

Research paper

Genetic determinants of freckle occurrence in the Spanish population: Towards ephelides prediction from human DNA samples

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Abstract

Prediction of human pigmentation traits, one of the most differentiable externally visible characteristics among individuals, from biological samples represents a useful tool in the field of forensic DNA phenotyping. In spite of freckling being a relatively common pigmentation characteristic in Europeans, little is known about the genetic basis of this largely genetically determined phenotype in southern European populations. In this work, we explored the predictive capacity of eight freckle and sunlight sensitivity-related genes in 458 individuals (266 non-freckled controls and 192 freckled cases) from Spain. Four loci were associated with freckling (*MC1R*, *IRF4*, *ASIP* and *BNC2*), and female sex was also found to be a predictive factor for having a freckling phenotype in our population. After identifying the most informative genetic variants responsible for human ephelides occurrence in our sample set, we developed a DNA-based freckle prediction model using a multivariate regression approach. Once developed, the capabilities of the prediction model were tested by a repeated 10-fold cross-validation approach. The proportion of correctly predicted individuals using the DNA-based freckle prediction model was 74.13%. The implementation of sex into the DNA-based freckle prediction model slightly improved the overall prediction accuracy by 2.19% (76.32%). Further evaluation of the newly-generated prediction model was performed by assessing the model's performance in a new cohort of 212 Spanish individuals, reaching a classification success rate of 74.61%. Validation of this prediction model may be carried out in larger populations, including samples from different European populations. Further research to validate and improve this newly-generated freckle prediction model will be needed before its forensic application. Together with DNA tests already validated for eye and hair colour prediction, this freckle prediction model may lead to a substantially more detailed physical description of unknown individuals from DNA found at the crime scene.

Keywords: Freckles; Ephelides; Externally visible traits; DNA-based prediction; Forensic science

1 Introduction

Identifying predictive biomarkers of human appearance traits is being systematically investigated by the forensic community with the purpose of individual identification merely from a biological sample [1]. Human pigmentation traits are some of the most differentiable externally visible characteristics among individuals. For this reason, researchers have been focused in the design of genetic prediction tests for eye, skin and hair colour variation [2-7]. However, DNA-based prediction of other human pigmentation traits under a strong genetic control, such as ephelides occurrence, has not been generated yet.

Ephelides (also known as freckles) are small, flat, pale-brown spots commonly observed in fair-skinned and/or red-haired individuals. Ephelides typically appear early in childhood, may increase in size, number and intensity during adolescence and partly disappear during the young adulthood period [8]. Although the development of these hyperpigmented spots may be triggered by exposure to sunlight, the occurrence of ephelides is largely genetically

determined [9]. The melanocortin-1 receptor (*MC1R*) gene seems to be the major contributor to the formation of freckles in European-origin individuals, independent of skin type and hair colour [10,11]. From all non-synonymous allelic variants found in the *MC1R* gene, six have been traditionally associated with a more severe phenotype, characterised by fair skin, red hair and freckling (known as the RHC phenotype): D84E, R142H, R151C, I155T, R160W and D294H [11,12]. Functional analyses have demonstrated that these *MC1R* genetic variants severely affect receptor function reducing stimulation of the pigmentation pathway. These six variants as well as other rare alleles that completely hamper MC1R function are known as ‘R’ alleles. Alternatively, weaker variants with lower penetrance are classified as ‘r’ alleles, and other rare non-synonymous variants that do not seem to have a noticeable effect in receptor function are defined as pseudoalleles [13].

The presence of alleles with impaired function ultimately results in an increased synthesis of pheomelanin (instead of eumelanin) in melanocytes [13–15]. Nevertheless, it is thought that other genes must contribute to freckling, since a significant percentage of the individuals with freckles do not harbour mutations in the *MC1R* gene. Accordingly, other genes have also been associated with ephelides occurrence via genome-wide association studies, including *IRF4*, *ASIP*, *TYR* and *BNC2* [16–18].

In this work, we analysed the role of eight genes previously associated with sunlight sensitivity in an ephelides case-control study. As far as we know, this study tackles for the first time the genetic basis of freckles in a southern European population (Spain), where the freckling genotype presents a reduced frequency compared to northern Europe. After performing an association study, we developed a multivariate regression approach where only the most informative loci responsible for ephelides were included, in order to predict human ephelides occurrence. To test the power of the newly-generated freckle prediction model in future forensic applications, we evaluated the model’s prediction performance in terms of accuracy, sensitivity and specificity by means of a cross-validation approach as well as an external validation using an independent sample.

2 Materials and methods

2.1 Study subjects and data collection

2.1.1 Original population

A total of 458 individuals (266 non-freckled controls and 192 freckled cases) were included in this study. Initially, unrelated participants were randomly selected, the percentage of the freckling phenotype being 21.86% in our population. These participants were recruited among the students and staff of the Jaume I University of Castellon. With the aim of performing a case-control association study, additional consecutive freckled volunteers were included in our study population. All individuals were Europeans of Spanish origin. Written informed consent was provided by all participants, and the study was approved by the Ethics Committee of the Jaume I University of Castellon (Castellon, Spain).

Under the supervision of a professional, each participant completed a standardised questionnaire to collect information on sex, age, pigmentation traits, history of childhood sunburns, Fitzpatrick’s skin type classification, and sun exposure habits. Details of ephelides occurrence both during the infancy or adolescence periods and in adulthood were collected in the questionnaire (an illustration is shown in Fig. S1 in Supplementary material in online version at DOI: [10.1016/j.fsigen.2017.11.013](https://doi.org/10.1016/j.fsigen.2017.11.013)).

2.1.2 Independent validation population

A second phase of the study was composed of 212 unrelated individuals (109 non-freckled controls and 103 freckled cases) of Spanish origin, also recruited among the students and staff of the Jaume I University of Castellon. All individuals gave a written informed consent and completed the standardised questionnaire used to collect phenotypic information.

2.2 DNA extraction

Genomic DNA was obtained from buccal swabs that were stored at –20 °C until sample processing. DNA was isolated using QIAamp DNA Mini Kit (Qiagen, Hilden, Germany), according to the manufacturer’s protocol. After DNA extraction, all samples were diluted to a concentration of 5 ng/µl in order to prepare them for **DNA**PCR amplification.

2.3 SNP selection and genotyping

Previous literature was used to **performselect** our candidate gene list. We **selectedopted for** genes previously associated with sensitivity to sunlight and/or freckling [8,11,16–20]. Eventually, eight SNPs located in seven pigmentation-related genes were selected and genotyped: rs4911442 located in the *ASIP* gene [8,16,20], rs2153271 in the *BNC2* gene [16], rs12896399 in *SLC24A4* [18], rs16891982 in *SLC45A2* [19], rs1393350 and rs1042602 in the *TYR* gene [18], rs12203592 in *IRF4* [18–20], and rs12821256 in the *KITLG* gene [18]. Other candidate SNPs mentioned in the cited literature were excluded due to SNP redundancy or to extremely low frequencies in the Spanish population. SNP codes, locations, ancestral and derived alleles and their frequencies were obtained from the Ensembl Variation database (<http://www.ensembl.org/info/genome/variation/index.html>).

Genotyping assays were performed by using KASP Genotyping Chemistry (LGC, Hoddesdon, United Kingdom). For *SLC45A2* rs16891982, TaqMan technology was used (Applied Biosystems, Foster City, USA). Genotyping analyses were carried out in a StepOnePlus™ Real-Time PCR System, with varying PCR conditions depending on the requirements of each probe. The genotype of each sample was determined by measuring allele-specific

fluorescence, using SDS v2.3 software for allelic discrimination (Applied Biosystems, Foster City, USA). For quality control, we included a negative control and a trio of samples with known genotype (major allele homozygous, heterozygous and rare allele homozygous) in each 96-well plate.

2.4 Sequencing of *MC1R* coding region

All DNA samples were analysed for the coding sequence of the *MC1R* gene by direct automated DNA sequencing, as previously described [21]. The primers used to amplify the *MC1R* coding region were: MC1R-F (5'-CAGCACCATGAACTAAGCAGGACACCTG-3') and MC1R-R (5'-AAGGGTCCGCGCTTCAACACTTTCAGAG-3'). Amplification was carried out by using Type-it™ Microsatellite PCR Kit (Qiagen, Hilden, Germany), following the manufacturer's instructions. PCR products were purified with EnzSAP (EdgeBio, Gaithersburg, USA) and subsequently sequenced by direct gene sequencing with the Sanger method. A sample with known *MC1R* genotype per 96-well plate was added for quality control. All sequencing results were analysed using SeqScape v2.5 software to align and detect all nucleotide changes. All detected sequence changes were confirmed manually.

Non-synonymous *MC1R* mutations were then defined as 'R', 'r' or 'p' (pseudoallele) alleles according to their impact on protein function (Table 1). R alleles included genetic variants that have been associated with the red hair colour (RHC) phenotype [22,23], and have been shown to cause a significant impairment of receptor function in previous functional *in vitro* or *in silico* analysis [21,24-29]. Genetic variants that have not been associated with the RHC phenotype [23,30,31], and have been shown to display partial loss of function or present a possibly damaging effect based on prediction analysis [25,27,32,33], were defined as r alleles. Variants in which receptor function is similar to the wild-type form were catalogued as p alleles. Individuals were classified based on the number of R and/or r alleles carried.

Table 1 *MC1R* variants with protein sequence alterations identified in the current study.

alt-text: Table 1

Variant	Nucleotide change	Functional analysis	Prediction analysis	Polyphen score	Reference
R alleles					
D84E	c.252C > A	Yes	-	-	Beaumont et al. [23]
R142H	c.425G > A	Yes	-	-	Beaumont et al. [23]
R151C	c.451C > T	Yes	-	-	Beaumont et al. [22]
I155T	c.464T > C	Yes	-	-	Beaumont et al. [23]
R160W	c.478C > T	Yes	-	-	Beaumont et al. [22]
D294H	c.880G > C	Yes	-	-	Beaumont et al. (2006) [22]
Q30X	c.88C > T	No	No	-	Guan et al. [24]
C35Y	c.104G > A	Yes	-	-	Fargnoli et al. [32] and Fernandez et al. [29]
S41F	c.122C > T	Yes	-	-	Pérez Oliva et al. [28]
S83P	c.247T > C	No	Yes	-	Kanetsky et al. [26] and Ibarrola-Villava et al. (2013) [36]
S83L	c.248C > T	Yes	-	-	Ozola et al. [27]
G89R	c.265G > C	Yes	-	-	Ozola et al. [27]
M128T	c.383T > C	Yes	-	-	Pérez Oliva et al. [28]
L135R	c.404T > G	No	No	1.000	NEW. Never reported/not found in SNPs databases
Y152X	c.456C > A	Yes	-	-	Fargnoli et al. [32]
Q233X	c.697C > T	No	No	-	Martinez-Cadenas et al. [35]
P256S	c.766C > T	No	Yes	-	Hu et al. (2014) Kanetsky et al. [26]

P268R	c.803C > G	No	Yes	-	Ibarrola-Villava et al. [25]
r alleles					
V60L	c.178G > T	Yes	-	-	Beaumont et al. [23] and Herraiz et al. [30]
V92M	c.274G > A	Yes	-	-	Beaumont et al. [23] and Herraiz et al. [30]
R163Q	c.488G > A	Yes	-	-	Beaumont et al. [23] and Nakayama et al. [31]
L24M	c.70C > A	No	No	0.863	NECTAR (http://nectarmutation.org)
F45L	c.133T > C	No	Yes	-	Ozola et al. [27] and Ibarrola-Villava et al. [25]
L46V	c.136C > G	No	No	0.741	Ensembl (www.ensembl.org)
R67Q	c.200G > A	No	Yes	-	Fargnoli et al. [32]
T95M	c.284C > T	Yes	-	-	Ozola et al. [27]
I120T	c.359T > C	No	Yes	-	Fargnoli et al. [32]
V122M	c.364G > A	Yes	-	-	Jimenez-Cervantes et al. [33]
V156A	c.467T > C	No	No	0.784	Kanetsky et al. [26]
V193L	c.577G > T	No	No	0.567	Ensembl (www.ensembl.org)
N279K	c.837C > A	No	Yes	-	Fargnoli et al. [32] and Ibarrola-Villava et al. [25]
p alleles					
G32R	c.94G > A	No	No	0.299	NEW. Never reported/not found in SNPs databases
S47T	c.140G > C	No	No	0.003	Gan-Or et al. (2016) Garcia-Borron et al. [57]
A57V	c.170C > T	No	No	0.001	Ensembl (www.ensembl.org)
G89E	c.265G > A	No	No	0.170	Ensembl (www.ensembl.org)
M128V	c.382A > G	No	No	0.019	NECTAR (http://nectarmutation.org)
A167T	c.499G > A	No	No	0.024	Ensembl (www.ensembl.org)
V208I	c.622G > A	No	No	0.002	Ensembl (www.ensembl.org)

Only variants with protein sequence alterations are shown. Previous in vitro or in silico functional analysis were used to classify *MC1R* variants into three categories. In the absence of previous published data, impact on receptor function was predicted by the PolyPhen programme. Genetic variants with PolyPhen scores lower than 0.50 were considered as p alleles (pseudoalleles with similar function compared to wild-type), while variants with scores from 0.50 to 0.95 were classified as r alleles (possibly damaging).

2.5 Association analysis

Association analyses were performed using the R software (<http://www.R-project.org>). All analyses performed were two-sided, and a significance level of 0.05 was considered for rejection of the null-hypothesis. The conservative Bonferroni correction was used to adjust the significance level for multiple testing ($P\text{-value} < 4.54 \times 10^{-3} = 0.05/11$). Unknown and missing values were excluded at each specific analysis.

For each polymorphism studied, Fisher's exact test was used to check for deviations from Hardy-Weinberg equilibrium (HWE) among controls. Minor allele frequencies (MAFs) for freckled and non-freckled individuals were estimated from our population.

Associations between the genotyped polymorphisms and the presence of ephelides were assessed according to the co-dominant model of inheritance via binary logistic regression. Genotype-related odds ratios (ORs), their corresponding 95% confidence intervals (CIs) and associated *P*-values were estimated. The proportion of the total variance in freckling explained by each genetic variant was estimated using Nagelkerke pseudo- R^2 statistic (R^2). This statistic parameter was used to rank the genetic variants included in the study based on their contribution to the freckling phenotype.

2.6 Prediction model

A multivariate logistic regression approach was applied to build the prediction models. The DNA-based freckle prediction model was developed using backward elimination of genetic variants based on the Akaike information criterion (AIC), being the optimal model the one with the smaller AIC value - best balance between goodness-of-fit and parsimony. For each iteration, the lowest predictor in the variable set is excluded. Then, the model is rebuilt, used to predict again freckle occurrence, and the quality of the newly generated model is re-tested. This model-building process is repeated until all remaining polymorphisms included in the prediction model have a statistically significant contribution, and the estimated information loss is minimised.

To assess the influence of sex in the freckling phenotype, the DNA-based prediction model was additionally adjusted by including sex, as well as the interaction between sex and each genetic variant as covariates in the multivariate logistic regression. As above, the optimal sex-adjusted prediction model was developed based on the AIC variable selection criteria.

Finally, statistical interactions between genetic variants were examined using the MDR software (<http://www.epistasis.org/>). The multifactor dimensionality reduction method is a powerful strategy for detecting and interpreting statistical locus-locus epistasis [34]. Dendrogram interaction graphs provided by MDR illustrate the presence, strength, and nature of epistatic effects. Pairwise genetic interactions were tested by including interactions into the logistic regression model.

2.7 Model evaluation

2.7.1 Internal evaluation phase - cross-validation

In order to evaluate the predictive ability of the freckle prediction models, we applied a repeated 10-fold cross-validation approach. Briefly, individuals were randomly divided into 10 equal data subsets. For each round of cross-validation, nine data subsets were used for performing the analysis (training set) while the remaining subset was used for validating the analysis (testing set). This process was run 10 times, using a different subset as testing set each time. This entire procedure was repeated 200 times, using different random partitions of the original sample to protect against chance divisions of the dataset. Data partition was carried out using the 'caret package' for R software, and the repeated cross-validation approach was performed with custom programmes written in R.

Receiver operating characteristic (ROC) curve analysis was adopted to evaluate the cross-validated performance of the prediction models. The Youden Index method was used to set the optimal cut-off point - the point on the curve at which (sensitivity + specificity-1) is maximised. For each prediction model, basic prediction accuracy parameters - including the area under the ROC curve (AUC), sensitivity, specificity and accuracy - were calculated from the confusion table reporting the number of true positives (TP), true negatives (TN), false positives (FP), and false negatives (FN). The final values reported were averaged from the 200 × 10 individual cross-validation curves to produce a single estimation. All calculations and plots were performed using the 'ROCR package' for R software.

2.7.2 External evaluation phase - independent population

In order to carry out the external evaluation, only the genetic variants that were significantly associated with freckles in the first phase of the study - *IRF4*, *ASIP*, *BNC2* and *MC1R* - were analysed in the newly collected independent samples.

This independent dataset was used to assess the performance of the newly-generated freckle prediction model regarding the correct classification of individuals as freckled or non-freckled. Subsequently, a comparison between predicted and observed data was performed, and the basic accuracy parameters (AUC, sensitivity, specificity and accuracy) were calculated from the confusion table by using the 'pROC package' for R software.

3 Results

Our original sample set comprised 266 non-freckled and 192 freckled individuals of Spanish ancestry (Table 2). The study cohort included 260 females (56.52%) and 198 males (43.04%). Interestingly, the prevalence of the freckling phenotype was remarkably higher in females than in males (68.75% vs 31.25%, *P*-value = 9.00×10^{-9}). As expected, freckled individuals were more likely to have skin phototypes I-II and childhood sunburns than non-freckled individuals, although no significant differences between them were found regarding sun exposure habits (Table 2).

Table 2 Classification of the Spanish individuals according to sex, skin phototype and sun exposure habits.

			N	%	MAF	HWE <i>P</i> -value	N	%	MAF				
<i>ASIP</i>	rs4911442	AA	233	87.59	0.062	0.264	153	79.69	0.109	0.044	reference	1.01	4
		AG	29	10.90			36	18.75			1.89 (1.11–3.21)		
		GG	2	0.75			3	1.56			2.28 (0.38–13.83)		
		ND	2	0.75			0	0.00					
<i>BNC2</i>	rs2153271	TT	68	25.56	0.476	0.222	61	31.77	0.406	0.067	reference	0.87	5
		CT	143	53.76			106	55.21			0.83 (0.54–1.27)		
		CC	55	20.68			25	13.02			0.51 (0.28–0.91)		
		ND	0	0.00			0	0.00					
<i>IRF4</i>	rs12203592	CC	207	77.82	0.117	1.000	100	52.08	0.275	1.81E–05*	reference	5.92	2
		CT	56	21.05			77	40.10			2.49 (1.57–3.94)		
		TT	3	1.13			14	7.29			6.56 (1.68–25.54)		
		ND	0	0.00			1	0.52					
<i>KITLG</i>	rs12821256	TT	243	91.35	0.041	1.000	175	91.15	0.034	0.417	reference	0.11	9
		CT	21	7.89			11	5.73			0.73 (0.34–1.56)		
		CC	0	0.00			0	0.00			–		
		ND	2	0.75			6	3.13					
<i>MC1R</i>	R alleles	0/0	214	80.45	–	–	110	57.29	–	7.05E–08*	reference	6.91	1
		0/1	38	14.29			64	33.33			3.26 (2.05–5.18)		
		1/1	2	0.75			13	6.77			11.61 (2.55–52.83)		
		ND	12	4.51			5	2.60					
<i>MC1R</i>	r alleles	0/0	154	57.89	–	–	92	47.92	–	0.039	reference	1.09	3
		0/1	86	32.33			77	40.10			1.49 (1.01–2.24)		
		1/1	14	5.26			18	9.38			2.15 (1.02–4.53)		
		ND	12	4.51			5	2.60					
<i>SLC24A4</i>	rs12896399	TT	92	34.59	0.439	0.033	58	30.21	0.479	0.521	reference	0.21	7
		GT	113	42.48			85	44.27			1.20 (0.77–1.85)		
		GG	60	22.56			49	25.52			1.33 (0.80–2.19)		
		ND	1	0.38			0	0.00					
<i>SLC45A2</i>	rs16891982	GG	209	78.57	0.133	0.002	154	80.21	0.130	0.690	reference	0.12	8
		CG	43	16.17			26	13.54			0.82 (0.48–1.39)		
		CC	14	5.26			12	6.25			1.16 (0.52–2.59)		

		ND	0	0.00			0	0.00					
<i>TYR</i>	rs1393350	GG	75	28.30	0.239	1.000	58	30.21	0.259	0.777	reference	0.08	10
		AG	134	50.57			102	53.13			1.10 (0.75–1.63)		
		AA	56	21.13			32	16.67			1.27 (0.58–2.78)		
		ND	1	0.38			0	0.00					
<i>TYR</i>	rs1042602	CC	154	57.89	0.464	0.902	105	54.97	0.432	0.485	reference	0.23	6
		AC	97	36.47			73	38.22			0.98 (0.64–1.51)		
		AA	15	5.64			13	6.81			0.74 (0.42–1.28)		
		ND	0	0.00			1	0.52					

SNP, single nucleotide polymorphism; N, number of individuals; %, percentage of individuals per group among the total; OR, odds ratio per minor allele; CI, confidence interval; R^2 , Nagelkerke pseudo- R^2 statistic.

* Indicates significant results at Bonferroni threshold of 4.50E-03.

^a P -value for the binary logistic regression association analysis according to the co-dominant model of inheritance. Bold indicates statistically significant results.

^b The proportion of total variance in freckling phenotype explained by the genetic variants was estimated using R^2 .

^c The genetic variants were ranked according to their importance in freckling phenotype.

Univariate association analyses were performed to assess the independent effects of each genetic variant on freckling occurrence in childhood (Table 3). R variants of the *MC1R* gene were the alleles most strongly associated with freckling (P -value = 7.05×10^{-8}), explaining 6.91% of the variance. The rs12203592 polymorphism in the *IRF4* gene presented the second strongest association with the presence of ephelides (P -value = 1.81×10^{-5}), explaining 5.92% of the variance. Weaker significant associations were also observed for r variants (P -value = 0.039) and rs4911442 in *ASIP* (P -value = 0.044), explaining around 1% of the variance ($R^2 = 1.09\%$ and $R^2 = 1.01\%$, respectively). No associations were found for the remaining six polymorphisms, although rs2153271 in *BNC2* showed a marginal significant association with freckling (P -value = 0.067).

A prediction model was then constructed based on multinomial logistic regression. To design the optimal model, we performed a step-wise analysis by iteratively excluding the lowest predictor from the multinomial logistic regression model. The genetic variants were ranked according to their impact on freckling. The DNA-based prediction model included four loci (*MC1R*, *IRF4*, *ASIP* and *BNC2*), which together explained about 30% of the freckling phenotype variance in our population (Table 4). A repeated 10-fold cross-validation approach was used to perform an internal validation of the DNA-based prediction model by ROC analysis, which tests the power of the prediction model [38]. The prediction accuracy of the tested DNA-based prediction model was 74.13% in the population, with an AUC of 0.771, a specificity of 82.00%, and a sensitivity of 63.51% (Fig. 1A).

Table 4 Multivariate logistic regression testing freckles association with genetic variants and sex.

alt-text: Table 4

Gene	SNP rs# No.	Variable	DNA-based prediction model		Sex-adjusted prediction model	
			OR (95% CI)	P -value ^a	OR (95% CI)	P -value ^a
<i>MC1R</i>	R alleles	1/0	4.18 (2.49–7.02)	6.66E-08	4.21 (2.48–7.15)	1.02E-07
		1/1	21.51 (4.50–102.79)	1.20E-04	21.94 (2.48–105.16)	1.12E-04
<i>IRF4</i>	rs12203592	CT	3.28 (2.05–5.26)	7.75E-07	3.26 (2.02–5.27)	1.43E-06
		TT	15.51 (3.26–73.76)	5.65E-04	14.63 (3.06–70.10)	7.81E-04
<i>MC1R</i>	r alleles	1/0	2.08 (1.30–3.32)	2.20E-03	2.05 (1.28–3.30)	2.92E-03
		1/1	3.45 (1.51–7.87)	3.20E-03	3.39 (1.43–7.99)	5.35E-03

<i>ASIP</i>	rs4911442	AG	2.26 (1.22–4.17)	9.27E-03	2.38 (1.28–4.45)	6.49E-03
		GG	3.65 (0.35–38.40)	0.281	5.15 (0.47–56.62)	0.181
<i>BNC2</i>	rs2153271	CT	0.91 (0.55–1.49)	0.706	0.98 (0.59–1.63)	0.937
		CC	0.45 (0.23–0.90)	0.023	0.48 (0.24–0.97)	0.040
		Male sex	–	–	0.49 (0.32–0.79)	2.70E-03

SNP, single nucleotide polymorphism; OR, odds ratio per minor allele; CI, confidence interval.

Optimal model selection procedure was based on the Akaike information criterion (AIC).

^a *P*-value for the multivariate logistic regression association analysis. Bold indicates significant results at Bonferroni threshold of 4.50E-03.

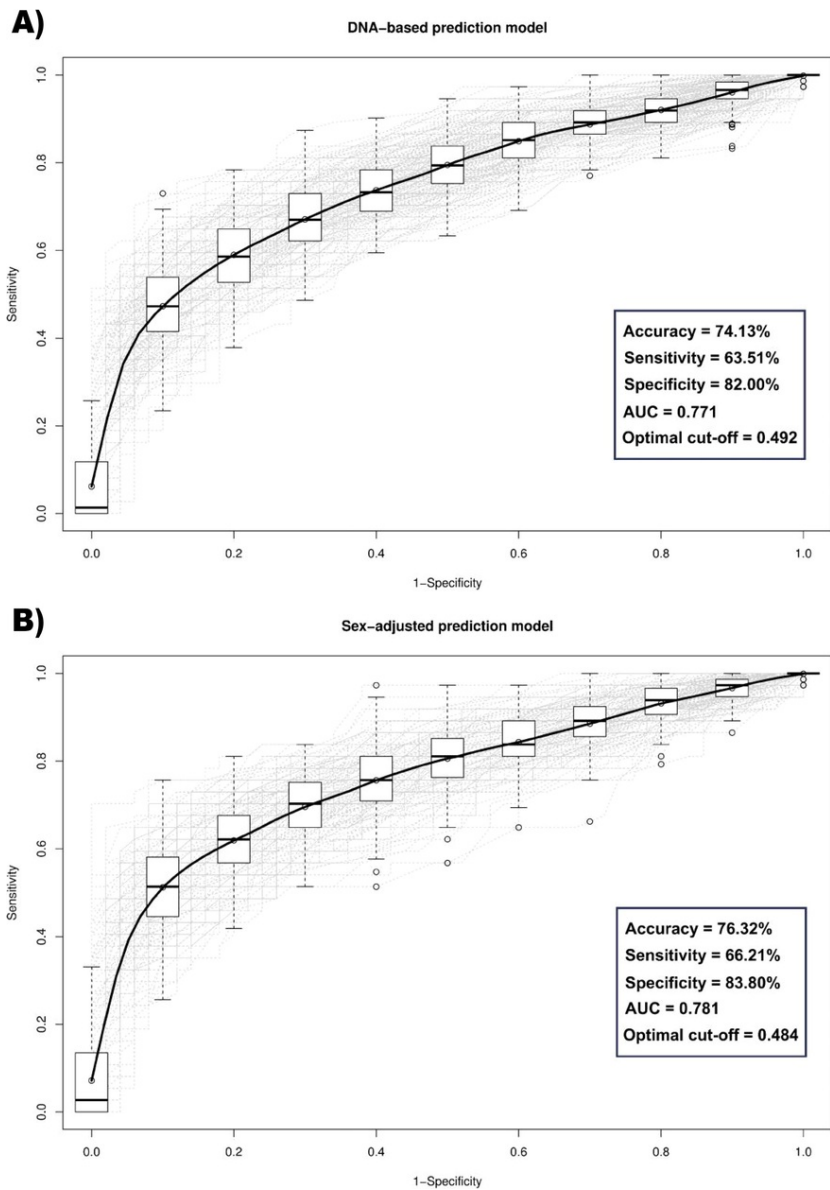


Fig. 1 Average receiver operating characteristic curves for repeated cross-validated model assessment of the A) DNA-based; and B) sex-adjusted freckle prediction models. Prediction models were developed with a Spanish population set using multivariate logistic regression. Repeated 10-fold cross-validation was used to assess the performance of the prediction models. Faint grey lines represent ROC curves for each of the individual cross-validation curves. Solid black line represents the overall average of all ROC curves. Mean values of the basic accuracy parameters are displayed. Optimal cut-off was selected for maximising Youden Index. AUC, area under the curve.

alt-text: Fig. 1

The inclusion of sex on the freckle prediction model appeared to increase the prediction accuracy by 2.19%, to a total of 76.32% (Fig. 1B). Slight increases in AUC (0.781), specificity (83.80%) and sensitivity (66.21%) values were observed when sex was taken into account. The results of the multivariate association analysis confirmed sex as an important variable in the estimation of the freckling phenotype (P -value = 2.70×10^{-3}). No significant

interaction between sex and other genetic predictors was found.

An external validation of the newly-generated prediction model, assessing the model generalisability [39], was also performed by collecting an independent cohort of 212 individuals - though only 193 individuals successfully genotyped for all loci were included in the external validation. The prediction accuracy of the freckle prediction model was 74.61% in the independent cohort, with an AUC of 0.809, a specificity of 89.12%, and a sensitivity of 59.79%. A total of 49 individuals were incorrectly predicted (misclassification rate of 25.39%) (Table 5).

Table 5 Accuracy of the sex-adjusted freckle prediction model obtained from an independent cohort of Spanish individuals.

alt-text: Table 5

Prediction model	Phenotype	Predicted phenotype		Total ^b	Prediction accuracy parameters			
		Freckled	Non-freckled		AUC	Sensitivity (%)	Specificity (%)	Accuracy (%)
Sex-based prediction model ^a	Freckled	55	37	92	0.809	59.79	89.12	74.61
	Non-freckled	12	89	101				
	Total	67	126	193				
	Fails (%)	12 (17.91)	37 (29.36)	49 (25.39)				

^a Optimal cut-off maximising Youden Index (0.6138) was used to classify individuals.

^b Genetic analyses of four loci were successfully performed in 193 out of 212 individuals (missing rate of 8.96%).

We also evaluated the prediction capacity of the sex-adjusted prediction model to determine the freckling phenotype in adulthood. Taking into account that freckles often disappear after adolescence or during young adulthood, the success rate of the prediction model was relatively good in our population (69.63%), with a high true negative rate (specificity of 80.70%) but a low true positive rate (sensitivity of 58.91%) (Fig. 2). Out of all six freckle predictors included in the model, only the r variants in *MC1R* did not show a significant contribution to freckling in adulthood (P -value > 0.05).

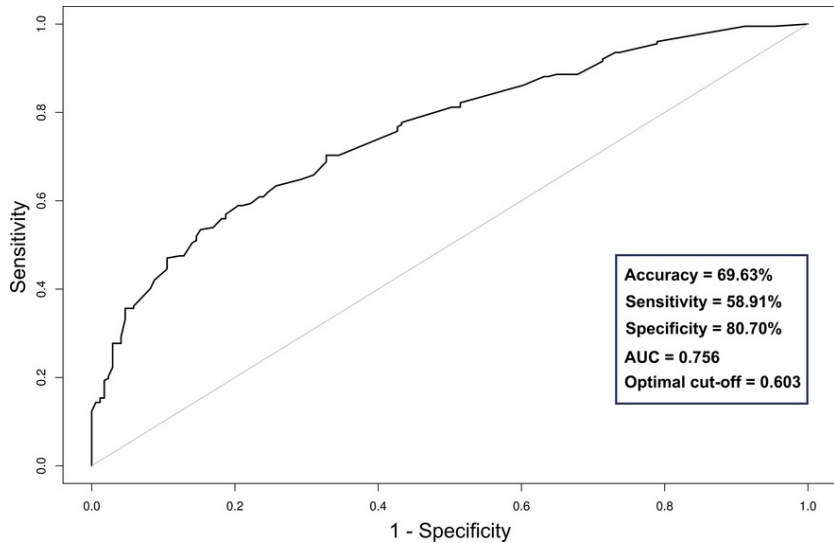


Fig. 2 Area under the receiver operating characteristic curves for the sex-adjusted freckle prediction model in adulthood. Individuals from both the first (model building and internal validation) and the second phase (external validation) of the study were pooled to perform this analysis. Using Youden Index, optimal cut-off was selected for maximising the sensitivity and specificity of the prediction model to classify individuals. AUC, area under the curve.

alt-text: Fig. 2

MDR analysis for locus-locus interaction detection indicated that the best freckle prediction model was composed by four genetic variables and sex (*BNC2* rs2153271 was not included). The locus-locus interaction analysis showed a redundant interaction between R variants in the *MC1R* gene and rs12203592 in the *IRF4* gene, denoted by the blue lines connecting these two genetic determinants in the dendrogram (Fig. S2 in Supplementary material in online version at DOI: [10.1016/j.fsigen.2017.11.013](https://doi.org/10.1016/j.fsigen.2017.11.013)). However, the inclusion of this genetic interaction in our freckle prediction model did not significantly modify the prediction capacity using a repeated 10-fold cross-validation approach (AUC = 0.768, specificity = 83.20%, sensitivity = 61.27%, and accuracy = 75.45%).

4 Discussion

In the current study, eight freckle and sunlight sensitivity-related genes were genotyped in 458 individuals from Spain, with the intention of analysing their putative implication in the appearance of ephelides, and its potential for future forensic applications. However, only the 438 individuals successfully genotyped for all loci were included in the development and cross-validation of the prediction model. Furthermore, an external evaluation of the freckle prediction model was performed on an extra 212 sample independent dataset by genotyping the genetic variants associated with freckling in the original population.

In the last few years, the genetic basis of ephelides has been adequately studied in populations of North European origin [11,16-18]. However, little is known about the genetic determinants of this pigmentation characteristic in southern European populations. Our results showed an association between ephelides and genetic variants in the *IRF4*, *MC1R* and *ASIP* genes in a Mediterranean population, confirming previous studies performed in North European populations [11,17,18]. R variants of the *MC1R* gene have been previously acknowledged as the most relevant locus associated with both freckling and sun sensitivity. The abnormal function of this receptor leads to a higher pheomelanin to eumelanin ratio, commonly resulting in the well-known freckle-generating RHC phenotype. *IRF4* has also been associated with freckling in several studies [8,18]. This interferon regulatory factor cooperates with MITF to activate the expression of tyrosinase in melanocytes, a function which seems to be impaired in carriers of the rs12203592*T derived allele [40]. The other main freckle-associated locus in this study, *ASIP*, antagonises the activation of MC1R, leading to a down-regulation of eumelanogenesis and an up-regulation of pheomelanogenesis. Variants in this gene, also previously linked to the RHC phenotype, may therefore have effects similar to variants of the *MC1R* gene [16,17].

The impact of *BNC2* rs2153271 on freckling was detected only after applying a multivariate association approach. The identification of the association between freckling and rs2153271, located in an intron of the *BNC2* gene, was recently discovered in a GWAS study performed in a population of North European ancestry [16], and was additionally correlated with acquired facial pigmented spots during aging [19].

Although rs1042602 and rs1393350, in the *TYR* gene, have been previously associated with freckling in two European populations of Icelandic and Dutch origin [18], a lack of association between these two SNPs and ephelides occurrence was observed in our Spanish population. Accordingly, no association with freckling was previously observed for a *TYR* melanoma-associated genetic variant (rs1126809) in a sample set comprising three Mediterranean populations from France, Spain and Italy [36]. However, that melanoma case-control study identified a moderate association between rs16891982 in *SLC45A2* and the absence of ephelides [36]. The inheritance of a set of genetic variants in *SLC45A2* was also associated with freckling in a population from Brazil [41]. However, we are not able to verify this association in the current study, perhaps due to the limited number of individuals included in the sample set.

This association study allows us to develop a freckle prediction model based on five genetic predictors: R variants in *MC1R*, *IRF4* rs12203592, r variants in *MC1R*, *ASIP* rs4911442 and *BNC2* rs2153271 (listed in order, from greater to lower genetic contribution). Predicting externally visible human traits from genotypes represents a potential valuable tool in forensic investigations [1]. However, DNA-based phenotyping needs a very demanding approach since externally visible characteristics are typically influenced by several genetic as well as environmental factors [42]. To date, DNA-based human eye colour prediction is the most accurate, advanced and applied test in forensic applications [3-5,43]. The IrisPlex system, a robust prediction model based on six eye colour SNP predictors, has been validated in numerous studies for forensic eye colour prediction [4,5,43]. Recently, a multiplex genotyping assay, called HIrisPlex, has been developed for simultaneous hair and eye colour prediction [6], being suitable and sufficiently sensitive for forensic use approval [44]. Currently, forensic genome-wide association studies are focused on increasing the genetic determinants of different phenotypes such as body height, hair shape, baldness, and facial variation, in order to reach more detailed predictions of an unknown person's appearance from the analysis of his/her DNA [16,45-48].

As a whole, the DNA-based freckle prediction model presents with high specificity (82.00%), but low sensitivity (63.51%). The low sensitivity levels achieved could be due to the fact that one third of the individuals carrying R variants did not display freckles, suggesting that other genetic determinants are also important in the appearance of freckles. Hopefully, future studies with increased sample sizes may add to the genetic factors influencing freckle occurrence, so that the model's prediction potential may be significantly improved.

Interestingly, we found a much higher prevalence of ephelides in females than in males. Previous studies have also stated discrepancies in human pigmentation and sunlight sensitivity traits between sexes [19,35,49-52]. Notably, higher freckle prevalence in females was previously observed in a GWAS study performed in a northern European population [18]. Additionally, Jacobs and cols. (2015) showed that females presented a much higher occurrence of facial sun spots than males, being the total variance explained by sex higher than any of the genetic variants studied [19].

The inclusion of sex in the IrisPlex model has been proposed to improve prediction accuracy mainly for intermediate eye colours [35,50,53,54]. Adding sex as a covariate in our DNA-based freckle prediction model slightly increased the prediction performance for ephelides prevalence. In particular, it presented with both higher specificity (83.80%) and sensitivity (66.21%) if compared to the prediction model based on the five freckle genetic predictors alone. Overall, this sex-adjusted freckle prediction model could explain a high proportion of the freckling phenotypic variance ($R^2 = 32.92\%$), which is quite large compared to other human complex traits [55].

The main purpose of a prediction model is to provide correct classification for new individuals, being therefore external validation a crucial phase of the model development process. Assessing both reproducibility (internal validation) and generalisability (external validation) of the developed prediction model are necessary in order to avoid overfitted models [56]. However, application of a prediction model should only be considered after proving adequate accuracies in an independent dataset. For this reason, the discrimination ability of the newly-generated prediction model was assessed by comparing the observed and predicted outcomes in a new dataset. As usually noted in most external validations, a slight decrease of the overall prediction accuracy was observed (from 76.32% to 74.61%).

In terms of prediction power, our freckle prediction model provided a similar accuracy than the one obtained for green/hazel eye colour prediction using IrisPlex (AUC = 0.809 and AUC = 0.76, respectively), although significantly lower than the accuracy level achieved for brown (AUC = 0.93) and blue eye colour prediction (AUC = 0.91) [5]. HIRISplex also presented equivalent prediction power for hair colour prediction, with an average accuracy of 73% [44], compared to 74.61% of our freckle prediction model. Notice that our results have been reached after a validation approach with a relatively small independent sample, while both IrisPlex and HIRISplex prediction models have been validated in independent large population samples.

After our preliminary validation, the freckle prediction model developed in this study has shown to display limited success, but it also appears promising for future applications in forensics. However, further research is needed to increase the correct prediction rate (sensitivity) of the model, since the percentage of freckled individuals predicted as non-freckled is considerably high (false negative rate of 40.21%).

For practical considerations, knowing that ephelides tend to disappear during aging due to undetermined reasons, we tested the accuracy of our prediction model to determinate the presence of ephelides in adulthood. The model presented an AUC of 0.756, a slightly lower value compared to the AUC value obtained for freckle prediction in childhood (0.781 for internal cross-validation and 0.809 for external validation). No significant contribution to freckling prediction in adulthood was noted for r variants, suggesting that other unknown predictors could have an independent additional impact on the freckling phenotype in adulthood. Interestingly, after adjustment by *IRF4* rs12203592 (the most associated genetic predictor for freckling in adulthood), the minor allele of *SLC45A2* rs16891982 was weakly associated with the absence of freckles in adulthood (OR = 0.64 (0.43-0.94), *P*-value = 0.023). As revealed by the MDR analysis, the effect of this *SLC45A2* variant may not be strongly influenced by a locus-locus interaction, since there is no interaction between *SLC45A2* and the rest of the freckle loci (Fig. S2B in Supplementary material in online version at DOI: [10.1016/j.fsigen.2017.11.013](https://doi.org/10.1016/j.fsigen.2017.11.013)).

Since the sample set used to build and evaluate this freckle prediction model was relatively small, the ability to accurately predict the presence of ephelides may be further improved by using larger studies. It is likely that phenotype prediction accuracy may be substantially improved by including undiscovered genetic variants at novel loci or by taking into account possible gene interactions affecting the freckling phenotype. Also, DNA samples from different populations of European origin should be included in further studies in order to validate this DNA predictive test for future forensic applications.

It is also important to note that predicting complex phenotypic traits from DNA studies remains a difficult task even if all genetic loci involved - and the interactions among them - are taken into account, since different environmental factors may always have a considerable effect (for example, the effect of UV exposure on freckle occurrence).

5 Conclusions

The main genetic variants involved in freckle appearance in the Spanish population are located in the *MC1R* and *IRF4* genes, with minor contributions from *ASIP*, *BNC2* and perhaps other as yet unknown freckle genes. However, the influence on freckling of different *MC1R* variants (R or r) is substantially different. As a result, the DNA-based model for freckle prediction developed in this work considers five genetic determinants: R variants of the *MC1R* gene, *IRF4* rs12203592, r variants of the *MC1R* gene, *ASIP* rs4911442 and *BNC2* rs2153271 - in order of greatest to lowest contribution - reaching a cross-validated prediction accuracy of up to 74.13%, a more than respectable percentage. When sex is added to the model, the cross-validated prediction accuracy reached is even higher, growing up to 76.32%.

Furthermore, the newly-generated freckle model was tested in an independent cohort reaching an acceptable specificity level of 89.12%. However, the sensitivity of the model is certainly improvable, attaining a percentage of around 60%.

For a more detailed pigmentation phenotype prediction, future research may focus on designing multiplex genetic analysis for simultaneously predicting different externally visible traits. Perhaps the inclusion of two more SNPs in the HIRISplex system, *BNC2* rs2153271 and *ASIP* rs4911442, could increase the forensic potential of this DNA-based prediction test to include freckle occurrence (as well as the current eye and hair colour prediction).

Acknowledgements

We are extremely grateful to all the volunteers for giving their consent to take part in this study, as well as to all the medical specialists for supervising phenotype collection of all samples. We also thank Rafael Velasco for providing us with the freckle photographs.

This work was supported by grant number [GV/2016/156](#) from the [Education Council of the Generalitat Valenciana](#), and co-funded by the [Jaume I University of Castellon](#). BH is funded by the Jaume I University of Castellon under a Predoctoral Research contract (No. 15721).

References

- [1]** M. Kayser, Forensic DNA phenotyping: predicting human appearance from crime scene material for investigative purposes, *Forensic Sci. Int. Genet.* **18**, 2015, 33–48, <https://doi.org/10.1016/j.fsigen.2015.02.003>.
- [2]** O. Maroñas, J. Söchtig, Y. Ruiz, C. Phillips, Á. Carracedo and M.V. Lareu, The genetics of skin, hair, and eye color variation and its relevance to forensic pigmentation predictive tests, *Forensic Sci. Rev.* **27**, 2015, 13–40.
- [3]** Y. Ruiz, C. Phillips, A. Gomez-Tato, J. Alvarez-Dios, M. Casares de Cal, R. Cruz, O. Maroñas, J. Söchtig, M. Fondevila, M.J. Rodriguez-Cid, A. Carracedo and M.V. Lareu, Further development of forensic eye color predictive tests *Forensic Sci. Int. Genet.* **7**, 2013, 28–40, <https://doi.org/10.1016/j.fsigen.2012.05.009>.
- [4]** S. Walsh, F. Liu, K.N. Ballantyne, M. van Oven, O. Lao and M. Kayser, IrisPlex: a sensitive DNA tool for accurate prediction of blue and brown eye colour in the absence of ancestry information, *Forensic Sci. Int. Genet.* **5**, 2011, 170–180, <https://doi.org/10.1016/j.fsigen.2010.02.004>.
- [5]** F. Liu, K. van Duijn, J.R. Vingerling, A. Hofman, A.G. Uitterlinden, A.C.J.W. Janssens and M. Kayser, Eye color and the prediction of complex phenotypes from genotypes, *Curr. Biol.* **19**, 2009, R192–193, <https://doi.org/10.1016/j.cub.2009.01.027>.
- [6]** S. Walsh, F. Liu, A. Wollstein, L. Kovatsi, A. Ralf, A. Kosiniak-Kamysz, W. Branicki and M. Kayser, The HIrisPlex system for simultaneous prediction of hair and eye colour from DNA, *Forensic Sci. Int. Genet.* **7**, 2013, 98–115, <https://doi.org/10.1016/j.fsigen.2012.07.005>.
- [7]** A. Pneuman, Z.M. Budimlija, T. Caragine, M. Prinz and E. Wurmbach, Verification of eye and skin color predictors in various populations, *Leg. Med. Tokyo Jpn.* **14**, 2012, 78–83, <https://doi.org/10.1016/j.legalmed.2011.12.005>.
- [8]** C. Praetorius, R.A. Sturm and E. Steingrimsson, Sun-induced freckling: ephelides and solar lentigines, *Pigment Cell Melanoma Res.* **27**, 2014, 339–350, <https://doi.org/10.1111/pcmr.12232>.
- [9]** V. Bataille, H. Snieder, A.J. MacGregor, P. Sasieni and T.D. Spector, Genetics of risk factors for melanoma: an adult twin study of nevi and freckles, *J. Natl. Cancer Inst.* **92**, 2000, 457–463.
- [10]** J.L. Rees, The genetics of sun sensitivity in humans, *Am. J. Hum. Genet.* **75**, 2004, 739–751, <https://doi.org/10.1086/425285>.
- [11]** M. Bastiaens, J. ter Huurne, N. Gruis, W. Bergman, R. Westendorp, B.J. Vermeer and J.N. Bouwes Bavinck, The melanocortin-1-receptor gene is the major freckle gene, *Hum. Mol. Genet.* **10**, 2001, 1701–1708.
- [12]** P. Valverde, E. Healy, I. Jackson, J.L. Rees and A.J. Thody, Variants of the melanocyte-stimulating hormone receptor gene are associated with red hair and fair skin in humans, *Nat. Genet.* **11**, 1995, 328–330, <https://doi.org/10.1038/ng1195-328>.
- [13]** J.C. García-Borrón, B.L. Sánchez-Laorden and C. Jiménez-Cervantes, Melanocortin-1 receptor structure and functional regulation, *Pigment Cell Res.* **18**, 2005, 393–410, <https://doi.org/10.1111/j.1600-0749.2005.00278.x>
- [14]** J.V. Schaffer and J.L. Bolognia, The melanocortin-1 receptor: red hair and beyond, *Arch. Dermatol.* **137**, 2001, 1477–1485.
- [15]** R.A. Sturm, R.D. Teasdale and N.F. Box, Human pigmentation genes: identification, structure and consequences of polymorphic variation, *Gene* **277**, 2001, 49–62.
- [16]** N. Eriksson, J.M. Macpherson, J.Y. Tung, L.S. Hon, B. Naughton, S. Saxonov, L. Avey, A. Wojcicki, I. Pe’er and J. Mountain, Web-based, participant-driven studies yield novel genetic associations for common traits, *PLoS Genet.* **6**, 2010, e1000993, <https://doi.org/10.1371/journal.pgen.1000993>.
- [17]** P. Sulem, D.F. Gudbjartsson, S.N. Stacey, A. Helgason, T. Rafnar, M. Jakobsdottir, S. Steinberg, S.A. Gudjonsson, A. Palsson, G. Thorleifsson, S. Pálsson, B. Sigurgeirsson, K. Thorisdottir, R. Ragnarsson, K.R. Benediksdottir, et al. Two newly identified genetic determinants of pigmentation in Europeans, *Nat. Genet.* **40**, 2008, 835–837, <https://doi.org/10.1038/ng.160>.
- [18]** P. Sulem, D.F. Gudbjartsson, S.N. Stacey, A. Helgason, T. Rafnar, K.P. Magnusson, A. Manolescu, A. Karason, A. Palsson, G. Thorleifsson, M. Jakobsdottir, S. Steinberg, S. Pálsson, F. Jonasson, B. Sigurgeirsson, et al., Genetic

determinants of hair, eye and skin pigmentation in Europeans, *Nat. Genet.* **39**, 2007, 1443–1452, <https://doi.org/10.1038/ng.2007.13>.

- [19]** L.C. Jacobs, M.A. Hamer, D.A. Gunn, J. Deelen, J.S. Lall, D. van Heemst, H.-W. Uh, A. Hofman, A.G. Uitterlinden, C.E.M. Griffiths, M. Beekman, P.E. Slagboom, M. Kayser, F. Liu and T. Nijsten, A genome-wide association study identifies the skin color genes IRF4, MC1R, ASIP, and BNC2 influencing facial pigmented spots, *J. Invest. Dermatol.* **135**, 2015, 1735–1742, <https://doi.org/10.1038/jid.2015.62>.
- [20]** L.C. Jacobs, A. Wollstein, O. Lao, A. Hofman, C.C. Klaver, A.G. Uitterlinden, T. Nijsten, M. Kayser and F. Liu, Comprehensive candidate gene study highlights UGT1A and BNC2 as new genes determining continuous skin color variation in Europeans, *Hum. Genet.* **132**, 2013, 147–158, <https://doi.org/10.1007/s00439-012-1232-9>.
- [21]** C. Martínez-Cadenas, S. López, G. Ribas, C. Flores, O. García, A. Sevilla, I. Smith-Zubiaga, M. Ibarrola-Villaba, M. del, M. Pino-Yanes, J. Gardeazabal, D. Boyano, A. García de Galdeano, N. Izagirre, C. de la Rúa and S. Alonso, Simultaneous purifying selection on the ancestral MC1R allele and positive selection on the melanoma-risk allele V60L in south Europeans, *Mol. Biol. Evol.* **30**, 2013, 2654–2665, <https://doi.org/10.1093/molbev/mst158>.
- [22]** K.A. Beaumont, R.A. Newton, D.J. Smit, J.H. Leonard, J.L. Stow and R.A. Sturm, Altered cell surface expression of human MC1R variant receptor alleles associated with red hair and skin cancer risk, *Hum. Mol. Genet.* **14**, 2005, 2145–2154, <https://doi.org/10.1093/hmg/ddi219>.
- [23]** K.A. Beaumont, S.N. Shekar, S.L. Shekar, R.A. Newton, M.R. James, J.L. Stow, D.L. Duffy and R.A. Sturm, Receptor function, dominant negative activity and phenotype correlations for MC1R variant alleles, *Hum. Mol. Genet.* **16**, 2007, 2249–2260, <https://doi.org/10.1093/hmg/ddm177>.
- [24]** X. Guan, J. Niu, Z. Liu, L.-E. Wang, C.I. Amos, J.E. Lee, J.E. Gershenwald, E.A. Grimm and Q. Wei, Variants in melanocortin 1 receptor gene contribute to risk of melanoma—a direct sequencing analysis in a Texas population, *Pigment Cell Melanoma Res.* **26**, 2013, 422–425, <https://doi.org/10.1111/pcmr.12070>.
- [25]** M. Ibarrola-Villava, M. Peña-Chilet, M.J. Llorca-Cardenosa, S. Oltra, C.-M. Cadenas, J. Bravo and G. Ribas, Modeling MC1R rare variants: a structural evaluation of variants detected in a Mediterranean case-control study *J. Invest. Dermatol.* **134**, 2014, 1146–1149, <https://doi.org/10.1038/jid.2013.469>.
- [26]** P.A. Kanetsky, T.R. Rebbeck, A.J. Hummer, S. Panossian, B.K. Armstrong, A. Krickler, L.D. Marrett, R.C. Millikan, S.B. Gruber, H.A. Culver, R. Zanetti, R.P. Gallagher, T. Dwyer, K. Busam, L. From, et al., Population-based study of natural variation in the melanocortin-1 receptor gene and melanoma, *Cancer Res.* **66**, 2006, 9330–9337, <https://doi.org/10.1158/0008-5472.CAN-06-1634>.
- [27]** A. Ozola, K. Azarjana, S. Doniņa, G. Proboka, I. Mandriķa, R. Petrovska, I. Cēma, O. Heisele, L. Enģele, B. Streinerte and D. Pjanova, Melanoma risk associated with MC1R gene variants in Latvia and the functional analysis of rare variants, *Cancer Genet.* **206**, 2013, 81–91, <https://doi.org/10.1016/j.cancergen.2013.01.002>.
- [28]** A.B. Pérez Oliva, L.P. Fernández, C. Detorre, C. Herráiz, J.A. Martínez-Escribano, J. Benítez, J.A. Lozano Teruel, J.C. García-Borrón, C. Jiménez-Cervantes and G. Ribas, Identification and functional analysis of novel variants of the human melanocortin 1 receptor found in melanoma patients, *Hum. Mutat.* **30**, 2009, 811–822, <https://doi.org/10.1002/humu.20971>.
- [29]** L. Fernandez, R. Milne, J. Bravo, J. Lopez, J. Avilés, M. Longo, J. Benítez, P. Lázaro and G. Ribas, MC1R: three novel variants identified in a malignant melanoma association study in the Spanish population, *Carcinogenesis* **28**, 2007, 1659–1664, <https://doi.org/10.1093/carcin/bgm084>.
- [30]** C. Herraiz, F. Journé, G. Ghanem, C. Jiménez-Cervantes and J.C. García-Borrón, Functional status and relationships of melanocortin 1 receptor signaling to the cAMP and extracellular signal-regulated protein kinases 1 and 2 pathways in human melanoma cells, *Int. J. Biochem. Cell Biol.* **44**, 2012, 2244–2252, <https://doi.org/10.1016/j.biocel.2012.09.008>.
- [31]** K. Nakayama, A. Soemantri, F. Jin, B. Dashnyam, R. Ohtsuka, P. Duanchang, M.N. Isa, W. Settheetham-Ishida, S. Harihara and T. Ishida, Identification of novel functional variants of the melanocortin 1 receptor gene originated from Asians, *Hum. Genet.* **119**, 2006, 322–330, <https://doi.org/10.1007/s00439-006-0141-1>.
- [32]** M.C. Fargnoli, E. Altobelli, G. Keller, S. Chimenti, H. Höfler and K. Peris, Contribution of melanocortin-1 receptor gene variants to sporadic cutaneous melanoma risk in a population in central Italy: a case-control study, *Melanoma Res.* **16**, 2006, 175–182, <https://doi.org/10.1097/01.cmr.0000198454.11580.b5>.
- [33]** C. Jiménez-Cervantes, S. Germer, P. González, J. Sánchez, C.O. Sánchez and J.C. García-Borrón, Thr40 and Met122 are new partial loss-of-function natural mutations of the human melanocortin 1 receptor, *FEBS Lett.* **508**, 2001, 44–48.
- [34]** J.H. Moore, J.C. Gilbert, C.-T. Tsai, F.-T. Chiang, T. Holden, N. Barney and B.C. White, A flexible computational framework for detecting, characterizing, and interpreting statistical patterns of epistasis in genetic studies

of human disease susceptibility, *J. Theor. Biol.* **241**, 2006, 252-261, <https://doi.org/10.1016/j.jtbi.2005.11.036>.

- [35] C. Martinez-Cadenas, M. Peña-Chilet, M. Ibarrola-Villava and G. Ribas, Gender is a major factor explaining discrepancies in eye colour prediction based on HERC2/OCA2 genotype and the IrisPlex model, *Forensic Sci. Int. Genet.* **7**, 2013, 453-460, <https://doi.org/10.1016/j.fsigen.2013.03.007>.
- [36] M. Ibarrola-Villava, H.-H. Hu, M. Guedj, L.P. Fernandez, V. Descamps, N. Basset-Seguín, M. Bagot, A. Bensussan, P. Saiag, M.C. Fargnoli, K. Peris, J.A. Aviles, A. Lluch, G. Ribas and N. Soufir, MC1R, SLC45A2 and TYR genetic variants involved in melanoma susceptibility in southern European populations: results from a meta-analysis, *Eur. J. Cancer-Oxf. Engl.* **1990** **48**, 2012, 2183-2191, <https://doi.org/10.1016/j.ejca.2012.03.006>. **48**, 2012, 2183-2191.
- [37] D.L. Duffy, Z.Z. Zhao, R.A. Sturm, N.K. Hayward, N.G. Martin and G.W. Montgomery, Multiple pigmentation gene polymorphisms account for a substantial proportion of risk of cutaneous malignant melanoma, *J. Invest. Dermatol.* **130**, 2010, 520-528, <https://doi.org/10.1038/jid.2009.258>.
- [38] E.W. Steyerberg, F.E. Harrell, G.J. Borsboom, M.J. Eijkemans, Y. Vergouwe and J.D. Habbema, Internal validation of predictive models: efficiency of some procedures for logistic regression analysis, *J. Clin. Epidemiol.* **54**, 2001, 774-781.
- [39] E.W. Steyerberg and Y. Vergouwe, Towards better clinical prediction models: seven steps for development and an ABCD for validation, *Eur. Heart J.* **35**, 2014, 1925-1931, <https://doi.org/10.1093/eurheartj/ehu207>.
- [40] C. Praetorius, C. Grill, S.N. Stacey, A.M. Metcalf, D.U. Gorkin, K.C. Robinson, E. Van Otterloo, R.S.Q. Kim, K. Bergsteinsdottir, M.H. Ogmundsdottir, E. Magnúsdottir, P.J. Mishra, S.R. Davis, T. Guo, M.R. Zaidi, et al., A polymorphism in IRF4 affects human pigmentation through a tyrosinase-dependent MITF/TFAP2A pathway, *Cell* **155**, 2013, <https://doi.org/10.1016/j.cell.2013.10.022>.
- [41] N.C. de A. Fracasso, E.S. de Andrade, C.E.V. Wiesel, C.C.F. Andrade, L.R. Zanão, M.S. da Silva, L.A. Marano, E.A. Donadi, E.C. Castelli, A.L. Simões and C.T. Mendes-Junior, Haplotypes from the SLC45A2 gene are associated with the presence of freckles and eye, hair and skin pigmentation in Brazil, *Leg. Med.* **25**, 2017, 43-51, <https://doi.org/10.1016/j.legalmed.2016.12.013>.
- [42] F. Liu, B. Wen and M. Kayser, Colorful DNA polymorphisms in humans, *Semin. Cell Dev. Biol.* **24**, 2013, 562-575, <https://doi.org/10.1016/j.semcd.2013.03.013>.
- [43] S. Walsh, A. Lindenbergh, S.B. Zuniga, T. Sijen, P. de Knijff, M. Kayser and K.N. Ballantyne, Developmental validation of the IrisPlex system: determination of blue and brown iris colour for forensic intelligence, *Forensic Sci. Int. Genet.* **5**, 2011, 464-471, <https://doi.org/10.1016/j.fsigen.2010.09.008>.
- [44] S. Walsh, L. Chaitanya, L. Clarisse, L. Wirken, J. Draus-Barini, L. Kovatsi, H. Maeda, T. Ishikawa, T. Sijen, P. de Knijff, W. Branicki, F. Liu and M. Kayser, Developmental validation of the HIrisPlex system: DNA-based eye and hair colour prediction for forensic and anthropological usage, *Forensic Sci. Int. Genet.* **9**, 2014, 150-161, <https://doi.org/10.1016/j.fsigen.2013.12.006>.
- [45] S. Heilmann, A.K. Kiefer, N. Fricker, D. Drichel, A.M. Hillmer, C. Herold, J.Y. Tung, N. Eriksson, S. Redler, R.C. Betz, R. Li, A. Káráson, D.R. Nyholt, K. Song, S.H. Vermeulen, et al., Androgenetic alopecia: identification of four genetic risk loci and evidence for the contribution of WNT signaling to its etiology, *J. Invest. Dermatol.* **133**, 2013, 1489-1496, <https://doi.org/10.1038/jid.2013.43>.
- [46] K. Adhikari, T. Fontanil, S. Cal, J. Mendoza-Revilla, M. Fuentes-Guajardo, J.-C. Chacón-Duque, F. Al-Saadi, J.A. Johansson, M. Quinto-Sanchez, V. Acuña-Alonso, C. Jaramillo, W. Arias, R. Barquera Lozano, G. Macín Pérez, J. Gómez-Valdés, et al., A genome-wide association scan in admixed Latin Americans identifies loci influencing facial and scalp hair features, *Nat. Commun.* **7**, 2016, 10815, <https://doi.org/10.1038/ncomms10815>.
- [47] F. Liu, A.E.J. Hendriks, A. Ralf, A.M. Boot, E. Benyi, L. Säwendahl, B.A. Oostra, C. van Duijn, A. Hofman, F. Rivadeneira, A.G. Uitterlinden, S.L.S. Drop and M. Kayser, Common DNA variants predict tall stature in Europeans, *Hum. Genet.* **133**, 2014, 587-597, <https://doi.org/10.1007/s00439-013-1394-0>.
- [48] P. Claes, D.K. Liberton, K. Daniels, K.M. Rosana, E.E. Quillen, L.N. Pearson, B. McEvoy, M. Bauchet, A.A. Zaidi, W. Yao, H. Tang, G.S. Barsh, D.M. Absher, D.A. Puts, J. Rocha, et al., Modeling 3D facial shape from DNA, *PLoS Genet.* **10**, 2014, e1004224, <https://doi.org/10.1371/journal.pgen.1004224>.
- [49] B. Hernando, M. Ibarrola-Villava, M. Peña-Chilet, S. Alonso, G. Ribas and C. Martinez-Cadenas, Sex and MC1R variants in human pigmentation: differences in tanning ability and sensitivity to sunlight between sexes, *J. Dermatol. Sci.* **84**, 2016, 346-348, <https://doi.org/10.1016/j.jdermsci.2016.09.004>.
- [50] E. Pośpiech, J. Karłowska-Pik, B. Ziemkiewicz, M. Kukla, M. Skowron, A. Wojas-Pelc and W. Branicki, Further evidence for population specific differences in the effect of DNA markers and gender on eye colour prediction in forensics, *Int. J. Legal Med.* **130**, 2016, 923-934, <https://doi.org/10.1007/s00414-016-1388-2>.

- [51] S.I. Candille, D.M. Absher, S. Beleza, M. Bauchet, B. McEvoy, N.A. Garrison, J.Z. Li, R.M. Myers, G.S. Barsh, H. Tang and M.D. Shriver, Genome-wide association studies of quantitatively measured skin, hair, and eye pigmentation in four European populations, *PLoS One* **7**, 2012, e48294, <https://doi.org/10.1371/journal.pone.0048294>.
- [52] B. Hernando, M. Ibarrola-Villava, L.P. Fernandez, M. Peña-Chilet, M. Llorca-Cardena, S.S. Oltra, S. Alonso, M.D. Boyano, C. Martinez-Cadenas and G. Ribas, Sex-specific genetic effects associated with pigmentation, sensitivity to sunlight, and melanoma in a population of Spanish origin, *Biol. Sex Differ.* **7**, 2016, 17, <https://doi.org/10.1186/s13293-016-0070-1>.
- [53] C. Pietroni, J.D. Andersen, P. Johansen, M.M. Andersen, S. Harder, R. Paulsen, C. Børsting and N. Morling, The effect of gender on eye colour variation in European populations and an evaluation of the IrisPlex prediction model, *Forensic Sci. Int. Genet.* **11**, 2014, 1-6, <https://doi.org/10.1016/j.fsigen.2014.02.002>.
- [54] C. Martinez-Cadenas, M. Peña-Chilet, M.J. Llorca-Cardena, R. Cervera, M. Ibarrola-Villava and G. Ribas, Gender and eye colour prediction discrepancies: a reply to criticisms, *Forensic Sci. Int. Genet.* **9**, 2014, e7-9, <https://doi.org/10.1016/j.fsigen.2013.10.002>.
- [55] A.R. Wood, T. Esko, J. Yang, S. Vedantam, T.H. Pers, S. Gustafsson, A.Y. Chu, K. Estrada, J. 'an Luan, Z. Kutalik, N. Amin, M.L. Buchkovich, D.C. Croteau-Chonka, F.R. Day, Y. Duan, et al., Defining the role of common variation in the genomic and biological architecture of adult human height, *Nat. Genet.* **46**, 2014, 1173-1186, <https://doi.org/10.1038/ng.3097>.
- [56] E.W. Steyerberg, S.E. Bleeker, H.A. Moll, D.E. Grobbee and K.G.M. Moons, Internal and external validation of predictive models: a simulation study of bias and precision in small samples, *J. Clin. Epidemiol.* **56**, 2003, 441-447 [57] J.C. Garcia-Borrón, Z. Abdel-Malek, C. Jiménez-Cervantes. MC1R, the cAMP pathway, and the response to solar UV: extending the horizon beyond pigmentation. *Pigment Cell Melanoma Res.* **27**, 2014, 699-720.

▼ E-Extra

For practical considerations, knowing that ephelides tend to disappear during aging due to undetermined reasons, we tested the accuracy of our prediction model to determine the presence of ephelides in adulthood. The model presented an AUC of 0.756, a slightly lower value compared to the AUC value obtained for freckle prediction in childhood (0.781 for internal cross-validation and 0.809 for external validation). No significant contribution to freckling prediction in adulthood was noted for *r* variants, suggesting that other unknown predictors could have an independent additional impact on the freckling phenotype in adulthood. Interestingly, after adjustment by *IRF4* rs12203592 (the most associated genetic predictor for freckling in adulthood), the minor allele of *SLC45A2* rs16891982 was weakly associated with the absence of freckles in adulthood (OR = 0.64 (0.43-0.94), *P*-value = 0.023). As revealed by the MDR analysis, the effect of this *SLC45A2* variant may not be strongly influenced by a locus-locus interaction, since there is no interaction between *SLC45A2* and the rest of the freckle loci (Fig. S2B).

▼ E-component

Under the supervision of a professional, each participant completed a standardised questionnaire to collect information on sex, age, pigmentation traits, history of childhood sunburns, Fitzpatrick's skin type classification, and sun exposure habits. Details of ephelides occurrence both during the infancy or adolescence periods and in adulthood were collected in the questionnaire (an illustration is shown in Fig. S1).

[Multimedia Component 1](#)

Fig. S1 Examples of heavily freckled, lightly freckled and non-freckled individuals of Mediterranean origin.

MDR analysis for locus-locus interaction detection indicated that the best freckle prediction model was composed by four genetic variables and sex (*BNC2* rs2153271 was not included). The locus-locus interaction analysis showed a redundant interaction between *R* variants in the *MC1R* gene and rs12203592 in the *IRF4* gene, denoted by the blue lines connecting these two genetic determinants in the dendrogram (Fig. S2). However, the inclusion of this genetic interaction in our freckle prediction model did not significantly modify the prediction capacity using a repeated 10-fold cross-validation approach (AUC = 0.768, specificity = 83.20%, sensitivity = 61.27%, and accuracy = 75.45%).

[Multimedia Component 2](#)

Fig. S2 Analysis using MDR software of locus-locus interactions influencing the freckling phenotype in A) childhood and B) adulthood. Blue lines shown in entropic dendrograms represent redundant interactions.

Highlights

- Prediction of human appearance from DNA is a useful tool to identify unknown persons.

- Genetic variants in *MC1R*, *IRF4*, *ASIP* and *BNC2* contribute to freckling in Spain.
 - A preliminary DNA-based prediction model for the presence of ephelides is developed.
 - Accuracy of the newly-generated freckle prediction model is reasonably high.
 - Further research is needed before practical use in forensics of the newly-generated freckle model.
-

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