



**UNIVERSITI PUTRA MALAYSIA**

**COMPARISON OF LACTIC ACID BACTERIA AND BIFIDOBACTERIA  
FROM HONEY STOMACHS AND HONEYCOMBS OF GIANT  
HONEYBEE (APIS DORSATA) IN KEDAH AND TERENGGANU,  
MALAYSIA**

**NASER TAJABADI**

**FP 2010 14**

**COMPARISON OF LACTIC ACID BACTERIA AND BIFIDOBACTERIA  
FROM HONEY STOMACHS AND HONEYCOMBS OF GIANT HONEYBEE  
(*APIS DORSATA*) IN KEDAH AND TERENGGANU, MALAYSIA**

**By**

**NASER TAJABADI**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfilment of the Requirements for the Degree of Master of Science**

**October 2010**



## **DEDICATION**

I wish to dedicate this thesis to my beloved family, my father, my mother, my son Hadi and my wife Faegheh who always understand and give me loving support.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

**COMPARISON OF LACTIC ACID BACTERIA AND BIFIDOBACTERIA  
FROM HONEY STOMACHS AND HONEYCOMBS OF GIANT HONEYBEE  
(*APIS DORSATA*) IN KEDAH AND TERENGGANU, MALAYSIA**

By

**NASER TAJABADI**

**October 2010**

**Chairman: Professor Dato Makhdzir Mardan, PhD**

**Faculty: Agriculture**

The isolation and identification of lactic acid bacteria (LAB) in the honey stomachs and honeycombs of Giant honeybee (*Apis dorsata*) from different areas in Malaysia were performed to prospect for beneficial bacteria or probiotics. Honey stomach samples of honeybee and honeycomb filled with honey were collected from *Apis dorsata* colonies in different bee trees from the rainforests of Pedu Lake in Kedah state and the agricultural region of Marang in Terengganu, Malaysia. The isolates were cultured anaerobically in different media of MRS agar, MRS broth and TPY broth at 37°C. Three hundred and fourteen isolates were obtained and identified based on biochemical tests, as LAB. In addition, 20 uncultured samples were identified as *Bifidobacterium* by using the nested polymerase chain reaction (PCR). The 16S rRNA genes from extracted DNA of bacterial colonies were amplified with PCR and nested PCR using universal



primers of 27F and 1492R for LAB, and for *Bifidobacterium* two pairs of genus specific primers (Lm26 paired with Lm3 and Bif164-F paired with Bif662-R) were utilized. All bacterial 16S rRNA genes were sequenced and entrusted in GenBank and were given specific accession numbers. Phylogenetic analysis of the 16S rRNA sequences showed that the novel LAB and *Bifidobacterium* isolates could be grouped into four different phylotypes. They were found to be composed of 37 *Lactobacillus*, 6 *Enterococcus*, 2 *Bifidobacterium* and 1 *Weissella* phylotypes. The results showed that among 334 isolates and sequences, *Lactobacillus* spp with 64.97% were found to represent the most common LAB in *Apis dorsata* honey stomachs and honeycombs followed by *Enterococcus* spp with 28.74%, *Bifidobacterium* with 5.99% and the rest 0.3% *Weissella*. Overall, the predominant lactobacilli species found in the total samples constituting approximately 40.55% of *Lactobacillus kunkeei*. *Lactobacillus kunkeei* (YH-15) related sequences (88) were the predominant lactobacilli followed by 33.18% of other *Lactobacillus* sp with related sequences (72). In honey stomach samples, the prevalence of *Lactobacillus* spp. was 53.21%. Interestingly, the predominant *Lactobacillus* species found in all honey stomach samples was *Lactobacillus plantarum*. (30.12%) followed by *Lactobacillus pentosus* (27.71%), *Lactobacillus kunkeei* (22.89%), *Lactobacillus* sp (15.66%), *Lactobacillus vermiform* (2.41%) and *Lactobacillus fermentum* (1.21%). Samples from the honey-filled honeycomb showed a prevalence of *Lactobacillus* spp. at 75.28 %. The predominant *Lactobacillus* species from all honeycomb samples constituting of *Lactobacillus kunkeei* (51.49%), *Lactobacillus* sp (44.03%) and *Lactobacillus alvei* (4.48%). *Enterococcus* spp. was isolated from every honey stomach (40.38%) and honeycomb (18.54%) of Kedah and



Terengganu samples. The predominant *Enterococcus* species from all honey stomach and honeycomb samples, constituting approximately 88.54% of total isolated *Enterococcus* spp., was *Enterococcus* sp and the rest was *Enterococcus faecalis* (11.46%). Whereas, Bifidobacteria and *Weissilla* account for about 5.99% and 0.3% respectively.

In conclusion, the diversity of total bacterial contents of honey stomachs and honeycombs in highland were somewhat different from the lowland data. This discrepancy reflects the corresponding difference associated with the high diversity of flower, nectar and pollen in forest area of Kedah state versus a low diversity of flower, nectar and pollen in agriculture area of Terengganu state.

Abstrak tesis yang dilemukan kepada Senat Universiti Putra Malaysia bagi memenuhi keperluan untuk ijazah Master Sains

**PERBANDINGAN BAKTERIA ASID LAKTIK DAN BIFIDOBACTERIA DARI MADU DAN SARANG LEBAH MADU APIS DORSATA DARI KEDAH DAN TERENGGANU, MALAYSIA**

Oleh

**NASER TAJABADI**

**Oktober 2010**

**Pengerusi: Profesor Dato Makhdzir Mardan, PhD**

**Fakulti: Pertanian**

Pemencilan dan pencirian bakteria asid laktik (LAB) dari perut madu dan sarang *Apis dorsata* dari pelbagai kawasan di Malaysia telah dijalankan bagi mendapatkan bakteria baik atau probiotik. Sampel perut madu dan sarang yang dipenuhi madu telah dikumpulkan dari koloni *Apis dorsata* dari pokok hutan hujan di Kedah dan kawasan pertanian di daerah Marang, Terengganu. Semua isolat dibiakkan secara anaerobik space media MRS agar, MRS broth dan TPY broth pada suhu 37°C. Sebanyak 314 isolat telah diperolehi dan dikenalpasti sebagai LAB menggunakan ujian biokimia. Space itu, sebanyak 20 sampel yang tidak boleh dibiakkan telah dikenaplasti sebagai *Bifidobacterium* dengan menggunakan PCR. Gen 16S rRNA dari DNA yang telah diekstrak dari koloni bakteria telah digandakan dengan kaedah PCR dan *nested* PCR



menggunakan primer *universal* 27F dan 1492R bagi LAB dan untuk *Bifidobacterium*, 2 pasang primer yang spesifik kepada genus *Bifidobacterium* (Lm26 berpasangan dengan Lm3 dan Bif164-F berpasangan dengan Bif662-R) telah digunakan. Semua urutan DNA gen 16S rRNA telah diujukkan dan disimpan di bank gen dengan diberi nombor capaian tertentu. Analisis pilogenetik untuk urutan 16S rDNA menunjukkan bahawa LAB dan *Bifidobacterium* boleh dibahagikan kepada 4 kumpulan pilotipik. Mereka terdiri dari 37 *Laktobasillus*, 6 *Enterokokkus*, 2 *Bifidobakterium* dan 1 *Weissella*. Keputusan menunjukkan bahawa diantara 334 isolat dan urutan, *Laktobasillus* spp dengan 64.97% adalah yang paling dominan LAB space perut madu dan sarang lebah *Apis dorsata* diikuti oleh *Enterococcus* spp dengan 28.74%, *Bifidobacterium* dengan 5.99% dan selebihnya 0.3% *Weissilla*. Secara keseluruhan, nya spesis *Laktobasillus* yang paling banyak ialah *L. kunkeei* (40.53%), diikuti oleh *Laktobasillus* sp. (33.18%). Didalam sampel perut madu, *Laktobacillus* spp. Hadir sebanyak 53.21%. Menariknya, *Laktobasillus* spesis yang dominan dari sampel perut madu adalah *Laktobasillus plantarum* (31.12%), diikuti oleh *Laktobacillus pentosus* (27.71%), *Laktobacillus kunkeei* (22.89%), *Laktobacillus* sp (15.66%), *Laktobacillus vermiform* (2.41%) dan *Laktobacillus fermentum* (1.21%). Sampel dari sarang yang dipenuhi madu pula menunjukkan kehadiran *Laktobacillus* spp. Sebanyak 75.28%. *Laktobacillus* yang dominan didalam sampel ini ialah *Laktobacillus kunkeei* (51.49%), *Laktobacillus* sp. (44.48%) dan *Laktobacillus alvei* (4.48%). *Enterococcus* telah dipencilkan dari setiap perut madu (40.38%) dan sarang (18.54%) dari sampel Kedah dan Terengganu. *Enterococcus* yang dominan dari semua sampel perut madu dan sarang yang mewakili sebanyak 88.54% ialah *Enterococcus* sp dan selebihnya ialah *Enterococcus faecalis*





(11.46%). Manakala, *Bifidobacterium* dan *Weisella* pula masing-masing sebanyak 5.99% dan 0.3%. Keputusan kami menunjukkan bahawa terdapat kepelbagaian yang besar space spesis bakteria dari perut madu dan sarang lebah *Apis dorsata*.

Kesimpulannya, kepelbagaian kandungan bakteria didalam perut madu dan sarang madu pada lebah dari tanah tinggi dan tanah rendah adalah berbeza. Perbezaan ini adalah saling berkait dan disebabkan oleh taburan atau kepelbagaian bunga, nektar dan polen yang berbeza diantara tanah tinggi di Kedah dan kawasan pertanian di Terengganu.

## ACKNOWLEDGEMENTS

Praise is to God, Who has spurred me on through his command: ‘Read! And thy Lord is Most Bountiful, He Who taught (the use of) the Pen’.

I am heartily thankful to my supervisor, Dato Prof Dr. Makhdzir Mardan, whose encouragement, guidance and support from the initial to the final level enabled me to develop an understanding of the subject.

Secondly my gratitude goes to Prof. Dr. Mohd Yazid Abdul Manap. He managed to keep the fragile balance between providing guidance and freedom. This allowed me to make mistakes, discover many aspects of science, and to explore a variety of different topics.

Thirdly, special thanks to Prof. Madya Dr. Shuhaimi Mustafa. He possesses the invaluable ability to provide – within a blink of an eye - excellent feedback to a sciatic problem, and re-establish both condense and motivation.

My heart-felt thanks and appreciation goes to my son Hadi and my wife Faegheh for their sacrifice, patience, understanding, help and encouragement throughout the study.

I would like to thank all students and members of the Food Biotechnology laboratory, past and present, for their friendship, support and advice.

I also give special thank to Dr. Amir Meimandipour and Mr. Arash Javanmard for they valuable professional advice upon consultation. Thanks to Assoc. Prof. Dr. Jothi Malar



Panandam and the members of her laboratory for allowing me to use their gel Doc. Also, thanks to Dr. Hamed Mirhossini for advice on statistical analysis.

I thank Prof. Dr. Halimi Mohd Saud, the Department Head of Agriculture Technology Faculty of Agriculture, and staff of Graduate School of Universiti Putra Malaysia for helping me in one way or another towards the completion of this study.

My deepest appreciation to my father, mother, brother, sisters and my in-law family members for their constant encouragement and also support during the course of the study.

I wish to express my thankfulness to my friends, Dr. Seyed Reza Hashemi, Rasoul Bahreini, Mohammad Mehdi Saberioon, Alireza Majidi, Morteza Karami, Mehdi Pasebani, Seyed Eemin Noorae, Abdoreza Soleimani and Seyed Nikbin for their encouragement throughout my study. Lastly, I offer my regards and blessings to all of those who supported me in any way in the completion of the project.



I certify that an Examination Committee has met on 1 October 2010 to conduct the final examination of Naser Tajabadi on his thesis entitled “Comparison of Lactic Acid Bacteria and Bifidobacteria from Honey Stomachs and Honeycombs of Giant Honeybee (*Apis dorsata* F.) of Kedah and Terengganu, Malaysia” in accordance with the University colleges Act 1971 and the Constitution of Universiti Putra Malaysia [P.U. (A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

**Halimi Mohd Saud, PhD**

Associate Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Chairman)

**Rosfarizan Mohamad, PhD**

Associate Professor  
Faculty of Biotechnology and Biomolecular Science  
Universiti Putra Malaysia  
(Internal Examiner)

**Fatimah Abu Bakar, PhD**

Associate Professor  
Faculty of Food Science and Technology  
Universiti Putra Malaysia  
(Internal Examiner)

**Abdul Manaf Ali, PhD**

Professor  
Faculty of Agriculture and Biotechnology  
Universiti Darul Iman Malaysia  
(External Examiner)

---

**BUJANG BIN KIM HUAT, PhD**

Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Makhdzir Mardan, PhD**

Dato, Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Chairman)

**Mohd Yazid Abd Manap, PhD**

Professor  
Faculty of Food Science and Technology  
Universiti Putra Malaysia  
(Member)

**Shuhaimi Mustafa, PhD**

Associated Professor  
Faculty of Biotechnology and Biomolecular Science  
Universiti Putra Malaysia  
(Member)

---

**HASANAH MOHD GHAZALI, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:



## **DECLARATION**

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.

---

**NASER TAJABADI**

Date 1 October 2010



## TABLE OF CONTENTS

	<b>Page</b>
<b>DEDICATION</b>	ii
<b>ABSTRACT</b>	iii
<b>ABSTRAK</b>	vi
<b>ACKNOWLEDGEMENTS</b>	ix
<b>APPROVAL SHEETS</b>	xi
<b>DECLARATION FORM</b>	xiii
<b>LIST OF TABLES</b>	xvii
<b>LIST OF FIGURES</b>	xviii
<b>LIST OF ABBREVIATIONS</b>	xx
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
<b>2 LITERTURE REVIEW</b>	<b>5</b>
2.1 Lactic Asid Bacteria	5
2.1.1 Genus <i>Lactobacillus</i>	5
2.1.2 Genus <i>Enterococcus</i>	7
2.1.3 Genus <i>Bifidobacterium</i>	7
2.1.4 Genus <i>Weissella</i>	8
2.2 Honeybee	9
2.2.1 Honeybee <i>Apis dorsata</i>	10
2.3 Honey	11
2.3.1 Honey Production and Collection	11
2.3.2 Composition of Honey	13
2.3.3 Honey as Medicine	15
2.3.4 Honey as Food	16
2.3.5 Honey as Prebiotic	17
2.4 Honeybee Gastrointestinal Tract	18
2.4.1 Indigenous Microflora in Honeybee Gastrointestinal Tract	20
2.4.2 Indigenous Microflora in Honey Stomach and Honeycomb	21
2.5 Probiotics	22
2.5.1 History and Definition of Probiotics	22
2.5.2 Isolation and Identification of LAB	23
2.5.3 Therapeutic Potential and Health Benefits of Probiotics	25
2.6 Traditional Approaches for the Analysis of Microbial Diversity	26
2.6.1 Morphological Methods use on Identification of Microflora Diversity	27
2.7 Molecular Techniques used to Identification Microflora Diversity	27
2.7.1 Methods for Testing DNA Purity and Quantification	28
2.7.2 Polymerase Chain Reaction (PCR)	30
2.7.3 Nested PCR	31



2.7.4	Sequencing	31
2.7.5	Sequence Alignment	32
2.7.6	Treeing Methods	33
<b>3</b>	<b>MATERIALS AND METHODS</b>	<b>35</b>
3.1	Sample Collections	35
3.2	Monitoring of pH	38
3.3	Culture Method	38
3.4	Biochemical Screening	39
3.4.1	Gram Stain Test	39
3.4.2	Catalase Test	40
3.5	Molecular Method	41
3.5.1	DNA Extraction	41
3.5.2	DNA Purity Checking and Quantification	42
3.5.3	PCR Primers	43
3.5.4	Polymerase Chain Reaction (PCR) and Program	44
3.5.5	Nested PCR	45
3.5.6	Gel Documentation	46
3.5.7	PCR Product Purification	47
3.5.8	DNA Sequencing and Blast	47
3.5.9	Analyzes of DNA Sequences using BioEdit	48
3.6	Nucleotide Sequence Accession Numbers	49
3.7	Statistical Analysis	49
<b>4</b>	<b>RESULTS AND DISCUSSION</b>	<b>50</b>
4.1	Isolation of Lactic Acid Bacteria	40
4.2	Phenotypic Identification	51
4.2.1	Morphological Examination	51
4.2.2	Gram Staining and Catalase Test	52
4.3	Genotypic Identification	56
4.3.1	Amplification of 16S rRNA Gene	57
4.3.2	Elution of PCR Product	59
4.3.3	Sequence analysis of the 16S rRNA gene	60
4.4	Phylogenetic Analysis of LAB	60
4.4.1	Phylogenetic Analysis of <i>Lactobacillus</i> Bacteria	61
4.4.2	Phylogenetic Analysis of <i>Enterococcus</i> Bacteria	83
4.4.3	Phylogenetic Analysis of <i>Bifidobacterium</i> in Honey Stomachs and Honeycombs	89
4.4.4	Phylogenetic Analysis of <i>Weissella</i>	93
<b>5</b>	<b>CONCLUSIONS AND RECOMMENDATION FOR FUTURE RESEARCHS</b>	<b>96</b>
	<b>REFERENCES</b>	<b>102</b>
	<b>APPENDICES</b>	<b>115</b>
	<b>BIODATA OF STUDENT</b>	<b>140</b>
	<b>LIST OF PUBLICATIONS</b>	<b>141</b>

