

Recommendations for the management of pulmonary fungal infections in patients with rheumatoid arthritis

M. Galli¹, S. Antinori¹, F. Atzeni², L. Meroni¹, A. Riva¹, C. Scirè³, F. Adorni⁴, L. Quartuccio⁵, M. Sebastiani⁶, P. Airò⁷, L. Bazzichi⁸, F. Cristini⁹, V. Del Bono¹⁰, A. Manfredi⁶, O. Viapiana¹¹, F. De Rosa¹², E. Favalli¹³, E. Petrelli¹⁴, C. Salvarani¹⁵, M. Govoni³, S. Corcione¹⁶, R. Scrivo¹⁷, L. Sarmati¹⁸, A. Lazzarin¹⁹, W. Grassi²⁰, C. Mastroianni²¹, G.B. Gaeta²², G. Ferraccioli²³, M. Cutolo²⁴, S. De Vita⁵, G. Lapadula²⁵, M. Matucci-Cerinic²⁶, O. Armignacco²⁷, P. Sarzi-Puttini²⁸

Authors' affiliations on page 1024.

Massimo Galli*, Spinello Antinori*, Fabiola Atzeni, Luca Meroni, Agostino Riva, Carlo A. Scirè, Fulvio Adorni, Luca Quartuccio, Marco Sebastiani, Paolo Airò, Laura Bazzichi, Francesco Cristini, Valerio Del Bono, Andreina Manfredi, Ombretta Viapiana, Francesco De Rosa, Ennio Favalli, Enzo Petrelli, Carlo Salvarani, Marcello Govoni, Silvia Corcione, Rossana Scrivo, Loredana Sarmati, Adriano Lazzarin, Walter Grassi, Claudio Mastroianni, Giovanni Battista Gaeta, Gianfranco Ferraccioli, Maurizio Cutolo, Salvatore De Vita, Giovanni Lapadula, Marco Matucci-Cerinic, Orlando Armignacco, Piercarlo Sarzi-Puttini

*These authors contributed equally to this paper.

Please address correspondence to: Massimo Galli, MD, Department of Clinical and Biomedical Sciences "Luigi Sacco", University of Milano, Luigi Sacco Hospital, 20127 Milan, Italy. E-mail: massimo.galli@unimi.it

Received on December 13, 2016; accepted in revised form on February 19, 2017.

Clin Exp Rheumatol 2017; 35: 1018-1028.

© Copyright CLINICAL AND EXPERIMENTAL RHEUMATOLOGY 2017.

Key words: rheumatoid arthritis, pneumonia, fungal infection, recommendations

The Italian group for the Study and Management of Infections in patients with Rheumatic diseases (ISMIR group) is jointly promoted by the Italian Society of Rheumatology (SIR) and the Italian Society of Infectious and Tropical Diseases (SIMIT), and is coordinated by Massimo Galli and Piercarlo Sarzi-Puttini.

Competing interests: none declared.

ABSTRACT

Often life-threatening pulmonary fungal infections (PFIs) can occur in patients with rheumatoid arthritis (RA) receiving disease-modifying anti-rheumatic drugs (DMARDs). Most of the data concerning PFIs in RA patients come from case reports and retrospective case series. Of the five most widely described PFIs, *Pneumocystis jirovecii* pneumonia (PJP) has rarely been seen outside Japan, pulmonary cryptococcosis has been diagnosed in only a small number of patients worldwide, pulmonary coccidioidomycosis has almost only been observed in endemic areas, the limited number of cases of pulmonary histoplasmosis have mainly occurred in the USA, and the rare cases of invasive pulmonary aspergillosis have only been encountered in leukopenic patients. Many aspects of the prophylaxis, diagnosis and treatment of PFIs in RA patients remain to be defined, as does the role of each DMARD in increasing the risk of infection, and the possibility of resuming biological and non-biological DMARD treatment after the infection has been cured. The recommendations for the management of PFIs described in this paper are the product of a consensus procedure promoted by the Italian group for the Study and Management of Infections in Patients with Rheumatic Diseases (the ISMIR group).

Introduction

The increasing number of immunocompromised patients and improvements in diagnostic methods have led to an increase in the number of diagnoses of

invasive pulmonary fungal infections (PFIs) (1). These have also been described in patients receiving biological tumour necrosis factor (TNF)- α inhibitors (TNFIs) (2-4), although the findings of meta-analyses of randomised controlled trials (RCTs) indicate that the real risk of opportunistic fungal infections is not very high. Bongartz *et al.* reported one case of histoplasmosis and one of coccidioidomycosis among 126 serious infections in patients with rheumatoid arthritis (RA) treated with infliximab or adalimumab (5), and a meta-analysis of 70 trials found a 1.7 excess risk of infections per 1000 patients treated with biological drugs due to a significantly increased risk of mycobacterial and viral infections, but not invasive fungal infections (4). Nevertheless, PFIs are often life-threatening, and many aspects of their prophylaxis, diagnosis and treatment in the particular setting of RA patients remain to be defined, as does the possibility of resuming treatment with disease-modifying anti-rheumatic drugs (DMARDs) after infection. For these reasons, the Italian group for the Study and Management of the Infections in Patients with Rheumatic Diseases (the ISMIR group) endorsed a national consensus process to review the available evidence and produce practical, hospital-wide recommendations.

Methods

The criteria of the Oxford Centre for Evidence-based Medicine (<http://www.cebm.net/index.aspx?o=1025>) were used to assess the quality of the

evidence and the strength of the recommendations.

Search strategy

The studies included in the evaluation had to be RCTs, observational studies (*i.e.* cross-sectional, non-interventional case-control or cohort studies), systematic reviews or case reports evaluating the risk of infections, opportunistic infections, and fungal infections in patients exposed to biological DMARDs (bDMARDs). A search was made of the MEDLINE and EMBASE databases from 1998 to May 2016 using the key words: “rheumatoid arthritis” or “arthritis” or “arthritides” or “polyarthritis” and “anti-TNF drugs” or “anti-TNF agents” or “anti-TNF therapy” or “anti-TNF blockers” or “infliximab” or “etanercept” or “adalimumab” and “fungal pulmonary infections”, “lung infections” “opportunistic infections” or “*Pneumocystis jirovecii* infection”, “*Cryptococcus neoformans* pulmonary infections” “*Coccidioides immitis* pulmonary infections” “*Histoplasma capsulatum* pulmonary infections”, “*Aspergillus* pulmonary infections”. There were no restrictions on publication status. The diagnosis of RA in the considered studies was made on the basis of the 1988 American College of Rheumatology (ACR) and ACR/European League Against Rheumatism (EULAR) criteria (6, 7); a diagnosis made after a clinical examination by a rheumatologist was also considered acceptable. Studies including subjects aged <18 years were discarded. This analysis considered the PFIs occurring in patients treated with bDMARDs or synthetic DMARDs (sDMARDs).

Pneumocystosis

Pneumocystis jirovecii (formerly known as *Pneumocystis carinii*) is a species-specific opportunistic pathogen classified as a fungus in the phylum Ascomyceta (8, 9). *P.jirovecii* pneumonia (PJP) can occur in deeply immunosuppressed patients (10-13), including RA patients in whom it can cause sudden and severe respiratory failure and death (14, 15). Before the introduction of bDMARDs, some cases of PJP were reported in patients who had been

treated with methotrexate (MTX) and steroids for a long time (16-18). TNF- α inhibition affects host defences against *P.jirovecii* infection in animal models (19-21) but, even after the introduction of bDMARDs, PJP remained an infrequent finding in RA patients in the USA and Europe (22-25).

A prospective study of the British Society for Rheumatology compared 13,905 patients treated with TNFs and 3,677 treated with sDMARD, and identified 17 cases of PJP cases: the incident rates were 0.2/1000 person-years of follow-up (95% confidence interval [CI] 1.2-3.3) in the TNFI cohort, and 0.11/1000 person-years follow-up (95% CI 0.3, 4.3) in the sDMARD cohort. The age-adjusted hazard ratio for PJP infection in the TNFI-treated patients was 2.3 (95% CI 0.5-10.1) (26). The situation seems to be different in Japan, where a total of 243 PJP cases had been reported in patients receiving MTX up to 2012 (15), and the post-marketing surveillance programme identified 16 cases in 7,091 patients receiving etanercept (0.23%) (27), 22 cases in 5,000 patients receiving infliximab (0.4%) (28), nine cases in 3,000 patients receiving adalimumab (0.3%) (29), and five cases in 3,881 patients receiving tocilizumab (0.28%) (30). Worldwide reports of PJP in RA patients receiving golimumab (15,31) and certolizumab pegol (32), or other biological drugs such as abatacept (15) and rituximab (33-36) are scarce and mainly anecdotal. The higher incidence of PJP in Japanese RA patients remains unexplained.

It is also unclear whether the use of bDMARDs rather than conventional treatments increases the risk of PJP, although one meta-analysis found no difference in the incidence of PJP between RA patients receiving bDMARDs and sDMARDs (mainly MTX) (4). Corticosteroid therapy can reduce the number and function of CD4⁺ T cells and thus increase the risk of PJP (10, 37-45), and it is known that CD4⁺ T cell counts of <200/ μ L are associated with the development of PJP in rheumatic patients (46, 47), although it may also occur at higher CD4 counts (14, 37, 41, 46, 48). It has been found that lymphopenia (<500/ μ L) during corticosteroid

therapy (45), lymphocyte counts of <1500/ μ L, and serum IgG levels of 1000 mg/dL (37) predict PJP infection in patients with rheumatic diseases. Komano *et al.* (49) found significantly lower levels of serum IgG and albumin in patients with PJP than in those without, and claimed that PJP was predicted by age (>65 years), prednisolone dose (\geq 6 mg/day), and pulmonary comorbidities, the role of which has since been confirmed (50).

In the British study mentioned above, the median time to PJP infection after starting TNFI treatment was 5.8 months (interquartile range 2.7-16.8) (26). The presentation of PJP in RA patients is preceded by variable degrees of fatigue and respiratory symptoms (15). Chest radiographs may be almost normal, but high-resolution CT scans can reveal diffuse ground-glass opacities without the interlobular septal boundaries typical of MTX-related interstitial pneumonia (13). The poor prognosis of PJP in RA patients has been associated with an older age, high 1-3 β -d-glucan levels, the use of MTX, hypoxaemia, bilateral lung findings, the need for mechanical ventilation, and low lymphocyte counts (37). It has been suggested that Japanese patients treated with bDMARDs are at increased risk of death (15), particularly those with pre-existing pulmonary lesions (51).

As *P.jirovecii* cannot be cultured, diagnosis of the infection relies on detecting it by means of optical microscopy in lung tissue or bronchoalveolar lavage (BAL) fluid. Searches in induced sputum are burdened by a high rate of false negative results (52) and, although the polymerase chain reaction (PCR) analysis of induced sputum is more sensitive, the method is not generally available in clinical practice. At the time of the introduction of TNF- α blocking agents, determining serum 1 \rightarrow 3- β -d-glucan levels (52, 53) was recommended in the Japanese guidelines for RA patients (54) but, in addition to being expensive, this test is pan-fungal and not specific for *P.jirovecii*.

The development of PJP in immunocompromised patients seems to be due to *de novo* infections, and the most likely way RA outpatients acquire the

infection is person-to-person transmission, and it has suggested that outpatient clinic waiting rooms may also play a role (15). The role of PJP prophylaxis in RA patients remains controversial. Universal routine prophylaxis is considered impracticable because of the long-term nature of anti-RA therapy (15), and it is known that PJP can occur after the discontinuation of primary prophylaxis in RA patients with lymphopenia (55, 56). A meta-analysis of studies of HIV-negative immunocompromised patients has shown that trimethoprim/sulfamethoxazole (TMP/SMX) prophylaxis is highly effective but it should only be considered whenever the risk of developing PJP is higher than the rate of adverse events associated with TMP/SMX administration (3.1% among adults) (57). It is therefore generally not recommended in RA patients (58), although the Japanese Ministry of Health recommendations (59) suggest short-term primary prophylaxis with TMP/SMX at a daily dose of 80-400 mg for 5-7 days (or two tablets three times a week) for all RA patients scheduled to start biological and/or non-biological anti-rheumatic therapy (60). Blood dyscrasia has been reported in patients simultaneously taking MTX and TMP/SMX at the standard therapeutic dose (61, 62), and so the use of TMP/SMX in patients treated with MTX is not recommended in the Canadian Initiative in Rheumatology guidelines (62).

TMP/SMX is the treatment of choice for *P. jirovecii* infections of any severity (1, 52). In the case of mild disease, oral TMP/SMX can be given at a dose of two double-strength tablets (160 mg of TMP and 800 mg of SMX) every eight hours. In severe cases, the drug should be administered intravenously at a TMP dose of 15-20 mg/kg and an SMX dose of 75-100 mg/kg every 6-8 hours. The alternative treatments for patients who are either intolerant or hypersensitive to TMP/SMX are intravenous pentamidine or intravenous clindamycin plus oral primaquine (52). High-dose corticosteroids greatly improve PJP outcomes in HIV-infected patients (63). It must also be remembered that the discontinuation of corticosteroid therapy in patients developing PJP can lead to

lung damage as a result of immune reconstitution (64).

P. jirovecii disappears after 7-10 days of TMP-SMX treatment in most cases (65), and no relapse of PJP was seen during an average follow-up of 22 months in patients treated with TMP/SMX for a mean of 17 days, even though they continued to receive immunosuppressants for their rheumatic diseases without secondary prophylaxis (39).

Questions and statements

When should PJP be suspected in RA patients?

PJP should enter in the differential diagnosis of interstitial disorders of the lung in RA patients. CII

Should patients with RA receive primary PJP prophylaxis?

PJP prophylaxis should be prescribed to patients with a CD4 cell count of <200/μL or a lymphocyte count of <500/μL, while carefully monitoring untoward effects. BII

Patients presenting three or more potential risk factors (an age of >65 years, a lymphocyte count of >500 μL<1500 μL, exposure to immunosuppressants and/or corticosteroids for >3 years, previous exposure to another biological drug, lung comorbidities, below normal serum albumin or serum IgG levels) merit particular attention, and can be considered for primary PJP prophylaxis on a case-by-case basis. CIII

What should be done if a patient is diagnosed with PJP?

The immediate withdrawal of corticosteroid therapy is not appropriate. On the contrary, intensification with high-dose corticosteroids should be considered during the acute phase of the disease. BII

It is appropriate to discontinue MTX, particularly in the case of serious infections requiring high-dose intravenous TMP/SMX. CIII

The discontinuation of bDMARDs should be considered in all patients, particularly in the case of lymphopenia or low CD4⁺ lymphocyte counts. DIII

Is re-treatment with bDMARDs possible in PJP survivors?

After the complete resolution of PJP symptoms and the completion of TMP/

SMX treatment, re-treatment with bDMARDs can be considered on a case-by-case basis. Secondary prophylaxis with TMP/SMX or pentamidine during re-treatment with bDMARDs can also be considered, particularly in lymphopenic patients. DIII

Cryptococcus

Cryptococcosis is a fungal infection caused by two species of yeast: *Cryptococcus neoformans* and *C. gattii* (66). *C. neoformans* infection is observed worldwide mainly in immunocompromised patients (67, 68); however, approximately 20% of the patients with a diagnosis of cryptococcosis have no apparent risk factor or underlying disease. *C. gattii* infection mainly occurs in immunocompetent hosts in endemic tropical regions, but it has also recently been reported in Vancouver Island (British Columbia) and the Pacific North-west of the USA (69).

Primary cryptococcal infection occurs in the lung following the inhalation of dried yeast cells or basidiospores from an environmental source such as avian guano, soil and trees. The disease may remain localised or disseminate through the bloodstream to the central nervous system (CNS) and invade the leptomeninges (67).

Pulmonary cryptococcosis was first described in MTX-treated RA patients in 1987 (70), and the first disseminated infection in an RA patient treated with infliximab and MTX was reported in 2002 (71). Up to 2008, 28 cases of TNFI-associated *Cryptococcus* infection had been reviewed (2). Analysis of the FDA's Adverse Event Reporting System (AERS) database from 1998 to the third quarter of 2002 showed that the incidence of cryptococcosis was similar in American patients treated with infliximab (5.1/10⁵) or etanercept (7.1/10⁵) (72), whereas no cases of cryptococcosis were detected in the 10,050 patients treated with adalimumab included in the US post-marketing database (73). However, four cases of cryptococcosis (two with pulmonary infections) were reported in 1,080 Spanish patients treated with adalimumab between 2003 and 2006 (74-76). Pulmonary cryptococcosis has also been

described in Japanese patients treated with corticosteroids and MTX including one receiving infliximab (77) and one treated with MTX and adalimumab (78). However, no cases were reported among more than 30,000 patients treated with bDMARDs enrolled in the 70 RCTs included in the meta-analysis of Kourbeti *et al.*, and only one case was detected among the control patients (4). Taken together, the available data do not indicate any significant difference in incidence of cryptococcosis depending on what bDMARD is used, or between patients treated with biological or non-biological drugs.

Computed tomography (CT) reveals nodules in 80% of RA patients with pulmonary cryptococcosis, consolidation in 30%, and cavitory lesions in 10%. Among the patients treated with TNFIs, the diagnosis was made on the basis of a biopsy or surgery in five cases (71, 75, 78-80) and by means of bronchoalveolar lavage in the others (81, 82). The serum cryptococcal capsular antigen test is highly sensitive and specific in the context of meningitis in other immunocompromised hosts (83), and was positive in 57% of RA patients with disseminated disease and pulmonary involvement (71, 80, 84, 85) and in about 40% of those with pulmonary infection alone (75, 79-81). In the presence of rheumatoid factor, false positive cryptococcal capsular antigen test results have been recorded in serum and cerebrospinal fluid (86), but these can be avoided by pre-treating serum with pronase or boiling it with EDTA for five minutes. Although the majority of patients so far described did not undergo a lumbar puncture to rule out meningitis, this is recommended in immunocompromised hosts because the fungus has a high propensity to disseminate (67, 85). In sub-Saharan Africa, where the burden of cryptococcal meningitis in HIV patients is the highest in the world, the use of the newly available lateral flow assay for the detection of cryptococcal antigen has been shown to be cost-effective (87, 88).

There are no specific recommendations regarding screening and prophylaxis for cryptococcus infection in patients receiving TNF- α blockers. All but one of the reported RA patients with pul-

monary cryptococcosis were treated with amphotericin B deoxycholate or fluconazole, followed by fluconazole maintenance therapy (71, 75, 80-84). According to the guidelines of the Infectious Diseases Society of America, immunocompromised patients with pulmonary cryptococcosis should be treated with fluconazole 400 mg/day for 6-12 months in the case of mild-moderate symptoms, whereas treatment such as that used for CNS disease is recommended in the case of severe pneumonia (89). The recommended regimen for HIV-negative transplantation patients is amphotericin B deoxycholate 0.7-1 mg/kg/day plus flucytosine 100 mg/kg/day for at least four weeks, followed by consolidation with fluconazole 400-800 mg/day for eight weeks, and 6-12 months of maintenance therapy with fluconazole 200 mg/day (89). Although it has not been demonstrated that liposomal amphotericin B (L-AMB) is more efficacious than the deoxycholate formulation, a number of guidelines and experts recommend it as first-choice therapy because it is less nephrotoxic (90-92). In the transplant setting, the use of L-AMB regardless of renal dysfunction has been suggested because it has less pro-inflammatory activity than the deoxycholate formulation (90-93). All of the patients so far described discontinued TNF- α blocker agents, and the treatment was resumed in only one case (80).

Questions and statements

When should pulmonary cryptococcosis be suspected?

Pulmonary cryptococcosis enters in the differential diagnosis of lung disorders in severely immunocompromised patients presenting nodular, consolidated or cavitory lesions of the lung. BI

What should be done in the case of patients with pulmonary cryptococcosis?

DMARD treatment should be discontinued. CIII

Is re-treatment with biological DMARDs possible in survivors?

It is currently unknown whether or not a patient with previous cryptococcosis can be safely re-treated with biological DMARDs. Most of the experts on the panel consider it unwise. CIII

Coccidiomycosis

Coccidiomycosis can be caused by two closely related dimorphic fungi: *Coccidioides immitis* and *C. posadasii* (94). *Coccidioides* species are endemic in the west and south-western part of the USA (particularly Arizona, California, Nevada, Texas and New Mexico), as well as in desert areas of Central America (Mexico, Guatemala and Honduras) (95) and South America (Argentina, Bolivia, Brazil, Paraguay and Venezuela) (94). The majority of cases are acquired by inhalation, and approximately 60% of the infections are asymptomatic. The infective forms are arthroconidia (or spore), which can remain viable for many years in soil under dry conditions.

Three hundred and forty-five cumulative cases of coccidiomycosis were identified in a random 5% sample of USA Medicare data from 1999 to 2008, including 21 patients (6.1%) with concomitant RA (95). A retrospective study of patients observed between January 2000 and June 2006 in Arizona revealed a diagnosis of coccidiomycosis in 3.1% of RA patients, and in 14.3% of those with ankylosing spondylitis (94). Among patients receiving TNFIs in endemic areas, the use of infliximab was associated with a 5.23 relative risk of coccidiomycosis (CI 95% 1.54-17.71) in comparison with other medications, and a significantly increased risk persisted even after adjusting for age and the use of MTX and prednisone (95).

Pulmonary involvement has been observed in 66-100% of coccidiomycosis cases diagnosed in patients with rheumatic diseases treated with immunosuppressive drugs (96-99). Some cases of lethal disseminated disease have been described (96-100), although exclusively pulmonary involvement has been found in 66-81.8% of patients. Rheumatic symptoms accompanying acute pulmonary disease can cause a clinical picture known as "desert rheumatism", which may lead to more aggressive treatment of the underlying rheumatic syndrome, sometime with serious consequences for the patient (98). The prevalence of coccidiomycosis in endemic areas is 1.1-9% (96-98), with incidence rates of 1% in the

first year after the diagnosis of RA, and 9% during the first five years after diagnosis, and respectively 2% and 12% one and five years after starting infliximab therapy (96). The risk is virtually absent in patients living in non-endemic areas, where the only reported RA case of pulmonary coccidioidomycosis occurred in a patient treated with infliximab who probably inhaled rock dust imported from Arizona (101).

Most cases of coccidioidomycosis have been diagnosed serologically, and some by means of cultures or histopathological demonstration (97, 102). Taroumian *et al.* made a definite diagnosis of pulmonary coccidioidomycosis by means of histology or culture in about 15% of their patients, but in all of those with disseminated disease, whereas serological tests (enzyme immunoassay, immunodiffusion or complement fixation) detected a previous asymptomatic infection in 14% (98). Some authors recommend that patients living in endemic areas should undergo a chest radiography and specific serological tests before starting TNFI therapy (103, 104), and a serological test every 3–4 months (105).

Azole prophylaxis has been suggested in patients treated with TNFIs who live in endemic regions. Immunocompromised patients with pulmonary coccidioidomycosis should be treated with an azole (fluconazole or itraconazole 400 mg/day) for 3–6 months or longer, depending on their clinical response (1). In the presence of lung cavities, the duration of antifungal therapy should be extended to 12–18 months. Patients presenting with primary coccidioidomycosis and signs or symptoms indicating possible CNS involvement should undergo a lumbar puncture to exclude meningitis. This is especially important as coccidioidomycosis meningitis should be treated with fluconazole 400–1000 mg/day or itraconazole 400–600 mg/day for life. The use of amphotericin B is generally reserved for the induction treatment of severe pulmonary or disseminated (non-meningeal) disease (1).

Fifty-nine percent of the patients in the retrospective study of Taroumian *et al.* stopped both sDMARDs and b-

DMARDs at the time coccidioidomycosis was diagnosed, 18% only stopped bDMARDs, and 22.7% continued their ongoing immunosuppressive treatment. The drugs were continued or had been resumed by 87% of the patients after a median follow-up of 12 months (range 1–72 months) (98). Most of the patients (93%) received antifungal therapy (usually fluconazole 400 mg/day), the median duration of which was 12 months (range 0–96 months).

A history of CNS infection is generally considered a contraindication to the resumption of DMARDs. In the other cases, fluconazole maintenance therapy is suggested if bDMARD therapy is resumed (100).

Questions and statements

Is any specific bDMARD associated with an increased risk of coccidioidomycosis?

The use of infliximab is associated with an increased risk of symptomatic coccidioidomycosis. CII

Are any specific interventions recommended before or during TNFI treatment?

Chest radiography and coccidioidal serology (IgM/IgG) are generally recommended in endemic areas before starting TNF- α antagonist treatment. CIII

What should be done in the case of patients with pulmonary coccidioidomycosis? Is re-treatment with biological DMARDs possible in survivors?

Immunosuppressive therapy should be stopped when coccidioidomycosis is diagnosed, but its resumption can be considered on an individual basis accompanied by fluconazole maintenance therapy. DIII

Histoplasmosis

Histoplasmosis is an endemic mycosis caused by two varieties of the dimorphic fungus *Histoplasma capsulatum* (Hc): *Hc var. capsulatum* in Mid-western and South-eastern areas of the USA, Latin America, Asia (China, India, Myanmar and Thailand), and Africa; and *Hc var. duboisii* in Africa (106). Outside these endemic areas, histoplasmosis is generally an imported infection, although small foci of autochthonous histoplasmosis have been described in

Italy (mainly in Lombardy and Emilia-Romagna) (107, 108). However, the fungus is probably more widespread than previously thought, and some experts recommend considering a diagnosis of histoplasmosis in all febrile and immunosuppressed patients, regardless of their geographical location or travelling history (109, 110). The natural habitat of the fungus is soil with a high nitrogen content, particularly in areas contaminated by bird or bat guano. Many of the described cases involve activities associated with soil excavation, building or renovation work, or visits to bat caves. Once inhaled, the mycelial form transforms itself into the pathogenic yeast in the lungs, and causes a wide range of clinical manifestations, including acute pulmonary infection or disseminated disease in about 5% of cases (mainly immunocompromised subjects) (106).

Histoplasmosis was first reported in RA patients in 2002 and, soon after, two case series from endemic areas of the United States highlighted its severe and life-threatening nature (111–113). The FDA's AERS database reported 42 cases of histoplasmosis between 1998 and 2002, with a six-fold higher ratio in patients treated with infliximab (39/233,000) than in those treated with etanercept (114). In the Mayo Clinic case series, 26 RA patients developed histoplasmosis over a period of eleven years (115): 81% were receiving MTX, 58% TNFIs, and 58% corticosteroids. The lung was the only site of infection in 54% of the patients, and 75% of the patients with disseminated disease had lung involvement. Antifungal therapy was administered to 92% of the patients, nine of whom (35%) received an initial L-AMB course (115). Hage *et al.* at Indiana University (116) found that 89% percent of the 19 patients who developed histoplasmosis while receiving TNFIs had disseminated disease (as is usual among immunocompromised hosts), and 79% showed lung involvement. A multicentre retrospective review of 98 patients diagnosed with histoplasmosis (including 52 patients with RA), 67.3% of whom were being treated with infliximab, found that concomitant corticosteroid use (OR 3.94,

Table I. Suggested screening for opportunistic pulmonary fungal infections before administering a biological DMARD to RA patients.

Pneumocystosis:	Consider total leukocyte count at baseline, and CD4 lymphocyte count in patients previously exposed to several DMARD combinations.
Cryptococcosis:	Consider chest radiography and serum cryptococcal antigen testing at baseline.
Histoplasmosis:	In endemic areas, consider chest radiography and urine histoplasmin antigen testing at baseline, and follow-up urine antigen testing every 3-4 months.
Coccidioidomycosis:	Chest radiography and serological IgM and IgG testing at baseline. Consider follow-up testing every 3-4 months for patients living in endemic areas.
Aspergillosis:	Consider absolute neutrophil count at baseline.

95% CI 1.06–14.60) and higher urine histoplasma antigen levels (OR, 1.14, 95% CI, 1.03–1.25) were independent predictors of severe disease. Two of the RA patients experienced a recurrence: one after eight months (two months after resuming adalimumab), and one after 14 months while still without biological drug treatment (117).

Transbronchial or open lung biopsies were used to diagnose 80% of the patients with histoplasmosis taking TNFIs (112). It has been suggested that the diagnostic work-up of symptomatic patients should include two or more blood cultures, and testing for *Histoplasma* antigenuria and antigenemia (116). The sensitivity of the *Histoplasma* antigen assay is 95% in patients on TNFIs (116), but the test is generally unavail-

able outside the USA. Serum *Aspergillus* galactomannan antigen test gives false-positive results in patients with disseminated histoplasmosis, and might be considered a surrogate diagnostic marker when no specific antigen test is available (118, 119). However, serum galactomannan antigen levels can be non-specifically high in patients with RA (120). Bronchoscopy with BAL and a biopsy are generally indicated for the diagnosis of pulmonary histoplasmosis, and specimens should be cultured and appropriately examined microscopically in order to identify the fungal pathogen. When available, the *Histoplasma* antigen test can also be used on BAL fluid. According to the most recent Infectious Diseases Society of America (IDSA) guidelines, antifungal

treatment is indicated for all patients (121). The recommended treatment regimen is itraconazole 200 mg three times daily for three days, followed by 200 mg twice daily for 12 months if the disease is mild or moderate; in patients with moderately severe or severe acute pulmonary histoplasmosis, it is an intravenous lipid formulation of amphotericin B (3–5 mg/kg/day for 1–2 weeks), followed by itraconazole 200 mg three times daily for three days, and then 200 mg twice daily for 12 weeks (1, 121). Monitoring itraconazole levels is generally recommended because they vary widely from patient to patient. In the patients described by Vergidis *et al.*, antifungal treatment was continued for a median of 11 months (range 3–27 months), and the median follow-up was 32 months (range 1–120 months) (117). The authors suggested that antifungal therapy can be discontinued after 12 months in patients who do not resume TNFIs. The FDA recommends discontinuing TNFIs in patients with a presumed or definite diagnosis of systemic fungal infections (122). Hage *et al.* (116) observed immune reconstitution inflammatory syndrome (IRIS) in 42% of the patients stopping TNFIs, but Vergidis *et al.* observed it in only two

Table II. Invasive pulmonary fungal diseases in RA patients treated with DMARDs: frequency and management.

Fungal disease	Frequency	Most frequently involved drugs [§]	Should DMARD treatment be discontinued?	Immune reconstitution syndrome	Can DMARDs be resumed after clinical resolution of fungal disease?
<i>Pneumocystosis</i>	Rare (except in Japan)	MTX and/or infliximab	Yes (possible steroid intensification in acute phase of PJP)	Possible in patients abruptly discontinuing corticosteroid treatment	Consider case-by-case (not recommended in the presence of lymphopenia or low CD4 cell count). Consider secondary prophylaxis with TMP/SMX
<i>Cryptococcosis</i>	Rare	None	Yes	Reported in one case (74)	Insufficient data, probably very hazardous
<i>Coccidioidomycosis</i>	Rare (only seen in the Americas)*	Infliximab	Yes	No data	Consider case-by-case, with fluconazole maintenance therapy
<i>Histoplasmosis</i>	Rare in Europe;** more frequent in the USA	Infliximab	Yes	Reported in 9% of cases (125)	Consider in patients with undetectable antigen levels and no signs of residual disease after prolonged anti-fungal therapy (≥12 months)
<i>Aspergillosis</i>	Very rare (only seen in neutropenic patients)	None	Yes	No data	Consider after reversal of neutropenia

[§]Statistical significance is not reached in any case

*In Europe, consider patients who have travelled to highly endemic areas.

**Consider in patients who have travelled to highly endemic areas; some autochthonous cases observed in Northern Italy (110, 111).

of their 52 patients (3.8%) (117). Furthermore, 25 of 74 patients (33.8%; the number of RA patients was not specified) resumed bDMARDs after a median of 12 months (range, 1–69 months). It has been suggested that patients resuming TNFI therapy should continue to receive itraconazole for as long as they are on treatment (123).

Questions and statements

Is any specific bDMARD associated with an increased risk of developing pulmonary histoplasmosis?

The risk of pulmonary histoplasmosis seems to be higher in patients treated with infliximab. DIII

What should be done in the case of patients who have pulmonary histoplasmosis?

Discontinuing immunosuppressive therapy should always be considered. If discontinued, monitor for IRIS. DIII

Is re-treatment with biological DMARDs possible?

*Resuming immunosuppressive treatment (including bDMARDs) can be considered after treatment with antifungal drugs for ≥ 12 months in patients who are *Histoplasma antigen* negative and do not show signs of residual disease. DIII*

Aspergillosis

Aspergillus spp. are ubiquitous environmental fungi capable of causing a wide variety of manifestations ranging from allergic aspergillosis to invasive disease (124). The first reported case of *A. fumigatus* pneumonia in a patient with RA was probably that of a 73-year-old woman treated with multiple immunosuppressive agents who possibly acquired the infection during hospital construction (125), and another involved concurrent infection with *A. fumigatus*, TB and herpes simplex (126). Other cases of aspergillus infection (20 associated with infliximab and 10 associated with etanercept) have been reported, but no clinical details were provided (127). TNFIs may increase the risk of developing aspergillosis by inhibiting neutrophil recruitment, and murine models have confirmed increased mortality due to aspergillus in the presence of TNF- α inhibition (127).

The main risk factor for invasive pulmonary aspergillosis is prolonged severe neutropenia.

Invasive pulmonary aspergillosis in RA patients should be treated in accordance with the international guidelines (128)

Questions and statements

What should be done in the case of patients who have invasive pulmonary aspergillosis? Discontinuing immunosuppressive therapy seems to be mandatory. DIII

Is re-treatment with biological DMARDs possible?

Re-treatment might be considered after a full course of antifungal therapy and the reversal of neutropenia. DIII

Conclusions

Although infrequent, a PFI occurring in patients with RA is a critical event that requires immediate therapeutic management decisions (Table I, II). The small number of cases reported in studies and the national registers of patients treated with bDMARDs does not allow the extrapolation of data from meta-analyses and limits the possibility of drawing up recommendations to clinical experience and expert opinion (129–131). It is also unclear whether the risk of developing a PFI is different depending on the administered TNFI. Although there is no evidence that the incidence of PFI is increasing despite the extended use of bDMARDs, the prolonged life expectancy of RA patients, the probable future increase in the use of bDMARDs and more complicated therapeutic regimens, and the resulting increase in the number of severe immunodepressed patients (132), make it necessary to pay special attention to the occurrence of PFIs.

Take-home messages

- Pulmonary fungal infections (PFIs) can occur in patients with rheumatoid arthritis (RA) receiving disease-modifying anti-rheumatic drugs (DMARDs).
- *Pneumocystis jirovecii* pneumonia (PJP) has rarely been seen outside Japan.
- Pulmonary cryptococcosis has been diagnosed in only a small number

of patients worldwide. Pulmonary coccidioidomycosis has almost only been observed in endemic areas.

- Many aspects of the prophylaxis, diagnosis and treatment of PFIs in RA patients remain to be defined.
- The possibility of resuming biological and non-biological DMARD treatment after the infection has been cured, it has been discussed.

Authors' affiliations

¹Clinica delle Malattie Infettive, Dept. of Biomedical and Clinical Sciences L. Sacco University Hospital, Milan;

²IRCCS Galeazzi Orthopaedic Institute, Milan;

³Department of Medical Sciences, UOC of Rheumatology, Santa Anna University Hospital, Ferrara;

⁴Institute of Biomedical Technologies, National Research Council, Milan;

⁵Department of Medical and Biological Sciences, Rheumatology Clinic, Santa Maria della Misericordia University-Hospital, Udine;

⁶Rheumatology Unit, Dept. of Internal Medicine, Azienda Ospedaliera ASMN, University of Modena and Reggio Emilia;

⁷Rheumatology, Allergology and Clinical Immunology Service, Spedali Civili and University, Brescia;

⁸Rheumatology Unit, Dept. of Clinical and Experimental Medicine, University of Pisa;

⁹Infectious Diseases Unit, Teaching Hospital S. Orsola-Malpighi, Alma Mater Studiorum University of Bologna;

¹⁰Clinica Malattie Infettive, IRCCS AOU San Martino-IST, Università di Genova;

¹¹Rheumatology Unit, Dept. of Medicine, University of Verona;

¹²Infectious Disease Unit, Amedeo di Savoia Hospital, University of Turin, Italy;

¹³G. Pini Orthopaedic Institute, Milan;

¹⁴Infectious Diseases Unit, San Salvatore Hospital, Pesaro;

¹⁵Rheumatology Unit, Dept. of Internal Medicine, Azienda Ospedaliera ASMN, Istituto di Ricovero e Cura a Carattere Scientifico, Reggio Emilia;

¹⁶Dept. of Medical Sciences, University of Turin, Infectious Diseases at Amedeo di Savoia Hospital, Turin;

¹⁷Rheumatology Unit, Sapienza University, Rome;

¹⁸Infectious Disease Unit, University of Rome Tor Vergata, Rome;

¹⁹Infectious Disease Unit, Vita e Salute University, Milan;

²⁰Rheumatology Unit, Polytechnic University of Marche, C. Urbani Hospital, Jesi, Italy;

²¹Infectious Disease Unit, Sapienza

University, Polo Pontino, ASL Latina;

²²Azienda Ospedaliera Universitaria Seconda Università di Napoli;

²³Division of Rheumatology, Institute of Rheumatology, Catholic University of the Sacred Heart, Rome;

²⁴Research Laboratory and Division of Clinical Rheumatology, University of Genova;

²⁵Rheumatology Unit, University of Bari;

²⁶Dept. of Experimental and Clinical Medicine, Division of Rheumatology, AOUC, University of Florence;

²⁷Infectious Diseases Unit, Belcolle Hospital, Viterbo;

²⁸Rheumatology Unit, L. Sacco University Hospital, Milan, Italy.

References

- LIMPER AH, KNOX KS, SAROSI GA *et al.*: An official American Thoracic Society Statement: treatment of fungal infections in adult pulmonary and critical care patients. *Am J Respir Crit Care Med* 2011; 183: 96-128.
- TSIODRAS S, SAMONIS G, BOUMPAS DT, KONTOYIANNIS DP: Fungal infections complicating tumor necrosis factor alpha blockade therapy. *Mayo Clin Proc* 2008; 83: 181-94.
- MASCHMEYER G, PATTERSON TF: New immunosuppressive agents and risk for invasive fungal infections. *Curr Infect Dis Reports* 2009; 11: 435-8.
- KOURBETI IS, ZIAKAS PD, MYLONAKIS E: Biologic therapies in rheumatoid arthritis and the risk of opportunistic infections: a meta-analysis. *Clin Infect Dis* 2014; 58: 1649-57.
- BONGARTZ T, SUTTON AJ, SWEETING MJ, BUCHAN I, MATTESON EL, MONTORI V: Anti-TNF antibody therapy in rheumatoid arthritis and the risk of serious infections and malignancies: systematic review and meta-analysis of rare harmful effects in randomized controlled trials. *JAMA* 2006; 295: 2275-85.
- HARRISON BJ, SYMMONS DP, BARRETT EM, SILMAN AJ: The performance of the 1987 ACR classification criteria for rheumatoid arthritis in a population based cohort of patients with early inflammatory polyarthritis. American Rheumatism Association. *J Rheumatol* 1998; 25: 2324-30.
- ALETAHA D, NEOGI T, SILMAN AJ *et al.*: Rheumatoid arthritis classification criteria: an American College of Rheumatology/European League Against Rheumatism collaborative initiative. *Arthritis Rheum* 2010; 62: 2569-81.
- EDMAN JC, KOVACS JA, MASUR H, SANTI DV, ELWOOD HJ, SOGIN ML: Ribosomal RNA sequence shows *Pneumocystis carinii* to be a member of the fungi. *Nature* 1988; 334: 519-22.
- STRINGER JR, BEARD CB, MILLER RF, WAKEFIELD AE: A new name (*Pneumocystis jirovecii*) for *Pneumocystis* from humans. *Emerg Infect Dis* 2002; 8: 891-6.
- AREND SM, KROON FP, VAN'T WOUT JW: *Pneumocystis carinii* pneumonia in patients without AIDS, 1980 through 1993: an analysis of 78 cases. *Arch Intern Med* 1995; 155: 2436-41.
- NUESCH R, BELLINI C, ZIMMERLI W: *Pneumocystis carinii* pneumonia in human immunodeficiency virus (HIV)-positive and HIV-negative immunocompromised patients. *Clin Infect Dis* 1999; 29: 1519-23.
- ROBLOT F, GODET C, LE MOAL G *et al.*: Analysis of underlying diseases and prognosis factors associated with *Pneumocystis carinii* pneumonia in immunocompromised HIV-negative patients. *Eur J Clin Microbiol Infect Dis* 2002; 21: 523-31.
- OVERGAARD UM, HELWEG-LARSEN J: *Pneumocystis jirovecii* pneumonia (PCP) in HIV-1-negative patients: a retrospective study 2002-2004. *Scand J Infect Dis* 2007; 39: 589-95.
- TOKUDA H, SAKAI F, YAMADA H *et al.*: Clinical and radiological features of *Pneumocystis* pneumonia in patients with rheumatoid arthritis, in comparison with methotrexate pneumonitis and *Pneumocystis* pneumonia in acquired immunodeficiency syndrome: a multicenter study. *Intern Med* 2008; 47: 915-23.
- MORI S, SUGIMOTO M: *Pneumocystis jirovecii* infection: an emerging threat to patients with rheumatoid arthritis. *Rheumatology* 2012; 51: 2120-30.
- PERRUQUET JL, HARRINGTON TM, DAVIS DE: *Pneumocystis carinii* pneumonia following methotrexate therapy for rheumatoid arthritis. *Arthritis Rheum* 1983; 26: 1291-2.
- KANEKO Y, SUWA A, IKEDA Y, HIRAKATA M: *Pneumocystis jirovecii* pneumonia associated with low-dose methotrexate treatment for rheumatoid arthritis: report of two cases and review of the literature *Mod Rheumatol* 2006; 16: 36-8.
- WARD MM, DONALD F: *Pneumocystis carinii* pneumonia in patients with connective tissue diseases: the role of hospital experience in diagnosis and mortality. *Arthritis Rheum* 1999; 42: 780-9.
- ZHANG C, WANG SH, LASBURY ME *et al.*: Toll-like receptor 2 mediates alveolar macrophage response to *Pneumocystis murina*. *Infect Immun* 2006; 74: 1857-64.
- DOWNING JF, KACHEL DL, PASULA R, MARTIN WJ II: Gamma interferon stimulates rat alveolar macrophages to kill *Pneumocystis carinii* by L-arginine- and tumor necrosis factor-dependent mechanisms. *Infect Immun* 1999; 67: 1347-52.
- KOLLS JK, LEI D, VAZQUEZ C *et al.*: Exacerbation of murine *Pneumocystis carinii* infection by adenoviral-mediated gene transfer of a TNF inhibitor. *Am J Respir Cell Mol Biol* 1997; 16: 112-8.
- KAUR N, MAHL TC: *Pneumocystis jirovecii* (carinii) pneumonia after infliximab therapy: a review of 84 cases. *Dig Dis Sci* 2007; 52: 1481-4.
- LOUIE GH, WANG Z, WARD MM: Trends in hospitalizations for *Pneumocystis jirovecii* pneumonia among patients with rheumatoid arthritis in the US:1996-2007. *Arthritis Rheum* 2010; 62: 3826-7.
- SALMON-CERON D, TUBACH F, LORTHO-
- ARY O *et al.*: Drug-specific risk of non-tuberculous opportunistic infections in patients receiving anti-TNF therapy reported to the 3-year prospective French Ratio registry. *Ann Rheum Dis* 2011; 70: 616-23.
- FAVALLI EG, DESIATI F, ATZENI F *et al.*: Serious infections during anti-TNF alpha treatment in rheumatoid arthritis patients. *Autoimmun Rev* 2009; 8: 266-73.
- BRUCE ES, KEARSLEY-FLEET L, WATSON KD, SYMMONS DP, HYRICH KL: Risk of *Pneumocystis jirovecii* pneumonia in patients with rheumatoid arthritis treated with inhibitors of tumour necrosis factor α : results from the British Society for Rheumatology Biologics Register for Rheumatoid Arthritis. *Rheumatology* (Oxford) 2016; 55: 1336-37.
- KOIKE T, HARIGAI M, INOKUMA S *et al.*: Postmarketing surveillance of safety and effectiveness of etanercept in Japanese patients with rheumatoid arthritis. *Mod Rheumatol* 2011; 21: 343-51.
- TAKEUCHI T, TATSUKI Y, NOGAMI Y *et al.*: Postmarketing surveillance of the safety profile of infliximab in 5000 Japanese patients with rheumatoid arthritis. *Ann Rheum Dis* 2009; 67: 189-94.
- KOIKE T, HARIGAI M, ISHIGURO N *et al.*: Safety and effectiveness of adalimumab in Japanese rheumatoid arthritis patients: post-marketing surveillance report of the first 3,000 patients. *Mod Rheumatol* 2012; 22: 498-508.
- KOIKE T, HARIGAI M, INOKUMA S *et al.*: Postmarketing surveillance of tocilizumab for rheumatoid arthritis in Japan: interim analysis of 3881 patients. *Ann Rheum Dis* 2011; 70: 2148-51.
- KAY J, FLEISCHMANN R, KEYSTONE E *et al.*: Golimumab 3-year safety update: an analysis of pooled data from the long-term extensions of randomised, double-blind, placebo-controlled trials conducted in patients with rheumatoid arthritis, psoriatic arthritis or ankylosing spondylitis. *Ann Rheum Dis* 2015; 74: 538-46.
- BYKERK VP, CUSH J, WINTHROP K *et al.*: Update on the safety profile of certolizumab pegol in rheumatoid arthritis: an integrated analysis from clinical trials. *Ann Rheum Dis* 2015; 74: 96-103.
- TEICHMANN LL, WOENCKHAUS M, VOGEL C, SALZBERGER B, SCHOLMERICH J, FLECK M: Fatal *Pneumocystis* pneumonia following rituximab administration for rheumatoid arthritis. *Rheumatology* (Oxford) 2008; 47: 1256-7.
- VAN VOLLENHOVEN RF, EMERY P, BINGHAM CO 3RD *et al.*: Long-term safety of rituximab in rheumatoid arthritis: 9.5-year follow-up of the global clinical trial programme with a focus on adverse events of interest in RA patients. *Ann Rheum Dis* 2013; 72: 1496-502.
- TAK PP, RIGBY W, RUBBERT-ROTH A *et al.*: Sustained inhibition of progressive joint damage with rituximab plus methotrexate in early active rheumatoid arthritis: 2-year results from the randomised controlled trial IMAGE. *Ann Rheum Dis* 2012; 71: 351-7.
- KAMEDA H, TOKUDA H, SAKAI F *et al.*:

- Clinical and radiological features of acute-onset diffuse interstitial lung diseases in patients with rheumatoid arthritis receiving treatment with biological agents: importance of *Pneumocystis* pneumonia in Japan revealed by a multicenter study. *Intern Med* 2011; 50: 305-13.
37. IIKUNI N, KITAHAMA M, OHTA S, OKAMOTO H, KAMATANI N, NISHINARITA M: Evaluation of *Pneumocystis* pneumonia infection risk factors in patients with connective tissue disease. *Mod Rheumatol* 2006; 16: 282-8.
 38. HAHN PY, LIMPER AH: The role of inflammation in respiratory impairment during *Pneumocystis carinii* pneumonia. *Semin Respir Infect* 2003; 18: 40-7.
 39. GODEAU B, COUTANT-PERRONNE V, LE THI HUONG D *et al.*: *Pneumocystis carinii* pneumonia in the course of connective tissue disease: report of 34 cases. *J Rheumatol* 1994; 21: 246-51.
 40. SAITO K, NAKAYAMADA S, NAKANO K *et al.*: Detection of *Pneumocystis carinii* by DNA amplification in patients with connective tissue diseases: re-evaluation of clinical features of *P. carinii* pneumonia in rheumatic diseases. *Rheumatology* 2004; 43: 479-85.
 41. SATO T, INOKUMA S, MAEZAWA R *et al.*: Clinical characteristics of *Pneumocystis carinii* pneumonia in patients with connective tissue diseases. *Mod Rheumatol* 2005; 15: 191-7.
 42. SEPKOWITZ KA, BROWN AE, ARMSTRONG: *Pneumocystis carinii* pneumonia without acquired immunodeficiency syndrome. More patients, same risk. *Arch Intern Med* 1995; 155: 1125-8.
 43. YALE SH, LIMPER AH: *Pneumocystis carinii* pneumonia in patients without acquired immunodeficiency syndrome: associated illness and prior corticosteroid therapy. *Mayo Clin Proc* 1996; 71: 5-13.
 44. SOWDEN E, CARMICHAEL AJ: Autoimmune inflammatory disorders, systemic corticosteroids and pneumocystis pneumonia: a strategy for prevention. *BMC Infect Dis* 2004; 4: 42.
 45. OGAWA J, HARIGAI M, NAGASAKA K, NAKAMURA T, MIYASAKA N: Prediction of and prophylaxis against *Pneumocystis* pneumonia in patients with connective tissue diseases undergoing medium- or high-dose corticosteroid therapy. *Mod Rheumatol* 2005; 15: 91-6.
 46. ENOMOTO T, AZUMA A, KOHNO A *et al.*: Differences in the clinical characteristics of *Pneumocystis jirovecii* pneumonia in immunocompromised patients with and without HIV infection. *Respirology* 2010; 15: 126-31.
 47. FESTIC E, GAJIC O, LIMPER AH, AKSAMIT TR: Acute respiratory failure due to *Pneumocystis* pneumonia in patients without human immunodeficiency virus infection: outcome and associated features. *Chest* 2005; 128: 573-9.
 48. MORI S, CHO I, SUGIMOTO M: A cluster of *Pneumocystis jirovecii* infection among outpatients with rheumatoid arthritis. *J Rheumatol* 2010; 37: 1547-8.
 49. KOMANO Y, HARIGAI M, KOIKE R *et al.*: *Pneumocystis jirovecii* pneumonia in patients with rheumatoid arthritis treated with infliximab: a retrospective review and case-control study of 21 patients. *Arthritis Rheum* 2009; 61: 305-12.
 50. KATSUYAMA T, SAITO K, KUBO S, NAWATA M, TANAKA Y: Prophylaxis for *Pneumocystis* pneumonia in patients with rheumatoid arthritis treated with biologics, based on risk factors found in a retrospective study. *Arthritis Res Ther* 2014; 16: R43.
 51. YOSHIDA Y, TAKAHASHI Y, MINEMURA N *et al.*: Prognosis of *Pneumocystis* pneumonia complicated in patients with rheumatoid arthritis (RA) and non-RA rheumatic diseases. *Mod Rheumatol* 2012; 22: 509-14.
 52. MORRIS A, NORRIS KA: Colonization by *Pneumocystis jirovecii* and Its Role in Disease. *Clin Microbiol Rev* 2012; 25: 297-317.
 53. YASUOKA A, TACHIKAWA N, SHIMADA K, KIMURA S, OKA S: (1→3) β -d glucan as a quantitative serological marker for *Pneumocystis carinii* pneumonia. *Clin Diagn Lab Immunol* 1996; 3: 197-9.
 54. KOIKE R, TAKEUCHI T, EGUCHI K, MIYASAKA N: Update on the Japanese guidelines for the use of infliximab and etanercept in rheumatoid arthritis. *Mod Rheumatol* 2007; 17: 451-58.
 55. MORI S, CHO I, SUGIMOTO M: A follow-up study of asymptomatic carriers of *Pneumocystis jirovecii* during immunosuppressive therapy for rheumatoid arthritis. *J Rheumatol* 2009; 36: 1600-5.
 56. SURYAPRASAD A, STONE JH: When is it safe to stop *Pneumocystis jirovecii* pneumonia prophylaxis? Insights from three cases complicating autoimmune diseases. *Arthritis Rheum* 2008; 59: 1034-9.
 57. GREEN H, PAUL M, VIDAL L, LEIBOVICI L: Prophylaxis for *Pneumocystis* pneumonia (PCP) in non-HIV immunocompromised patients (Review). Copyright © 2011 The Cochrane Collaboration. Published by John-Wiley & Sons, Ltd.
 58. RODRIGUEZ M, FISHMAN JA: Prevention of infection due to *Pneumocystis* spp. in human immunodeficiency virus-negative immunocompromised patients. *Clin Microbiol Rev* 2004; 17: 770-82.
 59. INOKUMA S, OKADA Y, KANEI M *et al.*: Prophylaxis of *Pneumocystis jirovecii* pneumonia associated with inflammatory rheumatic diseases. Clinical guideline: the Japanese Ministry of Health, Laboratory and Welfare Study Group on complications and treatment of immune diseases. In: HASHIMOTO H (Ed.): Japanese Ministry of Health, Labor and Welfare Study Group, Tokio 2004: 24-29 (in Japanese).
 60. MORI S, SUGIMOTO M: Organizing pneumonia in rheumatoid arthritis patients: a case-based review. *Clin Med Insights Circ Respir Pulm Med.* 2015; 9 (Suppl. 1): 69-80.
 61. AL-AWADHI A, DALE P, MCKENDRY R: Pancytopenia associated with low dose methotrexate therapy. A regional survey. *J Rheumatol* 1993; 20: 1121-5.
 62. KATCHAMART W, BOURRE-TESSIER J, DONKA T *et al.*: Canadian recommendations for use of methotrexate in patients with rheumatoid arthritis. *J Rheumatol* 2010; 37: 1422-30.
 63. BOZZETTE SA, SATTTLER FR, CHIU J *et al.*: A controlled trial of early adjunctive treatment with corticosteroids for *Pneumocystis carinii* pneumonia in the acquired immunodeficiency syndrome. California Collaborative Treatment Group. *N Engl J Med* 1990; 323: 1451-57.
 64. WU AK, CHENG VC, TANG BS *et al.*: The unmasking of *Pneumocystis jirovecii* pneumonia during reversal of immunosuppression: case reports and literature review. *BMC Infect Dis* 2004; 4: 57.
 65. SAITO K, NAKAYAMADA S, NAKANO K *et al.*: Detection of *Pneumocystis carinii* by DNA amplification in patients with connective tissue diseases: reevaluation of clinical features of *P. carinii* pneumonia in rheumatic diseases. *Rheumatology* 2004; 43: 479-85.
 66. KWON-CHUNG KJ, VARMA A: Do major species concepts support one, two or more species within *Cryptococcus neoformans*? *FEMS Yeast Res* 2006; 6: 574-87.
 67. LA HOZ RM, PAPPAS PG: Cryptococcal infections: changing epidemiology and implications for therapy. *Drugs* 2013; 73: 495-504.
 68. ANTINORI S: New insights into HIV/AIDS-associated cryptococcosis. *ISRN AIDS* 2013; 471363.
 69. BARTLETT KH, KIDD SE, KRONSTAD JW: The emergence of *Cryptococcus gattii* in British Columbia and the Pacific Northwest. *Curr Infect Dis* 2008; 10: 58-65.
 70. ALTZ-SMITH M, KENDALL LG JR, STAMM AM: Cryptococcosis associated with low-dose methotrexate for arthritis. *Am J Med* 1987; 83: 179-81.
 71. TRUE DG, PENMETCHA M, PECKAM SJ: Disseminated cryptococcal infection in rheumatoid arthritis treated with methotrexate and infliximab. *J Rheumatol* 2002; 29: 1561-63.
 72. FURST DE, WALLIS R, BRODER M, BEENHOUWER DO: Tumor necrosis factor antagonists: different kinetics and/or mechanisms of action may explain differences in the risk for developing granulomatous infection. *Semin Arthritis Rheum* 2006; 36: 159-67.
 73. SCHIFF MH, BURMESTER GR, KENT JD *et al.*: Safety analyses of adalimumab (HUMIRA) in global clinical trials and US postmarketing surveillance of patients with rheumatoid arthritis. *Ann Rheum Dis* 2006; 65: 889-94.
 74. DESCALZO MA, BIOBADASER STUDY GROUP: Spanish registry of adverse events of biologic therapies in rheumatic diseases (BIOBADASER). Report of the situation on January 2006. *Reumatologia Clinica* 2007; 3: 4-20 (article in Spanish).
 75. CADENA J, THOMPSON GR III, HO TT, MED-INA E, HUGHES DW, PATTERSON TF: Immune reconstitution inflammatory syndrome after cessation of the tumor necrosis factor α blocker adalimumab in cryptococcal pneumonia. *Diagn Microbiol Infect Dis* 2009; 64: 327-30.
 76. HORCAJADA JP, PENA JL, MARTINEZ-TABOADA VM *et al.*: Invasive cryptococcosis and adalimumab treatment. *Emerg Infect Dis* 2007; 13: 953-5.
 77. YANAGAWA N, SAKAI F, TAKEMURA T *et al.*: Pulmonary cryptococcosis in rheumatoid arthritis (RA) patients : comparison of

- imaging characteristics among RA, acquired immunodeficiency syndrome, and immunocompetent patients. *Eur J Radiol* 2013; 82: 2035-42.
78. IWATA T, NAGANOT, TOMITA M *et al.*: Adalimumab-associated pulmonary cryptococcosis. *Ann Thorac Cardiovasc Surg* 2011; 17: 390-3.
 79. HAGE CA, WOOD KL, WINER-MURAM HT, WILSON SJ, SAROSI G, KNOX KS: Pulmonary cryptococcosis after initiation of anti-tumor necrosis factor- α therapy. *Chest* 2003; 124: 2395-97.
 80. STARRETT WG, CZACHOR J, DALLAL M, DREHMER T: Cryptococcal pneumonia following treatment with infliximab for rheumatoid arthritis. In: Program and abstracts of the 40th annual meeting of the Infectious Diseases Society of America. October 24-27, 2002 (Abstract 374).
 81. SHRESTA RK, STOLLER JK, HONARI G, PROCOP GW, GORDON SM: Pneumonia due to *Cryptococcus neoformans* in a patient receiving infliximab: possible zoonotic transmission from a pet cockatiel. *Respir care* 2004; 49: 606-8.
 82. AREND SM, KUIJPER EJ, ALLAART CF, MULLER WH, VAN DISSEL JT: Cavitating pneumonia after treatment with infliximab and prednisone. *Eur J Clin Microbiol Infect Dis* 2004; 23: 638-41.
 83. ANTINORI S, RADICE A, GALIMBERTI L, MAGNI C, FASAN M, PARRAVICINI C: The role of cryptococcal antigen assay in diagnosis and monitoring of cryptococcal meningitis. *J Clin Microbiol* 2005; 43: 5828-9.
 84. MUÑOZ P, PALOMO J, GUINEA J, YAÑEZ J, GIANNELLA M, BOUZA E: Cryptococcal meningitis in a patient treated with infliximab. *Diagn Microbiol Infect Dis* 2007; 57: 443-6.
 85. KOZIC H, RIGGS K, RINGPFEIL F, LEE JB: Disseminated *Cryptococcus neoformans* after treatment with infliximab for rheumatoid arthritis. *J Am Acad Dermatol* 2008; 58 (Suppl. 1): S95-6.
 86. ENG RH, PERSON A: Serum cryptococcal antigen determination in the presence of rheumatoid factor. *J Clin Microbiol* 1981; 14: 700-2.
 87. LINDSLEY MA, MEKHA N, BAGGETT HC *et al.*: Evaluation of a newly developed lateral flow immunoassay for the diagnosis of cryptococcosis. *Clin Infect Dis* 2011; 53: 321-5.
 88. MEYA DB, MANABE YC, CASTELNUOVO B *et al.*: Cost-effectiveness of serum cryptococcal antigen screening to prevent deaths among HIV-infected persons with a CD4⁺ cell count <100 cells/ μ L who start HIV therapy in resource-limited settings. *Clin Infect Dis* 2010; 51: 448-55.
 89. PERFECT JR, DISMUKES WE, DROMER F *et al.*: Clinical practice guidelines for the management of cryptococcal disease: 2010 update by the Infectious Diseases Society of America. *Clin Infect Dis* 2010; 50: 291-322.
 90. BADDLEY JW, FORREST GN, AND THE AST INFECTIOUS DISEASES COMMUNITY OF PRACTICE: Cryptococcosis in solid organ transplantation. *Am J Transpl* 2013; 13 (Suppl. 4): 242-9.
 91. NELSON M, MANJI H, WILKINS E: Central nervous system opportunistic infections. *HIV Medicine* 2011; 12 (Suppl. 2): 8-24.
 92. SINGH N: How I treat cryptococcosis in organ transplant recipients. *Transplantation* 2012; 93: 17-21.
 93. ARNING M, KLICHE KO, HEER-SONDERHOFF AH, WEHMEIER A: Infusion-related toxicity of three different amphotericin B formulations and its relation to cytokine plasma levels. *Mycoses* 1995; 38: 459-65.
 94. NGUYEN C, BARKER BM, HOOVER S *et al.*: Recent advances in our understanding of the environmental, epidemiological, immunological, and clinical dimensions of Coccidioidomycosis. *Clin Microbiol Rev* 2013; 26: 505-25.
 95. BADDLEY JW, WINTHROP KL, PATKAR NP *et al.*: Geographic distribution of endemic fungal infections among older persons, United States. *Emerg Infect Dis* 2011; 17: 1664-9.
 96. MERTZ LE, BLAIR JE: Coccidioidomycosis in rheumatology patients. Incidence and potential risk factors. *Ann NY Acad Sci* 2007; 1111: 343-57.
 97. BERGSTROM L, YOCUM DE, AMPEL NM *et al.*: Increased risk of coccidioidomycosis in patients treated with tumor necrosis factor α antagonists. *Arthritis Rheum* 2004; 50: 1959-66.
 98. TAROUMIAN S, KNOWLES SL, LISSE JR *et al.*: Management of coccidioidomycosis in patients receiving biologic response modifiers or disease-modifying antirheumatic drugs. *Arthritis Care Res* 2012; 64: 1903-9.
 99. ROGAN MP, THOMAS K: Fatal miliary coccidioidomycosis in a patient receiving infliximab therapy: a case report. *J Med Case Reports* 2007; 1: 79.
 100. SMITH JA, KAUFFMAN CA: Endemic fungal infections in patients receiving tumor necrosis factor- α inhibitor therapy. *Drugs* 2009; 69: 1403-15.
 101. DWEIK M, BAETHEGE BA, DUARTE AG: Coccidioidomycosis pneumonia in a nonendemic area associated with infliximab. *South Med J* 2007; 100: 517-8.
 102. RACLETTE A: Coccidioidomycosis infections in patients on TNF- α inhibitors. *American Academy of Dermatology Annual Meeting* 2007; Feb 2-6, Washington DC.
 103. CRUM NF, LEDERMAN ER, WALLACE MR: Infections associated with tumor necrosis factor- α antagonists. *Medicine* 2005; 84: 291-302.
 104. GARCIA-VIDAL C, RODRIGUEZ-FERNADEZ S, TEIJON S *et al.*: Risk factors for opportunistic infections in infliximab-treated patients: the importance of screening in prevention. *Eur J Clin Microbiol Infect Dis* 2009; 28: 331-7.
 105. BLAIR EJ, DOUGLAS DD, MULLIGAN DC: Early results of targeted prophylaxis for coccidioidomycosis in patients undergoing orthotopic liver transplantation within an endemic area. *Transpl Infect Dis* 2003; 5: 3-8.
 106. KAUFFMAN CA: Histoplasmosis : a clinical and laboratory update. *Clin Microbiol Rev* 2007; 20: 115-32.
 107. ASHBEE HR, EVANS EGV, VIVIANI MA *et al.*: Histoplasmosis in Europe: report on epidemiological survey from the European Confederation of Medical Mycology Working Group. *Med Mycol* 2008; 46: 57-65.
 108. ANTINORI S, MAGNI C, NEBULONI M *et al.*: Histoplasmosis among human immunodeficiency virus-infected people in Europe: report of 4 cases and review of the literature. *Medicine* 2006; 85: 22-36.
 109. ANTINORI S: *Histoplasma capsulatum*: more widespread than previously thought. *Am J Trop Med Hyg* 2014; 90: 982-3.
 110. BAHR NC, ANTINORI S, WHEAT LJ, SAROSI GA: Histoplasmosis infections worldwide: thinking outside of the Ohio River valley. *Curr Trop Med Rep* 2015; 2: 70-80.
 111. NAKELCHIK M, MANGINO JE: Reactivation of histoplasmosis after treatment with infliximab. *Am J Med* 2002; 112: 78.
 112. LEE J-H, SLIFMAN NR, GERSHON SK *et al.*: Life-threatening histoplasmosis complicating immunotherapy with tumor necrosis factor α antagonists infliximab and etanercept. *Arthritis Rheum* 2002; 46: 2565-70.
 113. WOOD KL, HAGE CA, KNOX KS *et al.*: Histoplasmosis after treatment with anti-tumor necrosis factor- α therapy. *Am J Respir Crit Care Med* 2003; 167: 1279-82.
 114. WALLIS RS, BRODER MS, WONG JY, HANSON ME, BEENHOUWER DO: Granulomatous infectious diseases associated with tumor necrosis factor antagonists. *Clin Infect Dis* 2004; 38: 1261-5.
 115. OLSON TC, BONGARTZ T, CROWSON CS *et al.*: Histoplasmosis infection in patients with rheumatoid arthritis , 1998-2009. *BMC Infect Dis* 2011; 11: 145.
 116. HAGE CA, BOWYER S, TARVIN SE, HELPER D, KLEIMAN MB, WHEAT LJ: Recognition, diagnosis and treatment of histoplasmosis complicating tumor necrosis factor blocker therapy. *Clin Infect Dis* 2010; 50: 85-92.
 117. VERGIDIS P, AVERY RK, WHEAT LJ *et al.*: Histoplasmosis complicating tumor necrosis factor- α blocker therapy: a retrospective analysis of 98 cases. *Clin Infect Dis* 2015; 61: 409-17.
 118. WHEAT LJ, HACKETT E, DURKIN M *et al.*: Histoplasmosis-associated cross-reactivity in the BioRad Platelia Aspergillus enzyme immunoassay. *Clin Vaccine Immunol* 2007; 14: 638-40.
 119. RIVIERE S, DENIS B, BOUGNOUX M-E, LANTERNIER F, LECUIT M, LORTHOLARY O: Serum *Aspergillus* galactomannan for the management of disseminated histoplasmosis in AIDS. *Am J Trop Med Hyg* 2012; 87: 303-5.
 120. HORIE M, TAMIYA H, GOTO Y *et al.*: Non-specific elevation of serum *Aspergillus* galactomannan antigen levels in patients with rheumatoid arthritis. *Respir Investig* 2016; 54: 44-9.
 121. WHEAT LJ, FREIFELD AG, KLEIMAN MB *et al.*: Clinical practice guidelines for the management of patients with histoplasmosis: 2007 update by the Infectious Diseases Society of America. *Clin Infect Dis* 2007; 45: 807-25.
 122. US FOOD AND DRUG ADMINISTRATION: Information for healthcare professionals: cimzia (certolizumab pegol), enbrel (etanercept),

- humira (adalimumab), and remicade (infliximab). 4 September 2008. Available at: <http://www.fda.gov/drugs/drugsafety/postmarket-drugsafetyinformationforpatientsandproviders/ucm124185.htm>
123. SMITH JA, KAUFFMAN CA: Endemic fungal infections in patients receiving tumor necrosis factor- α inhibitor therapy. *Drugs* 2009; 69: 1403-15.
 124. WARRIS A, BJORNEKLETT A, GAUSTAD P: Invasive pulmonary aspergillosis associated with infliximab therapy. *N Engl J Med* 2001; 344: 1099-100.
 125. DE ROSA FG, BONORA S, DI PERRI G: Tuberculosis and treatment with infliximab. *N Engl J Med* 2002; 346: 623-26.
 126. VAN DER KLOOSTER JM, BOSMAN RJ, OUDEMANS-VAN STRAATEN HM, VAN DER SPOEL JI, WESTER JP, ZANDSTRA DF: Disseminated tuberculosis, pulmonary aspergillosis and cutaneous herpes simplex infection in a patient with infliximab and methotrexate. *Intensive Care Med* 2003; 29: 2327-9.
 127. WALLIS RS, BRODER MS, WONG JY, HANSON ME, BEENHOUWER DO: Granulomatous infectious diseases associated with tumor necrosis factor antagonists. *Clin Infect Dis* 2004; 38: 1261-65.
 128. DENNING DW, CADRANEL J, BEIGELMAN-AUBRY C *et al.*: European Society for Clinical Microbiology and Infectious Diseases and European Respiratory Society. Chronic pulmonary aspergillosis: rationale and clinical guidelines for diagnosis and management. *Eur Respir J* 2016; 47: 45-68.
 129. ATZENI F, SARZI-PUTTINI P, MUTTI A, BUGATTI S, CAVAGNA L, CAPORALI R: Long-term safety of abatacept in patients with rheumatoid arthritis. *Autoimmun Rev* 2013; 12: 1115-7.
 130. ATZENI F, BENUCCI M, SALLÌ S, BONGIOVANNI S, BOCCASSINI L, SARZI-PUTTINI P: Different effects of biological drugs in rheumatoid arthritis. *Autoimmun Rev* 2013; 12: 575-9.
 131. MERONI PL, VALENTINI G, AYALA F, CATTANEO A, VALESINI G: New strategies to address the pharmacodynamics and pharmacokinetics of tumor necrosis factor (TNF) inhibitors: A systematic analysis. *Autoimmun Rev* 2015; 14: 812-29.
 132. GALICIA-HERNÁNDEZ G, PARRA-SALCEDO F, UGARTE-MARTÍNEZ P *et al.*: Sustained moderate-to-high disease activity and higher Charlson score are predictors of incidental serious infection events in RA patients treated with conventional disease-modifying anti-rheumatic drugs: a cohort study in the treat-to-target era. *Clin Exp Rheumatol* 2016; 34: 261-9.