Acute and long-term effects of a single dose of MDMA on aggression in Dark Agouti rats

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Abstract

MDMA causes selective depletion of serotonergic terminals in experimental animals and the consequent decrease in synaptic 5-HT may, inter alia, increase impulsivity. To study the effects of MDMA upon brain function, the behaviour of male Dark Agouti rats exposed to MDMA (15 mg/kg i.p.), two 5-HT_{1B} agonists (CGS-12066A and CP-94,253, both 5 mg/kg i.p.) or saline were investigated in the resident-intruder test. Studies were performed in drug-naive rats and also in rats exposed to MDMA (15 mg/kg i.p.) 21 d earlier. In parallel experiments the functional neuroanatomy of MDMA effects were assessed using 2-deoxyglucose imaging of local cerebral metabolic rate of glucose utilization (LCMR_{Glu}) and neurotoxicity was assessed by measuring [³H]paroxetine binding. There was no significant difference in aggressive behaviour (biting, boxing, wrestling and their latencies) between drug-naive rats and rats previously exposed to MDMA 21 d earlier, despite reduced social behaviour, decreased LCMR_{Glu} in several brain areas involved in aggression, and reductions in paroxetine binding by 30-60% in the forebrain. CGS-12066A, CP-94,253 and acute MDMA produced marked decreases in aggressive behaviours, especially in biting, boxing and kicking found in drug-naive rats. In animals previously exposed to the drug, acute antiaggressive effects of MDMA were, in general, preserved as were MDMA-induced increases in LCMR_{Glu}. Our studies provide evidence that in the resident-intruder test, where social isolation is a requirement, aggressive behaviour and acute anti-aggressive effects of MDMA and 5-HT_{1B} receptor agonists remain intact 3 wk after a single dose of the drug despite significant damage to the serotonergic system.

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Introduction

The ring-substituted amphetamine derivative (\pm) -3,4-methylendioxymethamphetamine (MDMA; ecstasy) has become a widely abused psychoactive drug among young people and is popular for its mood-enhancing effects (Bond et al., 2004; Parrott, 2001; Steele et al., 1994). In the central nervous system (CNS), MDMA has characteristic, and well-documented two-phase effects upon monoaminergic systems. First, shortly after administration, there is an

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Tel.: +36-1-3915407 *Fax*: +36-1-3915408 *E-mail*: bag13638@mail.iif.hu acute release of serotonin (5-HT) (Green et al., 2003; Mechan et al., 2002; Shankaran and Gudelsky, 1998), dopamine (DA) and norepinephrine (Colado et al., 1999; Fitzgerald and Reid, 1990; Rothman et al., 2001; Yamamoto and Spanos, 1988). Second, in the brains of rodents and non-human primates, MDMA causes a long-term reduction in 5-HT, 5-hydroxyindolacetic acid (5-HIAA) concentration and 5-HT transporter density. Most investigators interpret the latter as a neurotoxic effect that involves the serotonergic axons and depletion of terminals (Battaglia et al., 1987; Colado et al., 1993; Green et al., 2003; Ricaurte et al., 2000; Schmidt, 1987).

There is strong evidence that 5-HT plays a crucial role in the regulation of impulsivity and aggression (Berman et al., 1997; Morgan, 1998), although other neurotransmitters, notably DA and GABA have also been linked to human and animal aggression (Miczek et al., 2002; Nelson and Chiavegatto, 2001). 5-HT release, or high 5-HT levels, are generally correlated with low aggressiveness (Nelson and Chiavegatto, 2001; van der Vegt et al., 2003). In contrast, serotonergic dysfunction, as defined by low cerebrospinal fluid (CSF) levels of 5-HT and 5-HIAA, has been associated with increased aggression (Clark and Neumaier, 2001; Scearce-Levie et al., 1999a).

Acute administration of MDMA has anti-aggressive effects both in humans and in mice (Curran et al., 2004; Miczek and Haney, 1994), presumably due to 5-HT release although increased impulsivity and aggression has been found amongst humans who report previous chronic heavy ecstasy use (Bond et al., 2004; Gerra et al., 1998; Morgan, 1998; Parrott et al., 2000). Although it is tempting to speculate that increased aggression in human users is a symptom of MDMAinduced neurotoxicity, the supporting evidence, whilst growing (Curran, 2000; Reneman et al., 2001; Ricaurte et al., 2000) is not yet totally compelling. It is possible that symptomatic neurotoxicity may be limited to heavy users, and/or to particularly susceptible individuals (McCann and Ricaurte, 1991), in which vulnerability may be genetically (Roiser et al., 2005) or environmentally determined. This may go some way towards explaining why MDMA-induced serotonergic neurotoxicity is not fully established in humans and shows great inter-individual variation. Nonetheless, it is perhaps timely to assess whether MDMA treatment in rats, within the dose range which causes a well-established long-term serotonergic deficit, is likely to elicit changes in aggressive behaviour and impulsivity which may parallel similar behavioural changes in humans.

The drug-induced long-term serotonergic loss may evoke functional changes to 5-HT receptors which, as well as other physiological functions in the brain, may play a role in modulating aggression and related behaviours. The 5-HT_{1B} receptor in particular is thought to have such a function, and a number of pharmacological studies have suggested that 5-HT_{1B} receptor activation reduces aggression (Clark and Neumaier, 2001; Muehlenkamp et al., 1995; Scearce-Levie et al., 1999a). Moreover, it is known that CGS-12066 and CP-94,253, two selective 5-HT_{1B} receptor agonists, have an anti-aggressive effect (Bell et al., 1995; Clark and Neumaier, 2001; Fish et al., 1999).

In our previous studies, we have reported that acute MDMA results in a generalized increase in metabolic activity in many brain areas, while in the longer term, chronic hypoactivity has been described (Balogh et al., 2004; Quate et al., 2004). These effects are more widespread than would be predicted solely on the

basis of serotonergic terminal distributions, reflecting the highly integrated nature of cerebral function, and may help in identifying brain areas that are involved in the control of aggressive behaviour both directly and indirectly. Despite the evidence of these and other studies on the acute effects, few studies have investigated the effects of MDMA on behaviour in animals previously exposed to the drug. In this study, we aimed to reassess previous results showing that acute MDMA decreases aggressive-type behaviour, and to test the hypothesis that 3 wk after a single, neurotoxic dose of MDMA, there would be a significant increase in aggression and impulsivity, an attenuation of the acute effects of further MDMA exposure, and changes in $5-HT_{1B}$ receptor function.

To these ends, the aim of this work was to examine the acute and long-term effects of a single dose of MDMA upon aggression, in drug-naive rats and rats previously exposed to the same dose 3 wk earlier, using the resident-intruder aggression test. A second set of experiments was designed to assess possible MDMA-evoked changes in 5-HT_{1B} receptor function in rats previously exposed to the drug, by measuring the acute effects of the 5-HT_{1B} selective agonist CGS-12066 (and its congener CP-94,253) upon aggression. In parallel to the behavioural studies we examined the acute and long-term effects of MDMA upon brain metabolism, in drug-naive and MDMA-pretreated animals, by measuring the local cerebral metabolic rate of glucose utilization (LCMR_{Glu}), in brain areas relevant to the regulation of aggression. In addition, [³H]paroxetine binding to 5-HT transporter sites was measured by autoradiography in order to provide evidence for long-term serotonergic changes in forebrain areas. These transporter sites are localized to 5-HT terminals and reductions in radioligand binding have been shown to correlate with anatomical evidence of terminal depletion (Battaglia et al., 1987).

Methods

Animals

All animal experiments were carried out in accordance with the European Communities Council Directive of 24 November 1986 (86/609/EEC) and the National Institutes of Health 'Principles of Laboratory Animal Care' (NIH Publication no. 85-23, revised 1985). In addition, specific national laws were adhered to [the Hungarian Governmental Regulation about animal studies of 31 December 1998 and the United Kingdom Animals (Scientific Procedures) Act, 1986]; permission was also obtained from local ethical committees.

Male Dark Agouti (DAg) rats (Harlan, Olac Ltd, Shaw's Farm, Blackthorn, Bicester, Oxon, UK, aged 4-5 wk and weighing 50-80 g upon arrival), were used in the experiments. Animals were housed under standard 12-h light-dark conditions (lights on 06:00 hours), with standard rat chow and water freely available (Balogh et al., 2004). Ambient temperature in the animal house was maintained at 21+1 °C. MDMA or saline pretreatment was administered at the age of 7 wk and the resident-intruder test was carried out 3 wk later when the resident animals were 10 wk old (weight 215 ± 2.50 g) and the intruders were 5–6 wk old (weight 80.65 ± 1.35 g). To establish the resident status of the rats, these animals were housed singly for the final 2 wk prior to the test (Stuerenburg et al., 2002). The same conditions were replicated as closely as possible for animals used in the measurement of LCMR_{Glu}, with the exception that animals were group-housed until the day of the final experiment.

Drugs

(±)3,4-methylenedioxymethamphetamine hydrochloride (MDMA, certified reference compound, purity >99.5%) was provided by Sanofi-Synthelabo-Chinoin, Budapest, Hungary. The MDMA was dissolved in saline and animals injected with a dose equivalent to 15 mg/kg free base. 5-propoxy-3-(1,2,3,6-tetrahydro-4-pyridinyl)-1H-pyrrolo[3,2-b]pyridine hydrochloride (CP-94,253; Tocris, Bristol, UK) was dissolved in distilled water at a dose of 5 mg/kg. 7-trifluoromethyl-4(4-methyl-1-piperazinyl)-pyrrolo[1,2-a]quinoxaline maleate (CGS-12066A; Sigma RBI, Steiheim, Germany) was dissolved in 10% HPCD (2-hydroxypropyl- β cyclodextrin; Fluka, Buchs, Switzerland).

Control rats received an injection of 0.9% NaCl (saline) or 10% HPCD (vehicle for CGS-12066A) in a volume of 1 ml/kg. All drugs were administered intraperitoneally (i.p.).

Experimental procedures

Resident-intruder test

Resident rats were pretreated with MDMA (15 mg/kg) or saline on a single occasion, 3 wk prior to the acute drug administrations (MDMA, CGS-12066, CP-94,253), which occurred 20 min prior to the introduction of the intruder to the home cage of the resident rat (Fish et al., 1999). Treatment groups consisted of 6–10 animals in the resident-intruder test. Each rat was tested only once.

The resident-intruder aggression test was conducted by introducing a small (intruder) rat into the cage of a larger (resident) rat (Muehlenkamp et al., 1995). The behaviour of the animals was recorded on videotape for 15 min immediately after the introduction of the smaller animal into the cage (Holmes et al., 2002). The record was scored manually at a later time using a computer program designed for the purpose, as previously described by us (Kantor et al., 2000). The data were calculated from the raw scores made by a trained observer and these were included in the results.

The following behavioural elements were scored: biting, boxing, kicking were scored and classified as impulsive aggressive behaviours, and the incidence of each parameter, as well as their sum were calculated. The latency of each behaviour (the time elapsed until the first event) was also recorded. An additional parameter, namely the latency of first impulsive aggressive behaviour (biting, boxing, and kicking) was also recorded. This was defined as the time elapsed until the appearance of the first element irrespective of whether this constituted biting or boxing or kicking. Duration and latency of wrestling were also scored and calculated. Within this parameter pushing and dominant posture were also included.

Duration of anogenital sniffing, grooming of the intruder, over-crawling and under-crossing were included and summarized within the social behaviours. From these behaviours, grooming the intruder was also analysed separately, scored and calculated.

The motor activity of each resident animal was observed and scored in each of the three 5-min periods and a single numerical score was assigned to each rat, for each period. Scoring was based on a five-point scale (0–4); 0 indicating lack of movement, and 4 indicating continuous motor activity. The three scores from each of the periods were summed, yielding a score in the range 0–12 for each resident animal. Exploratory behaviour was scored in a similar way to locomotor activity, using a five-point (0–4) scale. Scoring recorded the frequency of the following events: rearing, sniffing, exploration of the home cage.

For statistical analysis, STATISTICA 6.0 (Statsoft Inc., Tulsa, OK, USA) and GraphPad Prism (GraphPad Software Inc., San Diego, CA, USA) software were used. Tests for homogeneity of variances indicated that variances for duration of wrestling, social behaviour and grooming were not homogenous and thus, the Kruskal–Wallis non-parametric test was performed for these parameters while incidences and latencies of events, scores of motor activity and exploratory behaviour were analysed using non-parametric tests. Where statistical differences were detected, further comparisons were performed by Mann–Whitney *U* tests. For the analysis of pretreatment × treatment interactions two-way analysis of variance (ANOVA) was used for all parameters.

Measurement of LCMR_{Glu}

On the day of the experiment, 21 d after pretreatment with either MDMA or saline, rats were anaesthetized with halothane (maintained at 1% in a gas mixture of 70% nitrous oxide, 30% oxygen) and polythene cannulae were inserted into both femoral arteries (to allow sampling of arterial blood and monitoring of arterial blood pressure), and both femoral veins (for the injection of radiolabelled tracers and barbiturate at the termination of the experiment). An additional cannula was inserted i.p. via a trocar and held in place with a fine suture to allow the i.p. injection of MDMA. The incision sites were infiltrated with local anaesthetic and the wounds sutured closed. A loose-fitting, individually moulded plaster cast was applied around the hindquarters and pelvis and secured to a weighted platform. Thus, lightly restrained and supported, a rectal temperature probe was inserted and anaesthesia was discontinued. Animals were allowed to recover from anaesthesia for at least 2 h prior to further experimental manipulation.

The measurement of LCMR_{Glu} was performed using the fully quantitative [14C]-2-deoxyglucose (Sokoloff et al., 1977) autoradiographic technique. The measurement protocols were in complete accordance with the methodology as originally published (Sokoloff et al., 1977) and as previously detailed from this laboratory (Kelly et al., 1995; Quate et al., 2004). On the final experimental day, rats from both pretreatment groups were injected with MDMA (15 mg/ kg; saline pretreatment+acute MDMA group and MDMA pretreatment+acute MDMA) or saline as control (saline group) via the in-dwelling i.p. cannula. Fifteen minutes later, the measurement of LCMR_{Glu} was initiated with the intravenous injection of [14C]-2deoxyglucose (30 μ Ci in 0.75 ml saline) administered at a constant rate over the first 30 s of the experiment. Over the subsequent 45 min, a total of 14 timed arterial blood samples were collected at pre-determined intervals which were centrifuged in order to separate the plasma. Aliquots of plasma were removed for the determination of plasma ¹⁴C concentrations (20 μ l) and glucose levels $(10 \,\mu l)$ by liquid scintillation analysis and semi-automated glucose oxidase assay (Beckman, High Wycombe, Bucks, UK) respectively. At the end of the measurement period the rats were killed with a rapid intravenous injection of sodium pentobarbitone and the brains were immediately dissected out intact, and rapidly frozen in pre-cooled 2-methylbutane

(-45 °C). Frozen brains were mounted onto specimen holders and stored at -70 °C until sectioned in a cryostat for the preparation of autoradiograms.

Sections adjacent to those used for autoradiographic measurement of LCMR_{Glu} were thaw-mounted onto gelatine-covered glass slides and stored at -70 °C for subsequent [³H]paroxetine autoradiographic binding analysis of 5-HT uptake sites according to the protocol described by De Souza and Kuyatt (1987) but with the addition of two 5-min prewashes in buffer [50 mM Tris-HCl, 120 mm NaCl, 5 mm KCl (pH 7.7) at 23 $^\circ\text{C}]$ to remove ¹⁴C tracers from the tissue (De Souza and Kuyatt, 1987; Sharkey et al., 1991). Sections were then incubated in the same buffer containing a saturating concentration (250 pM) of [³H]paroxetine (specific activity 23.1 Ci/mmol; New England Nuclear DuPont, Stevenage, UK). Non-specific binding was defined in adjacent sections by [3H]paroxetine binding in the presence of 4 μ M citalopram. Following incubation, the sections were washed in buffer, dipped in deionized water and rapidly dried under a stream of cold air. The slides, together with a set of [3H]-containing standards (Amersham Microscales, Amersham, Bucks, UK) were apposed to X-ray film (Amersham Hyperfilm) in a light-tight cassette and stored at -70 °C for 4–6 wk. The films were processed according to the manufacturer's instructions.

Analysis of [¹⁴C]-2-deoxyglucose and [³H]paroxetine autoradiograms was performed using a computer-based image analysis system (MCID/M5+). Local tissue isotope concentrations were derived from the optical density of autoradiographic images of brain tissue, relative to appropriate [¹⁴C]- or [³H]containing standards. LCMR_{Glu} was calculated using the operational equation for the technique (Sokoloff et al., 1977), and radioligand binding was calculated from the tissue concentrations and the specific activity of the [³H]paroxetine. Data were analysed using Student's *t* test for grouped data with acceptable levels of significance set at *p* < 0.05.

Results

Long-term effects of MDMA

In rats exposed to MDMA 3 wk prior to the experiment there were no significant differences in the incidence of biting, boxing, the sum of biting, boxing and kicking or in the latencies of the three combined measures, and neither was there any difference in the duration of wrestling, or its latency compared to the control, drug-naive group (Figure 1). There was, however, a significant increase in kicking (H=18.912,



Figure 1. Acute effects of MDMA (15 mg/kg) upon incidence of biting (a), boxing (b), kicking (c), total incidence (d) and latency (e) of biting, boxing, kicking, duration (f) and latency (g) of wrestling, duration of social behaviour (h) and grooming (i) in drug-naive rats and in rats exposed to MDMA 3 wk earlier. Each group consisted of seven animals. Pretreatment × treatment interactions of ANOVA are significant where noted. * Significantly different from the appropriate control with the same pretreatment (p < 0.05). # Significantly different from the saline-pretreated group with the same acute treatment (p < 0.05). Sal, Saline.



Figure 2. Acute effects of MDMA (15 mg/kg) on motor activity and exploratory behaviour in drug-naive rats and in rats exposed to MDMA 3 wk earlier. Each group consisted of seven animals. * Significantly different from the appropriate control with the same pretreatment, (p < 0.05). Sal, Saline.

p < 0.05, Figure 1) and a small decrease in grooming and social behaviour (H = 24.473, p < 0.05 and H = 24.340, p < 0.01, for grooming and social behaviour respectively; Figure 1). Locomotor activity and exploratory behaviour were also unchanged in rats pretreated with MDMA (Figure 2).

Acute effects of MDMA in drug-naive rats and in rats previously exposed to MDMA

In drug-naive rats, acute MDMA treatment caused massive decreases in all measures of aggressive behaviour, namely, incidence of biting (H=12.749, p<0.01), boxing (H=19.829, p<0.001), kicking (H=18.912, p>0.05), and both the sum (H=21.602, p<0.001) and latency of the three measures combined (H=15.832, p<0.001). The duration of wrestling (H=21.237, p<0.01) was also reduced and subsequently, its latency was increased (H=20.350, p<0.01; Figure 1). Furthermore, a marked reduction

was also observed in other social behaviours (H=24.340, p<0.001) and grooming (H=24.473, p<0.001)p < 0.001, Figure 1) When young intruder animals were placed into the residents' home cages, both locomotor activity and exploratory behaviour of salinepretreated resident rats was markedly increased. This was not exclusively restricted to exploration of the intruder animal, but also the home cage itself was intensively explored. Increases in locomotor activity (H=20.541, p>0.05) and exploratory behaviour (H=20.957, p>0.05, Figure 2) were attenuated in saline-pretreated animals after acute exposure to MDMA. In the MDMA-pretreated rats, all acute effects of MDMA were preserved (Figures 1 and 2). The pretreatment × treatment interaction was significant only for the duration of social behaviours ($F_{1,24} = 6.394$, p = 0.0008) and grooming ($F_{1, 24} = 14.824$, p = 0.018).

Acute effects of CGS-12066 and CP-94,253 in drug-naive rats and in rats previously exposed to MDMA

Acute treatment with CGS-12066 reduced the incidence of biting (H=8.029, p<0.01) and boxing (H=12.372, p<0.01), the sum of biting, boxing and kicking (H=15.792, p<0.01), and the duration of wrestling (H=8.298, p<0.01) in drug-naive animals (Figure 3). Latencies of biting, boxing and kicking (H=12.022, p<0.01) were increased by the drug. Interestingly, the effects of the drug on biting, boxing and kicking were more robust than the effects upon wrestling. The compound also significantly decreased grooming (H = 11.617, p < 0.01) and the sum of all nonaggressive social behaviour (H = 6.107, p < 0.05; Figure 3). In contrast to the effects of acute MDMA, CGS-12066 significantly increased the motor activation elicited by the presence of the intruder (H=14.938,p < 0.01; Figure 4) and exploratory behaviour was also increased (H = 22.471, p > 0.05). The locomotor scores presented here reflect the time spent in locomotor activity. It was observed, however, that all motor activities, including exploration, social behaviour and different manifestations of aggressive behaviour were all performed at a noticeably increased speed in the presence of CGS-12066.

In rats previously exposed to MDMA, the acute effects of CGS-12066 were preserved with respect to most parameters (biting, boxing, duration of wrestling and grooming) although some of these, such as the effects on kicking, the sum of biting, boxing and kicking, as well as their latencies were more weakly expressed (Figure 3). In contrast, CGS-12066 treatment enhanced locomotor activity (H=14.938, p <0.05)



Figure 3. Acute effects of CGS-12066 (5 mg/kg) upon incidence of biting (a), boxing (b), kicking (c), total incidence (d) and latency (e) of biting, boxing, kicking, duration (f) and latency (g) of wrestling, duration of social behaviour (h) and grooming (i) in drug-naive rats and in rats exposed to MDMA 3 wk earlier. Each group consisted of 6–10 animals [Sal + Sal (n = 8), Sal + CGS (n = 10), MDMA + Sal (n = 6), MDMA + CGS (n = 10)]. * Significantly different from the appropriate control with the same pretreatment (p < 0.05). # Significantly different from the saline-pretreated group with the same acute treatment (p < 0.05). Sal, Saline; Veh, vehicle.



Figure 4. Acute effects of CGS-12066 (5 mg/kg) on motor activity and exploratory behaviour in drug-naive rats and in rats exposed to MDMA 3 wk earlier. Each group consisted of 6–10 animals [Sal+Sal (n=8), Sal+CGS (n=10), MDMA+Sal (n=6), MDMA+CGS (n=10)]. * Significantly different from the appropriate control with the same pretreatment (p <0.05). # Significantly different from the saline-pretreated group with the same acute treatment, (p <0.05). Sal, Saline.

and exploratory behaviour (H=22.471, p<0.001) in MDMA-pretreated animals (Figure 4). Significant pretreatment × treatment interaction was found in exploratory behaviour ($F_{1,32}$ =4.947, p=0.033).

The effects of CP-94,253 were, in general, comparable to those of CGS-12066, with similar interactions with MDMA pretreatment (data not shown).

Acute and long-term effects of MDMA on LCMR_{Glu}

The effects of acute MDMA on LCMR_{Glu} in those brain areas thought to be involved in aggression were measured in both drug-naive rats and in rats treated with MDMA 3 wk earlier. In rats pretreated with MDMA, and treated acutely with saline, there was a general trend towards decreased LCMR_{Glu} (Table 1), ranging from -3% in central amygdala (not significant) to -25% in medial amygdala (p < 0.05). Other significant decreases were observed in cingulate cortex, lateral septal nucleus, mediodorsal thalamic nucleus, periaqueductal grey matter, locus coeruleus and median raphe nucleus. In two areas of the brain, the hypothalamic attack area and the arcuate nucleus, LCMR_{Glu} was found to be increased in MDMA-pretreated rats, but these effects were small (9% and 6% respectively) and not statistically significant.

In keeping with previous observations, acute treatment with MDMA resulted in widespread increases in LCMR_{Glu} in drug-naive rats (Figure 5). The only exceptions were found in the hypothalamic attack area (-9%), anterior hypothalamic area (-4%) and median raphe (-5%) but the decreases in LCMR_{Glu} in these areas were relatively small and not significant. However a more substantial decrease in the cingulate cortex (-20%) was significantly different from that in rats treated acutely with saline.

The increases in LCMR_{Glu} elicited by acute MDMA in rats previously exposed to the drug were more widespread, and included areas in which the drug had been shown to decrease LCMR_{Glu} in previously drugnaive rats (most notably median raphe, anterior hypothalamus and cingulate cortex). Only in the hypothalamic attack area were decreases in LCMR_{Glu} (of similar magnitude) found in response to acute MDMA, irrespective of pretreatment. Elsewhere in the brain, there was evidence that the response to acute MDMA was potentiated by MDMA-pretreatment (Figure 5). This potentiation was most marked in the lateral septal nucleus (+30% vs. +70%) and locus coeruleus (+15% vs. +55%).

Long-term effects of MDMA on [³H]paroxetine binding

[³H]paroxetine binding was measured in brain tissue from animals exposed to saline or MDMA 3 wk previously (Table 2). No significant effects of MDMA were observed in the median raphe nucleus (2%), although significant reductions in binding were observed in the piriform cortex (-38%), cingulate cortex (-59%), septal nucleus (-20%), amygdala (-31%).

Discussion and conclusion

Acute MDMA treatment decreased aggressive-type behaviour as measured with the resident-intruder test, and the anti-aggressive effects of 5-HT_{1B} agonists were similar to those of MDMA. Acute MDMA treatment

Brain area	LCMR _{Glu} (µmol/100 g . min)		
	Saline pretreatment + acute saline	MDMA pretreatment + acute saline	% change
Median raphe nucleus	87±3	$70 \pm 2^{*}$	-20
Locus coeruleus	62 ± 2	$49 \pm 2^{*}$	-21
Periaqueductal grey	60 ± 2	$48 \pm 2^{*}$	-20
Mediodorsal thalamic nucleus	101 ± 4	83±3*	-18
Central amygdala	68 ± 7	66 ± 3	-3
Medial amygdala	53 ± 5	$40 \pm 2^{*}$	-25
Arcuate nucleus	54 ± 2	57 ± 1	6
Hypothalamic attack area	55 ± 2	60 ± 4	9
Anterior hypothalamic area	52 ± 1	49 ± 2	-6
Medial preoptic area	50 ± 3	45 ± 1	-10
Bed nucleus of the stria terminalis	52 ± 2	45 ± 2	-13
Lateral septal nucleus	60 ± 3	$50 \pm 2^*$	-17
Medial septal nucleus	48 ± 2	44 ± 2	-8
Piriform cortex	52 ± 4	48 ± 1	-8
Cingulate cortex	113 ± 4	$91 \pm 2^*$	-19

Table 1. Local cerebral metabolic rate of glucose utilization (LCMR_{Glu}) in saline and MDMA (15 mg/kg i.p.) pretreated animals

Data presented as mean ± S.E.M. Percentages are differences from saline control.

* Significantly different from saline control group (p < 0.05). Each group consisted of seven animals.



Figure 5. Acute effects of MDMA upon LCMR_{Glu} in drugnaive animals and in animals previously exposed to MDMA. Data are presented as mean percentage change from control \pm S.E.M. * Significantly different from saline pretreated animals (p < 0.05, Mann–Whitney U test). Each group consisted of seven animals. **•**, MDMA pretreatment + acute MDMA; \Box , saline pretreatment + acute MDMA.

increased brain metabolism in several areas including those involved in the regulation of aggression. Significant long-term decreases in paroxetine binding **Table 2.** [³H]Paroxetine binding in animals exposed to either saline or MDMA (15 mg/kg) 3 wk previously

Brain area	Saline	MDMA	Lesion %
Neocortex			
Piriform cortex	64 ± 5	$40\pm7^*$	-38
Cingulate cortex	66 ± 7	$27\pm4^*$	-59
Septal nucleus	64 ± 4	$51 \pm 3^{*}$	-20
Amygdala	210 ± 15	$144\pm10^*$	-31
Median raphe nucleus	170 ± 16	173 ± 21	2

Data are presented as mean binding density (fmol/mg tissue)±s.e.m.

Each group consisted of seven animals.

*Significantly different from saline control group (p < 0.05).

and brain metabolism were found in many brain areas, 3 wk after MDMA administration, although at this time-point aggressive-type behaviour was unaltered. Contrary to expectation, increases in brain metabolism induced by acute MDMA were not diminished in rats previously exposed to the drug. The anti-aggressive effects of 5-HT_{1B}-agonists in MDMA-pretreated rats were, in general, preserved and only modest changes were observed.

Acute effects of MDMA on aggression and LCMR_{Glu}

The data from the present study support previous observations that acute MDMA treatment decreases aggressive-type behaviour in rats, as measured in the resident-intruder test (Miczek et al., 1994; Navarro et al., 1999; Verheyden et al., 2002), as well as in human abusers (Curran et al., 2004). However, in the present study there were some differences in the effect of MDMA upon the separate parameters of aggression, for example in the frequency of biting, boxing and kicking. Closer examination of the data reveals that this is due, in the main, to differences in the manifestation of attack between individuals, with some animals preferring to kick, while others showed a greater tendency to box or bite.

Interestingly, changes were also found in other parameters not closely related to aggression. Introducing an intruder to the home cage usually results in marked increases in locomotor activity and exploratory behaviour from the resident animal, but this hyperlocomotion was significantly attenuated by acute MDMA treatment (Figure 2). Similarly, significant decreases were found in social behaviour (Figure 1). Thus, it appears that if the resident animal is treated with MDMA in advance of the introduction of the intruder, it is less affected by the presence of the intruder animal. In parallel studies on animals outwith the behavioural paradigm, i.e. where no stimulus or target for aggressive behaviour is available, MDMAinduced increases in local cerebral metabolic rate were found in several brain regions that regulate (either activate or inhibit) aggression, and also in other regions that regulate many other physiological functions. This is perhaps not surprising given the widespread anatomical distribution of serotonergic projections, and the possible involvement of both dopaminergic and noradrenergic systems in the pharmacological response to MDMA.

The long-term effects of MDMA on aggression, LCMR_{Glu} and paroxetine binding

In our study, MDMA was administered to Dark Agouti rats at a dose of 15 mg/kg. Although there are considerable strain differences in the susceptibility to MDMA-induced long-term serotonergic depletion, many authors suggests that a single, 15 mg/kg injection of MDMA in rats may more accurately model the possible exposure to the drug in humans, particularly when pharmacokinetic scaling is applied to body size and basal metabolic rate (Colado and Green, 1995; Malpass et al., 1999). In contrast, it is less likely that previously used models in which, typically, rats are treated with a higher dose (20 mg/kg) twice a day for four consecutive days adequately reflect human experience. Moreover, Dark Agouti rats show decreased microsomal CYP2D6 isoenzyme activity, which plays an important role in the metabolism of MDMA. As a consequence, the Dark Agouti rat is considered to be a poor or moderate metabolizer of the drug with MDMA pharmacokinetics that are similar to those which may be found in particularly vulnerable human users (Balogh et al., 2004; O'Shea et al., 1998; Quate et al., 2004). Such users might represent a genetically defined human sub-population in which clinical complications are more likely to occur.

In this study, significant decreases in paroxetine binding of between 30% and 60% were found in the neocortex, septal nucleus and amygdala, but not in the median raphe nucleus, 3 wk after a single exposure to MDMA. This finding is totally in agreement with previous data, which showed that a similar MDMA treatment caused long-term serotonergic depletion in the forebrain of Dark Agouti rats (Colado et al., 1998; O'Shea et al., 1998). Of particular interest in the context of our behavioural data, is the observation of decreased binding in some areas of the brain known to be involved in the regulation of aggression and impulsivity. However, despite the deleterious effect of MDMA on the serotonergic system, and previous evidence that depletion of 5-HT in the amygdala facilitates aggressive behaviour (Morgan, 2000), the present study indicates that manifestations of aggressive behaviours was not abnormal in rats pretreated with MDMA 3 wk earlier. For example the animals did not exhibit attacks to vulnerable areas on the target intruder, which is considered a pathological form of aggression (Halasz et al., 2002a,b). There was, however, evidence from our metabolic mapping studies that, in the resting state at least, MDMA pretreatment did result in altered functional activity in some elements of the amygdala, indicating that the treatment was not without effect.

The lack of evidence for increased aggression in this test may be explained in many ways. First, the level of aggression exhibited by rats in this test is normally relatively high. It could be difficult to detect further small increases in an already high aggressive state, which is described as an essential factor in this test paradigm (Navarro and Maldonado, 1999). Furthermore, chronic social isolation, which is necessary to confer resident status, has been reported to decrease 5-HT levels in forebrain areas in the long-term (Dalley et al., 2002; Fulford and Marsden, 1998; Robbins et al., 1996), and indeed this could be one explanation for the increased baseline aggression in the resident-intruder albeit in the absence of aggression-inducing stimuli. The decrease in metabolism was found in several brain areas, including those thought to mediate aggressive behaviour, and also in those that inhibit aggression. It is conceivable that the integration of these inhibitory and disinhibitory effects could confound our measures of aggression, or alternatively represent a compensatory mechanism to normalize function.

Human studies that have sought to identify MDMA-induced abnormalities are difficult to interpret due to inherent methodological complications with retrospective studies. These complications include inaccurate reporting of the doses MDMA taken, the duration and frequency of drug use, and nonreporting of the concurrent use of other substances, licit or illicit (Green et al., 1995; Morgan, 2000). However, some studies have demonstrated an increase in aggression and impulsivity in heavy ecstasy users (Bond et al., 2004; Gerra et al., 1998, 2000), which may suggest that in the long term MDMA might increase aggression in humans. However, no significant increase was found in light/moderate users (Morgan, 1998; Parrott, 2000) and earlier studies in humans have found that, although aggression increases 4 d after exposure to MDMA, there is no significant increase in aggressive behaviour 7 d after the exposure (Curran et al., 2004). Only McCann et al. (1994) reported that heavy ecstasy users had lower aggression than control participants.

Effects of a previous MDMA treatment on MDMA-induced increases in LCMR_{Glu} and on the behavioural effects of acute MDMA, CGS-12066 and CP-94,253 administration

Our results indicate that all of the acute behavioural effects of MDMA were preserved when a neurotoxic MDMA pretreatment was applied 3 wk before the test. The MDMA-induced depletion of serotonergic terminals, for which our [³H]paroxetine-binding data provide positive evidence in this study, could be assumed to reduce the availability of releasable 5-HT, and might be expected to result in an attenuated anti-aggressive response to acute MDMA treatment. However, these anti-aggressive effects or any possible changes caused by 5-HT depletion, may possibly be masked by the general inhibitory behaviour which is unaffected by neurotoxic MDMA pretreatment.

Acute MDMA caused a significantly greater increase in brain metabolism in pretreated rats compared to saline-treated animals. This could be explained in part by the lower baseline activity in the brain areas examined (as stated previously), so that metabolic rates after acute MDMA administration were largely similar in drug-naive rats and rats previously exposed to the drug. However, this cannot explain the divergent response found, for example in the cingulate cortex, where an MDMA-induced decrease in drug-naive rats is replaced by a significant MDMA-induced increase in rats previously exposed to the drug.

The acute anti-aggressive effects of 5-HT_{1B} agonists in drug-naive rats were more selective than those of MDMA, namely, changes in impulsive and aggressive behaviours such as biting, boxing, kicking were very marked. Furthermore, the effects of 5-HT_{1B} agonists on motor activation and exploratory behaviour induced by the presence of the intruder animal were quite different from those of MDMA, such that 5-HT_{1B} agonists caused increases in this motor activation, while MDMA caused a reduction. 5-HT_{1B} agonists evoked generally similar effects in MDMA-pretreated rats, compared to drug-naive rats. These data do not, therefore, indicate a significant sensitization of 5-HT_{1B} receptor function as a consequence of a long-term serotonergic depletion. A possible explanation is that despite the partial axonal loss, some, or all, of the serotonergic axons and terminals involved in antiaggressive behaviour may remain intact. Furthermore, 5-HT_{1B} receptors involved in this action may be located post-synaptically (Barnes and Sharp, 1999; Clark and Neumaier, 2001; Scearce-Levie et al., 1999b) or are heteroreceptors, located not only on serotonergic terminals, but also on terminals where they might inhibit the release of other neurotransmitters including GABA, acetylcholine or glutamate (Clark and Neumaier, 2001; Sexton et al., 1999). Alternatively, there is the possibility that non-serotonergic mechanisms are also involved in the acute effects of MDMA upon aggression, including dopamine, glutamate or GABA (Sexton et al., 1999). It is also possible that anxiety, reactivity and impulsivity confound the overt behavioural expression of MDMA-induced effects, as has previously been described after partial 5-HT depletion (Green et al., 2003; Harro, 2002). There was, however, a difference in drug-induced motor activation. The increase in locomotor activity and exploration was significantly enhanced in MDMA pretreated rats, suggesting that the involvement of 5-HT_{1B} receptors in locomotor activation differs from that in the regulation of aggression and impulsivity.

In conclusion, acute MDMA administration abolished aggressive behaviour in the resident-intruder test as expected. Unexpectedly, MDMA pretreatment and the consequent depletion of serotonergic terminals did not increase aggression. This may be explained by the confounding effects of social isolation, which is known to induce decrease in brain 5-HT concentration. Thus, the lack of increase in aggression may be explained by the neurochemical effects of social isolation that might be similar to the long-term effects exerted by MDMA. This suggestion is also supported by the fact that a previous exposure to MDMA failed to affect 5-HT_{1B} receptor function in these animals.

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Statement of Interest

None.

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