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Multi-Ethnic Meta-Analysis Identifies *RAI1* as a Possible Obstructive Sleep Apnea Related Quantitative Trait Locus in Men

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intellectual content.

Abstract

Obstructive sleep apnea (OSA) is a common heritable disorder displaying marked sexual dimorphism in disease prevalence and progression. Previous genetic association studies have identified a few genetic loci associated with OSA and related quantitative traits, but they have only focused on single ethnic groups and a large proportion of the heritability remains unexplained. The apnea hypopnea index (AHI) is a commonly used quantitative measure characterizing OSA severity. Since OSA differs by sex, and the pathophysiology of obstructive events differ in rapid eye movement (REM) and non-REM (NREM) sleep, we hypothesized that additional genetic association signals would be identified by analyzing the NREM/REM-specific AHI and by conducting sex-specific analyses in multi-ethnic samples. We performed genome-wide association tests for up to 19,733 participants of African-, Asian-, European-, and Hispanic/Latino-American ancestry in seven studies. We identified rs12936587 on chromosome 17 as a possible quantitative trait locus for NREM AHI in men ($N = 6,737$; $P = 1.7 \times 10^{-8}$), but not in women ($P = 0.77$). The association with NREM AHI was replicated in a physiological research study ($N = 67$; $P = 0.047$). This locus overlapping the *RAI1* gene and encompassing genes *PEMT1*, *SREBF1* and *RASDI*, was previously reported to be associated with coronary artery disease, lipid metabolism, and implicated in Potocki-Lupski Syndrome and Smith-Magenis Syndrome, which are characterized by abnormal sleep phenotypes. We also identified gene-by-sex interactions in suggestive association regions, suggesting that genetic variants for AHI appear to vary by sex, consistent with the clinical observations of strong sexual dimorphism.

Keywords: obstructive sleep apnea, genetics, genome-wide association studies, multi-ethnic, sexual dimorphism.

Introduction

Obstructive Sleep Apnea (OSA) is a complex chronic condition that affects more than 10% of the population, and is associated with cardio-metabolic and behavioral morbidity (1-3). The prevalence of OSA is particularly high in minority racial/ethnic groups such as those with African-, Asian- and Hispanic ancestry (4-7). Moreover, OSA is approximately 3-fold more prevalent in men as compared to women (8). In women, OSA severity is less likely to worsen in the supine compared to other sleeping positions (9) and more likely to worsen in rapid eye movement (REM) sleep, when neuromuscular tone and chemoreflexes are reduced (9, 10). These differences have been attributable to sex differences in airway-collapsibility, related to both differences in anatomy and respiratory chemosensitivity (11). An increase in OSA severity in women after menopause also suggests a role for sex hormones in influencing this disorder (12).

The severity of OSA is most often characterized by the apnea hypopnea index (AHI), defined as the number of apnea and hypopnea events per hour of sleep. AHI levels are highly heritable in African-Americans and European-Americans, with 30 to 40% of the variance explained by genetic factors (13, 14). Previous genetic studies have identified several genetic variants associated with AHI, although these findings were based on modest sample sizes or single ethnic groups, and largely have not been replicated across populations (15-18).

Large-scale genome-wide association studies (GWAS) have identified sexual dimorphism in genetic loci for traits associated with OSA, such as body fat distribution, particularly waist circumference and waist-to-hip ratio, each adjusted for body mass index (BMI) (19, 20).

Furthermore, measures of adiposity such as waist phenotypes have been shown to be regulated

by sexually dimorphic genes (19, 21). Animal models suggest that both gonadal hormones and X chromosome dose influence lipid levels (22). Despite strong clinical and epidemiological evidence for sex differences in OSA, prior genetic association studies were not sufficiently powered to study consistent sex differences in OSA in multi-ethnic samples (15-17).

We conducted genome-wide association studies in multi-ethnic samples from 7 cohorts to identify genetic variants with sex-specific association for AHI. Given differences in the physiological bases for OSA in REM and non-REM sleep (23), we performed analyses for AHI calculated for each sleep state (REM; non-REM). Although BMI is a significant risk factor for OSA, only 40% of the genetic variance for OSA is shared with BMI (14). Therefore, we adjusted for BMI in order to discover genetic loci acting independently of BMI, which may provide insights into novel etiological mechanisms. We focused on association signals that show concordant direction of effects across African-, Asian-, Hispanic/Latino- and European-Americans through BMI-independent pathways.

Materials and Methods

Study Subjects

We included seven cohorts in the discovery analyses: the Atherosclerosis Risk in Communities Study (ARIC, n=1,463 European-Americans), the Cleveland Family Study (CFS, n=731 African-Americans and 702 European-Americans), the Framingham Heart Study (FHS, n=646 European-Americans), the Hispanic Community Health Study / Study of Latinos (HCHS/SOL, n=11,317 Hispanic/Latino-Americans), the Multi-Ethnic Study of Atherosclerosis (MESA, n=490 African-Americans, 228 Asian-Americans, 707 European-Americans, and 458 Hispanic/Latino-

Americans), the Osteoporotic Fractures in Men Study (MrOS, n=2,209 European-Americans), and the Starr County Health Studies (Starr, n=782 Hispanic/Latino-Americans) (Table 1). An additional six cohort studies and data from one physiological research study were analyzed to examine for generalizability across samples. Details about the study subjects are provided in the online supplement.

Phenotypes and Covariates

OSA was quantified using the AHI, defined as the number of episodes of complete (apnea) or partial (hypopnea) cessations of airflow per hour of sleep (or recording time). In this study, sleep data from all seven discovery studies were scored in our Sleep Reading Center. Details of the sleep testing and scoring procedures for each cohort are provided in the online supplement. The primary phenotype was the AHI calculated across the total sleep (or recording) period (AHI-Total; AHI-T). All studies used a hypopnea definition that required a $\geq 3\%$ event-related desaturation. Covariates include age, sex and BMI. AHI measured during REM (AHI-R) and non-REM (AHI-N) sleep periods also were analyzed where available (ARIC, CFS, FHS, MESA and MrOS).

Genotyping and Quality Control

Study participants in ARIC, MESA, and Starr County were genotyped using the Affymetrix 6.0 array; CFS participants were genotyped using the Illumina OmniExpress, Affymetrix 6.0 and the ITMAT-Broad-CARe (IBC) (24) arrays; FHS participants were genotyped using the Affymetrix 500K mapping array and Illumina Omni5 array; HCHS/SOL participants were genotyped using the Illumina Omni 2.5M array with custom content; and MrOS participants were genotyped

using the Illumina Omni 1M array. Data from CFS, FHS, MESA, MrOS and Starr were phased using SHAPEIT (25) and imputed using IMPUTE2 (26) and a 1000 Genomes Project Phase 3 background (version 5, all populations, which contains haplotypes on 2,504 samples for a total of about 81.2 million polymorphic markers); ARIC and HCHS/SOL were imputed using a 1000 Genomes Project Phase 1 background. Single nucleotide polymorphism (SNP) strands were checked in Ensembl and with 1000 Genomes data in SHAPEIT. SNPs with an IMPUTE2 Info score less than 0.88, or a minor allele frequency less than 1% in each study cohort were excluded from analyses.

Statistical Analysis

Rank normalized age and sex-adjusted residuals were analyzed using linear mixed models with a genetic relatedness matrix (GRM) in GEMMA (27) to control for population stratification and relatedness, adjusting for BMI and BMI². Multi-ethnic meta-analyses were performed using the inverse variance weighted fixed-effects approach in METAL (28). Details on statistical analysis are provided in the online supplement.

Results

Demographics

Key characteristics of each cohort were presented in Table 1, with additional details in the online supplement. Across the seven distinct cohorts, data were available for 19,733 individuals, including 10,113 women. Participants were on average middle-aged to elderly and are overweight to obese. The proportion with moderate to severe sleep apnea (AHI \geq 15 events per hour) ranged from 11.7% to 54.8%. In general, the AHI varied with the mean age of the cohort

(higher in the older cohorts). Overall, the sample ancestry was 29.0% European, 6.2% African, 63.6% Hispanic and 1.2% Asian.

Sex-Combined and Sex-Stratified Analyses

The top results of the multi-ethnic meta-analyses were shown in Table 2. In sex-combined results, eight loci showed suggestive association ($P < 1.0 \times 10^{-6}$) with AHI-T, five loci with AHI-N, and two loci with AHI-R. These regions included rs146579140, where variation was associated with AHI-N at an almost significant level ($P = 8.8 \times 10^{-8}$). In sex-stratified results, six loci showed suggestive association with AHI-T, three loci with AHI-N, and three loci with AHI-R in women; eleven loci showed suggestive association with AHI-T, three loci with AHI-N, and two loci with AHI-R in men. In addition, there was one locus significantly associated with AHI-N in men on chromosome 17 (Figure 1), with a lead SNP rs12936587 ($P = 1.7 \times 10^{-8}$). This locus overlapped with the gene *RAI1* (Figure 1C), which codes retinoic acid induced 1 that has been implicated in Smith-Magenis Syndrome (SMS)(29). This lead SNP also showed suggestive association with AHI-N in sex-combined analysis, although the findings reflected associations in men and not women. Figure 2 showed that compared to men with a homozygous genotype of the ancestral allele (A), men with more risk alleles (G) had a higher age- and BMI-adjusted AHI-N on average, but there was no such pattern in women.

Gene-by-Sex Interaction

We performed gene-by-sex interaction analyses for top loci in Table 2 and identified thirteen gene-sex interactions in multi-ethnic meta-analyses, after Bonferroni correction to control for family-wise significance level of 0.05. Of these thirteen gene-sex interactions, twelve had

significant or suggestive association in men but not in women (including rs12936587 with AHI-N; interaction $P = 2.6 \times 10^{-5}$), and one had suggestive association in sex-combined results but neither in men nor in women (although the P value in men was still several orders of magnitude lower than in women). These results suggested there might be different genetic mechanisms for obstructive sleep apnea in women and men.

Expression Quantitative Trait Loci Databases

We examined our most significant SNPs in the *RAII* region (NCBI build 37 locations: chr17:17531709-17644364; $P < 1 \times 10^{-7}$) in expression quantitative trait loci (eQTLs) databases that associate SNPs with gene expression in specific cell lines and tissues (Table E1). Five of the eight genes associated with the *RAII* locus SNPs had minimum eQTL $P < 1 \times 10^{-6}$: *PEMT* (whole blood $P = 2.1 \times 10^{-20}$), *SREBF1* (whole blood $P = 5.0 \times 10^{-20}$), *RASDI* (monocyte $P = 6.8 \times 10^{-12}$), *RAII* (lymphoblastoid $P = 3.4 \times 10^{-7}$), and *TOMIL2* (pituitary $P = 9.4 \times 10^{-7}$).

Assessment of Generalizability in Independent Samples

In summary data provided by replication cohort studies (Table E2), we found no evidence for association with AHI-N for *RAII* in men ($P = 0.34$). However, a consistent direction of association was found in the Cardiovascular Health Study (CHS), the only replication cohort in which sleep studies were scored by the same Sleep Reading Center that scored data in the discovery cohorts.

We replicated the association with rs12936587 in an independent physiological research study of 67 individuals (70% male) (Table E3) studied with in-laboratory polysomnography for the

purposes of elucidating the physiology of OSA. Details about the study subjects are provided in the online supplement. The sample included 55 patients with moderate to severe OSA without other significant co-morbidities and 12 healthy controls. In this well phenotyped sample, after adjusting for age, sex and BMI, the risk G allele of rs12936587 was associated with increasing AHI-N ($P = 0.047$). The association was stronger when we restricted the analysis to AHI-N in the supine position ($P = 0.017$), when airway collapsibility is high.

Discussion

To our knowledge, this is the largest genome-wide analysis of AHI and the only multi-ethnic sex-specific AHI meta-analysis to date. It is also the first human genetic epidemiological study that has examined AHI in REM- and non-REM sleep. Analyses of rigorously collected quantitative sleep data and genome-wide genotype data identified several novel genetic regions with at least suggestive association evidence with each AHI measure. The most significant findings emerged from sex-specific and sleep state-specific analyses. Across all cohorts and race/ethnic groups, the most significant finding was for an association between a locus in *RAI1* in men for AHI measured in Non-REM sleep. Our results identify several biologically-plausible candidates for future functional studies, and highlight genetic variants that may specifically influence OSA propensity in REM vs non-REM sleep, which may have different associations in men and women. The finding of multiple significant gene-by-sex interactions further provides statistical evidence of distinct genetic mechanisms influencing OSA in men and women.

RAI1 is a promising candidate gene for OSA. It encodes a protein that is highly expressed in neuronal tissues and is involved in early neural differentiation and transcriptional regulation of

circadian clock components. Haploinsufficiency of the *RAII* gene has been implicated in Smith Magenis Syndrome (SMS) (29), a complex neurobehavioral disorder that is characterized by multiple craniofacial abnormalities, sleep disturbances, and obesity (30). The craniofacial features include a brachycephalic head form and mid face hypoplasia, which are anatomic risk factors for OSA (30, 31). A majority of individuals with SMS have significant sleep difficulties and disturbed sleep architecture and circadian rhythms (32) and excessive daytime sleepiness. Speech abnormalities, a hoarse voice, and airway hypotonia are also reported (33), suggesting a role of *RAII* in influencing upper airway function. Abnormalities in *RAII* also have been implicated in Potocki-Lupski Syndrome (PTLS) (34-36). PTLS patients often have developmental delay and mild dysmorphic facial features (34, 35), and can exhibit multiple neurological and cardiovascular abnormalities. Eight of the nine patients with PTLS in the initial study displayed central and/or obstructive sleep apnea (36). Both SMS and PTLS appear to involve the *RAII* gene on the short arm of chromosome 17 (37). A de novo *RAII* mutation has been reported in a boy with rapid-onset obesity with hypothalamic dysfunction, hypoventilation, and autonomic dysregulation. The individual displayed an AHI of 10 at age 5 and 27 at age 8, hypercholesterolemia, and macrocephaly (38). In mice, *Rai1* haploinsufficiency is associated with hyperphagia and obesity and abnormal expression of multiple genes in the hypothalamus, including *BDNF* (associated with behavioral and psychiatric morbidities) and *WNT9B* (associated with midfacial development) (39). Although the authors suggested the value in further investigating the role of *RAII* in growth, adiposity and behavior, our results also suggest value in considering sleep apnea as a relevant *RAII* phenotype.

RAII and other genes in the locus may be involved in OSA etiology. Multiple SNPs in the locus overlap epigenetic and/or expression quantitative trait loci (eQTL) evidence that may indicate

regulatory effects. The SNP rs12938840 ($P = 3.97 \times 10^{-7}$) overlaps enhancer regions in 127 Roadmap Epigenomics and ENCODE cell lines and in a further 129 samples of brain regions (40-43). Lead SNPs are associated with expression of five genes (minimum eQTL $P < 1 \times 10^{-6}$; Table E1), including *PEMT*, *SREBF1*, *RAII*, and *RASDI* (44-47). *SREBF1* (formerly *SREBP1*), an important cholesterol biosynthesis regulator, is activated in mice subjected to intermittent hypoxia, leading to hyperlipidemia (48). Activation of *Srebf1* in mouse type 2 alveolar cells leads to lipotoxicity, chronic pulmonary inflammation, and alveolar remodeling (49). *Pemt*, also involved with lipid metabolism, displays sex-specific effects in regulation of HDL and VLDL in mice (50). A waist-hip ratio GWAS association with rs4646404 at the *PEMT* locus was largely sex-specific (21). *RAII* and *RASDI* (formerly *DEXRASI*) regulate circadian rhythm (32, 51, 52). *Rai1* haploinsufficiency in mice leads to sex-specific differences in subcutaneous and abdominal fat distributions (39). The lead SNP rs12936587 (*RAII*) is also significantly associated with coronary artery disease. A sex-stratified analysis indicated that this result was almost entirely due to an association in men (53), providing an exciting avenue for investigating sex differences in not only OSA but also in the association between OSA with coronary artery disease (reported to be stronger in men compared to women) (2). These results also support the importance of future assessment of pleiotropy, specifically the influence of genetic variants that influence both OSA and cardio-metabolic disease and other co-occurring traits.

There are several possible explanations for stronger associations between the *RAII* locus and AHI-N in men compared to women. Men are more likely to have a higher AHI in non-REM sleep than women (9), which has been attributed to poorer neuromuscular compensatory mechanisms. Thus, genetic variants that further reduce airway patency or ventilatory stability in

sleep may have stronger effects in men due to underlying anatomic or physiological risk factors. Conversely, factors that protect women in NREM sleep from recurrent apneas, such as sex hormone-mediated modulation of respiratory chemosensitivity in NREM sleep, may attenuate effects of some genetic variants. It is also possible that sex steroids interact with genetic variants in *RAII* to differentially affect the development of the brain or craniofacial structures, or otherwise interact with genes regulated by sex hormones. *RAII* is upregulated by retinoic acid, which can interact with sex steroids. In the western mosquitofish, *Gambusia affinis*, retinoic acid controls sex-specific development of motor neurons within the spinal cord (54). Furthermore, it has been reported that male *Rai1*-transgenic mice are more growth retarded than are female transgenic mice (55). *Rai1* haploinsufficiency in mice leads to sex-specific differences in adiposity, with greater abdominal fat in females compared to males (39).

This study has several strengths, including the rigorous phenotyping for all discovery cohorts by a central Sleep Reading Center of the sleep studies to ensure high degrees of quality control. Participants in 5 of the 7 cohorts were studied using almost identical equipment and scoring techniques. Consistency of findings for our most significant finding was observed across the 5 distinct discovery cohorts with available data on NREM AHI as well as across all ethnic/racial groups, even when using data from alternative sleep apnea testing devices. The inclusion of multiple ethnic/race groups allowed leveraging different LD structures across populations to identify genetic variants consistently associated with the phenotypes across multiple ethnic/race groups. Genome-wide genotype data were available for the largest sample with OSA phenotypes to date.

The AHI was defined using standard approaches that are used commonly, are reproducible, and show heritability. Hypopneas minimally required a $\geq 3\%$ oxyhemoglobin desaturation. Although AHI levels are highly correlated regardless of hypopnea definition (56), it is possible that associations may have varied because of use of different measurement approaches. The strongest findings for the NREM AHI may not only reflect the specificity of this phenotype, but also the greater accuracy of AHI measures scored from polysomnograms that include electroencephalography recording. The power for replication was limited due to modest sample size (particularly for stage-specific results), although associations in the CHS European-Americans, which had undergone identical phenotyping as several of the discovery cohorts, provided evidence consistent with the discovery finding in the *RAII* region. In addition, in an independent in-laboratory physiological research study of carefully phenotyped individuals that explicitly recruited known cases of OSA without other significant co-morbidities, the association with NREM AHI in the *RAII* region was replicated in sex-combined analysis. This sample, however, was too small to test for sex-specific differences in associations. Although this observation needs to be cautiously interpreted, it is of interest that the strongest finding was for AHI in NREM sleep in the supine position. Men have a significantly higher proportion of apneas in NREM sleep than women, likely due to the occurrence of greater breathing instability in NREM sleep in men compared to women. Men also have more severe sleep apnea in the supine compared to non-supine position, attributed to the effects of positional-dependent airway collapsibility. In contrast, women show a REM-predominant pattern and less positional dependency (57). In other words, the lead SNP associated most strongly with a phenotype subtype most characteristic of “male” sleep apnea. The lack of significant association for this phenotype in our sex-combined discovery sample may reflect differences in the spectrum of

sleep apnea in the physiological study compared to the predominant community-based samples, where sleep apnea in women tends to be mild.

Our study, while identifying novel genetic pathways that may influence OSA, was not designed to identify specific mechanisms. In particular, we were not able to assess to what extent the genetic associations with AHI could be explained by craniofacial features, differences in body fat distribution (particularly, neck circumference) or physiological traits due to lack of information on specific intermediate phenotypes in most of the study samples.

In conclusion, we have identified from multi-ethnic meta-analyses several interesting biological candidates for sex-specific and sleep state-specific associations with AHI, the most widely used clinical measure for OSA. The approach underscores the value of sex-specific analyses in a trait such as OSA, for which there are significant differences in presentation and pathogenesis between men and women. It is widely recognized that the overall AHI likely reflects a heterogeneous set of phenotypes. The analysis of sleep state-specific (REM; Non-REM) findings allowed assessment of more specific OSA phenotypes (i.e., operating in the background of different levels of neuromuscular control) than the overall AHI. Further investigation of the *RAI1* regional association is particularly promising given its role in at least three congenital syndromes associated with sleep abnormalities and its influence on metabolic and physiological traits closely associated with OSA. However, future large-scale studies are warranted for replication and refinement of signals. These studies could lead to important insights into the underlying pathogenesis of the disorder, resulting in targeted treatments, as well as inform screening and risk

stratification. Additional insights into the genetic bases for OSA may be gleaned from further detailed phenotyping, including assessments of neuromuscular control of the airway.

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References

1. Marin JM, Carrizo SJ, Vicente E, Agusti AG. Long-term cardiovascular outcomes in men with obstructive sleep apnoea-hypopnoea with or without treatment with continuous positive airway pressure: An observational study. *Lancet*. 2005 Mar 19-25;365(9464):1046-53.
2. Gottlieb DJ, Yenokyan G, Newman AB, O'Connor GT, Punjabi NM, Quan SF, et al. Prospective study of obstructive sleep apnea and incident coronary heart disease and heart failure: The sleep heart health study. *Circulation*. 2010 Jul 27;122(4):352-60.
3. Kendzerska T, Gershon AS, Hawker G, Tomlinson G, Leung RS. Obstructive sleep apnea and incident diabetes. A historical cohort study. *Am J Respir Crit Care Med*. 2014 Jul 15;190(2):218-25.
4. Alkhasna A, Bhat A, Ladesich J, Barthel B, Bohnam AJ. Severity of obstructive sleep apnea between black and white patients. *Hosp Pract (1995)*. 2011 Oct;39(4):82-6.
5. Redline S, Sotres-Alvarez D, Loreda J, Hall M, Patel SR, Ramos A, et al. Sleep-disordered breathing in hispanic/latino individuals of diverse backgrounds. the hispanic community health study/study of latinos. *Am J Respir Crit Care Med*. 2014 Feb 1;189(3):335-44.

6. Pensuksan WC, Chen X, Lohsoonthorn V, Lertmaharit S, Gelaye B, Williams MA. High risk for obstructive sleep apnea in relation to hypertension among southeast asian young adults: Role of obesity as an effect modifier. *Am J Hypertens*. 2014 Feb;27(2):229-36.
7. Chen X, Wang R, Zee P, Lutsey PL, Javaheri S, Alcantara C, et al. Racial/ethnic differences in sleep disturbances: The multi-ethnic study of atherosclerosis (MESA). *Sleep*. 2015 Jun 1;38(6):877-88.
8. Young T, Palta M, Dempsey J, Skatrud J, Weber S, Badr S. The occurrence of sleep-disordered breathing among middle-aged adults. *N Engl J Med*. 1993 Apr 29;328(17):1230-5.
9. Mohsenin V. Effects of gender on upper airway collapsibility and severity of obstructive sleep apnea. *Sleep Med*. 2003 Nov;4(6):523-9.
10. Koo BB, Patel SR, Strohl K, Hoffstein V. Rapid eye movement-related sleep-disordered breathing: Influence of age and gender. *Chest*. 2008 Dec;134(6):1156-61.
11. Wimms AJ, Ketheeswaran S, Armitstead JP. Obstructive sleep apnea in women: Specific issues and interventions. Sydney, Australia: ResMed Science Center; 2014.
12. Hachul H, Frange C, Bezerra AG, Hirotsu C, Pires GN, Andersen ML, et al. The effect of menopause on objective sleep parameters: Data from an epidemiologic study in sao paulo, brazil. *Maturitas*. 2015 Feb;80(2):170-8.
13. Redline S, Tishler PV, Tosteson TD, Williamson J, Kump K, Browner I, et al. The familial aggregation of obstructive sleep apnea. *Am J Respir Crit Care Med*. 1995 Mar;151(3 Pt 1):682-7.
14. Patel SR, Larkin EK, Redline S. Shared genetic basis for obstructive sleep apnea and adiposity measures. *Int J Obes (Lond)*. 2008 May;32(5):795-800.
15. Larkin EK, Patel SR, Goodloe RJ, Li Y, Zhu X, Gray-McGuire C, et al. A candidate gene study of obstructive sleep apnea in european americans and african americans. *Am J Respir Crit Care Med*. 2010 Oct 1;182(7):947-53.
16. Patel SR, Goodloe R, De G, Kowgier M, Weng J, Buxbaum SG, et al. Association of genetic loci with sleep apnea in european americans and african-americans: The candidate gene association resource (CARE). *PLoS One*. 2012;7(11):e48836.
17. Yue W, Liu H, Zhang J, Zhang X, Wang X, Liu T, et al. Association study of serotonin transporter gene polymorphisms with obstructive sleep apnea syndrome in chinese han population. *Sleep*. 2008 Nov;31(11):1535-41.
18. Cade BE, Chen H, Stilp AM, Gleason KJ, Sofer T, Ancoli-Israel S, et al. Genetic associations with obstructive sleep apnea traits in hispanic/latino americans. *Am J Respir Crit Care Med*. 2016 Mar 15;194(7):886-97.

19. Randall JC, Winkler TW, Kutalik Z, Berndt SI, Jackson AU, Monda KL, et al. Sex-stratified genome-wide association studies including 270,000 individuals show sexual dimorphism in genetic loci for anthropometric traits. *PLoS Genet*. 2013 Jun;9(6):e1003500.
20. Winkler TW, Justice AE, Graff M, Barata L, Feitosa MF, Chu S, et al. The influence of age and sex on genetic associations with adult body size and shape: A large-scale genome-wide interaction study. *PLoS Genet*. 2015 Oct 1;11(10):e1005378.
21. Shungin D, Winkler TW, Croteau-Chonka DC, Ferreira T, Locke AE, Magi R, et al. New genetic loci link adipose and insulin biology to body fat distribution. *Nature*. 2015 Feb 12;518(7538):187-96.
22. Link JC, Chen X, Prien C, Borja MS, Hammerson B, Oda MN, et al. Increased high-density lipoprotein cholesterol levels in mice with XX versus XY sex chromosomes. *Arterioscler Thromb Vasc Biol*. 2015 Aug;35(8):1778-86.
23. Siddiqui F, Walters AS, Goldstein D, Lahey M, Desai H. Half of patients with obstructive sleep apnea have a higher NREM AHI than REM AHI. *Sleep Med*. 2006 Apr;7(3):281-5.
24. Keating BJ, Tischfield S, Murray SS, Bhangale T, Price TS, Glessner JT, et al. Concept, design and implementation of a cardiovascular gene-centric 50 k SNP array for large-scale genomic association studies. *PLoS One*. 2008;3(10):e3583.
25. Delaneau O, Marchini J, Zagury JF. A linear complexity phasing method for thousands of genomes. *Nat Methods*. 2011 Dec 4;9(2):179-81.
26. Howie BN, Donnelly P, Marchini J. A flexible and accurate genotype imputation method for the next generation of genome-wide association studies. *PLoS Genet*. 2009 Jun;5(6):e1000529.
27. Zhou X, Stephens M. Genome-wide efficient mixed-model analysis for association studies. *Nat Genet*. 2012 Jun 17;44(7):821-4.
28. Willer CJ, Li Y, Abecasis GR. METAL: Fast and efficient meta-analysis of genomewide association scans. *Bioinformatics*. 2010 Sep 1;26(17):2190-1.
29. Elsea SH, Girirajan S. Smith-magenis syndrome. *Eur J Hum Genet*. 2008 Apr;16(4):412-21.
30. Smith AC, Dykens E, Greenberg F. Sleep disturbance in smith-magenis syndrome (del 17 p11.2). *Am J Med Genet*. 1998 Mar 28;81(2):186-91.
31. Cakirer B, Hans MG, Graham G, Aylor J, Tishler PV, Redline S. The relationship between craniofacial morphology and obstructive sleep apnea in whites and in african-americans. *Am J Respir Crit Care Med*. 2001 Mar;163(4):947-50.

32. Williams SR, Zies D, Mullegama SV, Grotewiel MS, Elsea SH. Smith-magenis syndrome results in disruption of CLOCK gene transcription and reveals an integral role for RAI1 in the maintenance of circadian rhythmicity. *Am J Hum Genet.* 2012 Jun 8;90(6):941-9.
33. Gropman AL, Duncan WC, Smith AC. Neurologic and developmental features of the smith-magenis syndrome (del 17p11.2). *Pediatr Neurol.* 2006 May;34(5):337-50.
34. Brown A, Phelan MC, Patil S, Crawford E, Rogers RC, Schwartz C. Two patients with duplication of 17p11.2: The reciprocal of the smith-magenis syndrome deletion? *Am J Med Genet.* 1996 May 17;63(2):373-7.
35. Potocki L, Chen KS, Park SS, Osterholm DE, Withers MA, Kimonis V, et al. Molecular mechanism for duplication 17p11.2- the homologous recombination reciprocal of the smith-magenis microdeletion. *Nat Genet.* 2000 Jan;24(1):84-7.
36. Potocki L, Bi W, Treadwell-Deering D, Carvalho CM, Eifert A, Friedman EM, et al. Characterization of potocki-lupski syndrome (dup(17)(p11.2p11.2)) and delineation of a dosage-sensitive critical interval that can convey an autism phenotype. *Am J Hum Genet.* 2007 Apr;80(4):633-49.
37. Lupski JR, Stankiewicz P. Genomic disorders: Molecular mechanisms for rearrangements and conveyed phenotypes. *PLoS Genet.* 2005 Dec;1(6):e49.
38. Thaker VV, Esteves KM, Towne MC, Brownstein CA, James PM, Crowley L, et al. Whole exome sequencing identifies RAI1 mutation in a morbidly obese child diagnosed with ROHHAD syndrome. *J Clin Endocrinol Metab.* 2015 May;100(5):1723-30.
39. Burns B, Schmidt K, Williams SR, Kim S, Girirajan S, Elsea SH. Rai1 haploinsufficiency causes reduced bdnf expression resulting in hyperphagia, obesity and altered fat distribution in mice and humans with no evidence of metabolic syndrome. *Hum Mol Genet.* 2010 Oct 15;19(20):4026-42.
40. Roadmap Epigenomics Consortium, Kundaje A, Meuleman W, Ernst J, Bilenky M, Yen A, et al. Integrative analysis of 111 reference human epigenomes. *Nature.* 2015 Feb 19;518(7539):317-30.
41. ENCODE Project Consortium. An integrated encyclopedia of DNA elements in the human genome. *Nature.* 2012 Sep 6;489(7414):57-74.
42. Ward LD, Kellis M. HaploReg: A resource for exploring chromatin states, conservation, and regulatory motif alterations within sets of genetically linked variants. *Nucleic Acids Res.* 2012 Jan;40(Database issue):D930-4.
43. Vermunt MW, Reinink P, Korving J, de Bruijn E, Creyghton PM, Basak O, et al. Large-scale identification of coregulated enhancer networks in the adult human brain. *Cell Rep.* 2014 Oct 23;9(2):767-79.

44. Westra HJ, Peters MJ, Esko T, Yaghootkar H, Schurmann C, Kettunen J, et al. Systematic identification of trans eQTLs as putative drivers of known disease associations. *Nat Genet.* 2013 Oct;45(10):1238-43.
45. Zeller T, Wild P, Szymczak S, Rotival M, Schillert A, Castagne R, et al. Genetics and beyond--the transcriptome of human monocytes and disease susceptibility. *PLoS One.* 2010 May 18;5(5):e10693.
46. Lappalainen T, Sammeth M, Friedlander MR, 't Hoen PA, Monlong J, Rivas MA, et al. Transcriptome and genome sequencing uncovers functional variation in humans. *Nature.* 2013 Sep 26;501(7468):506-11.
47. GTEx Consortium. Human genomics. the genotype-tissue expression (GTEx) pilot analysis: Multitissue gene regulation in humans. *Science.* 2015 May 8;348(6235):648-60.
48. Li J, Nanayakkara A, Jun J, Savransky V, Polotsky VY. Effect of deficiency in SREBP cleavage-activating protein on lipid metabolism during intermittent hypoxia. *Physiol Genomics.* 2007 Oct 22;31(2):273-80.
49. Plantier L, Besnard V, Xu Y, Ikegami M, Wert SE, Hunt AN, et al. Activation of sterol-response element-binding proteins (SREBP) in alveolar type II cells enhances lipogenesis causing pulmonary lipotoxicity. *J Biol Chem.* 2012 Mar 23;287(13):10099-114.
50. Noga AA, Vance DE. A gender-specific role for phosphatidylethanolamine N-methyltransferase-derived phosphatidylcholine in the regulation of plasma high density and very low density lipoproteins in mice. *J Biol Chem.* 2003 Jun 13;278(24):21851-9.
51. Boone PM, Reiter RJ, Glaze DG, Tan DX, Lupski JR, Potocki L. Abnormal circadian rhythm of melatonin in smith-magenis syndrome patients with RAI1 point mutations. *Am J Med Genet A.* 2011 Aug;155A(8):2024-7.
52. Cheng HY, Dziema H, Papp J, Mathur DP, Koletar M, Ralph MR, et al. The molecular gatekeeper *Dexras1* sculpts the photic responsiveness of the mammalian circadian clock. *J Neurosci.* 2006 Dec 13;26(50):12984-95.
53. CARDIoGRAMplusC4D Consortium, Deloukas P, Kanoni S, Willenborg C, Farrall M, Assimes TL, et al. Large-scale association analysis identifies new risk loci for coronary artery disease. *Nat Genet.* 2013 Jan;45(1):25-33.
54. McCaffery PJ, Adams J, Maden M, Rosa-Molinar E. Too much of a good thing: Retinoic acid as an endogenous regulator of neural differentiation and exogenous teratogen. *Eur J Neurosci.* 2003 Aug;18(3):457-72.
55. Girirajan S, Patel N, Slager RE, Tokarz ME, Bucan M, Wiley JL, et al. How much is too much? phenotypic consequences of *Rai1* overexpression in mice. *Eur J Hum Genet.* 2008 Aug;16(8):941-54.

56. Redline S, Kapur VK, Sanders MH, Quan SF, Gottlieb DJ, Rapoport DM, et al. Effects of varying approaches for identifying respiratory disturbances on sleep apnea assessment. *Am J Respir Crit Care Med.* 2000 Feb;161(2 Pt 1):369-74.

57. Lozo T, Komnenov D, Badr MS, Mateika JH. Sex differences in sleep disordered breathing in adults. *Respir Physiol Neurobiol.* 2017 Nov;245:65-75.

Figures

Figure 1. Manhattan, quantile-quantile (Q-Q), and regional association plots of multi-ethnic meta-analysis results for AHI-N in men. A) The Manhattan plot shows minus log 10 p-values against genomic coordinates (NCBI build 37), and consecutive chromosomes were colored in black and grey alternately; B) The Q-Q plot shows observed minus log 10 p-values against expected values under no association; C) The regional association plot of multi-ethnic meta-analysis results for AHI-N in men near the *RAI1* gene on chromosome 17.

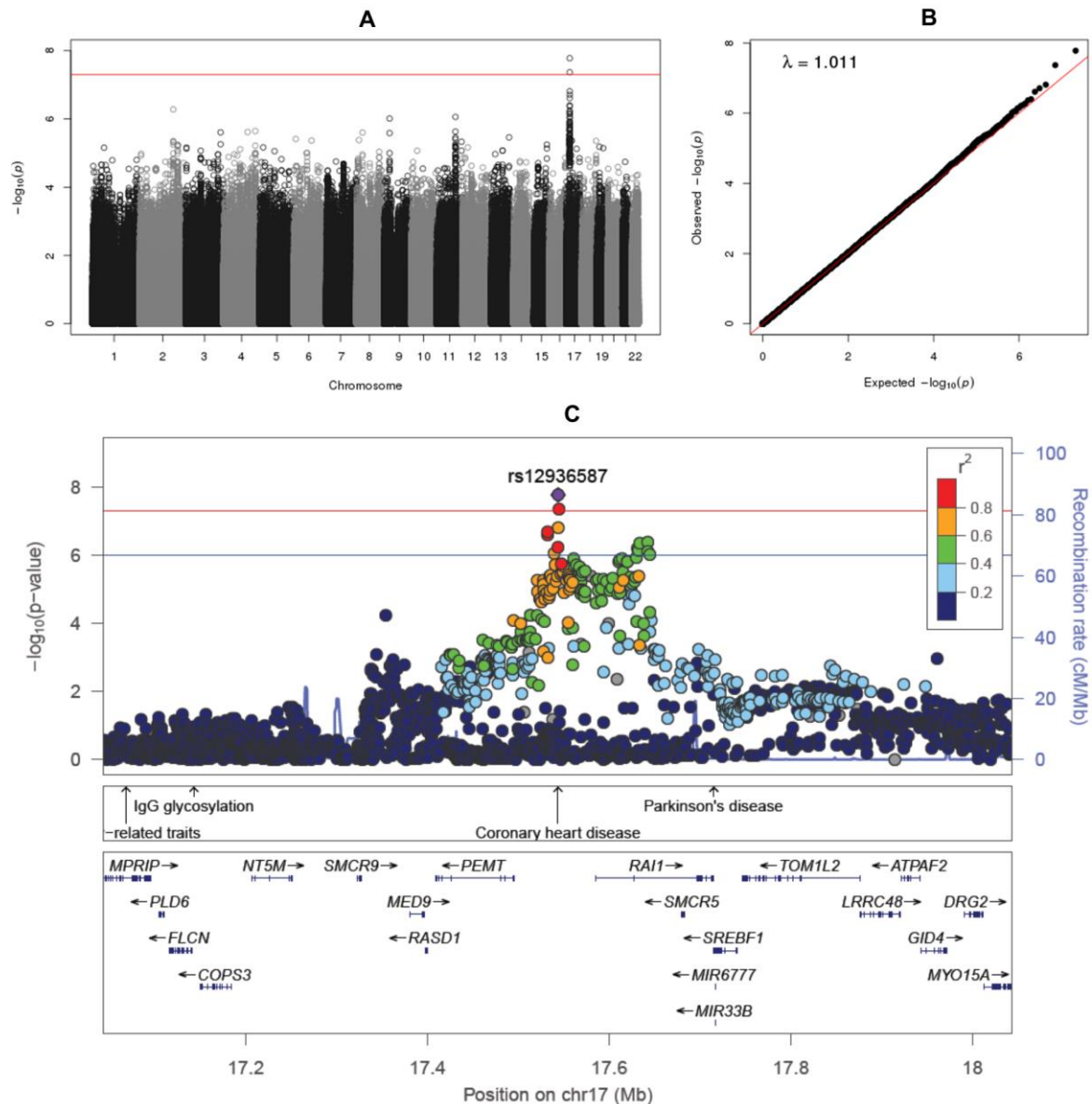
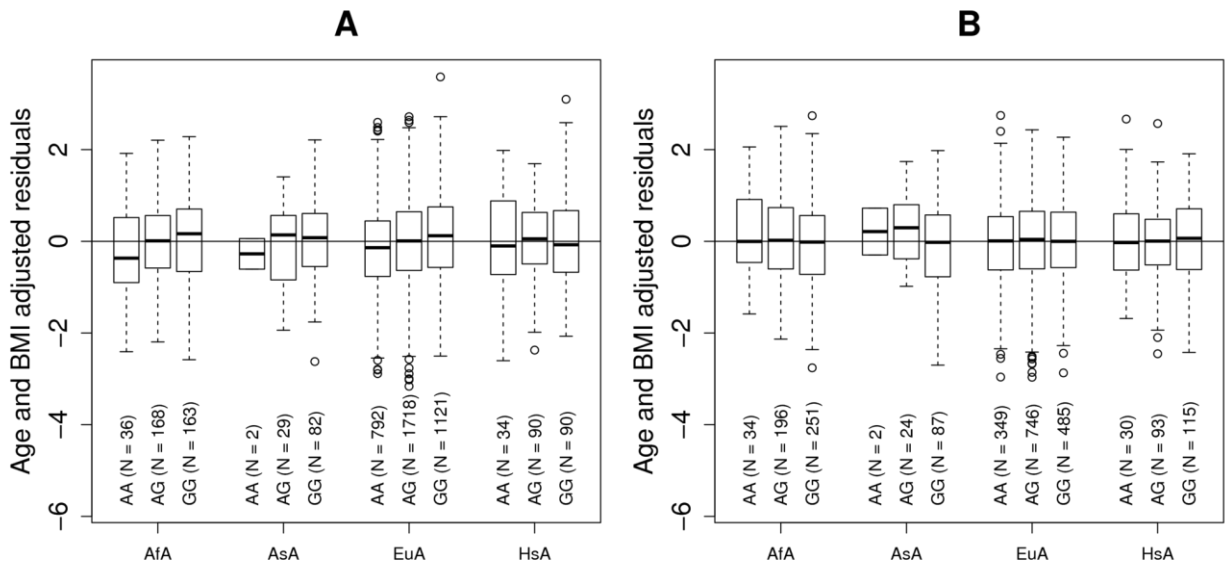


Figure 2. Sex differences in the distribution of BMI and BMI² adjusted residuals for study subjects with different genotypes (AA, AG, and GG) of rs12936587. The phenotype is rank normalized sex-stratified residuals of AHI-N adjusting for age, age². A) Men; B) Women. AfA: African-Americans; AsA: Asian-Americans; EuA: European-Americans; HsA: Hispanic/Latino-Americans.



Tables

Table 1. Sample description of study subjects in discovery cohorts.

Ethnic Group	Cohort	N	Age (years)	Percent Female	BMI (kg/m²)	Apnea Hypopnea Index	Percent OSA	AHI-N	AHI-R	AHI-R/AHI-N
African-Americans	CFS*	731	37.84 (19.44)	56.2	31.63 (9.69)	5.85 (19.70)	31.7	2.47 (12.19)	9.23 (30.87)	2.89 (8.70)
	MESA	490	69.13 (9.10)	54.3	30.42 (5.71)	13.34 (21.16)	46.5	8.64 (20.1)	30.97 (39.83)	2.64 (5.09)
Asian-Americans	MESA	228	68.13 (9.19)	50.4	24.08 (3.19)	13.97 (23.90)	47.4	11.61 (24.31)	21.77 (34.19)	1.68 (2.64)
European-Americans	ARIC	1,463	62.43 (5.69)	51.5	28.83 (5.13)	8.70 (15.50)	32.5	5.75 (13.78)	16.46 (27.68)	2.58 (4.69)
	CFS*	702	41.59 (19.45)	52.7	30.24 (8.66)	5.59 (18.99)	31.1	2.07 (13.14)	7.06 (21.20)	2.28 (7.40)
	FHS*	646	59.38 (8.97)	50.0	28.49 (5.01)	8.18 (14.51)	30.0	5.25 (13.66)	15.25 (23.60)	2.48 (4.98)
	MESA	707	68.52 (9.10)	53.6	27.99 (5.21)	12.62 (20.67)	44.0	9.61 (20.45)	22.31 (30.65)	1.80 (2.97)
	MrOS	2,209	76.68 (5.66)	0.0	27.22 (3.74)	12.73 (18.11)	43.7	10.93 (19.58)	18.30 (24.17)	1.53 (2.42)
Hispanic/Latino-Americans	HCHS/SOL	11,317	46.17 (13.79)	59.1	29.79 (6.00)	1.97 (6.20)	11.7	NA	NA	NA

Ethnic Group	Cohort	N	Age (years)	Percent Female	BMI (kg/m ²)	Apnea Hypopnea Index	Percent OSA	AHI-N	AHI-R	AHI-R/AHI-N
	MESA	458	68.34 (9.20)	52.8	30.07 (5.52)	16.94 (23.05)	54.8	12.16 (23.12)	30.00 (36.09)	2.13 (3.50)
	Starr	782	52.34 (11.29)	71.9	32.15 (6.78)	10.35 (17.18)	37.1	NA	NA	NA

Seven studies included 19,733 individuals with genotypes and phenotypes (1,221 African-Americans; 228 Asian-Americans; 5,727 European-Americans; 12,557 Hispanic/Latino-Americans). NREM- and REM-specific data are only available in a sample of the CFS data and were not collected in HCHS/SOL and Starr County. Mean (standard deviation) are listed for Age and BMI, and median (interquartile range) are listed for Apnea Hypopnea Index, AHI-N, AHI-R, and AHI-R/AHI-N. OSA: obstructive sleep apnea, defined as Apnea Hypopnea Index \geq 15. AHI-N: NREM-specific Apnea Hypopnea Index. AHI-R: REM-specific Apnea Hypopnea Index. *: Family cohorts.

Table 2. Significant and suggestive multi-ethnic meta-analysis results for AHI (total), AHI-NREM, and AHI-REM.

Trait	SNP	Chr	Locus	Alleles	Sex-combined			Women			Men			Sex diff
					N	Effect	P	N	Effect	P	N	Effect	P	P
AHI-T	rs76321756	1		C/T	2,381	0.130	9.7×10^{-5}	1,289	-0.017	7.0×10^{-1}	1,092	0.244	2.3×10^{-7}	2.7×10^{-6}
AHI-T	rs11897825	2	AC011752.1; AC067959.1	G/A	15,364	0.026	2.2×10^{-2}	7,650	0.080	3.3×10^{-7}	7,714	-0.014	3.6×10^{-1}	2.8×10^{-3}
AHI-T	rs999944	2		G/A	19,733	0.072	9.7×10^{-7}	10,113	0.063	2.0×10^{-3}	9,620	0.077	2.6×10^{-4}	5.5×10^{-3}
AHI-T	rs35520189	2		C/A	19,733	0.052	6.1×10^{-7}	10,113	0.064	5.8×10^{-6}	9,620	0.042	5.3×10^{-3}	5.9×10^{-1}
AHI-T	rs72149316:AATAA	2	SCN3A	GATAA/	1,679	0.428	6.0×10^{-7}	919	0.326	3.7×10^{-3}	764	0.606	5.9×10^{-7}	2.3×10^{-3}

				Sex-combined	Women	Men	Sex diff							
				AATAA										
AHI-T	rs34526934	2	HOXD-AS2;HOXD3;HOXD4;MIR10B	A/T	19,505	0.060	1.1×10 ⁻⁶	9,998	0.024	1.7×10 ⁻¹	9,507	0.087	3.6×10 ⁻⁷	1.8×10 ⁻³
AHI-T	rs62189527	2	HDAC4	C/T	12,665	0.174	4.6×10 ⁻⁵	7,384	0.067	2.3×10 ⁻¹	5,281	0.320	7.4×10 ⁻⁷	9.7×10⁻⁴
AHI-T	rs35001935:CGTG TGT	3	C3orf67; RP11-147N17.1	CGTGT/CGTGTG T	1,679	0.251	7.7×10 ⁻⁷	919	0.091	1.9×10 ⁻¹	760	0.339	1.3×10 ⁻⁶	3.3×10⁻⁴
AHI-T	chr3:166153534:D	3		A/AAC	2,461	0.249	3.4×10 ⁻⁵	1,481	0.376	6.1×10 ⁻⁷	980	0.157	9.4×10 ⁻²	3.2×10 ⁻¹
AHI-T	rs34188544	4		G/A	2,135	0.522	3.6×10 ⁻⁷	1,264	0.333	1.4×10 ⁻²	871	0.728	2.1×10 ⁻⁶	8.2×10 ⁻³
AHI-T	rs73352871	5		C/A	12,996	0.082	2.4×10 ⁻³	7,610	-0.025	4.8×10 ⁻¹	5,386	0.203	5.9×10 ⁻⁷	2.2×10⁻⁶
AHI-T	rs79987021	7		T/C	12,265	0.256	6.2×10 ⁻⁷	7,199	0.186	5.4×10 ⁻³	5,066	0.316	7.7×10 ⁻⁵	1.5×10 ⁻²
AHI-T	rs117004340	8	DLC1	T/A	19,504	0.079	2.2×10 ⁻⁵	9,997	-0.001	9.7×10 ⁻¹	9,507	0.136	1.8×10 ⁻⁷	9.2×10⁻⁴
AHI-T	rs3736021	8	ENTPD4	T/G	19,732	0.065	7.2×10 ⁻⁷	10,112	0.057	1.1×10 ⁻³	9,620	0.071	3.4×10 ⁻⁴	1.1×10 ⁻¹
AHI-T	rs35857674	8	SNTG1	A/G	19,732	0.024	2.2×10 ⁻²	10,112	0.070	7.9×10 ⁻⁷	9,620	-0.016	2.8×10 ⁻¹	5.2×10 ⁻³
AHI-T	rs117169866	10		G/A	18,512	0.150	5.6×10 ⁻⁷	9,436	0.116	7.0×10 ⁻³	9,396	0.158	1.1×10 ⁻⁴	6.7×10 ⁻²

					Sex-combined			Women			Men		Sex diff	
AHI-T	rs1387259	12	RP11-370I10.2; ZNF641	A/G	19,733	0.052	2.3×10 ⁻⁶	10,113	0.031	4.1×10 ⁻²	9,620	0.078	3.7×10 ⁻⁷	2.3×10 ⁻²
AHI-T	rs9600832	13	NBEA	C/T	19,733	0.064	1.2×10 ⁻⁵	10,113	0.012	5.4×10 ⁻¹	9,620	0.104	5.6×10 ⁻⁷	7.8×10⁻⁵
AHI-T	rs75900232	13		G/A	16,337	0.159	2.1×10 ⁻⁴	8,942	-0.037	5.2×10 ⁻¹	8,199	0.307	6.5×10 ⁻⁷	4.9×10⁻⁵
AHI-T	rs115432071	16	C16orf62	T/A	12,996	0.197	8.5×10 ⁻⁶	7,610	0.055	3.4×10 ⁻¹	5,386	0.346	3.0×10 ⁻⁷	1.6×10⁻⁴
AHI-T	rs4787347	16	HS3ST4	G/A	19,733	0.029	3.3×10 ⁻³	10,113	0.067	7.2×10 ⁻⁷	9,620	0.002	8.8×10 ⁻¹	1.9×10 ⁻¹
AHI-T	rs112190082	17	STX8	G/T	13,905	0.099	1.0×10 ⁻³	8,188	0.200	3.6×10 ⁻⁷	5,717	0.013	7.7×10 ⁻¹	3.4×10 ⁻²
AHI-T	rs113724004	17		T/C	19,505	0.069	5.9×10 ⁻⁵	9,998	0.020	4.0×10 ⁻¹	9,507	0.123	8.0×10 ⁻⁷	3.9×10⁻⁴
AHI-T	rs142002225	20		T/C	19,733	0.055	2.8×10 ⁻⁴	10,113	0.108	3.2×10 ⁻⁷	9,620	0.022	3.1×10 ⁻¹	3.4×10 ⁻¹
AHI-N	rs146579140	2		C/T	6,737	0.206	8.8×10 ⁻⁸	2,412	0.092	1.6×10 ⁻¹	4,212	0.237	5.2×10 ⁻⁷	1.3×10 ⁻²
AHI-N	rs10474877	5	ANKRD33B	T/C	6,737	0.039	1.1×10 ⁻¹	2,412	0.203	2.1×10 ⁻⁷	4,212	-0.015	6.2×10 ⁻¹	3.7×10 ⁻²
AHI-N	rs73686127	7		A/T	1,300	0.508	2.6×10 ⁻⁷	719	0.509	4.7×10 ⁻⁵	367	0.464	4.6×10 ⁻³	2.2×10 ⁻¹
AHI-N	rs10968431	9	LINGO2	G/A	1,300	0.239	5.3×10 ⁻⁴	719	-0.030	7.5×10 ⁻¹	581	0.474	9.7×10 ⁻⁷	1.3×10⁻⁵
AHI-N	rs79697311	11		C/A	6,029	0.215	5.2×10 ⁻³	1,295	0.719	6.9×10 ⁻⁷	3,885	0.176	6.9×10 ⁻²	4.2×10 ⁻¹
AHI-N	rs34174435	11	OR5AN1	GA/G	5,304	0.101	6.4×10 ⁻⁷	1,673	0.105	4.4×10 ⁻³	3,631	0.086	3.3×10 ⁻⁴	1.1×10 ⁻¹

					Sex-combined			Women			Men		Sex diff	
AHI-N	rs75482679	11	CNTN5	T/C	6,737	0.128	1.0×10^{-5}	2,412	-0.019	6.9×10^{-1}	4,325	0.176	8.7×10^{-7}	1.3×10^{-4}
AHI-N	rs72986876	11	<i>PDGFD</i>	A/T	6,511	0.223	7.8×10^{-7}	2,299	0.194	1.3×10^{-2}	4,212	0.217	7.9×10^{-5}	4.4×10^{-1}
AHI-N	rs116696666	16		A/T	1,300	0.290	1.6×10^{-3}	719	0.608	5.9×10^{-7}	581	0.136	3.1×10^{-1}	2.5×10^{-1}
AHI-N	rs12936587	17	RAI1	G/A	6,737	0.083	6.9×10^{-7}	2,412	-0.008	7.7×10^{-1}	4,325	0.116	1.7×10^{-8}	2.6×10^{-5}
AHI-R	chr1:74135031	1	RP4-788P17.1	A/T	1,546	0.310	9.8×10^{-7}	858	0.208	1.0×10^{-2}	688	0.403	5.2×10^{-5}	1.2×10^{-1}
AHI-R	rs72956768	4	TMEM154	G/T	1,290	0.254	6.4×10^{-4}	714	0.020	8.3×10^{-1}	576	0.563	5.2×10^{-7}	1.5×10^{-4}
AHI-R	rs35077018	7	<i>HUS1</i> ; <i>PKD1L1</i>	G/GCTA GTGCGT GCATGA ACTAGT TGGTCT GCAAGT ACAAGA TGTATA AATATA CAGGG GAAAA AACATC	1,290	0.198	5.5×10^{-4}	714	0.357	8.2×10^{-7}	576	0.049	5.9×10^{-1}	3.4×10^{-2}
AHI-R	rs9297743	8	RP11-89K10.1	G/A	6,700	0.127	5.5×10^{-4}	2,400	-0.037	5.1×10^{-1}	4,300	0.241	9.2×10^{-7}	2.6×10^{-3}
AHI-R	rs79697311	11		C/A	5,998	0.173	1.8×10^{-2}	1,289	0.692	4.4×10^{-7}	3,867	0.068	4.7×10^{-1}	4.6×10^{-2}

					Sex-combined			Women			Men		Sex diff	
AHI-R	rs201360344	12	<i>C12orf55</i>	G/GA	924	0.478	5.5×10^{-7}	498	0.516	3.2×10^{-5}	426	0.329	3.1×10^{-2}	4.1×10^{-1}
AHI-R	rs41408454	15		G/A	1,290	0.287	1.1×10^{-3}	714	0.523	8.8×10^{-7}	576	0.019	8.9×10^{-1}	1.2×10^{-2}

SNPs with significant ($p < 5.0 \times 10^{-8}$) and suggestive ($p < 1.0 \times 10^{-6}$) p-values. SNPs that were present in only one ethnic group were excluded. AHI-T: AHI (total). AHI-N: AHI during NREM. AHI-R: AHI during REM. Alleles: effect allele / non-effect allele.