



UNIVERSITI PUTRA MALAYSIA

***PREPARATION AND CHARACTERIZATION OF CORE-SHELL
COPPER NANOPARTICLES STABILIZED BY KAPPA CARRAGENAN
AND SODIUM ALGINATE***

HAJAR KHANEHZAEI

FS 2015 11



**PREPARATION AND CHARACTERIZATION OF CORE-SHELL
COPPER NANOPARTICLES STABILIZED BY KAPPA CARRAGENAN
AND SODIUM ALGINATE**

By

HAJAR KHANEHZAIE

**Thesis Submitted to the School of Graduate Studies Universiti Putra
Malaysia, in Fulfillment of the Requirement for the degree of Master of
Science**

May 2015

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of University Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purpose from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of University Putra Malaysia.

Copyright© Universiti Putra Malaysia



DEDICATION

To God, who gave me life and strength

To my family



© COPYRIGHT UPM

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

**PREPARATION AND CHARACTERIZATION OF CORE-SHELL
COPPER NANOPARTICLES STABILIZED BY KAPPA CARRAGENAN
AND SODIUM ALGINATE**

By

HAJAR KHANEHZA EI

May 2015

Chairman: Professor Mansor B. Ahmad, PhD
Faculty: Science

Among metal nanoparticles, copper nanoparticles (Cu-NPs) have recently attracted increased attention because of their low cost (in contrast to Au and Ag) and their usable properties. Rate of resistance of microorganisms to antibiotics is alarming. Pathogenic bacteria are becoming much resistant to antibiotics. Copper nanoparticles synthesis particularly in bio-based stabilizers can act as antimicrobial agents, which in turn could resist pathogenic microorganisms. In this work, synthesis and characterization of Cu-NPs and Cu@Cu₂O core shell nanoparticles (CSNPs) were carried out in the presence of two different marine polymers as stabilizers *Kappaphycus alvarezii* (*K. alvarezii*) and sodium alginate (SA) by two varied methods; chemical reduction and gamma irradiation. The aim of this research is to synthesize, characterize and test the antimicrobial activity of Cu and Cu₂O nanoparticles. The Cu@Cu₂O-CSNPs were synthesized by using chemical reduction method in *K. alvarezii* media. Also Cu-NPs and Cu@Cu₂O-CSNPs were prepared in sodium alginate by using same method. The synthesis carried out using aqueous medium in presence of hydrazinium hydroxide as reducing agent, CuSO₄.5H₂O as copper precursor and sodium hydroxide as pH moderator.

The effects of the concentration of *K. alvarezii* and sodium alginate as stabilizer were studied. Formation of nanoparticles was determined by UV-vis spectroscopy where surface plasmon absorption maxima can be observed at 390-600 nm from the UV-vis spectrum. The synthesized nanostructures were also characterized by X-ray diffraction (XRD). In addition; the morphology and structure of the nanoparticles were investigated by Transmission Electron Microscopy (TEM), emission scanning electron microscopy (FESEM) and Energy-dispersive X-ray spectroscopy (EDX). TEM results showed a gradual decrease of particle size from low concentration of *K. alvarezii* and sodium alginate to high concentration. The study clearly

showed that using various amounts of *K. alvarezii* and sodium alginate led to produce Cu@Cu₂O-CSNPs with different sizes and ratios of Cu:Cu₂O. For the *K. alvarezii*/Cu@Cu₂O-CSNPs, the Cu-NPs increased and the Cu₂O decreased with increasing the concentration of *K. alvarezii* until 0.2 wt.% in the Cu@Cu₂O-CSNPs. At the highest concentration of *K. alvarezii*, Cu decreased compare to Cu₂O. Moreover for the Cu@Cu₂O-CSNPs synthesized in sodium alginate with increasing the media concentration, the Cu increased and at the highest concentration of media (1.0 wt.%) only copper nanoparticles were produced. The size of the nanoparticles decreased as the amount of *K. alvarezii* and sodium alginate was increased.

Moreover the *K. alvarezii*/Cu@Cu₂O-CSNPs and SA/Cu-NPs were synthesized by using gamma irradiation method. The synthesis was done through γ -irradiation reduction of copper ions. The suspension was irradiated under γ -irradiation source ⁶⁰Co with doses of 5, 20, 40, 80, 100 and 120 kGy at room temperature. The effect of irradiation dosage was investigated. The UV-Vis spectroscopy results obtained for samples indicated that only the samples which were irradiated by 80, 100 and 120 kGy showed surface plasmon resonance (SPR) peaks in the range of 350-600 nm. It was due to the formation of *K. alvarezii*/Cu@Cu₂O-CSNPs and SA/Cu-NPs for these doses. TEM analysis indicated size and distribution of the nanoparticles varied in different doses of gamma irradiation. When the absorbed dose increased from 80 to 120 kGy the nanoparticle size decreased. Following synthesis of the nanoparticles, antibacterial analysis of the synthesized particles was investigated. Antimicrobial analysis conducted on the synthesized nanoparticles showed activity against all tested microorganisms.

Abstrak tesis yang dikemukakan kepada senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Master Sains

**PENYEDIAAN DAN PENCIRIAN NANOPARTIKEL KUPRUM TERAS
CANGKERANG YANG DISTABILKAN OLEH KAPPA CARRAGENAN
DAN NATRIUM ALGINAT**

Oleh

HAJAR KHANEHZAIE

Mei 2015

Pengerusi: Profesor Mansor B. Ahmad, PhD
Fakulti: Sains

Antara nanopartikel logam, nanopartikel tembaga (Cu-NPs) baru-baru ini telah menarik banyak perhatian kerana kos yang rendah (berbanding Au dan Ag) dan sifat-sifat bermanfaat mereka. Kadar rintangan mikroorganisma terhadap antibiotik adalah membimbangkan. Bakteria patogenik menjadi lebih tahan terhadap antibiotik. Nanopartikel tembaga sintesis terutamanya dalam penstabil berasaskan bio boleh bertindak sebagai agen anti-mikrob, yang seterusnya dapat menahan mikroorganisma patogenik.

Dalam kerja ini, sintesis dan pencirian nanopartikel Cu dan nanopartikel teras petala (CSNPs) Cu@Cu₂O telah dijalankan dengan menggunakan dua polimer marin yang berbeza sebagai penstabil, *Kappaphycus alvarezii* (*K. alvarezii*) dan natrium alginat (SA) oleh dua kaedah iaitu penurunan kimia dan radiasi gamma. Tujuan kajian ini adalah untuk mensintesis, mencirikan nanopartikel Cu dan Cu₂O dan menguji aktiviti antimikrobia nanopartikel. CSNPs Cu@Cu₂O telah disintesis dengan menggunakan kaedah penurunan kimia dalam media *K. alvarezii*. Cu-NPs dan nanopartikel teras petala Cu@Cu₂O juga telah disediakan dalam natrium alginat dengan menggunakan kaedah yang sama. Sintesis dijalankan dalam medium akueus dengan menggunakan hidrazinium hidroksida sebagai agen penurunan, CuSO₄.5H₂O sebagai prekursor tembaga dan natrium hidroksida sebagai pengawal pH.

Kesan kepekatan *K. alvarezii* dan natrium alginat sebagai penstabil telah dikaji. Pembentukan nanopartikel telah ditentukan oleh spektroskopi UV-nampak di mana penyerapan plasmon permukaan maksimum dapat dilihat pada 390-600 nm dari spektrum UV-nampak. Nanostruktur yang disintesis juga dicirikan dengan pembelauan sinar-X (XRD). Sebagai tambahan,

morfologi dan struktur nanopartikel telah disiasat dengan Mikroskopi transmisi electron (TEM), mikroskopi pengimbasan electron FESEM dan Tenaga-serakan sinar-x (EDX). Keputusan TEM menunjukkan saiz zarah berkurangan secara sedikit demi sedikit daripada kepekatan *K. alvarezii* dan natrium alginat rendah kepada kepekatan tinggi. Kajian ini jelas menunjukkan bahawa dengan memvariasikan jumlah *K. alvarezii* dan natrium alginat membawa kepada penghasilan CSNPs Cu@Cu₂O dengan saiz dan nisbah Cu:Cu₂O yang berbeza. Untuk CSNPs *K. alvarezii*/Cu@Cu₂O, Cu-NPs bertambah dan Cu₂O berkurangan dengan peningkatan kepekatan *K. alvarezii* sehingga berat 0.2% dalam nanopartikel Cu@Cu₂O. Pada kepekatan tertinggi *K. alvarezii*, Cu menurun berbanding Cu₂O. Tambahan pula nanopartikel Cu@Cu₂O yang disintesis dalam natrium alginat dengan peningkatan kepekatan media, Cu meningkat dan pada kepekatan tertinggi media (berat 1.0%) hanya nanopartikel tembaga dihasilkan. Saiz nanopartikel berkurangan sebagaimana jumlah *K. alvarezii* dan natrium alginat meningkat.

Selain itu, *K. alvarezii*/Cu@Cu₂O-CSNPs dan SA/Cu-NPs telah disintesis dengan menggunakan kaedah radiasi gamma. Sintesis itu dilakukan melalui penurunan ion tembaga oleh sinar- γ . Koloid itu telah diradiasi di bawah sinar- γ bersumberkan ⁶⁰Co dengan penyerapan dos daripada 5, 20, 40, 80, 100 dan 120 kGy pada suhu bilik. Keputusan spektroskopi UV-nampak diperolehi menunjukkan bahawa hanya sampel yang telah diradiasi sebanyak 80, 100 dan 120 kGy menunjukkan puncak plasmon permukaan resonans (SPR) dalam julat 350-600 nm. Ia adalah disebabkan oleh pembentukan *K. alvarezii*/Cu@Cu₂O-CSNPs dan SA/Cu-NPs bagi dos-dos tersebut. Analisis TEM menunjukkan saiz dan taburan nanopartikel berubah dalam dos radiasi gamma berbeza. Apabila dos yang diserap meningkat dari 80 kepada 120 kGy, saiz nanopartikel berkurangan.

Selepas mensintesis nanopartikel, analisis antibakteria zarah yang disintesis telah dikaji. Analisis antimikrobia dijalankan ke atas nanopartikel yang disintesis dan ianya menunjukkan aktiviti terhadap semua mikroorganisma yang diuji.

ACKNOWLEDGEMENTS

First of all, I will like to give praise to Almighty God for the wisdom and determination that he has bestowed upon me during this research project, and indeed, for keeping me healthy and safe throughout my life.

I would like to express my sincere gratitude to my supervisor, Prof. Dr. Mansor B. Ahmad for his guidance and support throughout this study, especially for his countless hours of reflecting, reading, encouraging, and most of all patience throughout the entire process. I would also like to thank my committee members: Prof. Mansor B. Ahmad and Norhazlin Zainuddin and were more than generous with their expertise and precious time.

I would like to acknowledge my school division for allowing me to conduct my research and providing all the necessary assistance requested. Special thanks are also given to all the staffs of development and human resources department for their support. I would like to express my deepest gratitude to Dr Kamyar Shameli for sharing with me their knowledge and giving their precious time to help me achieve my aim.

Appreciation is also given to my good friends for their help encouragement which keep me going and wish them all the best in their life.

Most of all, I would like to express my sweetest appreciation to my family for their affectionate support, patience and encouragement all time. Their prayers and good wishes always help me to be strong, especially in difficult times. I am very grateful and thankful to them.

I certify that a Thesis Examination Committee has met on 25 May 2015 to conduct the final examination of Hajar Khaneh Zaei on her thesis entitled "Preparation and Characterization of Core-Shell Copper Nanoparticles Stabilized by Kappa Carragenan and Sodium Alginate" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Mahiran binti Basri, PhD

Professor

Faculty of Science

Universiti Putra Malaysia

(Chairman)

Mohamad Zaki bin Abd Rahman, PhD

Associate Professor

Faculty of Science

Universiti Putra Malaysia

(Internal Examiner)

Sugeng Triwahyono, PhD

Associate Professor

Universiti Teknologi Malaysia

Malaysia

(External Examiner)



ZULKARNAIN ZAINAL, PhD

Professor and Deputy Dean

School of Graduate Studies

Universiti Putra Malaysia

Date: 17 June 2015

This thesis submitted to the Senate of Universiti Putra Malaysia has been accepted as fulfillment for the degree of Doctor of Philosophy.

The members of the Supervisory committee were as follows:

Mansor B. Ahmad, PhD

Professor

Faculty of Science

Universiti Putra Malaysia

(Chairman)

Norhazlin Zainuddin, PhD

Senior Lecturer

Faculty of Science

Universiti Putra Malaysia

(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- This thesis is my original work;
- Quotations, illustrations and citations have been duly referenced;
- This thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- Intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- Written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- There is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____

Date: _____

Name and Matric No: Hajar Khanehzaei (GS35367)

Declaration by Members of Supervisory Committee

This is to confirm that:

- The research conducted and the writing of this thesis was under our Supervision;
- Supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:
Name of
Chairman of
Supervisory
Committee:

Signature:
Name of
Member of
Supervisory
Committee:



TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAKT	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF FIGURES	xiii
LIST OF TABLES	xvi
LIST OF ABBREVIATIONS	xvii
CHAPTER	
1 INTRODUCTION	
1.1 Background	1
1.2 Problem Statement	2
1.3 Objectives	2
2 LITERATURE REVIEW	4
2.1 Nanomaterials	4
2.2 Nanoparticle synthesis	5
2.2.1 Physical methods	5
2.2.2 Chemical methods	6
2.2.3 Unstabilized chemical synthesis of metal nanoparticles	7
2.2.4 Stabilized chemical synthesis of metal nanoparticles	7
2.2.5 Green synthesis	9
2.3 Seaweeds	11
2.4 Stabilizers	13
2.5 Antibacterial activity of nanoparticles	14
2.5.1 Gram positive bacteria	15
2.5.2 Gram negative bacteria	15
3 MATERIALS AND METHODS	17
3.1 Materials	17
3.2 Methods	17
3.2.1 Preparation aqueous medium of <i>K. alvarezii</i>	17
3.2.2 Preparation of Cu@Cu ₂ O core shell nanoparticles by chemical reduction method in <i>K. alvarezii</i>	17
3.2.3 Preparation of Cu@Cu ₂ O-CSNPs and Cu-	18

	NPs by chemical reduction method in sodium alginate	
3.2.4	Preparation of Cu@Cu ₂ O-CSNPs by gamma irradiation method in <i>K. alvarezii</i>	18
3.2.5	Preparation of Cu-NPs by gamma irradiation method in Sodium alginate	18
3.3	Characterization	19
3.3.1	Ultra Violet-Visible Spectroscopy	19
3.3.2	X-ray Diffraction	19
3.3.3	Field Emission Scanning Electron Microscopy	19
3.3.4	Transmission Electron Microscopy	19
3.3.5	Energy-Dispersive X-ray spectroscopy	20
3.3.6	Fourier Transformed-Infrared Spectroscopy	20
3.4	Antibacterial test	20
4	RESULTS AND DISCUSSION	21
4.1	Preparation of Cu@Cu ₂ O core shell nanoparticles by chemical reduction method in <i>K. alvarezii</i>	21
4.2	Characterization of <i>K. alvarezii</i> / Cu@Cu ₂ O-CSNPs	23
4.2.1	UV-visible spectroscopy	23
4.2.2	X-ray diffraction	25
4.2.3	Transmission electron microscopy	26
4.2.4	Scanning electron microscopy	28
4.2.5	Energy dispersive X-ray spectroscopy	29
4.2.6	Fourier Transformed-Infrared Spectroscopy	30
4.3	Antibacterial activity of synthesized <i>K. alvarezii</i> /Cu@Cu ₂ O- CSNPs by chemical reduction	32
4.4	Preparation of Cu and Cu@Cu ₂ O core shell nanoparticles by chemical reduction method in sodium alginate	34
4.5	Characterization of SA /Cu@Cu ₂ O-CSNPs	36
4.5.1	UV-visible spectroscopy	36
4.5.2	X-ray diffraction	37
4.5.3	Transmission electron microscopy	38
4.5.4	Scanning electron microscopy	41
4.5.5	Energy-dispersive X-ray spectroscopy	42
4.5.6	Fourier Transformed-Infrared Spectroscopy	43
4.6	Antibacterial activity of Cu@Cu ₂ O-CSNPs and Cu-NPs synthesis by chemical reduction method in SA	44
4.7	Preparation of Cu@Cu ₂ O-CSNPs by gamma irradiation method in <i>K. alvarezii</i>	46

4.8	Characterization of <i>K. alvarezii</i> /Cu@Cu ₂ O- CSNPs prepared by gamma irradiation	47
4.8.1	UV-visible spectroscopy	47
4.8.2	X-ray diffraction	49
4.8.3	Transmission electron microscopy	50
4.8.4	Scanning electron microscopy	52
4.8.5	Energy-dispersive X-ray spectroscopy	54
4.8.6	Fourier Transformed-Infrared Spectroscopy	54
4.9	Antibacterial activity of synthesized <i>K.</i> <i>alvarezii</i> /Cu@Cu ₂ O- CSNPs by gamma irradiation	56
4.10	Preparation of Cu-NPs by gamma irradiation method in sodium alginate	57
4.11	Characterization of SA/Cu-NPs prepared by gamma Irradiation	59
4.11.1	UV-Visible Spectroscopy	59
4.11.2	X-ray diffraction	60
4.11.3	Transmission electron microscopy	61
4.11.4	Scanning electron microscopy	63
4.11.5	Energy-dispersive X-ray spectroscopy	64
4.11.6	Fourier Transformed-Infrared Spectroscopy	65
4.12	Antibacterial activity of synthesized SA/Cu-NPs by gamma Irradiation	66
5	CONCLUSIONS AND RECOMMENDATION	68
5.1	Conclusions	68
5.2	Recommendation for future research	69
	REFERENCES	70
	BIODATA OF STUDENT	80
	LIST OF PUBLICATIONS	81

--	--



© COP YRIGHT UPM

LIST OF FIGURES

Figure	Page
2.1. Kappaphycusalvarezii(source https://seaweedindustry.com/seaweed/type/kappaphycus-alvarezii)	11
2.2. Chemical structure of kappa carrageenan(Urano and Ina, 2007)	12
2.3. Chemical structure of sodium alginate) (Kashima and Imai, (2012)	13
4.1. Schematic illustration of the synthesized Cu@Cu ₂ O-CSNPs in <i>K. alvarezii</i> by chemical reduction method	22
4.2. Photograph of prepared <i>K. alvarezii</i> /Cu@Cu ₂ O-NPs at different Concentration of medium, respectively [0.05, 0.1, 0.2, 0.5 and 1.0(a-e) wt.%].	23
4.3. UV-vis absorption of <i>K. alvarezii</i> (a) and <i>K. alvarezii</i> /Cu@Cu ₂ ONPs at different concentration of medium, respectively [0.05(b), 0.1(c), 0.2(d), 0.5(e) and 1.0(f) wt.%].	24
4.4. XRD diffractogram of <i>K. alvarezii</i> (a) and <i>K. alvarezii</i> /Cu@Cu ₂ O-NPs at different concentration of medium, respectively [0.05(b), 0.1(c), 0.2(d), 0.5(e) and 1.0 (f) wt.%].	25
4.5. Transmission electron micrographs and particle size distribution of <i>K. alvarezii</i> /Cu@Cu ₂ O-NPs at different concentration of <i>K. alvarezii</i> medium, 0.05, 0.1, 0.2 and 1.0 wt % (a-d), respectively	27
4.6. TEM image of multi-single particles Cu@Cu ₂ O core shell at 0.2 wt.% of <i>K. alvarezii</i> medium	28
4.7. Field emission scanning electron micrographs of <i>K. alvarezii</i> /Cu@Cu ₂ O- NPs at different Concentration of <i>K. alvarezii</i> medium, respectively [0.05, 0.1, 0.2 and 1.0 wt.% (a-d)].	29
4.8. Energy-dispersive X-ray spectroscopy of <i>K. alvarezii</i> /Cu@Cu ₂ O-NPs at different concentration of <i>K. alvarezii</i> medium respectively [0.05, 0.1, 0.2 and 1.0 wt.% (a-d)].	30
4.9. Fourier transformed infrared spectra for <i>K. alvarezii</i> and <i>K. alvarezii</i> /Cu@Cu ₂ O-NPs at different concentration of <i>K. Alvarezii</i> medium, respectively [0.05, 0.2 and 1.0 wt.% (b-d)]	31
4.10. Images of Cu@Cu ₂ O-CSNPs in <i>K. alvarezii</i> media in (a) Methicillin Resistant Staphylococcus aureus (MRSA) (b) Staphylococcus aureus (S276) (c) Pseudomonas aeruginosa (15442) (d) Escherichia coli (E266) [S (<i>k. alvarezii</i>) S1, S2, S3, S4 and S4 (0.05, 0.1, 0.2, 0.5 and 1.0 wt.%	33
4.11. Schematic illustration of the synthesized Cu@Cu ₂ O-CSNPs in SA by chemical reduction method.	35
4.12. Photograph of prepared SA (a), Cu@Cu ₂ O-CSNPs (b-e), and Cu-NPs (f), at different percentage of SA medium,	36

respectively [0.05, 0.1, 0.2, 0.5 and 1.0 wt.%]	
4.13. UV-vis absorption of SA (a) SA/Cu@Cu ₂ O-NPs (b-e) and SA/Cu-NPs (f), at different percentage of SA medium, respectively [0.05, 0.1, 0.2, 0.5 and 1.0 wt.%]	37
4.14. XRD diffractogram of SA (a) Cu@Cu ₂ O-CSNPs (b-e) and Cu-NPs (f), at different percentage of SA medium, respectively [0.05, 0.1, 0.2, 0.5 and 1.0 wt.%].	38
4.15. Transmission electron micrographs and particle size distribution of Cu@Cu ₂ O-CSNPs (a-c) and Cu-NPs (d) at different percentage of SA medium, respectively [0.05, 0.1, 0.2 and 1.0 wt%].	40
4.16. TEM image for core shell structure of SA/Cu@Cu ₂ O at 0.2 wt.% of sodium alginate	41
4.17. Field emission scanning electron micrographs of Cu@Cu ₂ O-CSNPs (a-c) and Cu-NPs (d) at different percentage of SA medium, respectively [0.05, 0.1, 0.2 and 1.0 wt.%]	42
4.18. Energy-dispersive X-ray spectroscopy of Cu@Cu ₂ O-CSNPs (a-c) and Cu-NPs (d) at different percentage of SA medium [0.05, 0.1, 0.2 and 1.0 wt.%], respectively	42
4.19. Fourier transformed infrared spectra for SA (a), Cu@Cu ₂ O-CSNPs (b,c) and Cu-NPs (d) at different percentage of SA medium [0.05, 0.2 and 1.0 wt.%], respectively	43
4.20. Images of Cu@Cu ₂ O-CSNPs in sodium alginate media in (a) Methicillin Resistant Staphylococcus aureus (MRSA) (b) Staphylococcus aureus (S276) (c) Pseudomonas aeruginosa (15442) (d) Escherichia coli (E266) [SA(sodium alginate) SA1, SA2, SA3, SA4 and SA5 (0.05, 0.1, 0.2, 0.5 and 1.0 wt %)]	45
4.21. Photograph of prepared <i>K. alvarezii</i> /Cu@Cu ₂ O-CSNPs by different doses of gamma radiation, respectively [80,100 and 120 kGy]	46
4.22. UV-vis absorption of <i>K. alvarezii</i> /Cu@Cu ₂ O-CSNPs at different doses of gamma radiation, respectively [5(a), 40(b), 80(c), 100 (d) and 120(e) kGy]	48
4.23. XRD diffractogram of <i>K. alvarezii</i> (a) and <i>K. alvarezii</i> /Cu@Cu ₂ O-CSNPs at different doses of gamma radiation, respectively [80, 100 and 120 kGy (b-d)]	49
4.24. Transmission electron micrographs and particle size distribution of <i>K. alvarezii</i> /Cu@Cu ₂ O-CSNPs at different doses of gamma radiation 80, 100 and 120 kGy (a-c), respectively	51
4.25. TEM image of core shell structure of <i>K. alvarezii</i> /Cu@Cu ₂ O at 100 kGy of gamma radiation	52
4.26. Field emission scanning electron micrographs of Cu@Cu ₂ O-CSNPs (a- c) at different doses of gamma radiation, respectively [80, 90 and 120 kGy]	53
4.27. Energy-dispersive X-ray spectroscopy of <i>K</i>	54

alvarezii/alvarezii/Cu@Cu ₂ O-NPs at different doses of gamma radiation, respectively [80, 100 and 120 (a-c)]	
4.28. Fourier transformed infrared spectra <i>K. alvarezii</i> (a) and <i>K. alvarezii</i> /Cu@Cu ₂ O-CSNPs (b-d) at different doses of gamma radiation, respectively [5, 80 and 120 kGy]	55
4.29. Images of <i>K. alvarezii</i> /Cu@Cu ₂ O-CSNPs in (a) Methicillin Resistant <i>Staphylococcus aureus</i> (MRSA) (b) <i>Staphylococcus aureus</i> (S276) (c) <i>Pseudomonas aeruginosa</i> (15442) (d) <i>Escherichia coli</i> (E266) [1 (<i>K. alvarezii</i>) 2, 3,4,5 (5,80, 100 and 120 kGy)]	56
4.30. Schematic illustration of the synthesized Cu-NPs in SA by gamma radiation	58
4.31. Photograph of prepared SA/Cu-NPs by different doses of gamma radiation, respectively [80,100 and 120 kGy]	59
4.32. UV-vis absorption of Cu-NPs at different doses of gamma radiation, respectively [5(a), 40(b),80(c), 100(d) and 120(e) kGy]	60
4.33. XRD diffractogram of sodium alginate (a) Cu-NPs at different doses of gamma radiation, respectively [80, 100 and 120 kGy (b-d)].	61
4.34. Transmission electron micrographs and particle size distribution of Cu-NPs at different doses of gamma radiation 80, 100 and 120 kGy (a- c), respectively	62
4.35. Field emission scanning electron micrographs of Cu-NPs (a- c) at different doses of gamma radiation, respectively [80, 90 and 120 kGy]	63
4.36. Energy-dispersive X-ray spectroscopy of Cu-NPs at different doses of gamma radiation, respectively [80, 100 and 120 (a- c)]	64
4.37. Fourier transformed infrared spectra SA (a) and Cu-NPs (b- d) at different doses of gamma radiation, respectively [5, 80 and 120 kGy]	66
4.38. Images Cu-NPs in (a) Methicillin Resistant <i>Staphylococcus aureus</i> (MRSA) (b) <i>Staphylococcus aureus</i> (S276) (c) <i>Pseudomonas aeruginosa</i> (15442) (d) <i>Escherichia coli</i> (E266) [1 (SA) 2, 3,4,5 (5,80, 100 and 120 kGy)]	67

LIST OF TABLES

Table	Page
4.1. Antibacterial activities of <i>K. alvarezii</i> /Cu@Cu ₂ O-CSNPs against different bacteria	34
4.2. Antibacterial activities of SA /Cu@Cu ₂ O-CSNPs against different bacteria	46
4.3. Antibacterial activities of synthesized <i>K. alvarezii</i> /Cu@Cu ₂ O-CSNPs by gamma radiation against different bacteria	57
4.4. Antibacterial activities of synthesized SA /Cu by gamma radiation against different Bacteria	67

LIST OF ABBREVIATIONS

CSNPs	Core shell nanoparticles
<i>K. alvarezii</i>	<i>Kappaphycus alvarezii</i>
SA	Sodium alginate
NPs	Nanoparticles
SPR	Surface Plasmon Resonance
XRD	X-ray Diffraction
TEM	Transmission Electron Microscopy
FESEM	Field Emission Scanning Electron Microscopy
SEM	Scanning Electron Microscopy
EDX	Energy Dispersive X-ray
FT-IR	Fourier Transfer Infrared Spectroscopy
DIZ	Diameters of inhibition zone
MRSA	<i>Methicillin Resistant Staphylococcus aureus</i>

CHAPTER 1

INTRODUCTION

1.1 Background

Nanotechnology is one area which offers insight into materials at a nano levels. Nanotechnology deals with technology at atomic, molecular as well as macromolecular level which is in nano-scale. It also gives the basic knowledge of nanoparticles and their effect in association with their application and size (Roco, 2001). Nanoparticle sizes are specifically on interest in nanotechnology studies, as the sizes affect most of their special properties. The special properties of nanoparticles have varying results in various nanotechnology applications. There are different nanoparticle types for different applications such as metallic, non-metallic, carbon nano tubes as well as oxide nanoparticles (Yun *et al.*, 2008). Metallic nanoparticles have special properties which are because of a combination of big proportion of high energy surface atoms as compared with bulk solid as well as to the nanometer scale mean free path of electrons in any metal ($\sim 10\text{-}100$ nm for most metals at room temperature). These are useful in different applications including catalysis, optoelectronics, magnetic, biology as well as micro-electronics (Lijie Zhang and Webster, 2009).

At present, among metallic nanoparticles, synthesis of copper (both oxide and metallic) nanoparticles has gained the attention of scholars because of their usable properties and inexpensiveness.

Copper being one of the inexpensive materials, has gained the attention of researchers via its nanoparticle synthesis due to the similarity in terms of properties with silver and gold, which are expensive (Chattopadhyay and Patel, 2012). Copper is in fact regarded as an alternative for gold, silver as well as platinum in the areas of thermal conducting as well as micro-electronics. Also it has been reported in studies of antimicrobial susceptibility studies, that copper nanoparticles were shown to have increased antibacterial activity than its counterparts in silver (Valodkar *et al.*, 2012). This aids copper nanoparticles synthesis one major area of research in nanotechnology. Copper nanoparticles synthesis of various compositions with preferred sizes as well as shapes is crucial for exploring their applications in the field of nanotechnology (Wen *et al.*, 2011). Areas such as catalysis (Hoover *et al.*, 2006), micro-electronics and sensors (Yun *et al.*, 2008) greatly depend on the size and shapes of nanoparticles.

1.2 Problem statement

In the current environmental pollution being a challenging situation, using environment friendly materials is adamant in research work. Natural sources are being used to get biopolymers which are bio-degradable. They are non-toxic, easily available and also renewable which make them environmentally-friendly. Rate of resistance of microorganisms to antibiotics is surprising. Pathogenic bacteria are becoming much resistant to antibiotics, which are produced on a continuous basis for combating microorganism. At present, antibiotics are resisted by every single pathogenic organism, which makes the fight much challenging (Raffi *et al.*, 2010).

Nanotechnology is a crucial area for research in future with immense potential in terms of nano-medicine. Nanoparticles and their properties such as antimicrobial and smaller size make them able for application in various areas such as medicine and biology. They show an extensive range of properties such as biological properties which varies from their bulk materials. As such synthesis of copper nanoparticles varies from their bulk materials. As such, copper nanoparticles synthesis particularly in bio-based stabilizers can act as antimicrobial agents, which in turn could resist pathogenic microorganisms. For synthesis of copper nanoparticles the stabilization of nanoparticles is one of the major problems to applications and the production of copper nanoparticles is much more challenging than noble metals because copper nanoparticles are totally sensitive to aqueous solutions, and air is stable at these conditions. To avoid oxidation of copper nanoparticles during synthesis an inert environment, such as argon or nitrogen is used.

Stabilization of nanoparticles is difficult due to the high surface area to volume ratio and surface energy of nanoparticles. To obtain copper nanoparticles with high stability, Surfactants and polymers widely used as stabilizers. Many of these commonly used stabilization agents, such as Polyethylene glycol (PEG) and poly vinyl pyrrolidone (PVP) are organic chemicals which are probably toxic.

1.3 Objectives

Synthesis of copper (both oxide and metallic) nanoparticles is beneficial because of its applicability in various industries. Its similarity in terms of properties with noble and expensive metals such as platinum, silver and gold makes it a good alternative to these metals, mainly because of the ready availability as well as its inexpensive nature. Even though there are different Cu and Cu@Cu₂O nanoparticles synthesis approach, this study intends to explore the synthesis and characterization of Cu and Cu@Cu₂O core shell nanoparticles in the sodium alginate and *Kappaphycus alvarezii* (*K. alvarezii*) as

stabilizer via using two varying physical (γ -irradiation) as well as chemical approaches.

The specific objectives are:

1. To prepared (*K. alvarezii*) and sodium alginate stabilized Cu-NPs and Cu@ Cu₂O-CNSPs by chemical reduction and γ -irradiation methods.
2. To characterize Cu-NPs and Cu@Cu₂O-CSNPs using some of the different spectroscopy techniques.
3. To evaluate the antibacterial capability of the produced nanoparticles.



REFERENCES

- Aftab, T., Khan, M., Naeem, M., Idrees, M., Siddiqi, T., Varshney, L. (2014). Effect of irradiated sodium alginate and phosphorus on biomass and artemisinin production in *Artemisia annua*. *Carbohydrate Polymers*, 110396-404.
- Ahmed, A., Gajbhiye, N. S., Joshi, A. G. (2011). Shape controlled synthesis and characterization of Cu₂O nanostructures assisted by composite surfactants system. *Materials Chemistry and Physics*, 129(3): 740-745.
- Akhavan, A., Sheikh, N., Khoylou, F., Naimian, F., Ataeivarjovi, E. (2014). Synthesis of antimicrobial silver/hydroxyapatite nanocomposite by gamma irradiation. *Radiation Physics and Chemistry*, 9846-50.
- Armelao, L., Barreca, D., Bottaro, G., Gasparotto, A., Maccato, C., Maragno, C., Tondello, E., Štangar, U. L., Bergant, M., Mahne, D. (2007). Photocatalytic and antibacterial activity of TiO₂ and Au/TiO₂ nanosystems. *Nanotechnology*, 18(37): 375709.
- Aye, H. L., Choopun, S., Chairuangsi, T. (2010). Preparation of nanoparticles by laser ablation on copper target in distilled water. *Advanced Materials Research*, 9383-86.
- Balavandy, S. K., Shameli, K., Biak, D. R. B. A., Abidin, Z. Z. (2014). Stirring time effect of silver nanoparticles prepared in glutathione mediated by green method. *Chemistry Central Journal*, 8(1): 11.
- Biçer, M., Şişman, İ. (2010). Controlled synthesis of copper nano/microstructures using ascorbic acid in aqueous CTAB solution. *Powder Technology*, 198(2): 279-284.
- Chatterjee, A. K., Sarkar, R. K., Chattopadhyay, A. P., Aich, P., Chakraborty, R., Basu, T. (2012). A simple robust method for synthesis of metallic copper nanoparticles of high antibacterial potency against *E. coli*. *Nanotechnology*, 23(8): 085103.
- Chattopadhyay, D., Patel, B. (2012). Preparation, characterization and stabilization of nano sized copper particles. *Int. J. Pure Appl. Sci. Technol*, 9(1): 1-8.
- Chattopadhyay, K. (2009). *Introduction To Nanoscience And Nenotechnology*: PHI Learning Pvt. Ltd.

- Chen, D.-H., Hsieh, C.-H. (2002). Synthesis of nickel nanoparticles in aqueous cationic surfactant solutions. *Journal of Materials Chemistry*, 12(8): 2412-2415.
- Chen, P., Song, L., Liu, Y., Fang, Y.-e. (2007). Synthesis of silver nanoparticles by γ -ray irradiation in acetic water solution containing chitosan. *Radiation Physics and Chemistry*, 76(7): 1165-1168.
- Chen, Y., Munechika, K., Ginger, D. S. (2007). Dependence of fluorescence intensity on the spectral overlap between fluorophores and plasmon resonant single silver nanoparticles. *Nano letters*, 7(3): 690-696.
- Cho, K.-H., Park, J.-E., Osaka, T., Park, S.-G. (2005). The study of antimicrobial activity and preservative effects of nanosilver ingredient. *Electrochimica Acta*, 51(5): 956-960.
- Chung, Y.-C., Su, Y.-P., Chen, C.-C., Jia, G., Wang, H.-I., Wu, J. G., Lin, J.-G. (2004). Relationship between antibacterial activity of chitosan and surface characteristics of cell wall. *Acta Pharmacologica Sinica*, 25932-936.
- Cioffi, N., Rai, M. (2012). *Nano-Antimicrobials*: Springer.
- Dadgostar, N., Ferdous, S., Henneke, D. (2010). Colloidal synthesis of copper nanoparticles in a two-phase liquid-liquid system. *Materials Letters*, 64(1): 45-48.
- Dang, T. M. D., Le, T. T. T., Fribourg-Blanc, E., Dang, M. C. (2011). Synthesis and optical properties of copper nanoparticles prepared by a chemical reduction method. *Advances in Natural Sciences: Nanoscience and Nanotechnology*, 2(1): 015009.
- Dhanalekshmi, K., Meena, K. (2014). Comparison of antibacterial activities of Ag@ TiO₂ and Ag@ SiO₂ core-shell nanoparticles. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 128887-890.
- Dong, C., Cai, H., Zhang, X., Cao, C. (2014). Synthesis and characterization of monodisperse copper nanoparticles using gum acacia. *Physica E: Low-dimensional Systems and Nanostructures*, 5712-20.
- Gao, G., Wu, H., He, R., Cui, D. (2010). Corrosion inhibition during synthesis of Cu₂O nanoparticles by 1, 3-diaminopropylene in solution. *Corrosion Science*, 52(9): 2804-2812.
- Ghodselahi, T., Vesaghi, M., Shafiekhani, A. (2009). Study of surface plasmon resonance of Cu@ Cu₂O core-shell nanoparticles by Mie theory. *Journal of Physics D: Applied Physics*, 42(1): 015308.

- Ghodselahe, T., Vesaghi, M., Shafiekhani, A., Baghizadeh, A., Lameii, M. (2008). XPS study of the Cu@Cu₂O core-shell nanoparticles. *Applied Surface Science*, 255(5): 2730-2734.
- Gómez-Ordóñez, E., Rupérez, P. (2011). FTIR-ATR spectroscopy as a tool for polysaccharide identification in edible brown and red seaweeds. *Food Hydrocolloids*, 25(6): 1514-1520.
- Gopinath, M., Subbaiya, R., Selvam, M. M., Suresh, D. (2014). Synthesis of Copper Nanoparticles from Nerium oleander Leaf aqueous extract and its Antibacterial Activity. *Int. J. Curr. Microbiol. App. Sci*, 3(9): 814-818.
- Guzmán, M. G., Dille, J., Godet, S. (2009). Synthesis of silver nanoparticles by chemical reduction method and their antibacterial activity. *Int J Chem Biomol Eng*, 2(3): 104-111.
- Han, W.-K., Choi, J.-W., Hwang, G.-H., Hong, S.-J., Lee, J.-S., Kang, S.-G. (2006). Fabrication of Cu nano particles by direct electrochemical reduction from CuO nano particles. *Applied Surface Science*, 252(8): 2832-2838.
- Honary, S., Barabadi, H., Gharaei-Fathabad, E., Naghib, F. (2012). Green synthesis of copper oxide nanoparticles using *Penicillium aurantiogriseum*, *Penicillium citrinum* and *Penicillium waksmanii*. *Dig J Nanomater Bios*, 7999-1005.
- Hong, Z.-s., Cao, Y., Deng, J.-f. (2002). A convenient alcohothermal approach for low temperature synthesis of CuO nanoparticles. *Materials Letters*, 52(1): 34-38.
- Hoover, N. N., Auten, B. J., Chandler, B. D. (2006). Tuning supported catalyst reactivity with dendrimer-templated Pt-Cu nanoparticles. *The Journal of Physical Chemistry B*, 110(17): 8606-8612.
- Huang, H., Yan, F., Kek, Y., Chew, C., Xu, G., Ji, W., Oh, P., Tang, S. (1997). Synthesis, characterization, and nonlinear optical properties of copper nanoparticles. *Langmuir*, 13(2): 172-175.
- Jortner, J., Rao, C. (2002). Nanostructured advanced materials. Perspectives and directions. *Pure and applied chemistry*, 74(9): 1491-1506.
- Joshi, S., Patil, S., Iyer, V., Mahumuni, S. (1998). Radiation induced synthesis and characterization of copper nanoparticles. *Nanostructured materials*, 10(7): 1135-1144.

- Kannan, R., Stirk, W., Van Staden, J. (2013). Synthesis of silver nanoparticles using the seaweed *Codium capitatum* PC Silva (Chlorophyceae). *South African Journal of Botany*, 86:1-4.
- Kashima, K., Imai, M. (2012). Advanced Membrane Material from Marine Biological Polymer and Sensitive Molecular-Size Recognition for Promising Separation Technology.
- Kobayashi, Y., Abe, Y., Maeda, T., Yasuda, Y., Morita, T. (2014). A metal-metal bonding process using metallic copper nanoparticles produced by reduction of copper oxide nanoparticles. *Journal of Materials Research and Technology*, 3(2): 114-121.
- Krishnaraj, C., Jagan, E., Rajasekar, S., Selvakumar, P., Kalaichelvan, P., Mohan, N. (2010). Synthesis of silver nanoparticles using *Acalypha indica* leaf extracts and its antibacterial activity against water borne pathogens. *Colloids and Surfaces B: Biointerfaces*, 76(1): 50-56.
- Kruis, F. E., Fissan, H., Rellinghaus, B. (2000). Sintering and evaporation characteristics of gas-phase synthesis of size-selected PbS nanoparticles. *Materials Science and Engineering: B*, 69:329-334.
- Kumar, K. S., Ganesan, K., Rao, P. (2008). Antioxidant potential of solvent extracts of *Kappaphycus alvarezii* (Doty) Doty—An edible seaweed. *Food chemistry*, 107(1): 289-295.
- Kumar, P., Selvi, S. S., Govindaraju, M. (2013). Seaweed-mediated biosynthesis of silver nanoparticles using *Gracilaria corticata* for its antifungal activity against *Candida* spp. *Applied Nanoscience*, 3(6): 495-500.
- Leal, D., Matsuhira, B., Rossi, M., Caruso, F. (2008). FT-IR spectra of alginic acid block fractions in three species of brown seaweeds. *Carbohydrate research*, 343(2): 308-316.
- Lee, Y., Choi, J.-r., Lee, K. J., Stott, N. E., Kim, D. (2008). Large-scale synthesis of copper nanoparticles by chemically controlled reduction for applications of inkjet-printed electronics. *Nanotechnology*, 19(41): 415604.
- Li, S.-K., Guo, X., Wang, Y., Huang, F.-Z., Shen, Y.-H., Wang, X.-M., Xie, A.-J. (2011). Rapid synthesis of flower-like Cu₂O architectures in ionic liquids by the assistance of microwave irradiation with high photochemical activity. *Dalton Transactions*, 40(25): 6745-6750.

- Link, S., El-Sayed, M. A. (2003). Optical properties and ultrafast dynamics of metallic nanocrystals. *Annual review of physical chemistry*, 54(1): 331-366.
- Liu, X., Geng, B., Du, Q., Ma, J., Liu, X. (2007). Temperature-controlled self-assembled synthesis of CuO, Cu₂O and Cu nanoparticles through a single-precursor route. *Materials Science and Engineering: A*, 448(1): 7-14.
- Liu, Z., Bando, Y. (2003). A novel method for preparing copper nanorods and nanowires. *Advanced Materials*, 15(4): 303-305.
- Liu, Z., Gao, Y., Bando, Y. (2002). Highly effective metal vapor absorbents based on carbon nanotubes. *Applied physics letters*, 81(25): 4844-4846.
- Long, D., Wu, G., Chen, S. (2007). Preparation of oligochitosan stabilized silver nanoparticles by gamma irradiation. *Radiation Physics and Chemistry*, 76(7): 1126-1131.
- Longano, D., Ditaranto, N., Sabbatini, L., Torsi, L., Cioffi, N. (2012). Synthesis and antimicrobial activity of copper nanomaterials *Nano-Antimicrobials* (pp. 85-117): Springer.
- Ma, J., Lin, Y., Chen, X., Zhao, B., Zhang, J. (2014). Flow behavior, thixotropy and dynamical viscoelasticity of sodium alginate aqueous solutions. *Food Hydrocolloids*, 38: 119-128.
- Madigan, M. T., Martinko, J. M., Dunlap, P. V., Clark, D. P. (2008). Brock Biology of microorganisms 12th edn. *International Microbiology*, 1165-73.
- Mott, D., Galkowski, J., Wang, L., Luo, J., Zhong, C.-J. (2007). Synthesis of size-controlled and shaped copper nanoparticles. *Langmuir*, 23(10): 5740-5745.
- MubarakAli, D., Thajuddin, N., Jeganathan, K., Gunasekaran, M. (2011). Plant extract mediated synthesis of silver and gold nanoparticles and its antibacterial activity against clinically isolated pathogens. *Colloids and Surfaces B: Biointerfaces*, 85(2): 360-365.
- Muller, V., Rasp, M., Stefanić, G., Ba, J., Günther, S., Rathousky, J., Niederberger, M., Fattakhova-Rohlfing, D. (2009). Highly Conducting Nanosized Monodispersed Antimony-Doped Tin Oxide Particles Synthesized via Nonaqueous Sol- Gel Procedure. *Chemistry of materials*, 21(21): 5229-5236.

- Naghavi, K., Saion, E., Rezaee, K., Yunus, W. M. M. (2010). Influence of dose on particle size of colloidal silver nanoparticles synthesized by gamma radiation. *Radiation Physics and Chemistry*, 79(12): 1203-1208.
- Nasrollahzadeh, M., Sajadi, S. M., Khalaj, M. (2014). Green synthesis of copper nanoparticles using aqueous extract of the leaves of *Euphorbia esula* L and their catalytic activity for ligand-free Ullmann-coupling reaction and reduction of 4-nitrophenol. *RSC Advances*, 4(88): 47313-47318.
- Ponce, A. A., Klabunde, K. J. (2005). Chemical and catalytic activity of copper nanoparticles prepared via metal vapor synthesis. *Journal of Molecular Catalysis A: Chemical*, 225(1): 1-6.
- Rabea, E. I., Badawy, M. E.-T., Stevens, C. V., Smagghe, G., Steurbaut, W. (2003). Chitosan as antimicrobial agent: applications and mode of action. *Biomacromolecules*, 4(6): 1457-1465.
- Raffi, M., Mehrwan, S., Bhatti, T. M., Akhter, J. I., Hameed, A., Yawar, W., ul Hasan, M. M. (2010). Investigations into the antibacterial behavior of copper nanoparticles against *Escherichia coli*. *Annals of microbiology*, 60(1): 75-80.
- Ramanathan, R., Bhargava, S. K., Bansal, V. (2011). Biological synthesis of copper/copper oxide nanoparticles.
- Ramnani, S., Biswal, J., Sabharwal, S. (2007). Synthesis of silver nanoparticles supported on silica aerogel using gamma radiolysis. *Radiation Physics and Chemistry*, 76(8): 1290-1294.
- Ramyadevi, J., Jeyasubramanian, K., Marikani, A., Rajakumar, G., Rahuman, A. A. (2012). Synthesis and antimicrobial activity of copper nanoparticles. *Materials Letters*, 71114-116.
- Roco, M. C. (2001). International strategy for nanotechnology research. *Journal of Nanoparticle Research*, 3(5-6): 353-360.
- Rotello, V. M. (2004). *Nanoparticles: building blocks for nanotechnology*: Springer.
- Ruparelia, J. P., Chatterjee, A. K., Duttagupta, S. P., Mukherji, S. (2008). Strain specificity in antimicrobial activity of silver and copper nanoparticles. *Acta Biomaterialia*, 4(3): 707-716.
- Salavati-Niasari, M., Davar, F., Mir, N. (2008). Synthesis and characterization of metallic copper nanoparticles via thermal decomposition. *Polyhedron*, 27(17): 3514-3518.

- Salgueiro, A. M., Daniel-da-Silva, A. L., Girão, A. V., Pinheiro, P. C., Trindade, T. (2013). Unusual dye adsorption behavior of κ -carrageenan coated superparamagnetic nanoparticles. *Chemical Engineering Journal*, 229276-284.
- Sánchez-Ramírez, J., Vazquez-Lopez, C., Pal, U. (2002). Preparation and optical absorption of colloidal dispersion of Au/Cu nanoparticles. *Superficies y vacío*, 1516-18.
- Sartori, C., Finch, D. S., Ralph, B., Gilding, K. (1997). Determination of the cation content of alginate thin films by FTIR spectroscopy. *Polymer*, 38(1): 43-51.
- Sau, T. K., Murphy, C. J. (2004). Room temperature, high-yield synthesis of multiple shapes of gold nanoparticles in aqueous solution. *Journal of the American Chemical Society*, 126(28): 8648-8649.
- Sawai, J. (2003). Quantitative evaluation of antibacterial activities of metallic oxide powders (ZnO, MgO and CaO) by conductimetric assay. *Journal of Microbiological Methods*, 54(2): 177-182.
- Şen, M., Erboz, E. N. (2010). Determination of critical gelation conditions of κ -carrageenan by viscosimetric and FT-IR analyses. *Food research international*, 43(5): 1361-1364.
- Soon, A., Todorova, M., Delley, B., Stampfl, C. (2007). Thermodynamic stability and structure of copper oxide surfaces: A first-principles investigation. *Physical Review B*, 75(12): 125420.
- Surmawar, N. V., Thakare, S. R., Khaty, N. (2011). One-Pot, Single Step Green Synthesis of Copper Nanoparticles: SPR Nanoparticles. *International Journal of Green Nanotechnology*, 3(4): 302-308.
- Thakore, S., Valodkar, M., Soni, J. Y., Vyas, K., Jadeja, R. N., Devkar, R. V., Rathore, P. S. (2013). Synthesis and cytotoxicity evaluation of novel acylated starch nanoparticles. *Bioorganic chemistry*, 4626-30.
- Tian, K., Liu, C., Yang, H., Ren, X. (2012). In situ synthesis of copper nanoparticles/polystyrene composite. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 39712-15.
- Tsuji, T., Iryo, K., Nishimura, Y., Tsuji, M. (2001). Preparation of metal colloids by a laser ablation technique in solution: influence of laser wavelength on the ablation efficiency (II). *Journal of Photochemistry and Photobiology A: Chemistry*, 145(3): 201-207.

- Urano, T., Ina, S. (2007). Powdery mixture of a calcium hydroxide optionally containing magnesium hydroxide with one a carrageenan having a six-membered-ring galactose skeleton and bonded in equatorial conformation to a sulfate group and a hydroxyl group and being kneaded together with water at time of use: Google Patents.
- Usman, M. S., Ibrahim, N. A., Shameli, K., Zainuddin, N., Yunus, W. M. Z. W. (2012). Copper nanoparticles mediated by chitosan: synthesis and characterization via chemical methods. *Molecules*, 17(12): 14928-14936.
- Valodkar, M., Jadeja, R. N., Thounaojam, M. C., Devkar, R. V., Thakore, S. (2011a). Biocompatible synthesis of peptide capped copper nanoparticles and their biological effect on tumor cells. *Materials Chemistry and Physics*, 128(1): 83-89.
- Valodkar, M., Pal, A., Thakore, S. (2011b). Synthesis and characterization of cuprous oxide dendrites: New simplified green hydrothermal route. *Journal of Alloys and Compounds*, 509(2): 523-528.
- Valodkar, M., Rathore, P. S., Jadeja, R. N., Thounaojam, M., Devkar, R. V., Thakore, S. (2012). Cytotoxicity evaluation and antimicrobial studies of starch capped water soluble copper nanoparticles. *Journal of hazardous materials*, 201244-249.
- Wahab, R., Mishra, A., Yun, S.-I., Kim, Y.-S., Shin, H.-S. (2010). Antibacterial activity of ZnO nanoparticles prepared via non-hydrolytic solution route. *Applied microbiology and biotechnology*, 87(5): 1917-1925.
- Wang, C., Daimon, H., Lee, Y., Kim, J., Sun, S. (2007). Synthesis of monodisperse Pt nanocubes and their enhanced catalysis for oxygen reduction. *Journal of the American Chemical Society*, 129(22): 6974-6975.
- Warschkow, O., Chuasiripattana, K., Lyle, M., Delley, B., Stampfl, C. (2011). Cu/ZnO (0001) under oxidating and reducing conditions: A first-principles survey of surface structures. *Physical Review B*, 84(12): 125311.
- Wen, J., Li, J., Liu, S., Chen, Q.-y. (2011). Preparation of copper nanoparticles in a water/oleic acid mixed solvent via two-step reduction method. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 373(1): 29-35.
- Wu, C., Mosher, B. P., Zeng, T. (2006). One-step green route to narrowly dispersed copper nanocrystals. *Journal of Nanoparticle Research*, 8(6): 965-969.

- Wu, D., Chen, Z., Cai, K., Zhuo, D., Chen, J., Jiang, B. (2014). Investigation into the antibacterial activity of monodisperse BSA-conjugated zinc oxide nanoparticles. *Current Applied Physics*, 14(11): 1470-1475.
- Xiao, Q., Gu, X., Tan, S. (2014). Drying process of sodium alginate films studied by two-dimensional correlation ATR-FTIR spectroscopy. *Food chemistry*, 164:179-184.
- Xie, S.-Y., Ma, Z.-J., Wang, C.-F., Lin, S.-C., Jiang, Z.-Y., Huang, R.-B., Zheng, L.-S. (2004). Preparation and self-assembly of copper nanoparticles via discharge of copper rod electrodes in a surfactant solution: a combination of physical and chemical processes. *Journal of Solid State Chemistry*, 177(10): 3743-3747.
- Xiong, J., Wu, X.-d., Xue, Q.-j. (2013). Biomolecule-assisted synthesis of highly stable dispersions of water-soluble copper nanoparticles. *Journal of colloid and interface science*, 390(1): 41-46.
- Yallappa, S., Manjanna, J., Sindhe, M., Satyanarayan, N., Pramod, S., Nagaraja, K. (2013). Microwave assisted rapid synthesis and biological evaluation of stable copper nanoparticles using T. arjuna bark extract. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 110:108-115.
- Yun, J., Cho, K., Park, B., Kang, H. C., Ju, B.-K., Kim, S. (2008). Optical Heating of Ink-Jet Printable Ag and Ag-Cu Nanoparticles. *Japanese Journal of Applied Physics*, 47(6S): 5070.
- Zhang, L., Jiang, Y., Ding, Y., Povey, M., York, D. (2007). Investigation into the antibacterial behaviour of suspensions of ZnO nanoparticles (ZnO nanofluids). *Journal of Nanoparticle Research*, 9(3): 479-489.
- Zhang, L., Webster, T. J. (2009). Nanotechnology and nanomaterials: promises for improved tissue regeneration. *Nano Today*, 4(1): 66-80.
- Zhang, N., Gao, Y., Zhang, H., Feng, X., Cai, H., Liu, Y. (2010). Preparation and characterization of core-shell structure of SiO₂@ Cu antibacterial agent. *Colloids and Surfaces B: Biointerfaces*, 81(2): 537-543.
- Zhang, Q.-L., Yang, Z.-M., Ding, B.-J., Lan, X.-Z., Guo, Y.-J. (2010). Preparation of copper nanoparticles by chemical reduction method using potassium borohydride. *Transactions of Nonferrous Metals Society of China*, 20:240-s244.
- Zhang, Z., Che, H., Gao, J., Wang, Y., She, X., Sun, J., Gunawan, P., Zhong, Z., Su, F. (2012a). Shape-controlled synthesis of Cu₂O microparticles

and their catalytic performances in the Rochow reaction. *Catalysis Science & Technology*, 2(6): 1207-1212.

Zhang, Z., Che, H., Wang, Y., Gao, J., Ping, Y., Zhong, Z., Su, F. (2012b). Template-free synthesis of Cu@Cu₂O core-shell microspheres and their application as copper-based catalysts for dimethyldichlorosilane synthesis. *Chemical Engineering Journal*, 211421-431.

Zhao, X., Xia, Y., Li, Q., Ma, X., Quan, F., Geng, C., Han, Z. (2014). Microwave-assisted synthesis of silver nanoparticles using sodium alginate and their antibacterial activity. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 444180-188.

Zhou, Y., Yu, S. H., Wang, C. Y., Li, X. G., Zhu, Y. R., Chen, Z. Y. (1999). A novel ultraviolet irradiation photoreduction technique for the preparation of single-crystal Ag nanorods and Ag dendrites. *Advanced Materials*, 11(10): 850-852.