

A new genus, *Desertispora*, and a new species, *Diversispora sabulosa*, in the family Diversisporaceae (order Diversisporales, subphylum Glomeromycotina)

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Abstract

Phylogenetic analyses of sequences of the SSU–ITS–LSU nrDNA segment and the RPB1 gene showed that the arbuscular mycorrhizal fungus originally described as *Diversispora omaniana* does not belong to the genus *Diversispora*, but represents a separate clade at the rank of genus in the family Diversisporaceae of the order Diversisporales. The closest natural relatives of the fungus proved to be species of the genera *Corymbiglomus* and *Redeckera*. Consequently, the new genus was named *Desertispora*, and *Di. omaniana* was renamed *De. omaniana* comb. nov. In addition, the morphological and histochemical features of spores and mycorrhizal structures of a new *Diversispora* sp., *Di. sabulosa*, were described and the closest relatives of the species were determined based on phylogenetic analyses of sequences of the two loci mentioned above. The new fungus was grown in single-species cultures established from spores extracted from a trap culture inoculated with a mixture of the rhizosphere soil and root fragments of *Ammophila arenaria* that had colonized maritime sand dunes of the Curonian Spit located in the north of Lithuania. *Diversispora sabulosa* was never found before in many different sites of the world which were sampled during the last 34 years by the last author of the paper. Also, the lack of molecular sequences in public databases of identity $\geq 97\%$ to sequences of *Di. sabulosa* suggests that the fungus is rare on the Earth.

Keywords Arbuscular mycorrhizal fungi · Morphology · Molecular phylogeny · RPB1 · SSU–ITS–LSU nrDNA

Introduction

The subphylum Glomeromycotina (C. Walker & A. Schüßler) Spatafora & Stajich (phylum Mucoromycota Doweld;

Spatafora et al. 2016), comprising arbuscular mycorrhizal fungi (AMF), consists of four orders, of which the Diversisporales sensu C. Walker & A. Schüßler is distinguished by possessing species most diverse in morphology and histochemical properties of spores and mycorrhizal structures, and molecular phylogeny (Walker and Schüßler 2004; Schüßler and Walker 2010). Oehl et al. (2011a) divided the Diversisporales to erect Gigasporales Sieverd. et al. ord. nov. comprising AMF forming spores from a bulbous sporogenous hypha, called gigasporoid spores. Redecker et al. (2013) rejected the proposal as unsupported, but Castillo et al. (2016) and Pagano et al. (2016) accepted the erection of the Gigasporales. In the present study, we have not checked the accuracy of the decisions, but tested the hypothesis that the species originally described as *Diversispora omaniana* Symanczik, Błasz. & Al-Yahya'ei (Symanczik et al. 2014) represents an undescribed taxon in the Diversisporales. In order to determine the rank and position of the taxon, we concluded that the molecular properties of the species should be compared with representatives of as many families of the

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Diversisporales as possible. Therefore, we consider here the composition of the Diversisporales according to its original definition (Walker and Schüßler 2004; Schüßler and Walker 2010).

The widely understood Diversisporales contains five families, among which is the Diversisporaceae C. Walker & A. Schüßler with five genera. Three of the genera, *Corymbiglomus* Błaszk. & Chwat, *Diversispora* C. Walker & A. Schüßler, and *Redeckera*, are represented by species forming spores at the tip of a cylindrical or funnel-shaped sporogenous hypha (Schüßler and Walker 2010; Błaszkowski 2012; Błaszkowski and Chwat 2013). Spores of the monospecific genera *Otospora* Oehl, Palenz. & N. Ferrol and *Tricispora* Oehl et al. arise laterally on and inside, respectively, the neck of a sporiferous saccule (Palenzuela et al. 2008; Oehl et al. 2011c). Thus, the spore formation modes of *O. bareae* Palenz., N. Ferrol & Oehl and *T. nevadensis* (Palenz. et al.) Oehl et al., the latter originally described as *Entrophospora nevadensis* Palenz. et al. (Palenzuela et al. 2010), resemble those of spores of *Acaulospora laevis* Gerd. & Trappe and *Entrophospora infrequens* (I.R. Hall) R.N. Ames & R.W. Schneid., respectively, the type species of the genera *Acaulospora* Gerd. & Trappe and *Entrophospora* R.N. Ames & R.W. Schneid., respectively (Gerdemann and Trappe 1974; Ames and Schneider 1979; Schüßler and Walker 2010).

As mentioned above, spores of *Diversispora* spp. arise terminally on a cylindrical or funnel-shaped subtending hypha, thus similarly to glomoid spores of *Glomus macrocarpum* Tul. & C. Tul., the type species of the genus *Glomus* Tul. & C. Tul. and the Glomeromycotina (Schüßler and Walker 2010). In *Glomus* spp., all layers of the subtending hyphal wall and the spore wall form concurrently and the subtending hyphal wall layer continuous with the structural laminate spore wall layer arises far below the spore base and, thereby, the subtending hypha is persistent and its wall is colored similarly to the laminate spore wall layer (Oehl et al. 2011b; Błaszkowski 2012). In contrast, the subtending hyphal wall of all *Diversispora* spp., also those with dark-colored spores, is hyaline to pale-colored and continuous only with a spore wall layer or layers overlying the main structural laminate spore wall layer, or the components of the subtending hyphal wall are continuous with all these spore wall layers (Oehl et al. 2011b). However, the laminate spore wall layer probably arises de novo sequentially after the full differentiation of the spore wall layer(s), which cover them, and starts developing either at the level of the lower surface of the spore wall layer directly adhering to its upper surface or slightly below the spore base (Oehl et al. 2011b). Thus, in the region of the subtending hyphal lumen, the laminate spore wall layer either is not present in the subtending hyphal wall at all or this layer is a small part of the subtending hyphal wall. The result is that the subtending hyphal wall layer continuous with the laminate

spore wall layer does not create any or forms a weak physical support for the other subtending hyphal wall layer(s). Therefore, the spore subtending hypha of *Diversispora* spp. is usually fragile, and Oehl et al. (2011b) named spores of the genus diversisporoid.

Like in most species of the Glomeromycotina producing terminal, one-walled spores on a cylindrical or funnel-shaped sporogenous hypha, the spore wall structure of *Diversispora* spp. is simple and consists of two to four layers (Oehl et al. 2011b; Błaszkowski 2012). However, in contrast to many other genera of AMF forming spores in a similar way, spores of all *Diversispora* spp. do not react at all or rarely stain only faintly in Melzer's reagent, except for *Di. omaniana* (Symanczik et al. 2014), which is the main focus of this study.

Although the spore morphological and histochemical features discussed above clearly allow to separate *Diversispora* spp. from its closest natural relatives, i.e., *Corymbiglomus* and *Redeckera* spp. (see Figs. 1, 2, and 3; Błaszkowski 2012), the differentiation between *Diversispora* spp. and some species of the genus *Claroideoglomus* C. Walker & A. Schüßler of the family Claroideoglomeraceae C. Walker & A. Schüßler may be difficult and uncertain (Oehl et al. 2011b; Błaszkowski, pers. observ.). This also applies to species of many other clades of the Glomeromycotina, mainly those producing spores similar in the mode of formation and construction to spores of *Diversispora* spp. Therefore, the identification and characterization of AMF have to be based on combined morphological data and those obtained from molecular phylogenetic analyses, as, for example, Gamper et al. (2009) suggested.

Based on our observations, of the molecular markers used in studies dealing with the identification and phylogeny of AMF, the small subunit gene (part; thereafter named SSU), the internal transcribed spacer 1 (ITS1), the 5.8S rRNA gene, ITS2 rDNA (full; hereafter named ITS), and the large subunit (part; thereafter named LSU) rRNA gene (thereafter together named SSU–ITS–LSU) is best suited for molecular and phylogenetic analyses of AMF because it is easily, successfully, and faultlessly (in 100% in our studies) amplified by the primers designed by Krüger et al. (2009), which, moreover, amplify the DNA fragment of all clades of the Glomeromycotina. The only drawback of sequences of the SSU–ITS–LSU segment is their weak species resolving power with respect to some *Rhizoglomus* spp., in which the variability of the ITS part may reach 20% (Stockinger et al. 2009). To overcome the problem, Stockinger et al. (2014) came back to earlier findings of Redecker and Raab (2006), which proved that the information contained in the largest subunit of RNA polymerase II (RPB1) gene is less variable and its species resolution is higher, and, consequently, constructed numerous primers to amplify the RPB1 gene of the Glomeromycotina. Our analyses of RPB1 sequences also showed that their variability did not exceed 0.5%. However, the use of the RPB1

gene in studies of AMF is much more difficult than that of the SSU–ITS–LSU nrDNA segment, which is further outlined in the **Discussion** section. Nonetheless, in phylogenetic studies, it is widely recognized that the study of several loci substantially improves the reliability and resolution power of phylogenies (James et al. 2006).

Currently, the genus *Diversispora* is represented by 15 species of confirmed molecular phylogeny and four species, which Oehl et al. (2011b) included into the genus based only on morphological characters. However, the literature data and molecular sequences deposited in public databases prove that the vast majority of AMF living in the world remain unnamed and many of them belong in the genus *Diversispora* (Gamper et al. 2009; Balázs et al. 2015; Błaszowski et al. 2015b; Błaszowski, pers. observ.).

Diversispora omaniana was described from spores formed singly at the top of a cylindrical to funnel-shaped sporogenous hypha (Symanczik et al. 2014). Phylogenetic analyses of molecular sequences of selected species of the Glomeromycotina placed the fungus in a sister clade to a clade grouping four *Diversispora* spp. Unfortunately, in the analyses, no sequence of *Corymbiglomus* and *Redeckera* spp. was considered because the morphology of members of both genera and the ecology of *Redeckera* spp. differed essentially from those of *Di. omaniana* (see the section **Discussion** regarding the taxa). However, subsequent pilot molecular phylogenetic analyses suggested that *Di. omaniana* does not belong in the genus *Diversispora*, but should represent a separate clade in the Diversisporales.

In addition, we grew, in single-species cultures, an AMF of morphological features, prompting that it is an undescribed member of the genus *Diversispora*, which also confirmed the pilot molecular analyses mentioned above.

Consequently, considering the information presented above, the aims of this study were to reconstruct the true phylogeny of the “*omaniana*” fungus and describe a new species in the genus *Diversispora*.

Materials and methods

Origin of study material, establishment and growth of trap and single-species cultures, extraction of spores, and staining of mycorrhizal structures

Trap and single-species cultures of the “*omaniana*” fungus were established and grown as described by Symanczik et al. (2014).

Spores of *Di. sabulosa* were first extracted from a pot trap culture inoculated with the rhizosphere soil and root fragments of *Ammophila arenaria* (L.) Link that had colonized maritime sand dunes of the Curonian Spit, Lithuania. The spores were subsequently used to establish single-species cultures. The

trap and single-species cultures were established and grown as described by Błaszowski et al. (2012), using *Plantago lanceolata* L. as the host plant. Spores for morphological and molecular analyses and roots for studies of mycorrhizal structures were collected as described by Błaszowski et al. (2015a). Spores and roots came from five- to six-month-old cultures. Roots were stained following the protocol of Błaszowski (2012).

Microscopy and nomenclature

At least 50–100 spores of each species mounted in water, lactic acid, polyvinyl alcohol/lactic acid/glycerol (PVLG; Omar et al. 1979), and a mixture of PVLG and Melzer’s reagent (1:1, v/v) were examined to determine their morphological features and the phenotypic and histochemical characters of spore wall layers. The preparation of spores and mycorrhizal structures for study and photography were as described previously (Błaszowski 2012; Błaszowski et al. 2012). Types of spore wall layers are those defined by Błaszowski (2012), Stürmer and Morton (1997), and Walker (1983). Color names are from Kornerup and Wanscher (1983). Nomenclature of fungi and the authors of fungal names are from the Index Fungorum website (<http://www.indexfungorum.org/AuthorsOfFungalNames.htm>). Voucher specimens of the new species were mounted in PVLG and a mixture of PVLG and Melzer’s reagent (1:1, v/v) on slides and deposited at ETH in Zurich, Switzerland (Z + ZT; holotype), the Department of Ecology, Protection and Shaping of Environment (DEPSE), West Pomeranian University of Technology, Szczecin, and in the herbarium at Oregon State University (OSC) in Corvallis, Oregon, USA (isotypes).

Molecular phylogeny, DNA extraction, polymerase chain reaction, cloning, and DNA sequencing

Crude DNA of the “*omaniana*” fungus and the new *Diversispora* sp. was extracted from eight single spores of each fungus. Details of the treatment of the spores prior to polymerase chain reactions (PCRs), the conditions, and primers used in the PCRs to obtain SSU–ITS–LSU sequences were as those described in Błaszowski et al. (2015a, b), Krüger et al. (2009), and Symanczik et al. (2014).

In order to obtain RPB1 sequences of the two fungi, nested PCRs were performed in conditions recommended and with primers designed by Stockinger et al. (2014). The first PCR was performed with the primers RPB1-DR160fmix (a,b,c,d) and RPB1-HS189GPf, and the second with RPB1-HS189GPf and RPB1-DR1210r. In the second, nested PCR, 0.5 µL of the first PCR product was used as the template. Cloning and sequencing of PCR products to obtain both types of sequences were performed as described by Błaszowski et al. (2015a).

The sequences were deposited in GenBank (MG459179–MG459217).

Sequence alignment and phylogenetic analyses

After pilot molecular phylogenetic analyses, which confirmed the supposition that the phylogenetic position of the species originally described as *Di. omaniana* is erroneous and the AMF found by us is an undescribed *Diversispora* sp., three sequence sets were assembled, two with sequences of the SSU–ITS–LSU nrDNA region and one with sequences of the RPB1 gene. Sequence identity matrices of the most closely related species to the “*omaniana*” fungus and the new *Diversispora* sp. were done using the BioEdit software (Hall 1999). All comparisons were performed on sequences of the same length.

The first SSU–ITS–LSU sequence set was established to determine the taxonomic rank of the “*omaniana*” fungus among families of the Diversisporales sensu Walker and Schüßler (2004). Sixty-six sequences of the set characterized 26 species of seven families of the Diversisporales and *G. macrocarpum* of the family Glomeraceae in the order Glomerales J.B. Morton & Benny, which served as an outgroup (Fig. 1). Most sequences (28, 2–5 each) represented 11 species known to belong in the Diversisporaceae. The “*omaniana*” fungus was represented by seven sequences.

The second SSU–ITS–LSU sequence set mainly aimed at finding the phylogenetic position of the putative new *Diversispora* sp. among described species of the *Diversispora* clade. It was also supposed to support the conclusion of the existence of a new taxon represented by the “*omaniana*” fungus in the Diversisporales. The set consisted of 78 sequences of 13 certain *Diversispora* spp., the “*omaniana*” fungus, and the newly described *Di. sabulosa*, as well as *C. corymbiforme* Błaszk. & Chwat, three *Redeckera* spp., and *Pacispora scintillans* (S.L. Rose & Trappe) Sieverd. & Oehl ex C. Walker, Vestberg & A. Schüßler, of which the latter served as an outgroup taxon (Fig. 2). Each species was represented by 1–7 sequences. All sequences used in the alignment covered the whole SSU–ITS–LSU nrDNA segment amplified by the primers of Krüger et al. (2009), except the LSU sequences of *Di. celata* C. Walker, Gamper & A. Schüßler and the SSU–ITS sequences of *R. fulvum* (Berk. & Broome) C. Walker & A. Schüßler and *R. pulvinatum* (Henn.) C. Walker & A. Schüßler.

The RPB1 set contained 20 sequences of ten *Diversispora* spp., including the new *Di. sabulosa*, four sequences of the “*omaniana*” fungus, and 2–3 sequences each of representatives of the genera *Corymbiglomus*, *Redeckera*, and *Acaulospora*, with *A. laevis* being the outgroup taxon (Fig. 3).

All sequence sets were aligned separately with MAFFT v.7 using the auto option (<http://mafft.cbrc.jp/alignment/server>). In the SSU–ITS–LSU sets, indels were coded by means of

the simple indel coding algorithm (Simmons et al. 2001) as implemented in GapCoder (Young and Healy 2003), and these binary character sets were added to the nucleotide alignments, as described and justified by Błaszkowski et al. (2015c). The RPB1 set comprised only sequences of the RPB1 gene. Bayesian (BI) phylogenetic analyses of the SSU–ITS–LSU and RPB1 sequences were conducted with MrBayes 3.1.2 (Huelsenbeck and Ronquist 2001). In all analyses, the GTR nucleotide substitution model was used, which was selected by jModelTest (Posada 2008), considering the selection of the Akaike information criterion. Four Markov chains were run for 5,000,000 generations, sampling every 100 steps, with a burn-in at 7500 sampled trees. Maximum likelihood (ML) phylogenetic analyses of sequences of the two loci were carried out with the raxmlGUI (Silvestro and Michalak 2012) implementation of RAXML (Stamatakis 2014), using the GTRGAMMA algorithm. Rapid bootstrap analysis with 1000 replicates was used to determine the support of the branches. In both BI and ML analyses of the SSU–ITS–LSU sequences, the sequence sets were divided into four partitions. The generated phylogenetic trees were visualized and edited in MEGA6 (Tamura et al. 2013).

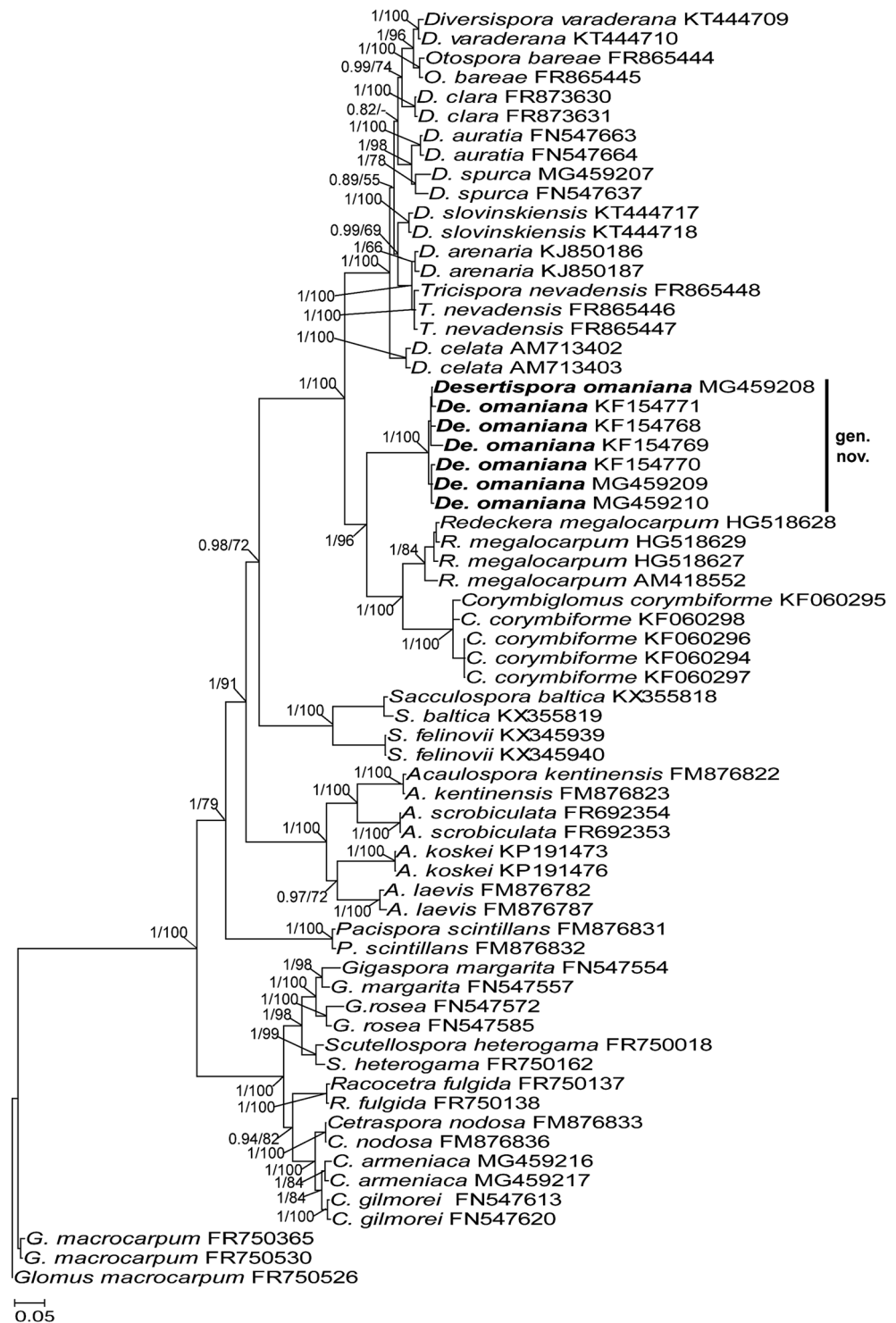
Results

General data and phylogeny

To confirm the supposition of the earlier erroneously established phylogenetic position of the species originally described as *Di. omaniana* and, consequently, to give the fungus the appropriate status within the Glomeromycotina, as well as to determine the position of the putative undescribed *Diversispora* sp. among other *Diversispora* spp. of known molecular phylogeny, three sequence sets were assembled, two with sequences of the SSU–ITS–LSU nrDNA region and one with sequences of the RPB1 gene.

Bayesian and ML analyses of SSU–ITS–LSU sequences of the first sequence set with representatives of all families of the Diversisporales sensu Walker and Schüßler (2004) grouped the “*omaniana*” fungus in a separate clade at the rank of genus, whose sister clade contained sequences of *C. corymbiforme* and *R. megalocarpum* distributed separately in two subclades, each at the rank of genus (Fig. 1). The three generic clades connected a node with a large sister clade clustering sequences of *Diversispora* spp., *O. bareae* and *T. nevadensis*, which represented the Diversisporaceae clade, together with the “*omaniana*” fungus, *C. corymbiforme*, and *R. megalocarpum*. The Diversisporaceae clade, as well as the “*omaniana*” and *C. corymbiforme* clades, were fully supported in both analyses (BI = 1.0, ML = 100%). The supports of the *R. megalocarpum* clade and the node connecting the *C. corymbiforme* and *R. megalocarpum* clades with the

Fig. 1 A 50% majority rule consensus phylogram inferred from a Bayesian analysis of SSU–ITS–LSU nrDNA sequences of *Desertispora omaniana* comb. nov., 25 other species of arbuscular mycorrhizal fungi (AMF) of the order Diversisporales, and *Glomus macrocarpum* as the outgroup. The new combination is in **bold** and followed by its GenBank accession numbers. The Bayesian posterior probabilities ≥ 0.50 and maximum likelihood (ML) bootstrap values $\geq 50\%$ are shown near the branches, respectively. The bar indicates 0.05 expected change per site per branch

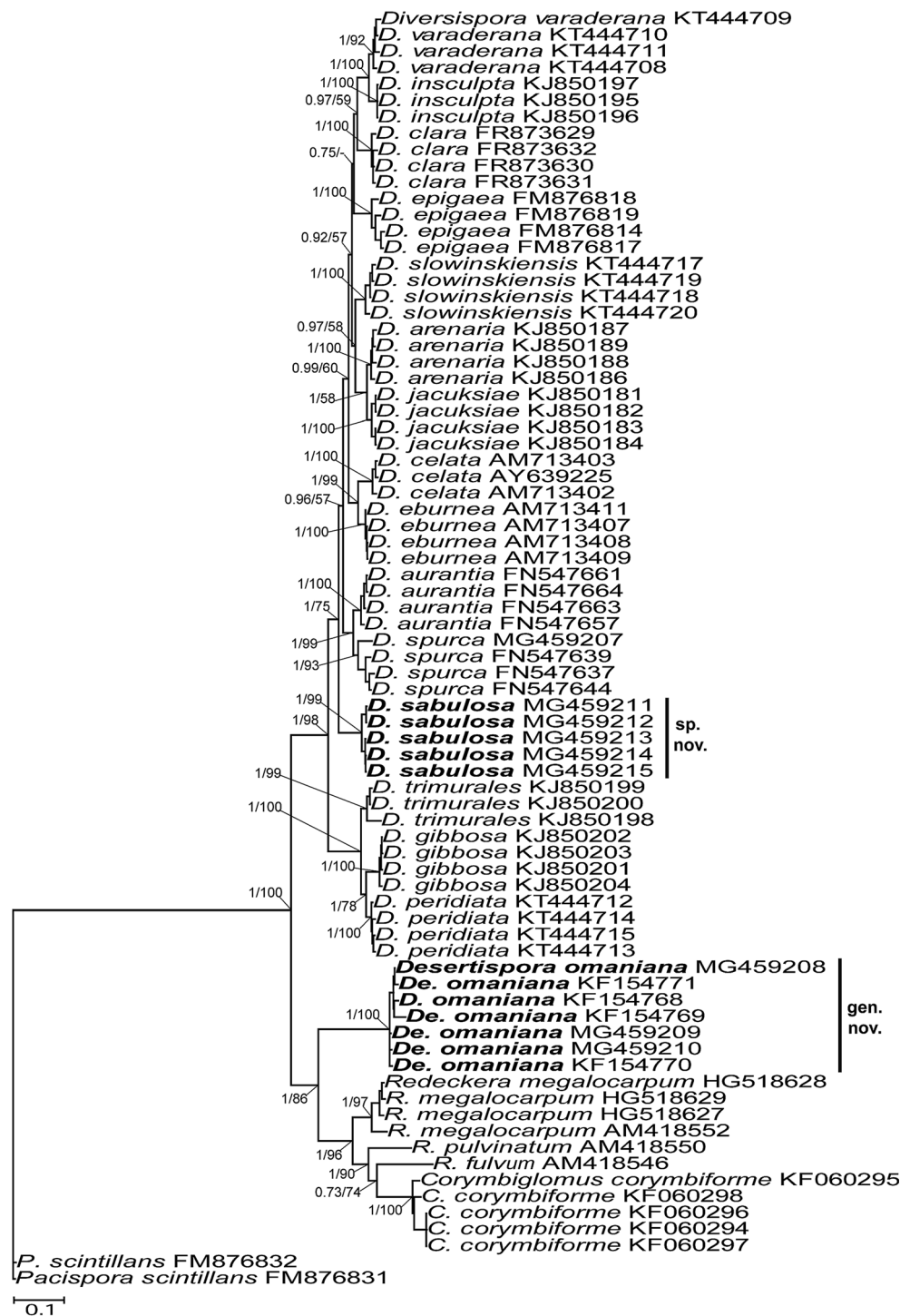


“*omaniana*” clade were also full and strong. The alignment of the sequence set had a length of 2057 characters, of which 72.2% were phylogenetic informative. The identity values of the seven SSU–ITS–LSU sequences of the “*omaniana*” fungus calculated with BioEdit (Hall 1999) were 96.7–99.4%.

Bayesian and ML analyses of the second set with SSU–ITS–LSU sequences confirmed the novelty of the

Diversispora sp. tested in this study and placed the species in an own clade positioned below a clade with *Di. aurantia* (Błaszk. et al.) C. Walker & A. Schüßler and *D. spurca* distributed in two subclades and above a clade with three monophyletic groups represented by *Di. gibbosa*, *Di. peridiata* Błaszk. et al., and *Di. trimurales* (Koske & Halvorson) C. Walker & A. Schüßler (Fig. 2). The *Di. sabulosa* clade was

Fig. 2 A 50% majority rule consensus phylogram inferred from a Bayesian analysis of SSU–ITS–LSU nrDNA sequences of *Desertispora omaniana* comb. nov., *Desertispora sabulosa* sp. nov., 18 other species of AMF of the order Diversisporales, and *Pacispora scintillans* as the outgroup. The new combination and new species are in **bold** and followed by their GenBank accession numbers. The Bayesian posterior probabilities ≥ 0.50 and ML bootstrap values $\geq 50\%$ are shown near the branches, respectively. The bar indicates 0.1 expected change per site per branch

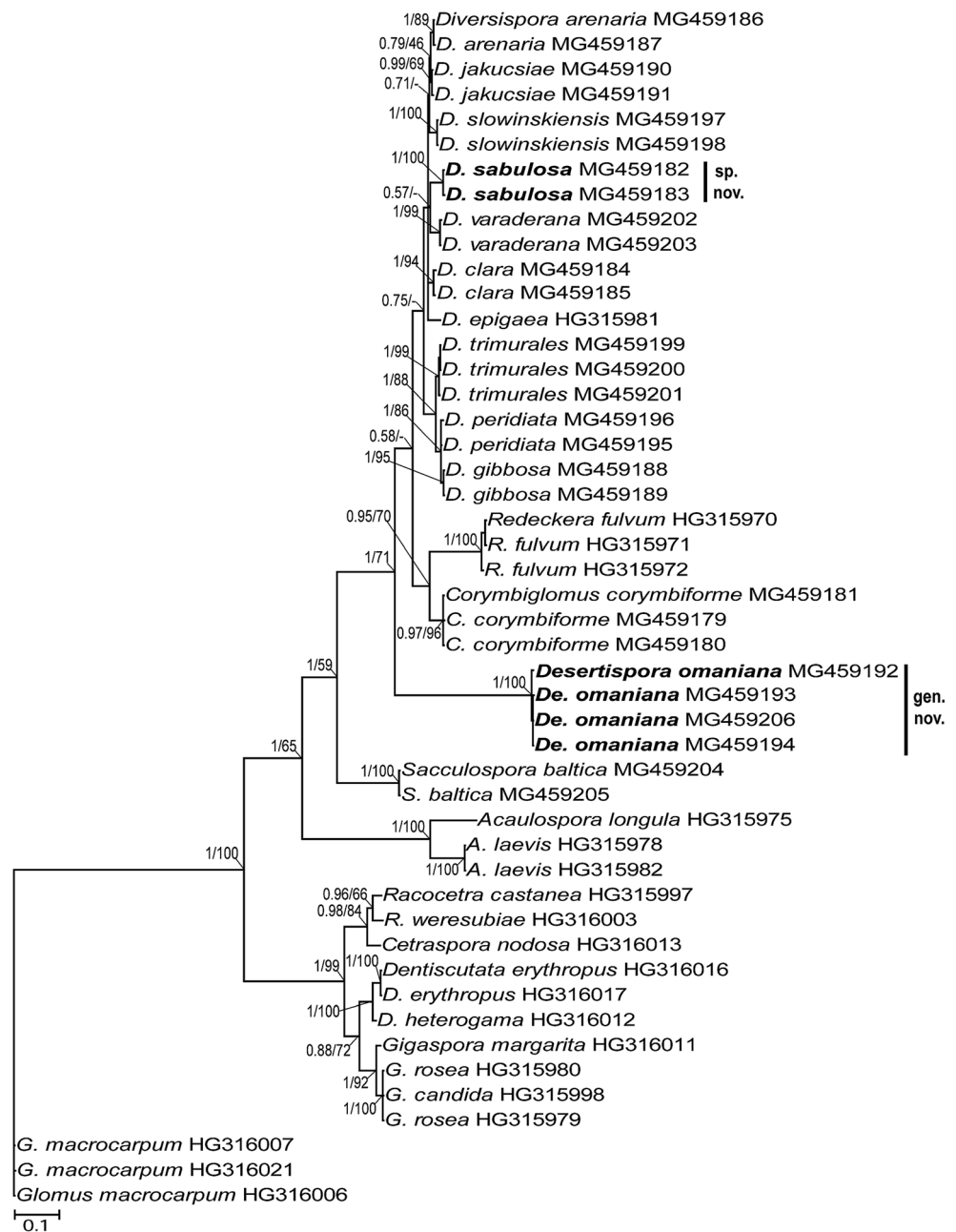


fully (BI = 1.0) and highly (ML = 99%) supported in the analyses. The identity values of the five SSU–ITS–LSU sequences of *Di. sabulosa* ranged from 98.0% to 99.8%.

Also, the analyses of the second sequence set proved that the basal AMF group of the Diversisporaceae consists of two sister genera *Corymbiglomus* and *Redeckera*, and a new genus represented by the “*omaniana*” fungus (Fig. 2). The “*omaniana*” clade was fully supported in both BI and ML

analyses. The large clade *C. corymbiforme*/*Redeckera* spp./“*omaniana*”, as well as the clades *C. corymbiforme* only and *Redeckera* spp. only, also obtained full or high support values. Importantly, the node connecting the *C. corymbiforme*/*Redeckera* spp./“*omaniana*” clade with the clade comprising all *Diversispora* spp. of known molecular phylogeny, and, thereby, representing the family Diversisporaceae, was also fully supported in both analyses.

Fig. 3 A 50% majority rule consensus phylogram inferred from a Bayesian analysis of RPB1 sequences of *Desertispora omaniana* comb. nov., *Desertispora sabulosa* sp. nov., 22 other species of AMF of the order Diversisporales, and *Glomus macrocarpum* as the outgroup. The new combination and species are in **bold** and followed by their GenBank accession numbers. The Bayesian posterior probabilities ≥ 0.50 and ML bootstrap values $\geq 50\%$ are shown near the branches, respectively. The bar indicates 0.1 expected change per site per branch



The alignment had a length of 1927 characters, of which 80.38% were phylogenetic informative.

In the tree with RPB1 sequences, the new *Di. sabulosa* was also located in a separate clade, fully supported in BI and ML analyses, but the clade was embedded between a clade with *Di. arenaria* and *Di. jakucsiae* distributed in two well-supported subclades and a fully supported clade with *Di. slowinskiensis* (Fig. 3). The positions of the “*omaniana*” clade and the clades with *C. corymbiforme* and *R. fulvum* relative to each other and the other analyzed members of the Diversisporaceae were identical to the positions of the taxa visualized in the SSU–ITS–LSU trees (Figs. 1 and 2). In both BI and ML analyses, the “*omaniana*” clade received full

support values. The clades with *C. corymbiforme* and *R. fulvum* were fully or very strongly supported, and the support values of the Diversisporaceae clade were 1.0 and 71% in BI and ML analyses, respectively. The length of the alignment was 829 characters, of which 71.41% were phylogenetic informative. The identity values of the two RPB1 sequences of *Di. sabulosa* and the four RPB1 sequences of the “*omaniana*” fungus were 99.8% and 99.3–99.8%, respectively.

The identities of the SSU–ITS–LSU sequences of “*omaniana*” vs. *C. corymbiforme*, “*omaniana*” vs. *R. megalocarpum*, and *C. corymbiforme* vs. *R. megalocarpum* calculated with BioEdit (Hall 1999) were 79.7–81.8%, 83.0–83.8%, and 86.4–87.6%, respectively. Of

the *Diversispora* spp. most closely related to *Di. sabulosa* sp. nov. (see above and Fig. 2), *Di. aurantia* had the highest identity of the SSU–ITS–LSU sequences: 92.2–92.7%.

The identities of the RPB1 sequences of “*omaniana*” vs. *C. corymbiforme*, “*omaniana*” vs. *R. fulvum*, and *C. corymbiforme* vs. *R. fulvum* were 84.6–84.9%, 81.4–82.1%, and 92.0–92.5%, respectively. The identities of the RPB1 sequences of the new *Diversispora* sp. and its closest relatives *Di. arenaria*, *Di. jakucsiae*, and *Di. slowinskiensis* (Fig. 3) ranged from 96.1% (*Di. arenaria*) to 96.7% (*Di. jakucsiae*).

Thus, the phylogenetic analyses discussed above unambiguously proved that the “*omaniana*” fungus should represent a new taxon at the rank of genus belonging to the family Diversisporaceae and the AMF found by us is a new species in the genus *Diversispora*. Therefore, the taxa are described below as *Desertispora* gen. nov., with *De. omaniana* comb. nov., and *Di. sabulosa* sp. nov.

Taxonomy

Erection of a new genus

Desertispora Błaszk., Kozłowska, Ryszka, Al-Yahya’ei & Symanczik, gen. nov., MB 823563. Figs. 4, 5

Type species: Desertispora omaniana (Symanczik, Błaszk. & Al-Yahya’ei) Symanczik, Błaszk., Kozłowska & Al-Yahya’ei.

Etymology: Desertispora, referring to the desert habitat in which the fungus was originally found.

Diagnosis: Differs from other genera in the Diversisporaceae and Diversisporales, whose species form spores at the tip of a cylindrical or funnel-shaped sporogenous hypha, in the possession of a spore wall with a structural laminate layer reacting in Melzer’s reagent, and in having the specific sequences of the nrDNA regions of ITS1: GAAATTTAATTCCCGGGAATCTTGTTCCCG, ITS2: AAAATGAATTCTGGTTCAAGTCAAAAACGTTC TATGC and TCATTATGTTCCACCT, and the LSU gene: CGAAGTCCAATCAGATACGCTCTCG and ACATTCGT GGGTCTTGACGACCACGGTTAGAGCGTTC, as well as the RPB1 gene: for example, TCTCTTCTTCCC TCTCTTCTTCTTCTTCC, CAAATTCATCATG TTTTCATTACTTATTCATATCACGCGGAAAAT, and CTCCACCCACCCCGGAGCCAAGTACGTGATTA GGGATACCGGTGAAAGGATCGAC.

Description: Spores formed singly in soil, develop blastically at the tip of sporogenous hyphae. Spores with one subtending hypha. Spore wall composed of three layers (layers 1–3). Layer 1 evanescent, roughened, hyaline. Layer 2 permanent, pliable, uniform (not composed of sublayers), smooth, hyaline to pastel yellow (2A4). Layer 3 laminate, smooth, hyaline. In Melzer’s reagent, layer 1 always remains

nonreactive, layer 2 usually darkens to pale yellow (3A3), and layer 3 usually stains pinkish (9A2) to dull red (11C3). *Subtending hypha* cylindrical to funnel-shaped, rarely constricted at the spore base, not breaking up in crushed spores. *Wall of subtending hypha* composed of three layers continuous with spore wall layers 1–3. *Pore* open or occluded by a curved septum continuous with some innermost laminae of spore wall layer 3; septum positioned up to 15.7 µm below the spore base. Spore contents of a hyaline to brownish yellow (5C8), sticky, opaque substance.

Desertispora omaniana (Symanczik, Błaszk. & Al-Yahya’ei) Symanczik, Błaszk., Kozłowska & Al-Yahya’ei, comb. nov., MB 823564. Figs. 4, 5

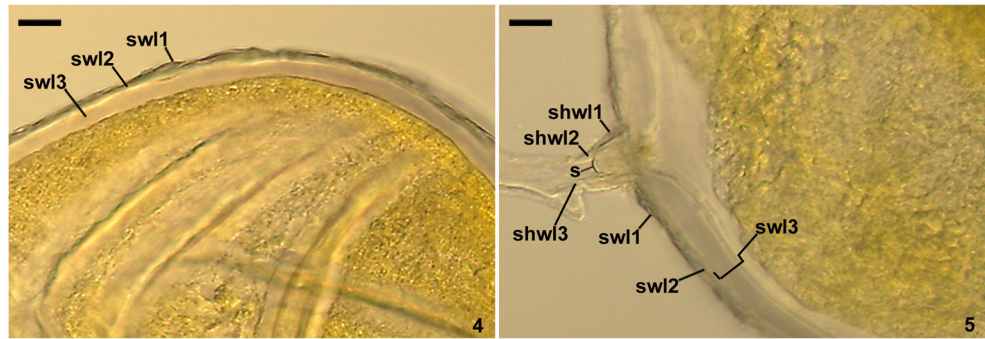
Basionym: Diversispora omaniana Symanczik, Błaszk. & Al-Yahya’ei. Mycologia 106: 247, 2014.

Spores formed singly in soil, develop blastically at the tip of sporogenous hyphae. Spores hyaline to brownish yellow (5C8); globose to subglobose; (85–)136(–170) µm diam.; rarely egg-shaped; 110–160 × 130–180 µm; with one subtending hypha. Spore wall composed of three layers (layers 1–3; Figs. 4, 5). Layer 1 evanescent, roughened, hyaline, (1.0–)1.9(–2.8) µm thick when intact, usually more or less deteriorated in mature spores, frequently partly or completely sloughed in older specimens. Layer 2 permanent, pliable, uniform (not composed of sublayers), smooth, hyaline to pastel yellow (2A4), (1.5–)2.4(–3.8) µm thick. Layer 3 laminate, smooth, hyaline, (4.8–)8.9(–22.0) µm thick, frequently stratifying into groups of or single laminae in crushed spores. In Melzer’s reagent, layer 1 always remains nonreactive, layer 2 usually darkens to pale yellow (3A3), and layer 3 usually stains pinkish (9A2) to dull red (11C3; Fig. 4). *Subtending hypha* hyaline to pastel yellow (2A4); cylindrical to funnel-shaped, rarely constricted at the spore base; (10.8–)14.3(–23.5) µm wide at the spore base, not breaking up in crushed spores (Fig. 5). *Wall of subtending hypha* hyaline to pastel yellow (2A4); (2.3–)4.2(–5.5) µm thick at the spore base; composed of three layers continuous with spore wall layers 1–3 (Fig. 5). *Pore* (3.0–)7.1(–14.3) µm diam., open or occluded by a curved septum, 1.0–1.3 mm thick, continuous with some innermost laminae of spore wall layer 3; septum positioned up to 15.7 µm below the spore base (Fig. 5). Spore content of a hyaline to brownish yellow (5C8), sticky, opaque substance (Figs. 4, 5).

Specimens examined: Holotype: 3222 (DEPSE), *isotypes:* 3221, 3223–3243, and two slides at OSC (Symanczik et al. 2014).

Distribution and habitat: To date, found only based on spores extracted from trap cultures inoculated with the rhizosphere soils of *Salvadora persica* Wall., *Tetraena qatarense* Beier & Thulin. and unrecognized grasses, in undisturbed natural field located at Al-Kamel in the Al-Sharqya region of Oman (22°14′911″N, 59°10′53″E; Symanczik et al.

Figs. 4, 5 Spores of *Desertispora omaniana*. 4. Spore wall layers (swl) 1–3. 5. Subtending hyphal wall layers (shwl) 1–3 continuous with spore wall layers (swl) 1–3 and septum (s) in the lumen of the subtending hypha. 4. Spore in PVLG + Melzer's reagent. 5. Spore in PVLG. 4, 5. Differential interference microscopy. Bars: 4, 5 = 10 μm



2014). BLAST queries did not show any SSU–ITS–LSU sequence of identity at least 97% to the SSU–ITS–LSU sequences of *De. omaniana*. The highest identity of the RPB1 sequences of *De. omaniana* was found in comparison with the RPB1 HG315981 sequence of *Di. epigaea*, which was only 86%.

Description of a new species

Diversispora sabulosa Błaszcz. & Kozłowska, sp. nov., MB 823568. Figs. 6–13

Holotype: ZT Myc 58,675 (Z + ZT), isotypes: 3593–3601 (DEPSE), and OSC 161509, OSC 161510 (OSC).

Etymology: Latin, *sabulosa*, referring to the sandy habitat in which the fungus exists.

Sporocarps unknown. Spores formed singly in soil (Figs. 6–12); arise blastically at the tip of sporogenous hyphae developed from mycorrhizal extraradical hyphae. **Spores** deep yellow (4A8) to orange yellow (4B8), rarely brownish yellow (5C8); globose to subglobose; (17–)51(–67) μm diam.; frequently ovoid; 80–110 \times 95–130 μm ; with one subtending hypha (Figs. 6–12). **Spore wall** consists of two layers (Figs. 7–12). Layer 1, forming the spore surface, uniform (not divided into visible sublayers), permanent, semi-flexible, hyaline to light yellow (4A5), (1.2–)2.5(–3.6) μm thick, usually tightly adherent to the upper surface of layer 2, sometimes separating from it, especially in vigorously crushed spores (Figs. 7–12). Layer 2 laminate, permanent, smooth, deep yellow (4A8) to orange yellow (4B8), rarely brownish yellow (5C8), (3.8–)5.2(–7.5) μm thick, consisting of very thin, < 0.5 μm thick, laminae, tightly adherent to each other (Figs. 7–12). Layers 1 and 2 do not show amyloid or dextrinoid reaction in Melzer's reagent (Figs. 9, 11, 12). **Subtending hypha** hyaline to pale yellow (3A3), rarely light yellow (4A4); straight or recurved, cylindrical to slightly funnel-shaped, sometimes constricted at the spore base; (5.6–)6.9(–8.5) μm wide at the spore base (Figs. 10–12). **Wall of subtending hypha** hyaline to pale yellow (3A3), rarely light yellow (4A4); (1.6–)2.5(–4.3) μm thick at the spore base; continuous with spore wall layers 1 and 2 (Figs. 10–12). **Pore** (1.4–)2.4(–3.0) μm diam., open or occluded by a straight

or curved septum, formed by subtending hyphal wall layer 2; septum positioned up to 5.4 μm below the spore base (Figs. 10–12). **Germination** unknown.

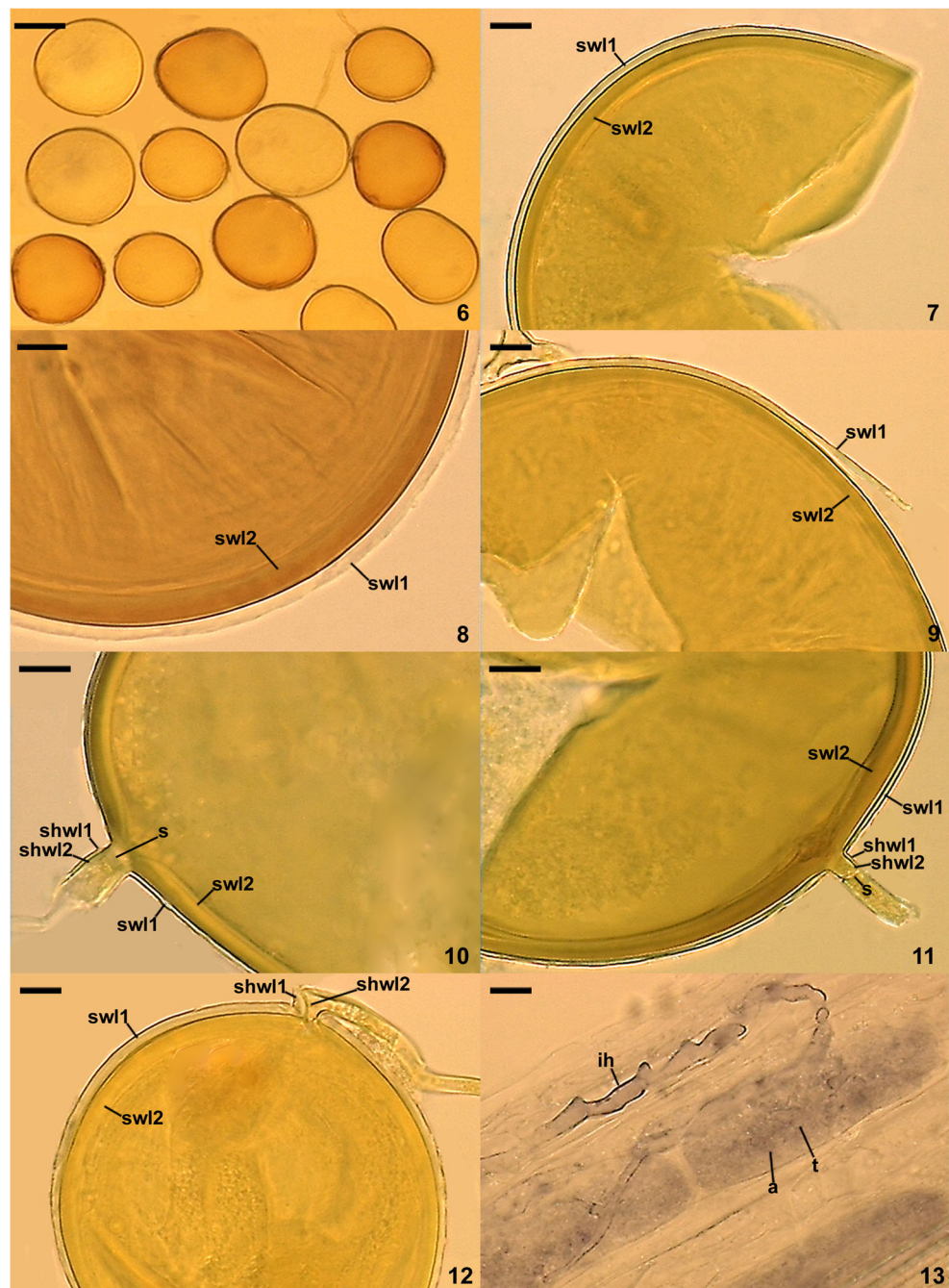
Mycorrhizal associations: The presence of spores of *Di. sabulosa* in a trap culture inoculated with the rhizosphere soil of *Ammophila arenaria* (L.) Link suggested that the AMF forms a symbiotic association with the plant. However, no molecular analysis was performed on its roots to confirm the supposition. The field soil \times root sample was collected on 31 September 2013.

In single-species cultures with *P. lanceolata* as the host plant, *Di. sabulosa* formed mycorrhiza with arbuscules and intraradical and extraradical hyphae (Fig. 13). No vesicles were found. Arbuscules were uniformly distributed along the root fragments examined (Fig. 13). Intraradical hyphae were straight, slightly recurved, or formed Y-shaped branches and coils (Fig. 13). Extraradical hyphae occurred infrequently or abundantly, depending on the root fragment examined. In 0.1% Trypan blue, all of the structures stained faintly [violet white (15A2) to pale violet (15A3); Fig. 13].

Distribution and habitat: Despite that J. Błaszczowski personally examined the occurrence of AMF in ca. 2500 field-collected rhizosphere soil samples and ca. 3000 trap cultures inoculated with cultivated and natural soils collected in different regions of Africa, Asia, Brazil, Europe, and the USA, *Di. sabulosa* was found only in one trap culture representing mobile maritime dunes of the Curonian Spit located in the north of Lithuania (55°26'N, 21°04'E). The dunes are the highest sand dunes in Europe: their average height is 35 m, but some attain a height of 60 m (https://en.wikipedia.org/wiki/Curonian_Spit). Since 2000, the Curonian Spit has been on UNESCO's World Heritage List under cultural criteria "V". The vegetation on the Curonian Spit is varied and unique due to the specific climatic conditions of the sea coast (http://daugenis.mch.mii.lt/apsamogitia/LANKYTINOS_VIETOS/nerija.en.htm). About 700 plant species were found on the spit, among which the prevailing plants are those well adapted to infertile and unstable sands, such as, for example, *A. arenaria*.

BLAST queries confirmed the rare occurrence of *Di. sabulosa* in the world determined on the basis of studies of

Figs. 6–13 Spores and mycorrhizal structures of *Diversispora sabulosa*. 6. Intact spores. 7–9. Spore wall layers (swl) 1 and 2. 10, 11. Spore wall layers (swl) 1 and 2 continuous with wall layers (shwl) 1 and 2 of the cylindrical subtending hypha and a septum (s) enclosing its lumen. 12. Spore wall layers (swl) 1 and 2 continuous with wall layers (shwl) 1 and 2 of the subtending hypha constricted at the spore base. 13. Arbuscules (a) with a trunk (t) and intraradical hypha (ih). 6–8, 10, 13. Spores in PVLG. 9, 11, 12. Spores in PVLG + Melzer's reagent. 6–13. Differential interference microscopy. Bars: 6 = 50 μ m, 7–13 = 10 μ m



the presence of spores of AMF in field soils and trap cultures: in public databases, there is no sequence of the SSU-ITS-LSU nrDNA segment and the RPB1 gene of identity $\geq 97\%$.

Discussion

The investigations discussed in this paper confirmed our suppositions that the AMF originally described as *Di. omaniana* does not belong in the genus *Diversispora*, but represents a new taxon in the family Diversisporaceae (Figs. 1, 2, and 3),

which was described here as *Desertispora* gen. nov. These studies also proved that the putative undescribed *Diversispora* sp. that we found in maritime sand dunes of the Curonian Spit, Lithuania, is a new species of the genus (Figs. 2 and 3), here named *Di. sabulosa*.

Despite that the features of *De. omaniana*, especially the histochemical traits of its spores, do not fit the characters defining the genus *Diversispora* and other clades of the order Diversisporales comprising species producing spores terminally on a cylindrical or funnel-shaped sporogenous hypha (Oehl et al. 2011b; Schüßler and Walker 2010; Symanczik

et al. 2014) and *Di. sabulosa* in morphology is unique compared to the other *Diversispora* spp., the certain determination of the phylogenetic position of the two species within the Diversisporales was possible only after molecular phylogenetic analyses of their sequences. The analyses proved that the new genus *Desertispora* is most closely related to the genera *Corymbiglomus* and *Redeckera* and showed the closest relatives of *Di. sabulosa* (Figs. 1, 2, and 3).

Morphologically, *De. omaniana* differs clearly from *Corymbiglomus* spp. and *Redeckera* spp. *Corymbiglomus corymbiforme*, whose sequences were used in the molecular phylogenetic analyses discussed here (Figs. 1, 2, and 3), forms spores mainly in corymbiform clusters, rarely singly (Błaszowski 1995, 2012), whereas spores of *De. omaniana* arise only singly (Figs. 4, 5; Symanczik et al. 2014). Moreover, *C. corymbiforme* spores are surrounded by a hyphal mantle, their spore wall layer 3 is flexible to semi-flexible, and none of the three spore wall layers stains in Melzer's reagent. In *De. omaniana*, spores are naked (not covered by hyphae; Figs. 4, 5), and their laminate spore wall 3 usually stains pinkish (9A2) to dull red (11C3) in Melzer's reagent (Fig. 4). Spores of *C. globiferum* (Koske & C. Walker) Błasz. & Chwat and *C. tortuosum* (N.C. Schenck & G.S. Sm.) Błasz. & Chwat are also covered with a peridium, do not react in Melzer's reagent, and their spore wall is one-layered (*C. tortuosum*) or three-layered with a semi-flexible layer 3 (*C. globiferum*; Koske and Walker 1986; Błaszowski 2012). The last of the described *Corymbiglomus* spp., *C. pacificum* Oehl et al., produces single naked spores, thus similarly as *De. omaniana* does (Figs. 4, 5), but spores of the former species have a more complex subcellular structure, which, according to Medina et al. (2014), consists of two walls, each with three layers.

Similarly as in comparison to *C. corymbiforme*, *De. omaniana* differs substantially from *Redeckera* spp. All of the known *Redeckera* spp. have been found to form spores mainly in epigeous compact sporocarps with a peridium (Gerdemann and Trappe 1974; Redecker et al. 2007), rarely singly (Oehl et al. 2011b). The ability to form mycorrhiza by *Redeckera* spp. remains unknown because many attempts to grow them in culture have failed (Redecker et al. 2007). The spore wall of *Redeckera* spp. consists of only one laminate layer not reacting in Melzer's reagent. Their spore subtending hypha, whose wall is one-layered, is usually fragile, inflated slightly below the spore base, and its lumen is closed by a straight, broad septum positioned at the level of the spore base; it is rarely open (Gerdemann and Trappe 1974; Redecker et al. 2007; Oehl et al. 2011b; Błaszowski 2012). In contrast, *De. omaniana* produced only hypogeous ectocarpic spores (Figs. 4, 5) and the species was grown in single-species cultures, which were established from only one spore (Symanczik et al. 2014). The spore wall of *De. omaniana* comprises three layers (Figs. 4, 5), of which the

laminate layer 3 usually stains in Melzer's reagent (Fig. 4). The subtending hypha of *De. omaniana* is persistent, consists of three layers continuous with spore wall layers 1–3, is cylindrical to funnel-shaped without an inflation, and its lumen is closed by a curved septum positioned up to 15.7 µm below the spore base (Fig. 5); in young specimens, it is frequently open. Probably, the only characters uniting *De. omaniana* and *Redeckera* spp. are those of the spore content. In *De. omaniana* (Figs. 4, 5) and, for example, in *R. fulvum* and *R. megalocarpum*, the spore content is colorless to yellow-colored, sticky, and opaque (Błaszowski 2012).

Apart from the clear morphological differences discussed above, *De. omaniana*, *Corymbiglomus* spp., and *Redeckera* spp. are separated by a large molecular distance (see the above section entitled **General data and phylogeny**; Błaszowski 2012; Błaszowski and Chwat 2013).

The most distinctive morphological character of *Di. sabulosa* is its spore wall layer 1, which is permanent, uniform, usually colorless, and relatively thick when compared with the laminate spore wall layer 2 (Figs. 7–12).

The separateness of *Di. sabulosa* unambiguously proved the performed molecular phylogenetic analyses, particularly those of sequences of the SSU–ITS–LSU nrDNA segment, which considered all *Diversispora* spp. of known natural phylogeny. The analyses clustered *Di. sabulosa* sequences in a separate clade, nested between a clade with *Di. aurantia* and *Di. spurca*, and a clade with sequences of *Di. gibbosa*, *Di. peridiata*, and *Di. trimurales* (Fig. 2). However, of the five *Diversispora* spp., only *Di. peridiata* produces spores with a two-layered spore wall and the two layers are permanent (Błaszowski et al. 2015b). Several morphological characters clearly distinguish *Di. sabulosa* from *Di. peridiata*. Apart from single spores, *Di. peridiata* produces spores in clusters and the spores are occasionally covered with peridium-like hyphae. *Diversispora sabulosa* forms only single spores without a peridium (Figs. 6–12). Spores of *Di. peridiata* are 1.5–2.1-fold larger when globose, and their spore wall layer 1 and the subtending hyphal wall at the spore base are 1.5–1.7-fold and 1.4–2.8-fold thicker, respectively. In addition, spores of the species are lighter [yellowish white (4A2) to maize yellow (4A6)], and their spore wall layer 1 may be much darker [brownish yellow (5C8)] and is never as hyaline as in *D. sabulosa* (Figs. 7–12).

The BI and ML phylogenetic analyses with sequences of the RPBI gene also located *Di. sabulosa* in a separate, fully supported clade, but the clade was embedded between a clade with *Di. arenaria* and *Di. jakucsiae* distributed in two subclades and a clade with *Di. slowinskiensis* (Fig. 3), which, in the SSU–ITS–LSU tree, were placed far from the new species (Fig. 2). However, these relationships need to be treated with caution because the RPBI analyses did not consider sequences of *Di. aurantia* and *Di. spurca* included in the SSU–ITS–LSU analyses (Figs. 1 and 2); many attempts to obtain

them failed. Nevertheless, morphologically, *Di. sabulosa* differs substantially from *Di. arenaria*, *Di. jakucsiae*, and *Di. slowinskiensis*, for example, in the spore wall structure, which is two-layered in the former new AMF (Figs. 7–12), and three-layered in the three latter species (Błaszowski et al. 2001, 2015b; Balázs et al. 2015). The molecular distance between the species is also large (see the above section entitled **General data and phylogeny**).

As mentioned in the **Introduction** section, the use of the RPB1 gene in studies of AMF still encounters many limitations. Unfortunately, the primers designed by Stockinger et al. (2014) do not amplify all clades of the subphylum, and many of them either amplify only short or much shorter than expected fragments of the RPB1 gene, such as, for example, those recommended for *Diversispora* spp., or do not work at all (pers. observ.). However, the limited usability of the primers mainly results from that they amplify only particular and not all clades of the Glomeromycotina. Thus, the recognition of molecular properties of targeted AMF based on the RPB1 gene has to be preceded by the identification of their position within the subphylum, using, for example, the primers by Krüger et al. (2009).

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References

- Ames RN, Schneider RW (1979) *Entrophospora*, a new genus in the Endogonaceae. *Mycotaxon* 8:347–352
- Balázs TK, Błaszowski J, Chwat G, Góralska A, Gáspár BK, Lukács AF, Kovács GM (2015) Spore-based study of arbuscular mycorrhizal fungi of semiarid sandy areas in Hungary, with *Diversispora jakucsiae* sp. nov. *Mycol Prog* 14:1021. <https://doi.org/10.1007/s11557-014-1021-z>
- Błaszowski J (1995) *Glomus corymbiforme*, a new species in Glomales from Poland. *Mycologia* 87:732–737. <https://doi.org/10.2307/3760819>
- Błaszowski J (2012) Glomeromycota. W. Szafer Institute of Botany, Polish Academy of Sciences, Kraków
- Błaszowski J, Chwat G (2013) *Septoglomus deserticola* emended and new combinations in the emended definition of the family Diversisporaceae. *Acta Mycol* 48(1):89–103. <https://doi.org/10.5586/am.2013.011>
- Błaszowski J, Tadych M, Madej T (2001) *Glomus arenarium*, a new species in Glomales (Zygomycetes). *Acta Soc Bot Pol* 70:97–101. <https://doi.org/10.5586/asbp.2001.013>
- Błaszowski J, Kovács GM, Gáspár BK, Balázs TK, Buscot F, Ryszka P (2012) The arbuscular mycorrhizal *Paraglomus majewskii* sp. nov. represents a distinct basal lineage in Glomeromycota. *Mycologia* 104:148–156. <https://doi.org/10.3852/10-430>
- Błaszowski J, Chwat G, Symanczik S, Góralska A (2015a) *Dominikia duoreactiva* sp. nov. and *Dominikia difficilevidera* sp. nov., two new species in the Glomeromycota. *Botany* 93:389–396. <https://doi.org/10.1139/cjb-2015-0016>
- Błaszowski J, Furrázola E, Chwat G, Góralska A, Lukács AF, Kovács GM (2015b) Three new arbuscular mycorrhizal *Diversispora* species in Glomeromycota. *Mycol Prog* 14:105. <https://doi.org/10.1007/s11557-015-1122-3>
- Błaszowski J, Chwat G, Góralska A, Ryszka P, Kovács GM (2015c) Two new genera, *Dominikia* and *Kamienskia*, and *D. disticha* sp. nov. in Glomeromycota. *Nova Hedwigia* 100:225–238. https://doi.org/10.1127/nova_hedwigia/2014/0216
- Castillo CG, Borie F, Oehl F, Sieverding E (2016) Arbuscular mycorrhizal fungi biodiversity: prospecting in Southern-Central zone of Chile. A review. *J Soil Sci Plant Nutr* 16:400–422. <https://doi.org/10.4067/S0718-95162016005000036>
- Gamper HA, Walker C, Schüßler A (2009) *Diversispora celata* sp. nov.: molecular ecology and phylotaxonomy of an inconspicuous arbuscular mycorrhizal fungus. *New Phytol* 182:495–506. <https://doi.org/10.1111/j.1469-8137.2008.02750.x>
- Gerdemann JW, Trappe JM (1974) The Endogonaceae in the Pacific Northwest. *Mycol Mem* 5:1–76
- Hall TA (1999) BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symp Ser* 41:95–98
- Huelsenbeck JP, Ronquist F (2001) MRBAYES: Bayesian inference of phylogenetic trees. *Bioinformatics* 17:754–755. <https://doi.org/10.1093/bioinformatics/17.8.754>
- James TY, Kauff F, Schoch CL, Matheny PB, Hofstetter V, Cox CJ et al (2006) Reconstructing the early evolution of Fungi using a six-gene phylogeny. *Nature* 443:818–822. <https://doi.org/10.1038/nature05110>
- Kornerup A, Wanscher JH (1983) *Methuen handbook of colour*, 3rd edn. E. Methuen, London
- Koske RE, Walker C (1986) *Glomus globiferum*: a new species of Endogonaceae with a hyphal peridium. *Mycotaxon* 26:133–142
- Krüger M, Stockinger H, Krüger C, Schüßler A (2009) DNA-based species level detection of Glomeromycota: one PCR primer set for all arbuscular mycorrhizal fungi. *New Phytol* 183:212–223. <https://doi.org/10.1111/j.1469-8137.2009.02835.x>
- Medina J, Cornejo P, Borie F, Meier S, Palenzuela J, Vieira HEE, Ferreira ACA, da Silva GA, Sánchez-Castro I, Oehl F (2014) *Corymbiglomus pacificum*, a new glomeromycete from a saline lakeshore in Chile. *Mycotaxon* 127:173–183. <https://doi.org/10.5248/127.173>
- Oehl F, da Silva GA, Goto BT, Maia LC, Sieverding E (2011a) Glomeromycota: two new classes and a new order. *Mycotaxon* 116:365–379. <https://doi.org/10.5248/116.365>
- Oehl F, da Silva GA, Goto BT, Sieverding E (2011b) Glomeromycota: three new genera and glomoid species reorganized. *Mycotaxon* 116:75–120. <https://doi.org/10.5248/116.75>
- Oehl F, da Silva GA, Sánchez-Castro I, Goto BT, Maia LC, Vieira HEE, Barea J-M, Sieverding E, Palenzuela J (2011c) Revision of Glomeromycetes with entrophosporoid and glomoid spore formation with three new genera. *Mycotaxon* 117:297–316. <https://doi.org/10.5248/117.297>
- Omar MB, Bolland L, Heather WA (1979) A permanent mounting medium for fungi. *Bull Br Mycol Soc* 13:31–32. [https://doi.org/10.1016/S0007-1528\(79\)80038-3](https://doi.org/10.1016/S0007-1528(79)80038-3)
- Pagano MC, Oehl F, Silva GA, Maia LC, Silva DK, Cabello MN (2016) Advances in arbuscular mycorrhizal taxonomy. In: Pagano M (ed) Recent advances on mycorrhizal fungi. Springer International Publishing, Switzerland, pp 15–21. https://doi.org/10.1007/978-3-319-24355-9_2

- Palenzuela J, Ferrol N, Boller T, Azcón-Aguilar C, Oehl F (2008) *Otospora bareai*, a new fungal species in the Glomeromycetes from a dolomitic shrub land in Sierra de Baza National Park (Granada, Spain). *Mycologia* 100:296–305. <https://doi.org/10.1080/15572536.2008.11832484>
- Palenzuela J, Barea JM, Ferrol N, Azcón-Aguilar C, Oehl F (2010) *Entrophospora nevadensis*, a new arbuscular mycorrhizal fungus from Sierra Nevada National Park (southeastern Spain). *Mycologia* 102(3):624–632. <https://doi.org/10.3852/09-145>
- Posada D (2008) jModelTest: phylogenetic model averaging. *Mol Biol Evol* 25:1253–1256. <https://doi.org/10.1093/molbev/msn083>
- Redecker D, Raab PA (2006) Phylogeny of the Glomeromycota (arbuscular mycorrhizal fungi): recent developments and new gene markers. *Mycologia* 98:885–895. <https://doi.org/10.1080/15572536.2006.11832618>
- Redecker D, Raab PA, Oehl F, Camacho FJ, Courtecuisse R (2007) A novel clade of sporocarp-forming species of glomeromycotan fungi in the Diversisporales lineage. *Mycol Prog* 6:35–44. <https://doi.org/10.1007/s11557-007-0524-2>
- Redecker D, Schüßler A, Stockinger H, Stürmer SL, Morton JB, Walker C (2013) An evidence-based consensus for the classification of arbuscular mycorrhizal fungi (Glomeromycota). *Mycorrhiza* 23: 515–531. <https://doi.org/10.1007/s00572-013-0486-y>
- Schüßler A, Walker C (2010) The Glomeromycota. A species list with new families and new genera. Royal Botanic Garden Edinburgh, Gloucester
- Silvestro D, Michalak I (2012) raxmlGUI: a graphical front-end for RAxML. *Org Divers Evol* 12:335–337. <https://doi.org/10.1007/s13127-011-0056-0>
- Simmons MP, Ochoterena H, Carr TG (2001) Incorporation, relative homoplasy, and effect of gap characters in sequence-based phylogenetic analyses. *Syst Biol* 50:454–462. <https://doi.org/10.1080/106351501300318049>
- Spatafora JW, Chang Y, Benny GL, Lazarus K, Smith ME, Berbee ML, Bonito G, Corradi N, Grigoriev I, Gryganskyi A, James TY, O'Donnell K, Roberson RW, Taylor TN, Uehling J, Vilgalys R, White MM, Stajich JE (2016) A phylum-level phylogenetic classification of zygomycete fungi based on genome-scale data. *Mycologia* 108:1028–1046. <https://doi.org/10.3852/16-042>
- Stamatakis A (2014) RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* 30:1312–1313. <https://doi.org/10.1093/bioinformatics/btu033>
- Stockinger H, Walker C, Schüßler A (2009) ‘*Glomus intraradices* DAOM197198’, a model fungus in arbuscular mycorrhiza research, is not *Glomus intraradices*. *New Phytol* 183:1176–1187. <https://doi.org/10.1111/j.1469-8137.2009.02874.x>
- Stockinger H, Peyret-Guzzon M, Koenig S, Bouffaud M-L, Redecker D (2014) The largest subunit of RNA polymerase II as a new marker gene to study assemblages of arbuscular mycorrhizal fungi in the field. *PLoS One* 9(10):e107783. <https://doi.org/10.1371/journal.pone.0107783>
- Stürmer SL, Morton JB (1997) Developmental patterns defining morphological characters in spores of four species in *Glomus*. *Mycologia* 89:72–81. <https://doi.org/10.2307/3761174>
- Symanczik S, Błaszczkowski J, Chwat G, Boller T, Wiemken A, Al-Yahya'ei MN (2014) Three new species of arbuscular mycorrhizal fungi discovered at one location in a desert of Oman: *Diversispora omaniana*, *Septoglomus nakheelum* and *Rhizophagus arabicus*. *Mycologia* 106(2):243–259. <https://doi.org/10.3852/106.2.243>
- Tamura K, Stecher G, Peterson D, Filipiński A, Kumar S (2013) MEGA6: molecular evolutionary genetics analysis version 6.0. *Mol Biol Evol* 30:2725–2729. <https://doi.org/10.1093/molbev/mst197>
- Walker C (1983) Taxonomic concepts in the Endogonaceae; spore wall characteristics in species descriptions. *Mycotaxon* 18:443–455
- Walker C, Schüßler A (2004) Nomenclatural clarifications and new taxa in the Glomeromycota *Pacispora*. *Mycol Res* 108:981–982. <https://doi.org/10.1017/S0953756204231173>
- Young ND, Healy J (2003) GapCoder automates the use of indel characters in phylogenetic analysis. *BMC Bioinformatics* 4:6. <https://doi.org/10.1186/1471-2105-4-6>