

UNIVERSITI PUTRA MALAYSIA

MOLECULAR CLONING AND FUNCTIONAL STUDIES OF SILICON-RESPONSIVE SERINE-RICH PROTEIN TRANSCRIPTS FROM MANGROVE PLANT, Rhizophora apiculata (Blume)

MAHBOD SAHEBI

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Ву

MAHBOD SAHEBI

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

February 2014

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DEDICATED TO

My Father Mansoor Sahebi; My Mother Mehri Sahebi; My beloved wife Parisa Azizi; My Brother and sister Abstract of thesis submitted to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for Doctor of Philosophy

MOLECULAR CLONING AND FUNCTIONAL STUDIES OF SILICON-RESPONSIVE SERINE-RICH PROTEIN TRANSCRIPTS FROM MANGROVE PLANT, Rhizophora apiculata (Blume)

By

MAHBOD SAHEBI

February 2014

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Institute : Tropical Agriculture

Silicon (Si) is one of the most plentiful elements found in the soil. Silicon plays an important role in decreasing susceptibility of plants against variety of different biotic and abiotic stresses. Mangrove plant (Rhizophora apiculata) is able to accumulate, and process Si to generate biosilica. Therefore, it would be a beneficial source for genetic manipulation of susceptible plants in the stress conditions. The objectives of the study were (i) to identify and characterize of a Si responsive gene in mangrove, (ii) to analyze the expression levels of a gene encoding serine-rich protein, and (iii) Functional studies of serine-rich protein in Arabidopsis thaliana. Three different methods and RNeasy plant mini kit were used to extract nucleic acids. The Suppression Subtractive Hybridization (SSH) technique was used to remove transcripts from proteins which were not involved in Si accumulation. Specific primer was designed to get full-length CDS of Semi-quantitative RT-PCR and real-time PCR were serine-rich protein. performed to examine its expression level under the control and treatment conditions. The Gateway Technology was used to construct entry and the expression vectors. Transformation of Arabidopsis thaliana with serine-rich protein gene was performed using Agrobacterium-mediated transformation by the floral-dip method. Energy-dispersive X-ray spectroscopy and high performance liquid chromatography were used to measure the quantity of Si and serine amino acid, respectively. Modified CTAB and SDS were quick and reliable methods for isolation of total RNA from the roots and leaves of mangrove, respectively. Of the sequences obtained from cDNA library, four were 97% similar to serine-rich protein gene of groundnut (Arachis hypogaea). Full-length of the serine-rich protein cDNA obtained through amplification of the cDNA template using specific primers. The

expression levels of *serine-rich protein* transcript were generally higher in the Si treated mangrove plants than untreated plants. The amount of serine amino acid of transgenic *Arabidopsis* has increased significantly from 1.02 mg g⁻¹ in wild-type plants to 37.76 mg g⁻¹. In addition, concentration of Si in the leaves and roots of transgenic plant was significantly higher than that in the wild type (P<0.01). This study successfully determined the Si responsive transcript related to *serine-rich protein* in mangrove plant (*R. apiculata*).



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

PENGKLONAN MOLEKUL DAN PENCIRIAN FUNGSI TRANSKRIP PROTEIN SILIKON-RESPONSIF SERINE-KAYA DARI PADA TUMBUHAN BAKAU, Rhizophora apiculata (Blume)

Oleh

MAHBOD SAHEBI

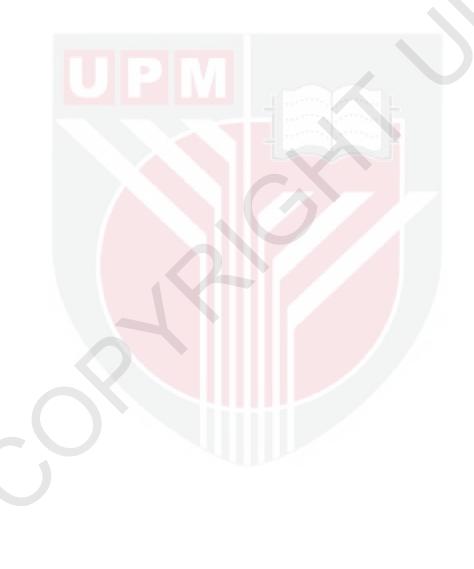
Februari 2014

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Silikon (Si) adalah salah satu elemen yang paling banyak didapati dalam tanah. Silikon memainkan peranan yang penting dalam mengurangkan kerentanan tumbuhan terhadap pelbagai tekanan biotik dan abiotik. Tumbuhan bakau (Rhizophora apiculata) mampu mengumpul dan memproses Si untuk menjana biosilika. Oleh itu, ia berupaya menjadi sumber bermanfaat untuk memanipulasikan genetik tumbuhan yang terdedah kepada tekanan persekitaran. Objektif kajian ini adalah (i) untuk mengenalpasti dan mencirikan gen Si-responsif dalam tumbuhan bakau, (ii) untuk mengesan tahap pengekespresan gen yang mengekod protein serine-kaya, dan (iii) kajian fungsi protein serine-kaya dalam Arabidopsis thaliana. Tiga kaedah yang berbeza dan RNeasy plant mini kit telah digunakan untuk mengeluarkan asid nukleik. Teknik Suppression Subtractive Hybridization (SSH) telah digunakan untuk mengasingkan protein transkrip yang tidak terlibat dalam pengumpulan Si. Buku asas khusus yang telah direka untuk mendapatkan CDS penuh panjang protein serine yang kaya. Separuh-kuantitatif RT-PCR dan tepat masa PCR telah dijalankan untuk mengkaji tahap ungkapan di bawah kawalan dan rawatan syarat-syarat. Gateway teknologi telah digunakan untuk fungsi pembinaan vektor. Transformasi Arabidopsis thaliana dengan protein serinekaya gen telah dilakukan melalui Agrobacterium dengan keadah pencelupan bunga. Tenaga-serakan X-ray spektroskopi dan kromatografi cecair prestasi tinggi telah digunakan untuk mengukur kualiti Si dan asid amino serine. CTAB dan SDS yang diubahsuai merupakan dua kaedah yang cepat dan sesuai untuk pengeluaran asid nukleik RNA daripada akar dan daun bakau. Empat urutan yang diperolehi daripada jumlah perpustakaan cDNA menunjukkan 97% persamaan dengan jujukan penuh protein serine-kaya gen dari kacang tanah (Arachis hypogaea). Jujukan penuh protein serine-kaya diperolehi melalui amplifikasi template cDNA

menggunakan primer tertentu. Hasil kajian menunjukkan bahawa tahap pengekespresan serine-kaya protein transkrip adalah lebih tinggi dalam tumbuhan bakau yang dirawat dengan Si berbanding dengan yang tidak dirawat. Jumlah asid amino serine dalam transgenik *Arabidopsis* telah meningkat dengan ketara dari 1.02 mg g⁻¹ dalam tumbuh-tumbuhan jenis liar sehingga 37.76 mg g⁻¹. Di samping itu, kandungan Si dalam daun dan akar tumbuhan transgenik adalah jauh lebih tinggi daripada jenis liar (P < 0.01). Kajian ini berjaya menentukan Si-responsif transkrip adalah berkaitan dengan *protein serine-kaya* dalam tumbuhan bakau (*R. apiculata*).



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Last but not least, very special thanks to my loving father and mother for their endless love and support.

I certify that a Thesis Examination Committee has met on 13 February 2014 to conduct the final examination of Mahbod Sahebi on his thesis entitled "Molecular Cloning and Functional Studies of Silicon-Responsive Serine-Rich Protein Transcripts from Mangrove Plant, *Rhizophora apiculata* (Blume)" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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LIST OF ABBREVIATIONS

A. tumefaciens Agrobacterium. tumefaciens

Asp Aspartic acid
Arg Arginine
Ala Alanine
aa Amino acid
Bp Base Pairs

cDNA Complementary DNA

CTAB Hexacetyltrimethyl ammonium

bromide

DEPC
Diethyl pyrocarbonate
DNA
Deoxyribonucleic acid
DNase
dNTPs
Ds
Double-stranded

EDTA Ethylene diamine tetra acetic acid
EDX Energy-dispersive X-ray spectroscopy

EtBr Ethidium bromide

G Gram

Glu Glutamic acid
Gly Glycine
His Histidine
Hr

HCl Hydrochloric acid
HPLC High performance liquid

Ile Isoleucine kb Kilo base-pair

L Liter

LB Luria-bertani
LiCl Lithium chloride

Lys
Leu
Leucine
M
Molar
min
Minure
Met
Methionine
mg
Milligram

mg g⁻¹ Miligram per gram

mL Milliliter
mM Millimolar
mRNA Massenger RNA
NaCl Sodium chloride

NCBI National Center for Biotechnology

Information

ng Nanogram
OD Optical density
ORF Open reading frame

OHHydroxide

PCR Polymerase chain reactions **PVP** Polyvinylpyrrolidone

Proline Pro

Phe Phenylalanine **RNA** Ribonucleic acid RTRoom Tempreture

Reverse transcriptase polymerase chain RT-PCR

reaction

Ribonuclease **RNase** Gravity g (rcf)

ROS Reactive oxygen species

SSH Suppression Subtractive Hybridization

SDS Sodium dodecyl sulphate

Sec Second

Scanning electron microscope **SEM**

Serine Ser Si Silicon

Single-stranded SS

Species spp

TAE Tris acetate EDTA TBE Tris borate EDTA

Tris-EDTA TE T-DNA Transfer DNA Threonine Thr Tyr **Tyrosine** Val Valine

X-Gal 5-bromo-4-chloro-3-indolyl-β-D-

galactopyranoside

 $\mu g \mu L^{-1}$ Microgram per microlitre

μL Microliter μg °C Microgram

Degree Centigrade

% Percentage

CHAPTER 1

INTRODUCTION

1.1 General Introduction

One of the highly prevalent elements among the soil ingredients is silicon (Si). Silicon is considered as a non-essential nutrient element for a large number of plant species. Absorption of Si by plants may protect them against a variety of different abiotic stresses including heavy metal toxicity (Neumann and Zur Nieden, 2001), salinity (Tahir *et al.*, 2006), drought (Lux *et al.*, 2002), disproportion of soil nutrients (Jianfeng and Takahashi, 1990; Ma, 2004) and climate changes (Agarie *et al.*, 1998; Ma *et al.*, 2001b). Besides, absorption of Si may increase tolerance of plants to some biotic stresses such as pathogens and pests (Ishiguro, 2001; Meyer and Keeping, 2001). Hence, Si can have an effect on both the yield and quality of agricultural crops.

Several processes such as addition, transformation, deduction, and translocation of different particles are involved in soil formation. Silicate minerals are the most important factors in chemical transformation inside the soil. Minerals rich in Si differ in the intensity and period of numerous specific processes involved in soil formation (Korndörfer and Lepsch, Role of Si as a fertilizer has been reported widely by both horticulturists and agronomists in certain soils leading to increased crop yield and quality (Epstein, 2001). Silicon is the eighth common element in the world which is found in compounds, and it hardly ever occurs as an untainted free element in the nature. The most abundant element in the soil after the oxygen is Si appears in two forms silica and oxides of silicon. The Si is dispersed widely as different forms of silica in the sands, plants, dusts, and planetoids. Silicate minerals form the major portion of the Earth's crust (Ma, 2004). Silicon is considered as a useful element for plant formation and growth, and its absorption by plants helps them to overcome different abiotic and biotic stresses (Ma, 2004). Silicon is used in different industrial fields, such as industrial building in tainted form and extracted through a few steps of processing of the natural compounds to make ceramic brick, concrete. Pure Si is also used in aluminum-casting, making fumed silica, and steel refining; extremely purified Si is used in semiconductor electronics. In biology field tiny trace of Si acts as a vital element for animals (Nielsen, 1984). A variety of microorganisms, such as diatoms use Si to form their structures. Besides to all above-mentioned roles of Si, it has an important role in plants especially in grass metabolism.

Biogenic silica is made by diatoms in the environment, however, it seems that there is a lack in structural control of the process. producing of inorganic silica at ambient conditions needs extreme temperatures and pHs (Iler, 1979). These limitations of producing silica in the environment encouraged material scientists to synthesis biomimetic silica and use it in industrial as well as electronic devices (Morse, 1999; Tacke, 1999; Vrieling et al., 1999). Use of organic molecules has been suggested in the silica biomineralization process (Kinrade et al., 1999; Zhou et al., 1999; Perry and Keeling-Tucker, 2000; Sahai and Tossell, 2001), however, complexes of organic Si have been assumed to be effective on Si uptake and transportation (Hildebrand et al., 1997; Da Silva and Williams, 2001; Sahai and Tossell, 2001). In the current study, Si has been selected due to its important function in crops and sustainable agricultural systems. The molecular mechanisms of Si uptake in most plant species, except for some cereal such as rice and maize, are poorly examined. Silicon absorption and transportation mechanism most probably is genetically different between plants even between different species of the same genus. For instance, three different genes have been identified as Si uptake and transporter genes in the roots and leaves of rice (Ma et al., 2008), while their role and localization are different in maize (Mitani et al., 2009).

Mangrove as woody plants, growing in tropical and sub tropical areas, have a high range of adaptability to different harsh environmental situations and pathogens as well. Because of their particular habitat, mangrove plants may be providing a valuable source of genes responsible for tolerance to a wide range of biotic and abiotic stresses. It has been reported that mangrove plants are able to absorb large amount of Si from the soil through their specific roots and transfer to shoot parts. Therefore, this hypotheses is derived that mangrove would be a beneficial source for genetic manipulation of susceptible plants exposed to stress condition to absorb and accumulate more silicon. Mangrove plants are able to successfully grow in intertidal areas, where sediments usually formed under shortage of essential plant nutrients, oxygen, and accretion of soluble phytotoxins, such as H₂S, CH₄, Fe²⁺ and Mn²⁺(Ponnamperuma, 1984). This is attributed to their anatomical adaptation leading to transport sufficient amount of oxygen to below-ground parts of the roots (Koncalová, 1990; Kludze et al., 1993; Youssef and Saenger, 1996; Cheng et al., 2010).

Mangrove plants are able to accumulate, store and process Si to generate biosilica, which theoretically supposed to be similar to what happens in sponges and diatoms. To understand *in vivo* silica formation and study the required environment for biosilica occurrence, one approach is the isolation and characterization of different sequences of amino-acids and

their structure, association to their related protein functions, followed by investigation of the role of proteins in silica formation *in vitro* (Kauss *et al.*, 2003).

This study investigated the gene responsible for Si accumulation and transportation in mangrove seedling after treatments with Si, using suppression subtractive hybridization (SSH) as a molecular biology technique to make a subtractive cDNA library and examine the expression levels of Si responsive gene. The use of SSH technique lead to remove proteins with the same regulatory functions than those involved in Si uptake and accumulation. There is no information available on the literature relating to the mechanisms in association with Si uptake by roots of mangrove plants. The mechanism of Si accumulation and transportation in mangrove plants should be different with other higher plants and may be controlled by different genes.

The organic substances related to biogenic silica synthesis involves glycoproteins and polysaccharides enriched in hydroxyl terminated group amino acids containing serine, glycine, threonine, glutamic acid and aspartic acid (Hecky et al., 1973; Swift and Wheeler, 1992; Vrieling et al., 1999). Although it is assumed that biosilica can be produced by organisms, exact details of relevant procedures have not been discovered yet. Organic environment includes carbohydrates, lipids, proteins, metal ions and phenolic components in plants, which probably play an elementary role in producing biosilica (Perry and Lu, 1992; Harrison, 1996). Several studies have been done to examine the effect of different amino-acids including (glutamine, serine, lysine, proline, threonine, aspartic acid, asparagines, and arginine) and their homopeptides on silica formation (Coradin and Livage, 2001; Belton et al., 2004).

This is assumed that the *serine- rich protein* may be involved in Si uptake, transport and silica nucleation in matrix-mediated as a stable intermediate. Theses complexes supposed to involve C-O-Si covalent bonds or H- bonds with Si in different coordination forms (Quadra or Penta) (Swift and Wheeler, 1992; Lobel *et al.*, 1996; Kinrade *et al.*, 1999; Da Silva and Williams, 2001).

The primary objective of this thesis was to isolate and characterize novel silicon-responsive genes in mangrove (*Rhizophora apiculata*) roots. Further objective was to investigate the role of protein rich in serine with respect to the gene expression regulation in order to decide if serine-rich protein mediates changes in gene expression. The specific objectives of this study were:

1. To analyze silicon induced changes of gene expression in root and leaves of mangrove.

- 2. To generate Expressed Sequence Tag (EST) library from mangrove after Si treatment.
- 3. To obtain full-length cDNAs clones corresponding to the identified silicon responsive complete cDNA.
- 4. To determine the role of protein rich in serine in accumulation of silicon in roots of transformed *Arabidopsis thaliana*.

This study not only grants genetically information involved in Si absorption by roots of mangrove, it also provides significant information resulted in inducing *serine-rich protein* gene in *Arabidopsis thaliana*. Although so far the molecular role of *serine-rich protein* has not been reported in plants, the role of *serine-rich protein* has been widely studied biochemically and provided highlights information on silica formation.

BIBLIOGRAPHY

- Agarie, S., Hanaoka, N., Ueno, O., Miyazaki, A., Kubota, F., Agata, W., et al. (1998). Effects of silicon on tolerance to water deficit and heat stress in rice plants (*Oryza sativa* L.), monitored by electrolyte leakage. *Plant Production Science*, 1 (2): 96-103.
- Ahmad, R., Zaheer, S. H., and Ismail, S. (1992). Role of silicon in salt tolerance of wheat (*Triticum aestivum* L.). *Plant Science*, 85 (1): 43-50.
- Al-aghabary, K., Zhu, Z., and Shi, Q. (2005). Influence of silicon supply on chlorophyll content, chlorophyll fluorescence, and antioxidative enzyme activities in tomato plants under salt stress. *Journal of Plant Nutrition*, 27 (12): 2101-2115.
- Alongi, D., Sasekumar, A., Chong, V., Pfitzner, J., Trott, L., Tirendi, F., et al. (2004). Sediment accumulation and organic material flux in a managed mangrove ecosystem: estimates of land-ocean-atmosphere exchange in peninsular Malaysia. *Marine Geology*, 208 (2): 383-402.
- Alongi, D. M., Clough, B. F., Dixon, P., and Tirendi, F. (2003). Nutrient partitioning and storage in arid-zone forests of the mangroves Rhizophora stylosa and Avicennia marina. *Trees-Structure and Function*, 17 (1): 51-60.
- Altschul, S. F., Gish, W., Miller, W., Myers, E. W., and Lipman, D. J. (1990). Basic local alignment search tool. *Journal of Molecular Biology*, 215 (3): 403-410.
- Altschul, S. F., Madden, T. L., Schäffer, A. A., Zhang, J., Zhang, Z., Miller, W., et al. (1997). Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Research*, 25 (17): 3389-3402.
- Aly, A. H., Debbab, A., Kjer, J., and Proksch, P. (2010). Fungal endophytes from higher plants: a prolific source of phytochemicals and other bioactive natural products. *Fungal Diversity*, 41 (1): 1-16.
- Armstrong, J., and Armstrong, W. (1988). Phragmites australis-a preliminary study of soil-oxidizing sites and internal gas transport pathways. *New Phytologist*: 373-382.

- Armstrong, J., Armstrong, W., and Beckett, P. M. (1992). Phragmites australis: Venturi-and humidity-induced pressure flows enhance rhizome aeration and rhizosphere oxidation. *New Phytologist*, 120 (2): 197-207.
- Ashraf, M. (2009). Biotechnological approach of improving plant salt tolerance using antioxidants as markers. *Biotechnology Advances*, 27 (1): 84-93.
- Ashraf, M., Afzal, M., Ahmed, R., Mujeeb, F., Sarwar, A., and Ali, L. (2010). Alleviation of detrimental effects of NaCl by silicon nutrition in salt-sensitive and salt-tolerant genotypes of sugarcane (*Saccharum officinarum* L.). *Plant and Soil*, 326 (1-2): 381-391.
- Ball, M. C. (1988). Ecophysiology of mangroves. *Trees-Structure and Function*, 2 (3): 129-142.
- Ball, M. C., Mulkey, S. S., Chazdon, R. L., and Smith, A. P. (1996). Comparative ecophysiology of mangrove forest and tropical lowland moist rainforest.
- Barber, D. A., and Shone, M. G. T. (1966). The absorption of silica from aqueous solutions by plants. *Journal of Experimental Botany*, 17 (3): 569-578.
- Bartlett, J. G., Alves, S. C., Smedley, M., Snape, J. W., and Harwood, W. A. (2008). High-throughput Agrobacterium-mediated barley transformation. *Plant Methods*, 4 (1): 22.
- Baylis, A. D., Gragopoulou, C., Davidson, K. J., and Birchall, J. D. (1994). Effects of silicon on the toxicity of aluminium to soybean. *Communications in Soil Science & Plant Analysis*, 25 (5-6): 537-546.
- Bélanger, R. R., Benhamou, N., and Menzies, J. G. (2003). Cytological evidence of an active role of silicon in wheat resistance to powdery mildew (*Blumeria graminis f.* sp. *tritici*). *Phytopathology*, 93 (4): 402-412.
- Belton, D., Paine, G., Patwardhan, S. V., and Perry, C. C. (2004). Towards an understanding of (bio) silicification: the role of amino acids and lysine oligomers in silicification. *Journal of Materials Chemistry*, 14 (14): 2231-2241.

- Berger, W. H., Smetacek, V. S., and Wefer, G. (1989). Ocean productivity and paleoproductivity-an overview. *Productivity of the ocean: present and past*(44): 1-34.
- Binns, A. N., and Thomashow, M. F. (1988). Cell biology of Agrobacterium infection and transformation of plants. *Annual Reviews in Microbiology*, 42 (1): 575-606.
- Birchall, J. D. (1990). Role of silicon in biology. *Chemistry in Britain*, 26 (2): 141-144.
- Bohnert, H. J., Nelson, D. E., and Jensen, R. G. (1995). Adaptations to environmental stresses. *The Plant Cell*, 7 (7): 1099.
- Bowen, P., Menzies, J., Ehret, D., Samuels, L., and Glass, A. D. M. (1992). Soluble silicon sprays inhibit powdery mildew development on grape leaves. *Journal of the American Society for Horticultural Science*, 117 (6): 906-912.
- Bradbury, M., and Ahmad, R. (1990). The effect of silicon on the growth of Prosopis juliflora growing in saline soil. *Plant and Soil*, 125 (1): 71-74.
- Buatong, J., Phongpaichit, S., Rukachaisirikul, V., and Sakayaroj, J. (2011). Antimicrobial activity of crude extracts from mangrove fungal endophytes. *World Journal of Microbiology and Biotechnology*, 27 (12): 3005-3008.
- Cai, K., Gao, D., Luo, S., Zeng, R., Yang, J., and Zhu, X. (2008). Physiological and cytological mechanisms of silicon-induced resistance in rice against blast disease. *Physiologia Plantarum*, 134 (2): 324-333.
- Casey, W. H., Kinrade, S. D., Knight, C. T. G., Rains, D. W., and Epstein, E. (2004). Aqueous silicate complexes in wheat, Triticum aestivum L. *Plant, Cell & Environment*, 27 (1): 51-54.
- Chan, K. L., Ho, C. L., Namasivayam, P., and Napis, S. (2007). A simple and rapid method for RNA isolation from plant tissues with high phenolic compounds and polysaccharides. *Nature Protocol*, 184.
- Chen, G., Zhu, Y., Wang, H. Z., Wang, S. J., and Zhang, R. Q. (2007). The metabolites of a mangrove endophytic fungus, Penicillium thomi. *Journal of Asian natural products research*, 9 (2): 159-164.

- Chen, H. M., Zheng, C. R., Tu, C., and Shen, Z. G. (2000). Chemical methods and phytoremediation of soil contaminated with heavy metals. *Chemosphere*, 41 (1): 229-234.
- Cheng, H., Liu, Y., Tam, N. F. Y., Wang, X., Li, S. Y., Chen, G. Z., et al. (2010). The role of radial oxygen loss and root anatomy on zinc uptake and tolerance in mangrove seedlings. *Environmental Pollution*, 158 (5): 1189-1196.
- Chérif, M., Asselin, A., and Bélanger, R. R. (1994). Defense responses induced by soluble silicon in cucumber roots infected by *Pythium* spp. *Phytopathology*, 84 (3): 236-242.
- Chérif, M., Benhamou, N., Menzies, J. G., and Bélanger, R. R. (1992). Silicon induced resistance in cucumber plants against *Pythium ultimum*. *Physiological and Molecular Plant Pathology*, 41 (6): 411-425.
- Colmer, T. D., Cox, M. C. H., and Voesenek, L. (2006). Root aeration in rice (*Oryza sativa*): evaluation of oxygen, carbon dioxide, and ethylene as possible regulators of root acclimatizations. *New Phytologist*, 170 (4): 767-778.
- Conesa, A., Götz, S., Garcia-Gomez, J. M., Terol, J., Talon, M., and Robles, M. (2005). Blast2GO: a universal tool for annotation, visualization and analysis in functional genomics research. *Bioinformatics*, 21 (18): 3674-3676.
- Coradin, T., and Livage, J. (2001). Effect of some amino acids and peptides on silicic acid polymerization. *Colloids and Surfaces B: Biointerfaces*, 21 (4): 329-336.
- Curnow, P., Senior, L., Knight, M. J., Thamatrakoln, K., Hildebrand, M., and Booth, P. J. (2012). Expression, purification, and reconstitution of a diatom silicon transporter. *Biochemistry*, *51* (18): 3776-3785.
- da Cunha, K. P. V., and do Nascimento, C. W. A. (2009). Silicon effects on metal tolerance and structural changes in maize (*Zea mays* L.) grown on a cadmium and zinc enriched soil. *Water, Air, and Soil Pollution*, 197 (1-4): 323-330.
- Da Silva, J. J. R. F., and Williams, R. J. P. (2001). *The biological chemistry of the elements: the inorganic chemistry of life*: Oxford University Press.

- Dahdouh-Guebas, F., Jayatissa, L. P., Di Nitto, D., Bosire, J. O., Lo Seen, D., and Koedam, N. (2005). How effective were mangroves as a defence against the recent tsunami? *Current Biology*, *15* (12): R443-R447.
- Danielsen, F., Sørensen, M. K., Olwig, M. F., Selvam, V., Parish, F., Burgess, N. D., et al. (2005). The Asian tsunami: a protective role for coastal vegetation. *Science(Washington)*, 310 (5748): 643.
- Dasgupta, N., Nandy, P., Sengupta, C., and Das, S. (2012). Protein and enzymes regulations towards salt tolerance of some Indian mangroves in relation to adaptation. *Trees*, 26 (2): 377-391.
- Dasgupta, N., Nandy, P., Tiwari, C., and Das, S. (2010). Salinity-imposed changes of some isozymes and total leaf protein expression in five mangroves from two different habitats. *Journal of Plant Interactions*, 5 (3): 211-221.
- Datnoff, L. E., Seebold, K. W., and Correa-V, F. J. (2001). The use of silicon for integrated disease management: reducing fungicide applications and enhancing host plant resistance *Silicon in Agriculture* (Vol. 8, pp. 171-184): Elsevier.
- de Montellano, P. R. O. (2004). Cytochrome P450: structure, mechanism, and biochemistry: Springer.
- Diatchenko, L., Lau, Y. F., Campbell, A. P., Chenchik, A., Moqadam, F., Huang, B., et al. (1996). Suppression subtractive hybridization: a method for generating differentially regulated or tissue-specific cDNA probes and libraries. *Proceedings of the National Academy of Sciences*, 93 (12): 6025-6030.
- Dodd, R. S., Afzal Rafii, Z., and Bousquet-Melou, A. (2001). Evolutionary divergence in the pan-Atlantic mangrove Avicennia germinans. *New Phytologist*, 145 (1): 115-125.
- Doncheva, S. N., Poschenrieder, C., Stoyanova, Z., Georgieva, K., Velichkova, M., and Barcelób, J. (2009). Silicon amelioration of manganese toxicity in Mn-sensitive and Mn-tolerant maize varieties. *Environmental and Experimental Botany*, 65 (2): 189-197.
- Dragišić Maksimović, J., Bogdanović, J., Maksimović, V., and Nikolic, M. (2007). Silicon modulates the metabolism and utilization of phenolic compounds in cucumber (*Cucumis sativus* L.) grown at excess manganese. *Journal of Plant Nutrition and Soil Science*, 170 (6): 739-744.

- Duke, N. C., Benzie, J. A. H., Goodall, J. A., and Ballment, E. R. (1998). Genetic structure and evolution of species in the mangrove genus Avicennia (Avicenniaceae) in the Indo-West Pacific. *Evolution*: 1612-1626.
- Epstein, E. (1994). The anomaly of silicon in plant biology. *Proceedings of the National Academy of Sciences*, 91 (1): 11-17.
- Epstein, E. (1999). Silicon. Annual Review of Plant Physiology and Plant Molecular Biology, 50: 641-664.
- Epstein, E. (2001). Silicon in plants: Facts vs. concepts *Silicon in Agriculture* (Vol. 8, pp. 1-15): Elsevier.
- Espinoza, C., Medina, C., Somerville, S., and Arce-Johnson, P. (2007). Senescence-associated genes induced during compatible viral interactions with grapevine and Arabidopsis. *Journal of Experimental Botany*, *58* (12): 3197-3212.
- Exley, C., and Birchall, J. D. (1993). A mechanism of hydroxyaluminosilicate formation. *Polyhedron*, 12 (9): 1007-1017.
- Faivre, R. O., Cardle, L., Marshall, D., Viola, R., and Taylor, M. A. (2004). Changes in gene expression during meristem activation processes in *Solanum tuberosum* with a focus on the regulation of an auxin response factor gene. *Journal of Experimental Botany*, 397: 613-622.
- Fan, T. W. M., Colmer, T. D., Lane, A. N., and Higashi, R. M. (1993). Determination of Metabolites by 1H NMR and GC: Analysis for Organic Osmolytes in Crude Tissue Extracts. *Analytical Biochemistry*, 214 (1): 260-271.
- Fauteux, F., Chain, F., Belzile, F., Menzies, J. G., and Bélanger, R. R. (2006). The protective role of silicon in the Arabidopsis-powdery mildew pathosystem. *Proceedings of the National Academy of Sciences*, 103 (46): 17554-17559.
- Fawe, A., Abou-Zaid, M., Menzies, J. G., and Bélanger, R. R. (1998). Silicon-mediated accumulation of flavonoid phytoalexins in cucumber. *Phytopathology*, 88 (5): 396-401.
- Feng, J., Shi, Q., Wang, X., Wei, M., Yang, F., and Xu, H. (2010). Silicon supplementation ameliorated the inhibition of photosynthesis and

- nitrate metabolism by cadmium (Cd) toxicity in *Cucumis sativus* L. *Scientia Horticulturae*, 123 (4): 521-530.
- Galvez, L., and Clark, R. B. (1991). Effects of silicon on growth and mineral composition of sorghum (*Sorghum bicolor*) grown with toxic levels of aluminium *Plant-Soil Interactions at Low pH* (pp. 815-823): Springer.
- Galvez, L., Clark, R. B., Gourley, L. M., and Maranville, J. W. (1987). Silicon interactions with manganese and aluminum toxicity in sorghum. *Journal of Plant Nutrition*, 10 (9-16): 1139-1147.
- Galvez, L., Clark, R. B., Gourley, L. M., and Maranville, J. W. (1989). Effects of silicon on mineral composition of sorghum grown with excess manganese 1. *Journal of Plant Nutrition*, 12 (5): 547-561.
- Ganesan, G., Sankararamasubramanian, H. M., Narayanan, J. M., Sivaprakash, K. R., and Parida, A. (2008). Transcript level characterization of a cDNA encoding stress regulated NAC transcription factor in the mangrove plant Avicennia marina. *Plant Physiology and Biochemistry*, 46 (10): 928-934.
- Gao, X., Zou, C., Wang, L., and Zhang, F. (2005). Silicon improves water use efficiency in maize plants. *Journal of Plant Nutrition*, 27 (8): 1457-1470.
- Gelvin, S. B. (2000). Agrobacterium and plant genes involved in T-DNA transfer and integration. *Annual Review of Plant Biology*, 51 (1): 223-256.
- Giri, C., Ochieng, E., Tieszen, L. L., Zhu, Z., Singh, A., Loveland, T., et al. (2011). Status and distribution of mangrove forests of the world using earth observation satellite data. *Global Ecol. Biogeogr.*, 20: 154-159.
- Gong, H. J., Randall, D. P., and Flowers, T. J. (2006). Silicon deposition in the root reduces sodium uptake in rice (*Oryza sativa* L.) seedlings by reducing bypass flow. *Plant, Cell & Environment*, 29 (10): 1970-1979.
- Gonzalez-Mendoza, D., Quiroz Moreno, A., and Zapata-Perez, O. (2008). An improved method for the isolation of total RNA from Avicennia germinans leaves. *Zeitschrift Fur Naturforschung Section A-A Journal of Physical Sciences*, 63 (11): 124.

- Greil, P. (2001). Biomorphous ceramics from lignocellulosics. *Journal of the European Ceramic Society*, 21 (2): 105-118.
- Guével, M. H., Menzies, J. G., and Bélanger, R. R. (2007). Effect of root and foliar applications of soluble silicon on powdery mildew control and growth of wheat plants. *European Journal of Plant Pathology*, 119 (4): 429-436.
- Gurskaya, N. G., Diatchenko, L., Chenchik, A., Siebert, P. D., Khaspekov, G. L., Lukyanov, K. A., *et al.* (1996). Equalizing cDNA subtraction based on selective suppression of polymerase chain reaction: cloning of Jurkat cell transcripts induced by phytohemaglutinin and phorbol 12-myristate 13-acetate. *Analytical Biochemistry*, 240 (1): 90-97.
- Hammond, K. E., Evans, D. E., and Hodson, M. J. (1995). Aluminium/silicon interactions in barley (*Hordeum vulgare* L.) seedlings. *Plant and Soil*, 173 (1): 89-95.
- Hara, E., Kato, T., Nakada, S., Sekiya, S., and Oda, K. (1991). Subtractive cDNA cloning using oligo (dT) 30-latex and PCR: isolation of cDNA clones specific to undifferentiated human embryonal carcinoma cells. *Nucleic Acids Research*, 19 (25): 7097-7104.
- Harrison, C. C. (1996). Evidence for intramineral macromolecules containing protein from plant silicas. *Phytochemistry*, 41 (1): 37-42.
- Hashemi, A., Abdolzadeh, A., and Sadeghipour, H. R. (2010). Beneficial effects of silicon nutrition in alleviating salinity stress in hydroponically grown canola, *Brassica napus* L., plants. *Soil Science & Plant Nutrition*, 56 (2): 244-253.
- Hecky, R. E., Mopper, K., Kilham, P., and Degens, E. T. (1973). The amino acid and sugar composition of diatom cell-walls. *Marine Biology*, 19 (4): 323-331.
- Hedrick, S. M., Cohen, D. I., Nielsen, E. A., and Davis, M. M. (1984). Isolation of cDNA clones encoding T cell-specific membrane-associated proteins. *Nature*, 308 (5955): 149-153.
- Hibino, T., Meng, Y. L., Kawamitsu, Y., Uehara, N., Matsuda, N., Tanaka, Y., et al. (2001). Molecular cloning and functional characterization of two kinds of betaine-aldehyde dehydrogenase in betaine-accumulating mangrove Avicennia marina (Forsk.) Vierh. *Plant Molecular Biology*, 45 (3): 353-363.

- Higuchi, R., Dollinger, G., Walsh, P. S., and Griffth, R. (1992). Simultaneous amplification and detection of specific DNA-sequences. *Bio-Technology*, 10 (4): 413–417.
- Hildebrand, M., Volcani, B. E., Gassmann, W., and Schroeder, J. I. (1997). A gene family of silicon transporters. *Nature*, *385* (6618): 688.
- Hodson, M. J., White, P. J., Mead, A., and Broadley, M. R. (2005). Phylogenetic variation in the silicon composition of plants. *Annals of Botany*, 96 (6): 1027-1046.
- Hogarth, P., and Hogarth, P. J. (2007). The biology of mangroves and seagrasses: Oxford University Press.
- Horiguchi, T. (1988). Mechanism of manganese toxicity and tolerance of plants: IV. Effects of silicon on alleviation of manganese toxicity of rice plants. *Soil Science and Plant Nutrition*, 34 (1): 65-73.
- Horiguchi, T., and Morita, S. (1987). Mechanism of manganese toxicity and tolerance of plants VI. effect of silicon on alleviation of manganese toxicity of barley. *Journal of Plant Nutrition*, 10 (17): 2299-2310.
- Horst, W. J., and Marschner, H. (1978). Effect of silicon on manganese tolerance of bean plants (*Phaseolus vulgaris* L.). *Plant and Soil*, 50 (1-3): 287-303.
- Huang, X., Dai, S., Meng, L., and Zheng, G. (2006). The application of suppression subtractive hybridization (SSH) on isolating plant different genes. *Molecular Plant Breeding*, 5: 735–746.
- Huang, X., and Madan, A. (1999). CAP3: A DNA sequence assembly program. *Genome Research*, 9 (9): 868-877.
- Huang, Z., Cai, X., Shao, C., She, Z., Xia, X., Chen, Y., et al. (2008). Chemistry and weak antimicrobial activities of phomopsins produced by mangrove endophytic fungus Phomopsis sp. ZSU-H76. *Phytochemistry*, 69 (7): 1604-1608.
- Hubank, M., and Schatz, D. G. (1994). Identifying differences in mRNA expression by representational difference analysis of cDNA. *Nucleic Acids Research*, 22 (25): 5640-5648.

- Ii, A. G. P. (2003). An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG II. *Botanical Journal of the Linnean Society*, 141: 399-436.
- Iler, R. K. (1979). The chemistry of silica: solubility, polymerization, colloid and surface properties, and biochemistry: Wiley New York.
- Ishiguro, K. (2001). Review of research in Japan on the roles of silicon in conferring resistance against rice blast *Silicon in Agriculture* (Vol. 8, pp. 277-291): Elsevier.
- Iwasaki, K., Maier, P., Fecht, M., and Horst, W. J. (2002a). Effects of silicon supply on apoplastic manganese concentrations in leaves and their relation to manganese tolerance in cowpea (*Vigna unguiculata* (L.) Walp.). *Plant and Soil*, 238 (2): 281-288.
- Iwasaki, K., Maier, P., Fecht, M., and Horst, W. J. (2002b). Leaf apoplastic silicon enhances manganese tolerance of cowpea (*Vigna unguiculata*). *Journal of Plant Physiology*, 159 (2): 167-173.
- Iwasaki, K., and Matsumura, A. (1999). Effect of silicon on alleviation of manganese toxicity in pumpkin (*Cucurbita moschata* Duch cv. Shintosa). *Soil Science and Plant Nutrition*, 45 (4): 909-920.
- Jarvis, S. C. (1987). The uptake and transport of silicon by perennial ryegrass and wheat. *Plant and Soil*, 97 (3): 429-437.
- Jayaraman, A., Puranik, S., Rai, N. K., Vidapu, S., Sahu, P. P., Lata, C., et al. (2008). cDNA-AFLP analysis reveals differential gene expression in response to salt stress in foxtail millet (*Setaria italica* L.). *Molecular Biotechnology*, 40 (3): 241-251.
- Jennerjahn, T. C., and Ittekkot, V. (2002). Relevance of mangroves for the production and deposition of organic matter along tropical continental margins. *Naturwissenschaften*, 89 (1): 23-30.
- Jianfeng, M. A., and Takahashi, E. (1990). The effect of silicic acid on rice in a P-deficient soil. *Plant and Soil*, 126 (1): 121-125.
- Kanto, T. (2002). Research of silicate for improvement of plant defense against pathogens in Japan. Paper presented at the Second Silicon in Agriculture Conference, Kyoto, Japan.

- Karimi, M., Inzé, D., and Depicker, A. (2002). GATEWAYTM vectors for Agrobacterium-mediated plant transformation. *Trends in Plant Science*, 7 (5): 193-195.
- Kathiresan, K., and Bingham, B. L. (2001). Biology of mangroves and mangrove ecosystems. *Advances in Marine Biology*, 40: 81-251.
- Kathiresan, K., and Rajendran, N. (2005). Coastal mangrove forests mitigated tsunami. *Estuarine, Coastal and Shelf Science, 65* (3): 601-606.
- Kaufman, P. B., Dayanandan, P., Takeoka, Y., Bigelow, W. C., Jones, J. D., and Iler, R. (1981). Silica in shoots of higher plants *Silicon and Siliceous Structures in Biological Systems* (pp. 409-449): Springer.
- Kauss, H., Seehaus, K., Franke, R., Gilbert, S., Dietrich, R. A., and Kröger, N. (2003). Silica deposition by a strongly cationic proline-rich protein from systemically resistant cucumber plants. *The Plant Journal*, 33 (1): 87-95.
- Kavitha, K., George, S., Venkataraman, G., and Parida, A. (2010). A salt-inducible chloroplastic monodehydroascorbate reductase from halophyte Avicennia marina confers salt stress tolerance on transgenic plants. *Biochimie*, 92 (10): 1321-1329.
- Kaya, C., Tuna, L., and Higgs, D. (2006). Effect of silicon on plant growth and mineral nutrition of maize grown under water-stress conditions. *Journal of Plant Nutrition*, 29 (8): 1469-1480.
- Khalid, R. A., and Silva, J. A. (1980). Residual effect of calcium silicate on Ph, phosphorus, and aluminum in a tropical soil profile. *Soil Science and Plant Nutrition*, 26 (1): 87-98.
- Kidd, P. S., Llugany, M., Poschenrieder, C., Gunse, B., and Barcelo, J. (2001). The role of root exudates in aluminium resistance and silicon-induced amelioration of aluminium toxicity in three varieties of maize (*Zea mays* L.). *Journal of Experimental Botany*, 52 (359): 1339-1352.
- Kiefer, E., Heller, W., and Ernst, D. (2000). A simple and efficient protocol for isolation of functional RNA from plant tissues rich in secondary metabolites. *Plant Molecular Biology Reporter*, *18* (1): 33-39.

- Kim, S. G., Kim, K. W., Park, E. W., and Choi, D. (2002). Silicon-induced cell wall fortification of rice leaves: a possible cellular mechanism of enhanced host resistance to blast. *Phytopathology*, *92* (10): 1095-1103.
- Kinrade, S. D., Del Nin, J. W., Schach, A. S., Sloan, T. A., Wilson, K. L., and Knight, C. T. G. (1999). Stable five-and six-coordinated silicate anions in aqueous solution. *Science*, *285* (5433): 1542-1545.
- Kitaya, Y., Yabuki, K., Kiyota, M., Tani, A., Hirano, T., and Aiga, I. (2002). Gas exchange and oxygen concentration in pneumatophores and prop roots of four mangrove species. *Trees-Structure and Function*, 16 (2): 155-158.
- Kludze, H. K., DeLaune, R. D., and Patrick, W. H. (1993). Aerenchyma formation and methane and oxygen exchange in rice. *Soil Science Society of America Journal*, *57* (2): 386-391.
- Kluthcouski, J., and Nelson, L. E. (1980). The effect of silicon on the manganese nutrition of soybeans (*Glycine max* (L.) Merrill). *Plant and Soil*, 56 (1): 157-160.
- Koncalová, H. (1990). Anatomical adaptations to waterlogging in roots of wetland graminoids: limitations and drawbacks. *Aquatic Botany, 38* (1): 127-134.
- Korndörfer, G. H., and Lepsch, I. (2001). Effect of silicon on plant growth and crop yield *Silicon in Agriculture* (Vol. 8, pp. 133-147): Elsevier.
- Kröger, N., Lorenz, S., Brunner, E., and Sumper, M. (2002). Self-assembly of highly phosphorylated silaffins and their function in biosilica morphogenesis. *Science*, 298 (5593): 584-586.
- Kulkarni, V. A., Jagtap, T. G., Mhalsekar, N. M., and Naik, A. N. (2010). Biological and environmental characteristics of mangrove habitats from Manori creek, West Coast, India. *Environmental Monitoring and Assessment*, 168 (1-4): 587-596.
- Lacerda, L. D. (1993). Conservation and sustainable utilization of mangrove forests in Latin America and African regions (Part 1: Latin America). Conservation and Sustainable Utilization of mangrove Forests in Latin America and Africa Regions: Part I-Latin America.
- Lakshmi, M., Rajalakshmi, S., Parani, M., Anuratha, C. S., and Parida, A. (1997). Molecular phylogeny of mangroves I. Use of molecular markers in assessing the intraspecific genetic variability in the

- mangrove species Acanthus ilicifolius Linn.(Acanthaceae). *TAG Theoretical and Applied Genetics*, 94 (8): 1121-1127.
- Lanning, F. C. (1963). Plant Constituents, Silicon in Rice. *Journal of Agricultural and Food Chemistry*, 11 (5): 435-437.
- Lewin, J., and Reimann, B. E. F. (1969). Silicon and plant growth. *Annual Review of Plant Physiology*, 20 (1): 289-304.
- Liang-zoo, S. H. U., and Ying-hui, L. I. U. (2001). Effects of Silicon on Membrane Lipid Peroxidation and Protective Systems in The Leaves of Maize Seedlings Under Salt Stress [J]. *Journal of Xiamen University*, 6: 021.
- Liang, Y. (1999). Effects of silicon on enzyme activity and sodium, potassium and calcium concentration in barley under salt stress. *Plant and Soil*, 209 (2): 217-224.
- Liang, Y., Chen, Q., Liu, Q., Zhang, W., and Ding, R. (2003). Exogenous silicon (Si) increases antioxidant enzyme activity and reduces lipid peroxidation in roots of salt-stressed barley (*Hordeum vulgare* L.). *Journal of Plant Physiology*, 160 (10): 1157-1164.
- Liang, Y., Shen, Q., Shen, Z., and Ma, T. (1996). Effects of silicon on salinity tolerance of two barley cultivars. *Journal of Plant Nutrition*, 19 (1): 173-183.
- Liang, Y., Si, J., and Römheld, V. (2005a). Silicon uptake and transport is an active process in *Cucumis sativus*. *New Phytologist*, 167 (3): 797-804.
- Liang, Y., Wong, J. W. C., and Wei, L. (2005b). Silicon-mediated enhancement of cadmium tolerance in maize (*Zea mays* L.) grown in cadmium contaminated soil. *Chemosphere*, *58* (4): 475-483.
- Liang, Y., Zhang, W., Chen, Q., Liu, Y., and Ding, R. (2006). Effect of exogenous silicon (Si) on H+-ATPase activity, phospholipids and fluidity of plasma membrane in leaves of salt-stressed barley (Hordeum vulgare L.). Environmental and Experimental Botany, 57 (3): 212-219.
- Liang, Y. C., Sun, W. C., Si, J., and Römheld, V. (2005c). Effects of foliarand root-applied silicon on the enhancement of induced resistance to powdery mildew in *Cucumis sativus*. *Plant Pathology*, 54 (5): 678-685.

- Livak, K. J., and Schmittgen, T. D. (2001). Analysis of Relative Gene Expression Data Using Real-Time Quantitative PCR and the 2–ΔΔCT Method. *methods*, 25 (4): 402-408.
- Lobel, K. D., West, J. K., and Hench, L. L. (1996). Computational model for protein-mediated biomineralization of the diatom frustule. *Marine Biology*, 126 (3): 353-360.
- Lukyanov, S., Rebrikov, D., and Buzdin, A. (2007). Suppression Subtractive Hybridization. *Nucleic Acids Hybridization Modern Applications*: 53-84.
- Lumsdon, D. G., and Farmer, V. C. (1995). Solubility characteristics of protoA• imogolite sols: how silicic acid can de-toxify aluminium solutions. *European Journal of Soil Science*, 46 (2): 179-186.
- Lux, A., Luxová, M., Abe, J., Tanimoto, E., Hattori, T., and Inanaga, S. (2003). The dynamics of silicon deposition in the sorghum root endodermis. *New Phytologist*, *158* (3): 437-441.
- Lux, A., Luxova, M., Hattori, T., Inanaga, S., and Sugimoto, Y. (2002). Silicification in sorghum (Sorghum bicolor) cultivars with different drought tolerance. *Physiologia Plantarum*, 115 (1): 87-92.
- Ma, J., and Takahashi, E. (1990). Effect of silicon on the growth and phosphorus uptake of rice. *Plant and Soil*, 126 (1): 115-119.
- Ma, J., and Takahashi, E. (2002). Soil, fertilizer and plant silicon research in Japan. Amsterdam: Elsevier
- Ma, J. F. (2003). Functions of silicon in higher plants *Silicon Biomineralization* (pp. 127-147): Springer.
- Ma, J. F. (2004). Role of silicon in enhancing the resistance of plants to biotic and abiotic stresses. *Soil Science and Plant Nutrition*, 50 (1): 11-18.
- Ma, J. F., Goto, S., Tamai, K., and Ichii, M. (2001a). Role of root hairs and lateral roots in silicon uptake by rice. *Plant Physiology*, 127 (4): 1773-1780.
- Ma, J. F., Mitani, N., Nagao, S., Konishi, S., Tamai, K., Iwashita, T., *et al.* (2004). Characterization of the silicon uptake system and molecular mapping of the silicon transporter gene in rice. *Plant Physiology*, 136 (2): 3284-3289.

- Ma, J. F., Miyake, Y., and Takahashi, E. (2001b). Silicon as a beneficial element for crop plants *Silicon in Agriculture* (Vol. 8, pp. 17-39): Elsevier.
- Ma, J. F., Tamai, K., Ichii, M., and Wu, G. F. (2002). A rice mutant defective in Si uptake. *Plant Physiology*, 130 (4): 2111-2117.
- Ma, J. F., Tamai, K., Yamaji, N., Mitani, N., Konishi, S., Katsuhara, M., et al. (2006). A silicon transporter in rice. *Nature*, 440 (7084): 688-691.
- Ma, J. F., and Yamaji, N. (2006). Silicon uptake and accumulation in higher plants. *Trends in Plant Science*, 11 (8): 392-397.
- Ma, J. F., Yamaji, N., Mitani, N., Tamai, K., Konishi, S., Fujiwara, T., et al. (2007). An efflux transporter of silicon in rice. *Nature*, 448 (7150): 209-212.
- Ma, J. F., Yamaji, N., Mitani, N., Xu, X.-Y., Su, Y.-H., McGrath, S. P., et al. (2008). Transporters of arsenite in rice and their role in arsenic accumulation in rice grain. *Proceedings of the National Academy of Sciences*, 105 (29): 9931-9935.
- Maguire, T. L., Saenger, P., Baverstock, P., and Henry, R. (2001). Microsatellite analysis of genetic structure in the mangrove species Avicennia marina (Forsk.) Vierh.(Avicenniaceae). *Molecular Ecology*, 9 (11): 1853-1862.
- Mann, S., and Ozin, G. A. (1996). Synthesis of inorganic materials with complex form. *Nature*, 382 (6589): 313-318.
- Massel, S. R., Furukawa, K., and Brinkman, R. M. (1999). Surface wave propagation in mangrove forests. *Fluid Dynamics Research*, 24 (4): 219-249.
- Matichenkov, V., and Bocharnikova, E. (2004). Si in horticultural industry *Production Practices and Quality Assessment of Food Crops* (Vol. 2, pp. 217-228): Springer.
- Matichenkov, V. V. (Ed.). (1990). *Amorphous oxide of silicon in soddy podzolic soil and its influence on plants*: Moscow State University.
- Matichenkov, V. V., Ammosova, Y. M., and Bocharnikova, E. A. (1997). The method for determination of plant available silica in soil. *Agrochemistry*, 1: 76-84.

- Matichenkov, V. V., and Calvert, D. V. (2002). Silicon as a beneficial element for sugarcane. *Journal American Society of Sugarcane Technologists*, 22 (2): 21-30.
- Matichenkov, V. V., and Snyder, G. H. (1996). The mobile silicon compounds in some South Florida soils. *Eurasian soil science*, 12: 1165-1173.
- Matoh, T. r., Kairusmee, P., and Takahashi, E. (1986). Salt-induced damage to rice plants and alleviation effect of silicate. *Soil Science and Plant Nutrition*, 32 (2): 295-304.
- Matychenkov, V. V., and Ammosova, Y. M. (1996). Effect of amorphous silica on some properties of a sod-podzolic acid. *Eurasian soil science*, 28 (10): 87-99.
- Matychenkov, V. V., Pinskiy, D. L., and Bocharnikova, Y. A. (1995). Influence of mechanical compaction of soils on the state and form of available silicon. *Eurasian soil science*, 27 (12): 58-67.
- May, G. D., Afza, R., Mason, H. S., Wiecko, A., Novak, F. J., and Arntzen, C. J. (1995). Generation of transgenic banana (Musa acuminata) plants via Agrobacterium-mediated transformation. *Nature Biotechnology*, 13 (5): 486-492.
- Mazda, Y., Kobashi, D., and Okada, S. (2005). Tidal-scale hydrodynamics within mangrove swamps. *Wetlands Ecology and Management, 13* (6): 647-655.
- Menzies, J., Bowen, P., Ehret, D., and Glass, A. D. M. (1992). Foliar applications of potassium silicate reduce severity of powdery mildew on cucumber, muskmelon, and zucchini squash. *Journal of the American Society for Horticultural Science*, 117 (6): 902-905.
- Menzies, J. G., Ehret, D. L., Glass, A. D. M., and Samuels, A. L. (1991). The influence of silicon on cytological interactions between *Sphaerotheca fuliginea* and *Cucumis sativus*. *Physiological and Molecular Plant Pathology*, 39 (6): 403-414.
- Meyer, J. H., and Keeping, M. G. (2001). Past, present and future research of the role of silicon for sugarcane in southern Africa *Silicon in Agriculture* (Vol. 8, pp. 257-275): Elsevier.

- Mikael, K., Jose, M. A., Martin, B., Amin, F., Jiri, J., Kristina, L., et al. (2006). The real-time polymerase chain reaction. *Molecular Aspects of Medicine*, 27: 95-125.
- Mitani, N., and Ma, J. F. (2005). Uptake system of silicon in different plant species. *Journal of Experimental Botany*, *56* (414): 1255-1261.
- Mitani, N., Ma, J. F., and Iwashita, T. (2005). Identification of the silicon form in xylem sap of rice (*Oryza sativa* L.). *Plant and Cell Physiology*, 46 (2): 279-283.
- Mitani, N., Yamaji, N., and Ma, J. F. (2008). Characterization of substrate specificity of a rice silicon transporter, Lsi1. *Pflügers Archiv-European Journal of Physiology*, 456 (4): 679-686.
- Mitani, N., Yamaji, N., and Ma, J. F. (2009). Identification of maize silicon influx transporters. *Plant and Cell Physiology*, *50* (1): 5-12.
- Mittova, V., Guy, M., Tal, M., and Volokita, M. (2004). Salinity upregulates the antioxidative system in root mitochondria and peroxisomes of the wild salt-tolerant tomato species *Lycopersicon pennellii*. *Journal of Experimental Botany*, 55 (399): 1105-1113.
- Miyama, M., Shimizu, H., Sugiyama, M., and Hanagata, N. (2006). Sequencing and analysis of 14,842 expressed sequence tags of burma mangrove, Bruguiera gymnorrhiza. *Plant Science*, 171 (2): 234-241.
- Miyama, M., and Tada, Y. (2008). Transcriptional and physiological study of the response of Burma mangrove (Bruguiera gymnorhiza) to salt and osmotic stress. *Plant Molecular Biology*, 68 (1): 119-129.
- Moussa, H. R. (2006). Influence of exogenous application of silicon on physiological response of salt-stressed maize (*Zea mays* L.). *International Journal of Agriculture and Biology, 8*: 293-297.
- Müller, W. E. G., Wang, X., Wiens, M., Schloßmacher, U., Jochum, K. P., and Schröder, H. C. (2011). Hardening of bio-silica in sponge spicules involves an aging process after its enzymatic polycondensation: Evidence for an aquaporin-mediated water absorption. *Biochimica et Biophysica Acta (BBA)-General Subjects, 1810* (7): 713-726.
- Nandy, P., Das, S., Ghose, M., and Spooner-Hart, R. (2007). Effects of salinity on photosynthesis, leaf anatomy, ion accumulation and

- photosynthetic nitrogen use efficiency in five Indian mangroves. *Wetlands Ecology and Management, 15* (4): 347-357.
- Nandy, P., Dasgupta, N., and Das, S. (2009). Differential expression of physiological and biochemical characters of some Indian mangroves towards salt tolerance. *Physiology and Molecular Biology of Plants*, 15 (2): 151-160.
- Narusaka, Y., Narusaka, M., Yamasaki, S., and Iwabuchi, M. (2012). Methods to Transfer Foreign Genes to Plants.
- Neumann, D., and Zur Nieden, U. (2001). Silicon and heavy metal tolerance of higher plants. *Phytochemistry*, *56* (7): 685-692.
- Nguyen, P. D., Ho, C. L., Harikrishna, J. A., Wong, M. C. V. L., and Rahim, R. A. (2006). Generation and analysis of expressed sequence tags from the mangrove plant, Acanthus ebracteatus Vahl. *Tree Genetics & Genomes*, 2 (4): 196-201.
- Nielsen, F. H. (1984). Ultratrace elements in nutrition. *Annual Review of Nutrition*, 4 (1): 21-41.
- Nikolic, M., Nikolic, N., Liang, Y., Kirkby, E. A., and Römheld, V. (2007). Germanium-68 as an adequate tracer for silicon transport in plants. Characterization of silicon uptake in different crop species. *Plant Physiology*, 143 (1): 495-503.
- Nwugo, C. C., and Huerta, A. J. (2008). Effects of silicon nutrition on cadmium uptake, growth and photosynthesis of rice plants exposed to low-level cadmium. *Plant and Soil*, 311 (1-2): 73-86.
- Ohyama, N. (1985). Amelioration of cold weather damage of rice by silicate fertilizer application. *Agriculture Horticulture*, 60: 1385-1389.
- Oku, H., Baba, S., Koga, H., Takara, K., and Iwasaki, H. (2003). Lipid composition of mangrove and its relevance to salt tolerance. *Journal of Plant Research*, 116 (1): 37-45.
- Okuda, A., and Takahashi, E. (1962). Effect of silicon supply on the injuries due to excessive amounts of Fe, Mn, Cu, As, Al, Co of barley and rice plant. *Soil Science and Plant Nutrition*, 33: 1-8.
- Ong, E. (1982). Mangroves and aquaculture in Malaysia. *Ambio, 11* (5): 252-257.

- Orlov, D. S. (1985). Humus acids of soils: Oxonian Press.
- Parida, A. K., and Jha, B. (2010). Salt tolerance mechanisms in mangroves: a review. *Trees*, 24 (2): 199-217.
- Patrícia Vieira da Cunha, K., Williams Araújo do Nascimento, C., and José da Silva, A. (2008). Silicon alleviates the toxicity of cadmium and zinc for maize (*Zea mays* L.) grown on a contaminated soil. *Journal of Plant Nutrition and Soil Science*, 171 (6): 849-853.
- Peaslee, D. E., and Frink, C. R. (1969). Influence of Silicie Acid on Uptake of Mn, Al, Zn, and Cu by Tomatoes (*Lycopersicum esculentum*) Grown on an Acid Soil. *Soil Science Society of America Journal*, 33 (4): 569-571.
- Perry, C. C., and Keeling-Tucker, T. (2000). Biosilicification: the role of the organic matrix in structure control. *JBIC Journal of Biological Inorganic Chemistry*, **5** (5): 537-550.
- Perry, C. C., and Lu, Y. (1992). Preparation of silicas from silicon complexes: role of cellulose in polymerisation and aggregation control. *J. Chem. Soc., Faraday Trans.*, 88 (19): 2915-2921.
- Peters, F. A. R., Datnoff, L. E., Korndörfer, G. H., Seebold, K. W., and Rush, M. C. (2001). Effect of silicon and host resistance on sheath blight development in rice. *Plant Disease*, 85 (8): 827-832.
- Pi, N., Tam, N. F. Y., Wu, Y., and Wong, M. H. (2009). Root anatomy and spatial pattern of radial oxygen loss of eight true mangrove species. *Aquatic Botany*, 90 (3): 222-230.
- Ponnamperuma, F. N. (1984). Effects of flooding on soils.
- Popp, M. (1984). Chemical composition of Australian mangroves II. Low molecular weight carbohydrates. *Zeitschrift für Pflanzenphysiologie*, 113 (5): 411-421.
- Poulsen, N., Sumper, M., and Kröger, N. (Eds.). (2003) Proceedings of the National Academy of Sciences of the United States of America (Vols. 100).
- Raven, J. A. (2001). Silicon transport at the cell and tissue level *Silicon in Agriculture* (Vol. 8, pp. 41-55): Elsevier.
- Ravikumar, S., Inbaneson, S. J., Suganthi, P., Venkatesan, M., and Ramu, A. (2011). Mangrove plants as a source of lead compounds for the

- development of new antiplasmodial drugs from South East coast of India. *Parasitology Research*, 108 (6): 1405-1410.
- Rebrikov, D. V., Desai, S. M., Siebert, P. D., and Lukyanov, S. A. (2004). Suppression subtractive hybridization *Gene Expression Profiling* (pp. 107-134): Springer.
- Rémus-Borel, W., Menzies, J. G., and Bélanger, R. R. (2005). Silicon induces antifungal compounds in powdery mildew-infected wheat. *Physiological and Molecular Plant Pathology, 66* (3): 108-115.
- Resende, R. S., Rodrigues, F. Á. v., Costa, R. V., and Silva, D. D. (2013). Silicon and fungicide effects on anthracnose in moderately resistant and susceptible Sorghum Lines. *Journal of Phytopathology*, 161 (1): 11-17.
- Richmond, K. E., and Sussman, M. (2003). Got silicon? The non-essential beneficial plant nutrient. *Current Opinion in Plant Biology*, 6 (3): 268-272.
- Robertson, A. I., and Duke, N. C. (1990). Mangrove fish-communities in tropical Queensland, Australia: spatial and temporal patterns in densities, biomass and community structure. *Marine Biology*, 104 (3): 369-379.
- Rodrigues, F. Á., Benhamou, N., Datnoff, L. E., Jones, J. B., and Bélanger, R. R. (2003a). Ultrastructural and cytochemical aspects of siliconmediated rice blast resistance. *Phytopathology*, 93 (5): 535-546.
- Rodrigues, F. A., and Datnoff, L. E. (2005). Silicon and rice disease management. *Fitopatologia Brasileira*, 30 (5): 457-469.
- Rodrigues, F. Á., Vale, F. X. R., Datnoff, L. E., Prabhu, A. S., and Korndörfer, G. H. (2003b). Effect of rice growth stages and silicon on sheath blight development. *Phytopathology*, 93 (3): 256-261.
- Rodrigues, F. A., Vale, F. X. R., Korndörfer, G. H., Prabhu, A. S., Datnoff, L. E., Oliveira, A. M. A., *et al.* (2003c). Influence of silicon on sheath blight of rice in Brazil. *Crop Protection*, 22 (1): 23-29.
- Rogalla, H., and Römheld, V. (2002). Role of leaf apoplast in silicon-mediated manganese tolerance of *Cucumis sativus* L. *Plant, Cell & Environment*, 25 (4): 549-555.

- Romero-Aranda, M. R., Jurado, O., and Cuartero, J. (2006). Silicon alleviates the deleterious salt effect on tomato plant growth by improving plant water status. *Journal of Plant Physiology*, 163 (8): 847-855.
- Romero-Aranda, R., Soria, T., and Cuartero, J. (2001). Tomato plant-water uptake and plant-water relationships under saline growth conditions. *Plant Science*, 160 (2): 265-272.
- Ruan, Y., Gilmore, J., and Conner, T. (1998). Towards *Arabidopsis* genome analysis: monitoring expression profiles of 1400 genes using cDNA microarrays. *The Plant Journal*, 15 (6): 821-833.
- Rubio-Pina, J. A., and Zapata-Perez, O. (2011). Isolation of total RNA from tissues rich in polyphenols and polysaccharides of mangrove plants. *Electronic Journal of Biotechnology*, 14 (5): 11-11.
- Saenger, P. (2003). Mangrove ecology, silviculture and conservation
- Sahai, N., and Tossell, J. A. (2001). Formation energies and NMR chemical shifts calculated for putative serine-silicate complexes in silica biomineralization. *Geochimica et Cosmochimica Acta, 65* (13): 2043-2053.
- Salzman, R. A., Fujita, T., Zhu-Salzman, K., Hasegawa, P. M., and Bressan, R. A. (1999). An improved RNA isolation method for plant tissues containing high levels of phenolic compounds or carbohydrates. *Plant Molecular Biology Reporter*, *17* (1): 11-17.
- Samuels, A. L., Glass, A. D. M., Ehret, D. L., and Menzies, J. G. (1991). Mobility and deposition of silicon in cucumber plants. *Plant, Cell & Environment*, 14 (5): 485-492.
- Savant, N. K., Snyder, G. H., and Datnoff, L. E. (1996). Silicon management and sustainable rice production. *Advances in Agronomy*, *58*: 151-199.
- Savvas, D., Giotis, D., Chatzieustratiou, E., Bakea, M., and Patakioutas, G. (2009). Silicon supply in soilless cultivations of zucchini alleviates stress induced by salinity and powdery mildew infections. *Environmental and Experimental Botany*, 65 (1): 11-17.
- Scheffel, A., Poulsen, N., Shian, S., and Kröger, N. (2011). *Nanopatterned protein microrings from a diatom that direct silica morphogenesis*. Paper presented at the Proceedings of the National Academy of Sciences of the United States of America.

- Schmidt, H. (1995). Die Bedeutung der Mangroven für tropische Küstengewässer: Beispiel Brasilien. *Geogr Rundsch*, 47: 128-132.
- Schultz, D. J., Craig, R., Cox-Foster, D. L., Mumma, R. O., and Medford, J. I. (1994). RNA isolation from recalcitrant plant tissue. *Plant Molecular Biology Reporter*, 12 (4): 310-316.
- Seebold Jr, K. W., Datnoff, L. E., Correa-Victoria, F. J., Kucharek, T. A., and Snyder, G. H. (2004). Effects of silicon and fungicides on the control of leaf and neck blast in upland rice. *Plant Disease*, *88* (3): 253-258.
- Seebold, K. W., Datnoff, L. E., Correa-Victoria, F. J., Kucharek, T. A., and Snyder, G. H. (2000). Effect of silicon rate and host resistance on blast, scald, and yield of upland rice. *Plant Disease*, 84 (8): 871-876.
- Shi, G., Cai, Q., Liu, C., and Wu, L. (2010). Silicon alleviates cadmium toxicity in peanut plants in relation to cadmium distribution and stimulation of antioxidative enzymes. *Plant Growth Regulation*, 61 (1): 45-52.
- Shi, Q., Bao, Z., Zhu, Z., He, Y., Qian, Q., and Yu, J. (2005a). Silicon-mediated alleviation of Mn toxicity in Cucumis sativus in relation to activities of superoxide dismutase and ascorbate peroxidase. *Phytochemistry*, 66 (13): 1551-1559.
- Shi, Q., and Zhu, Z. (2008). Effects of exogenous salicylic acid on manganese toxicity, element contents and antioxidative system in cucumber. *Environmental and Experimental Botany*, 63 (1): 317-326.
- Shi, X.-h., Wang, H., and Zhang, F. S. (2006). Research on the mechanism of silica improving the resistance of rice seedlings to Cd. *Journal of Agro-Environment Science*, 25 (5): 1112-1116.
- Shi, X., Zhang, C., Wang, H., and Zhang, F. (2005b). Effect of Si on the distribution of Cd in rice seedlings. *Plant and Soil*, 272 (1-2): 53-60.
- Shinozaki, K., and Yamaguchi-Shinozaki, K. (1997). Gene expression and signal transduction in water-stress response. *Plant Physiology*, 115 (2): 327.
- Skov, M. W., and Hartnoll, R. G. (2002). Paradoxical selective feeding on a low-nutrient diet: why do mangrove crabs eat leaves? *Oecologia*, 131 (1): 1-7.

- Snedaker, S. C. (1984). Mangroves: A summary of knowledge with emphasis on Pakistan. *Marine geology and oceanography of Arabian Sea and Coastal Pakistan*: 255-262.
- Snyder, G. H., Matichenkov, V. V., and Datnoff, L. E. (2006). Silicon *Plant Nutrition* (pp. 551-562). Belle Glade, Florida.
- Song, A., Li, Z., Zhang, J., Xue, G., Fan, F., and Liang, Y. (2009). Siliconenhanced resistance to cadmium toxicity in Brassica chinensis L. is attributed to Si-suppressed cadmium uptake and transport and Sienhanced antioxidant defense capacity. *Journal of Hazardous Materials*, 172 (1): 74-83.
- Soukup, A., Armstrong, W., Schreiber, L., Franke, R., and Votrubová, O. (2007). Apoplastic barriers to radial oxygen loss and solute penetration: a chemical and functional comparison of the exodermis of two wetland species, Phragmites australis and Glyceria maxima. *New Phytologist*, 173 (2): 264-278.
- Sujatha, G., Reddy, G. P., and Murthy, M. M. K. (1987). Effect of certain biochemical factors on expression of resistance of rice varieties to brown plant hopper (Nilaparvata Lugens Stal.). Agricultural University.
- Sun, H.-J., Uchii, S., Watanabe, S., and Ezura, H. (2006). A highly efficient transformation protocol for Micro-Tom, a model cultivar for tomato functional genomics. *Plant and Cell Physiology*, 47 (3): 426-431.
- Swift, D. M., and Wheeler, A. P. (1992). Evidence of An Organic Matrix From Diatom Biosilica1. *Journal of Phycology*, 28 (2): 202-209.
- Taggart, M. A., Mateo, R., Charnock, J. M., Bahrami, F., Green, A. J., and Meharg, A. A. (2009). Arsenic rich iron plaque on macrophyte roots-an ecotoxicological risk? *Environmental Pollution*, 157 (3): 946-954.
- Tahir, M. A., Rahmatullah, T., Aziz, T., Ashraf, M., Kanwal, S., and Maqsood, M. A. (2006). Beneficial effects of silicon in wheat (*Triticum aestivum* L.) under salinity stress. *Pakistan Journal of Botany*, 38: 1715-1722.
- Takahashi, E., Ma, J. F., and Miyake, Y. (1990). The possibility of silicon as an essential element for higher plants. *comments on agricultural and food chemistry*, 2 (2): 99-102.

- Tamai, K., and Ma, J. F. (2003). Characterization of silicon uptake by rice roots. *New Phytologist*, 158 (3): 431-436.
- Thampanya, U., Vermaat, J., Sinsakul, S., and Panapitukkul, N. (2006). Coastal erosion and mangrove progradation of Southern Thailand. *Estuarine, Coastal and Shelf Science, 68* (1): 75-85.
- Tuna, A. L., Kaya, C., Higgs, D., Murillo-Amador, B., Aydemir, S., and Girgin, A. R. (2008). Silicon improves salinity tolerance in wheat plants. *Environmental and Experimental Botany*, 62 (1): 10-16.
- Tzfira, T., Jensen, C. S., Wang, W., Zuker, A., Vinocur, B., Altman, A., et al. (1997). Transgenic Populus tremula: a step-by-step protocol for its Agrobacterium-mediated transformation. *Plant Molecular Biology Reporter*, 15 (3): 219-235.
- Tzfira, T., Li, J., Lacroix, B. t., and Citovsky, V. (2004). *Agrobacterium* T-DNA integration: molecules and models. *Trends in Genetics*, 20 (8): 375-383.
- Vaculík, M., Lux, A., Luxová, M., Tanimoto, E., and Lichtscheidl, I. (2009). Silicon mitigates cadmium inhibitory effects in young maize plants. *Environmental and Experimental Botany, 67* (1): 52-58.
- Van der Vorm, P. D. J. (1980). Uptake of Si by five plant species, as influenced by variations in Si-supply. *Plant and Soil*, *56* (1): 153-156.
- Vasanthaiah, H. K. N., Katam, R., and Sheikh, M. B. (2008). Efficient protocol for isolation of functional RNA from different grape tissue rich in polyphenols and polysaccharides for gene expression studies. *Electronic Journal of Biotechnology*, 11 (3): 42-51.
- Vasellati, V., Oesterheld, M., Medan, D., and Loreti, J. (2001). Effects of flooding and drought on the anatomy of Paspalum dilatatum. *Annals of Botany, 88* (3): 355-360.
- Visser, E. J. W., Colmer, T. D., Blom, C., and Voesenek, L. (2000). Changes in growth, porosity, and radial oxygen loss from adventitious roots of selected mono-and dicotyledonous wetland species with contrasting types of aerenchyma. *Plant, Cell & Environment*, 23 (11): 1237-1245.
- Vlamis, J., and Williams, D. E. (1967). Manganese and silicon interaction in the Gramineae. *Plant and Soil*, 27 (1): 131-140.

- Vrieling, E. G., Beelen, T. P. M., van Santen, R. A., and Gieskes, W. W. C. (1999). Diatom silicon biomineralization as an inspirational source of new approaches to silica production. *Journal of Biotechnology*, 70 (1): 39-51.
- Wang, Z., and Brown, D. D. (1991). A gene expression screen. *Proceedings* of the National Academy of Sciences, 88 (24): 11505-11509.
- Waterken, L., Bienfait, A., and Peeters, A. (1981). Callose et silice epidermiques. Rapports avec la transpiration cuticulaire. *Cellule*, 73.
- Werner, A., and Stelzer, R. (1990). Physiological responses of the mangrove Rhizophora mangle grown in the absence and presence of NaCl. *Plant, Cell & Environment, 13* (3): 243-255.
- Williams, D. E., and Vlamis, J. (1957). The effect of silicon on yield and manganese-54 uptake and distribution in the leaves of barley plants grown in culture solutions. *Plant Physiology*, 32 (5): 404.
- Woesz, A., Weaver, J. C., Kazanci, M., Dauphin, Y., Aizenberg, J., Morse, D. E., *et al.* (2006). Micromechanical properties of biological silica in skeletons of deep-sea sponges. *Journal of Materials Research*, 21 (08): 2068-2078.
- Woodhead, M., Taylor, M. A., Davies, H. V., Brennan, R. M., and McNicol, R. J. (1997). Isolation of RNA from blackcurrant (Ribes nigrum L.) fruit. *Molecular Biotechnology*, 7 (1): 1-4.
- Yamaji, N., Mitatni, N., and Ma, J. F. (2008). A transporter regulating silicon distribution in rice shoots. *The Plant Cell*, 20 (5): 1381-1389.
- Yamanaka, T., Miyama, M., and Tada, Y. (2009). Transcriptome profiling of the mangrove plant Bruguiera gymnorhiza and identification of salt tolerance genes by Agrobacterium functional screening. *Bioscience Biotechnology and Biochemistry*, 73 (2): 304-310.
- Yang, G., Zhou, R., Tang, T., and Shi, S. (2008). Simple and efficient isolation of high-quality total RNA from Hibiscus tiliaceus, a mangrove associate and its relatives. *Preparative Biochemistry & Biotechnology*, 38 (3): 257-264.
- Yeo, A. R., Flowers, S. A., Rao, G., Welfare, K., Senanayake, N., and Flowers, T. J. (1999). Silicon reduces sodium uptake in rice (*Oryza sativa* L.) in saline conditions and this is accounted for by a

- reduction in the transpirational bypass flow. *Plant, Cell & Environment, 22* (5): 559-565.
- Yildirim, E., Turan, M., and Guvenc, I. (2008). Effect of foliar salicylic acid applications on growth, chlorophyll, and mineral content of cucumber grown under salt stress. *Journal of Plant Nutrition*, 31 (3): 593-612.
- Yongchao, L., and Ruixing, D. (2002). Influence of silicon on microdistribution of mineral ions in roots of salt-stressed barley as associated with salt tolerance in plants. *Life Sciences*, 45 (3): 298-308.
- Yoshida, S. (1965). Chemical aspects of the role of silicon in physiology of the rice plant *Bulletin of the National Institute of Agricultural Sciences Bull* (Vol. Ser B (15), pp. 1-58). Japan.
- Yoshida, S. (1975). *The physiology of silicon in rice, Technical Bulletin-Asian and Pacific Council*. Paper presented at the Food and Fertilizer Technology Center.
- Yoshida, S., Ohnishi, Y., and Kitagishi, K. (1962). Chemical forms, mobility and deposition of silicon in rice plant. *Soil Science and Plant Nutrition*, 8 (3): 15-21.
- Youssef, T., and Saenger, P. (1996). Anatomical adaptive strategies to flooding and rhizosphere oxidation in mangrove seedlings. *Australian Journal of Botany*, 44 (3): 297-313.
- Zeng, H. C., Deng, L. H., and Zhang, C. F. (2006). Cloning of Salt Tolerance• Related cDNAs from the Mangrove Plant Sesuvium portulacastrum L. *Journal of Integrative Plant Biology*, 48 (8): 952-957.
- Zhang, C., Wang, L., Nie, Q., Zhang, W., and Zhang, F. (2008). Long-term effects of exogenous silicon on cadmium translocation and toxicity in rice (*Oryza sativa* L.). *Environmental and Experimental Botany*, 62 (3): 300-307.
- Zhang, W.-W., Jian, G.-L., Jiang, T.-F., Wang, S.-Z., Qi, F.-J., and Xu, S.-C. (2012). Cotton gene expression profiles in resistant Gossypium hirsutum cv. Zhongzhimian KV1 responding to Verticillium dahliae strain V991 infection. *Molecular Biology Reports*, 39 (10): 9765-9774.
- Zhou, Y., Shimizu, K., Cha, J. N., Stucky, G. D., and Morse, D. E. (1999). Efficient catalysis of polysiloxane synthesis by silicatein α requires

- specific hydroxy and imidazole functionalities. *Angewandte Chemie International Edition*, 38 (6): 779-782.
- Zhu, Z., Wei, G., Li, J., Qian, Q., and Yu, J. (2004). Silicon alleviates salt stress and increases antioxidant enzymes activity in leaves of salt-stressed cucumber (*Cucumis sativus* L.). *Plant Science*, 167 (3): 527-533.
- Zipper, H., Brunner, H., Bernhagen, J., and Vitzthum, F. (2004). Investigations on DNA intercalation and surface binding by SYBR Green I, its structure determination and methodological implications. *Nucleic Acids Research*, 32 (12): 103.
- Zupan, J. R., and Zambryski, P. (1995). Transfer of T-DNA from Agrobacterium to the plant cell. *Plant Physiology*, 107 (4): 1041.