

#### **UNIVERSITI PUTRA MALAYSIA**

# IMMUNE RESPONSES TO A LIVE VIRUS VACCINE CANDIDATE AND PROTECTION AGAINST PSEUDORABIES VIRUS INFECTION

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FPV 1999 5



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ABDEL-WAHID SAEED ALI BABIKER

DOCTOR OF PHILOSOPHY UNIVERSITI PUTRA MALAYSIA

1999



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# By ABDEL-WAHID SAEED ALI BABIKER

Thesis Submitted in Fulfilment of the Requirements for the Degree of Doctor of Philosophy in the Faculty of Veterinary Medicine Universiti Putra Malaysia

December 1999



#### **DEDICATED TO MY**

PARENTS, SAEED & SITTANA
BROTHERS, ABDEL-GADIR & ALI
SISTERS, UMMUHANI, KHADIJA, AISHA, EHSAN, RAHMA, IGBAL,
FATIMA & SHAHINDA

AND TO MY WIFE, AWADIA

FOR THEIR PATIENCE, CONSTANT ENCOURAGEMENT AND SUPPORT



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#### LIST OF ABBREVIATIONS

Ab(s) Antibody(ies)

ABTS 2,2-azino-bis (3-ethylbenzthiazoline-6-Sulfonic acid)

AD/ADV Aujeszky's Disease/Aujeszky's Disease Virus

ATV Antibiotic Trypsin Versin BHV Bovine herpesvirus

bp Base Pair

BSA Bovine serum albumin °C Degree Celcius

CEF Chicken Embryo Fibroblast

cm Centimeter

cm<sup>2</sup> Centimeter square
CMI Cell-mediated immunity
CNS Central nervous system

CO<sub>2</sub> Carbon dioxide
CPE Cytopathic effect
CPS Cyclophosphamide
D.W Distilled water

DNA Deoxyribonucleic Acid

Dr. Doctor

DTH Delayed-type Hypersensitivity

DXM Dexamethasone e.g. For example EBV Epstein Barr Virus

EDTA Ethylene Diamine Tetra-acetate

EHV Equine Herpesvirus

ELISA Enzyme Linked Immunosorbent Assay

f.p. Footpad

FC Final concentration FCS Foetal Calf Serum FLM Flumethasone

g Gram

H<sub>2</sub>O<sub>2</sub> Hydrogen peroxide

h/hrs Hour/Hours

HIS Hyperimmune Serum HSV Herpes-simplex Virus IE Immediate early

i.e. That is

i.m. Intramusculari.n. Intranasali.p. Intraperitoneal

IBRV Infectious Bovine Rhinotracheitis Virus

IgA Immunoglobulin A
IgG Immunoglobulin G
IgM Immunoglobulin M
IR Inverted repeat



Kbp Kilobase pairs Kda Kilodalton

L15 Leibovitz's 15 media

LAT Latency associated transcripts

M Molar

Mab Monoclonal antibody

MLV/ MLVs Modified live vaccine/ Modified live vaccines

M.W Molecular Weight

mg Milligram

MHC Major histocompatibility complex

min/ mins Minute/ Minutes

ml Millilitre mM Millimolar

N-Ab(s) Neutralizing antibody(ies)

NCEM Negative Contrast Electron Microscope

nm Nanometer
O.D. Optical Density
p.f.u. Plaque Forming Unit
p.i. Post-Inoculation

PAGE Polyacrylamide Gel Electrophoresis

PBS Phosphate Buffer Saline

PBST Phosphate Buffer Saline Tween 20

PCR Polymerase chain reaction pH Hydrogen-ion Concentration

PrV Pseudorabies Virus

RE Restriction Endonuclease

RNA Ribonucleic Acid Rpm Revolution per minute

s.c. Subcutaneous

SDS Sodium Dodecyl Sulphate

Sec Sec

SNT Serum Neutralization Test

TE Tris-EDTA

TK Thymidine kinase TNE Tris-NaCl-EDTA

UL Unique long region of PrV genome

UPM Universiti Putra Malaysia

US Unique short region of PrV genome

UV Ultraviolet V/V Volume/Volume

Vero Cell line derived from kidney tissue of green African monkey

VZV Varicella Zoster Virus

w/v
w/w
Weight/Volume
w/w
μg
Microgram
μl
Microlitre
%
Percent



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the degree of Doctor of Philosophy

IMMUNE RESPONSES TO A LIVE VIRUS VACCINE CANDIDATE AND PROTECTION AGAINST PSEUDORABIES VIRUS INFECTION

By

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December 1999

Chairman: Mohd. Azmi Mohd. Lila, Ph.D.

Faculty: Veterinary Medicine

A comparative study of three Malaysian pseudorabies virus (PrV) isolates, one Malaysian PrV strain and one American reference PrV was carried out using restriction endonucleases. Variations were observed among these viruses when their DNAs were digested with only *Kpn* I and *Bam*H I enzymes. The Malaysian strain (PrV-mA1p) showed restriction pattern which was different from its parental virus. An obvious differences were observed between the reference virus and the Malaysian PrVs. When the SDS-PAGE technique was employed, no significant variations in protein profiles of the PrVs but variations in their immunogenic proteins were observed when they were reacted against homologous and heterologous antisera in Western blotting. The viruses were suggested to belong to the same serologic sub-type using SNT and ELISA tests.

Experiments in mice demonstrated that all the viruses were pathogenic to various extents while PrV-mA1p was found to be non-pathogenic even when mice were infected with 10<sup>8</sup> p.f.u. The study showed that 100% protection was obtained following immunization of mice with at least 10<sup>5</sup> p.f.u. However, the

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protection level was highly dose and route of immunization dependent. Antibody (Ab) responses to PrV-mAlp were correlated with the protection levels obtained and demonstrated to persist in mouse serum for five consecutive months.

Low levels of neutralizing Ab (N-Ab) were detected in mice following immunization with PrV-mA1p depending on the dose and route of inoculation. They were correlated with the protection levels but not with the ELISA Ab titres. Higher DTH responses were observed in mice immunized with PrV-mA1p especially via intranasal route (i.n.). A second dose of immunization slightly increased the DTH response. A significant (p<0.05) increase of PrV-mA1p growth was observed in mice tissues treated with immunosuppressive agents. However, the highest virus titres were detected in cyclophosphamide (CPS)-treated mice followed by dexamethasone (DXM)- and flumethasone (FLM)-treated mice. A significant (p<0.01) decline in Ab titres following treatment of mice with all the drugs was noted. However, the highest decline was observed in CPS-treated mice. PrV-mA1p was reactivated in low titres from latently infected nervous tissues of mice using CPS and DXM but not from respiratory tissues.

Increases in the protection levels and Ab responses were observed in mice immunized with the algammulin-adjuvanated PrV-mAlp. PrV-mAlp was also found to be non-pathogenic for swine following inoculation of piglets with up to  $10^8$  p.f.u. but displayed high levels of immunogenicity in piglets, hence the virus was highly suitable to be proposed as a live virus vaccine against PrV infection.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan Ijazah Doktor Falsafah

GERAKBALAS IMUN TERHADAP CALON VAKSIN VIRUS HIDUP DAN PERLINDUNGAN TERHADAP JANGKITAN VIRUS PSEUDORABIES

Oleh

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Desember 1999

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Suatu kajian bandingan terhadap pencilan virus pseudorabies Malaysia (PrV), satu strain PrV Malaysia dan satu PrV rujukan Amerika telah dijalankan mengguna endonuklease pengehadan. Perbezaan telah dicerap di kalangan virus ini apabila DNAnya dicernakan dengan enzim *Kpn* I dan *Bam*H 1 sahaja. Strain Malaysia (PrV-mA1p) menunjukkan pola pengehadan yang berbeza daripada virus induknya. Suatu perberzaan yang nyata dicerapkan di antara virus rujukan dan PrV Malaysia. Apabila teknik SDS-PAGE digunakan, tiada perbezaan tererti didapati dalam profil protein PrV tetapi perbezaan dalam protein imunogen telah dicerapkan apabila ianya ditindakkan dengan antiserum homologus dan heterologus dalam sap Western. Dengan mengguna ujian SNT dan ELISA, virus-virus ini disarankan adalah daripada sub-tip serologi yang sama.

Ujikaji pada mencit menunjukkan yang kesemua virus ini patogen pada tahap yang berbeza-beza, sambil PrV-mAlp pula didapati bukan patogen walaupun mencit ini telah dijangkitkan dengan 10<sup>8</sup> p.f.u. Kajian ini menunjukkan

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yang perlindungan 100% diperolehi berikutan pengimunan mencit dengan sekurang-kurangnya 10<sup>5</sup> p.f.u. Bagaimanapun, aras perlindungan ini paling bersandarkan dos dan cara pengimunan. Gerak balas antibodi (Ab) terhadap PrV-mA1p berkorelasikan dengan aras perlindungan dan ini telah ditunjuk berterusan dalam serum mencit selama 5 bulan.

Aras Ab peneutralan (N-Ab) yang rendah telah dikesan dalam mencit berikutan pengimunan dengan PrV-mA1p, iaitu bergantung kepada dos dan cara penginokulatan. Ini berkorelasi dengan aras perlindungan tetapi bukan dengan titer Ab ELISA. Gerak balas DTH dicerap lebih tinggi dalam mencit terimun dengan PrV-mA1p, khususnya dengan cara intranasum (i.n.). Satu dos pengimunan kedua meningkatkan sedikit gerak balas DTH ini. Peningkatan tererti (p<0.05) pertumbuhan PrV-mA1p dicerapkan dalam tisu mencit yang diperlakukan dengan agen imunotindas. Bagaimanapun, titer virus paling tinggi dikesan dalam mencit terperlaku siklofosfamida (CPS) diikuti dengan mencit terperlaku deksametason (DXM) dan flumetason (FLM). Penurunan tererti (P<0.01) dalam titer Ab berikutan perlakuan mencit dengan kesemua drug ini telah dilihat. Bagaimanapun, penurunan paling besar dicerapkan dalam mencit terperlaku CPS. PrV-mA1p telah diaktif semula sekadar titer rendah daripada tisu saraf mencit terjangkit pendam mengguna CPS dan DXM, tetapi perkara ini tidak berlaku daripada tisu pernafasan.

Peningkatan dalam aras perlindungan dan gerak balas Ab dicerapkan dalam mencit yang terimun dengan PrV-mA1p teradjuvan algamulin. PrV-mA1p



juga didapati bukan patogen untuk babi berikutan penginokulatan anaknya dengan setinggi 10<sup>8</sup> p.f.u., tetapi telah menunjukkan keimunogenan aras tinggi dalam anak babi tersebut, dan justeru itu virus ini adalah paling sesuai disarankan sebagai vaksin virus hidup terhadap jangkitan PrV.



#### CHAPTER I

#### **INTRODUCTION**

Pseudorabies virus (PrV) is a neurotropic herpesvirus that causes pseudorabies in domestic and wild animals (Gustafson, 1986; Pensaert and Kluge, 1989). The first description of PrV infectivity was made in 1902 by Aujeszky, a Hungarian scientist (Aujeszky, 1902) hence the name Aujeszky's disease. Pigs are relatively resistant to PrV and are the natural host. Other secondary hosts include cattle, sheep, dogs and cats (Pensaert and Kluge, 1989). Laboratory animals such as rabbits and mice can be infected (Nash *et al.*, 1980).

Aujeszky's disease is characterized by inflammation of the respiratory tract and a severe encephalomyelitis (Crandell, 1982; Eich, 1983). The common route of PrV entry is the nasopharyngeal region. The virus is able to replicate in cells of nasal and pharyngeal mucosa (Pol et al., 1989). After the infection of the peripheral neural cells, the virus is transported to the central nervous system, where it causes a severe encephalitis which is fatal in young pigs (Baskerville et al., 1973; Dow and McFerran, 1962). Clinical signs of pseudorabies depend on the virulence of the virus strain, the infective dose and the age of the animal affected. Infection in older pigs results in respiratory and reproductive disorders. Infection with mild strains of the virus results in an inapparent infection (Kluge et al., 1992).

