

Library

The University of Bradford Institutional Repository

http://bradscholars.brad.ac.uk

This work is made available online in accordance with publisher policies. Please refer to the repository record for this item and our Policy Document available from the repository home page for further information.

To see the final version of this work please visit the publisher's website. Access to the published online version may require a subscription.

Link to publisher's version: http://dx.doi.org/10.1128/genomeA.00095-15

Citation: Chan K, Chen JW, Chang C et al (2015) Draft genome sequence of *Lysinibacillus* sp. strain A1, isolated from Malaysian tropical soil. Genome Announcements. 3(2): e00095-15.

Copyright statement: © 2015 Chan et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.





Draft Genome Sequence of *Lysinibacillus* sp. Strain A1, Isolated from Malaysian Tropical Soil

© Kok-Gan Chan, a Jian Woon Chen, a Chien-Yi Chang, b,c Wai-Fong Yin, a Xin-Yue Chana

Division of Genetics and Molecular Biology, Institute of Biological Sciences, Faculty of Science, University of Malaya, Kuala Lumpur, Malaysia^a; Interdisciplinary Computing and Complex BioSystems (ICOS) Research Group, School of Computing Science, Newcastle University, Newcastle upon Tyne, United Kingdom^b; Centre for Bacterial Cell Biology, Medical School, Newcastle University, Newcastle upon Tyne, United Kingdom^c

In this work, we describe the genome of *Lysinibacillus* sp. strain A1, which was isolated from tropical soil. Analysis of its genome sequence shows the presence of a gene encoding for a putative peptidase responsible for nitrogen compounds.

Received 24 January 2015 Accepted 18 February 2015 Published 26 March 2015

Citation Chan K-G, Chen JW, Chang C-Y, Yin W-F, Chan X-Y. 2015. Draft genome sequence of *Lysinibacillus* sp. strain A1, isolated from Malaysian tropical soil. Genome Announc 3(2):e00095-15. doi:10.1128/genomeA.00095-15.

Copyright © 2015 Chan et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license. Address correspondence to Kok-Gan Chan, kokqan@um.edu.my.

Lysinibacillus spp. are Gram-positive rod bacteria that can survive in a wide range of extreme environments, including heavy metal-contaminated soil (1–5). Even though Lysinibacillus sp. is well known for its bioremediation potential, knowledge about its biodegradation ability is limited. In this study, we analyzed the bacterial genome of Lysinibacillus sp. strain A1. In addition, we determined the peptidase-coding gene from its whole-genome sequence.

Lysinibacillus sp. strain A1 was isolated from soil surface (Rimba Ilmu, Kuala Lumpur, Malaysia) using KGm medium and cultivated in Luria-Bertani medium (6, 7). The genomic DNA was isolated using a MasterPure DNA purification kit (Epicenter, USA) (8). The extracted DNA was quantified and qualified using Qubit version 2.0 (Invitrogen, USA) and Nanodrop (Thermo Scientific, USA) (9). The high-quality DNA was sent for nextgeneration sequencing (NGS) library preparation using a Nextera DNA sample preparation kit (Illumina, USA) and sequenced using a MiSeq 600-cycle sequencing kit (version 3) on a MiSeq platform (Illumina, USA) (9). The preliminary analysis was performed with CLC Genomic Workbench version 7.5, while the annotation are performed using the NCBI prokaryote genome annotation pipeline (version 2.9) and NCBI BLAST against the nr database (10, 11).

The NGS of the *Lysinibacillus* sp. strain A1 genome by MiSeq generated 1.2 million paired-end reads. These reads were trimmed and assembled into 57 contigs with average coverage of 42-fold and an N_{50} of 206,495 bp. The genome size is 4.75 Mbps with 37.45% G+C content. This genome carried 4,449 coding DNA sequences (CDS) and coded for 4,693 genes and 159 pseudogenes.

A peptidase-coding gene was detected in contig 1 in the draft genome of *Lysinibacillus* sp. strain A1. The length of this gene is 1,089 bp, and it is located at a position between 182,713 and 183,801 bp at contig 1. Based on the gene alignment and comparison, it is classified into peptidase M28. It has been reported that peptidase is used by both macro- and microorganisms to break down protein by hydrolyzing its peptide bond in order to acquire nutrients (12). The cocatalytic active site of peptidase M28 was

formed by two zinc ions (13, 14). To our knowledge, the production of peptidase M28 by *Lysinibacillus* has not been reported. Thus, our work on the genome and peptidase gene of *Lysinibacillus* sp. strain A1 will lead to further understanding on the function of this protein hydrolysis enzyme.

Nucleotide sequence accession numbers. This draft genome was deposited into DDBJ/EMBL/GenBank under the accession number JSZM00000000. The version described in this paper is the first version, JSZM01000000.

ACKNOWLEDGMENTS

Kok-Gan Chan thanks the UM High Impact Research Grants (UM-MOHE HIR Grant UM C/625/1/HIR/MOHE/CHAN/01, no. A000001-50001; UM-MOHE HIR Grant UM C/625/1/HIR/MOHE/CHAN/14/1, no. H-50001-A000027) for financial support.

REFERENCES

- 1. Kong D, Wang Y, Zhao B, Li Y, Song J, Zhai Y, Zhang C, Wang H, Chen X, Zhao B, Ruan Z. 2014. *Lysinibacillus halotolerans* sp. nov., isolated from saline-alkaline soil. Int J Syst Evol Microbiol 64:2593–2598. http://dx.doi.org/10.1099/ijs.0.061465-0.
- He M, Li X, Liu H, Miller SJ, Wang G, Rensing C. 2011. Characterization and genomic analysis of a highly chromate resistant and reducing bacterial strain *Lysinibacillus fusiformis* ZC1. J Hazard Mater 185: 682–688. http://dx.doi.org/10.1016/j.jhazmat.2010.09.072.
- Miwa H, Ahmed I, Yokota A, Fujiwara T. 2009. Lysinibacillus parviboronicapiens sp. nov., a low-boron-containing bacterium isolated from soil. Int J Syst Evol Microbiol 59:1427–1432. http://dx.doi.org/10.1099/ ijs.0.65455-0.
- Rahman A, Nahar N, Nawani NN, Jass J, Desale P, Kapadnis BP, Hossain K, Saha AK, Ghosh S, Olsson B, Mandal A. 2014. Isolation and characterization of a *Lysinibacillus* strain B1-CDA showing potential for bioremediation of arsenics from contaminated water. J Environ Sci Health A Tox Hazard Subst Environ Eng 49:1349–1360. http://dx.doi.org/ 10.1080/10934529.2014.928247.
- Kumar M, Yadav AN, Tiwari R, Prasanna R, Saxena AK. 2014. Deciphering the diversity of culturable thermotolerant bacteria from Manikaran hot springs. Ann Microbiol 64:741–751. http://dx.doi.org/10.1007/s13213-013-0709-7.
- Chen JW, Koh C-L, Sam C-K, Yin W-F, Chan K-G. 2013. Short chain N-acyl homoserine lactone production by soil isolate Burkholderia sp.

- strain A9. Sensors 13:13217-13227. http://dx.doi.org/10.3390/s131013217.
- Chan K-G, Yin W-F, Sam C-K, Koh C-L. 2009. A novel medium for the isolation of *N*-acylhomoserine lactone-degrading bacteria. J Ind Microbiol Biotechnol 36:247–251. http://dx.doi.org/10.1007/s10295 -008-0491-x.
- Han-Jen RE, Wai-Fong Y, Kok-Gan C. 2013. Pandoraea sp. RB-44, a novel quorum sensing soil bacterium. Sensors 13:14121–14132. http:// dx.doi.org/10.3390/s131014121.
- Chan X-Y, Chua KH, Yin W-F, Puthucheary SD, Chan K-G. 2014. Whole-genome analysis of *Aeromonas hydrophila* strain 187, exhibiting quorum-sensing activity. Genome Announc 2(6):e01360-01314. http://dx.doi.org/10.1128/genomeA.01360-14.
- NCBI Resource Coordinators. 2014. Database resources of the national center for biotechnology information. Nucleic Acids Res 42:D7–D17. http://dx.doi.org/10.1093/nar/gkt1146.
- 11. McCulloch JA, de Oliveira VM, de Almeida Pina AV, Pérez-Chaparro

- PJ, de Almeida LM, de Vasconcelos JM, de Oliveira LF, da Silva DE, Rogez HL, Cretenet M, Mamizuka EM, Nunes MR. 2014. Complete genome sequence of *Lactococcus lactis* strain ai06, an endophyte of the Amazonian açaí palm. Genome Announc 2:e01225-01214. http://dx.doi.org/10.1128/genomeA.01225-14.
- 12. Kooi C, Sokol PA. 1996. Differentiation of thermolysins and serralysins by monoclonal antibodies. J Med Microbiol 45:219–225. http://dx.doi.org/10.1099/00222615-45-3-219.
- Tsukamoto T, Flanary JM, Rojas C, Slusher BS, Valiaeva N, Coward JK. 2002. Phosphonate and phosphinate analogues of N-acylated γ-glutamylglutamate: potent inhibitors of glutamate carboxypeptidase II. Bioorg Med Chem Lett 12:2189–2192. http://dx.doi.org/10.1016/S0960-894X(02)00360-8.
- Fundoiano-Hershcovitz Y, Rabinovitch L, Langut Y, Reiland V, Shoham G, Shoham Y. 2004. Identification of the catalytic residues in the double-zinc aminopeptidase from *Streptomyces griseus*. FEBS Lett 571: 192–196. http://dx.doi.org/10.1016/j.febslet.2004.07.001.