Malaysian Journal of Medicine and Health Sciences Vol. 2(2) June 2006: 1-26

Invited Review

Cancer Immunotherapy: Current Progress and Applications

Pooi Pooi Leong & Heng Fong Seow

Immunology Unit, Department of Pathology, Faculty of Medicine and Health Sciences Universiti Putra Malaysia, 43400 UPM, Selangor, Malaysia

ABSTRACT

Cancer immunotherapy is a form of treatment protocol for cancer patients that has been studied intensively over the last two decades. The undesirable side effects during the course of conventional treatment has lead to the development of immunotherapy as an alternative treatment modality. This approach encompasses the use of three different strategies with various immunotherapeutic modalities including (i) cytokines and monoclonal antibodies; (ii) activation of antigen presentation cells (APC) by using antigen-specific peptides or sources of antigens such as tumour lysate; and finally (iii) adoptive transfer of ex vivo activated autologous cytotoxic T-cells. Due to specific-targeting by antigen-specific monoclonal antibodies, dendritic cells and activated CD8+ T-cells, immunotherapy can eliminate tumour cells efficiently but the normal tissues are unaffected. Despite years of investigation, the outcome of immunotherapy-based clinical trials are inconsistent with very low response rates from patients. Several mechanisms have been proposed to contribute to this failure including the presence of regulatory T-cells (Treg), immunomodulatory cytokines, and aberrant gene expression in tumour cells. This review summarises information from about 140 articles and review papers. In addition, it also provides an update on recent trends in combinational immunotherapy with conventional therapy and encouraging results have been obtained. Reevaluation of previous studies is necessary to fine-tune the design and approach of immunotherapy to ensure better treatment outcomes.

Keywords: Immunotherapy, cancer, combinational immunotherapy

INTRODUCTION

Immunotherapy is a form of clinical management procedure to enhance the defence mechanism of patients either specifically or non-specifically to fight against disease. Historically, the idea of immunotherapy was initiated a long time ago in the 1890s by Coley who attempted to use bacterial toxins to treat cancer patients.^[1] Tumour regression was observed in those patients who received the injection. This could have been due to the effect of cytokines upon induction of a non-specific stimulation of the immune response towards the foreign antigen, the bacterial toxin.^[1] Until today, Coley's idea of a general stimulation of the immune system by using bacterial agents has been applied to malignancy. For example, Bacillus Calmette Guerin (BCG) has been tested and used as an adjuvant together with either immunotherapy or chemotherapy to treat patients with superficial bladder cancers.^[2] In addition, the potency of using virus-derived substances and various chemicals have also been investigated.^[3, 4]

There are three different approaches of immunotherapy to enhance the cellular immune system for immunosurveillance (*Fig. 1*). Firstly, synthetic cytokines, interleukins or antibodies are used to enhance the immune system in general. For example, interleukin-2 (IL-2) and interferon γ and α have been used to treat systemic malignancy such as multiple myeloma^[5] and breast cancer.^[6] Secondly, the immune system can be activated specifically by optimising antigen presentation. Administration of biologic modifiers such as antigen-specific peptide or whole tumour cells are able to enhance endogenous antigen presentation of the epitope by professional antigen presenting (APC) cells to CD8⁺ T cells. The third approach is to restore the immune system by adoptive transfer of *ex vivo* activated effector cells. The effector cells will first be stimulated against the tumour antigens of tumour cells *in vitro* before being infused back into the patients. Examples of immunotherapeutic agents that have been used in various immunotherapeutic settings are shown in Table 1.

BENEFITS OF IMMUNOTHERAPY

Radiotherapy, chemotherapy and surgery are the standard protocols to treat or to ensure better quality of time for patients with neoplasms. Surgery helps to reduce tumour burden, radiotherapy eliminates residual tumour cells in the surgical field and chemotherapy eliminates micro-metastatic disease both systematically as well as in the surgical field.^[7] Despite their roles in tumour tissue eradication, these conventional protocols are associated with a number of side effects that may slowly deteriorate the normal physiological functions especially the immune defence mechanism of the patients. Besides eradication of tumour cells, both radiotherapy and chemotherapy can kill normal proliferating cells, including those in the bone marrow, hair follicles and lining of the gastrointestinal tract. During the course of conventional treatments, patients are vulnerable to infection as the immune system of the patient is weak due to toxicity.

To date, immunotherapy has moved from test tubes in the laboratory to the preclinical and clinical level.^[8] Table 2 shows the list of clinical trials that involve immunotherapy. Immunotherapy helps to overcome several obstacles faced by conventional treatments, including, lack of specificity and toxicity.^[9] Furthermore, immunotherapy has the ability to provide memory response to combat tumour reoccurrence.^[7] Immunotherapy may provide a better quality of time and reduce the psychological burden for the patients. As for those patients who fail in their conventional therapy, when death appears to be inevitable, immunotherapy may provide better treatment outcomes for them.

It is without doubt that surgery is a useful approach for cancer treatment. However, it has been shown that surgical procedures can lead to post-operative immunosuppression, which may facilitate dissemination of tumour cells and outgrowth of minimal residual disease or micro metastasis.^[10] Therefore, strategies for stimulation of anti-tumour immune responses are particularly crucial to prevent metastasis formation and reduce risk of relapse. In patients with esophageal cancer who underwent trasthoracic esophaectomy, *ex-vivo* activated lymphokine-activated killer (LAK) cells were transferred just after surgery to overcome the post-operative immunosuppression.^[11] According to Yamagchi and colleagues, LAK cells transfer can significantly increase the helper and cytotoxic T-cell populations in post-operative patients. In addition, the LAK cell treated group had a trend of reduction for post-





Malaysian Journal of Medicine and Health Sciences Vol. 2(2) June 2006

Immuntherapeutic agents	Reference
Dendritic cells	
- loaded with peptide	[98]
- loaded with tumour lysate	[99]
- pulse with necrotic tumour cells	[100]
- loaded with irradiated tumour cells	[101]
- pulsed with recombinant viral vector	[102]
- pulsed with fusion protein	[103]
- transfected with recombinant viral vector	[104]
Monoclonal antibody	
- antibody alone (anti-CCR4, anti-CD25, anti-CD20, etc)	[105]
- fused with bacterial toxin	[98,106]
- bispecific	[107]
- genetically engineered	[62]
- radioactive labeled	[17]
Whole cell vaccine	
- irradiated tumour cells	[108]
- irradiated tumour cells transduced with viral vector	[109]
- modified by virus	[110]
Come me dified	
outotoxia T colls (Supermetural T coll)	[54 111]
dendritic cells	[34,111]
	[03]
Bacterial gene therapy	
- recombinant idiotype Fab fragment	[112]
- recombinant viral vector	[87]
Immune cells	
- peripheral gamma delta T cells	[113]
- autologous cytokine induced killer (CIK)	[114]
- natural killer (NK)	[97]
- lymphokine activated killer (LAK)	[11]
- activated CD8 ⁺ or CD4 ⁺ or CD3 ⁺ lymphocyte	[115,116]
Peptide	[117, 118]
Non-specific immunostimulatory factor (IL-2)	[19]

Table 1. List of Immunotherapeutic agents used

operative remote infection such as pneumonia and surgical site infection when compared to the control group.^[11] In a study conducted on patients who underwent radical surgery for colorectal cancer (Duke' B and C), it was found that pre-operative interleukin-2 (IL-2) administration had greatly reduced post-operative immunodeficiency with improved prognosis.^[12] This would be due to early activation of the antineoplastic immune system to counteract the growth of minimal residue disease and prevent late disease progression.^[12]

Besides surgery, radiotherapy is also an important treatment modality for cancer patients. However, ionising radiation acts both as a carcinogen as well as a therapeutic agent.^[13] Besides the killing of cancer cells, it usually causes induction of intensive stress and DNA damage to the surrounding tissues that lead to production of pro-inflammatory cytokines including TGF-B,^[14] reactive oxygen species (ROS),^[13] epidermal growth factors and fibroblast growth factors.^[15] The series of these events may lead to persistence of inflammatory reaction in the microenvironment and later carcinogenesis. [13] Thus, targeted radiotherapy is essential to ensure the efficacy of treatment. Conjugation of tumourassociated antigens (TAAs)-specific monoclonal antibodies with radioisotopes helped to achieve a better response and prognosis.^[16] Radioimmunotherapy of prostate cancer using radiolabeled anti-prostate-specific membrane antigen monoclonal antibody (J591), showed excellent targeting of soft-tissue and bone metastasis with limited toxicity of manageable myelotoxicity.^[17] A phase I/II clinical trial using the novel ^[131]I (iodine)-labeled Hab18G/ CD147-specific monoclonal antibody Fab', fragment on patients with hepatocellular carcinoma has yielded an encouraging clinical outcome with improved survival rate of progression-free in the patients after either one or two cycles of treatment (p<0.0001).^[18]

CHALLENGES IN THE FIELD OF IMMUNOTHERAPY

The first mentioned immunotherapeutic approach using the non-specific immunostimulatory agents including IFN- γ , IFN- α and rIL-2 has been associated with systemic toxicity and autoimmune-like symptoms due to the over-activated immune system.^[19,20] Administration of IL-2 in patients with advanced melanoma has shown no evidence of improved survival or disease remission. ^[21] Toxicity complications caused by systemic administration of cytokines such as IL-2 and IFN- α have been documented in patients with metastatic renal cell carcinoma, ^[19] melanoma,^[21] low-grade lymphoid neoplasm,^[22] and prostate cancer.^[23]

Despite the use of imuunostimulatory cytokines, monoclonal antibodies (mAbs), which mainly target tumour associated antigens (TAAs), are also widely used in immunotherapy. Tumour cells that are 'tagged' by the TAAs specific mAbs may help to recruit and provoke host effector mechanisms to eradicate the targeted tumour cells.^[9] However, some mAbs fail to respond with expected outcomes such as, trastuzumab (anti-Her/2neu).^[24] The failure of antibody-based immunotherapy may mainly be due to low or loss of TAAs expression and alteration of the immunodominant epitope.

The evolution of the immune evasion during tumorigenesis further complicates the situation. Cancer immunoediting is a concept that focuses on the shaping of neoplasm and has been proposed by Dunn *et al.* in 2002.^[25] In this process, the immune system will first undergo vigorous tumour killing, then come to the process of immune sculpturing to allow the survival of low immunogenic tumour variants and finally lead to tumour escape mechanism.^[25] Down regulation or loss of TAA expression has been reported both in human neoplasm and animal models such as melanoma,^[26] colorectal carcinoma ^[27] and murine retinoblastoma.^[28] This phenomenon is often associated with repeated treatments ^[29] and the presence of *in vivo* immunoselection pressure for immunoresistant tumor variants.^[30, 31] According to Matsui and colleagues, aggressive immunotherapy with peptide-stimulated DUC18 T cells in tumours bearing BALB/c mice led to relapse of the disease with re-growth

	Table 2. List of recent clinical trials that are re	clated to applications of immuno	therapy	
Disease	Agent used	Response	Toxicity	Reference
Non-Hodgkin's lymphoma	Epratuzumb (anti-CD22 MAb) + rituximab (anti-CD20)	47% response rate	-nil-	[105]
Prostate cancer	Provenge (DC pulsed with PA2024, a recombinant PSA) + bevacizumab	Induction of immune response towards PA2024	-nil-	[102]
Advance melanoma	Peptide, tumour lysate or both peptide & tumour lysate pulsed dendritic cells	Tumour reduction (1/16), DTH response (10/16)	Autosensitisation dermatitis-like eruption	[66]
Recurrent malignant glioma	Autologous whole cell tumour + GM-CSF + anti-CD3 activated lymphocyte	DTH response towards tumour (17/18)	-nil-	[108]
Metastasis, asymptomatic hormone refractory prostate cancer (Phase III)	Sipuleucel-T (APC8015)	Survival advantage	-nil-	[119]
Breast cancer (Phase III)	Oxidised mannan-MUC1 [ISRCTN71711835]	No recurrence (0/16), anti- MUC-1(9/13), MUC-1 specific response (4/10)	-nil-	[120]
Hormone-naive prostate cancer (Phase I/II)	Irradiated, allogeneic, prostate cancer cells with GM-CSF gene transduced	Decrease prostate-specific antigen (16/21), infiltration of DC and macrophage, oligoclonal antibodies	-nil-	[109]
Metastatic renal cancer (Phase I)	Autologous DC pulsed with HLA-A2 specific peptides + low dose IL-2	Regression (6/20), MUC-1 specific T-cells response at peripheral	-nil-	[86]
Gliomas and melanoma (Pilot trial)	Irradiated tumour cells tranduced with recombinant retroviral vector with B7-2 and GM-CSF	No specific anti-tumour immunity	Inflammation response	[121]
Advance B-cell lymphoma (Phase I)	Recombinant Fab fragment of <i>E coli</i> + lipid-based adjuvant	Vaccine specific mAb (5/17) anti- Fab specific T-cell response (8/17)	Mild	[112]
Solid malignancy (Phase I/II)	HLA-A2402- restricted modified WT1 (Wilms' tumour gene) peptide in Montanide ISA51 adjuvant	Potential anti-tumour effect	Acceptable	[117]
Acute myeloid leukemia (Phase I/II)	Autologous dendritic-like leukemic cells (DLLC)	Anti-leukaemic T-cells response	Autoimmunity, eczema (1/22)	[122]
Metastatic renal carcinoma	IL-2 + IFN- α	Only subset of patients responded	Fever, asthenia, nausea /emesis, skin disorder, hypotension, diarrhea	[19]

6

Malaysian Journal of Medicine and Health Sciences Vol. 2(2) June 2006

Malignant Mesothelioma (MM)	Autologous MM tumour lysate with recombinant GM-CSF	Anti-MM immune response (7/22), no major tumour regression	-nil-	[123]
Hormone refractory prostate cancer (HRPC) (Phase I)	DC loaded with HLA-A0201 restricted peptide derived from PSA, PSMA, surviving, prostein, trp-p8	Decrease serum PSA (1/8), antigen specific CD8 + activation against survivin, prostein and PSMA	Local skin reaction	[124]
Adenocarcinoma (Phase I)	Autologous DC pulse with mannan-MUC-1 fusion protein	Tumour stabilisation (2/10)	-nil-	[103]
Metastatic melanoma	Recombinant vaccinia virus with B7.1, ICAM-1 and LFA-3 (rV-TRICOM)	Anti-vaccinia antibody and T-cell response	Mild fatigue, myalgia, vitiligo	[87]
Cervical intraepithelial neoplasia	HPV 16 L1 VLP vaccine	High level protection against HPV16 infection for at least 3.5 years after immunisation	-nil-	[125]
T-cell large granular lymphocyte leukemia	Mikbeta1 (murine MAb against CD122)	Down-regulation of receptors from surface of leukemic cell.	-nil-	[126]
Advance non-small cell lung cancer (NSCLC) (Phase I)	Epidermal growth factor (EGF) – based vaccine	Good mAb response against EGF (15/43)	Fever, chills, nausea, vomiting, flushing	[127]
Stage II melanoma	DC pulsed with Melan/MART-lantigen	Increase antigen specific CD4 ⁺ and CD8 ⁺ T-cell response, long-live tumour antigen specific DTH reactivity	-ni-	[128]
Multiple melanoma	Idiotype-pulsed allogeneic DC (alloDC) + soluble protein Id conjugated with KLH (Id-KLH)	Anti-KLH antibody response, secretion of cytokines (IL-2 , IL-10, TNF- α , IL-6, IFN- γ)	-nil-	[129]
Multiple metastasis (Phase I)	DC pulsed with autologous necrotic tumour cells + OKT3 and IL-2 activated lymphocytes	DTH hypersensitivity reported, prolonged survival time	Low grade fever	[130]
Respectable stage III melanoma (randomised double-blind trial)	Newcastle disease virus-modified autologous melanoma cell lysate + IL-2	No remarkable differences between verum and placebo group	-nil-	[110]
Metastatic melanoma (Phase I/II)	Anti-CTLA-4 +IL-2	Objective tumour response (8/36), no synergistic effect observed	Autoimmune toxicity (grade II/IV),	[20]
Prostate cancer	PSA146 peptide (HLA-A2-specific PSA peptide)+GM-CSF	Positive DTH response, T-cell immunity developed	-nil-	[118]
<i>Note:</i> Due to limitations of space, or prostate-specific antigen; 'DTH', de 'E coli', <i>Escherichia coli</i> ; 'IFN-o', i tumour	Ily summarised data from articles that have been published layed-type hypersensitivity; 'GM-CSF', granulocyte macro interferon alpha; 'PSMA', prostate-specific membrane anti	in <i>Pubmed</i> in the year 2006 are summari: ophage colony stimulating factor; 'IL', in gen; ' HPV', Human papillomavirus; ' KJ	sed in the table. ['DC', dend tterleukin; 'HLA', human le LH', keyhole limpet hemocy	ritic cell; 'PSA', ucocyte antigen; /anin; ' TNF- α ',

of tumour cells and loss of TAAs expression. ^[32] Other tumour escape mechanisms are shown in *Fig. 2*.

The infiltration and priming efficiency of effector cells into the tumour micro-environment may not guarantee anti-tumour immunity.^[33, 34] The presence of large numbers of transgenic or *in-vitro* expanded tumour antigen-specific T cells do not consistently cause regression of tumour either in animals or patients.^[35, 36, 37] It is most likely that this phenomenon may be caused by the trafficking properties of the activated T cells as well as the tumour environment.^[38] Subpopulations of activated T cells may lose expression of CD62L and CCR7 that enable them to adhere to the peripheral basement membrane^[35] and trafficking toward the inflammatory chemokine such as CCL3 and CCL5.^[39, 40]

The use of ex-vivo cultured effector cells and autologous tumour cells, which is the third immunotherapeutic approach as mentioned earlier, is highly labour intensive. In addition, these cells are not always successfully grown in vitro.^[38] To generate effector cells for adoptive transfer, novel cell culture methodologies are needed to improve the frequency of specific T cells. Studies have shown that T cells isolated from different anatomical sites including peripheral blood, solid tumour, liquid tumour, and tumour involved or tumour free lymph nodes have different growth signals.^[41] Tumour infiltrating lymphocytes (TILs) can grow more consistently and to high levels than those isolated from peripheral blood. ^[41, 42] These differences may be reflected by different maturation state, nature of antigenic stimulus, frequency of re-stimulation, growth factors and co-stimulation of the T cells at different sites including the expression of chemokine receptors.^[43] As for adoptive transfer with autologous tumour cells, large quantities of cells are needed. This attempt aims to immunise the patient with all the antigens that are endogenously expressed by their tumour.^[7] However, some patients may have small tumour size and this can limit the quantity of cells available for study. Thus, the procedure for expansion of cells will become too costly for the patients. ^[7] In cases where the tumour lesion is not accessible, this approach will not be possible.

The presence of immunosuppressive cytokines such as, TGF-β and IL-10 within the tumour (as shown in Fig. 2) may disturb the maturation of dendritic cells as well as effector T cells generation and function.^[44] For example, injection of either apoptotic or non-apoptotic tumour cells may not necessarily cause dendritic cell maturation but may provoke tolerance of tumour immunity.^[45, 46, 47] There are many other cells types, which may have immunostimulatory effects, upon contact with apoptotic cells. Other than dendritic cells, macrophages, which are more commonly present at the tumour site have the ability to engulf apoptotic cells to cause secretion of a wide range of anti-inflammatory cytokines, including interleukin (IL)-10 and TGF-B.^[48, 49, 50] In contrast, macrophages also respond to non-apoptotic cells but to a lesser extent to produce pro-inflammatory cytokines such as TNF- α and IL-1 β .^[48, 49, 50] Occasionally, apoptotic cells that have not been efficiently phagocytosed may lead to secondary necrosis, which may cause autoimmunity.^[51] A study showed that mice bearing mutations resulting in defective phagocytosis of apoptotic cells have a greater tendency to develop autoimmunity than the null type mice.^[52] Therefore, strategies to block phagocyte function or the inhibitory effect of cytokines produced during phagocytosis have been carried out. These apoptotic cells are found to be significantly more immunogenic to the dendritic cells.^[53]



Figure 2. Proposed tumour escape mechanisms: (1) Local Dendritic cells (DCs) may be immature and unable to up-take, process, or present antigens. These DCs may be inhibited from migrating to regional lymph nodes or may actually induce tolerance, especially when presenting self-antigens. (2) Regulatory T cells which infiltrate the tumor site are able to mediate suppression of antigen-primed T cells. (3) The presence of helper CD4 T cell may skew the immune response toward a Th2 phenotype by secretion of Th2 cytokines such as IL-6, TGF- β and IL-10. This phenomenon inhibits the initiation of Th1 T cells and effective cellular immunity. (4) Tumor cells and the surrounding stromal may release a number of suppressive cytokines such as IL-6, IL-10, and TGF-â. This creates an environment that is not conducive to local immunity, which allows tumor cells to escape. (5) Tumor cells may express aberrant MHC class I molecules or â2-microglobulins, resulting in inadequate antigen presentation that prevent recognition of tumors by effector T cells. (6) Mutations may cause aberrant gene expression in components of signaling pathways as well as tumour antigen in tumour cells, resulting in abnormal cell behaviours and inadequate antigen presentation to activate the effector T cells. (Source: Adapted from www.innovitaresearch.org/news/04051401.html)

STRATEGIES IN IMMUNOTHERAPY

Despite expression of a wide range of chemokines, cytokines with various immunomodulatory effects have also being found. One possible explanation is that counteractive cytokines such as TNF- α , IL-10, TGF- β are expressed simultaneously at the tumour site. Thus, activated effector cells are poorly attracted to the tumour site. To overcome this problem, steps have been taken to block the inhibitory effect of the cytokine and to genetically modify T cells to express a chemokine receptor for adoptive transfer has significantly enhanced the T cell trafficking to the tumour and anti-tumour efficiency. ^[54] In addition, modification of tumour cells to express CCL3 and CCL20 has generated effective anti-tumour immune response. ^[44, 55, 56, 57] Therefore, a strategy to generate T cells with appropriate trafficking properties as well as modification of the tumour cells may help to enhance the efficiency of immunotherapy.

The efficiency of immunotherapy depends greatly on the recognition of antigen bearing tumour cells by the antigen-specific effector population. Non-specific approaches to stimulate the immune system have been carried out by using either autologous or allogenic tumour cells expressing shared antigens, purified defined tumour antigens and epitopes have shown some promising results in small trials. ^[58, 59, 60] Non-specific immunostimulatory agents, including cytokines (IFN- γ and TNF- α), bacterial toxin (BCG) ^[3,4] and immunostimulatory mAbs ^[9] have been used to up-regulate the immunity of the patients. High-dose IFN- γ has been used to treat patients with high-risk melanoma. ^[61] The tumoricidal ability of genetically modified tumour cells and cytotoxic T-cells has been tested out recently in several clinical trials. ^[62, 63] Randomised clinical trials of autologous GM-CSF –producing prostate cancer cells, APC8015 (Provenge) has demonstrated improved survival in patients with hormone-refractory prostate cancer. ^[64]

In addition, several strategies have been formulated to overcome the toxic effect of cytokine-based immunotherapy. O'Brien and colleagues have co-administered the rIL-2 with tauolidine in stage IV melanoma patients to reduce the toxicity without diminishing the therapeutic effect. ^[65] In addition, ReGel, an aqueous-based polymer has been used as an IL-2 delivery tool for cancer therapy. ^[66] The outcome of the study is very encouraging with sustained IL-2 tumoricidal effect and decreased IL-2 toxicity. ^[66] In addition, selection of alternative cytokines that have similar anti-tumour effect but lesser toxicity has been tested. Oniki and colleagues have recently used a combination of two IL-12-related cytokines, IL-23 and IL-27 to treat mice with poorly immunogenic B16F10 melanoma. Their study showed that systemic administration of these cytokines induced a protective anti-tumour activity which involved both the CD8⁺ T-cell and natural killer (NK) cells with minimal toxicity as compared to IL-12 administration. ^[67] Furthermore, a novel cytokine molecule, which combines desired cytokine activities to render toxic effect, has been developed by Acres & colleagues. ^[68] An IL-2/IL-18 recombinant molecule which has a novel lymphocyte-stimulating activity with reduced IL-2 associated toxicities in murine models has been tested. ^[68]

Lastly, aberrant expression of tumour-associated antigen in tumour cells has directed the development of mAbs against cell-surface receptors of the immune system. ^[9] This strategy helps to overcome the problem of loss or low antigen expression in tumour cells and to boost or block the effects of immunosuppressive cytokines present in the tumour environment. ^[69] These immunostimulating antibodies are designed against a number of molecules controlling the positive and negative signals in T cell activation. ^[70,71] In animal models, antibodies against CD40, ^[72] 4-1BB, ^[73] CTLA-4 ^[72,74] and CD25 ^[75] have been shown to provoke powerful tumour specific CTL responses capable of eradicating established tumour mass.

CURRENT TRENDS IN IMMUNOTHERAPY

New combinational immunotherapies are currently being tested to improve the efficacy of traditional treatment modalities. Combination of intratumoural dendritic cells with systemic cytotoxic therapies, such as chemotherapy and radiotherapy, have been reported in several studies. ^[76, 77, 78] In these models, the tumour cells were first killed by the chemotherapeutic drugs which was followed by the administration of dendritic cells to react with the apoptotic tumour cells to provoke anti-tumour immunity. ^[76, 77, 78] The combinational therapy was significantly more effective than either agent alone. ^[76, 77, 78, 79] Using combinational therapy, lower dose treatment regimens can be prescribed to the patients. ^[80] This innovative approach helps to avoid the higher medication level that causes undesirable side-effects when a single treatment modality is prescribed. [Please refer to Table 3 for the list of the clinical trials involve combinational immunotherapy]

The issues of tumour immunosuppression and lack of identified tumour-associated antigens (TAAs) are critical determining factors for successful immunotherapy.^[81] High amounts of immunoregulatory lymphocytes (Treg), including CD4+CD25+ and CD4+CD25+Foxp3+ subpopulations, IL-10 producing TR1 lymphocytes and tumour necrosis factor beta (TGF- β) producing TH3 lymphocytes have been detected at the tumour microenvironment which can inhibit the immune response and promote tumour outgrowth. ^[82] In addition, high numbers of monocytes, macrophages and neutrophils have been associated with very poor survival in patients with metastatic renal cell carcinoma.^[83] Several groups of researchers have developed therapeutic approaches based on the concept of combinational immunotherapy to tackle these problems. For example, in patients with Burkitt-type lymphoma (BL) or lymphoblastic leukemia (B-ALL), the use of hyperfractionated cyclophosphamide, vincristine, doxorubicin and dexamethasome (hyper-CVAD) together with rituximab (anti- CD20 monoclonal antibody) helped to achieve complete remission in 86% (24/28) of the evaluated patients. ^[84] Chemotherapeutic drugs not only eradicate tumour cell but they can also help to control Treg accumulation at the tumour microenvironment. [85] At the same time, histamine, which can help to protect NK and T cells against oxidative damage by inhibiting the formation and release of the phagocyte-derived reactive oxygen species, can also be used for immunotherapy.^[83] Co-administration of histamine and IL-2 help to increase intratumoural CD56 NK cells (p=0.008) and CD8⁺ T cells (p=0.019) with manageable toxicity compared to the baseline as well as treatment with IL-2 alone. [83] In an in vivo model of B16/BL6 melanoma, prescription of anti-CTLA-4 and a GM-CSF- transduced tumour cell vaccine (Gvax) helped to change the intratumour balance of Treg and effector T-cells to one which favors tumour rejection. [86] In this model, anti-CTLA-4 prevents release of inhibitory substances on T cell activation and proliferation, whereas,

	Table 3. List of recent clinical	trials that are related to app	dications of combinational imn	nunotherapy	
Types	Disease	Agent used	Response	Toxicity	Reference
Radio-immunotherapy (Phase I)	Metastatic CEA-producing malignancies	(90)Y labeled cT84.66 anti-CEA conjugate to DTPA	Human anti-chimeric antibody (8/13)	Haematologic toxicity	[131]
Radio-immunotherapy (Phase I/II)	Hepatocellular carcinoma	¹³¹ I(iodine) labeled HAb 18G/CD147- specific mAb Fab' fragment	Improved survival rate	-'n-	[18]
Radio-immunotherapy	Medullary thyroid carcinoma (MTC)	Anti-CEA/anti-DTPA- indium BsMAb + ¹³¹ I-labeled bivalent hapten	Longer overall survival	Haematologic toxicityto bone related / bone marrow metastasis	[132]
Chemo-immunotherapy	Burkitt-type lymphoma (BL) & acute lympho- blastic leukemia (B-ALL)	Rituximab + hyper- CVAD (hyper- fractionated cyclo- phosphamide, vincristine, doxorubicin, dexamethasone)	Improved overall survival rate in adult BL and B-ALL	-ii-	[84]
Radio- immunotherapy	Advance lung cancer necrosis therapy	Iodine- 131- labeled tumour chimeric antibody (¹³¹ I-chTNT)	Promising results	-ini-	[133]
Chemo-immunotheray	Metastatic renal cell carcinoma, melanoma	Thalidomide + rIL-2	Durable and active response	Mild to moderate (sommolence, constipation, neuropathy, rash, flu-like symptoms, fluid retention, hypotension, hypothyroidism	[134, 65]
Chemo-immunotherapy	Extensive stage small cell lung cancer	DC transfected with full-length wild- type p53 gene in adenoviral	p53 specific T-cell response 57.1%), anti-adenovirus mAb, high-rate objective	-nil-	[104]

Malaysian Journal of Medicine and Health Sciences Vol. 2(2) June 2006

		vector + chemotherapy	chemotherapy response (61.7%)		
Radio-immunotherapy	Recurrent primary and metastatic malignant brain tumour (Phase II)	lodine- 131-labeled murine antitenascin MAb 81C6 (¹³¹ I-m81C6)		Acute reversible/ irreversible neuro- toxicity, acute haematologic toxicity	[82]
External radiotherapy + Radio-immunotherapy	Non-small cell lung cancer (NSCLC)	¹¹¹ In-F(ab')2 -HMFG1	Residence time and adsorbed dose in tumour are low	-nil-	[135]
Radio-immunotherapy (Phase II)	Relapsed or refractory indolent non -Hodgkin's lymphoma (NHL)	iodine-131 (¹³¹ J) -rituximab chimeric anti-CD20 antibody	Overall response (76%) with 53% complete response	Neutropenia, thrombocyto- penia (Grade IV)	[136]
Chemotherapy + Radio- immunotherapy (Phase II)	Advanced follicular lymphoma	Cyclophosp- hamide, doxorubicin, vincristine, and prednisone (CHOP)+ tositumomab/iodine I-131 tositumomab	Overall response rate (91%) with 69% complete remission	-nil-	[137]
Radio-immunotherapy (Phase I)	CEA-expressing tumors [nonmedullary thyroid carcinoma (non-MTC), medullary thyroid carcinoma (MTC)]	anti-carcino-embryonic antigen (CEA) hMN-14 x m734 bispecific antibody (BsmAb) + (131)I-di- diethylenetriamine pentaacetic acid (DTPA)- indium hapten	Better response in MTC group than in non-MTC group patients)	Hematologic toxicity (higher in MTC patients than in non-MTC	[138]
Chemo-immunotherapy (Phase I)	Chronic lymphocytic leukemia	Pentostatin, cyclophos- phamide and rituximab response	Response rate (91%) with 41% complete	Modest haematologic toxicity	[139]
Chemo-immunotherapy (randomised clinical trial)	Progressive metastatic renal cell carcinoma	Interleukin-2/interferon- alpha2a/13-retinoic acid	No survival advantage	-nil-	[140]
Chemo-immunotherapy (Phase II)	Metastatic renal cell carcinoma	Vinorelbine and IL-2 (low dose)	Improved overall survival rate	-nil-	[141]
<i>Note:</i> Due to limitations of carcinoembryonic antigen; '	space, data summarised from article DTPA', Diethylenetriaminepentaace	s that have been published in <i>F</i> state; 'mAb', monoclonal antibu	^D ubmed in the year 2006 are summar ody]	ised in the table. ['CE/	ν,

Malaysian Journal of Medicine and Health Sciences Vol. 2(2) June 2006

Gvax primes the tumour-reactive effector T-cells which ultimately leads to T-cell activation and proliferation.^[86]

Another innovative approach to tackle the problem with immunodeficiency and immunosuppression of patients is to up-regulate the immune system by using compounds or fragments from microorganisms or recombinant bacterial or viral vectors.^[87] Local delivery of vaccinia virus expressing multiple co-stimulatory molecules including B7.1 (CD80), ICAM-1 and LFA-3 (rV-TRICOM) in patients with metastatic melanoma and breast cancer have been shown to enhance T-cell responses to TAA to a level far greater than any one or two applications of the co-stimulatory molecules in combination.^[87, 88] Recently, immunotoxinbased therapy has been evaluated in several clinical trials in both haema-tological malignancies and solid tumours.^[89] According to reports from several clinical trials, bacterial toxin such as Diphteria toxin (DT, 580 amino acid) or Pseudomonas aeruginosa exotoxin A (PE, 613 amino acid) that fused to the antibody Fv fragments has been shown to be a powerful antibody with selectivity to cell killing.^[89] Immunotoxins with anti-CD25 fused to PE or DT was able to eliminate the Tregs population in the microenvironment, hence, preventing immune tolerance.^[90,91] According to results from meta-analysis of randomised trials, intravesical BCG administration effectively reduced the recurrence rate after transurethral resection of patients with superficial bladder cancers.^[92]

Strategies have been made to encounter the technical problem of how to deliver molecules to the targeted destination. Dendritic cell transplantation has been widely used to overcome this problem. Autologous dendritic cells, which are extracted from the patients, are cultivated together with the tumour-specific antigen to be injected back into the patients. In order to further enhance the efficiency of this protocol, several groups have developed ordinary differential equation (ODE) systems by using mathematical models to predict the kinetics of the treatment, the influence of the treatment towards the normal physiologic function and the optimal schedule of injection of an immunotherapeutic agent against cancer.^[80, 93] These system characteristics are useful to gain a better understanding of the dynamic of a tumour-specific system which can ultimately help to guide the development of combinational therapies for elimination of the entire tumour.^[80]

In addition, the choices of clinical adjuvants used for delivery of the therapeutic agents to the target site are crucial. Due to the rapid degradation of the administrated immunodominant epitopes from tumour associated antigens (TAAs) *in vivo*, the efficacy and immunogenicity of the TAAs are low. Thus, this results in failure to mount an effective immune response to the target tumour cells.^[94] In addition, improper use of an adjuvant may create undesirable secondary reactions that may increase the risk beneficial ratio for the patients.^[95] The use of lipid- based adjuvants including liposome, virosome, ^[94] oil in water emulsion and bacterial immunostimulating compounds ^[95] have greatly enhanced the delivery as well as the therapeutic efficiency of the therapeutic agents including TAAs-specific peptides.^[94]

CONCLUSION

Immunotherapy of cancer has re-emerged vigorously in the 1990s. More than 50 immunotherapeutic regimens are currently under clinical testing and more than 400 clinical

trials have been conducted.^[95] Nevertheless, the clinical outcome and the response rate of the patients towards the testing agents are not sufficiently encouraging to prescribe it as a standard treatment regimen in a clinical setting. Through fundamental basic research in immunology, we will be able to figure out the 'bad guy' that is responsible for such a failure. Immune dominant tolerance by regulatory T-cells and immuno-suppressive role of Th2 cytokines are some examples of tumour escape mechanisms. Researchers have found numerous innovative approaches to overcome these barriers including the use of receptorspecific monoclonal antibodies to block the suppressive effect of Treg (Intrabodies) and the application of radio-labeled monoclonal antibodies to specifically guide the radioactive treatment to the target tumour cells. Recently, the role of innate immune response in tumour surveillance has been intensively studied. Clinical trials using natural killers against the solid tumour has been conducted.^[96] The inherent complexity of the immune system as a network of multiple interactions and redundant control loops amongst a huge diversity of components may either create another barrier or open other options for immunotherapy. Reevaluation of previous clinical trials has suggested that combinational immunotherapy with conventional cancer therapy have a better tumoricidal effect. A change in design and ultimate goals of clinical trials will be needed to ensure long term disease stabilisation and a better treatment outcome.

ACKNOWLEDGEMENT

We would like to acknowledge the financial support for this project from the Ministry of Science, Technology and Innovation (IRPA Project No. 06-02-04-0636-PR0054105-030).

REFERENCES

- Richardson MA, Ramirez T, Russell NC, Moye LA. Coley toxins immunotherapy: a retrospective review. Altern Ther Health Med 1999; 5(3): 42-47.
- [2] Morabito F, Rossi R, Graziano ME, Ferrando U, Lancini V, Cretarola E, Conti G, Luporini AC, Muto G, Castelli E, D'Urso L, Razionale P, Lissoni G, Simone M, Francesca F, Sommariva M, Casu M, Hurle R. Multicenter study on the use of gemcitabine to prevent recurrence of multiple-recurring superficial bladder tumors following intravesical antiblastic agents and/or BCG: evaluation of tolerance. Arch Ital Urol Androl 2006; 78(1): 1-4.
- [3] Schlag P, Manasterski M, Gerneth T, Hohenberger P, Dueck M, Herfarth C, Liebrich W, Schirrmacher V. Active specific immunotherapy with Newcastle-disease-virus-modified autologous tumor cells following resection of liver metastases in colorectal cancer. First evaluation of clinical response of a phase II-trial. Cancer Immunol Immunother 1992; 35(5): 325-330.
- [4] Dufour P, Lang JM, Giron C, Duclos B, Haehnel P, Jaeck D, Jung JM, Oberling F. Sodium dithiocarb as adjuvant immunotherapy for high risk breast cancer: a randomised study. Biotherapy 1993; 6(1): 9-12
- [5] Kaminska T, Dmoszynska A, Cioch M, Hus I, Jawniak D, Szuster-Ciesielska A, Kandefer-Szerszen M. Interferon gamma as immunomodulator in a patient with multiple myeloma. Arch Immunol Ther Exp (Warsz) 1999; 47(2): 107-112.

- [6] Kamamura Y, Takahashi K, Komaki K, Monden Y. Effects of interferon-alpha and gamma on development of LAK activity from mononuclear cells in breast cancer patients. J Med Invest 1998; 45(1-4): 71-75.
- [7] Yannelli JR, Wroblewski JM. On the road to a tumor cell vaccine: 20 years of cellular immunotherapy. Vaccine 2004; 15; 23(1): 97-113.
- [8] Bremers AJ, Parmiani G. Immunotherapy for colon cancer. Lancet 1999; 353(9163): 1524-1525.
- [9] Gray JC, Johnson PW, Glennie MJ. Therapeutic potential of immunostimulatory monoclonal antibodies. Clin Sci (Lond) 2006; 111(2): 93-106.
- [10] Oosterling SJ, van der Bij GJ, Mels AK, Beelen RH, Meijer S, van Egmond M, van Leeuwen PA. Perioperative IFN-alpha to avoid surgically induced immune suppression in colorectal cancer patients. Histol Histopathol 2006; 21(7): 753-760.
- [11] Yamaguchi Y, Hihara J, Hironaka K, Ohshita A, Okita R, Okawaki M, Matsuura K, Nagamine I, Ikeda T, Ohara M, Hamai Y. Postoperative immunosuppression cascade and immunotherapy using lymphokine-activated killer cells for patients with esophageal cancer: possible application for compensatory anti-inflammatory response syndrome. Oncol Rep 2006; 15(4): 895-901.
- [12] Brivio F, Fumagalli L, Lissoni P, Nardone A, Nespoli L, Fattori L, Denova M, Chiarelli M, Nespoli A. Pre-operative immunoprophylaxis with interleukin-2 may improve prognosis in radical surgery for colorectal cancer stage B-C. Anticancer Res 2006; 26(1B): 599-603.
- [13] Barcellos-Hoff MH, Park C, Wright EG. Radiation and the microenvironment tumorigenesis and therapy. Nat Rev Cancer 2005; 5(11): 867-875.
- [14] McBride WH, Chiang CS, Olson JL, Wang CC, Hong JH, Pajonk F, Dougherty GJ, Iwamoto KS, Pervan M, Liao YP. A sense of danger from radiation. Radiat Res 2004; 162(1): 1-19.
- [15] Dent P, Yacoub A, Fisher PB, Hagan MP, Grant S. MAPK pathways in radiation responses. Oncogene 2003; 22(37): 5885-5896.
- [16] Paganelli G, Bartolomei M, Grana C, Ferrari M, Rocca P, Chinol M. Radioimmunotherapy of brain tumor. Neurol Res 2006; 28(5): 518-522.
- [17] David KA, Milowsky MI, Kostakoglu L, Vallabhajosula S, Goldsmith SJ, Nanus DM, Bander NH. Clinical utility of radiolabeled monoclonal antibodies in prostate cancer. Clin Genitourin Cancer 2006; 4(4): 249-256.
- [18] Chen ZN, Mi L, Xu J, Song F, Zhang Q, Zhang Z, Xing JL, Bian HJ, Jiang JL, Wang XH, Shang P, Qian AR, Zhang SH, Li L, Li Y, Feng Q, Yu XL, Feng Y, Yang XM, Tian R, Wu ZB, Leng N, Mo TS, Kuang AR, Tan TZ, Li YC, Liang DR, Lu WS, Miao J, Xu GH, Zhang ZH, Nan KJ, Han J, Liu QG, Zhang HX, Zhu P. Targeting radioimmunotherapy of hepatocellular carcinoma with iodine (1311) metuximab injection: clinical phase I/II trials. Int J Radiat Oncol Biol Phys 2006; 65(2): 435-444.
- [19] Culine S, Iborra F, Mottet N, Avances C, de Graeve B, Volpe P, Vignoud J, Bringer JP, Marroncle M, Le Pellec L, Ayuso D, Jansen E, Faix A, Rebillard X. Subcutaneous interleukin-2 and interferon-alpha in metastatic renal cell carcinoma: results of a French regional experience in Languedoc. Am J Clin Oncol 2006; 29(2): 148-152.

- [20] Maker AV, Phan GQ, Attia P, Yang JC, Sherry RM, Topalian SL, Kammula US, Royal RE, Haworth LR, Levy C, Kleiner D, Mavroukakis SA, Yellin M, Rosenberg SA. Tumor regression and autoimmunity in patients treated with cytotoxic T lymphocyte-associated antigen 4 blockade and interleukin 2: a phase I/II study. Ann Surg Oncol 2005; 12(12): 1005-1016.
- [21] Tarhini AA, Agarwala SS. Cutaneous melanoma: available therapy for metastatic disease. Dermatol Ther 2006; 19(1): 19-25.
- [22] Waselenko JK, Burrows A, Nelson DA, Lucas M, Ekstrand J, Edenfield WJ, Myhand RC. Post-transplant interleukin-2 in patients with low-grade lymphoid neoplasms previously treated with fludarabine is limited by hematologic toxicity. Ann Hematol 2003; 82(9): 552-557.
- [23] Trudel S, Trachtenberg J, Toi A, Sweet J, Li ZH, Jewett M, Tshilias J, Zhuang LH, Hitt M, Wan Y, Gauldie J, Graham FL, Dancey J, Stewart AK. A phase I trial of adenovector-mediated delivery of interleukin-2 (AdIL-2) in high-risk localised prostate cancer. Cancer Gene Ther 2003; 10(10): 755-763.
- [24] Glennie MJ, Johnson PW. Clinical trials of antibody therapy. Immunol Today 2000; 21(8): 403-410.
- [25] Dunn GP, Bruce AT, Ikeda H, Old LJ, Schreiber RD. Cancer immunoediting: from immunosurveillance to tumor escape. Nat Immunol 2002; 3(11): 991-998.
- [26] Trefzer U, Hofmann M, Reinke S, Guo YJ, Audring H, Spagnoli G, Sterry W. Concordant loss of melanoma differentiation antigens in synchronous and asynchronous melanoma metastases: implications for immunotherapy. Melanoma Res 2006; 16(2): 137-145.
- [27] Kim JC, Roh SA, Lee KH, Namgung H, Kim JR, Kim JS. Genetic and pathologic changes associated with lymphovascular invasion of colorectal adenocarcinoma. Clin Exp Metastasis 2005; 22(5):421-428.
- [28] Dimaras H, Coburn B, Pajovic S, Gallie BL. Loss of p75 neurotrophin receptor expression accompanies malignant progression to human and murine retinoblastoma. Mol Carcinog 2006; 45(5): 333-343.
- [29] Jager E, Ringhoffer M, Karbach J, Arand M, Oesch F, Knuth A. Inverse relationship of melanocyte differentiation antigen expression in melanoma tissues and CD8+ cytotoxic-Tcell responses: evidence for immunoselection of antigen-loss variants in vivo. Int J Cancer 1996; 66(4): 470-476.
- [30] Sanchez-Perez L, Kottke T, Diaz RM, Ahmed A, Thompson J, Chong H, Melcher A, Holmen S, Daniels G, Vile RG. Potent selection of antigen loss variants of B16 melanoma following inflammatory killing of melanocytes *in vivo*. Cancer Res 2005; 65(5): 2009-2017.
- [31] Khong HT, Wang QJ, Rosenberg SA. Identification of multiple antigens recognised by tumorinfiltrating lymphocytes from a single patient: tumor escape by antigen loss and loss of MHC expression. J Immunother 2004; 27(3): 184-190.
- [32] Matsui K, O'Mara LA, Allen PM. Successful elimination of large established tumors and avoidance of antigen-loss variants by aggressive adoptive T cell immunotherapy. Int Immunol 2003; 15(7): 797-805.

- [33] Clark DA, Hirte HW, Buick RN. Human ovarian carcinoma: evidence for patient-related differences in susceptibility to cytotoxic effectors that attack different cellular subpopulations within a tumour. Br J Cancer 1988; 58(4): 415-418.
- [34] Limmer A, Sacher T, Alferink J, Kretschmar M, Schonrich G, Nichterlein T, Arnold B, Hammerling GJ. Failure to induce organ-specific autoimmunity by breaking of tolerance: importance of the microenvironment. Eur J Immunol 1998; 28(8): 2395-2406.
- [35] Hermans IF, Daish A, Yang J, Ritchie DS, Ronchese F. Antigen expressed on tumor cells fails to elicit an immune response, even in the presence of increased numbers of tumor-specific cytotoxic T lymphocyte precursors. Cancer Res 1998; 58(17):3909-3917.
- [36] Melief CJ, Kast WM. Cytotoxic T lymphocyte therapy of cancer and tumor escape mechanisms. Semin Cancer Biol 1991; 2(5): 347-354
- [37] Speiser DE, Miranda R, Zakarian A, Bachmann MF, McKall-Faienza K, Odermatt B, Hanahan D, Zinkernagel RM, Ohashi PS. Self antigens expressed by solid tumors do not efficiently stimulate naive or activated T cells: implications for immunotherapy. J Exp Med 1997; 186(5): 645-653.
- [38] Mirenda V, Millington O, Lechler RI, Scott D, Hernandez-Fuentes MP, Read J, Tan PH, George AJ, Garside P, Marelli-Berg FM. Tolerant T cells display impaired trafficking ability. Eur J Immunol 2005; 35(7): 2146-2156.
- [39] Tomiyama H, Matsuda T, Takiguchi M. Differentiation of human CD8⁺ T cells from a memory to memory/effector phenotype. J Immunol 2002; 168(11):5538-5550.
- [40] Nansen A, Marker O, Bartholdy C, Thomsen AR. CCR2+ and CCR5+ CD8+ T cells increase during viral infection and migrate to sites of infection. Eur J Immunol 2000; 30(7):1797-1806.
- [41] Maleckar JR, Friddell CS, Sferruzza A, Thurman GB, Lewko WM, West WH, Oldham RK, Yannelli JR. Activation and expansion of tumor-derived activated cells for therapeutic use. J Natl Cancer Inst 1989; 81(21):1655-1660.
- [42] Yannelli JR, Hyatt C, McConnell S, Hines K, Jacknin L, Parker L, Sanders M, Rosenberg SA. Growth of tumor-infiltrating lymphocytes from human solid cancers: summary of a 5-year experience. Int J Cancer 1996; 65(4): 413-421.
- [43] Geginat J, Sallusto F, Lanzavecchia A. Cytokine-driven proliferation and differentiation of human naive, central memory and effector memory CD4+ T cells. Pathol Biol (Paris) 2003; 51(2): 64-66.
- [44] Crittenden MR, Thanarajasingam U, Vile RG, Gough MJ. Intratumoral immunotherapy: using the tumour against itself. Immunology 2005; 114(1):11-22.
- [45] Steinman L. Despite epitope spreading in the pathogenesis of autoimmune disease, highly restricted approaches to immune therapy may still succeed [with a hedge on this bet] J Autoimmun 2000; 14(4): 278-282.
- [46] Melero I, Vile RG, Colombo MP. Feeding dendritic cells with tumor antigens: self-service buffet or a la carte? Gene Ther 2000; 7(14):1167-1170.

- [47] Sauter B, Albert ML, Francisco L, Larsson M, Somersan S, Bhardwaj N. Consequences of cell death: exposure to necrotic tumor cells, but not primary tissue cells or apoptotic cells, induces the maturation of immunostimulatory dendritic cells. J Exp Med 2000; 191(3): 423-434.
- [48] Fadok VA, McDonald PP, Bratton DL, Henson PM. Regulation of macrophage cytokine production by phagocytosis of apoptotic and post-apoptotic cells. Biochem Soc Trans 1998; 26(4): 653-656.
- [49] Reiter I, Krammer B, Schwamberger G. Cutting edge: differential effect of apoptotic versus necrotic tumor cells on macrophage antitumor activities. J Immunol 1999; 163(4):1730-1732.
- [50] Gough MJ, Melcher AA, Ahmed A, Crittenden MR, Riddle DS, Linardakis E, Ruchatz AN, Emiliusen LM, Vile RG. Macrophages orchestrate the immune response to tumor cell death. Cancer Res 2001; 61(19):7240-7247.
- [51] Wu L, Vandenabeele S, Georgopoulos K. Derivation of dendritic cells from myeloid and lymphoid precursors. Int Rev Immunol 2001; 20(1):117-135.
- [52] Cohen PL, Caricchio R, Abraham V, Camenisch TD, Jennette JC, Roubey RA, Earp HS, Matsushima G, Reap EA. Delayed apoptotic cell clearance and lupus-like autoimmunity in mice lacking the c-mer membrane tyrosine kinase. J Exp Med 2002; 196(1):135-140.
- [53] Ronchetti A, Rovere P, Iezzi G, Galati G, Heltai S, Protti MP, Garancini MP, Manfredi AA, Rugarli C, Bellone M. Immunogenicity of apoptotic cells in vivo: role of antigen load, antigenpresenting cells, and cytokines. J Immunol 1999; 163(1):130-136.
- [54] Kershaw MH, Wang G, Westwood JA, Pachynski RK, Tiffany HL, Marincola FM, Wang E, Young HA, Murphy PM, Hwu P. Redirecting migration of T cells to chemokine secreted from tumors by genetic modification with CXCR2. Hum Gene Ther 2002; 13(16):1971-1980.
- [55] Tolba KA, Bowers WJ, Muller J, Housekneckt V, Giuliano RE, Federoff HJ, Rosenblatt JD. Herpes simplex virus (HSV) amplicon-mediated codelivery of secondary lymphoid tissue chemokine and CD40L results in augmented antitumor activity. Cancer Res 2002; 62(22):6545-6551.
- [56] Dranoff G, Jaffee E, Lazenby A, Golumbek P, Levitsky H, Brose K, Jackson V, Hamada H, Pardoll D, Mulligan RC. Vaccination with irradiated tumor cells engineered to secrete murine granulocyte-macrophage colony-stimulating factor stimulates potent, specific, and long-lasting anti-tumor immunity. Proc Natl Acad Sci U S A 1993; 90(8):3539-3543.
- [57] Van Deventer HW, Serody JS, McKinnon KP, Clements C, Brickey WJ, Ting JP. Transfection of macrophage inflammatory protein 1 alpha into B16 F10 melanoma cells inhibits growth of pulmonary metastases but not subcutaneous tumors. J Immunol 2002; 169(3):1634-1639.
- [58] McIllmurray MB, Reeves WG, Langman MJ, Deane M, Embleton MJ. Active immunotherapy in malignant melanoma. Br Med J 1978; 1(6112):579.
- [59] Sondak VK, Liu PY, Tuthill RJ, Kempf RA, Unger JM, Sosman JA, Thompson JA, Weiss GR, Redman BG, Jakowatz JG, Noyes RD, Flaherty LE. Adjuvant immunotherapy of resected, intermediate-thickness, node-negative melanoma with an allogeneic tumor vaccine: overall results of a randomised trial of the Southwest Oncology Group. J Clin Oncol 2002; 20(8): 2058-2066.

- [60] Slingluff CL Jr, Yamshchikov G, Neese P, Galavotti H, Eastham S, Engelhard VH, Kittlesen D, Deacon D, Hibbitts S, Grosh WW, Petroni G, Cohen R, Wiernasz C, Patterson JW, Conway BP, Ross WG. Phase I trial of a melanoma vaccine with gp100 (280-288) peptide and tetanus helper peptide in adjuvant: immunologic and clinical outcomes. Clin Cancer Res 2001; 7(10): 3012-3024.
- [61] Cole BF, Gelber RD, Kirkwood JM, Goldhirsch A, Barylak E, Borden E. Quality-of-lifeadjusted survival analysis of interferon alfa-2b adjuvant treatment of high-risk resected cutaneous melanoma: an Eastern Cooperative Oncology Group study. J Clin Oncol 1996; 14(10): 2666-2673.
- [62] Lin AM, Hershberg RM, Small EJ. Immunotherapy for prostate cancer using prostatic acid phosphatase loaded antigen presenting cells. Urol Oncol 2006; 24(5): 434-441.
- [63] Sangro B, Melero I, Qian C, Prieto J. Gene therapy of cancer based on interleukin 12. Curr Gene Ther 2005; 5(6): 573-581.
- [64] Dawson NA. New molecular targets in advanced prostate cancer. Expert Rev Anticancer Ther 2006; 6(7): 993-1002.
- [65] O'Brien GC, Cahill RA, Bouchier-Hayes DJ, Redmond HP. Co-immunotherapy with interleukin-2 and taurolidine for progressive metastatic melanoma. Ir J Med Sci 2006; 175(1): 10-14.
- [66] Samlowski WE, McGregor JR, Jurek M, Baudys M, Zentner GM, Fowers KD. ReGel polymer-based delivery of interleukin-2 as a cancer treatment. J Immunother 2006; 29(5): 524-535.
- [67] Oniki S, Nagai H, Horikawa T, Furukawa J, Belladonna ML, Yoshimoto T, Hara I, Nishigori C. Interleukin-23 and interleukin-27 exert quite different antitumor and vaccine effects on poorly immunogenic melanoma. Cancer Res 2006; 66(12): 6395-6404.
- [68] Acres B, Gantzer M, Remy C, Futin N, Accart N, Chaloin O, Hoebeke J, Balloul JM, Paul S. Fusokine interleukin-2/interleukin-18, a novel potent innate and adaptive immune stimulator with decreased toxicity. Cancer Res 2005; 65(20): 9536-9546.
- [69] Zhu M, Wei MF, Liu F, Shi HF, Wang G. Interleukin-10 modified dendritic cells induce allohyporesponsiveness and prolong small intestine allograft survival. World J Gastroenterol 2003; 9(11): 2509-2512.
- [70] Rothstein DM, Sayegh MH. T-cell costimulatory pathways in allograft rejection and tolerance. Immunol Rev 2003; 196: 85-108.
- [71] Watts TH, DeBenedette MA. T cell co-stimulatory molecules other than CD28. Curr Opin Immunol 1999; 11(3): 286-293.
- [72] Fu F, Li W, Lu L, Thomson AW, Fung JJ, Qian S. Systemic administration of CTLA4-Ig or anti-CD40 ligand antibody inhibits second-set rejection of mouse liver allografts. Transplant Proc 1999; 31(1-2): 1244.
- [73] Melero I, Shuford WW, Newby SA, Aruffo A, Ledbetter JA, Hellstrom KE, Mittler RS, Chen L. Monoclonal antibodies against the 4-1BB T-cell activation molecule eradicate established tumors. Nat Med 1997; (6): 682-685.

- [74] Leach DR, Krummel MF, Allison JP. Enhancement of antitumor immunity by CTLA-4 blockade. Science 1996; 271(5256): 1734-1736.
- [75] Onizuka S, Tawara I, Shimizu J, Sakaguchi S, Fujita T, Nakayama E. Tumor rejection by in vivo administration of anti-CD25 (interleukin-2 receptor alpha) monoclonal antibody. Cancer Res 1999; 59(13): 3128-3133.
- [76] Tong Y, Song W, Crystal RG. Combined intratumoral injection of bone marrow-derived dendritic cells and systemic chemotherapy to treat pre-existing murine tumors. Cancer Res 2001; 61(20): 7530-7535.
- [77] Tanaka H, Shimizu K, Hayashi T, Shu S. Therapeutic immune response induced by electrofusion of dendritic and tumor cells. Cell Immunol 2002; 220(1): 1-12.
- [78] Shin JY, Lee SK, Kang CD, Chung JS, Lee EY, Seo SY, Lee SY, Baek SY, Kim BS, Kim JB, Yoon S. Antitumor effect of intratumoral administration of dendritic cell combination with vincristine chemotherapy in a murine fibrosarcoma model. Histol Histopathol 2003; 18(2): 435-447.
- [79] Wheeler CJ, Yu JS, Black KL. Cellular immunity in the treatment of brain tumors. Clin Neurosurg 2004; 51: 132-139.
- [80] De Pillis LG, Gu W, Radunskaya AE. Mixed immunotherapy and chemotherapy of tumors: modeling, applications and biological interpretations. J Theor Biol 2006; 238(4): 841-862.
- [81] Amato RJ. Vaccine therapy for renal cell carcinoma. Rev Urol. 2003; 5(2): 65-71.
- [82] Reardon DA, Akabani G, Coleman RE, Friedman AH, Friedman HS, Herndon JE 2nd, McLendon RE, Pegram CN, Provenzale JM, Quinn JA, Rich JN, Vredenburgh JJ, Desjardins A, Gururangan S, Badruddoja M, Dowell JM, Wong TZ, Zhao XG, Zalutsky MR, Bigner DD. Salvage radioimmunotherapy with murine iodine-131-labeled antitenascin monoclonal antibody 81C6 for patients with recurrent primary and metastatic malignant brain tumors: phase II study results. J Clin Oncol 2006; 24(1): 115-122.
- [83] Donskov F, Hokland M, Marcussen N, Torp Madsen HH, von der Maase H. Monocytes and neutrophils as 'bad guys' for the outcome of interleukin-2 with and without histamine in metastatic renal cell carcinoma—results from a randomised phase II trial. Br J Cancer 2006; 94(2): 218-226.
- [84] Thomas DA, Faderl S, O'Brien S, Bueso-Ramos C, Cortes J, Garcia-Manero G, Giles FJ, Verstovsek S, Wierda WG, Pierce SA, Shan J, Brandt M, Hagemeister FB, Keating MJ, Cabanillas F, Kantarjian H. Chemoimmunotherapy with hyper-CVAD plus rituximab for the treatment of adult Burkitt and Burkitt-type lymphoma or acute lymphoblastic leukemia. Cancer 2006; 106(7): 1569-1580.
- [85] Lopez M, Aguilera R, Perez C, Mendoza-Naranjo A, Pereda C, Ramirez M, Ferrada C, Aguillon JC, Salazar-Onfray F. The role of regulatory T lymphocytes in the induced immune response mediated by biological vaccines. Immunobiology 2006; 211(1-2): 127-136.
- [86] Quezada SA, Peggs KS, Curran MA, Allison JP. CTLA4 blockade and GM-CSF combination immunotherapy alters the intratumor balance of effector and regulatory T cells. J Clin Invest 2006; 116(7): 1935-1945.
- [87] Kaufman HL, Cohen S, Cheung K, DeRaffele G, Mitcham J, Moroziewicz D, Schlom J, Hesdorffer C. Local delivery of vaccinia virus expressing multiple costimulatory molecules for the treatment of established tumors. Hum Gene Ther 2006; 17(2): 239-244.

- [88] Garnett CT, Greiner JW, Tsang KY, Kudo-Saito C, Grosenbach DW, Chakraborty M, Gulley JL, Arlen PM, Schlom J, Hodge JW. TRICOM vector based cancer vaccines. Curr Pharm Des 2006; 12(3):351-361.
- [89] Pastan I, Hassan R, Fitzgerald DJ, Kreitman RJ. Immunotoxin therapy of cancer. Nat Rev Cancer 2006; 6(7):559-565.
- [90] Kreitman RJ, Wilson WH, Robbins D, Margulies I, Stetler-Stevenson M, Waldmann TA, Pastan I. Responses in refractory hairy cell leukemia to a recombinant immunotoxin. Blood. 1999; 94(10):3340-3348.
- [91] Foss FM, Bacha P, Osann KE, Demierre MF, Bell T, Kuzel T. Biological correlates of acute hypersensitivity events with DAB (389) IL-2 (denileukin diffitox, ONTAK) in cutaneous Tcell lymphoma: decreased frequency and severity with steroid premedication. Clin Lymphoma 2001; 1(4):298-302.
- [92] Han RF, Pan JG. Can intravesical bacillus Calmette-Guerin reduce recurrence in patients with superficial bladder cancer? A meta-analysis of randomized trials. Urology 2006; 67(6):1216-1223.
- [93] Castiglione F, Piccoli B. Optimal control in a model of dendritic cell transfection cancer immunotherapy. Bull Math Biol 2006; 68(2):255-274.
- [94] Adamina M, Schumacher R, Zajac P, Weber WP, Rosenthal R, Groeper C, Feder C, Zurbriggen R, Amacker M, Spagnoli GC, Oertli D, Heberer M. Advanced liposomal vectors as cancer vaccines in melanoma immunotherapy. J Liposome Res 2006; 16(3):195-204.
- [95] Aucouturier J, Ascarateil S, Dupuis L. The use of oil adjuvants in therapeutic vaccines. Vaccine 2006; 24 Suppl 2:S2-44-45.
- [96] Lage A, Perez R, Fernandez LE. Therapeutic cancer vaccines: at midway between immunology and pharmacology. Curr Cancer Drug Targets 2005; 5(8):611-627.
- [97] Boyiadzis M, Foon KA. Natural killer cells: from bench to cancer therapy. Expert Opin Biol Ther 2006; 6(10):967-970.
- [98] Wierecky J, Mueller M, Brossart P. Dendritic cell-based cancer immunotherapy targeting MUC-1. Cancer Immunol Immunother 2006; 55(1):63-67.
- [99] Nakai N, Asai J, Ueda E, Takenaka H, Katoh N, Kishimoto S. Vaccination of Japanese patients with advanced melanoma with peptide, tumor lysate or both peptide and tumor lysate-pulsed mature, monocyte-derived dendritic cells. J Dermatol 2006; 33(7):462-472.
- [100] Katano M. Development of therapeutic strategies based on immunology against tumors Fukuoka Igaku Zasshi 2006; 97(5):131-139.
- [101] Imura K, Ueda Y, Hayashi T, Itoh T, Shimizu K, Tamai H, Yano Y, Naito K, Kohara J, Nakane K, Matsuura Y, Takeda A, Takeda T, Kawai K, Yamagishi H. Induction of cytotoxic T lymphocytes against human cancer cell lines using dendritic cell-tumor cell hybrids generated by a newly developed electrofusion technique. Int J Oncol 2006; 29(3):531-539.
- [102] Rini BI, Weinberg V, Fong L, Conry S, Hershberg RM, Small EJ. Combination immunotherapy with prostatic acid phosphatase pulsed antigen-presenting cells (provenge) plus bevacizumab in patients with serologic progression of prostate cancer after definitive local therapy. Cancer 2006; 107(1):67-74.

- [103] Loveland BE, Zhao A, White S, Gan H, Hamilton K, Xing PX, Pietersz GA, Apostolopoulos V, Vaughan H, Karanikas V, Kyriakou P, McKenzie IF, Mitchell PL. Mannan-MUC1-pulsed dendritic cell immunotherapy: a phase I trial in patients with adenocarcinoma. Clin Cancer Res 2006; 12(3 Pt 1):869-877.
- [104] Antonia SJ, Mirza N, Fricke I, Chiappori A, Thompson P, Williams N, Bepler G, Simon G, Janssen W, Lee JH, Menander K, Chada S, Gabrilovich DI. Combination of p53 cancer vaccine with chemotherapy in patients with extensive stage small cell lung cancer. Clin Cancer Res 2006; 12(3 Pt 1):878-887.
- [105] Strauss SJ, Morschhauser F, Rech J, Repp R, Solal-Celigny P, Zinzani PL, Engert A, Coiffier B, Hoelzer DF, Wegener WA, Teoh NK, Goldenberg DM, Lister TA. Multicenter phase II trial of immunotherapy with the humanized anti-CD22 antibody, epratuzumab, in combination with rituximab, in refractory or recurrent non-Hodgkin's lymphoma. J Clin Oncol 2006; 24(24):3880-3886.
- [106] King GD, Curtin JF, Candolfi M, Kroeger K, Lowenstein PR, Castro MG. Gene therapy and targeted toxins for glioma. Curr Gene Ther 2005; 5(6):535-557.
- [107] Zhou J, Chen J, Zhong R, Mokotoff M, Shultz LD, Ball ED. Targeting gastrin-releasing peptide receptors on small cell lung cancer cells with a bispecific molecule that activates polyclonal T lymphocytes. Clin Cancer Res 2006; 12(7 Pt 1):2224-2231.
- [108] Sloan AE, Dansey R, Zamorano L, Barger G, Hamm C, Diaz F, Baynes R, Wood G. Adoptive immunotherapy in patients with recurrent malignant glioma: preliminary results of using autologous whole-tumor vaccine plus granulocyte-macrophage colony-stimulating factor and adoptive transfer of anti-CD3-activated lymphocytes. Neurosurg Focus 2000; 9(6):e9.
- [109] Simons JW, Sacks N. Granulocyte-macrophage colony-stimulating factor-transduced allogeneic cancer cellular immunotherapy: the GVAX vaccine for prostate cancer. Urol Oncol 2006; 24(5):419-424.
- [110] Voit C, Kron M, Schwurzer-Voit M, Sterry W. Intradermal injection of Newcastle disease virus-modified autologous melanoma cell lysate and interleukin-2 for adjuvant treatment of melanoma patients with resectable stage III disease. J Dtsch Dermatol Ges 2003; 1(2):120-125.
- [111] Cooper LJ, Ausubel L, Gutierrez M, Stephan S, Shakeley R, Olivares S, Serrano LM, Burton L, Jensen MC, Forman SJ, DiGiusto DL. Manufacturing of gene-modified cytotoxic T lymphocytes for autologous cellular therapy for lymphoma. Cytotherapy 2006; 8(2):105-117.
- [112] Bertinetti C, Zirlik K, Heining-Mikesch K, Ihorst G, Dierbach H, Waller CF, Veelken H. Phase I trial of a novel intradermal idiotype vaccine in patients with advanced B-cell lymphoma: specific immune responses despite profound immunosuppression. Cancer Res 2006; 66(8):4496-4502.
- [113] Viey E, Fromont G, Escudier B, Morel Y, Da Rocha S, Chouaib S, Caignard A. Phosphostimactivated gamma delta T cells kill autologous metastatic renal cell carcinoma. J Immunol 2005; 174(3):1338-1347.
- [114] Jiang J, Xu N, Wu C, Deng H, Lu M, Li M, Xu B, Wu J, Wang R, Xu J, Nilsson-Ehle P. Treatment of advanced gastric cancer by chemotherapy combined with autologous cytokineinduced killer cells. Anticancer Res 2006; 26(3B):2237-2242.

- [115] Levine BL, Bernstein WB, Aronson NE, Schlienger K, Cotte J, Perfetto S, Humphries MJ, Ratto-Kim S, Birx DL, Steffens C, Landay A, Carroll RG, June CH. Adoptive transfer of costimulated CD4+ T cells induces expansion of peripheral T cells and decreased CCR5 expression in HIV infection. Nat Med 2002 8(1): 47-53.
- [116] Thompson JA, Figlin RA, Sifri-Steele C, Berenson RJ, Frohlich MW. A phase I trial of CD3/ CD28-activated T cells (Xcellerated T cells) and interleukin-2 in patients with metastatic renal cell carcinoma. Clin Cancer Res 2003; 9(10 Pt 1):3562-3570.
- [117] Morita S, Oka Y, Tsuboi A, Kawakami M, Maruno M, Izumoto S, Osaki T, Taguchi T, Ueda T, Myoui A, Nishida S, Shirakata T, Ohno S, Oji Y, Aozasa K, Hatazawa J, Udaka K, Yoshikawa H, Yoshimine T, Noguchi S, Kawase I, Nakatsuka S, Sugiyama H, Sakamoto J. A phase I/II trial of a WT1 (Wilms' tumor gene) peptide vaccine in patients with solid malignancy: safety assessment based on the phase I data. Jpn J Clin Oncol 2006; 36(4): 231-236.
- [118] Perambakam S, Hallmeyer S, Reddy S, Mahmud N, Bressler L, DeChristopher P, Mahmud D, Nunez R, Sosman JA, Peace DJ. Induction of specific T cell immunity in patients with prostate cancer by vaccination with PSA146-154 peptide. Cancer Immunol Immunother 2006; 55(9):1033-1042.
- [119] Small EJ, Schellhammer PF, Higano CS, Redfern CH, Nemunaitis JJ, Valone FH, Verjee SS, Jones LA, Hershberg RM. Placebo-controlled phase III trial of immunologic therapy with sipuleucel-T (APC8015) in patients with metastatic, asymptomatic hormone refractory prostate cancer. J Clin Oncol 2006; 24(19):3089-3094.
- [120] Apostolopoulos V, Pietersz GA, Tsibanis A, Tsikkinis A, Drakaki H, Loveland BE, Piddlesden SJ, Plebanski M, Pouniotis DS, Alexis MN, McKenzie IF, Vassilaros S. Pilot phase III immunotherapy study in early-stage breast cancer patients using oxidized mannan-MUC1 [ISRCTN71711835]. Breast Cancer Res. 2006; 8(3):R27.
- [121] Parney IF, Chang LJ, Farr-Jones MA, Hao C, Smylie M, Petruk KC. Technical hurdles in a pilot clinical trial of combined B7-2 and GM-CSF immunogene therapy for glioblastomas and melanomas. J Neurooncol 2006; 78(1):71-80.
- [122] Roddie H, Klammer M, Thomas C, Thomson R, Atkinson A, Sproul A, Waterfall M, Samuel K, Yin J, Johnson P, Turner M. Phase I/II study of vaccination with dendritic-like leukaemia cells for the immunotherapy of acute myeloid leukaemia. Br J Haematol 2006; 133(2):152-157.
- [123] Powell A, Creaney J, Broomfield S, Van Bruggen I, Robinson B. Recombinant GM-CSF plus autologous tumor cells as a vaccine for patients with mesothelioma. Lung Cancer 2006; 52(2):189-197.
- [124] Fuessel S, Meye A, Schmitz M, Zastrow S, Linne C, Richter K, Lobel B, Hakenberg OW, Hoelig K, Rieber EP, Wirth MP. Vaccination of hormone-refractory prostate cancer patients with peptide cocktail-loaded dendritic cells: results of a phase I clinical trial. Prostate 2006; 66(8): 811-821.
- [125] Mao C, Koutsky LA, Ault KA, Wheeler CM, Brown DR, Wiley DJ, Alvarez FB, Bautista OM, Jansen KU, Barr E. Efficacy of human papillomavirus-16 vaccine to prevent cervical intraepithelial neoplasia: a randomized controlled trial. Obstet Gynecol 2006; 107(1):18-27.

- [126] Morris JC, Janik JE, White JD, Fleisher TA, Brown M, Tsudo M, Goldman CK, Bryant B, Petrus M, Top L, Lee CC, Gao W, Waldmann TA. Preclinical and phase I clinical trial of blockade of IL-15 using Mikbeta1 monoclonal antibody in T cell large granular lymphocyte leukemia. Proc Natl Acad Sci U S A 2006; 103(2): 401-406.
- [127] Ramos TC, Vinageras EN, Ferrer MC, Verdecia BG, Rupale IL, Perez LM, Marinello GG, Rodriguez RP, Davila AL. Treatment of NSCLC patients with an EGF-based cancer vaccine: report of a Phase I trial. Cancer Biol Ther 2006; 5(2):145-149.
- [128] Tuettenberg A, Becker C, Huter E, Knop J, Enk AH, Jonuleit H. Induction of strong and persistent MelanA/MART-1-specific immune responses by adjuvant dendritic cell-based vaccination of stage II melanoma patients. Int J Cancer 2006; 118(10):2617-2627.
- [129] Bendandi M, Rodriguez-Calvillo M, Inoges S, Lopez-Diaz de Cerio A, Perez-Simon JA, Rodriguez-Caballero A, Garcia-Montero A, Almeida J, Zabalegui N, Giraldo P, San Miguel J, Orfao A. Combined vaccination with idiotype-pulsed allogeneic dendritic cells and soluble protein idiotype for multiple myeloma patients relapsing after reduced-intensity conditioning allogeneic stem cell transplantation. Leuk Lymphoma 2006; 47(1): 29-37.
- [130] Katano M, Morisaki T, Koga K, Nakamura M, Onishi H, Matsumoto K, Tasaki A, Nakashima H, Akiyoshi T, Nakamura M. Combination therapy with tumor cell-pulsed dendritic cells and activated lymphocytes for patients with disseminated carcinomas. Anticancer Res 2005; 25(6A): 3771-3776.
- [131] Wong JY, Chu DZ, Williams LE, Liu A, Zhan J, Yamauchi DM, Wilczynski S, Wu AM, Yazaki PJ, Shively JE, Leong L, Raubitschek AA. A phase I trial of (90)Y-DOTA-anti-CEA chimeric T84.66 (cT84.66) radioimmunotherapy in patients with metastatic CEA-producing malignancies. Cancer Biother Radiopharm 2006; 21(2): 88-100
- [132] Chatal JF, Campion L, Kraeber-Bodere F, Bardet S, Vuillez JP, Charbonnel B, Rohmer V, Chang CH, Sharkey RM, Goldenberg DM, Barbet J; French Endocrine Tumor Group. Survival improvement in patients with medullary thyroid carcinoma who undergo pretargeted anticarcinoembryonic-antigen radioimmunotherapy: a collaborative study with the French Endocrine Tumor Group. J Clin Oncol 2006; 24(11): 1705-1711.
- [133] Yu L, Ju DW, Chen W, Li T, Xu Z, Jiang C, Chen S, Tao Q, Ye D, Hu P, Khawli LA, Taylor CR, Epstein AL. 131I-chTNT radioimmunotherapy of 43 patients with advanced lung cancer. Cancer Biother Radiopharm 2006; 21(1): 5-14.
- [134] Amato RJ, Morgan M, Rawat A. Phase I/II study of thalidomide in combination with interleukin-2 in patients with metastatic renal cell carcinoma. Cancer 2006; 106(7): 1498-506.
- [135] Garkavij M, Samarzija M, Ewers SB, Jakopovic M, Tezak S, Tennvall J. Concurrent radiotherapy and tumor targeting with 111In-HMFG1-F(ab')2 in patients with MUC1positive non-small cell lung cancer. Anticancer Res 2005; 25(6C): 4663-4671.
- [136] Leahy MF, Seymour JF, Hicks RJ, Turner JH. Multicenter phase II clinical study of iodine-131-rituximab radioimmunotherapy in relapsed or refractory indolent non-Hodgkin's lymphoma. J Clin Oncol 2006; 24(27): 4418-4425.

- [137] Press OW, Unger JM, Braziel RM, Maloney DG, Miller TP, Leblanc M, Fisher RI; Southwest Oncology Group. Phase II trial of CHOP chemotherapy followed by tositumomab/iodine I-131 tositumomab for previously untreated follicular non-Hodgkin's lymphoma: five-year follow-up of Southwest Oncology Group Protocol S9911. J Clin Oncol 2006; 24(25): 4143-4149.
- [138] Kraeber-Bodere F, Rousseau C, Bodet-Milin C, Ferrer L, Faivre-Chauvet A, Campion L, Vuillez JP, Devillers A, Chang CH, Goldenberg DM, Chatal JF, Barbet J. Targeting, Toxicity, and Efficacy of 2-Step, Pretargeted Radioimmunotherapy Using a Chimeric Bispecific Antibody and 131I-Labeled Bivalent Hapten in a Phase I Optimization Clinical Trial. J Nucl Med 2006; 47(2): 247-255.
- [139] Kay NE, Geyer SM, Call TG, Shanafelt TD, Zent CS, Jelinek DF, Tschumper R, Bone ND, Dewald GW, Lin TS, Heerema NA, Smith L, Grever MR, Byrd JC. Combination chemoimmunotherapy with pentostatin, cyclophosphamide and rituximab shows significant clinical activity with low accompanying toxicity in previously untreated B-chronic lymphocytic leukemia. Blood 2007; 109 (2): 405-411.
- [140] Atzpodien J, Kirchner H, Rebmann U, Soder M, Gertenbach U, Siebels M, Roigas J, Raschke R, Salm S, Schwindl B, Muller SC, Hauser S, Leiber C, Huland E, Heinzer H, Siemer S, Metzner B, Heynemann H, Fornara P, Reitz M. Interleukin-2/interferon-alpha2a/13-retinoic acid-based chemoimmunotherapy in advanced renal cell carcinoma: results of a prospectively randomised trial of the German Cooperative Renal Carcinoma Chemoimmunotherapy Group (DGCIN). Br J Cancer 2006; 95(4): 463-469.
- [141] Mencoboni MP, Tredici S, Varaldo M, Queirolo G, Durand F, Rebella L, Galbusera V, Pannacciulli IM, Ghio R. Chemoimmunotherapy with low dose vinorelbine and interleukin-2 in treatment of patients with metastatic renal cell carcinoma. Neoplasma 2006; 53(4): 333-336.