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High temporal resolution sampling reveals reef fish settlement is highly clustered

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15 Abstract

16 Coral reef fish larvae settle on reefs predominantly at night around the new-moon phase, after an early developmental period spent in the pelagic environment. Most sampling is conducted 17 across whole nights, and any studies that have examined the frequency of arrival within 18 nights have typically been limited to coarse sampling time scales of 1–5 hours. Here, we 19 20 present results for arrival numbers of fish caught between dusk and midnight from light traps 21 sampled every 15 min at an Indonesian coral reef, providing the finest temporal resolution for 22 this type of study to date. A Spatial Analysis by Distance IndicEs (SADIE) analysis, adapted 23 to temporal data, revealed clustering of reef arrival times for many species, with an increase 24 in catches immediately after dusk dropping off towards midnight. Importantly, the timing of 25 clusters differed among species indicating that different factors determine the timing of 26 arrival among taxa. Our results support the hypothesis that larval behaviour influences the 27 timing of arrival at a coral reef for different fish species.

Keywords: Coral reefs; fish larvae; larval behaviour; larval settlement; SADIE analysis; IndoPacific.

30 Introduction

31 Most coral reef fish spend their early life stage as larvae in the open ocean before returning to reefs to 32 settle (Montgomery et al. 2001; Kingsford et al. 2002; Leis and McCormick 2002). Settlement stage 33 larvae move onto the reef predominantly at night (Robertson et al. 1988; Stobutzki and Bellwood 34 1998), with a few species potentially also settling to the reef during the daytime (e.g. Dufour and 35 Galzin 1993; Kingsford 2001). Settlement is higher around the new and third quarter moon, when 36 moonlight is weak (Milicich 1988; Meekan et al. 1993; Milicich and Doherty 1994; Sponaugle and 37 Cowen 1997; Leis and Carson-Ewart 1999; D'Alessandro et al. 2007;). Nocturnal settlement has 38 been suggested as a life strategy for avoidance of predators feeding on fish larvae at the reef (Hamner 39 et al. 1988), which could otherwise have a significant impact on the already high mortality rate of 40 settlement stage fishes (Almany and Webster 2006; Dytham and Simpson 2007). The high risk of 41 mortality typical of this life stage of demersal reef fishes is also thought to drive other behaviours in 42 larval fish including auditory and olfactory orientation (Atema et al. 2002; Simpson et al. 2004, 2005) 43 and schooling (Pitcher 1986; Leis 2006; Codling et al. 2007; Simpson et al. 2013; Irisson et al. 2015). 44 Because of the small size of larval fish and their tendency to settle during low-light levels, it is 45 challenging to directly observe larval behaviour in situ, which often has to be inferred from controlled 46 environments (e.g. olfactory preference Dixson et al. 2008; swimming speed Fisher and Bellwood 47 2003) or *in situ* observations made during the daytime (Dixson *et al.* 2014; Simpson *et al.* 2005; Leis 48 et al. 2015). From nocturnal sampling methods that are highly restricted in space and time it should be 49 possible to make informed speculations on the behaviours that drive settlement patterns. However, 50 only a few studies have examined the arrival patterns of fish over single night periods (Dufour et al. 51 1996; McIlwain 1997; Wilson 2003), with the highest temporal resolution being used typically in the range of one recording every 1–2 hours. Whilst this can provide us with useful information on when 52 53 the main settlement peaks occur during the night, it is not sufficient temporal resolution to capture 54 arrival of individual cohorts of fish. In the present study we assessed arrival of fish larvae to 55 Indonesian reefs using light traps sampled every 15 min: a finer temporal resolution than any previous 56 sampling method used in this context. The light trap sampling technique is stage selective, capturing

57 phototactic larval species at a competent settlement stage in high numbers. It does have species 58 sampling biases dependent on the degree of phototactic response and the swimming abilities of the 59 individual larvae (Choat et al. 1993). Overall, however, this technique lends itself well to the study of 60 larval settlement, providing large catches and ease of replication (Milichich 1988; Choat et al. 1993; 61 Wilson 2001, 2003). The focus of this study was not to examine the arrival rate of settlement stage 62 fish for this region (which would also be of interest), as it does not cover lunar, seasonal, or yearly 63 temporal variation, but to explore at high temporal resolution the arrival of fish within single nights. 64 We hypothesised that: a) settlement stage fish would not arrive at a constant rate in the light traps but 65 would display a high degree of clustering in the catches due to behaviour influencing settlement 66 times; and b) that the arrival rate of different species would follow different arrival patterns if 67 hydrodynamic forces were not the main drivers of clustering patterns.

68 Materials and Methods

69 Sites

70 Sampling was carried out at two sites offshore from the coral reefs of Hoga Island, Southeast 71 Sulawesi, Indonesia (Fig.1), selected based on their different water flow regimes. Buoy 1 is a site 72 located on the reef in the channel between Hoga Island and Kaledupa Island through which a large 73 body mass of water moves daily with the tides. Buoy 5 is located off the western reef of the island and 74 is relatively sheltered although the large changes in tidal range ($\sim 2m$) can result in relatively strong 75 water currents during tide flows towards and away from the shore. There were differences in the 76 acoustic composition of the reef sound spectra and some of the coral composition of the reef (Piercy 77 2015), but no other known significant differences between them.



Figure 1. Sites of light trap sampling off Hoga Island, Indonesia (5° 28' 20" S; 123° 45' 25" E). Buoy
1 is located in the channel between Hoga Island and Kaledupa (bottom left corner of this map) where
the water current is strongest. Buoy 5 is located in a central position along the Hoga Island western
reef and is dominated by inshore and offshore currents as the reef flat (45° grey bars) fills and drains.

84 Sampling

85 Sampling was carried out between the third quarter moon, 24 July 2011, and three days after the new 86 moon, 3 August 2011 (new moon was 31 July 2011) between dusk and midnight (18:00–00:00). The 87 focus of this study was on collecting high temporal resolution data rather than lunar or seasonal 88 changes that have been previously extensively examined (e.g., Robertson et al. 1988; Stobutzki and 89 Bellwood 1998; Dufour and Galzin 1993; Kingsford 2001). This "snapshot" approach is 90 representative of a typical new moon recruitment period but may differ in terms of abundance and 91 species composition compared to other times of the year. Throughout the night time period, light 92 levels remained low as the moon had not yet risen with the exception of the last two sampling dates (1

93 and 3 August) when the moon did not set until 19:30 and 21:00 respectively. However, considering 94 the proximity of these dates to the new moon phase the maximum fraction of the moon illuminated 95 was 17.8% (3 August) with the moon at a maximum altitude of 40° (18:15 2011-08-03) estimated 96 using the MoonAngle function in R (package oce, version 0.9-20), with half or more of the sampling 97 time being undertaken in the absence of any moonlight. Buoy 1 was sampled on nights of the 24 July 98 2011, and 1 and 3 August 2011, while B5 was sampled on 26, 28 and 30 July 2011. 99 Sampling was conducted using two Stobutzki and Bellwood (1997) light traps, suspended 1 m below 100 the water surface and fishing alternatively every 15 min throughout the sampling periods. The traps 101 were fitted with a 40 W 12 V fluorescent tube light connected to a 12 V 12 Ah lead acid battery 102 charged to >13 V. A boat was moored to a buoy to which the light trap was attached during 103 deployment at 1.5 m below the sea surface. The boat was attached to the mooring using 30 m of rope 104 to maintain it at a distance from the light trap and mooring and to minimise disturbance to recruiting 105 fish. Every 15 min the boat was hauled closer to the first light trap, and the second light trap was 106 deployed immediately prior to the first one being extracted from the water to maintain a constant light 107 source in the water column. The contents of the first light trap were emptied via a funnel into a 108 10x10x10 cm container with mesh netting openings on the sides and sealed with a mesh netting cap. 109 The operation was carried out within a 20 L bucket containing fresh seawater 10 cm deep to maintain 110 the fish alive and re-capture any escaped fish during the emptying operation. The container was then 111 placed in a fish mesh holder over the side of the boat. Once all 24 samples over the night period had 112 been collected, the mesh containers were placed inside a 100 L polystyrene container filled with fresh 113 seawater, brought back to the shore, and kept aerated using two airstones. The next day, fish from 114 each container were sedated using a mixture of clove oil, 90% ethanol, and sea water in a ratio of 115 1:3:6 respectively (see ethical note in Simpson et al. 2008). The fish from each container were spread 116 out on a gridded tray with 1 cm of water and photographed lying on their side using a Samsung S860 117 8.1 MP digital camera (Samsung Electronics Co., Ltd., Suwon, South Korea) and allowed to recover 118 in a 50 L aerated plastic tank containing fresh seawater before being released back onto the reef at 119 dusk. Fish were identified from the photos to family level using the guide by Leis and Carson-Ewart

120 (2000), and to genera and species where feasible. Fish that were difficult to identify were preserved in 121 70% ethanol after being anaesthetised and photographed using a Veho VMS-1 USB microscope with 122 x200 maximum magnification (Veho Electronics, Hampshire, UK). All work was carried out under 123 permits held by D. Smith and issued by the Indonesian Minister for Research and Development. 124 *Clustering analysis* 125 To test for non-randomness and provide indices for the degree of clustering on the temporal 126 distribution of fish counts, the Spatial Analysis by Distance IndicEs (SADIE) methodology (Perry et 127 al. 1999) was applied using the SADIEShell program (Open Source under GNU General Public 128 License, V3). This analysis is usually used to test for spatial clustering of species in a two-129 dimensional grid, but can also be applied to one-dimensional data sets such as quadrats positioned in 130 line along a transect (Perry et al. 2002). Since the calculation of the indices is based on a "spatial" 131 matrix of count data, to transpose this problem to a one-dimensional (1-D) context an artificial extra 132 dimension was included where units on the x axis were represented by the times of sampling and their 133 position on the y axis was assumed to be a constant (i.e. y = 1; Perry, pers. comm.). Hence, for I = 134 1,..., n cells were of the form $(x_i, 1)$, each containing an observed sample count. This method 135 calculates three indices for the counts of data before testing them against random permutations of the 136 counts to provide the probability p of the measures of aggregation against the randomised ones (Perry 1998; Perry et al. 1999). The first measure is the index of aggregation (Perry 1998), which quantifies 137 138 the degree of effort required for each cell to reach an even distribution across all cells. The index of 139 aggregation, *Ia*, is defined as:

140

$$Ia = D/Ea \tag{1}$$

where *D* is the distance to regularity, defined as the minimum value of the total distance that the *ith* individual sample unit at position (x_i , 1) would have to move, from one unit to the next, so that all units contained an identical count. *Ea* is the arithmetic mean distance to regularity for the randomised samples.

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The second index is the *distance to crowding index* (Perry 1998), a measure of the minimum effort required for the counts of each cell to move into a single cell. The distance to crowding index, J_a , is defined as

$$J_a = F_a/C \tag{2}$$

where *C* is the distance to crowding, defined as the minimum value of the total distance that the ith individual sample unit at position (x_i, I) must move so that all are congregated in one unit. F_a is the arithmetic mean distance to crowding for the randomised samples. This index increases in value as the distance to crowding *C* decreases, (i.e. a high index value indicates a more clustered group, with a lower distance needed for all data to move to the same sampling point). The index is more powerful at detecting a cluster than the index of aggregation I_a but cannot be interpreted correctly if more than one cluster is present.

The final measure is the *degree of clustering*. Similar to the distance to regularity, it calculates the effort required for each cell to reach an even distribution among the cells. However, in this case it computes the degree to which each data point influences the overall clustering, by calculating the strength of inflow and outflow from one cell to another in order to reach an even count between cells. For donor unit *i*, at position (x_i , *l*), the outflow to the *j* of n_i receiver units, $j = 1, ..., n_i$, at position (x_j , *l*), is denoted as v_{ij} . The distance of this flow d_{ij} is

162
$$d_{ij} = \sqrt{(x_i - x_j)^2 + (y_i - y_j)^2} = |x_i - x_j|$$
(3)

163 The average distance of outflow from unit *i*, weighted by the magnitude of each individual flow, is 164 denoted as Y_i , where $Y_i = \sum_j d_{ij} v_{ij} / \sum_j v_{ij}$. The same calculation is carried out for inflows.

The distances for inflows and outflows need to be standardised in order to obtain the dimensionless
clustering indices. For outflows, a standardised and dimensionless index of clustering, v_i, is then given
by:

$$v_i = Y_{io}Y_{ij}Y_{ij}Y_{ij}$$

169 where Y_i is the distance of the flow; i Y is the expected value of the average absolute flow distance for 170 the *i* unit, assuming a random arrangement of the observed counts among the observed sample units, 171 where the outflows are computed for each count that is randomly assigned to the *i* Unit. Similarly, 172 instead of following the unit i, through the counts randomised to it, we can follow the count c through 173 its randomisations to different units, where Y is the expected value of the average flow distance for 174 the observed count c; $_{0}$ Y is the expected value of the overall average absolute distance of flow for all 175 points and counts in the randomisations. By convention, inflows are given negative scores and 176 outflows positive scores and the average of their absolute values is used as the clustering index. Only 177 taxa for which at least an average of one individual per sampling time was collected on any one day 178 (i.e. > 24 individuals) were included in the analysis.

179 *Association analysis*

180 Temporal association between the arrival rates of different fish taxa was measured using the SADIE

181 association index. This method first calculates the similarity between the clustering indices of the two

182 populations at each time point then averages those similarities to provide an overall measure of

183 association: the correlation coefficient.

184 The measure of local spatial association for unit *i* is given by:

185
$$\chi_i = \frac{n(z_{i1} - q_1)(z_{i2} - q_2)}{\sqrt{\sum_k (z_{i1} - q_1)^2 \sum_k (z_{i2} - q_2)^2}}$$
(5)

where z_{k1} denotes the clustering indices of the first set of data, with mean q_1 , and z_{k2} denotes the clustering indices for the second set of data, with mean q_2 . The overall spatial association is the mean of these local values:

$$X = \sum_{i \chi_i} / n \tag{6}$$

where X is the correlation coefficient between the clustering indices of each set. The correlation is not performed directly on the counts because large count values would contribute disproportionately to the correlation coefficient. The significance of the association or disassociation is calculated by permutation of the counts. Since the randomisation process produces a histogram with the probability for both association and disassociation, the α value to accept or reject the null hypothesis is divided between the two ends of the distribution. Thus $\alpha = 0.05$ is split between the two extremities so that the null hypothesis is rejected either for p > 0.975 (significant disassociation) or p < 0.025 (significant association).

198 Autocorrelation is a property present in all clustering scenarios and could increase the chance of 199 finding non-existent associations and dis-associations because of the lack of independence of the 200 samples from one another. To minimise this effect and as recommended in Perry et al. (2002), a 201 Dutilleul adjustment was applied to account for the degree of autocorrelation and reduce the effective 202 sample size (Dutilleul 1993). To do this, where necessary, the sets were detrended, and the degrees of 203 freedom corrected for correlation, M-2, where M is the effective sample size. The critical values are inflated by a scale factor of $\sqrt{\frac{M-3}{n-3}}$ and the significance of the randomisation test adjusted accordingly. 204 205 An extreme example of this would be if all samples are so strongly autocorrelated that we only need 206 one sample in order to estimate the size of all the remaining samples, effectively reducing the sample 207 size to one.

208 Results

209 During the 144 light trap deployments and collections a total of 2,187 individual fish were caught, 210 representing 28 families, of which some Pomacentridae, Apogonidae, and Sygnathidae could be 211 identified to genus level and even separated into species. The species themselves could rarely be 212 identified due to lack of guides on species level identification for fish at the larval stage for this region 213 of high biodiversity; therefore some species may have been grouped under the same taxon. 214 No fish were caught during the first sampling period on any night (18:00–18:15) and only one 215 individual fish from the genus Spratelloides was caught on the second sampling period (18:15–18:30) 216 over the six sampling days. More fish were caught at B1 (1,656) compared to B5 (531 individuals; 217 Chi-square test, $\chi^2 = 37.8$, p<0.001, d.f. = 2).

218 Clustering

219 All taxa that met the inclusion criteria on at least one day (>24 individuals caught on that sampling 220 night), except Gobiidae and Synodontidae, displayed significant temporal clusters for at least one of 221 the measures for clustering (degree of clustering, index of aggregation and distance to crowding; 222 Table 1 and Figs. 2-5). The significance level of the degree of clustering and the index of aggregation 223 was in agreement for all taxa (i.e. they were either both significant or both non-significant). However, 224 the significance level of the distance to crowding index occasionally differed from the other two 225 indices. A significant clustering effect was observed in *Corythoichthys* sp.1 (family Sygnathidae) on 226 24 July 2011, Abudefduf sp.1 (family Pomacentridae) on 1 August 2011, and in Apogon sp.1 (family 227 Apogonidae), Apogon sp.3, Chromis sp. (family Pomacentridae) on 3 August 2011 according to the 228 distance to crowding but not the degree of clustering or the index of aggregation (Table 1 and Figs. 2 229 and 3). This is likely due to the greater sensitivity of the former index compared to the latter indices 230 when a single cluster is present. In contrast, the Gobiidae were significantly clustered according to the 231 degree of clustering and the index of aggregation on 30 July 2011 and 1 August 2011, but not 232 according to the distance to crowding index. This is likely due to the fact that the latter index only 233 provides meaningful results in the presence of a single cluster in the data, whilst two or three temporal 234 clusters appear to be present in Gobiidae catches on those days (Figs. 5a, b).

Table 1. Clustering indices from the SADIE methodology for fish taxa that met the minimum catch requirement (minimum 24 fish of the taxa caught over the

sampling night). Significant clustering indices are highlighted in bold with the significance level in brackets below (underlined).

Species or nearest identifiable taxon	Site	Day	# individuals	Degree of clustering (p)	Index of aggregation (p)	Distance to crowding index (p)
Apogon sp. 1	Buoy 1	2011-08-03	40	1.70 (0.069)	1.72 (0.063)	1.90 <u>(0.001)</u>
Apogon sp. 2	Buoy 1	2011-08-03	48	1.73 <u>(0.012)</u>	1.87 <u>(0.032)</u>	2.11 (0.001)
Apogon sp. 3	Buoy 1	2011-08-01	41	1.99 <u>(0.032)</u>	1.93 <u>(0.034)</u>	2.11 <u>(<0.001)</u>
		2011-08-03	52	1.44 (0.143)	1.59 (0.090)	1.82 <u>(0.003)</u>
Chromis sp.	Buoy 1	2011-07-24	245	2.18 <u>(0.015)</u>	2.29 <u>(0.010)</u>	1.64 <u>(<0.001)</u>
		2011-08-01	133	1.97 <u>(0.035)</u>	2.07 <u>(0.022)</u>	1.87 <u>(<0.001)</u>
		2011-08-03	133	1.57 (0.097)	1.68 (0.072)	1.76 <u>(<0.001)</u>
Gobiidae	Buoy 1	2011-08-01	42	<u> 1.99 (0.025)</u>	1.92 <u>(0.026)</u>	1.20 (0.209)
		2011-08-03	27	0.75 (0.685)	0.77 (0.654)	1.18 (0.173)
	Buoy 5	2011-07-26	63	1.39 (0.101)	1.30 (0.140)	0.96 (0.611)
		2011-07-28	38	1.45 (0.128)	1.58 (0.091)	1.39 (0.061)
		2011-07-30	50	1.90 <u>(0.021)</u>	1.80 <u>(0.035)</u>	1.36 (0.178)
Holocentridae	Buoy 1	2011-07-24	31	2.68 <u>(0.002)</u>	2.52 <u>(0.002)</u>	2.80 <u>(<0.001)</u>
Pomacentrus sp. 1	Buoy 1	2011-07-24	29	2.47 <u>(0.005)</u>	2.78 <u>(0.001)</u>	1.81 <u>(0.001)</u>
Corythoichthys sp.1	Buoy 1	2011-07-24	105	1.38 (0.140)	1.30 (0.180)	2.51 (0.001)
Synodontidae	Buoy 1	2011-07-24	31	0.45 (0.999)	0.51 (0.994)	0.69 (0.920)
Abudefduf sp.1	Buoy 1	2011-07-24	248	1.742 (0.060)	1.93 <u>(0.033)</u>	2.10 <u>(<0.001)</u>
		2011-08-01	53	2.12 <u>(0.022)</u>	2.30 <u>(0.011)</u>	2.17 <u>(<0.001)</u>
	Buoy 5	2011-07-26	27	1.23 (0.233)	1.23 (0.235)	1.49 (0.011)

238



241

Figure 2. Arrival numbers of different Pomacentrid taxa at Buoy 1 on three different nights. a) Arrival
numbers of *Chromis sp.*, the most abundant taxon; b) *Abudefduf* sp.1, the second most abundant
taxon; and c) *Pomacentrus* sp.1. The scales for the fish counts vary between panels. The time period
of the high (H.T.) and low (L.T) tides are indicated above their respective periods and the time of the
moon set is indicated with a dashed line and moon symbol. For 24 July there was no moon in the sky
during the sampling period. Nights on which a significant temporal cluster in arrival numbers was
present for at least one of the clustering indices presented in Table 1 are indicated by an asterisk (*).



Figure 3. Arrival numbers of different Apogonid taxa at Buoy 1 on three different nights. a) Arrival
numbers of *Apogon* sp.1; b) *Apogon* sp.2; and c) *Apogon* sp.3. Explanation of the symbols is given in
Fig. 2.



Figure 4. Arrival numbers of selected taxa at Buoy 1 on three different nights. The taxa presented all

256 met the selection criteria of >24 individuals on a single sampling night: a) Chorythoichtys sp.1

257 (Family: Sygnathidae); b) Holocentridae; and c) Synodontidae. Explanation of the symbols is given in

258 Fig. 2

259



Figure 5. Arrival numbers on six different nights of Gobiidae at: a) Buoy 1; and b) Buoy 5.

Explanation of the symbols is given in Fig. 2. Where >20 individuals were caught at a single sample

264 point, the number of fish caught is indicated above the bar.

265 Associations

266 There were significant associations in the arrival numbers among taxa of the same family (e.g.

267 Pomacentridae: Chromis sp. associated with Abudefduf sp.1 and Pomacentrus sp.1, and Apogonidae:

268 Apogon sp.1 associated with Apogon sp.3), but also between taxa from different families (e.g.

269 *Chromis sp.* associated with *Apogon* sp.1, and *Chromis sp.* associated with Gobiidae; Table 2). Most

association analyses between taxa only met the inclusion criteria for one of the sampling days (both

taxa with >24 individuals caught on the day). Exceptions to this were comparisons between *Chromis*

sp. and *Abudefduf* sp.1 (significantly associated on both days; X = 0.45, p = 0.02, and X = 0.68, p < 0.02, x = 0.02, y = 0

273 0.001 on 24 July 2011 and 1 August 2011 respectively), and *Chromis sp.* and Gobiidae, which were

significantly associated on 3 August 2011 (X = 0.44, p = 0.02) but not on 1 August 2011, when the

275 Gobiidae arrived in more than one cluster (Fig. 6).

Table 2. Association indices between temporal arrival numbers of different fish taxa. The

comparisons are made only between taxa on days for which both had a total catch of >24 individuals

278 (i.e. average of >1 individual per sampling period). The symbol N/A denotes where no comparisons

were possible because one species in each pair had fewer than 24 individuals. The date is indicated in

- the top line of the box, followed by the association index below with its significance level and
- 281 Dutilleul adjusted sample size in brackets. Significant associations are highlighted in bold.



	Chromis sp.	Apogon sp.1	Apogon sp.2	Apogon sp.3	Abudefduf sp.1	Pomacentrus sp.1	Corythoichthys sp.1	Synodontidae	Holocentridae	Significant associations per testable pairs
Chromis sp.	-	-	-	-	-	-	-	-	-	4/9
Apogon sp. 1	2011-08-03 0.37 (0.092; 24)	-	-	-	-	-	-	-	-	1/4
Apogon sp.2	2011-08-03 0.13 (0.56; 21.7)	2011-08-03 0.35 (0.14, 22.9)	-	-	-	-	-	-	-	0/4
Apogon sp.3	2011-08-01 0.71 (<0.001, 24)	2011-08-03 0.69 (0.004, 19.8)	2011-08-03 0.36 (0.16, 19.8)	-	-	-	-	-	-	2 / 4
Abudefduf sp.1	2011-07-24 0.45 (0.04, 22.8)	N/A	N/A	N/A	· ·	-	-	-	-	1/6
	2011-08-01 0.68 (<0.001, 22.9)									
Pomacentrus sp.1	2011-07-24 0.45 (0.04, 22.4)	N/A	N/A	N/A	2011-07-24 -0.12 (0.60, 23.7)	-	-	-	-	1/5
Corythoichthys sp.1	2011-07-24 0.26 (0.32, 24)	N/A	N/A	N/A	2011-07-24 0.54 (0.14, 11.1)	2011-07-24 0.01 (0.8, 21.7)	-	-	-	0/5
Synodontidae	2011-07-24 0.33 (0.24, 21.2)	N/A	N/A	N/A	2011-07-24- 0.10 (0.64, 21.6)	2011-07-24 —0.29 (0.08, 20.9)	2011-07-24 -0.18 (0.20, 21.7)	-	-	0/5
Holocentridae	2011-07-24 0.33 (0.16, 23.8)	N/A	N/A	N/A	2011-07-24 0.58 (0.06, 13)	2011-07-24 0.04 (0.82, 23.3)	2011-07-24 0.45 (0.12, 18.5)	2011-07-24 —0.21 (0.14, 23.1)	-	0/5
Gobiidae	2011-08-01 -0.01 (0.98, 19.8)	2011-08-03 0.35 (0.14, 22.9)	2011-08-03 -0.14 (0.62, 20.3)	2011-08-01 0.002 (0.90, 24)	2011-07-26 -0.16 (0.72, 20.2)	N/A	N/A	N/A	N/A	1/5
	2011-08-03 0.44 (0.04, 23.9)			2011-08-03 0.15 (0.52, 18.6)	2011-08-01 0.001 (0.90, 19.3)					

283 Discussion

284 To our knowledge, this is the first study to present high temporal resolution arrival numbers of 285 settlement stage fish. In this study clear temporal clustering was widespread among the ten most 286 abundant taxa over single night sampling periods, usually involving a single cluster for taxa identified 287 to genus level. Importantly, the arrival pattern in the light traps across the night often differed among 288 taxa, which suggests that different mechanisms could underlie their settlement timing. For the taxa 289 identified Pomacentridae taxa, 82-90% were caught between 19:00 and 22:00 on all sampling days at 290 Buoy 1, while the three taxa from the next most abundant family, Apogonidae, arrived later in the 291 evening, with 80-85% caught between 21:00 and 23:00 on 3 August but earlier for Apogonidae sp.3 292 on 1 August (19:00 - 22:00).

293 The fact that the Gobiidae were present in higher numbers at the site with lower flow could be a 294 strategy to increase their chances of settlement. Indeed, their smaller size compared to other taxa 295 collected is likely to result in slower swimming speeds (reviewed in Leis 2010). This would make 296 active swimming more energetically demanding due to the higher viscosity environment faced by 297 smaller fish (Bellwood and Fisher 2001) and the higher water flow, hence diminishing their settlement 298 success. Although larvae from the Gobiidae family did not appear to arrive at the traps in clusters, this 299 could be a result of the low taxonomic identification level. In fact, an outlier in Fig. 5b where 45 300 individuals (39% of the total Gobiidae caught on that sampling night) were caught in a single 15 min 301 sampling period, hints at possible high levels of clustering in a narrow time frame. A possible 302 explanation for this high catch could be that the Gobiidae were shoaling, as observed in laboratory 303 and field studies for larvae in this family (Breitburg 1989; Breitburg 1991). This may also be true for 304 the arrival of Synodontidae, which displayed no significant pattern of settlement but did recruit in 305 high numbers on a single sampling period on 24 July (Fig. 4c). However, these conclusions remain 306 speculative.

Comparisons among species forms an important part of this study. Setting aside taxa identified only to
family level, where lack of associations might be due to multiple species composition of the catches,
the arrival patterns of *Apogon* sp.2 and *Chorythoichthys* sp.1 were not similar to any of the other taxa.

310 There was also a lack of association between arrival patterns of two Pomacentrid species, *Abudefduf*311 sp.1 and *Pomacentrus* sp.1.

312 Settlement stage larvae of *Chromis* sp. have been observed to settle in small groups during the 313 daytime (Nolan 1975). Chromis atripectorialis improve orientation consistency, are able to more 314 accurately maintain a bearing, and swim faster when they school compared to movement as 315 individuals (Irisson *et al.* 2015). This could provide an important survival advantage to larvae that 316 school as this behaviour would enable them to reach a settlement site faster, therefore reducing the 317 time spent in the pelagic environment where they incur high mortality rates (Houde and Zastroe 318 1993). Persistent aggregations have been found to occur in the splitnose rockfish, Sebastes diploproa, 319 where 11.6% of settlement stage larvae were siblings (Ottmann et al. 2016). Interactions among group 320 members may play a role in maintaining these aggregations from spawning to settlement and could 321 provide the basis for clustering patterns observed in this study. It is important to note, however, that 322 there are a number of other mechanisms which could drive the observed clustering patterns, including 323 concentration of larvae in particular locations due to mesoscale eddies (Shulzitski et al. 2016). It is 324 also unclear how larvae would be able to maintain a school at night without being able to use vision to 325 see other group members, although other senses such as lateral line sensing (Faucher et al. 2010) or 326 vocal communication (Staaterman et al. 2014) may play a role.

327 There are insufficient sampling days in this study to determine whether different arrival patterns 328 observed among taxa are explained by random arrival of patches of larvae or driven by behaviours 329 aimed at improving settlement for a particular species. Why different taxa have different patterns of 330 arrival could, for example, be due to a trade-off between finding limited suitable settlement habitat 331 and avoiding predators. For *Corythoichthys* sp.1, that is widespread around the island and settle in 332 seagrass habitat between the reef crest and the shore (pers. obs.), avoidance of predators may be a 333 more important factor than limited habitat availability compared to species that recruit to specific 334 coral types like many pomacentrids. If this were the case, it might explain why they recruit only later 335 in the night when no light is available to predators (Johannes 1978; Dytham and Simpson 2007), and 336 the narrower time frame over which the recruitment occurred in this study (Fig.4a). The latter could

337	increase their chances of passing through the "wall of mouths" (Hamner et al. 1988) awaiting them at
338	the reef by achieving safety in numbers according to group theory (Bertram 1978).
339	This study furthers our understanding of the manner in which settlement stage fish recruit to the reef
340	in this critical transition phase, but cannot explain why different species display clustering patterns,
341	whether group behaviour mediates the temporal clustering or how and when the clustering is initiated.
342	Aggregations of larvae prior to settlement have been documented (Patterson et al. 2005), however,
343	whether this is due to environmental processes (e.g. eddies) that concentrate larvae in particular
344	locations or whether behaviour mediates the observed clustering patterns, will require further studies
345	that directly observe larvae during this critical life period
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