Influence of dietary palm oil on growth and carcass composition of *Heterobranchus longifilis*.

Danish fishment contrars approximately 10% fish oil as purchased, which m

^{1*}**Babalola, T.O.O.,** ¹**Adebayo, M.A.,** ²**Apata, D.F. and** ³**Omotosho, J.S.** ¹National Institute for Freshwater Fisheries Research, P.M.B. 6006, New-Bussa, Niger State ²Department of Animal Production, University of Ilorin, Ilorin, Kwara State. ³Department of Zoology, University of Ilorin, Ilorin, Kwara State.

r day, sub-divided into three equal feeds at 09.00, 15:00 and 21:00 h daily

ABSTRACT notice and most algorithming in original definition and the indicated by a second se

Supplies of marine fish oil (FO) are limited and continued growth in aquaculture production dictates that substitutes must be found that do not compromise fish health and growth. This study investigated the suitability of palm oil (PO) as a replacement for FO (cod liver oil) in diets of H. longifilis. Triplicate groups of H. longifilis fingerlings were fed five practical-type diets in which the added lipid was either 100% FO and 0% PO; 75% FO and 25% PO; 50% FO and 50% PO; 25% FO and 75% PO; or 0% FO and 100% PO for 8 weeks. There were no significant effects of diet on growth rate or feed conversion ratio. Lipid deposition was highest in the liver of fish fed 100% PO. No significant differences were observed between dietary treatments for moisture, protein and ash content in H. longifilis fingerlings. This study suggests that PO can be used successfully as a substitute for cod liver oil in diets for H. longifilis.

Key words: Heterobranchus longifilis, palm oil, cod liver oil, growth, carcass composition.

*Correspondence: tobabalola@yahoo.com. National Institute for Freshwater Fisheries Research, P.M.B. 6006, New-Bussa, Niger State, Nigeria.Introduction

Fish oil replacement has been given considerable attention in recent years in the light of perceived situation of world fisheries and aquaculture and potential limitations on fish oil supplies for aquaculture feeds (Caballere *et al.*, 2002; Bell *et al.*, 2003; Glencross *et al.*, 2003; Regost *et al.*, 2003). The steady increase of aquaculture production over the last two decades has resulted in an increased utilization of fish meal and fish oil. Aquafeeds currently use about 70% of the global supply of fish oil and by the year 2010, fish oil used in aquaculture is estimated to reach about 97% of the world supply (Tacon 2003). Fish oil is produced from small marine pelagic fish and represents a finite fishery resource. Over the past decade, global fish oil production has reached a plateau and is not expected to increase beyond current levels. In order to sustain its rapid development, the aquaculture industry cannot continue to rely on finite stocks of marine pelagic fish for fish oil supply. The substitution of fish oil with alternative oil sources is therefore imperative for the successful expansion of the industry.

Palm oil is currently second behind soybean oil in world seed oil production tonnage (O'Mara, 1998). Crude palm oil is the richest natural sources of â-carotene, tocopherols and tocotrienols which function as natural antioxidants. These confer beneficial effects to growth and flesh quality when fish are fed high levels of palm oil in their diets (Lim *et al.*, 2001). Palm oil has been evaluated in the diets for salmonids (Tortensen *et al.*, 2000; Bell *et al.*, 2002), catfishes (Legendre *et al.*, 1995; Lim *et al.*, 2001; Ng *et al.*, 2000; Ochang *et al.*, 2007) and tilapias (Ng *et al.*, 2001; Ochang *et al.*, 2007). Palm oil can meet the energy requirements of fish by providing easily oxidized fatty acids and at the same time, generate flesh fatty acid composition that are beneficial to the consumer.

The aim of the present study was to evaluate the influence of partial and/or total fish oil replacement with palm oil on growth and carcass composition of *H. longifilis*.

Materials and methods

Three hundred *H. longifilis* fingerlings were obtained from the hatchery of National Institute for Freshwater Fisheries Research (NIFFR), New-Bussa, Nigeria and used in this study. Prior to the commencement of the experiment, fish were acclimatized to the new environmental condition on a commercial catfish feed for two weeks in a mini-flow-through system.

Danish fish meal was used as the primary protein source in the test diets because of its prevalent use in

diets fed to cultured catfish. Danish fishmeal contains approximately 10% fish oil as purchased, which might provide sufficient HUFA to meet the essential fatty acid requirement of *H longifilis* and obviate the effects of the lipid of interest. Therefore, the fishmeal was exhaustively extracted to reduce the endogenous lipid to trace amounts before inclusion in the diets. Five *iso*-nitrogenous, *iso*-lipidic, *iso*-caloric experimental diets were formulated with 6% lipid originating from 100% fish oil (FO) and 0% (PO); 75% FO and 25% PO; 50% FO and 50% PO; 25% FO and 75% PO and 0% FO and 100% PO (Table 1). The diets were made into pellet with meat mincer through 2mm die, sundried, packed in polythene bags, sealed and stored at -20°C until used.

Fish were randomly assigned into groups of twenty per tank. Each dietary treatment had three replications and the experiment was conducted for 10 weeks. The fish were individually weighed at the beginning and at the end of the experiment and bulk-weighed by tank fortnightly in-between. Fortnight bulk weights were used to adjust the daily feed ration for the following week. Fish were offered 50 gkg⁻¹ of their body weight per day, sub divided into three equal feeds at 09:00, 15:00 and 21:00 h daily.

Faecal matter was collected once a day at about 8:00 am before feeding commenced during the later part of the experiment. Faeces collection was performed by siphoning materials from the bottom of tank. At the termination of the experiment, five fish were taken from each replication for determination of whole body composition, liver and muscle lipid.

The nutrient composition of the experimental diets, muscle and liver samples was determined by proximate analysis. Moisture was determined by thermal drying to constant weight in an oven at 110°C for 24 h. Crude protein was determined using the Kjeldhal method (Association of Official Analytical Chemists, 1990), Total lipid of diets, liver and muscle tissue was extracted according to Folch et al. (1957) and quantified gravimetrically.

The data were subjected to analysis of variance (ANOVA) and if significant (P<0.05) differences were found, Duncan's' multiple range test (Duncan, 1955) was used to rank the group using SPSS version 10.0 (SPSS, 1997). The data are presented as mean \pm S.E.M. of three replicate groups.

RESULTS

The mean final weights of H. longifilis fingerling fed all levels of palm oil substitution for fish oil were not significantly different. Similar trend were evident for other growth parameters [average feed intake (AVFI), feed conversion ratio (FCR) and specific growth rates (SGR)] (Table 2)

There were no significant differences between AVF1 or FCR for all groups. The lowest FCR however $(1.90 \pm)$ was observed for 25 % PO treatment, while AVF1 ranged from $21.02 \pm (25 \% \text{ PO})$ to $25.55 \pm (0 \% \text{ PO})$. Protein efficiency ratio (PER) ranged from $1.01 \pm (75 \% \text{ PO})$ $1.17 \pm (25 \% \text{ PO})$ and did not differ significantly between the dietary treatments.

Data on the composition of muscle and liver of the experimental fish are reported in Table 3. There were no significant differences observed between dietary treatments for muscle moisture, protein, total lipid and ash content of *H. longifilis* fed the experimental diets. The lipid content of the liver of fish fed 100 % PO was significantly higher than those fed the other diets.

DISCUSSIONS

The result of the present study suggest that PO can be used to completely replace fish oil with minimal adverse effect on *H. longifilis* fingerlings growth, as reported for other fish species (Martino *et al.*, 2002; Regost *et al.*, 2003; Turchini *et al.*, 2003; Francis *et al.*, 2006; Ochang *et al.*, 2007). This is evident by the weight gain and feed conversion of fish fed 25 %, 50 %, 75 %, and 100 % PO diets which ranged from $5.57 \pm to 6.70 \pm and 1.90 \pm to 2.07 \pm respectively, with no significant differences from fish fed the control (FO) diet, and generally comparable to previous results on tropical catfish (Ng$ *et al.*, 2000; Ochang*et al.*, 2007). The no significant difference between SGR and FCR of the experimental diets indicated that fish were able to digest the diets and convert the diets into body tissue with the same degree of efficiency.

Although muscle lipid levels in this study were not significantly different, higher level of lipid was observed in fish fed 0 % PO (100 % FO) compared with fish fed the other diets. Conversely, the muscle protein content was lower in fish fed 0 % PO, the second highest muscle lipid level, found in fish fed 50 % PO, was correlated with the second lowest protein content. This relationship between muscle protein and lipid content has been observed in studies with salmonids (Reinitz and Hitzel, 1980; Jokumsen and Aisted, 1990; Bell *et al.*, 2002). Lipid deposition in the liver was significantly affected by dietary treatment. The fish group fed 100 % PO had the highest liver lipid content. This observation is similar to that of Caballero *et al.*, (2004), that the reduction of dietary essential fatty acids due to the inclusion of vegetable oils in the diets tends to promote fat accumulation in the liver of seabream.

In conclusion, the replacement of cod liver oil by palm oil as lipid supplement in the diet permitted a clear improvement of growth and FCR of *H. longifilis*. This indicates that PO can effectively replace FO in the diet of *H. longifilis*.

REFERENCES

- AOAC (1990). In: Helrich, K. (Ed.), Official Methods of Analysis of the Association of Official Analytical Chemists. Association of Official Analytical Chemists, Arlington, VA, USA.
- Bell, J.G., R.J. Henderson, D.R. Tocher, F. McGhee, J.R. Dick, A. Porter, R.P. Smullen and J.R. Sargent (2002). Substituting fish oil with crude palm oil in the diet of Atlantic salmon (*Salmo salar*) affects muscle fatty acid composition and hepatic fatty acid metabolism. J. Nutr., 132: 222-230.
- Bell, J.G., McGhee, F., Campbell, P.J., Sargent, J.R. (2003). Rapeseed oil as an alternative to marine fish oil in diets of post-smolt Atlantic salmon (*Salmo salar*): changes in flesh fatty acid composition and effectiveness of subsequent fish oil "wash out". Aquaculture 218: 515528.
- Caballero, M.J., Obach, A., Rosenlund, G., Montero, D., Gisvold, M., Izquierdo, M.S. (2002). Impact of different dietary lipid sources on growth, lipid digestibility, tissue fatty acid composition and histology of rainbow trout, Oncorhynchus mykiss. Aquaculture214: 253271.
- Caballero, M.J., Izquierdo, M.S., Kjørsvik, E., Ferna'ndez, E., Rosenlund, G. (2004). Histological alterations in the liver of sea bream, *Sparus aurata* L., caused by short- or long-term feeding with vegetable oils. Recovery of normal morphology after feeding fish oil as the sole lipid source. J. Fish Dis. 27,:31541.
- Duncan, D.B. (1955). Multiple range and multiple (F) test. Biometrics, 11:1-42.
- Folch, J., Lees, M., Stanley-Sloane, G.H. (1957). A simple method for the isolation purification of total lipids from animal tissues. J. Biol. Chem. 226: 497 507
- Francis, D.S.; Turchini, G.M.; Jones, P.L. and De Silva, S.S. (2006). Effects of dietary oil source on growth and fillet fatty acid composition of Murray cod, *Maccullochella peelii peelii*. *Aquaculture*, **253**: 547 556.
- Glencross, B., Hawkins, W., Curnow, J. (2003). Evaluation of canola oils as alternative lipid resources in diets for juvenile red seabream, *Pagrus auratus*. Aquac. Nutr. 9: 305315.
- Jokumsen, A. and Aisted, N. (1990). Ørredvackst l relation til forholdet mellem protein og fedt l foderet. Medd. forsøgsdambruget 80:1-15.
- Legendre, M., N. Kerdchuan, G. Corraze and P. Bergot, (1995). Larval rearing of an African catfish *Heterobranchus longifilis* Teleostei, Clariidae : effect of dietary lipids on growth, survival and fatty acid composition of fry. *Aquat. Living Resour*, 8: 355-363.
- Lim, P.K., P.L. Boey and W.K. Ng, (2001). Dietary palm oil level affects growth performance, protein retention and tissue vitamin E concentration of African catfish, *Clarias gariepinus. Aquaculture*, 202: 101-112.
- O'Mara, C.J. (1998). U.S. oil seed industry looks at trade issues. Inform. 9: 132-136.
- Martino, R.C., Cyrino, J.E.P., Portz, L., Trugo, L.C. (2002). Performance and fatty acid composition of surubim (*Pseudoplatystoma coruscans*) fed diets with animal and plant lipids. *Aquaculture* 209: 233246.
- Ng, W.K., M.C. Tee and P.L. Boey (2000). Evaluation of crude palm oil and refined palmolein as dietary lipids in pelleted feeds for a tropical bagrid catfish *Mytus nemurus* (Cuvier and Valenciennes). *Aquacult. Res.*, **31**: 337-347.
- Ng, W.K, P.K. Lim and H. Sidek (2001). The influence of dietary lipid source on growth, muscle fatty acid composition and erythrocyte osmotic fragility on hybrid tilapia. *Fish Physiol. Biochem.*, 25: 301-310.
- Ng, W.K., P.K. Lim and P.L. Bocy (2003).Dietary lipid and palm oil source affects growth, fatty acid composition and muscle alpha-tocopherol concentration of African catfish, *Clarias gariepinus*. *Aquaculture*, **215**: 229-243.
- Ochang, S.N.; Fagbenro, O.A. and Adebayo O. T. (2007). Growth Performance, Body Composition,
- Haematology and Product Quality of the African Catfish (*Clarias gariepinus*) Fed Diets with Palm Oil *Pakistan Journal of Nutrition.* **6**: 452-459
- Ochang, S.N.; Fagbenro, O.A. and Adebayo O. T. (2007). Influence of dietary palm oil on growth response, carcass composition, haematology and organoleptic properties of juvenile Nile tilapia, *Oreochromis* niloticus. Pakistan Journal of Nutrition. 6: 424-429

Regost, C., J. Arzel, M. Cardinal, G. Rosenlund and S.J. Kaushik (2003). Total replacement of fish oil by

soybean or linseed oil with a return to fish oil in Turbot (*Psetta maxima*) II. Flesh quality properties. *Aquaculture*, **220** : 737-747.

- Reinitz, G., Hitzel, F. (1980). Formulation of practical diets for rainbow trout based on desired performance and body composition. Aquaculture 19: 243 252.SPSS, (1997). SPSS Base 7.5 for Window. SPSS, 444 N. Michigan Avenue, Chicago, IL, USA.
 - Tacon, A.G.J. (2003) Global trends in aquaculture and compound aquafeed production. In: International Aquafeed Directory and Buyers' Guide 2003 (Tacon, A.G.J. ed.), pp. 823. Turret RAI, Uxbridge, Middlesex, UK.
 - Tortensen, B.E., O. Lie and L. Froyland (2000). Lipidmetabolism and tissue composition in Atlantic salmon (Salmo salar L.) effects of capelin oil, palm oil and oleic-enriched sunflower oil as dietary lipid sources. Lipids, 35: 653-664.
 - Turchini, G.M., Mentasti, T., Frøyland, L., Orban, E., Caprino, F., Moretti, V.M., Valfre, F. (2003). Effects of alternative dietary lipid sources on performance, tissue chemical composition, mitochondrial fatty acid oxidation capabilities and sensory characteristics in brown trout (Salmo trutta L.). Aquaculture 225.:251267. M. Kuthalos L. M. Lio Zeo J. C. Grenardo M. D. Englaszo M. A. Roku O. L. ban nohizaquata 🖵 🗤

			Diets	oils Recovery [54]	vegetablet Die 27 - 1
Ingredients	0 % PO	25 % PO	50 % PO	75 % PO	100 % PO
Fish meal (Danish)	398	398	398	398	398
Soybean meal	313	313	ban 313 ann	313	313
Corn flour (Maize)	177	177	177	177	177
Cassava starch	20	20	20	20	20
Methionine	an isbito di	noiaei10 l azke	evber 10 . Wet	10	10
*Vit./Min. Premix	20	Jog 20 M b	18 os 20	20	20
Salt (NaCl)	1.5	1.5	1.5	1.5	1.5
Vitamin C	0.5	0.5	0.5	0.5	0.5
Fish oil	60	45	30	15	0
Palm oil and a star that but	Perf Omance	200215	30	1 9 45 om	60
Nutrient contents (%)	th animal and	ov study byl da	an what o the	ter part part of a second	al-della
Moisture online beniler b	6.23 abi	6.38	6.56	6.31	6.42
Crude protein	45.08	44.98	45.02	45.06	44.99
Crude lipid	12.52	12.50	12.53	12.56	12.45
Ash	8.75	8.82	8.56	8.32	8.40
Crude fibre	1.45	1.42	1.38	1.41	1.40
Nitrogen free extract	25.97	25.90	25.95	26.34	26.34
Gross energy (MJ/kg)**	19.92	19.88	19.91	20.00	19.87

: Following G.A. and Adebayo U.T. (2007), fail acrue of dietary raim of lot crossin easy

253

Vitamin/mineral premix supplied the following (per kg of diet): calcium, 4500mg; phosphorus, 4200mg; potassium, 1700mg; magnesium, 400mg; iron, 30mg; zinc, 30mg; manganese, 20mg; copper, 5mg; iodine, 1mg; selenium, 0.25mg; vitamin A, 5000IU; vitamin D, 2000IU; DL--tocopherol acetate, 100mg; menadione, 15mg; thiamine hydrochloride, 5mg; riboflavin, 10mg; pyridoxine hydrochloride, 10mg. Panthothenic acid, 35mg; nicotinic acid, 50mg; biotin, 0.5mg; folic cid, 2mg; ascorbic acid, 200mg; inositol, 250mg; choline, 400mg; vitamin B₁₂, 0.1mg and ethoxyquin, 60mg.

**Based on 23.4 MJ/kg protein, 39.2 MJ/kg lipid and 17.2 MJ/kg NFE

 Table 2. Growth performance of *H. longifilis* fed the experimental diets containing increasing levels of palm oil (PO) for 10 weeks.

Parameters	0 % PO	25 % PO	50 % PO	75 % PO	100 % PO
Initial weight (g)	5.71 ± 0.17	5.48 ± 0.06	5.44 ± 0.17	5.50± 0.10	5.53 ± 0.23
Final weight (g)	1213134 1.21	1.005.050.24	0.1214-1050 0.97	01973232 0.28	0.1131.359± 0.140.
Weight gain (g)	6.70 ± 0.19	5.57 ± 0.08	6.06 ± 0.12	5.80 ± 0.10	5.86 ± 0.11
SGR (% day ⁻¹)	2.07 ± 0.13	1.97 ± 0.02	2.02 ± 0.14	1.89 ± 0.05	2.00 ± 0.07
erage feed intake (g)	25.55 ± 2.77	21.02 ± 0.31	23.27 ± 0.96	23.01 ± 2.08	22.52 ± 0.79
Feed conversion ratio	2.07 ± 0.08 B	1.90±0.03	2.04 ± 0.05	2.03 ± 0.06	1.98 ±0.01 ↔
Protein efficiency ratio	1.0刀≇ 0.03	0.0371≠10.01 0	. 0 11101⊕0.05 C	.05010± 0.07	○1.121.4.20.05

Table 3. Proximate composition of muscle and liver lipid concentration of *Heterobranchus longifilis* fed diets containing increasing levels of palm oil (PO) for 10 weeks.

Parameters	olem do % PO	25 % PO	50 % PO	75 % PO	100 % PO
Parameters	0 % PO	25 % PO	50 % PO	75 % PO	100 % PO
Moisture	72.37 ± 0.18	72.4 ± 0.12	72.37 ± 0.09	72.30 ± 0.17	72.43 ± 0.18
Lipid) muinero sciuni	4.43 ± 0.15	$^{\circ}$ 4.26 \pm 0.02 $^{\circ}$	$4.37\pm0.09^{\circ}$	4.27 ± 0.09	4.20 ± 0.06
Protein d goulde ou dei	17.04 ± 0.22	17.18 ± 0.15	17.12 ± 0.28	19.16 ± 0.21	17.24 ± 0.15
Ash could have to a	5.67 ± 0.09	5.67 ± 0.09	5.70 ± 0.10	5.77 ± 0.09	5.70 = 0.06
Liver lipid	3.31 ± 0.01^{a}	3.34 ± 0.01^{a}	$3.50\pm0.01^{\circ}$	$3.61\pm0.01^\circ$	3.84 ± 0.03^{d}

his would enable us come up with maximum time required of fish out of water to remain qualifatively

Data are mean \pm S.E.M. (n=3): means with different superscripts are significantly different (P<0.05).

Material and Methods

335

254