

Evaluation of removal efficiency of phenol from synthetic aqueous solutions by *Citrullus colocynthis* seed ash

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Abstract

Background: Phenol is a prevalent pollutant found in many industrial wastewaters, and it is paid singular attention because of its special features like high toxicity, carcinogenic properties, and vital gathering ability that affects the health of humans and the environment. One of the most important technologies for the removal of phenol is the use of adsorbents. The current study investigated the removal of phenol from synthetic aqueous solutions using *Citrullus colocynthis* seed ash.

Methods: This study is experimental and was conducted on a pilot scale. The efficiency of phenol removal by *C. colocynthis* seed ash was evaluated in a batch system, and different parameters such as initial concentration of phenol (10, 20, 50, and 80 mg/L), contact time (2, 5, 10, and 30 minutes), pH (2–12), adsorbent dose (0.5, 1, 3, 5, and 10 g/L), and temperature were studied. Excel software was used for data analysis. The adsorption process was modeled with Freundlich and Langmuir isotherms at controlled temperatures.

Results: The results showed that the highest removal rate of phenol was obtained at a pH of 2 (83.4%), initial phenol concentration of 20 ppm (66.4%), adsorbent dose of 5 g/L (86.8%), and contact time of 10 minutes. The evaluation of correlation coefficients showed that the phenol adsorbed by *C. colocynthis* seed ash was in greater accordance with the Freundlich model than the Langmuir model.

Conclusion: In general, the results of this study revealed that *C. colocynthis* seed ash has suitable potential for use in removing phenol from aqueous solutions on operation and practical scales due to its low cost and easy access.

Keywords: Phenol, Adsorption, *Citrullus colocynthis*

Citation: Salari S, Afsharnia M, Moteallemi A, Ghasemi M. Evaluation of removal efficiency of phenol from synthetic aqueous solutions by *Citrullus colocynthis* seed ash. Environmental Health Engineering and Management Journal 2018; 5(1): 49–55. doi: 10.15171/EHEM.2018.07.

Article History:

Received: 11 September 2017

Accepted: 20 January 2018

ePublished: 29 January 2018

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Introduction

Phenol is one of the organic water pollutants with the chemical structure C_6H_5OH that occur because of industrialization (Figure 1) (1). This compound has very high solubility in water and many other organic solvents, and after dissolving in water it creates very little acidity (2). Naturally, phenol is produced from coal tar or gasoline distillation; artificially, it is created through heating sodium benzene sulfate with hydrous soda under high pressure (3). This matter and its derivatives are used in various industries such as resin and plastics, colors, pesticides, pharmaceuticals, oil refineries, and the steel, aluminum, lead, washes, synthetic textiles, and leather industries (4). Phenol is a major pollutant of industrial wastewater. Its compounds produce complexes with tangible tastes and odors in the presence of chlorine (5). The symptoms of acute phenol poisoning include xerostomia, darkening

of the color of urine, and the destruction of organs and muscles. Long-term exposure to phenol vapors can cause anorexia, weakness, headache, muscle pain, and jaundice (6,7). Due to the high absorption of phenol through the skin of the hand or forehead (60%–90%), in high concentrations, it can lead to death (8,9). Diagnosis, determination, and measurement of phenol compounds in water resources and monitoring their environmental effects play key roles in the control and impact reduction of these combinations (10). According to the standard guidelines of Iran, the maximum allowed concentration of phenol compounds in superficial water, agriculture, and irrigation is 1 mg/L (11). The World Health Organization (WHO) considers the maximum allowable concentration of phenol in drinking water to be 0.002 mg/L (12). Also, according to the United States Environmental Protection Agency (USEPA), the permissible concentrations of



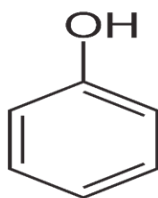


Figure 1. Chemical structure of phenol.

phenol in drinking water and industry effluent are less than 1 mg/L and 500 µg/L, respectively (13). Proper treatment based on existing standards must be carried out so as to reduce the harmful environmental effects of phenol combinations. To date, different methods have been used to treat phenol compounds, such as chemical oxidation, photochemical destruction, precipitation, ion exchange, wet oxidation, electrochemical methods, radiation, membranous filtration, enzymatic filtration, adsorption, and photocatalytic destruction (3,14). Most of these methods have disadvantages like high treatment costs, needs for additive treatment, production of hazardous by-products, high energy consumption, and low efficiency. Among different treatment methods, the adsorption process is one of the simplest and most effective methods for removing poisonous pollutants from aqueous environments (15). In the last two decades, many studies have been carried out on the adsorption of organic compounds with low degradability. In these studies, researchers used produced wastes such as activated carbon (15), coal (16), sugar cane ash (17), agricultural waste (18), and rice bran ash (19) as potential adsorbents. In one study, Bazrafshan et al investigated the efficiency of *Moringa peregrina* tree shell ash as a natural adsorbent for the removal of phenol from aqueous solutions. The effect of important variables such as adsorbent dose, primary phenol concentration, and solution pH were evaluated. The highest values of removal were obtained at a pH of 6, initial concentration of 100 mg/L, and adsorbent dose of 0.4 g/L. Data experiments were better represented by the Langmuir isotherm than the Freundlich model (20). *Citrullus colocynthis* is a plant native to southern Iran and Gonabad city in Razavi Khorasan province. This plant has a bitter taste and is most commonly used in Iranian traditional medicine for the treatment of diabetes and the reduction of blood glucose levels. Activating the plant is very easy, and, until now, no comprehensive study has been conducted using *Citrullus colocynthis* as adsorbent. Thus, the present study applied the seed ash of *C. colocynthis* as an affordable and available adsorbent for the removal of phenol from synthetic aqueous solutions.

Methods

Preparation of adsorbent

Citrullus colocynthis seeds were collected from the vicinity of Gonabad city, Razavi Khorasan province, Iran. The seeds were separated and washed with distilled water to

remove impurities like soluble, colored compounds and sand. Then they were dried in a drying oven (80°C) and electric furnaces (550°C) for 4 hours. The ash of the seeds was extracted and sieved through standard sieves of 20, 30, 40, 50, and 100 mesh to obtain particle size (21). The best results were obtained using 20 mesh.

XRD (X-ray Diffraction, D6792-PHILIPS) was used to determine the adsorbent crystalline phases (Figure 2).

Batch adsorption experiments

Synthetic solution was made by a stock solution containing phenol. Experimental solutions containing the desired concentrations of phenol were prepared by diluting the stock solution (50 mL). Then, a known mass of *C. colocynthis* seed ash (adsorbent dose) was added to the solution. Batch adsorption experiments were performed in a mechanical shaker to agitate the solutions. Finally, using Whatman grade 42 filter paper, the solutions were filtered and the phenol concentrations were analyzed (22). The amount of phenol adsorbed was calculated using the following formula:

$$q_e = \frac{(C_0 - C_e)V}{M}$$

where C_0 is the initial phenol concentration and C_e is the secondary concentration after reaching the balance, V is the volume of the solution (L), and M is the used amount of adsorbent (g).

Method of measuring phenol concentration

The residual phenol concentration was measured using spectrophotometry at a wavelength of 500 nm in accordance with the standard guidelines for water and wastewater (23).

Effect of initial phenol concentration, pH, adsorbent dose, and contact time

In this study, the effects of different variables such as pH, adsorbent dosage, and initial phenol concentration were investigated (10,16,19,22). In the first stage, the initial pH of samples in the range of 2-12 was examined. In this stage, the adsorbent dose, initial phenol concentration, and contact time were 3 g/L, 50 mg/L, and 60 minutes,

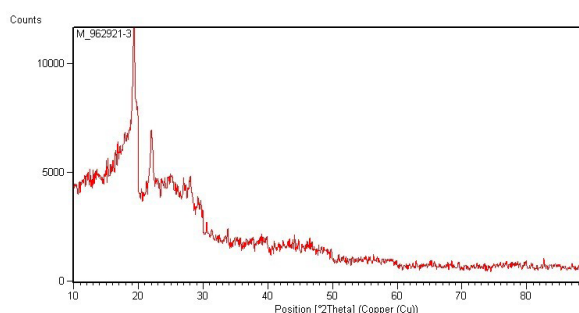


Figure 2. Characteristics of *Citrullus colocynthis* seed: XRD pattern.

respectively. After determining the optimal pH, the effects of adsorbent dose at concentrations of 0.5, 1, 3, 5, and 10 mg/L, phenol concentration of 50 mg/L, and contact time of 60 minutes were tested. Then, the effects of initial phenol concentrations (10, 20, 50, 80, and 100 mg/L) at different times (2, 5, 10, 30, and 60 minutes) were examined. The rate of phenol elimination was obtained as a percentage:

$$\% = \frac{C_0 - C_f}{C_0} \cdot 100$$

where C_0 (mg/L) is the initial concentration of phenol and C_f (mg/L) is the final concentration of phenol (after adsorption). All experimental tests were done based on the primary pilot, and all tests were repeated 2 times; the number of analyzed samples was 200.

Adsorption isotherms

Adsorption isotherms describe the interactive behavior between an adsorbent and the adsorbent. Isotherms are important to optimizing the use of adsorbents and showing the relationship between the phenol concentration of the soluble and the phenol absorption rate by the solid phase, when the two phases are in balance. For this, the Langmuir and Freundlich adsorption models were used to analyze the experimental equilibrium data for the adsorption of phenol by *C. colocynthis* seed ash. To evaluate data significant to the Langmuir and Freundlich isotherms, the regression coefficient (R²) was used.

The Langmuir isotherm

The Langmuir isotherm is valid for a homogeneous system in which all sorption sites are identical. It is determined using the following formula (24):

$$\frac{C_e}{q_e} = \frac{1}{Q_m \cdot b} + \frac{1}{Q_m} C_e$$

where C_e is the equilibrium concentration of the solution (mg/L), q_e is the value of adsorbate adsorbed per unit mass of adsorbent (mg/g), b (L/mg) is the Langmuir adsorption constant, and Q_m (mg/g) is the maximum amount adsorbed.

The Freundlich isotherm

This isotherm describes reversible adsorption in a heterogeneous system (24) and is calculated by the following formula:

$$\log q_e = \log K_f + \frac{1}{n} \log C_e$$

where K_f and n are Freundlich constants, $K_f((\text{mg/g})(\text{L/mg})^{1/n})$ is the adsorption capacity of the adsorbent, and n shows how favorable the adsorption process is. The values of $1/n < 1$ indicate a normal Friedrich isotherm and cooperative adsorption.

Results

Chemical constituents of *Citrullus colocynthis*

The presence of the following compounds has been proven in *C. colocynthis* seeds by phytochemical analysis: proteins = $13.99 \pm 0.06\%$, crude fibers = $46.73 \pm 0.15\%$, moisture = $6.43 \pm 0.15\%$, α -tocopherol = 1.90 ± 0.020 g/100 g, and fixed oil = 17%-28.5% with a high proportion of unsaturated fatty acids (79.80%), mainly linoleic acid, oleic acid, a low percentage of saturated, total saturated 20.20%, and a very low level of n-3 poly-unsaturated FA (0.5%). The seed oil of *Citrullus colocynthis* has the following properties: iodine value = 114.46 g I₂/100 g, density at 15°C = 905.3:kg/m³, kinematic viscosity at 40°C = 31.52 mm²/s, saponification value = 204.44 mg KOH/g, acid value = 0.98 mg KOH/g, free fatty acid = 0.49%, caloric value = 39.37 MJ/kg, color = 5Y + 0.4R, and average molecular weight = 874 g (25).

The mineral contents of fermented and unfermented *Citrullus colocynthis* include: Ca 0.250 ± 0.04 , Mg 0.139 ± 0.041 , K 0.244 ± 0.04 , Na 0.36 ± 0.02 , P 0.176 ± 0.022 mg/kg; and Ca 0.341 ± 0.18 , Mg 0.167 ± 0.12 , K 0.327 ± 0.10 , Na 0.034 ± 0.16 and P 0.097 ± 0.14 mg/kg, respectively (26).

Effect of pH

Solution pH is one of the most important parameters affecting the chemical properties of both absorbing and adsorbing. In this study, pH in the range of 2-12 was used. As presented in Figure 3, the removal efficiency was decreased when the pH value was increased. This result can be explained by the increase in OH ions and their competitive effect with the phenol anion. As seen in Figure 3, the highest and lowest removal rates were observed in the pH values of 2 (83.4%) and 12 (66.4%), respectively (Figure 3).

Effect of adsorbent dose on removal efficiency

To determine the effect of adsorbent dosage on the removal efficiency of phenol, adsorbent doses of 0.5, 1, 3, 5, and 10 g/L, initial phenol concentration of 50 mg/L, pH: 2, and 60 minutes contact time were considered. As shown in Figure 4, the phenol adsorption rate increased with

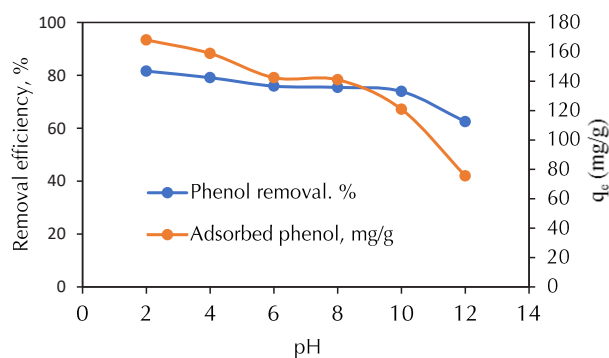


Figure 3. Effects of pH on phenol removal efficiency (phenol concentration 50 mg/L, adsorbent dose 3 g/L, contact time 60 min).

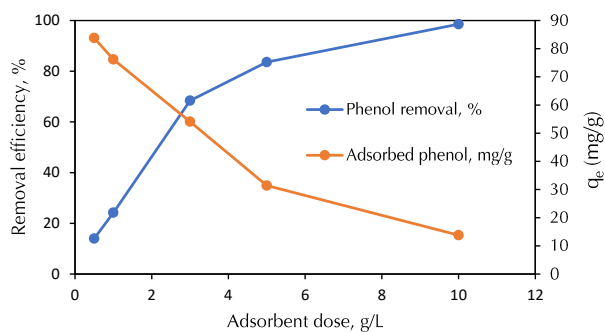


Figure 4. Effects of adsorbent dose on adsorption of phenol by *Citrullus colocynthis* seed ash (phenol concentration 50 mg/L, pH 2, contact time 60 min).

phenol concentration because of the increased number of sorption sites. Thus, the removal of phenol depends on its concentration.

Effects of initial phenol concentration on efficiency of adsorption process

The effects of primary phenol concentration (10, 20, 50, 80, and 100 mg/L) at contact times of 2, 5, 10, 30, and 60 minutes and an adsorbent dose of 3 g/L were evaluated. Figure 5 shows that by increasing the initial phenol concentration, the phenol removal efficiency was decreased. According to Figure 6, the highest removal efficiency of phenol was obtained in the first 10 minutes

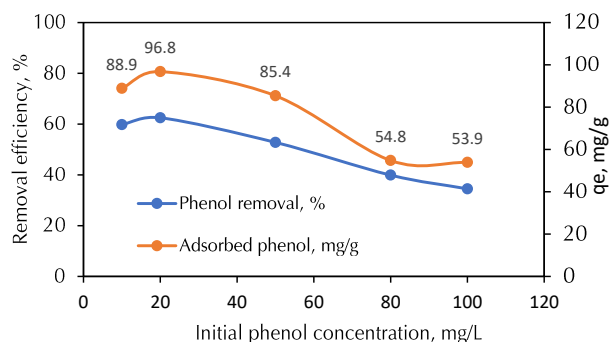


Figure 5. Effects of initial phenol concentration on adsorption process by *Citrullus colocynthis* seed ash (adsorbent dose of 3 g/L, pH 2).

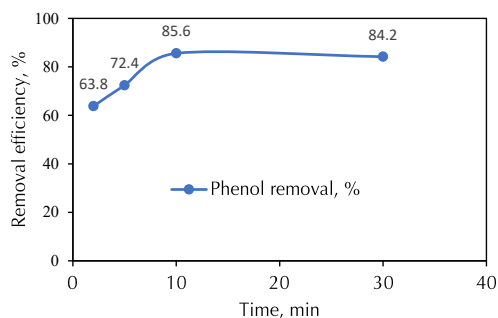


Figure 6. Effect of contact time on adsorption process by *Citrullus colocynthis* seed ash (Phenol concentration of 20 ppm, adsorbent dose of 5 g/L, pH 2).

of the test (Figure 6), and the removal efficiency remained almost constant as it reached equilibrium values.

Langmuir and Freundlich isotherms

A regression coefficient (R^2) higher than 0.9 was obtained for both models. The results showed that the Freundlich model is more appropriate for concluding the balanced adsorption of phenol on the adsorbent than the Langmuir model (Figures 7 and 8). Obtained isotherm parameters for phenol adsorption are presented in Table 1.

X-ray diffraction results

One of the most powerful techniques for characterizing crystalline materials is X-ray diffraction (XRD). XRD provides information on crystal structure and texture, phase, mean grain size, crystallinity, strain, and crystal defects (27). The constructive interference of a monochromatic beam of X-rays that diffract at specific angles from each set of lattice planes produce XRD peaks. The distribution of atoms within the lattice determines the peaks' intensities. As seen in Figure 2, the two broad diffraction peaks observed at around 18° and 22° vividly show that the adsorption process made little change in the surface structure of the adsorbent (27) and included compounds such as $C_{18}H_{42}CL_3N_6O_2$ and $C_{18}H_{42}CL_3LAN_6O_{21}$.

Discussion

Effects of pH

The pH of a solution is one of the most important and

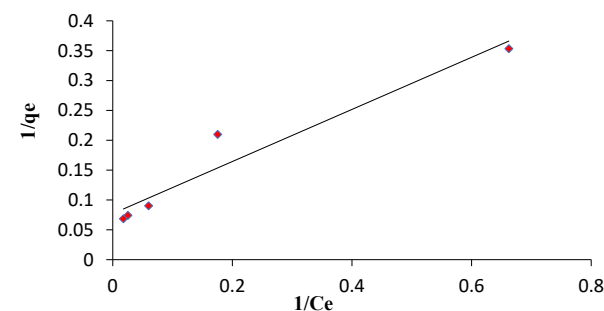


Figure 7. Appropriateness of the Langmuir model to phenol adsorption ($R^2 = 0.9348$).

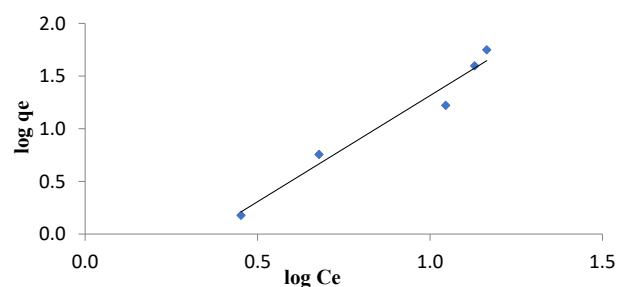


Figure 8. Appropriateness of the Freundlich model to phenol adsorption ($R^2 = 0.9674$).

Table 1. Obtained isotherm parameters for phenol adsorption

Langmuir isotherm			Freundlich isotherm		
q_m (mg g ⁻¹)	K_L (L mg ⁻¹)	R^2	k_f	n	R^2
13.33	0.092	0.9348	1.75	1.51	0.9674

effective parameters of the adsorption process, because it affects the electrical charge on the adsorbent surface and the ionization degree of the adsorbate (28). According to Figure 3, the removal efficiency of phenol decreased when pH was increased. The highest and lowest removal efficiency rates were 83.4% in pH=2 and 64.1% in pH = 12, respectively. These results can be due to the increase in OH ions and their competitive effects with phenol anion. The ion section of phenol is calculated by the following formula:

$$\phi_{ions} = \frac{1}{[1 + 10^{(PKa - PH)}]}$$

According to the formula above, ϕ_{ions} is increased by increasing pH. Phenol is a weak acid with PKa=10 that is greatly ionized in solutions. These ions are negatively charged, and because the adsorbent surface is positively charged in low values of pH, a strong attraction is created between the positive and negative ions (29). Senturk et al studied the removal rate of phenol from aqueous solutions by adsorption onto organomodified Tirebolu bentonite (30). Their results revealed that phenol adsorption was constant in pH values of 3-9, but decreased suddenly in a pH higher than 9. Similar results were presented for the adsorption of phenol by means of activated carbon, bentonite, and bagasse fly ash, by Halhouli et al (31), Banat et al. (32), and Srivastava et al, respectively (33).

Effects of adsorbent dose

According to Figure 4, the removal efficiency was increased when adsorbent dose was increased because of the increase in adsorbent surface area and the availability of more adsorption sites (31). The optimal elimination rate of phenol was 86.8 % (5 g/L). By increasing the amount of adsorbent to more than 5 g/L, phenol desorption occurred in the solution. The low adsorption rate can be explained by the lack of phenol concentration in high values of adsorbent (from 5-10 g/L). Similar results were achieved by Cengiz and Cavas (34) and SenthilKumar et al (35). However, these findings differed from the results of a study by Rahmani et al., which showed that by increasing the adsorbent dose from 0.25 g to 1 g, the residual phenol values were decreased from 40 mg/L to 1 mg/L. Moreover, the results showed that the amount of adsorbed contaminant per mass unit was decreased by increasing adsorbent values. In accordance with these results, the suitable dose of adsorbent was determined to be 0.5 g (36).

Effects of contact time and primary phenol concentration

The results of this study showed that the efficiency of

phenol removal was increased by increasing contact time in the first 10 minutes of the test (Figure 6). The increase in the rate of phenol removal was probably due to the limited empty spaces of adsorbent in high phenol concentrations. Also, these results showed that phenol adsorption is a fast process, because the highest amount of the considered concentrations of phenol was absorbed in the first 10 minutes of the test. The results of a study conducted by Dehghani et al to evaluate the efficiency of multiwalled carbon nanotubes in the removal of phenol from aqueous solutions revealed that phenol removal was increased when contact time was increased. The current study results differed with these in that Dehghani obtained maximum adsorption in the first 30 minutes of the test (37). The cause of this event is the existence of many available empty sites for adsorption at first; with the passage of time, the occupation of the remained sites becomes difficult because of the repulsive force between the solute molecules between the solid phase and the solution (38). In another study conducted by Liao et al on the adsorption of chlorothenols by multiwalled carbon, an equilibrium time of 30 minutes was obtained (39).

As seen in Figure 5, the effect of initial phenol concentration on the adsorption process showed that by increasing phenol concentration, removal efficiency was decreased. The highest and lowest phenol removal rates were obtained in concentrations of 20 ppm (66.4%) and 100 ppm (33.2%), respectively, because the initial phenol concentration acts as a driving force, and the resistance of solution mass transfer increases in the adsorbent surrounding the liquid layer and adsorbent surface. Also, increasing phenol concentration decreased the ratio of available surface to the number of adsorbed moles, and that led to a reduction in removal efficiency and an increase in residual phenol in the solution. Senturk et al reported similar results (30).

Phenol adsorption isotherm (Langmuir and Freundlich)

Several different models are used to describe adsorption. One of the most important models is adsorption isotherms. In this study, the Langmuir and Freundlich models were used to describe the relationship between the amount of adsorbed phenol and its equilibrium concentration in the solution (40). Adsorption isotherm studies were conducted under optimized conditions (pH= 2, adsorbent dosage=5 g/L), phenol concentrations ranging from 10 to 80 mg/L, and a contact time of 10 minutes. The investigation of correlation coefficients showed that the obtained regression coefficient was higher than 0.9 for both models, and the phenol adsorption by *C. colocynthis*

seed ash showed more suitability with the Freundlich model than the Langmuir model (Figures 7 and 8). Similar findings were reported by Feng et al. Equilibrium data was in greater accordance with the Freundlich model than the Langmuir model (41). This finding was also reported in the literature for phenol removal by other adsorbents (42,43).

Conclusion

In the current study, it was observed that *Citrullus colocynthis* seed ash can be used effectively for the removal of phenol from aqueous solutions. Among all parameters, pH had an important role in the adsorption process by this adsorbent. Phenol adsorption was greater in low pH values than in high values of pH. The results further revealed that the removal efficiency of phenol was increased by increasing adsorbent dose, contact time, and temperature. The evaluation of correlation coefficients showed that phenol adsorption by *Citrullus colocynthis* seed ash was in greater accordance with the Freundlich model than the Langmuir model. Thus, *C. colocynthis* seed ash has suitable potential for use in the removal of phenol on operation and practical scales due to its low cost and easy access.

Acknowledgments

The authors express their gratitude to the Gonabad University of Medical Sciences for the financial support of this project (Project number: 95/79).

Ethical issues

The authors hereby certify that all data collected during the study is as stated in this manuscript, and no data from the study has been or will be published elsewhere separately.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

All authors contributed equally and were involved in the study design, data collection, and article approval.

References

1. Razeghi N, Hashemi SH, Jadidi KH, Aghapoor K. Performance of Mesopore MCM-41 with a Nanoporous Structure in Removing Phenol from Water. *Journal of Water and Wastewater* 2015; 25(6): 2-9. [In Persian].
2. Saeedi M, Pajooheshfar SP. Investigation of phenol adsorption from contained water by activated carbon and carbon of almonds and walnut skin. *Quarterly Journal of Environmental Science and Technology* 2008; 10(4): 218-31. [In Persian].
3. Afsharnia M, Saeidi M, Zarei A, Narooie MR, Biglari H. Phenol removal from aqueous environment by adsorption onto pomegranate peel carbon. *Electro physic* 2016; 8(11): 3248-56.
4. Nakamoto S, Machida N. Phenol removal from aqueous solutions by peroxidase-catalyzed reaction using additives. *Water Res* 1992; 26(1): 49-54. doi: 10.1016/0043-1354(92)90110-P.
5. Caetano M, Valderrama C, Farran A, Cortina JL. Phenol removal from aqueous solution by adsorption and ion exchange mechanisms onto polymeric resins. *J Colloid Interface Sci* 2009; 338(2): 402-9. doi: 10.1016/j.jcis.2009.06.062.
6. Tor A, Cengeloglu Y, Aydin ME, Ersoz M. Removal of phenol from aqueous phase by using neutralized red mud. *J Colloid Interface Sci* 2006; 300(2): 498-503. doi: 10.1016/j.jcis.2006.04.054.
7. Brown VK, Box VL, Simpson BJ. Decontamination procedures for skin exposed to phenolic substances. *Arch Environ Health* 1975; 30(1): 1-6. doi: 10.1080/00039896.1975.10666623.
8. Baker EL, Landrigan PJ, Bertozzi PE, Field PH, Basteys BJ, Skinner HG. Phenol poisoning due to contaminated drinking water. *Arch Environ Health* 1978; 33(2): 89-94. doi: 10.1080/00039896.1978.10667314.
9. Babich H, Davis DL. Phenol: a review of environmental and health risks. *Regul Toxicol Pharmacol* 1981; 1(1): 90-109. doi: 10.1016/0273-2300(81)90071-4.
10. Maleki A, Mahvi H. Application of agricultural waste products for removal of phenol in aqueous systems. *Medical Journal of Hormozgan University* 2007; 10(4): 393-9. [In Persian].
11. World Health Organization. *Guidelines for Drinking-Water Quality*. Geneva: WHO; 2004.
12. Roostaei N, Tezel FH. Removal of phenol from aqueous solutions by adsorption. *J Environ Manage* 2004; 70(2): 157-64. doi: 10.1016/j.jenvman.2003.11.004.
13. Wu C, Liu X, Wei D, Fan J, Wang L. Photosonochemical degradation of phenol in water. *Water Res* 2001; 35(16): 3927-33. doi: 10.1016/S0043-1354(01)00133-6.
14. Mukherjee S, Kumar S, Misra AK, Fan M. Removal of phenols from water environment by activated carbon, bagasse ash and wood charcoal. *Chem Eng J* 2007; 129(1-3): 133-42. doi: 10.1016/j.cej.2006.10.030.
15. Potgieter JH, Bada SO, Potgieter-Vermaak SS. Adsorptive removal of various phenols from water by South African coal fly ash. *Water Sa* 2009; 35(1): 89-96.
16. Malakootian M, Mesdaghinia A, Rezaei S. Efficiency of ortho-chlorophenol removal from aqueous solutions using activated Fly Ash of Zaraned Fossil Fuel Power Plant. *Scientific Journal of School of Public Health and Institute of Public Health Research* 2014; 12(2): 81-92. [In Persian].
17. Daifullah AA, Girgis BS. Removal of some substituted phenols by activated carbon obtained from agricultural waste. *Water Res* 1998; 32(4): 1169-77. doi: 10.1016/S0043-1354(97)00310-2.
18. Gholizadeh A, Kermani M, Gholami M, Farzadkia M, Yaghmaeian K. Removal efficiency, adsorption kinetics and isotherms of phenolic compounds from aqueous solution using rice bran ash. *Asian J Chem* 2013; 25(7): 3871-3878.
19. Manshouri M, Daraei H, Yazdanbakhsh AR. The feasibility study of ostrich feathers in removing phenol from aqueous solution. *J Kermanshah Univ Med Sci* 2012; 16(5): 359-66.
20. Bazrafshan E, Kordmostafapour F, Faridi H, Barikbin B. Application of Moringa Peregrina seed extract as a natural coagulant for Phenol removal from aqueous solution. *Journal of Birjand University of Medical Sciences* 2013;

- 19(4): 389-98. [In Persian].
21. Bazrafshan E, Biglari H, Mahvi A. Performance evaluation of electrocoagulation process for phenol removal from aqueous solutions. *Fresenius Environmental Bulletin* 2012; 21(2): 364-71.
 22. Zarei A, Bazrafshan E, Faridi H, Khaksefidi R, Alizadeh M. The evaluation of removal efficiency of phenol from aqueous solutions using *Moringa peregrina* tree shell ash. *Iranian Journal of Health Sciences* 2013; 1(1): 65-74. doi: 10.18869/acadpub.jhs.1.1.65.
 23. American Public Health Association (APHA). Standard Methods for examination of water and wastewater. Washington: American Public Health Association; 2012
 24. Mall ID, Srivastava VC, Agarwal NK, Mishra IM. Adsorptive removal of malachite green dye from aqueous solution by bagasse fly ash and activated carbon-kinetic study and equilibrium isotherm analyses. *Colloids Surf A Physicochem Eng Asp* 2005; 264(1-3): 17-28. doi: 10.1016/j.colsurfa.2005.03.027.
 25. Dyanati R, Yousefi Z, Yazdani Cherati J, Balarak D. Investigating phenol absorption from aqueous solution by dried azolla. *Journal of Mazandaran University of Medical Sciences* 2013; 22(2): 13-20. [In Persian].
 26. Nath K, Panchani S, Bhakhar MS, Chatrola S. Preparation of activated carbon from dried pods of *Prosopis cineraria* with zinc chloride activation for the removal of phenol. *Environ Sci Pollut Res Int* 2013; 20(6): 4030-45. doi: 10.1007/s11356-012-1325-y.
 27. Shrestha S. Chemical, structural and elemental characterization of biosorbents using FE-SEM, SEM-EDX, XRD/XRPD and ATR-FTIR Techniques. *J Chem Eng Process Technol* 2016; 7(3):2-11. doi: 10.4172/2157-7048.1000295.
 28. Giwa S, Abdullah LC, Adam NM. Investigating "Egusi" (*Citrullus Colocynthis* L.) seed oil as potential biodiesel feedstock. *Energies* 2010; 3(4): 607-18. doi: 10.3390/en3040607.
 29. Al-Snafi AE. Chemical constituents and pharmacological effects of *Citrullus colocynthis* - A review. *IOSR J Pharm* 2016; 6(3): 57-67.
 30. Senturk HB, Ozdes D, Gundogdu A, Duran C, Soylak M. Removal of phenol from aqueous solutions by adsorption onto organomodified Tirebolu bentonite: equilibrium, kinetic and thermodynamic study. *J Hazard Mater* 2009; 172(1): 353-62. doi: 10.1016/j.jhazmat.2009.07.019.
 31. Halhouli KA, Darwish NA, Al-Dhoon NM. Effects of pH and inorganic salts on the adsorption of phenol from aqueous systems on activated decolorizing charcoal. *Sep Sci Technol* 1995; 30(17): 3313-24. doi: 10.1080/01496399508013147.
 32. Banat FA, Al-Bashir B, Al-Asheh S, Hayajneh O. Adsorption of phenol by bentonite. *Environ Pollut* 2000; 107(3): 391-8. doi: 10.1016/S0269-7491(99)00173-6.
 33. Srivastava VC, Swamy MM, Mall ID, Prasad B, Mishra IM. Adsorptive removal of phenol by bagasse fly ash and activated carbon: equilibrium, kinetics and thermodynamics. *Colloids Surf A Physicochem Eng Asp* 2006; 272(1-2): 89-104. doi: 10.1016/j.colsurfa.2005.07.016.
 34. Cengiz S, Cavas L. Removal of methylene blue by invasive marine seaweed: *Caulerpa racemosa* var. *cylindracea*. *Bioresour Technol* 2008; 99(7): 2357-63. doi: 10.1016/j.biortech.2007.05.011.
 35. SenthilKumar P, Ramalingam S, Sathyaselvabala V, Dinesh Kirupha S, Sivanesan S. Removal of copper (II) ions from aqueous solution by adsorption using cashew nut shell. *Desalination* 2011; 266(1-3): 63-71. doi: 10.1016/j.desal.2010.08.003.
 36. Rahmani A, Asgari G, Barjasteh Asgari F, Hedayati Kamran E, Alijani F. Investigation of phenol removal from aqueous solutions using copper-impregnated pumice. *Avicenna Journal of Clinical Medicine* 2011; 17(4): 50-6. [In Persian].
 37. Dehghani MH, Alimohammadi M, Mahvi AH, Rastkari N, Mostofi M, Gholami M. Performance of multiwall carbon nanotubes for removal phenol from aqueous solutions. *Iranian Journal of Health and Environment* 2014; 6(4): 491-502. [In Persian].
 38. Wang L, Zhang J, Zhao R, Zhang C, Li C, Li Y. Adsorption of 2,4-dichlorophenol on Mn-modified activated carbon prepared from *Polygonum orientale* Linn. *Desalination* 2011; 266(1-3): 175-81. doi: 10.1016/j.desal.2010.08.022.
 39. Liao Q, Sun J, Gao L. Adsorption of chlorophenols by multi-walled carbon nanotubes treated with HNO₃ and NH₃. *Carbon* 2008; 46(3): 553-5. doi: 10.1016/j.carbon.2007.12.009.
 40. Gholizadeh A, Kermani M, Gholami M, Farzadkia M. Kinetic and isotherm studies of adsorption and biosorption processes in the removal of phenolic compounds from aqueous solutions: comparative study. *J Environ Health Sci Eng* 2013; 11(1): 29. doi: 10.1186/2052-336x-11-29.
 41. Feng J, Qiao K, Pei L, Lv J, Xie S. Using activated carbon prepared from *Typha orientalis* Presl to remove phenol from aqueous solutions. *Ecol Eng* 2015; 84: 209-17. doi: 10.1016/j.ecoleng.2015.09.028.
 42. Viswanathan N, Meenakshi S. Enriched fluoride sorption using alumina/chitosan composite. *J Hazard Mater* 2010; 178(1-3): 226-32. doi: 10.1016/j.jhazmat.2010.01.067.
 43. Dou X, Zhang Y, Wang H, Wang T, Wang Y. Performance of granular zirconium-iron oxide in the removal of fluoride from drinking water. *Water Res* 2011; 45(12): 3571-8. doi: 10.1016/j.watres.2011.04.002.