, which occurs by a multi-step proton-coupled electron transfer mechanism. Using a photooxidant-labeled  $\beta_2$  subunit of *Escherichia coli* class Ia RNR, we demonstrate photoinitiated oxidation of a tyrosine in an  $\alpha_2$ : $\beta_2$  complex, which results in substrate turnover. Using site-directed mutations of the redox-active tyrosines at the subunit interface—Y<sub>356</sub>F( $\beta$ ) and Y<sub>731</sub>F( $\alpha$ )—this oxidation is identified to be localized on Y<sub>356</sub>. The rate of Y<sub>356</sub> oxidation depends on the presence of Y<sub>731</sub> across the interface. This observation supports the proposal that unidirectional PCET across the Y<sub>356</sub>( $\beta$ )–Y<sub>731</sub>( $\alpha$ )–Y<sub>730</sub>( $\alpha$ ) triad is crucial to radical transport in RNR.

Ribonucleotide reductases (RNRs) catalyze the conversion of nucleoside diphosphates (NDPs) to deoxynucleoside diphosphates (dNDPs) in all organisms. As such, they are largely responsible for maintaining the balanced cellular pools of DNA precursors.<sup>1</sup> While NDP reduction occurs by a highly conserved radical-based mechanism in the class I and II RNRs, the class Ia enzyme represents a unique and remarkable example of radical transport in biology.

Class Ia RNR consists of two homodimeric subunits,  $\alpha_2$  and  $\beta_2$ . Turnover requires reversible radical transport from the resting location of the radical, Y<sub>122</sub>( $\beta$ ), to the active site, C<sub>439</sub>( $\alpha$ ). These sites are separated by 35 Å, as predicted by a docking model<sup>2</sup> of both subunits and recently confirmed by pulsed electron-electron double resonance (PELDOR)<sup>3</sup> and small-angle X-ray scattering.<sup>4</sup> Any single-step radical transport mechanism over this distance is incongruent with the observed rate of turnover (2–10 s<sup>-1</sup>). Consequently, multi-step radical transfer through a pathway involving a number of highly conserved redox-active aromatic amino acids has been proposed, Y<sub>122</sub>→[W<sub>48</sub>?]→Y<sub>356</sub> in  $\beta_2$  to Y<sub>731</sub>→Y<sub>730</sub>→C<sub>439</sub> in  $\alpha_2$ , Figure 1.<sup>7,8</sup> Amino-acid radical generation requires removal of both a proton and an electron. One-electron oxidized or deprotonated intermediates are thermodynamically inaccessible under physiological conditions, implicating a proton-coupled electron transfer (PCET) mechanism for radical initiation and transport.

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### ASSOCIATED CONTENT

#### Supporting Information

Experimental methods; complete control experiments; data analysis description. This material is available free of charge via the Internet at <http://pubs.acs.org>.

In an effort to probe the mechanism of the long-range radical transport, we have developed a series of photochemical RNR constructs.<sup>9</sup> In these constructs, the  $\beta_2$  subunit is replaced by a short peptide encompassing the 20 C-terminal residues of the  $\beta_2$  protein, which includes the binding determinant for  $\alpha_2$  as well as a conserved tyrosine residue in a position analogous to Y<sub>356</sub>.<sup>10</sup> By appending a photooxidant to the N-terminus of this peptide, tyrosyl radicals can be photochemically generated, thereby enabling detailed studies of photoinitiated substrate turnover,<sup>11</sup> spectroscopic observation of photogenerated radicals,<sup>12</sup> and direct measurement of radical injection rates into  $\alpha_2$ .<sup>13</sup> In using these peptide constructs, we have found that the weak association between the peptide and the  $\alpha_2$  subunit ( $K_D \sim 10 \mu\text{M}$ ), and the conformational flexibility of the bound peptide,<sup>14</sup> inhibit radical injection and complicate the examination of PCET kinetics.

To address these challenges and to develop a faithful construct of the natural system, we have recently developed a full-length photochemical  $\beta_2$  subunit.<sup>15</sup> In this construct, a bromomethylpyridyl rhenium(I) tricarbonyl phenanthroline complex selectively modifies a single surface-accessible cysteine variant at position 355 of *E. coli* class Ia  $\beta_2$  to yield a labeled subunit ([Re]- $\beta_2$ ). Upon excitation of the [Re] complex, a transient tyrosyl radical forms and can be spectroscopically detected.<sup>15</sup> We now report the photochemical generation of  $\bullet\text{Y}_{356}$  in an intact  $\alpha_2:\beta_2$  complex, which is competent for photoinitiated substrate turnover. In order to decipher the nature of tyrosine oxidation processes by [Re] at the subunit interface, the Y<sub>356</sub>F variant ([Re]-Y<sub>356</sub>F- $\beta_2$ ) was prepared. By performing a comparative transient kinetics study of  $\alpha_2:[\text{Re}]-\text{Y}_{356}\text{F}(\beta_2)$  and [Re]- $\beta_2:\text{Y}_{731}\text{F}(\alpha_2)$  constructs, a photooxidation process to furnish radicals is ascribed to interfacial tyrosine radical transport.

The [Re] group does not preclude binding of [Re]- $\beta_2$  to  $\alpha_2$ . A competitive inhibition assay (Figure S1)<sup>10</sup> reveals that the observed dissociation constant ( $K_D([\text{Re}]-\beta_2) = 0.71(8) \mu\text{M}$ ) is comparable to that observed for aminotyrosine (NH<sub>2</sub>Y) substituted  $\beta_2$ .<sup>16</sup> With binding established, RNR activity assays were performed on the unlabeled S<sub>355</sub>C- $\beta_2$  and labeled [Re]- $\beta_2$  to ascertain the effect of labeling on activity. The 1,100 U/mg (Figure S2) activity of unlabeled S<sub>355</sub>C- $\beta_2$  is less than  $\beta_2$  (7,600 U/mg),<sup>17</sup> and decreases further upon labeling to 122 U/mg. The low observed activity of [Re]- $\beta_2$ , despite only a modest reduction in subunit binding affinity, suggests a subtle as opposed to a large perturbation to the protein structure. To avoid complications from steady-state activity, [Re]- $\beta_2$  was treated with hydroxyurea (HU) to reduce the native radical cofactor ( $\bullet\text{Y}_{122}$ ), yielding met-[Re]- $\beta_2$ . This form was used for all experiments except the above activity assays and is subsequently referred to as [Re]- $\beta_2$  for convenience.

The efficacy of photooxidant labeling is revealed by activity measurements of [Re]- $\beta_2$  under illumination ( $\lambda > 325 \text{ nm}$ ) and in the presence pre-reduced wt- $\alpha_2$ , where wt- $\alpha_2$  are reduced, but there is no additional reductant available. Figure 2 shows that dCDP product is formed at 0.47(3) dCDP/ $\alpha_2$  (pH 7.6) or 0.11(2) dCDP/ $\alpha_2$  (pH 8.3) under photochemical conditions. The identity of the product was confirmed by an HPLC assay (Figure S3). This product yield is less than that observed previously for photoRNRs consisting of a peptide surrogate for  $\beta_2$  in the presence of wt- $\alpha_2$  ( $\sim 0.2$  dCDP/ $\alpha_2$ ).<sup>12</sup> However, we note that in the peptide surrogate, position 356 on the peptide was occupied by 3,5-difluorotyrosine (3,5-F<sub>2</sub>Y), as opposed to tyrosine. Greater radical production yield at pH 8.3 is expected for 3,5-F<sub>2</sub>Y because the amino acid is deprotonated, and tyrosine is not. Hence, 3,5-F<sub>2</sub>Y is more rapidly oxidized than tyrosine, despite having similar reduction potentials at this pH. Consistent with previous peptide-based systems,<sup>12</sup> the phototurnover yield of the  $\alpha_2:[\text{Re}]-\beta_2$  construct is significantly greater and time-dependent at pH 7.6 (Figure S4).

Having established that [Re]- $\beta_2$  is photochemically active for substrate turnover, we sought to generate a radical within the  $\alpha_2$ : [Re]- $\beta_2$  complex. Photoexcitation ( $\lambda = 355$  nm) of [Re]- $\beta_2$  or [Re]-Y<sub>356</sub>F- $\beta_2$  yields transient absorption features (Figure S5) consistent with the <sup>3</sup>MLCT excited state ([Re<sup>I</sup>]\*),<sup>15</sup> but no signatures corresponding to tyrosyl radical are observed. Scheme 1 shows a photophysical model for the system. In the absence of tyrosine (left), the observed lifetime,  $\tau_o$  is defined by nonradiative ( $k_{NR}$ ) and radiative ( $k_R$ ) decay rate constants as follows,

$$1/\tau_o = k_R + k_{NR}. \quad (1)$$

For the kinetics associated with the  $\beta_2$  subunit,  $\tau_o$  is given by [Re]-Y<sub>356</sub>F- $\beta_2$ . For rate constant calculations, measurement of  $\tau_o$  differs from measurement of the observed lifetime owing to the presence of Y<sub>356</sub>, as indicated in Table 1. The presence of Y<sub>356</sub> in  $\beta_2$  adds an additional decay pathway via charge separation ( $k_{CS}$ ), which may be determined from the observed lifetime ( $\tau_{obs}$ ),

$$k_{CS} = 1/\tau_{obs} - 1/\tau_o. \quad (2)$$

where  $\tau_{obs}$  is inversely related to the observed rate constant ( $k_{obs}$ ) (i.e.,  $k_{obs} = 1/\tau_{obs}$ ). The lifetime of [Re<sup>I</sup>]\* in [Re]- $\beta_2$  is significantly shorter than that for the construct with Y<sub>356</sub> replaced by F, Table 1. We ascribe the shortened lifetime of [Re<sup>I</sup>]\* in [Re]- $\beta_2$  to quenching of [Re<sup>I</sup>]\* by Y<sub>356</sub> (Scheme 1). These data suggest that the inability to detect tyrosine radical signal is due to rapid charge recombination of the [Re<sup>0</sup>]-•Y charge-separated state ( $k_{CR} > k_{CS}$ ). Using the lifetime of [Re<sup>I</sup>]\* in [Re]-Y<sub>356</sub>F- $\beta_2$  to provide  $\tau_o$ , the rate constant for charge separation,  $k_{CS}$ , is calculated from eq. (2) to be  $k_{CS} = 8.37(66) \times 10^5$  s<sup>-1</sup>. Previously reported charge recombination rate constants for rhenium polypyridine tyrosine systems exceed  $10^7$  s<sup>-1</sup>.<sup>18</sup> Accordingly,  $k_{CR}$  appears to be much greater than  $k_{CS}$ , thus accounting for our inability to observe tyrosyl radical in the transient absorption spectrum.

In the presence of wt- $\alpha_2$ , the lifetime of [Re<sup>I</sup>]\* is also quenched within the intact  $\alpha_2$ : [Re]- $\beta_2$  complex. Under these conditions, 96% of [Re]- $\beta_2$  is bound to  $\alpha_2$ . In this  $\alpha_2$ : [Re]- $\beta_2$  complex, the lifetime of [Re<sup>I</sup>]\* increases for both [Re]- $\beta_2$  and [Re]-Y<sub>356</sub>F- $\beta_2$  (Table 1, Figure S6) because the [Re] center resides in the hydrophobic environment of the protein.<sup>14</sup> Notwithstanding, the lifetime of  $\alpha_2$ : [Re]- $\beta_2$  is shorter than that of the [Re]-Y<sub>356</sub>F- $\beta_2$  complex and a charge separation rate constant—again assigned to Y<sub>356</sub> oxidation—is calculated to be  $k_{CS} = 4.05(10) \times 10^5$  s<sup>-1</sup>.

The charge separation reaction is affected by perturbations of the radical transport pathway in  $\alpha_2$ . When Y<sub>731</sub>( $\alpha$ ) is replaced by phenylalanine, the rate constant for Y<sub>356</sub> oxidation decreases to  $3.28(11) \times 10^5$  s<sup>-1</sup>. This difference corresponds to a 23% enhancement in Y<sub>356</sub>( $\beta$ ) oxidation when Y<sub>731</sub>( $\alpha$ ) is present. At pH 8.3, a similar phenomenon is observed (Table S1, Figure S7), with faster charge separation and a rate enhancement due to presence of Y<sub>731</sub>( $\alpha$ ) of 14%. Studies of this process at pH 8.3 were performed to maximize tyrosine oxidation at higher pH; studies at pH 7.6 correspond to the regime where greater phototurnover is observed.

This decrease in the rate of Y<sub>356</sub> oxidation in the presence of F<sub>731</sub> provides evidence for direct interaction between Y<sub>356</sub>( $\beta$ ) and Y<sub>731</sub>( $\alpha$ ) in the assembled interface. This result is consistent with our previous work showing that oxidation of  $\alpha_2$  by a photogenerated 2,3,6-trifluorotyrosine radical requires both Y<sub>731</sub> and Y<sub>730</sub>.<sup>13</sup> This observation led to the proposal that the Y<sub>731</sub>( $\alpha$ )-Y<sub>730</sub>( $\alpha$ ) dyad is an important element for radical transport. The current observation that Y<sub>356</sub> photooxidation is modulated by Y<sub>731</sub> suggests a significant interaction between Y<sub>356</sub> and Y<sub>731</sub>, across the  $\alpha_2$ : $\beta_2$  interface. The present work therefore suggests

further elaboration to our previous model of a coupled tyrosine dyad to a triad—Y<sub>356</sub>(β)–Y<sub>731</sub>(α)–Y<sub>730</sub>(α)—which is responsible for mediating intersubunit radical transport as well as radical transport through α<sub>2</sub> to C<sub>439</sub>(α) at the active site. This result is in line with the recent observation that a new radical, generated by using a highly oxidizing •NO<sub>2</sub>Y<sub>122</sub>(β), equilibrates over Y<sub>356</sub>, Y<sub>730</sub>, and Y<sub>731</sub>.<sup>19</sup>

Given the mechanistic hypothesis that unidirectional PCET operates in the α<sub>2</sub> subunit,<sup>7</sup> even a slight perturbation of the Y<sub>356</sub>(β)–Y<sub>731</sub>(α)–Y<sub>730</sub>(α) triad is likely to have an effect on radical transport. Several related studies have provided insight into the nature of interactions between adjacent pathway tyrosine residues. Hydrogen-bonding interactions involving the tyrosine phenol groups of these residues are a crucial structural element. Such a hydrogen-bonded network is consistent with our observations that removal of one hydroxyl group disrupts the oxidation of an adjacent residue. In support of this model, rapid mixing studies of wt-β<sub>2</sub> and NH<sub>2</sub>Y<sub>730</sub>-α<sub>2</sub> (in the presence of CDP and ATP) results in generation of •NH<sub>2</sub>Y<sub>730</sub>(α) and concomitant loss of •Y<sub>122</sub>(β).<sup>20</sup> Examination of this trapped •NH<sub>2</sub>Y<sub>730</sub> radical by electron-nuclear double resonance and density functional theory suggests a detailed model for the hydrogen-bonding network in α<sub>2</sub> including •NH<sub>2</sub>Y<sub>730</sub>, C<sub>439</sub>, Y<sub>731</sub> and a key water molecule.<sup>21</sup>

The importance of hydrogen bonding on the PCET oxidation of phenols has been well established in studies of small molecule models,<sup>22–29</sup> as well in natural systems.<sup>30</sup> Hydrogen-bonding is critical to the reduction of the redox-active tyrosine Y<sub>Z</sub> in photosystem II, and plays a key role in the different oxidation states of the oxygen evolving complex.

Conformational changes may also occur upon substitution of nearby amino acids of the tyrosine triad. An effect of this type has been previously revealed by the NH<sub>2</sub>Y<sub>730</sub>-α<sub>2</sub> crystal structure; the observed electron density suggests a second conformation for Y<sub>731</sub>.<sup>17</sup> In the case of an F to Y mutation, a change in conformation may well result from the loss of a hydrogen bond to the substituted residue. A change in the conformation of a tyrosine residue could significantly alter the distance between residues as well as the local environment of the redox-active phenol group.

With the knowledge that [Re]-β<sub>2</sub> is capable of photochemical Y<sub>356</sub> oxidation to generate an on-pathway radical, we sought to circumvent the limitation imposed by  $k_{CR} > k_{CS}$  by employing the flash-quench methodology.<sup>31</sup> When [Re]-β<sub>2</sub> is in the presence of Ru<sup>III</sup>(NH<sub>3</sub>)<sub>6</sub>Cl<sub>3</sub>, [Re<sup>I</sup>]\* is rapidly quenched to reveal signals for a tyrosyl radical. As previously observed for the rhenium-bipyridine photopeptide system, increased concentrations of quencher result in larger observed signals but also in diminished •Y lifetimes, Figure S8.<sup>13</sup> In the presence of the α<sub>2</sub> subunit, less efficient quenching by Ru<sup>III</sup> is observed, requiring a concentration of 10 mM to obtain adequate radical signals.

The addition of 10 mM Ru<sup>III</sup>(NH<sub>3</sub>)<sub>6</sub>Cl<sub>3</sub> quenches [Re<sup>I</sup>]\*; in the absence of α<sub>2</sub>, the [Re]-β<sub>2</sub> lifetime is 65 ns. Addition of wt-α<sub>2</sub> to the solution results in a longer [Re<sup>I</sup>]\* lifetime (137 ns) owing to the decreased solution exposure of the [Re] complex and an attendant decrease in bimolecular quenching by Ru<sup>III</sup>. The sensitivity of the [Re<sup>I</sup>]\* lifetime to α<sub>2</sub> provided a method for determining  $K_D$  for [Re]-β<sub>2</sub> under flash quench conditions in lieu of the usual coupled assay, which cannot be performed in the presence of Ru<sup>III</sup>(NH<sub>3</sub>)<sub>6</sub>Cl<sub>3</sub> (Figure S9). Lifetime analysis of the binding shows that the presence of Ru<sup>III</sup>(NH<sub>3</sub>)<sub>6</sub>Cl<sub>3</sub> does not significantly perturb binding of [Re]-β<sub>2</sub> ( $K_D$ ([Re]-β<sub>2</sub>) = 0.68(21) μM), (Figure S10).

One potential issue when performing experiments at high concentrations of protein (up to 50 μM [Re]-β<sub>2</sub> and 75 μM α<sub>2</sub>), is the formation of assemblies larger than α<sub>2</sub>:β<sub>2</sub>, as previously observed for the inhibited form of *E. coli* Class Ia RNR (formed in the presence of dATP).<sup>4</sup>

Under different conditions, such oligomers may not retain an intact subunit interface, precluding the generation of on-pathway radical or its subsequent transport. With 10  $\mu\text{M}$  of each subunit, even in the absence of dATP, species larger than  $\alpha_2\beta_2$  are observed by analytical ultracentrifugation.<sup>4</sup> Nevertheless, PELDOR experiments performed at high concentration (100  $\mu\text{M}$  each of  $\alpha_2\beta_2$ ), including those previously discussed,<sup>19</sup> show on-pathway radicals are generated under these conditions, confirming that radical transport is intact at high concentrations.

In the presence of both wt- $\alpha_2$  and  $\text{Ru}^{\text{III}}(\text{NH}_3)_6\text{Cl}_3$ , photoexcitation of  $[\text{Re}]-\beta_2$  results in a typical  $\bullet\text{Y}$  absorption feature at 410 nm (Figure 3). Under these experimental conditions, 97% of  $[\text{Re}]-\beta_2$  in solution is bound to  $\alpha_2$ . This observation confirms that  $[\text{Re}]$  is capable of generating  $\bullet\text{Y}$  in an assembled  $\alpha_2\beta_2$  complex. As in the unquenched experiments above,  $[\text{Re}]-\text{Y}_{356}\text{F}-\beta_2$  and  $\text{Y}_{731}\text{F}-\alpha_2$  were employed in combination with  $[\text{Re}]-\beta_2$  and wt- $\alpha_2$  to determine the identity of the photogenerated radical. The presence of visible signals for  $\bullet\text{Y}$  in the absence of  $\text{Y}_{356}$  (Figure 3) indicates that the observed signal is not entirely due to  $\bullet\text{Y}_{356}$ . In fact, signals due to  $\bullet\text{Y}$  radical persist even in the case where neither interface tyrosine is present, (Figure S11) suggesting that oxidation of an off-pathway tyrosine is also occurring.

Despite these side-reactions under flash-quench conditions, the relative amplitudes of  $\bullet\text{Y}$  indicate that a significant fraction of the observed signal is due to  $\bullet\text{Y}_{356}$ . In all cases where  $\text{Y}_{356}$  is present, the observed amplitude of  $\bullet\text{Y}$  is greater than the amplitudes observed in any case where  $\text{Y}_{356}$  is absent (Figure S12). On the basis of single-wavelength kinetics (Figure S11), ~44% of the signal is due to  $\bullet\text{Y}_{356}$ . This amplitude corresponds to an estimated quantum yield of 1.2% for overall  $\bullet\text{Y}$  formation and 0.5% for  $\text{Y}_{356}$ , calculated as previously reported.<sup>15</sup> The presence of the off-pathway radical precludes direct study of radical transport kinetics within the triad.

In this work, we have observed rates of tyrosine oxidation in the range of  $10^5$ – $10^6$   $\text{s}^{-1}$ , consistent with PCET oxidation of protonated tyrosine. Future experiments will target the site-specific installation of a  $\text{F}_n\text{Y}$  at  $\text{Y}_{356}(\beta)$  with established methods<sup>32</sup> in an effort to out-compete off-pathway oxidations by  $[\text{Re}]$ , improve the yield of photogenerated radical and to provide a spectroscopically distinguishable radical signal. The increased acidity of fluorotyrosines allows for their deprotonation, enabling oxidation by electron transfer with rate constants of  $10^7$ – $10^9$   $\text{s}^{-1}$ .<sup>18</sup> Such rapid charge separation will enable observation of  $\bullet\text{F}_n\text{Y}$  without the necessity of implementing the flash-quench technique. These observed side reactions are expected to play a less significant role in the experiments directly involving  $[\text{Re}^{\text{I}}]^*$  owing to the decreased lifetime and reduction potential of  $[\text{Re}^{\text{I}}]^*$  as compared to flash-quench generated  $[\text{Re}^{\text{II}}]$ .

The photogeneration of  $\bullet\text{Y}_{356}$  within an assembled  $\alpha_2\beta_2$  complex represents a key advance in the study of PCET in RNRs. The observation of a radical along the  $\alpha_2$  pathway is enabled with the advantages of added fidelity to the natural system and improved subunit binding. This result also opens the door for the study of photoinitiated PCET along the radical transport pathway in the  $\beta_2$  subunit in the fully assembled protein complex, a longstanding goal of our studies in RNR.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.



## Acknowledgments

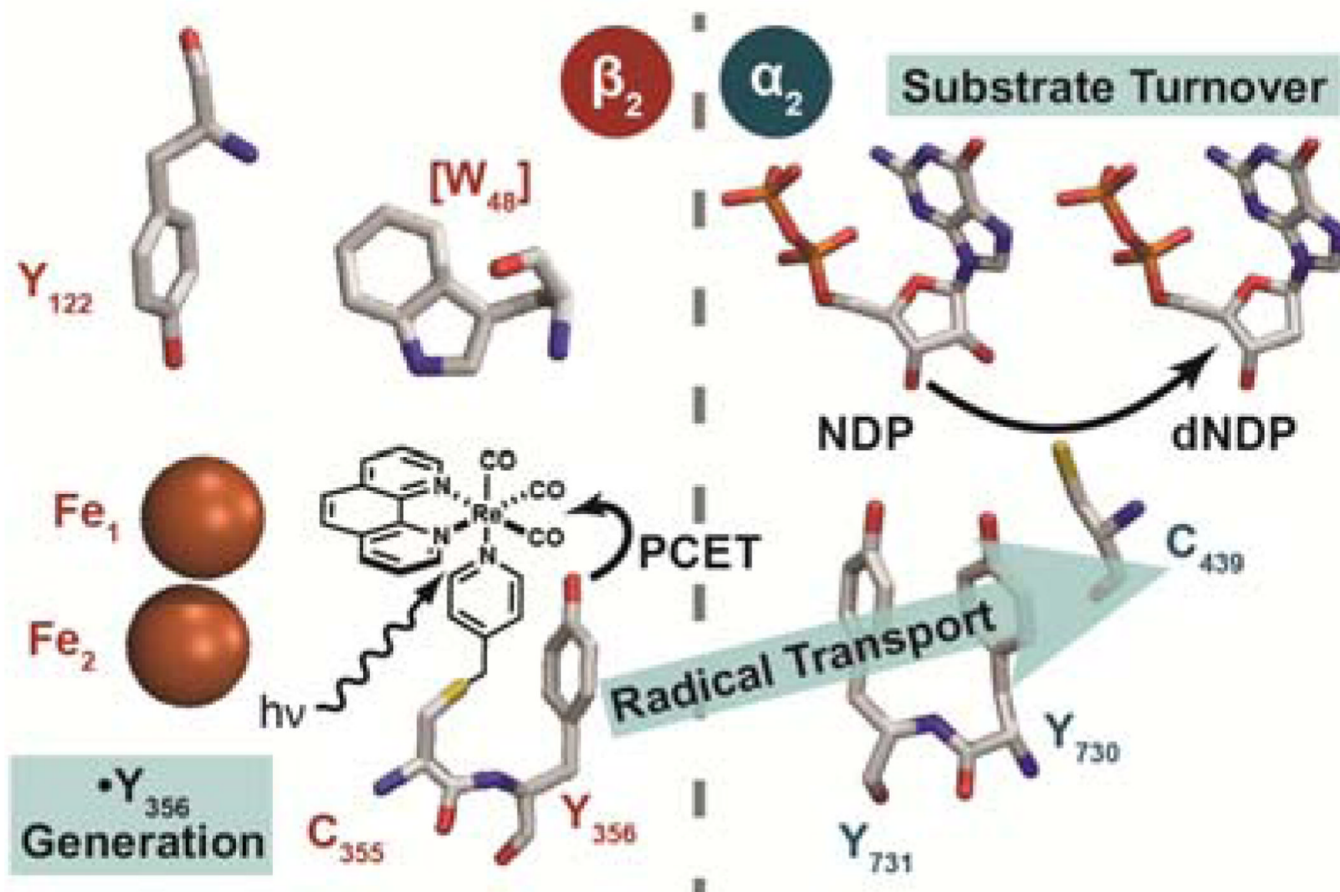
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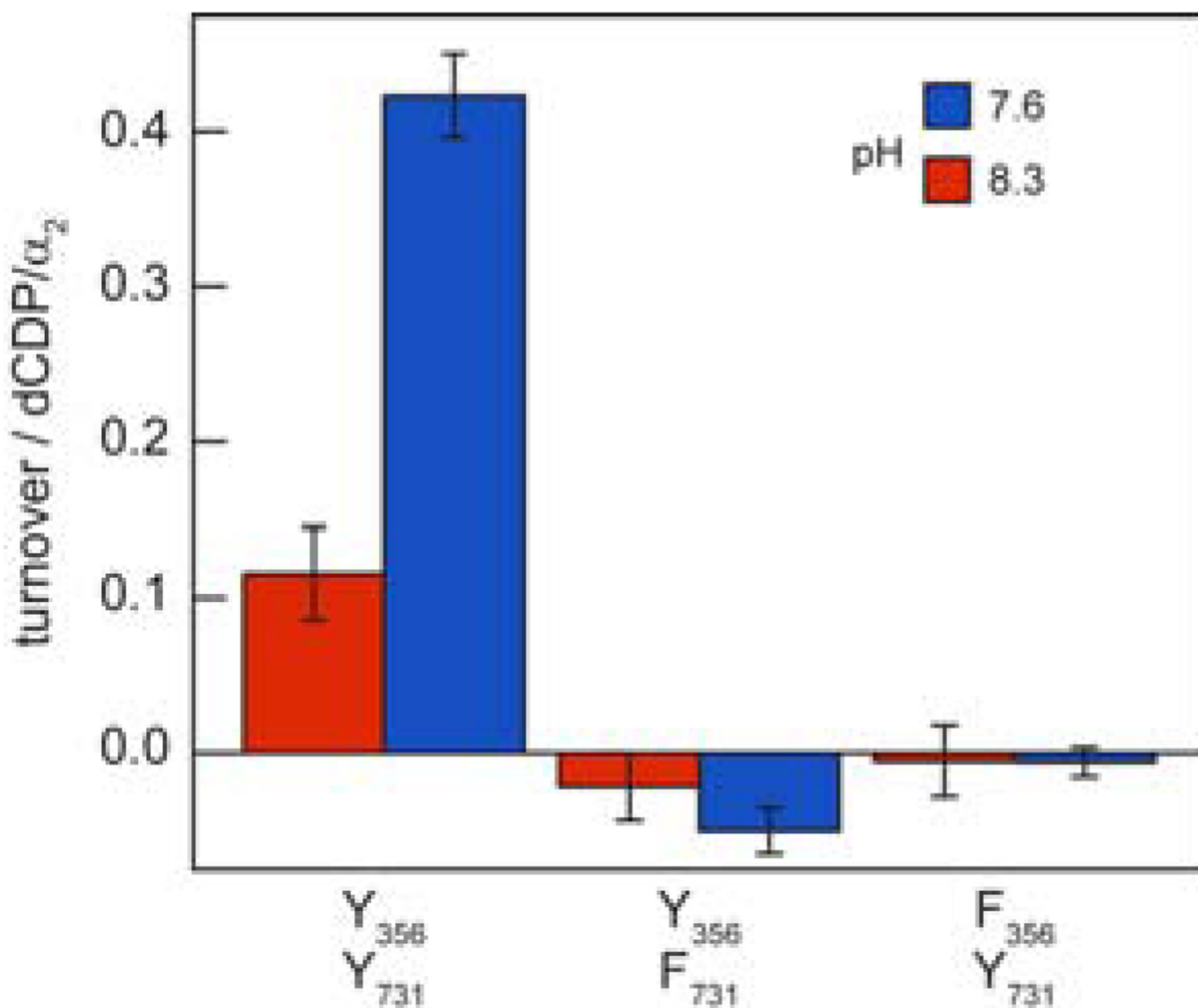
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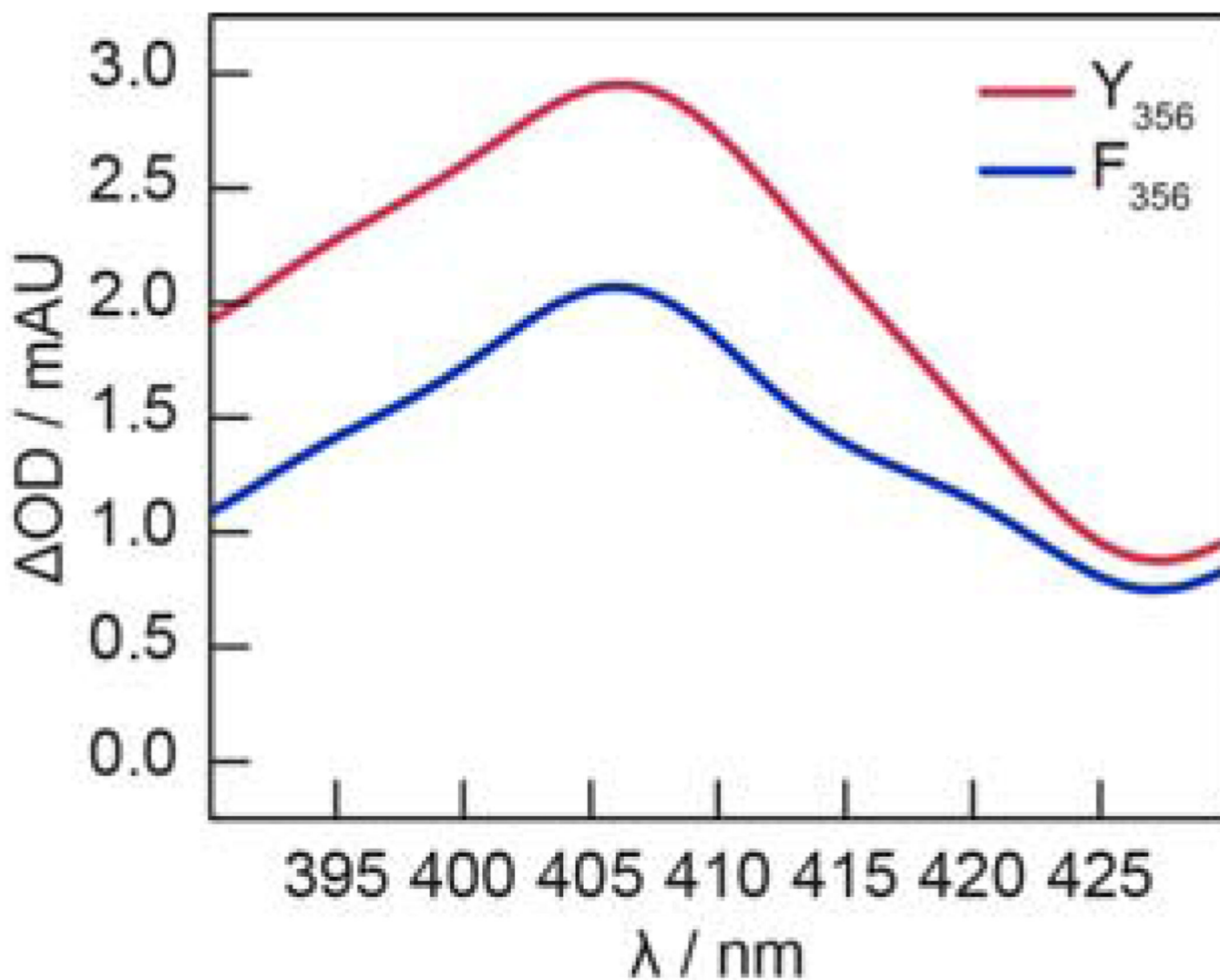


**Figure 1.** An intact, photochemical RNR where a radical is generated directly at  $Y_{356}(\beta)$  and is poised to follow the proposed radical transport pathway through  $\alpha_2$  to the active site. Figure prepared from PDB codes 4R1R<sup>5</sup> ( $\alpha_2$ ) and 1MXR<sup>6</sup> ( $\beta_2$ ).

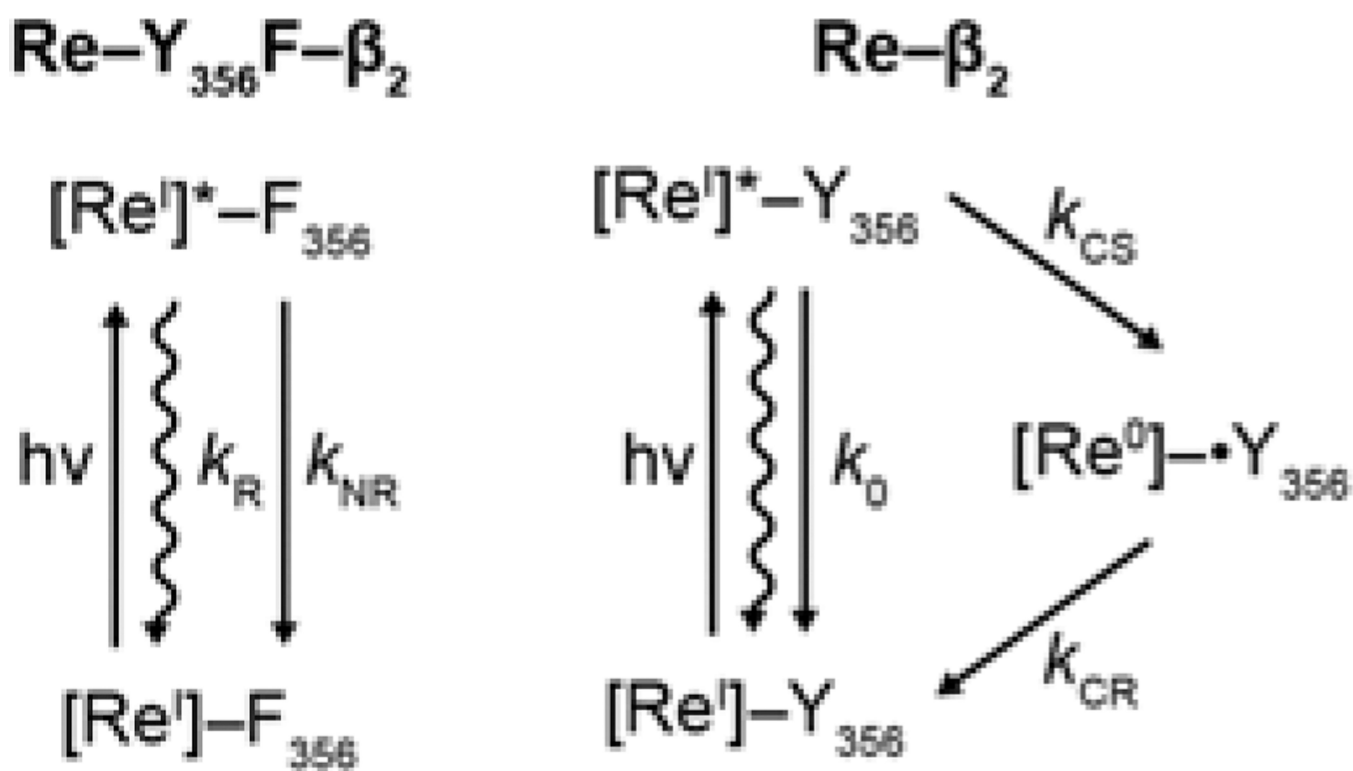


**Figure 2.**

Photochemical substrate turnover in the  $\alpha_2$ :[Re]- $\beta_2$  complex formed from 50  $\mu$ M met-[Re]- $\beta_2$  or met-[Re]-Y<sub>356</sub>F- $\beta_2$  and 20  $\mu$ M pre-reduced wt- $\alpha_2$  or Y<sub>731</sub>F- $\alpha_2$  (as indicated) at pH 8.3 (50 mM borate, 15 mM MgSO<sub>4</sub>, 0.2 mM [<sup>3</sup>H]-CDP (10750 cpm/nmol), 3 mM ATP, 5% glycerol) or pH 7.6 (50 mM HEPES, 15 mM MgSO<sub>4</sub>, 0.2 mM [<sup>3</sup>H]-CDP (22713 cpm/nmol), 3 mM ATP),  $\lambda_e > 325$  nm.



**Figure 3.** Spectral observation of a photogenerated,  $Y_{356}$ -centered tyrosine radical in an  $\alpha_2:[Re]-\beta_2$  complex. Transient absorption spectra measured 5  $\mu s$  after excitation on samples of 50  $\mu M$  met-[Re]- $\beta_2$  or met-[Re]- $Y_{356}F-\beta_2$  (as indicated), 75  $\mu M$  wt- $\alpha_2$ , and 10 mM  $Ru^{III}(NH_3)_6Cl_3$  in 50 mM borate, 15 mM  $MgSO_4$ , 1 mM CDP, 3 mM ATP, 5% glycerol, pH 8.3,  $\lambda_{exc} = 355$  nm.



**Scheme 1.**  
Excited-State Deactivation Pathways for  $[\text{Re}^{\text{I}}]^*$

**Table 1**Tyrosine Dependent Excited-State Quenching at the  $\alpha_2:\beta_2$  Interface (pH = 7.6)

$\beta_{356}$	$\alpha_{731}^a$	$\tau / \text{ns}^b$	$k_{\text{CS}} / 10^5 \text{ s}^{-1} c$
Y	–	403(31) (= $\tau_{\text{obs}}$ )	8.37(66)
F	–	607(7) (= $\tau_o$ )	
Y	Y	543(13) (= $\tau_{\text{obs}}$ )	4.05(10)
F	Y	696(5) (= $\tau_o$ )	
Y	F	590(17) (= $\tau_{\text{obs}}$ )	3.28(11)
F	F	732(11) (= $\tau_o$ )	

<sup>a</sup>Emission lifetimes measured on samples of 10  $\mu\text{M}$  met-[Re]- $\beta_2$  or met-[Re]-Y356F- $\beta_2$  and 25  $\mu\text{M}$  wt- $\alpha_2$  or Y731F- $\alpha_2$  (as indicated) in 50 mM HEPES, 15 mM  $\text{MgSO}_4$ , 1 mM CDP, 3 mM ATP, pH 7.6,  $\lambda_{\text{exc}} = 355 \text{ nm}$ ,  $\lambda_{\text{det}} = 600 \text{ nm}$ . Representative decay traces are shown in Figure S6.

<sup>b</sup>Errors shown in parentheses represent  $2\sigma$  from triplicate measurements on independently prepared samples.

<sup>c</sup> $k_{\text{CS}}$  is determined from eq. (2). Errors shown in parentheses represent  $2\sigma$  propagated error of corresponding lifetimes.