

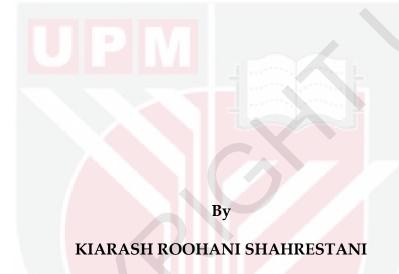
UNIVERSITI PUTRA MALAYSIA

CHARACTERIZATION OF NEWCASTLE DISEASE VIRUS (NDV)
ISOLATED FROM NDV VACCINATED BROILER FARMS AND
INVESTIGATION OF VACCINE EFFICACY AGAINST
CHALLENGE WITH VELOGENIC GENOTYPE VII NDV

KIARASH ROOHANI SHAHRESTANI



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Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

September 2014

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DEDICATION

This thesis is dedicated to my parents who have supported me all the way since the beginning of my life and my dear sister. Also, this thesis is dedicated to my beloved wife who has been a great source of motivation and inspiration.

Finally, this thesis is dedicated to all those who believe in the richness of learning.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the Degree of Master of Science

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By

KIARASH ROOHANI SHAHRESTANI

September 2014

Chairman: Abdul Rahman Omar, DVM, PhD

Faculty: Institute of Bioscience

Vaccines to control Newcastle disease (ND) were introduced more than 60 years ago. Despite that, ND is still one of the most significant avian diseases affecting major poultry farms in various countries. Class II of Newcastle disease viruses (NDV) can be divided into 10 different genotypes based on the F gene. However, since 1990s genotype VII NDV is the predominant velogenic NDV circulating in South-East Asia causing outbreaks even in well vaccinated flocks. Various factors such as inappropriate vaccination scheme, concurrent infections, immunosuppression and presence of variant NDV have been implicated as the probable causes of the outbreak. The current NDV vaccines comprised of genotype I and/or II viruses. Through the use of reverse genetic technology, genotype—matched NDV vaccine has been used in certain countries. However, the efficacy of this vaccine against the circulating velogenic genotype VII NDV is not well characterized.

This study focused on the isolation and characterization of velogenic NDV from NDV vaccinated chickens in Malaysia. Sequencing and phylogenetic analysis based on the F gene of five NDV isolates (IBS001 to IBS005) showed that the viruses belong to genotype VII and sub-genotype VIId of NDV, with F cleavage site motif of ¹¹²RRRKRF¹¹⁷. In addition, sequencing of the C-terminus of the HN gene revealed that, viruses lack extension and encoded a typical amino acid sequence length of virulent NDV. Hence, molecular characterization based on the F and HN genes indicated the viruses (IBS001 to IBS005) belong to velogenic genotype VII NDV.

One of the isolates, IBS002, was further characterized based on sequencing of the complete length of F and HN genes and pairwise comparisons between different genotypes. A maximum distance was detected between IBS002 and LaSota with nucleotide/amino acids variation between 17.71% to 18.67% for

F gene and 20.89% to 23.37% for HN gene. Nucleotide/amino acids variations of 8.79% to 9.77% for F gene and 9.17% to 11.60% for HN genes were detected between isolate IBS002 and genotype VII Dalguban N+vaccine. In addition, IBS002 has a mean death time (MDT) of 51.2 hours and intracerebral pathogenicity index (ICPI) of 1.76, further confirming that the virus is a velogenic strain.

Both genotype matched (Dalguban N+) and mismatched (LaSota and Avinew) vaccines induced 100% protection against mortality and severe clinical symptoms following challenge with 10⁵ ELD₅₀ of IBS002. Vaccinated chickens also showed significant (P<0.05) lower pathogenicity scores although there was no significant (P<0.05) difference among the vaccinated groups. However, sentinel birds of Avinew and Dalguban N+ groups showed lower pathogenicity score compared to sentinels in LaSota group (P<0.05). Furthermore, Avinew and Dalguban N+ vaccinated chickens shed significantly (P<0.05) less virus after challenge and the viral load decreased faster than LaSota group. Moreover, sentinel birds mortality in LaSota vaccinated and non-vaccinated groups were significantly (P<0.05) higher than Avinew and Dalguban N+ vaccinated groups suggesting the importance of genotype matched vaccine (Dalguban N+) and enteric based NDV vaccine (Avinew) in inducing vaccine induced immunity. Vaccine that matched with the hemagglutination-inhibition (HI) test's antigen induced significantly (P<0.05) higher antibody compared to vaccine from other genotypes where a 2 Log₂ difference were detected when genotype VII and genotype II NDV antigens were used to detect homologous and heterologous HI titters.

Immunophenotyping study showed significant increased (P<0.01) in KUL-1+ macrophages in PBMCs and splenocytes of control challenged birds. On the other hand, CD3+/CD4+ and CD3+/CD8+ T cells in spleen of different vaccinated groups were increased upon challenge suggesting the possible involvement of these cells in curtailing virus replication.

In conclusion, isolated NDVs were classified as velogenic strains and belonged to genotype VIId of class II of NDV. Both genotype matched and mismatched NDV vaccines were able to confer protection against challenge with velogenic genotype VII NDV. However, genotype matched and enteric based NDV vaccines seems to be able to confer a more complete protection against virus shedding and transmission to susceptible chickens following challenged with velogenic genotype VII NDV.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Sarjana Sains

PENCIRIAN VIRUS PENYAKIT SAMPAR (NDV) DARI LADANG AYAM PEDAGING YANG DISUNTIK VAKSIN NDV DAN SIASATAN EFIKASI VAKSIN TERHADAP CABARAN DENGAN NDV GENOTIP VII VELOGENIK

Oleh

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September 2014

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Walaupun lebih 60 tahun, vaksin bagi mengawal penyakit sampar (ND) telah diperkenalkan, ND masih merupakan salah satu penyakit unggas yang mempunyai impak besar terhadap ladang ayam di setiap pelusuk negara. Virus penyakit sampar (NDV) kelas II boleh dibahagikan kepada 10 genotip yang berbeza berdasarkan kepada gen F. Walau bagaimanapun, sejak tahun 1990an NDV genotip VII adalah virus velogenik utama yang tersebar di Asia Tenggara dan menyebabkan wabak mahupun dalam ayam yang disuntik vaksin. Pelbagai faktor yang menyumbang kepada penularan wabak ini termasuklah skim vaksinasi yang tidak sesuai, jangkitan secara serentak, imunotindas dan juga kemunculan virus penyakit sampar (NDV) varian. Vaksin NDV yang ada pada masa kini terdiri daripada virus genotip I dan/atau genotip II. Walau bagaimanapun, melalui penggunaan teknologi genetik terbalik, vaksin NDV genotip-padanan telah digunakan di negaranegara tertentu. Tetapi, efikasi vaksin ini terhadap NDV genotip VII beredar velogenik tidak dicirikan dengan begitu jelas lagi.

Kajian ini bertumpu kepada pengasingan dan pencirian NDV velogenik daripada ayam yang disuntik vaksin di Malaysia. Penjujukan dan analisis filogenetik berdasarkan gen F daripada lima isolat NDV (IBS001 hingga IBS005) menunjukkan bahawa virus-virus ini terdiri daripada NDV genotip VII dan sub-genotip VIId dengan tapak belahan F bermotifkan ¹¹²RRRKRF¹¹⁷. Selain daripada itu, jujukan terminus C gen HN menunjukkan bahawa virus-virus ini tidak tidak mempunyai lanjutan jujukan dengan panjang jujukan asid amino yang kerap ditemui bagi NDV virulen. Oleh itu, pencirian molekul berdasarkan gen F dan HN menunjukkan virus-virus ini (IBS001 hingga IBS005) tergolong dalam genotip VII NDV velogenik. Salah satu virus yang diasingkan, IBS002, telah dilanjutkan penciriannya berdasarkan jujukan lengkap gen F dan HN, dan seterusnya dibandingkan di antara genotip yang berbeza. Satu perbezaan jarak maksimum dikesan di antara IBS002 dan

LaSota dengan nukleotida/asid amino, daripada 17.71% kepada 18.67% bagi gen F dan 20.89% kepada 23.37% bagi gen HN. Variasi nukleotida/asid amino daripada 8.79% kepada 9.77% untuk gen F dan 9.17% kepada 11.60% bagi gen HN telah ditemui di antara IBS002 dan vaksin genotip VII Dalguban N+. Di samping itu, IBS002 mempunyai purata masa kematian (MDT) iaitu 51.2 jam dan indeks patogenisiti intraserebrum (ICPI) iaitu 1.76 seterusnya mengesahkan bahawa virus ini tergolong dalam kumpulan NDV velogenik.

Kedua-dua vaksin genotip-padanan (Dalguban N+) dan tidak padanan (LaSota dan Avinew), memberi perlindungan 100% terhadap kematian dan gejala klinikal yang teruk ekoran cabaran dengan IBS002 pada sukatan virus 10⁵ ELD₅₀. Ayam yang disuntik vaksin menunjukkan bacaan skor patogenisiti rendah yang signifikan (P<0.05), manakala tiada sebarang perbezaan yang signifikan (P<0.05) didapati di kalangan kumpulan ayam yang disuntik vaksin. Walau bagaimanapun, ayam sentinel daripada kumpulan Avinew dan Dalguban N+ menunjukkan skor patogenisiti yang lebih rendah berbanding dengan sentinel dalam kumpulan LaSota (P<0.05). Selain itu, ayam yang telah disuntik vaksin Avinew dan Dalguban N+ menunjukkan peluruhan virus yang rendah (P<0.05) selepas cabaran dan kuantiti virus menurun dengan lebih cepat daripada kumpulan LaSota. Tambahan lagi, skor kematian ayam sentinel dalam kumpulan vaksin LaSota dan kumpulan yang tidak disuntik vaksin (P<0.05) adalah sangat tinggi jika dibandingkan dengan Avinew dan Dalguban N+ mencadangkan kepentingan vaksin genotip-padanan (Dalguban N+) dan vaksin berasas enterik dalam mengaruh keimunan vaksin. Vaksin yang sepadan dengan antigen ujian perencatan-hemagglutinin (HI) menunjukkan titter antibodi yang lebih tinggi (P<0.05) berbanding dengan vaksin dari genotip lain di mana perbezaan 2 Log2 dikesan pada titter antibodi HI homolog dan heterolog bila virus genotip VII dan II digunakan sebagai antigen.

Kajian imunofenotip menunjukkan ayam disuntik vaksin LaSota mempunyai peratus makrofaj KUL-1+ dan IgM+ B limfosit yang lebih tinggi berbanding dengan kumpulan disuntik vaksin lain. Walaupun begitu, tiada perbezaan yang ketara dikesan pada kedua-dua sel T limpa, CD3+/CD4+ dan CD3 +/CD8+ dikalangan kumpulan yang disuntik vaksin yang berbeza. Peningkatan yang ketara (P<0.01) makrofaj KUL-1+ dalam PBMCs dan limpa telah dikesan dalam kumpulan kawalan yang dicabar. Sungguhpun begitu, peningkatan yang ketara pada jumlah sel T (P<0.01) limpa dari ayam yang disuntik vaksin dan dicabar dengan IBS002 mungkin menunjukkan peranan penting sel ini dalam mengekang replikasi virus. Kesimpulannya, kedua-dua genotip-padanan dan tidak padanan dapat memberikan perlindungan terhadap cabaran dengan NDV genotip VII velogenik. Walau bagaimanapun, vaksin NDV padanan dan berasaskan enterik dapat memberikan perlindungan yang lebih lengkap terhadap peluruhan virus dan transmisi virus kepada perumah rentan.

ACKNOWLEGEMENTS

This research project would not have been possible without the support of many people. The author wishes to express his gratitude to his supervisor, Prof. Dr. Abdul Rahman Omar who was abundantly helpful and offered invaluable assistance, support and guidance. Deepest gratitude are also due to the members of the supervisory committees, Prof. Datin Paduka Dr. Aini Ideris, Prof. Dr. Mohd Hair Bejo, Dr. Tan Sheau Wei and Dr. Yeap Swee Keong without whose knowledge and assistance, this study would not have been successful.

Special thanks also to all the graduate students and friends, especially group members; Dr. Davoud Jazayeri, Dr. Mehdi Rasouli Piroziyan, Mozhgan Mollayee, Farhana Zaini, Nor Yasmin, Amanda Teo, Kristeen Teo, Aimi Jamil, Dilan and many others for sharing their ideas, information and invaluable assistance. In addition, many thanks to all the staff of Institute of Bioscience, especially staffs at Laboratory of Vaccines and Immunotherapeutics for their assistance and kindness.

The author would also like to convey thanks to the Ministry of Higher Education (MOHE), Government of Malaysia for providing Institute of Bioscience Higher Institution Center of Excellence (IBS HICoE) grant, Institute of BioScience for providing the financial means and laboratory facilities and Prof. Dr. Abdul Rahman Omar for providing the Special Graduate Research Assistantship, under MOHE grant.

The author wishes to express his love and gratitude to his beloved families; for their understanding and endless love, through the duration of the study.

I certify that a Thesis Examination Committee has met on 23 September 2014 to conduct the final examination of Kiarash Roohani Shahrestani on his thesis entitled "Characterization of Newcastle Disease Virus (NDV) Isolated from NDV-Vaccinated Broiler Farms and Investigation of Vaccine Efficacy Against Challenge with Velogenic Genotype VII NDV" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

Aa Amino Acid

APMV Avian Paramyxovirus

BLAST Basic Local alignment Search Tool

BSA Bovine Serum Albumin DNA Deoxyribonucleic Acid

dsRNA Double strand Deoxyribonucleic Acid

EDTA Ethylene-Diamine-Tetraacetic Acid

ELD₅₀ Mean Egg Lethal Dose

ELISA Enzyme-linked immunosorbent assays

F Fusion Protein
F0 Fusion Protein 0
F1 Fusion Protein 1
F2 Fusion Protein 2

GIT Gastrointestinal Tract

HI Hemagglutination Inhibition
HN Hemagglutinin-Neuraminidase

IBD Infectious Bursal Disease

ICPI Intra Cerebral Pathogenicity Index
IVPI Intra Venus Pathogenicity Index

Large Polymerase Protein

M Matrix Protein

MAB Monoclonal Antibodies

MDT Mean Death Time

MEGA Molecular Evolutionary Genetics Analysis
NCBI National Center for Biotechnology Information

ND Newcastle Disease

NDV Newcastle Disease Virus NP Nucleocapsid Protein

NV-ND Neurotropic Velogenic Newcastle Disease

OIE Office International des Epizooties

P Phosphoprotein Protein

PBMC Peripheral Blood Mononuclear Cells

PBS Phosphate Buffered Saline PCR Polymerase Chain Reaction

rHVT Turkey Herpesvirus-based Recombinant Vaccine

RNA Ribonucleic Acid

Rpm Revolutions Per Minute RT Reverse Transcription

RT-PCR Reverse Transcription-Polymerase Chain Reaction

SPF Specific-Pathogen-Free

TAE Tris-Acetate-EDTA

VG/GA Villegas-Glisson/University of Georgia VV-ND Viscerotropic Velogenic Newcastle Disease



CHAPTER 1

INTRODUCTION

After more than 80 years since the first discovery of Newcastle disease (ND) in England and Java (Kraneveld, 1926; Doyle, 1927) and the introduction of vaccine in 1950s to control the disease (Alexander, 2008), ND is still the most significant avian disease that continue to cause huge economical loss to the poultry industry. Because of its worldwide outbreaks and geographical spread, World Organization for Animal Health (OIE) has listed ND as a notifiable disease (OIE, 2013). The disease is caused by Newcastle disease virus (NDV), a highly infectious agent capable of causing high mortality in non-vaccinated chickens, beside the subclinical forms of ND in vaccinated and/or NDV exposure flocks which could have synergist effect with other bacterial or viral infections and cause more severe disease and bigger economic losses (Capua and Alexander, 2009).

Newcastle disease virus (NDV) is an avian paramyxovirus serotype-1 a member of sub-family Paramyxovirinae Paramyxoviridae from virus order Mononegavirales (Mayo 2002; Fauquet and Fargette 2005). The virus has a negative sense, single stranded RNA genome which encodes for 6 genes (Lamb and Parks, 2006). Although all APMV-1 viruses are from one serotype, they share different genome structures and have been divided into different genotypes (Diel et al., 2012). NDVs are categorized based on their F gene into 2 classes. Class I NDV were mostly isolated from water fowls (Anatidae) and shore birds whilst, Class II NDVs are able to induce disease in poultry, and are divided into 10 genotypes (Miller et al., 2010). Some of virulent NDV isolates from different countries in Europe during 1990s were found not belonged into any of known genotypes by that days, hence, were classified as genotype VII NDVs (Lomniczi et al., 1998). These NDVs are believed to be originated from East-Asia (Lomniczi et al., 1998), later spread to Europe, Africa, Middle-East and South Africa (Wang et al., 2006; Bogovavlenskiy et al., 2009). Recent studies have shown that presently genotype VII is the predominant circulating NDV in South-East Asia causing major outbreaks including in NDV vaccinated flocks (Cho et al., 2008a; Tan et al., 2010a; Yi et al., 2011; Umali et al., 2013).

Since the discovery of NDV, many efforts have been implemented to control ND in poultry industry. Beside biosecurity and good farm husbandry practices, ND is controlled through vaccination. Currently, several different vaccines are available commercially to control ND in both commercial chicken farms and backyard village chickens. Most of these vaccines belong to genotype II of class II of NDVs (Chong *et al.*, 2010). In Malaysia, like most of other countries, farmers are utilizing low pathogenic NDVs namely lentogenic NDV such as Hitchner B1 and LaSota, as live vaccines for ND prevention. Beside these two live vaccines, other NDV vaccine strains such

are S, Ulster 2C, NDV-6/10 and enteric vaccine strain VG-GA have also been used (Aini, 2006). Various type of genetically engineered ND vaccines have been developed and tested experimentally. However, only a few recombinant NDV vaccines are available commercially, namely, herpesvirus turkey virus (HVT) based NDV vaccine (Palya *et al.*, 2012). These vaccines have shown promising results in conferring protection against challenge with velogenic NDV.

Although it is possible to estimate vaccine efficacy through laboratory scale experiments, it would be very difficult to evaluate the vaccine efficacy in the field (Chulan et al., 1982). Hence, vaccination failure following NDV vaccination has been reported. Among the underlying factors that contribute to poor NDV vaccine induced immunity are inappropriate vaccination dose and regime, presence of concurrent infection especially immunosuppressive agents such as chicken infectious anaemia (CAV), infectious bursal disease (IBD) or Marek's disease, nutritional deficiencies as well as mycotoxins in feed being the probably reason(s) for the break in the vaccine induced immunity (Saif, 1991; Zhang et al., 2012; Habibian et al., 2013). However, recent studies have shown that, commercial NDV vaccines provide different level of protections against challenged with different genotypes of NDV (Hu et al., 2009; Miller et al., 2009) raising the importance of relatedness between vaccine and field strains of NDV. Furthermore, it has been demonstrated that, LaSota vaccine, a genotype II NDV is not effective in reducing virus shedding and clinical signs upon challenge with genotype VII isolates compared to reverse genetic designed genotype VII vaccine (Cho et al., 2008b; Hu et al., 2009). However, the importance of virus shedding and transmission to susceptible chickens is not well defined. Numerous studies have shown that NDV vaccine was able to provide protection against mortality (disease immunity) but unable to provide sterilising immunity by preventing infection following challenged with velogenic NDV (Ezema et al., 2009; Cornax et al., 2012).

In Malaysia, since 2000, genotype VII NDV has been reported from both non-vaccinated and vaccinated chicken flocks (Maizan *et al.*, 2001; Berhanu *et al.*, 2010; Tan *et al.*, 2010a). Recent survey indicated that genotype VII NDV is still circulating among the poultry flocks in Malaysia despite the extensive use of LaSota based vaccines (unpublished data). However, limited studies have been carried out in accessing the ability of genotype II (LaSota, B1, VG/GA, Avinew) (genotype mismatched vaccine) and genotype VII (genotype matched vaccine) in conferring protection against challenge with velogenic genotype VII in specific-pathogen-free (SPF) and commercial chicken flocks.

Hence, the first hypothesis of this study is that the genotype of NDV isolated from ND outbreaks from vaccinated broiler farms belongs to velogenic genotype VII that is highly pathogenic in chickens. The second hypothesis of

this study is vaccination with genotype matched NDV will provide better protection against genotype VII NDV challenge.

To address both hypotheses, the specific objectives of this study are:

- To isolate and characterize Newcastle disease virus (NDV) from Newcastle disease outbreaks originated from NDV vaccinated farms based on biological and molecular analysis.
- To investigate the efficacy of genotype matched and genotype mismatched NDV vaccines against velogenic genotype VII challenge in specific-pathogen-free chickens based on serological, biological and immunological analysis.



REFERENCES

- Abolnik, C., R. Horner, S. Bisschop, M. Parker, M. Romito and G. Viljoen (2004). "A phylogenetic study of South African Newcastle disease virus strains isolated between 1990 and 2002 suggests epidemiological origins in the far east." <u>Archives of Virology</u> **149**(3): 603-619.
- Adair, B. M. (2000). "Immunopathogenesis of chicken anemia virus infection." <u>Developmental & Comparative Immunology</u> **24**(2-3): 247-255.
- Ahmed, K. A., V. K. Saxena, A. Ara, K. B. Singh, N. R. Sundaresan, M. Saxena and T. J. Rasool (2007). "Immune response to Newcastle disease virus in chicken lines divergently selected for cutaneous hypersensitivity." International Journal of Immunogenetics **34**(6): 445-455.
- Aini, I. (2006). Newcastle disease. <u>Disease of Poultry in Southeast Asia.</u> Serdang, Malaysia, Malindo Printers Sdn. Bhd: 225-233.
- Aini, I., A. L. Ibrahim and P. B. Spradbrow (1990). "Field trials of a food-based vaccine to protect village chickens against Newcastle disease." Research in Veterinary Science 49(2): 216-219.
- Al-Habeeb, M. A., M. Mohamed and S. Sharawi (2013). "Detection and characterization of Newcastle disease virus in clinical samples using real time RT-PCR and melting curve analysis based on matrix and fusion genes amplification." <u>Veterinary World</u> **6**(5): 239-243.
- Aldous, E., J. Mynn, J. Banks and D. Alexander (2003a). "A molecular epidemiological study of avian paramyxovirus type 1 (Newcastle disease virus) isolates by phylogenetic analysis of a partial nucleotide sequence of the fusion protein gene." Avian Pathology **32**(3): 237-255.
- Aldous, E. W. and D. J. Alexander (2001). "Detection and differentiation of Newcastle disease virus (avian paramyxovirus type 1)." <u>Avian Pathology</u> **30**(2): 117-128.
- Aldous, E. W., J. K. Mynn, J. Banks and D. J. Alexander (2003). "A molecular epidemiological study of avian paramyxovirus type 1 (Newcastle disease virus) isolates by phylogenetic analysis of a partial nucleotide sequence of the fusion protein gene." <u>Avian Pathology</u> **32**(3): 239-256.
- Alexander, D. J. (1988a). Newcastle disease. D. J. Alexander. Boston, Kluwer Academic Publishers: 256–272.
- Alexander, D. J. (1988b). Newcastle disease diagnosis. <u>Newcastle disease</u>, Springer: 147-160.
- Alexander, D. J. (1995). "The epidemiology and control of avian influenza and Newcastle disease." <u>Journal of Comparative Pathology</u> **112**(2): 105-126.
- Alexander, D. J. (2000). "Newcastle disease and other avian paramyxoviruses." <u>Revue Scientifique et Technique-Office International des Epizooties</u> **19**(2): 443-462.
- Alexander, D. J. (2001). "Gordon memorial lecture. Newcastle disease." British Poultry Science **42**(1): 5-22.

- Alexander, D. J. (2008). Newcastle disease, other avian paramyxoviruses, and pneumovirus infections. <u>Diseases of Poultry</u>. Y. M. Saif, Wiley-Blackwell.
- Alexander, D. J. (2010). "Newcastle disease. Chapter 2.3. 14." <u>Manual of Diagnostic Tests and Vaccines for Terrestrial Animals</u>: 576-589.
- Alexander, D. J. (2011). "Newcastle disease in the European Union 2000 to 2009." Avian Pathology **40**(6): 547-558.
- Alexander, D. J., E. W. Aldous and C. M. Fuller (2012). "The long view: A selective review of 40 years of Newcastle disease research." <u>Avian Pathology</u> **41**(4): 329-335.
- Alexander, D. J. and W. H. Allan (1974). "Newcastle disease virus pathotypes." <u>Avian Pathology</u> **3**(4): 269-278.
- Allan, W. and L. Borland (1979). "The stress index: A method for indicating the pathogenicity of vaccinal Newcastle disease virus when administered by aerosol." <u>Avian Pathology</u> 8(4): 401-409.
- Allan, W. H. and R. E. Gough (1974). "A standard haemagglutination inhibition test for Newcastle disease. (1). A comparison of macro and micro methods." <u>Veterinary Record</u> 95(6): 120-123.
- Allan, W. H., J. E. Lancaster and B. Toth (1978). Newcastle disease vaccines, their production and use, Food and Agriculture Organization of the United Nations.
- Aslam, S. M., J. D. Garlich and M. A. Qureshi (1998). "Vitamin D deficiency alters the immune responses of broiler chicks." <u>Poultry Science</u> 77(6): 842-849.
- Awan, M. A., M. J. Otte and A. D. James (1994). "The epidemiology of Newcastle disease in rural poultry: A review." <u>Avian Pathology</u> **23**(3): 405-423.
- Ballagi-Pordany, A., E. Wehmann, J. Herczeg, S. Belak and B. Lomniczi (1996). "Identification and grouping of Newcastle disease virus strains by restriction site analysis of a region from the F gene." <u>Archives of Virology 141(2): 243-261.</u>
- Battisti, A. J., G. Meng, D. C. Winkler, L. W. McGinnes, P. Plevka, A. C. Steven, T. G. Morrison and M. G. Rossmann (2012). "Structure and assembly of a paramyxovirus matrix protein." <u>Proceedings of the National Academy of Sciences of the United States</u> **109**(35): 13996-14000.
- Beard, C. and R. Hanson (1984). "Newcastle disease". <u>Diseases of Poultry.</u>, M. S. Hofstad, H. J. Barns, B. W. Clarck, W. M. Reid, and H. W. Voder, Iowa State University Press, Ames, 1984, 452
- Beard, C. W. and B. C. Easterday (1967). "The influence of the route of administration of Newcastle disease virus on host response. I. Serological and virus isolation studies." <u>The Journal of Infectious Diseases</u> **117**(1): 55-61.
- Bennejean, G. (1988). Newcastle disease: Control policies. <u>Newcastle disease</u>, Springer: 303-317.

- Bennejean, G., M. Guittet, J. Picault, J. Bouquet, B. Devaux, D. Gaudry and Y. Moreau (1978). "Vccination of one-day-old chicks against Newcastle disease using inactivated oil adjuvant vaccine and/or live vaccine." Avian Pathology 7(1): 15-27.
- Berhanu, A. (2010). Molecular characterization of malaysian isolates of Newcastle disease viruses and the development of a duplex RT-PCR for simultaneous diagnosis of Newcastle disease and avian influenza. Master of Science, University of Putra Malaysia.
- Berhanu, A., A. Ideris, A. R. Omar and M. H. Bejo (2010). "Molecular characterization of partial fusion gene and c-terminus extension length of haemagglutinin-neuraminidase gene of recently isolated Newcastle disease virus isolates in malaysia." <u>Virology Journal</u> 7(1): 183.
- Bogoyavlenskiy, A., V. Berezin, A. Prilipov, E. Usachev, O. Lyapina, I. Korotetskiy, I. Zaitceva, S. Asanova, A. Kydyrmanov and K. Daulbaeva (2009). "Newcastle disease outbreaks in Kazakhstan and Kyrgyzstan during 1998, 2000, 2001, 2003, 2004, and 2005 were caused by viruses of the genotypes VII b and VII d." <u>Virus Genes</u> **39**(1): 94-101.
- Boursnell, M. E., P. F. Green, A. C. Samson, J. I. Campbell, A. Deuter, R. W. Peters, N. S. Millar, P. T. Emmerson and M. M. Binns (1990). "A recombinant fowlpox virus expressing the hemagglutinin-neuraminidase gene of Newcastle disease virus (NDV) protects chickens against challenge by NDV." <u>Virology Journal</u> **178**(1): 297-300.
- Box, P. G. and I. G. Furminger (1975). "Newcastle disease antibody levels in chickens after vaccination with oil emulsion adjuvant killed vaccine." Veterinary Record **96**(5): 108-111.
- Brandly, C. A., H. E. Moses, E. Jones, and E. L.Jungherr (1946). "Immunization of chickens against Newcastle disease." <u>American Journal of Veterinary Research</u> 7: 3107–3132.
- Brown, J., R. Resurreccion and T. Dickson (1990). "The relationship between the hemagglutination-inhibition test and the enzyme-linked immunosorbent assay for the detection of antibody to Newcastle disease." <u>Avian Diseases</u>: 585-587.
- Brugh Jr, M., C. Beard and W. Wilkes (1978). "The influence of test conditions on Newcastle disease hemagglutination-inhibition titers." <u>Avian Diseases</u>: 320-328.
- Bwala, D. G., C. Abolnik, A. Van Wyk, E. Cornelius and S. P. Bisschop (2009). Efficacy of a genotype 2 Newcastle disease vaccine (Avinew®) against challenge with highly virulent genotypes 5d and 3d. <u>Journal of the South African Veterinary Association</u> 80(3): 174-178.
- Calnek, B., R. Harris, C. Buscaglia, K. Schat and B. Lucio (1998). "Relationship between the immunosuppressive potential and the pathotype of marek's disease virus isolates." Avian Diseases: 124-132.

- Cannon, M. J. and P. H. Russell (1986). "Secondary in vitro stimulation of specific cytotoxic cells to Newcastle disease virus in chickens." <u>Avian Pathology</u> **15**(4): 731-740.
- Capua, I. and D. J. Alexander (2009). <u>Avian influenza and Newcastle disease:</u> <u>A field and laboratory manual</u>, Springer.
- Cattoli, G., A. Fusaro, I. Monne, S. Molia, A. Le Menach, B. Maregeya, A. Nchare, I. Bangana, A. G. Maina, J. N. Koffi, H. Thiam, O. E. Bezeid, A. Salviato, R. Nisi, C. Terregino and I. Capua (2010). "Emergence of a new genetic lineage of Newcastle disease virus in west and central Africa-implications for diagnosis and control." <u>Veterinary Microbiology</u> **142**(3-4): 168-176.
- Chambers, P., N. S. Millar, R. W. Bingham and P. T. Emmerson (1986). "Molecular cloning of complementary DNA to Newcastle disease virus, and nucleotide sequence analysis of the junction between the genes encoding the haemagglutinin-neuraminidase and the large protein." Journal of General Virology 67(3): 475-486.
- Cho, S. H., S. J. Kim and H. J. Kwon (2007). "Genomic sequence of an antigenic variant Newcastle disease virus isolated in Korea." <u>Virus Genes</u> 35(2): 293-302.
- Cho, S. H., H. J. Kwon, T. E. Kim, J. H. Kim, H. S. Yoo and S. J. Kim (2008a). "Variation of a Newcastle disease virus hemagglutinin-neuraminidase linear epitope." Journal of Clinical Microbiology **46**(4): 1541-1544.
- Cho, S. H., H. J. Kwon, T. E. Kim, J. H. Kim, H. S. Yoo, M. H. Park, Y. H. Park and S. J. Kim (2008b). "Characterization of a recombinant Newcastle disease virus vaccine strain." <u>Clinical and Vaccine Immunology</u> **15**(10): 1572-1579.
- Choi, K.-S., S.-J. Kye, J.-Y. Kim and H.-S. Lee (2013). "Genetic and antigenic variation of shedding viruses from vaccinated chickens after challenge with virulent Newcastle disease virus." Avian Diseases 57(2): 303-306.
- Chong, Y. L., A. Padhi, P. J. Hudson and M. Poss (2010). "The effect of vaccination on the evolution and population dynamics of avian paramyxovirus-1." PLoS Pathogen 6(4): e1000872.
- Chulan, U., A. L. Ibrahim, A. M. Babjee and A. R. Sheikh-Omar (1982). "Vaccination against Newcastle disease." <u>Tropical Animal Health and Production</u> **14**(3): 177-184.
- Collins, M. S., J. Bashiruddin and D. J. Alexander (1993). "Deduced amino acid sequences at the fusion protein cleavage site of Newcastle disease viruses showing variation in antigenicity and pathogenicity." <u>Archives of Virology</u> **128**(3): 363-370.
- Collins, M. S., I. Strong and D. J. Alexander (1996). "Pathogenicity and phylogenetic evaluation of the variant Newcastle disease viruses termed "pigeon PMV-1 viruses" based on the nucleotide sequence of the fusion protein gene." <u>Archives of Virology</u> **141**(3-4): 635-647.
- Cornax, I., P. J. Miller and C. L. Afonso (2012). "Characterization of live Lasota vaccine strain-induced protection in chickens upon early

- challenge with a virulent Newcastle disease virus of heterologous genotype." <u>Avian Diseases</u> **56**(3): 464-470.
- Cvelic-Cabrilo, V., H. Mazija, Z. Bidin and W. L. Ragland (1992). "Correlation of haemagglutination inhibition and enzyme-linked immunosorbent assays for antibodies to Newcastle disease virus." <u>Avian Pathology</u> **21**(3): 509-512.
- Czegledi, A., D. Ujvari, E. Somogyi, E. Wehmann, O. Werner and B. Lomniczi (2006). "Third genome size category of avian paramyxovirus serotype 1 (Newcastle disease virus) and evolutionary implications." Virus Research 120(1-2): 36-48.
- De Boer, G., D. Van Roozelaar, R. Moormann, S. Jeurissen, J. Van Den Wijngaard, F. Hilbink and G. Koch (1994). "Interaction between chicken anaemia virus and live Newcastle disease vaccine." <u>Avian Pathology</u> **23**(2): 263-275.
- de Jong, M.D., Simmons, C.P., Thanh, T.T., Hien, V.M., Smith, G.J., Chau, T.N., Hoang, D.M., Chau, N.V., Khanh, T.H., Dong, V.C., Qui, P.T., Cam, B.V., Ha do, Q., Guan, Y., Peiris, J.S., Chinh, N.T., Hien, T.T., Farrar, J., 2006. Fatal outcome of human influenza A (H5N1) is associated with high viralload and hypercytokinemia. National Medicine 12 (10), 1203–1207.
- de Leeuw, O. S., G. Koch, L. Hartog, N. Ravenshorst and B. P. Peeters (2005). "Virulence of Newcastle disease virus is determined by the cleavage site of the fusion protein and by both the stem region and globular head of the haemagglutinin-neuraminidase protein." <u>Journal of General Virology</u> **86**(Pt 6): 1759-1769.
- Degefa, T., L. Dadi, A. Yami, G. M. K and M. Nassir (2004). "Technical and economic evaluation of different methods of Newcastle disease vaccine administration." <u>Journal of Veterinary Medicine</u>. A, Physiology, Pathology, Clinical Medicine **51**(7-8): 365-369.
- Diel, D. G., L. H. da Silva, H. Liu, Z. Wang, P. J. Miller and C. L. Afonso (2012). "Genetic diversity of avian paramyxovirus type 1: Proposal for a unified nomenclature and classification system of Newcastle disease virus genotypes." <u>Infection, Genetics and Evolution</u> **12**(8): 1770-1779.
- Doyle, T. (1927). "A hitherto unrecorded disease of fowls due to a filter-passing virus." Journal of Comparative Pathology **40**: 144-169.
- Esaki, M., A. Godoy, S. C. Rosenberger, Y. Gardin, A. Yasuda and K. Moore Dorsey (2013). "Protection and antibody response caused by turkey herpesvirus vector Newcastle disease vaccine." <u>Avian Diseases</u> **57**(4): 750-755.
- Estevez, C., D. King, B. Seal and Q. Yu (2007). "Evaluation of Newcastle disease virus chimeras expressing the hemagglutinin-neuraminidase protein of velogenic strains in the context of a mesogenic recombinant virus backbone." <u>Virus Research</u> **129**(2): 182-190.

- Ezema, W., J. Okoye and J. Nwanta (2009). "Lasota vaccination may not protect against the lesions of velogenic Newcastle disease in chickens." Tropical Animal Health and Production **41**(4): 477-484.
- Farkas, T., M. Antal, L. Sami, P. German, S. Kecskemeti, G. Kardos, S. Belak and I. Kiss (2007). "Rapid and simultaneous detection of avian influenza and Newcastle disease viruses by duplex polymerase chain reaction assay." Zoonoses and Public Health 54(1): 38-43.
- Fauquet, C. M. and D. Fargette (2005). "International committee on taxonomy of viruses and the 3,142 unassigned species." <u>Virology Journal</u> **2**: 64.
- Gabal, M. A. and A. H. Azzam (1998). "Interaction of aflatoxin in the feed and immunization against selected infectious diseases in poultry. Effect on one-day-old layer chicks simultaneously vaccinated against Newcastle disease, infectious bronchitis and infectious bursal disease."

 <u>Avian Pathology</u> **27**(3): 290-295.
- Garten, W., W. Berk, Y. Nagai, R. Rott and H. D. Klenk (1980). "Mutational changes of the protease susceptibility of glycoprotein F of Newcastle disease virus: Effects on pathogenicity." <u>Journal of General Virology</u> **50**(1): 135-147.
- Gauld, L. W., R. P. Hanson, W. H. Thompson and S. K. Sinha (1974). "Observations on a natural cycle of La Crosse virus (California group) in Southwestern Wisconsin." <u>The American Journal of Tropical Medicine and Hygiene</u> **23**(5): 983-992.
- Gharaibeh, S. and K. Mahmoud (2013). "Decay of maternal antibodies in broiler chickens." <u>Poultry Science</u> **92**(9): 2333-2336.
- Glisson, J., P. Villegas, L. Dufour, L. Christensen and D. Page (1990). Characterization of VG/GA Newcastle disease virus as a vaccine candidate. Proceedings of the 25th National Meeting on Poultry Health and Condemnations, Maryland, Ocean City USA.
- Goldhaft, T. M. (1980). "Guest editorial: Historical note on the origin of the Lasota strain of Newcastle disease virus." <u>Avian Diseases</u> **24**(2): 297.
- Gordon, S. and P. R. Taylor (2005). "Monocyte and macrophage heterogeneity." Nature Reviews Immunology 5(12): 953-964.
- Gotoh, B., T. Ogasawara, T. Toyoda, N. Inocencio, M. Hamaguchi and Y. Nagai (1990). "An endoprotease homologous to the blood clotting factor X as a determinant of viral tropism in chick embryo." <u>The EMBO journal</u> **9**(12): 4189.
- Gotoh, B., Y. Ohnishi, N. M. Inocencio, E. Esaki, K. Nakayama, P. J. Barr, G. Thomas and Y. Nagai (1992). "Mammalian subtilisin-related proteinases in cleavage activation of the paramyxovirus fusion glycoprotein: Superiority furin/PACE to PC2 or PC1/PC3" Journal of Virology **66**(11): 6391-6397.
- Grimes, S. E. (2002). "A basic laboratory manual for the small-scale production and testing of I-2 Newcastle disease vaccine." <u>RAP publication</u> **136**.

- Gudnason, H., M. Dufva, D. D. Bang and A. Wolff (2007). "Comparison of multiple DNA dyes for real-time PCR: Effects of dye concentration and sequence composition on DNA amplification and melting temperature." Nucleic acids research 35(19): e127.
- Habibian, M., S. Ghazi, M. M. Moeini and A. Abdolmohammadi (2013). "Effects of dietary selenium and vitamin E on immune response and biological blood parameters of broilers reared under thermoneutral or heat stress conditions." International Journal of Biometeorology: 1-12.
- Hall, T. (2005). "Bioedit v 7.0. 5." <u>Ibis Therapeutics, a division of Isis</u> Pharmaceuticals, Carlsbad.
- Hamal, K., S. Burgess, I. Pevzner and G. Erf (2006). "Maternal antibody transfer from dams to their egg yolks, egg whites, and chicks in meat lines of chickens." <u>Poultry Science</u> **85**(8): 1364-1372.
- Hanson, R. P. (1974). "The reemergence of Newcastle disease." <u>Advances in Veterinary Science & Comparative Medicine</u> **18**(0): 213-229.
- Hanson, R. P. and C. A. Brandly (1955). "Identification of vaccine strains of Newcastle disease virus." <u>Science</u> **122**(3160): 156-157.
- Hanson, R. P., J. Spalatin and G. S. Jacobson (1973). "The viscerotropic pathotype of Newcastle disease virus." <u>Avian Diseases</u> **17**(2): 354-361.
- Harrison, L., C. Brown, C. Afonso, J. Zhang and L. Susta (2011). "Early occurrence of apoptosis in lymphoid tissues from chickens infected with strains of Newcastle disease virus of varying virulence." <u>Journal of Comparative Pathology</u> **145**(4): 327-335.
- Hassett, D. E. and J. L. Whitton (1996). "DNA immunization." <u>Trends in Microbiology</u> **4**(8): 307-312.
- He, F., J. He, H. Guo, M. Yang, Z. Chen and S. Yu (2012). "The cloning F gene of pigeon paramyxovirus type I (PPMV-1) Pl strain and the study on DNA vaccine." Journal of Animal and Veterinary Advances 11(22): 4210-4216.
- Heid, C. A., J. Stevens, K. J. Livak and P. M. Williams (1996). "Real time quantitative PCR." Genome Research 6(10): 986-994.
- Heller, E. D., D. B. Nathan and M. Perek (1977). "The transfer of Newcastle disease serum antibody from the laying hen to the egg and the chick."

 <u>Research in Veterinary Science</u> **22**(3): 376-379.
- Hilbink, F., M. Vertommen, J. Tibben and W. van't Veer (1982). "The fluorescent antibody technique in the diagnosis of a number of poultry diseases: Manufacture of conjugates and use (author's transl)]." <u>Tijdschrift voor diergeneeskunde</u> **107**(5): 167.
- Hines, N. L. and C. L. Miller (2012). "Avian paramyxovirus serotype-1: A review of disease distribution, clinical symptoms, and laboratory diagnostics." <u>Veterinary Medicine International</u> **2012**: 708216.
- Hironori, S., H. Seisuke, I. Nobuhiro, I. Yasutada and K. Masao (1987). "Nucleotide sequence of the hemagglutinin-neuraminidase gene of Newcastle disease virus avirulent strain D26: Evidence for a longer coding region with a carboxyl terminal extension as compared to virulent strains." Virus Research 8(3): 217-232.

- Hitchner, S. and E. Johnson (1948a). "A virus of low virulence for immunizing fowls against Newcastle disease; Avian pneumoencephalitis." <u>Veterinary Medicine</u> **43**(12): 525.
- Hitchner, S. B. (2004). "History of biological control of poultry diseases in the USA." <u>Avian Diseases</u> **48**(1): 1-8.
- Hitchner, S. B., C. Domermuth, H. Purchase and J. Williams (1975). <u>Isolation and identification of avian pathogens</u>, Arnold Printing Corporation: 271-276.
- Hitchner, S. B. and E. P. Johnson (1948b). "A virus of low virulence for immunizing fowls against Newcastle disease; Avian pneumoencephalitis." <u>Journal of Veterinary Medicine Series A</u> **43**(12): 525-530.
- Holmes, H. C. (1979). "Virus-neutralizing antibody in sera and secretions of the upper and lower respiratory tract of chickens inoculated with live and inactivated Newcastle disease virus." <u>Journal of Comparative Pathology</u> **89**(1): 21-29.
- Hu, S., H. Ma, Y. Wu, W. Liu, X. Wang, Y. Liu and X. Liu (2009). "A vaccine candidate of attenuated genotype VII Newcastle disease virus generated by reverse genetics." <u>Vaccine</u> **27**(6): 904-910.
- Hu, X., R. Ray and R. Compans (1992). "Functional interactions between the fusion protein and hemagglutinin-neuraminidase of human parainfluenza viruses." <u>Journal of Virology</u> **66**(3): 1528-1534.
- Hu, Z., J. Hu, S. Hu, X. Liu, X. Wang and J. Zhu (2012). "Strong innate immune response and cell death in chicken splenocytes infected with genotype VII d Newcastle disease virus." <u>Virology Journal</u> 9(1): 208.
- Ibrahim, A. L., U. Chulan and A. M. Babjee (1980). "The immune response of chickens vaccinated against Newcastle disease with live Newcastle disease V4 vaccine." <u>Australian Veterinary Journal</u> **56**(1): 29-33.
- Johnson, M., I. Zaretskaya, Y. Raytselis, Y. Merezhuk, S. McGinnis and T. L. Madden (2008). "Ncbi blast: A better web interface." Nucleic Acids Research 36(suppl 2): W5-W9.
- Josefsen, M. H., C. Löfström, T. Hansen, E. Reynisson and J. Hoorfar (2012). "Instrumentation and fluorescent chemistries used in quantitative polymerase chain reaction." Quantitative Real-Time PCR in Applied Microbiology: 27.
- Juul-Madsen, H. R., B. Viertlböeck, S. Härtle, A. L. Smith and T. W. Göbel (2014). Innate immune responses. <u>Avian Immunology (second edition)</u>. A. S. Karel, K. Bernd, B. K. Peter Kaiser. Boston, Academic Press: 121-147.
- Kapczynski, D. R., C. L. Afonso and P. J. Miller (2013). "Immune responses of poultry to Newcastle disease virus." <u>Developmental & Comparative Immunology</u> **41**(3): 447-453.
- Kapczynski, D. R. and D. J. King (2005). "Protection of chickens against overt clinical disease and determination of viral shedding following vaccination with commercially available Newcastle disease virus

- vaccines upon challenge with highly virulent virus from the California 2002 exotic Newcastle disease outbreak." <u>Vaccine</u> **23**(26): 3424-3433.
- Karaca, K., J. M. Sharma, B. J. Winslow, D. E. Junker, S. Reddy, M. Cochran and J. McMillen (1998). "Recombinant fowlpox viruses coexpressing chicken type I IFN and Newcastle disease virus HN and F genes: Influence of IFN on protective efficacy and humoral responses of chickens following in-ovo or post-hatch administration of recombinant viruses." <u>Vaccine</u> **16**(16): 1496-1503.
- Ke, G. M., S. W. Yu, C. H. Ho, P. Y. Chu, L. Y. Ke, K. H. Lin, Y. C. Tsai, H. J. Liu and M. Y. Lin (2010). "Characterization of newly emerging Newcastle disease viruses isolated during 2002-2008 in Taiwan." <u>Virus Research</u> **147**(2): 247-257.
- Kido, H., Y. Yokogoshi, K. Sakai, M. Tashiro, Y. Kishino, A. Fukutomi and N. Katunuma (1992). "Isolation and characterization of a novel trypsin-like protease found in rat bronchiolar epithelial Clara cells. A possible activator of the viral fusion glycoprotein." <u>Journal of Biological Chemistry</u> **267**(19): 13573-13579.
- Kim, L. M., C. L. Afonso and D. L. Suarez (2006). "Effect of probe-site mismatches on detection of virulent Newcastle disease viruses using a fusion-gene real-time reverse transcription polymerase chain reaction test." Journal of Veterinary Diagnostic Investigation 18(6): 519-528.
- Kim, L. M., D. J. King, P. E. Curry, D. L. Suarez, D. E. Swayne, D. E. Stallknecht, R. D. Slemons, J. C. Pedersen, D. A. Senne and K. Winker (2007a). "Phylogenetic diversity among low-virulence Newcastle disease viruses from waterfowl and shorebirds and comparison of genotype distributions to those of poultry-origin isolates." <u>Journal of Virology</u> 81(22): 12641-12653.
- Kim, L. M., D. J. King, D. L. Suarez, C. W. Wong and C. L. Afonso (2007b). "Characterization of class I Newcastle disease virus isolates from Hong Kong live bird markets and detection using real-time reverse transcription-PCR." Journal of Clinical Microbiology 45(4): 1310-1314.
- Kim, S.-H., S. Nayak, A. Paldurai, B. Nayak, A. Samuel, G. L. Aplogan, K. A. Awoume, R. J. Webby, M. F. Ducatez and P. L. Collins (2012).
 "Complete genome sequence of a novel Newcastle disease virus strain isolated from a chicken in West Africa." <u>Journal of Virology</u> 86(20): 11394-11395.
- Kolakofsky, D., T. Pelet, D. Garcin, S. Hausmann, J. Curran and L. Roux (1998). "Paramyxovirus RNA synthesis and the requirement for hexamer genome length: The rule of six revisited." <u>Journal of Virology</u> **72**(2): 891-899.
- Kommers, G. D., D. J. King, B. S. Seal, K. P. Carmichael and C. C. Brown (2002). "Pathogenesis of six pigeon-origin isolates of Newcastle disease virus for domestic chickens." <u>Veterinary Pathology</u> **39**(3): 353-362.
- Kouwenhoven, B. (1993). "Newcastle disease." <u>Virus infections of birds</u>: 341-361.

- Kraneveld, F. (1926). "A poultry disease in the Dutch East Indies." <u>Ned Indisch Bl Diergeneeskd</u> **38**: 448-450.
- Krishnamurthy, S. and S. K. Samal (1998). "Nucleotide sequences of the trailer, nucleocapsid protein gene and intergenic regions of Newcastle disease virus strain Beaudette C and completion of the entire genome sequence." <u>Journal of General Virology</u> **79 (Pt 10)**(10): 2419-2424.
- Kwon, H. J., S. H. Cho, Y. J. Ahn, S. H. Seo, K. S. Choi and S. J. Kim (2003). "Molecular epidemiology of Newcastle disease in Republic of Korea." <u>Veterinary Microbiology</u> **95**(1-2): 39-48.
- Lai, M. C. and A. Ibrahim (1987). Velogenic viscerotropic Newcastle disease virus. Newcastle Disease in Poultry: A New Food Pellet Vaccine. J. W. Copland. Australia, ACIAR: 33-34.
- Lam, K. M. (1996). "Newcastle disease virus-induced apoptosis in the peripheral blood mononuclear cells of chickens." <u>Journal of Comparative Pathology</u> **114**(1): 63-71.
- Lamb, R. A. and G. D. Parks (2006). Paramyxoviridae: The viruses and their replication. <u>Fields virology</u>. B. N. Field. USA, Lippincott Williams & Wilkins: 1449-1491.
- Lambrecht, B., M. Gonze, G. Meulemans and T. P. van den Berg (2004). "Assessment of the cell-mediated immune response in chickens by detection of chicken interferon-gamma in response to mitogen and recall Newcastle disease viral antigen stimulation." <u>Avian Pathology</u> 33(3): 343-350.
- Lessard, M., D. Hutchings and N. A. Cave (1997). "Cell-mediated and humoral immune responses in broiler chickens maintained on diets containing different levels of vitamin A." <u>Poultry Science</u> **76**(10): 1368-1378.
- Letellier, C., A. Burny and G. Meulemans (1991). "Construction of a pigeonpox virus recombinant: Expression of the Newcastle disease virus (NDV) fusion glycoprotein and protection of chickens against NDV challenge." Archives of Virology **118**(1-2): 43-56.
- Li, Y., M. S. Collins, G. C. Whitelam and D. J. Alexander (2002). "Rapid pathotyping of Newcastle disease virus using a single-chain FV displayed on phage against the c-terminal end of the F2 polypeptide." Archives of Virology **147**(10): 2025-2037.
- Liang, R., D. J. Cao, J. Q. Li, J. Chen, X. Guo, F. F. Zhuang and M. X. Duan (2002). "Newcastle disease outbreaks in western China were caused by the genotypes VII a and VIII." <u>Veterinary Microbiology</u> **87**(3): 193-203.
- Lindh, E., C. Ek-Kommonen, V. M. Vaananen, J. Alasaari, A. Vaheri, O. Vapalahti and A. Huovilainen (2012). "Molecular epidemiology of outbreak-associated and wild-waterfowl-derived Newcastle disease virus strains in Finland, including a novel class I genotype." <u>Journal of Clinical Microbiology</u> **50**(11): 3664-3673.
- Liu, H., Z. Wang, Y. Wu, C. Sun, D. Zheng, T. Xu and J. Li (2008). "Molecular characterization and phylogenetic analysis of new Newcastle disease

- virus isolates from the mainland of China." Research in Veterinary Science 85(3): 612-616.
- Liu, X. F., H. Q. Wan, X. X. Ni, Y. T. Wu and W. B. Liu (2003). "Pathotypical and genotypical characterization of strains of Newcastle disease virus isolated from outbreaks in chicken and goose flocks in some regions of China during 1985–2001." <u>Archives of Virology</u> **148**(7): 1387-1403.
- Lockaby, S. B., F. J. Hoerr, A. C. Ellis and M. S. Yu (1993). "Immunohistochemical detection of Newcastle disease virus in chickens." <u>Avian Diseases</u> **37**(2): 433-437.
- Lomniczi, B., E. Wehmann, J. Herczeg, A. Ballagi-Pordany, E. Kaleta, O. Werner, G. Meulemans, P. Jorgensen, A. Mante and A. Gielkens (1998). "Newcastle disease outbreaks in recent years in western Europe were caused by an old (VI) and a novel genotype (VII)."

 <u>Archives of Virology</u> 143(1): 49-64.
- Mahon, P. J., A. M. Mirza and R. M. Iorio (2011). "Role of the two sialic acid binding sites on the Newcastle disease virus hn protein in triggering the interaction with the F protein required for the promotion of fusion." <u>Journal of Virology</u> **85**(22): 12079-12082.
- Maizan, M., S. Sharifah, G. Ong, R. Fuad and M. Suriani (2001). Genetic and phylogenetic analysis of Newcastle disease virus isolated from recent outbreaks. Proceedings of the 2nd International congress/13th VAM congress and CVA-Australasia/Oceania regional symposium, Kuala Lumpur, Malaysia.
- Malik, Y. S., D. P. Patnayak and S. M. Goyal (2004). "Detection of three avian respiratory viruses by single-tube multiplex reverse transcription-polymerase chain reaction assay." <u>Journal of Veterinary Diagnostic Investigation</u> **16**(3): 244-248.
- Malkinson, M. and P. A. Small, Jr. (1977). "Local immunity against Newcastle disease virus in the newly hatched chicken's respiratory tract." <u>Infection and Immunity</u> **16**(2): 587-592.
- Marangon, S. and L. Busani (2007). "The use of vaccination in poultry production." Revue Scientifique et Technique-Office International des Epizooties 26(1): 265.
- Mase, M., K. Imai, Y. Sanada, N. Sanada, N. Yuasa, T. Imada, K. Tsukamoto and S. Yamaguchi (2002). "Phylogenetic analysis of Newcastle disease virus genotypes isolated in japan." <u>Journal of Clinical Microbiology</u> **40**(10): 3826-3830.
- Mayahi, M., F. Talazadeh and M. Sina (2013). "Immune response in broilers given combinations of live and killed Newcastle disease vaccine viruses." Online Journal of Veterinary Research 17(8): 426-431.
- Mayo, M. A. (2002). "A summary of taxonomic changes recently approved by ICTV." <u>Archives of Virology</u> **147**(8): 1655-1663.
- McFerran, J. and R. McCracken (1988). Newcastle disease. <u>Newcastle Disease</u>, Springer: 161-183.

- McGinnes, L. W., H. Pantua, J. Reitter and T. G. Morrison (2006). "Newcastle disease virus: Propagation, quantification, and storage." <u>Current Protocols in Microbiology</u> Chapter 15: Unit 15F 12.
- Meulemans, G. (1988). Control by vaccination. <u>Newcastle disease</u>, Springer: 318-332.
- Meulemans, G., M. Gonze, M. C. Carlier, P. Petit, A. Burny and L. Long (1986). "Protective effects of HN and F glycoprotein-specific monoclonal antibodies on experimental Newcastle disease." <u>Avian Pathology</u> **15**(4): 761-768.
- Meulemans, G., C. Letellier, M. Gonze, M. C. Carlier and A. Burny (1988).

 "Newcastle disease virus F glycoprotein expressed from a recombinant vaccinia virus vector protects chickens against live-virus challenge." Avian Pathology 17(4): 821-827.
- Miller, P. J., C. L. Afonso, J. El Attrache, K. M. Dorsey, S. C. Courtney, Z. Guo and D. R. Kapczynski (2013). "Effects of Newcastle disease virus vaccine antibodies on the shedding and transmission of challenge viruses." <u>Developmental & Comparative Immunology</u> **41**(4): 505-513.
- Miller, P. J., E. L. Decanini and C. L. Afonso (2010). "Newcastle disease: Evolution of genotypes and the related diagnostic challenges." Infection, Genetics and Evolution 10(1): 26-35.
- Miller, P. J., C. Estevez, Q. Yu, D. L. Suarez and D. J. King (2009). "Comparison of viral shedding following vaccination with inactivated and live Newcastle disease vaccines formulated with wild-type and recombinant viruses." Avian Diseases 53(1): 39-49.
- Miller, P. J., D. J. King, C. L. Afonso and D. L. Suarez (2007). "Antigenic differences among Newcastle disease virus strains of different genotypes used in vaccine formulation affect viral shedding after a virulent challenge." <u>Vaccine</u> 25(41): 7238-7246.
- Morgan, R. W., J. Gelb, Jr., C. S. Schreurs, D. Lutticken, J. K. Rosenberger and P. J. Sondermeijer (1992). "Protection of chickens from Newcastle and Marek's diseases with a recombinant herpesvirus of turkeys vaccine expressing the Newcastle disease virus fusion protein." <u>Avian Diseases</u> 36(4): 858-870.
- Munir, M., M. Cortey, M. Abbas, Z. U. Qureshi, F. Afzal, M. Z. Shabbir, M. T. Khan, S. Ahmed, S. Ahmad, C. Baule, K. Stahl, S. Zohari and M. Berg (2012). "Biological characterization and phylogenetic analysis of a novel genetic group of Newcastle disease virus isolated from outbreaks in commercial poultry and from backyard poultry flocks in Pakistan." <u>Infection, Genetics and Evolution</u> **12**(5): 1010-1019.
- Nagai, Y. and H.-D. Klenk (1977). "Activation of precursors to both glycoproteins of Newcastle disease virus by proteolytic cleavage." <u>Virology Journal</u> 77(1): 125-134.
- Nagy, E., P. J. Krell, G. C. Dulac and J. B. Derbyshire (1991). "Vaccination against Newcastle disease with a recombinant baculovirus

- hemagglutinin-neuraminidase subunit vaccine." <u>Avian Diseases</u> **35**(3): 585-590.
- Nasrin, M., M. Z. Khan, M. N. Siddiqi and M. A. Masum (2013). "Mobilization of immunoglobulin (Ig)-containing plasma cells in harderian gland, cecal tonsil and trachea of broilers vaccinated with Newcastle disease vaccine." <u>Tissue & Cell</u> **45**(3): 191-197.
- Nunes, J., A. Vasconcelos, M. Jorge, E. Guimaraes, T. Paixao, N. Martins and J. Resende (2002). "Comparative morphometric analysis of vaccinal virulence of some lentogenic strains of Newcastle disease virus in tracheas of SPF chickens." <u>Arquivo Brasileiro de Medicina Veterinária e Zootecnia</u> **54**(4): 335-339.
- Ogasawara, T., B. Gotoh, H. Suzuki, J. Asaka, K. Shimokata, R. Rott and Y. Nagai (1992). "Expression of factor X and its significance for the determination of paramyxovirus tropism in the chick embryo." <u>EMBO Journal</u> **11**(2): 467-472.
- OIE (2000). Manual of standard for diagnostic tests and vaccines, OIE Publication. Pp.
- OIE (2012). Newcastle disease. OIE Terrestrial Manual, OIE
- OIE.(2013)."Exceptional epidemiological events." from http://www.oie.int/wahis_2/public/wahid.php/Countryinformatio n/Countryreports.
- Palya, V., I. Kiss, T. Tatar-Kis, T. Mato, B. Felfoldi and Y. Gardin (2012). "Advancement in vaccination against newcastle disease: Recombinant HVT NDV provides high clinical protection and reduces challenge virus shedding with the absence of vaccine reactions." <u>Avian Diseases</u> 56(2): 282-287.
- Park, M. S., M. L. Shaw, J. Munoz-Jordan, J. F. Cros, T. Nakaya, N. Bouvier, P. Palese, A. Garcia-Sastre and C. F. Basler (2003). "Newcastle disease virus (NDV)-based assay demonstrates interferon-antagonist activity for the NDV V protein and the Nipah virus V, W, and C proteins." <u>Journal of Virology</u> 77(2): 1501-1511.
- Parry, S. and I. Aitken (1977). "Local immunity in the respiratory tract of the chicken. The secretory immune response to Newcastle disease virus and the role of IgA." <u>Veterinary Microbiology</u> **2**(2): 143-165.
- Peeters, B., P. Verbruggen, F. Nelissen and O. de Leeuw (2004). "The P gene of Newcastle disease virus does not encode an accessory X protein." <u>Journal of General Virology</u> **85**(8): 2375-2378.
- Peroulis-Kourtis, I., K. O'Riley, D. Grix, R. J. Condron and C. Ainsworth (2002). "Molecular characterisation of Victorian Newcastle disease virus isolates from 1976 to 1999." <u>Australian Veterinary Journal</u> **80**(7): 422-424.
- Perozo, F., R. Marcano and C. L. Afonso (2012). "Biological and phylogenetic characterization of a genotype VII Newcastle disease virus from Venezuela: Efficacy of field vaccination." <u>Journal of Clinical Microbiology</u> **50**(4): 1204-1208.

- Perozo, F., P. Villegas, R. Dolz, C. L. Afonso and L. B. Purvis (2008a). "The VG/GA strain of Newcastle disease virus: Mucosal immunity, protection against lethal challenge and molecular analysis." <u>Avian Pathology</u> **37**(3): 237-245.
- Perozo, F., P. Villegas, C. Estevez, I. R. Alvarado, L. B. Purvis and E. Saume (2008b). "Avian adeno-associated virus-based expression of Newcastle disease virus hemagglutinin-neuraminidase protein for poultry vaccination." Avian Diseases **52**(2): 253-259.
- Qin, Z.-M., L.-T. Tan, H.-Y. Xu, B.-C. Ma, Y.-L. Wang, X.-Y. Yuan and W.-J. Liu (2008). "Pathotypical characterization and molecular epidemiology of Newcastle disease virus isolates from different hosts in china from 1996 to 2005." <u>Journal of Clinical Microbiology</u> **46**(2): 601-611.
- Rahman, M., A. Bari, M. Giasuddin, M. Islam, J. Alam and G. Sil (2002). "Evaluation of maternal and humoral immunity against Newcastle disease virus in chicken." <u>International Journal of Poultry Sciences</u> 1(5): 161-163.
- Rasoli, M., S. K. Yeap, S. W. Tan, H. Moeini, A. Ideris, M. H. Bejo, N. B. Alitheen, P. Kaiser and A. R. Omar (2014). "Alteration in lymphocyte responses, cytokine and chemokine profiles in chickens infected with genotype VII and VIII velogenic Newcastle disease virus."

 <u>Comparative Immunology, Microbiology & Infectious Diseases</u> 37(1): 11-21.
- Rautenschlein, S., J. M. Sharma, B. J. Winslow, J. McMillen, D. Junker and M. Cochran (1999). "Embryo vaccination of turkeys against Newcastle disease infection with recombinant fowlpox virus constructs containing interferons as adjuvants." <u>Vaccine</u> **18**(5): 426-433.
- Reed, L. J. and H. Muench (1938). "A simple method of estimating fifty per cent endpoints." <u>American Journal of Epidemiology</u> **27**(3): 493-497.
- Rehmani, S. F. (1996). "Newcastle disease vaccination: A comparison of vaccines and routes of administration in pakistan." <u>Preventive Veterinary Medicine</u> **25**(3–4): 241-248.
- Reis-Filho, J. S. (2009). "Next-generation sequencing." <u>Breast cancer research</u>: <u>BCR</u> **11 Suppl 3**(Suppl 3): S12.
- Reynolds, D. L. and A. D. Maraqa (2000a). "Protective immunity against Newcastle disease: The role of antibodies specific to Newcastle disease virus polypeptides." <u>Avian Diseases</u> **44**(1): 138-144.
- Reynolds, D. L. and A. D. Maraqa (2000b). "Protective immunity against newcastle disease: The role of cell-mediated immunity." <u>Avian Diseases</u> **44**(1): 145-154.
- Romer-Oberdorfer, A., O. Werner, J. Veits, T. Mebatsion and T. C. Mettenleiter (2003). "Contribution of the length of the HN protein and the sequence of the F protein cleavage site to Newcastle disease virus pathogenicity." <u>Journal of General Virology</u> **84**(Pt 11): 3121-3129.
- Rosseel, T., B. Lambrecht, F. Vandenbussche, T. Van Den Berg and S. Van Borm (2011). "Identification and complete genome sequencing of paramyxoviruses in mallard ducks (Anas platyrhynchos) using

- random access amplification and next generation sequencing technologies." <u>Virology Journal</u> **8**(1): 463.
- Rue, C. A., L. Susta, I. Cornax, C. C. Brown, D. R. Kapczynski, D. L. Suarez, D. J. King, P. J. Miller and C. L. Afonso (2011). "Virulent Newcastle disease virus elicits a strong innate immune response in chickens." <u>Journal of General Virology</u> 92(4): 931-939.
- Russell, P. (1988). Monoclonal antibodies in research, diagnosis and epizootiology of Newcastle disease. <u>Newcastle disease</u>, Springer: 131-146.
- Russell, P. H. (1993). "Newcastle disease virus: Virus replication in the Harderian gland stimulates lacrimal IgA; the yolk sac provides early lacrimal IgG." <u>Veterinary Immunology and Immunopathology</u> **37**(2): 151-163.
- Russell, P. H., P. N. Dwivedi and T. F. Davison (1997). "The effects of cyclosporin A and cyclophosphamide on the populations of B and T cells and virus in the harderian gland of chickens vaccinated with the Hitchner B1 strain of Newcastle disease virus." Veterinary Immunology and Immunopathology 60(1-2): 171-185.
- Russell, P. H., P. C. Griffiths and M. J. Cannon (1983). "A microwell immunoperoxidase test for screening hybridomas and for diagnosing Newcastle disease virus and Sendai virus." <u>Journal of Immunological Methods</u> **61**(2): 165-170.
- Saif, Y. M. (1991). "Immunosuppression induced by infectious bursal disease virus." Veterinary Immunology and Immunopathology **30**(1): 45-50.
- Saitou, N. and M. Nei (1987). "The neighbor-joining method: A new method for reconstructing phylogenetic trees." Molecular Biology and Evolution 4(4): 406-425.
- Sakaguchi, T., T. Toyoda, B. Gotoh, N. M. Inocencio, K. Kuma, T. Miyata and Y. Nagai (1989). "Newcastle disease virus evolution. I. Multiple lineages defined by sequence variability of the hemagglutinin-neuraminidase gene." Virology Journal 169(2): 260-272.
- Sawant, P. M., P. C. Verma, P. K. Subudhi, U. Chaturvedi, M. Singh, R. Kumar and A. K. Tiwari (2011). "Immunomodulation of bivalent Newcastle disease DNA vaccine induced immune response by codelivery of chicken IFN-gamma and IL-4 genes." <u>Veterinary Immunology and Immunopathology</u> **144**(1-2): 36-44.
- Scheid, A. and P. W. Choppin (1973). "Isolation and purification of the envelope proteins of Newcastle disease virus." <u>Journal of Virology</u> **11**(2): 263-271.
- Seal, B. S., D. J. King and H. S. Sellers (2000). "The avian response to Newcastle disease virus." <u>Developmental & Comparative Immunology</u> **24**(2-3): 257-268.
- Sharma, J. M. (1999). "Introduction to poultry vaccines and immunity." <u>Journal of Advanced Veterinary Medicine</u> **41**: 481-494.
- SHI, X.-m., S. HE, M. WANG, G.-h. YANG, W.-w. ZENG, H.-y. CUI, G.-z. TONG and Y.-f. WANG (2009). "Evaluation of immune efficacy with

- gene modified DNA vaccines based on HN gene of Newcastle disease virus." Chinese Journal of Animal and Veterinary Sciences 10: 017.
- Sick, C., U. Schultz, U. Munster, J. Meier, B. Kaspers and P. Staeheli (1998). "Promoter structures and differential responses to viral and nonviral inducers of chicken type I interferon genes." <u>The Journal of Biological Chemistry</u> **273**(16): 9749-9754.
- Singh, K. V. and A. El-Zein (1978). "Viral proliferation patterns of a velogenic (VLT), a mesogenic (Komarov), and a lentogenic (F) strain of Newcastle disease virus." <u>Poultry Science</u> **57**(6): 1563-1566.
- Spradbrow, P. (1988). Geographical distribution. <u>Newcastle disease</u>, Springer: 247-255.
- Spradbrow, P., A. Ibrahim, A. Mustaffa-Babjee and S. Kim (1978). "Use of an avirulent Australian strain of Newcastle disease virus as a vaccine." <u>Avian Diseases</u>: 329-335.
- Spradbrow, P. B., M. MacKenzie and S. E. Grimes (1995). "Recent isolates of Newcastle disease virus in Australia." <u>Veterinary Microbiology</u> **46**(1-3): 21-28.
- Steward, M., I. B. Vipond, N. S. Millar and P. T. Emmerson (1993). "RNA editing in Newcastle disease virus." <u>Journal of General Virology</u> **74** (**Pt 12**)(12): 2539-2547.
- Sun, H. L., Y. F. Wang, G. Z. Tong, P. J. Zhang, D. Y. Miao, H. D. Zhi and M. Wang (2008). "Protection of chickens from Newcastle disease and infectious laryngotracheitis with a recombinant fowlpox virus coexpressing the F, HN genes of Newcastle disease virus and gb gene of infectious laryngotracheitis virus." <u>Avian Diseases</u> 52(1): 111-117.
- Sundaresan, N. R., K. A. Ahmed, V. K. Saxena, K. V. Sastry, M. Saxena, A. B. Pramod, M. Nath, K. B. Singh, T. J. Rasool, A. K. DevRoy and R. V. Singh (2005). "Differential expression of inducible nitric oxide synthase and cytokine mRNA in chicken lines divergent for cutaneous hypersensitivity response." <u>Veterinary Immunology and Immunopathology</u> **108**(3-4): 373-385.
- Tamura, K., D. Peterson, N. Peterson, G. Stecher, M. Nei and S. Kumar (2011). "Mega5: Molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods." <u>Molecular Biology and Evolution</u> **28**(10): 2731-2739.
- Tan, S. W., A. Ideris, A. R. Omar, K. Yusoff and M. Hair-Bejo (2009). "Detection and differentiation of velogenic and lentogenic Newcastle disease viruses using sybr green I real-time PCR with nucleocapsid gene-specific primers." <u>Journal of Virological Methods</u> **160**(1): 149-156.
- Tan, S. W., A. Ideris, A. R. Omar, K. Yusoff and M. Hair-Bejo (2010a). "Sequence and phylogenetic analysis of Newcastle disease virus genotypes isolated in Malaysia between 2004 and 2005." <u>Archives of Virology</u> **155**(1): 63-70.
- Tan, S. W., A. Ideris, A. R. Omar, K. Yusoff and M. Hair-Bejo (2010b). "Sequence and phylogenetic analysis of Newcastle disease virus

- genotypes isolated in Malaysia between 2004 and 2005." <u>Archives of Virology</u> **155**(1): 63-70.
- Tan, S. W., A. R. Omar, A. Ideris, K. Yusoff and W. Tan (2004). "Detection of Newcastle disease virus using a Sybr green I real time polymerase chain reaction." <u>Acta Virologica</u> **48**(1): 23-28.
- Terregino, C. and I. Capua (2009a). Clinical traits and pathology of Newcastle disease infection and guidelines for farm visit and differential diagnosis. <u>Avian Influenza and Newcastle Disease: A field and laboratory manual</u>: 113-122.
- Terregino, C. and I. Capua (2009b). Conventional diagnosis of Newcastle disease virus infection. <u>Avian Influenza and Newcastle disease: A field and laboratory manual</u>: 123-125.
- Thekisoe, M. M., P. A. Mbati and S. P. Bisschop (2004). "Different approaches to the vaccination of free ranging village chickens against Newcastle disease in Qwa-Qwa, South Africa." <u>Veterinary Microbiology</u> **101**(1): 23-30.
- Timms, L. and D. J. Alexander (1977). "Cell-mediated immune response of chickens to Newcastle disease vaccines." <u>Avian Pathology</u> **6**(1): 51-59.
- Umali, D. V., H. Ito, T. Suzuki, K. Shirota, H. Katoh and T. Ito (2013). "Molecular epidemiology of Newcastle disease virus isolates from vaccinated commercial poultry farms in non-epidemic areas of Japan." Virology Journal 10(1): 330.
- Umino, Y., T. Kohama, T. Sato and A. Sugiura (1990). "Protective effect of monoclonal antibodies to Newcastle disease virus in passive immunization." Journal of General Virology **71**(5): 1199-1203.
- van Boven, M., A. Bouma, T. H. Fabri, E. Katsma, L. Hartog and G. Koch (2008). "Herd immunity to Newcastle disease virus in poultry by vaccination." <u>Avian Pathology</u> **37**(1): 1-5.
- Volkova, V., R. Wills, S. Hubbard, D. Magee, J. Byrd and R. Bailey (2011).

 "Associations between vaccinations against protozoal and viral infections and Salmonella in broiler flocks." Epidemiology and Infection 139(2): 206-215.
- Wang, Z., H. Liu, J. Xu, J. Bao, D. Zheng, C. Sun, R. Wei, C. Song and J. Chen (2006). "Genotyping of Newcastle disease viruses isolated from 2002 to 2004 in China." <u>Annals of the New York Academy of Sciences</u> **1081**(1): 228-239.
- Wilkinson, M. (1991). "Purification of rna." <u>Essential molecular biology: a</u> practical approach 1: 69-87.
- Wise, M. G., H. S. Sellers, R. Alvarez and B. S. Seal (2004a). "Rna-dependent rna polymerase gene analysis of worldwide Newcastle disease virus isolates representing different virulence types and their phylogenetic relationship with other members of the paramyxoviridae." <u>Virus Research</u> **104**(1): 71-80.
- Wise, M. G., D. L. Suarez, B. S. Seal, J. C. Pedersen, D. A. Senne, D. J. King, D. R. Kapczynski and E. Spackman (2004b). "Development of a real-time reverse-transcription PCR for detection of Newcastle disease virus

- RNA in clinical samples." <u>Journal of Clinical Microbiology</u> **42**(1): 329-338.
- Xiao, S., A. Paldurai, B. Nayak, A. Samuel, E. E. Bharoto, T. Y. Prajitno, P. L. Collins and S. K. Samal (2012). "Complete genome sequences of Newcastle disease virus strains circulating in chicken populations of indonesia." <u>Journal of Virology</u> 86(10): 5969-5970.
- Xie, Z. X., X. F. Tang, J. B. Dong, J. B. Liu, Y. S. Pang, X. W. Deng and Z. Q. Xie (2006). "Sequencing and analysis of the genomes of three Newcastle disease viruses isolated from Guangxi." <u>Virologica Sinica</u> **21**(3): 261.
- Yan, Z., Y. Du, Q. Zhao, R. Fan, W. Guo, R. Ma, X. Wang and R. Zhu (2011).

 "Mucosal immune responses against live Newcastle disease vaccine in immunosuppressed chickens."

 Pakistan Veterinary Journal 31: 280-286.
- Yang, C. Y., H. K. Shieh, Y. L. Lin and P. C. Chang (1999). "Newcastle disease virus isolated from recent outbreaks in Taiwan phylogenetically related to viruses (genotype VII) from recent outbreaks in western Europe." <u>Avian Diseases</u> **43**(1): 125-130.
- Yi, J., C. Liu, B. Chen and S. Wu (2011). "Molecular characterization of a virulent genotype VII d strain of Newcastle disease virus from farmed chickens in Shanghai." <u>Avian Diseases</u> 55(2): 279-284.
- Yohannes, T., A. K. Sharma, S. D. Singh and T. K. Goswami (2012). "Immunopathological effects of experimental T-2 mycotoxocosis in broiler chicken co-infected with infectious bronchitis virus (IBV)." Veterinary Immunology and Immunopathology 146(3-4): 245-253.
- Yu, L., Z. Wang, Y. Jiang, L. Chang and J. Kwang (2001). "Characterization of newly emerging Newcastle disease virus isolates from the People's republic of China and Taiwan." <u>Journal of Clinical Microbiology</u> 39(10): 3512-3519.
- Zhang, Z. W., Q. H. Wang, J. L. Zhang, S. Li, X. L. Wang and S. W. Xu (2012). "Effects of oxidative stress on immunosuppression induced by selenium deficiency in chickens." <u>Biological Trace Element Research</u> **149**(3): 352-361.