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# Lipid Pathway Deregulation in Advanced Prostate Cancer

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# <u>Abstract</u>

The link between prostate cancer (PC) development and lipid metabolism is well established, with AR intimately involved in a number of lipogenic processes involving SREBP1, PPARG, FASN, ACC, ACLY and SCD1. Recently, there is growing evidence implicating the role of obesity and peri-prostatic adipose tissue (PPAT) in PC aggressiveness and related mortality, suggesting the importance of lipid pathways in both localised and disseminated disease. A number of promising agents are in development to target the lipogenic axis in PC, and the likelihood is that these agents will form part of combination drug strategies, with targeting of multiple metabolic pathways (e.g. FASN and CPT1), or in combination with AR pathway inhibitors (SCD1 and AR). Introduction

 Prostate cancer (PC) is the commonest adult male cancer in the developed world, and the second leading cause of cancer related death in men (1). The majority of patients are likely to die with, rather than from PC, making it important to identify key pathways that confer poor prognosis, thus minimising overtreatment.

Huggins *et al* in 1941 demonstrated that PC epithelial growth and survival was dependent upon androgens (2). Androgen deprivation therapy (or ADT) is often the first line treatment in patients with advanced disease. This is achieved via drug treatments designed to block androgen activity, either by direct suppression of the Luteinising Hormone Releasing Hormone (LHRH) or Androgen Receptor (AR) axis, thus, mimicking surgical castration. With time, PC develops resistance to these treatments and the disease progresses to CRPC (castrate resistant PC) form, which is uniformly fatal (3). Thus, a need exists to identify other signalling pathways in PC development and progression, and design specific treatments that exploit the dependencies and vulnerabilities of CRPC.

In 1953 Medes and colleagues (4) observed a relationship between lipid metabolism and cancer. They demonstrated that cancer tissues could generate fatty acids (FAs) and phospholipids through cellular *de novo* lipogenesis, and were not solely reliant upon lipid/FA uptake from the environment. This is turn provided the support required for the excessive growth and proliferation, which is a hallmark of cancer. The use of FAs in the cell can be utilised for the

generation of energy via their breakdown by β-oxidation to generate ATP. The energy demands within cancer cells are much higher than that for non-cancerous cells, at least in part to support uncensored growth and proliferation. However, the use of FAs to synthesise lipids is of equal importance. Membrane synthesis, which is a pre-requisite of growth and cell division, is linked to G1 phase of cell cycle (5). In G1 phase, cell cycle arrest results from suppressed expression of key lipid metabolism genes such as Fatty Acid Synthase (FASN) and Acetyl-CoA Carboxylase (ACC) (6). Besides membrane synthesis, lipogenesis is necessary for other functions (7). For instance, *de novo* synthesis of mono-unsaturated and saturated lipids plays key roles in signal transduction, intracellular trafficking, cell polarisation and migration (8-10). Each of these processes are often hijacked and deregulated in cancer cells to promote their survival. As such it is not unreasonable to think that disruption or blocking of the lipid metabolism pathways would be detrimental to tumour cell growth, proliferation and ultimately survival, thus representing potential therapeutic targets.

 Whilst androgens have long been established as a key player in PC, it has also been observed that advanced prostate tumours accumulate lipid droplets (11). It is now recognised that androgens may play a role in this due to effects they have on lipid metabolism (11, 12). Androgens regulate the mRNA and protein expression of one of the key regulators of lipid metabolism, the sterol regulatory element-binding protein (SREBP). SREBPs not only increase lipid metabolism, but also increase cholesterol metabolism (13), which in turn can aid androgen synthesis. It has also been observed that as well as utilising FA's for *de novo* lipogenesis, PC cells tend to use fatty acids over glucose as an energetic substrate

through increased  $\beta$ -oxidation and this is not the case in non-tumorigenic cell lines (14), making lipid metabolism an attractive as well as specific avenue for treatment.

Key Regulators of Lipid Metabolism in PC

#### Sterol Regulatory Element-Binding Protein 1 (SREBP1)

SREBP1 is a master regulator in FA metabolism. It controls the transcription of ATP Citrate Lyase (ACYL), ACC, Stearoyl-CoA Desaturase 1 (SCD1) and FASN. Un-regulated SREBP activation has been linked to obesity, fatty liver disease, insulin resistance, autoimmune diseases, as well as cancer development (15). It is frequently overexpressed in many cancers and is highly associated with increased tumorgenicity and invasion.

As previously mentioned, in PC androgens increase the activity of SREBP (13). This has recently been attributed to androgen receptor (AR) mediated transcription of SREBP-cleavage activating protein (SCAP). SCAP binds to SREBP, and a complex translocates from the endoplasmic reticulum (ER) to the Golgi apparatus, where the complex is cleaved by the proteases SP1&2, thus releasing SREBP from SCAP, with the N-terminal DNA binding and transcriptional activation domains of SREBP exposed for transcriptional functions on its target genes (16, 17). This situation is further compounded by the ability of SREBP, once activated, to further enhance AR expression through binding to a SRE (sterol regulatory element) present in the AR gene (18). SREBP has also been identified as an oxygen sensor in yeast (19). The SREBP pathway can monitor oxygen-dependent sterol synthesis as a measure of oxygen

availability, and control a transcriptional program required for adaptation to hypoxia, which is frequently found in solid tumours such as those of the prostate.

In addition to response to oxygen and androgens levels, SREBP is also activated by the AKT/PI3K pathway (20, 21). AKT signalling in a PTEN null environment (a situation common in PC with PTEN loss being a driver mutation in the disease) increases SREBP expression which in turn up-regulates expression of the Low-density Lipoprotein (LDL) receptor, thus increasing the uptake of particles, containing cholesteryl esters (CE). Depletion of this CE storage led to an impairment in PC aggressiveness, has been observed to attenuate cell growth, both *in vitro* and *in vivo* through limitation of the uptake of essential fatty acids (11).

In summary, SREBP is crucial factor in PC progression and interference with its associated pathways in PC may be a possible avenue for treatment of advanced disease. Physiologic inhibitory mechanisms already exist within the cell to prevent over-activation of SREBP. AMPK can phosphorylate SREBP, which prevents the proteolytic action of SP1 and SP2 in the Golgi apparatus, thus preventing SREBP1 activation (22). Another negative regulator of SREBP is Farnesoid X receptor (FXR). Upon its activation through ligand binding of Chenodexycholic acid (CDCA), FXR reduces the mRNA and proteins levels of SREBP, which in turn attenuates SREBP controlled lipid metabolism and consequently reduces tumour growth and proliferation (23). It has been observed that FXR inhibits co-activator recruitment to the SREBP promoter thereby reducing its expression and consequently affecting various downstream

effectors (24). Interestingly, FXR may also further impact on PC via upregulation of PTEN (25).

Regardless, as a master regulator of lipid metabolism it has been shown that by attenuating its function tumour growth and proliferation are diminished and thus it remains as an attractive drug target.

#### Peroxisome proliferator-activated receptor gamma (PPARG)

PPARG is a transcription factor belonging to the nuclear hormone receptor superfamily. It is known to have roles in adipocyte differentiation, lipid metabolism, peripheral glucose utilisation and inflammatory response. It has two isoforms PPARG1 and PPARG2. PPARG1 is expressed in most tissues whilst PPARG2 is present in adipocytes. Previous studies have demonstrated its role as a tumour suppressor in a variety of cancers, showing that upon treatment with PPARG agonists that proliferation of tumour cells is reduced (26-30). It was also thought to be the case in PC (31), however our work (32) and that of others (33) has challenged this view.

In our forward genetic screen using a murine transgenic mouse prostate cancer model driven by *Pten* deletion (32), PPARG was found to promote metastatic PC by associated up-regulation of the lipid metabolism pathways, more specifically those involved in *de novo* lipogenesis. Additionally, inhibition of PPARG supressed tumour growth and down-regulated the lipid synthesis pathway genes. PPARG levels were observed to correlate strongly with that of FASN, a key enzyme in the lipid synthesis pathway, and that high PPARG/FASN levels along with PTEN loss conferred poor prognosis. This finding could be used

therapeutically to stratify patients, identifying those with more aggressive
disease who would benefit from a PPARG/FASN derived treatment program.
Furthermore, our work as well as that of others suggests that PPARG does not
affect the initiation of the primary tumour (34), at least in the mouse models
examined, but has a role more specific to the development of aggressive
metastatic disease (32). In a separate study, a link was also established between
PPARG and PC progression, identifying Fatty acid binding protein 5 (FABP5) as a
potential agonist for PPARG, with increasing FABP5 and PPARG levels
correlating with disease severity (35). This is in line with earlier work where
FABP5 was found to be positively associated with an invasive more aggressive
phenotype, which could be abrogated by addition of PPARG inhibitor GW9662,
leading the authors to surmise that the metastatic effects they observed through
FABP5 over-expression resulted from an FABP5 delivery of fatty acid ligand to
nuclear membrane bound PPARG resulting in its activation (36).

 A further study also made the prostate specific observation regarding the role of PPARG as an oncogene (33) Whilst attempting to elucidate the mechanism of long-term warfarin (a vitamin K antagonist) and its role in reducing the risk of PC, they established a functional link between warfarin, AR and PPARG function. Their study demonstrated that warfarin could inhibit AR transcriptional activity, independent of its  $\gamma$ -carboxylation, through inhibition of PPARG signalling. This suggests that PPARG can act as a regulator of AR, with its inhibition causing reduction in PC growth and proliferation via AR. However, the authors were not able to demonstrate a direct effect of AR on PPARG. This is at odds with another recent study that demonstrates that AR can regulate the activity of PPARG,

showing that AR normally functions to supress PPARG expression within AR positive PC cells (37). These conflicting observations may simply be because of context and cell line differences, but highlight the need for further investigation into the intersecting regulatory pathways of PPARG and AR, as any potential therapy designed around these axes may lead to a worsening of the disease rather alleviating it. Indeed, it may be that a two-pronged approach is required, targeting both AR and PPARG simultaneously.

As PPARG has been a therapeutic target in disease areas other than cancer, there are already agents available in the clinic known to target PPARG. Thiazolidinediones (TZDs) or 'glitazones' are agonists of PPARG used in the treatment of type 2 diabetes, through improvements of insulin sensitivity (38). However, the concentration at which the TZDs are used to treat diabetes are far higher than the concentration required for full PPARG activation and thus the mechanism may in fact be due to a PPARG independent effect (39, 40). Given the link between diabetes and obesity, and the emerging evidence of the role of obesity and PC (to be discussed later) it is worth re-considering the potential risks of using a drug known to activate PPARG in (these obese and diabetic) men. PPARG represents an exciting new target for cancer therapy, but further investigation is needed to identify the subgroup of patients who would benefit from this targeted treatment.

#### Fatty acid synthase (FASN)

 FASN) is a key component of the lipid synthesis pathway and has been implicated in many cancers (41, 42). FAs are essential constituents of membrane

lipids, and are an essential substrate for energy metabolism. There are two sources of FAs for animal metabolism, namely exogenous (dietary) FAs and endogenous (FASN synthesised) FAs. FASN synthesises long-chain FAs from acetyl-CoA and malonyl-CoA, producing the 16C FA, palmitate (42). In healthy individuals, FASN has minimal effect since there is adequate levels of FA available from dietary fat. Thus, most normal cells will preferentially utilise circulating FA for the synthesis of new structural lipids. In normal conditions, FASN converts excess carbohydrate into FAs, which are then esterified to triacylglycerols that can be stored (and if needed provide energy via βoxidation).

 FASN has been shown to be one of the downstream effectors of the PTEN/PI3K/AKT pathway in the PC cell line LNCaP (43). Similarly, Migita et al demonstrated that forced overexpression of FASN increased cell proliferation both *in vitro* and *in vivo*, dependent on the presence of AR in the PC cells (44). Knock down of FASN in the same cells triggered apoptosis, suggesting that FASN can act as an oncogene in the presence of AR, and that FASN exerts its oncogenic influence by inhibiting apoptosis.

P300 (also known as EP300 or E1a binding protein 300) is an acetlytransferase that acts as a transcription co-activator and has been linked to PC growth. It is known to acetylate histone H3 lysine 27 (H3K27Ac) within the FASN gene promoter region, and studies have demonstrated that it acts to increase FASN expression, driving lipid accumulation and PC cell growth (45). Immumohistochemical (IHC) studies of FASN expression suggest that it is one of the earliest and commonest events in the development of PC (46, 47). As the

disease progresses, FASN levels correlate with Gleason Scores (tumour differentiation) and PSA levels (48).

 Upon epithelial to mesenchymal transition (EMT), a process crucial for metastasis, FASN levels appear to rise along with increases in lipid droplet and triacylglycerides (TAG) formation in DU145 PC cells (49). It remains unclear what role FASN plays in EMT or what the TAGs contained inside the lipid droplets are doing, but it is possible that the accumulation of stored TAGs may contribute to EMT through provision of fuel source with the generation of ATP as well as biomass for membrane synthesis (5). Inhibition of FASN has been observed to suppress both proliferation and key EMT phenotypes including cell adhesion, migration and invasion (50). FASN knockdown is observed to reduce the synthesis of phospholipids and triglycerides but not cholesterol (6).

Androgens have been observed to induce FASN expression and subsequent lipid accumulation *in vitro* in multiple PC cell lines (51). It is probable that this effect is presided over by SREBP and/or PPARG, however other androgen regulated factors may also have a role to play. Androgens may exert their effect on FASN through their ability to increase expression of ubiquitin-specific protease-2a (USP2a), an isopeptidase, which is able to stabilise FASN by deubiquitinating it at a preproteasomal level (52). Thus, androgens can induce FASN expression both through activation of SREBP and PPARG, but also further downstream by stabilisation of the resultant protein, allowing PC cells to achieve even greater levels of FASN expression. Aside from androgens, FASN expression has also been linked hypoxia due through the activation of Akt and SREBP1 in breast cancer, another hormone driven cancer (53).

The specific oncogenic nature of FASN in PC seems to mark it out as an ideal candidate for drug development (41). Its increased levels and function correlating with the most aggressive forms of the disease give promise that such a treatment would potentially be useful at all stages, from chemoprevention up to even the most severe cases.

#### Stearoyl-CoA desaturase 1 (SCD1)

SCD1 ( $\Delta$ -9-destaurase) is an endoplasmic reticulum (ER) enzyme that catalyses the rate-limiting step in the formation of mono-unsaturated FAs (MUFAs) from stearoyl-CoA and palmitoyl-CoA (54). These MUFAs (oleate and palmitoleate) are major components of membrane phospholipids and cholesterol esters. SCD1 is a key enzyme in FA metabolism, introducing a double bond at the  $\Delta$ 9 position in newly synthesised FAs.

Like FASN, SCD1 expression is regulated by SREBP. FASN acts upstream to produce saturated FAs, which SCD1 can then unsaturate. A recent study demonstrated that SCD1 inhibition altered the cellular lipid composition, and importantly impeded cell viability in the absence of exogenous lipids (55). Inhibition also altered cardiolipin composition, leading to the release of cytochrome C and induction of apoptosis. Silencing of SCD1 expression in a prostate orthograft model using LNCaP cells efficiently blocked tumour growth and significantly increased animal survival (55). This corresponds with previous studies where pharmacological inhibition of SCD1 impaired lipid synthesis by depleting MUFA and slowed PC xenograft growth in nude mice (56, 57).

Despite this it has also been observed that loss of SCD1 function can induce increased ER- and oxidative stress, bought about by accumulation of saturated fatty acids in membrane phospholipids, which induces an unfolded protein response (UPR) (58). Indeed, it has been shown that PC cells have increased levels of membrane lipid saturation which may protect from free radicals and chemotherapeutics (59). Intriguingly, out with its direct role in lipid metabolism, proteolytic cleavage of SCD1 protein generates a small peptide that has been shown to can exert a positive effect on the transcriptional activity of AR (60).

 The role of SCD1 in PC therefore seems twofold. Firstly, to function in its capacity as a desaturase to increase the levels of mono-unsaturated lipids in the cancer cell. This can meet the increased need for these lipids in rapidly dividing and growing cells. Secondly, upon proteolytic cleavage of SCD1, a small peptide fragment can enhance AR mediated signalling, thus further promoting PC growth and proliferation. If a therapy can be designed around this peptide it may be represent an opportunity to attenuate the effects of AR on PC.

## ATP Citrate Lyase (ACLY) and Acetyl-CoA Carboxylase (ACC)

ACC and ACLY are both up-stream of FASN in the lipid synthesis pathway. ACLY is responsible for the conversion of citrate (derived from the TCA cycle and metabolism of glucose) to acetyl CoA. Linking glucose metabolism to FA synthesis, ACC then takes the acetyl CoA produced by ACLY and converts it to malonyl-CoA, which can then be fed to FASN to generate saturated FA. Knockdown or chemical inhibition of either of these two enzymes has been

shown to inhibit the growth of a variety of solid tumours, including PC (6, 61-64). Both ACC and ACLY expression has been linked to androgens (13).
Reduction in ACLY levels by RNAi and the inhibitor SB-204990 has been observed to cause a dramatic reduction in growth of human PC3 orthografts.
This is due to their higher rate of glycolysis, and correspondingly high rate of glucose-dependent lipid synthesis, making them sensitive to ACLY inhibition (61).

 A recent study has demonstrated that it is possible to target the ACLY-AMPK-AR axes to sensitise CRPC cells to AR antagonism (65). A combined pharmacological approach with an AR antagonist and ACLY inhibition in CRPC cells promotes energetic stress and AMPK activation, resulting in further suppression of AR levels and target gene expression, inhibition of proliferation, and apoptosis.

#### Gross effects of the adiposity within the tumour microenvironment

Obesity is a risk factor in many cancers, including PC. Levels in males in developed countries are set to rise to 83% by 2025 (66). A recent meta-analysis has demonstrated that whilst not significantly correlated with PC incidence (RR, 1.00; 95% CI, 0.95–1.06), obesity correlates strongly with increased risk of developing aggressive PC (RR, 1.14; 95% CI, 1.04–1.25) and PC specific mortality (RR, 1.24; 95% CI, 1.15–2.33) (67).

Knowing that androgens are major drivers of PC, it is surprising that high BMI and visceral/subcutaneous fat content actually inversely correlates with testosterone levels (68). Consequently, in obese men, testosterone (androgen) levels are reduced. Therefore, it is surprising that obesity, as a low testosterone phenotype, correlates with PC growth and development.

The fat deposit closest to the prostate is the peri-prostatic adipose tissue (PPAT), which is found surrounding the prostate. PPAT volume, measured both on Magnetic Resonance Imaging (MRI) and Ultrasound (US), has been established as a potential biomarker for PC aggressiveness (69, 70). Periprostatic fat volume was found to be highest in patients at highest risk of developing Castrate Resistant Prostate Cancer (CRPC). This highlights a potential role of PPAT in predicting the effectiveness of ADT treatment.

 It has been observed that PC cells grown in conditioned media (CM) from PPAT have a significant increase in proliferation and motility *in vitro* (71, 72). This effect was specific to PPAT, with factors derived from alternative adipose CM sources showing minimal effect. The specific "adipokines" causing these effects in the PPAT CM has only recently begun to be elucidated. Matrix metalloproteinase (MMP) activity, known to be required for migration and metastasis, has been associated with PPAT (72). PPAT is able to promote tumour growth and migration through increased matrix metalloproteinase activity of MMP2 and MMP9, which are released into the tumour microenvironment (72). Furthermore, the expression level of the adipokine receptor CCR3 was found to increase in tandem with increasing volume of PPAT (69). Similarly, secretion of CCL7 from periprostatic adipocytes was found to promote the migration of CCR3 expressing PC cells *in vitro* and *in vivo* (73). In obesity, there is higher secretion of CCL7 by cancer associated adipocytes (CAA), which may mechanistically promote the development of locally advanced disease. The increased migration of PC cells was inhibited with suppression of the CCR3/CCL7 axis. Clinically, increasing expression of CCR3 is associated with

higher Gleason Sum Score, higher pathological tumour (T) stage, lymph node invasion and an increased risk of biochemical recurrence (73).

A variety of pro-inflammatory cytokines and chemokines are found to increase relative to levels of obesity; one such chemokine IL-6 has been shown to be associated with PC (74, 75) with increasing levels correlating with advanced aggressive castrate resistant metastatic disease (76, 77). IL-6 is produced by the adipose tissue surrounding the prostate, such as the PPAT, and is involved in regulation of proliferative responses and cell death (78). Following migration of tumour cells along this chemokine (IL-6) gradient and upon contact with the PPAT, it appears that PPAT volume is reduced, which may be due to re-modelling of the PPAT by the invading tumour (73). However, given that tumour cells have been observed to induce lipolysis in neighbouring adipocytes and thus parasitise their lipid stores to fuel tumour growth and proliferation (79, 80), it is also possible that this loss of PPAT upon contact with tumour is a result of the tumour utilising the fuel stored there to grow and divide.

Another fat deposit utilised by PC cells is bone marrow adipocytes. Given bone is a site to which PC preferentially metastasises, it raises the question as to whether this is related to the presence of the marrow fat cells. It is similarly hypothesised the reason that PC metastasises at a later stage compared to other cancers is because of the relative abundance of locally accessible fat stores, and only when these are exhausted do the PC cell metastasise to within proximity of local lipid rich marrow fat cells. These PC cells can then induce the marrow fat cells to undergo lipolysis, releasing free FAs and glycerol, the latter of which can

then feed into the glycolytic pathway of the PC cells (81). Highlighting the impact of tumour micro-environment, bone marrow adipocytes can alter the gene expression profile of PC cells to enhance utilisation of the glycolytic pathway with concurrent increase in lactate production, indicating a shift to a glycolytic metabolic profile, which is consistent with the Warburg Phenotype (81).

#### Fat metabolism targeted treatments for prostate cancer

 With the evidence linking PC to lipid metabolism growing, a number of treatment strategies targeting various stages of the pathway have been investigated.

Silibinin is compound that is isolated from the seeds of the milk thistle plant and is widely consumed for the liver health benefits it offers, including its use as a potential treatment in PC (82). Silibinin activates AMPK, which in turn phosphorylates SREBP preventing SREBP cleavage and its subsequent nuclear translocation and resultant activation of SREBP target genes (83, 84). This reduces lipid and cholesterol content in PC cells compared to benign prostate epithelial cells, making it a PC specific treatment option (84). Inactivation of SREBP1 by silibinin causes downstream reduction in expression levels and activities of multiple lipid and cholesterol metabolic genes; among them are FASN, ACYL, ACC, AMACR (an isomerase involved in the β-oxidation pathway of fatty acids) and HMGCR (an enzyme that is the rate limiting step in the mevalonate pathway that produces cholesterol) (83). Thus, silibinin acts to inhibit both lipid metabolism and cholesterol synthesis through SREBP

inhibition, halting proliferation and inducing cell cycle arrest, as well as preventing the development of androgen resistance in PC cells (84).

Another molecule, Fatostatin, a synthetic diarylthiazole derivative, is known to block adipogeneis through inhibition of SREBP (85). It has been observed to bind to SCAP, the escort protein of SREBP, blocking the ER-Golgi translocation of SREBP, and thus preventing its activation (85). Whilst this work has not been performed in PC, it represents another potential mechanism for blocking SREBP activation (86).

When considering PPARG we have already mentioned the differing opinions upon its role as an oncogene or a tumour suppressor. The evidence supporting its role as a tumour suppressor advocates the use of PPARG synthetic agonists for treatment of PC. Thiazolidinediones (TZDs) are synthetic PPARG agonists and have been successfully used in the treatment of type II diabetes, for review see (87). The premise behind their use being that upon treatment with TZD's PPARG is activated and in a dose dependant manner relieving the effects of hyperglycaemia. However further research has discovered that the concentrations at which TZD's are being used to treat diabetes is far higher than required for the full activation of PPARG (88). When applied at more physiologic (lower) concentrations for full PPARG activation, TZD was in fact protective against apoptosis, possibly through enhancing the cells' ability to maintain the mitochondrial membrane potential (88). This suggests that treatment with TZD's at high concentrations is not necessarily resulting in specifically PPARG driven effect and indeed the activation of PPARG in this context may be

counterintuitive. Recent work now appears to suggest that PPARG activation may be tumourigenic in PC (32, 33), and compounding this with the fact that those suffering type II diabetes are often overweight/obese, then an activation of PPARG by TZD could accelerate tumourigenesis.

Conflicting evidence surrounds the role of statins in PC, reviewed in (89). Studies of PC cell lines and animal models have shown that statins have antitumourigenic potential, by inhibiting proliferation and growth of PC cells (90, 91). Recent Danish registry based studies also demonstrated a positive role of statins in reducing PC mortality, both pre-and post-diagnosis (92, 93). A UK registry based study found that post-diagnosis statin use was associated with reduced PC mortality, particularly among patients who had used it prior to the diagnosis of PC (94). Two meta-analyses failed to find an association between statin use and PC recurrence among patients following radical prostatectomy or radiotherapy (95, 96). In contrast, a more recent meta-analysis demonstrated up to 25% risk reductions for PSA recurrence, and both PC-specific and overall survival (97). The recent Finnish Randomised Study for PC screening showed no reduction in PC mortality with pre-diagnosis statin use, whereas post-diagnosis use was associated with reduced mortality, especially in patients on Androgen Deprivation Treatment (ADT) (98). Another recent study examining selection bias found that once this was accounted for, statin use within 6 months of cancer diagnosis did *not* appear to improve 3-year cancer specific survival or overall survival (99).

FASN is now accepted as a *bona fide* oncogene. Inhibition of FASN has been found to cause selective apoptosis of cancer cell in multiple cancer types (100), however the mechanism behind this remains unclear. It is possible that loss of FASN affects membrane function, DNA replication or inhibition of anti-apoptotic proteins and/or the accumulation of Malonyl-CoA (101). The selective apoptotic effects caused by loss of FASN activity on cancer cells make it an attractive target for therapy. Cerulenin, a naturally derived inhibitor of FASN, is produced by a fungus *Cephalosporum caerulens*. Cerulenin binds the B-ketoacyl synthase domain of FASN to suppress its function. It is highly potent but is also unstable with toxic side effects. The synthetic analogue C75 of cerulenin was developed with a better side effect profile and greater stability (102). However, the major side effect of both C75 (and cerlenin) is dramatic and rapid weight loss seemingly resulting from stimulation of carnitine palmitoyltransferase-1 (CPT1), which activates mitochondrial fatty acid oxidation, is the limiting factor in developing these agents as cancer therapies (102, 103).

 Novel combination strategies with co-inhibition of FASN and AMPK have also been explored in pre-clinical models, with the use of AMPK inhibitor compound C (cC) and C75. Blocking lipid synthesis with concurrent AMPK inhibition, results in accumulation of toxic metabolites such as malonyl-CoA and NADPH as well as generating of toxic reactive oxygen species (ROS) inducing apoptosis and arrest of tumour cell proliferation (104).

Another naturally occurring FASN inhibitor exists in the form of various plant flavonoids, one of which is found in green tea, namely Epigallocatechin-3-gallate (EGCG). EGCG has been shown to block the formation of tumours in a range of

animal models (105). Similar to treatments with C75 and cerlenin, EGCG treatment also resulted excessive and speedy weight loss, possibly through activation of CPT1 (103). The use of CPT1 inhibitors such as etomoxir result in reduced PC growth *in vitro* and *in vivo*, so combination therapy with FASN inhibitors may allow inhibition of tumour growth whilst mitigating the unwanted side effects of weight loss. (106).

Orlistat, an anti-obesity drug, has also been found to inhibit the thioesterase domain of FASN; thus halting PC cell proliferation, inducing apoptosis and reducing tumour cell growth in nude mice (107). In its current formulation, Orlistat is limited as an anti-cancer therapy. It has a poor solubility and bioavailability, and when given orally is only functional in the areas it directly comes into contact with, inhibiting pancreatic lipases in the gut (107).

TVB-2640 is the first-in-class, small molecule reversible inhibitor of FASN that demonstrates Phase I clinical efficacy in KRAS mutant NSCLC, ovarian and breast cancer (108, 109). In this trial, prolonged stable disease was seen with monotherapy. In addition, when given in combination with paclitaxel, there is evidence of prolonged stable disease in both NSCLC and breast cancer patients, with a confirmed partial response in an patient with peritoneal serous carcinoma (108).

## **Conclusions**

The link between PC and fat metabolism is well established, with AR intimately involved as up- and down-stream factors (mediators) for a number of metabolic

enzymes. Furthermore, the evidence surrounding the risk of developing more advanced and aggressive PC with increased obesity and gross fat volume surrounding the prostate suggests the importance of lipid pathways not only on primary tumour growth but also on the development of advanced and metastatic disease.

Despite this, there is a paucity of agents in clinical trials for PC. It is likely that these agents will form part of combination drug strategies, with targeting of multiple metabolic pathways (e.g. FASN and CPT1), or in combination with AR pathway inhibitors (SCD1 and AR). Pre-clinical studies suggest this may improve "cancer kill" whilst reducing the toxic side effect profile.

1321	
1322	
1323	Poforoncos
1324	Kelefences
1325	
1326	1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2015. CA Cancer J Clin.
1327	2015;65(1):5-29.
1328	
1329	2. Huggins CaH, C.V. Studies on Prostatic Cancer: I. The effect of castration,
1330	of estrogen, and of androgen injection on serum phosphatases in metastatic
1331	carcinoma of the prostate. Cancer Res. 1941;1(4):293 -297.
1332	
1333	3. Feldman BI, Feldman D. The development of androgen-independent
1334	prostate cancer Nat Rev Cancer $2001\cdot1(1)\cdot34-45$
1335	
1336	A Medes C. Thomas A. Weinhouse S. Metabolism of neoplastic tissue, IV. A
1337	4. Medes d, Hiomas A, Weinhouse S. Metabolishi of heoplastic tissue. IV. A
1338	study of lipid synthesis in neoplastic tissue slices in vitro. Cancer Res.
1220	1953;13(1):27-29.
1339	
1340	5. Jackowski S. Coordination of membrane phospholipid synthesis with the
1341	cell cycle. J Biol Chem. 1994;269(5):3858-3867.
1342	
1343	6. Brusselmans K, De Schrijver E, Verhoeven G, Swinnen JV. RNA
1344	interference-mediated silencing of the acetyl-CoA-carboxylase-alpha gene
1345	induces growth inhibition and apoptosis of prostate cancer cells. Cancer Res.
1346	2005·65(15)·6719-6725
1347	2003,03(13).0713 0723.
1348	7 Swinnen IV Vanderboydong F. Flgamal AA. Felen M. Vergaeren I. Joniau S.
1349	Van Dennel II. Deart I. Coorsens V. Herne W. Verheeren C. Selective estivation of
1350	van Poppei H, Baert L, Goossens K, Heyns W, Vernoeven G. Selective activation of
1351	the fatty acid synthesis pathway in human prostate cancer. Int J Cancer.
1352	2000;88(2):176-179.
1353	
1354	8. Simons K, Toomre D. Lipid rafts and signal transduction. Nat Rev Mol Cell
1355	Biol. 2000;1(1):31-39.
1356	
1357	9. Manes S, Mira E, Gomez-Mouton C, Lacalle RA, Keller P, Labrador JP,
1358	Martinez AC. Membrane raft microdomains mediate front-rear polarity in
1359	migrating cells. EMBO I. 1999:18(22):6211-6220.
1360	
1361	10 Ikonen F. Simons K. Protein and linid sorting from the trans-Golgi
1362	notwork to the plasma membrane in polarized calls. Somin Coll Day Biol
1363	
1364	1998;9(5):503-509.
1365	
1366	11. Yue S, Li J, Lee SY, Lee HJ, Snao T, Song B, Cheng L, Masterson TA, Liu X,
1267	Ratliff TL, Cheng JX. Cholesteryl ester accumulation induced by PTEN loss and
1007	PI3K/AKT activation underlies human prostate cancer aggressiveness. Cell
1300	Metab. 2014;19(3):393-406.
1369	
13/0	12. Swinnen JV, Van Veldhoven PP, Esquenet M, Heyns W, Verhoeven G.
13/1	Androgens markedly stimulate the accumulation of neutral lipids in the human
1372	prostatic adenocarcinoma cell line LNCaP Endocrinology 1996.137(10).4468-
13/3	4474
13/4	1 1/ 1.
1375	
1376	
1377	
1378	
1379	23
1380	

1382		
1383	12 Continuous IV Illatia IV Usana IV Varia succe C. Coordinate regulation of	
1384	13. Swinnen JV, Ulrix W, Heyns W, Vernoeven G. Coordinate regulation of	
1385	lipogenic gene expression by androgens: evidence for a cascade mechanism	
1386	involving sterol regulatory element binding proteins. Proc Natl Acad Sci U S A.	
1207	1997:94(24):12975-12980	
1307		
1388	14 Lin V. Zuskier I.C. Chaseni NV. Dominant untake of fatty agid over gluce.	~~
1389	14. Liu Y, Zuckier LS, Ghesani NV. Dominant uptake of fatty actu over giucos	se
1390	by prostate cells: a potential new diagnostic and therapeutic approach.	
1391	Anticancer Res. 2010;30(2):369-374.	
1392		
1393	15. Shao W. Espenshade PL Expanding roles for SREBP in metabolism. Cell	
1394	Motoh $2012 \cdot 16(A) \cdot A1A - A19$	
1205	Metab. 2012,10(4).414-41).	
1395		
1396	16. Heemers H, verrijdt G, Organe S, Claessens F, Heyns W, vernoeven G,	
1397	Swinnen JV. Identification of an androgen response element in intron 8 of the	
1398	sterol regulatory element-binding protein cleavage-activating protein gene	
1399	allowing direct regulation by the androgen recentor I Biol Chem	
1400	$2004.270(20).30880_30887$	
1401	2004,279(29).30000-30007.	
1402		
1/03	17. Nohturfft A, Zhang SC. Coordination of lipid metabolism in membrane	
1403	biogenesis. Annu Rev Cell Dev Biol. 2009;25:539-566.	
1404		
1405	18. Huang WC. Zhau HE. Chung LW. Androgen receptor survival signaling is	s
1406	blocked by anti-beta2-microglobulin monoclonal antibody via a MAPK/linoger	nic
1407	biotked by and bedaz microgrobulin monocional and body via a Min K/npoger	пс (
1408	pathway in numan prostate cancer cells. J Biol Chem. 2010;285(11):/94/-/950	ь.
1409		_
1410	19. Hughes AL, Todd BL, Espenshade PJ. SREBP pathway responds to stero	ls
1411	and functions as an oxygen sensor in fission yeast. Cell. 2005;120(6):831-842.	
1/10		
1412	20 Porstmann T. Griffiths B. Chung YL. Delnuech O. Griffiths IR. Downward	I
1413	Schulzo A DKR / Alt induces transcription of anzumes involved in cholestorel a	nd
1414	Schulze A. I KD/AKt muutes it anschiption of enzymes myoryeu in choresteror a	nu
1415	fatty acid biosynthesis via activation of SREBP. Uncogene. 2005;24(43):6465-	
1416	6481.	
1417		
1418	21. Duvel K, Yecies JL, Menon S, Raman P, Lipovsky AI, Souza AL,	
1419	Triantafellow E. Ma O. Gorski R. Cleaver S. Vander Heiden MG. MacKeigan IP	
1420	Finan DM Click CD Murnhy I.O. Manning DD Activation of a matabalia gana	
1/20	Final PM, Chsh CD, Mulphy LO, Maining DD. Activation of a metabolic gene	4
1421	regulatory network downstream of mTOR complex 1. Mol Cell. 2010;39(2):17.	1-
1422	183.	
1423		
1424	22. Li Y, Xu S, Mihaylova MM, Zheng B, Hou X, Jiang B, Park O, Luo Z, Lefai E	,
1425	Shvy IY, Gao B, Wierzbicki M, Verheuren TI, Shaw RI, Cohen RA, Zang M, AMPK	
1426	nhoenhorylates and inhibits CDERD activity to attenuate honatic stastesis and	
1427	phosphorylates and minutes SKEDF activity to attenuate nepatic steatosis and	
1428	atherosclerosis in diet-induced insulin-resistant mice. Cell Metab.	
1429	2011;13(4):376-388.	
1430		
1/31	23. Liu N, Zhao J, Wang J, Teng H, Fu Y, Yuan H. Farnesoid X receptor ligand	
1422	CDCA suppresses human prostate cancer cells growth by inhibiting lipid	
1432	metabolism via targeting starol rosponse alament hinding protain 1 Am I Trar	nel
1433	netabolishi via targeting steroi response cicilicit bilitilig proteili 1. Alli j 11dli	131
1434	$Kes. 2010; \delta(11): 5110-5124.$	
1435		
1436		
1437		
1438		
1439		24

1441	
1442	
1443	
1444	24. Watanabe M, Houten SM, Wang L, Moschetta A, Mangelsdorf DJ, Heyman
1445	RA, Moore DD, Auwerx J. Bile acids lower triglyceride levels via a pathway
1446	involving FXR, SHP, and SREBP-1c, I Clin Invest, 2004:113(10):1408-1418.
1440	
1447	25 Liu I Tong SI Wang X Ou LX Farnesoid X recentor inhibits LNcaP cell
1448	proliferation via the unregulation of DTEN. Fur They Mod. 2014.0(4):1200-1212
1449	promeration via the upregulation of PTEN. Exp Ther Med. 2014;6(4):1209-1212.
1450	
1451	26. Sarraf P, Mueller E, Smith WM, Wright HM, Kum JB, Aaltonen LA, de la
1452	Chapelle A, Spiegelman BM, Eng C. Loss-of-function mutations in PPAR gamma
1453	associated with human colon cancer. Mol Cell. 1999;3(6):799-804.
1454	
1455	27. Sarraf P. Mueller E. Jones D. King FI. DeAngelo DJ. Partridge IB. Holden SA.
1456	Chen LB Singer S Eletcher C Sniegelman BM Differentiation and reversal of
1457	malignant changes in colon concer through DDAD gamma Nat Mod
1/58	1000 A(0) 10AC 1052
1450	1998;4(9):1046-1052.
1409	
1460	28. Tsubouchi Y, Sano H, Kawahito Y, Mukai S, Yamada R, Kohno M, Inoue K,
1461	Hla T, Kondo M. Inhibition of human lung cancer cell growth by the peroxisome
1462	proliferator-activated receptor-gamma agonists through induction of apoptosis.
1463	Riochem Biophys Res Commun 2000-270(2)-400-405
1464	
1465	20 Kulko MH Domotri CD Sharplace NE Dyap DD Shiydacani D Clark IS
1466	25. Kuike Mii, Deilieu I GD, Shai piess NE, Kyali DF, Shivuasani K, Gai K JS,
1467	Spiegelman BM, Kim H, Mayer RJ, Fuchs CS. A phase II study of trogitazone, an
1468	activator of the PPARgamma receptor, in patients with chemotherapy-resistant
1469	metastatic colorectal cancer. Cancer J. 2002;8(5):395-399.
1470	
1/71	30. Wang G, Cao R, Wang Y, Qian G, Dan HC, Jiang W, Ju L, Wu M, Xiao Y, Wang
1470	X Simvastatin induces cell cycle arrest and inhibits proliferation of bladder
1472	cancer cells via PPARgamma signalling nathway. Sci Ren. 2016:6:35783
1473	cancer cens via i i Argannia signannig pathway. Sei Rep. 2010,0.35703.
1474	21 Mueller E. Smith M. Sarref D. Kroll T. Aiver A. Kaufman DS. Oh W. Demetri
1475	51. Mueller E, Sillui M, Saltar P, Kroli T, Alyer A, Kauillan DS, Oli W, Delleu T
1476	G, Figg WD, Zhou XP, Eng C, Spiegelman BM, Kantoff PW. Effects of ligand
1477	activation of peroxisome proliferator-activated receptor gamma in human
1478	prostate cancer. Proc Natl Acad Sci U S A. 2000;97(20):10990-10995.
1479	
1480	32. Ahmad I, Mui E, Galbraith L, Patel R, Tan EH, Salji M, Rust AG, Repiscak P,
1481	Hedley A. Markert E. Loveridge C. van der Weyden L. Edwards I. Sansom OL
1482	Adams DI Leung HV Sleening Beauty screen reveals Phara activation in
1483	matastatic prostate capcor. Dree Natl Acad Cai II C A 2016.112(20).0200 0205
1484	metastatic prostate cancer. Proc Nati Acad Sci U S A. 2016;113(29):8290-8295.
1485	
1486	33. Tew BY, Hong TB, Otto-Duessel M, Elix C, Castro E, He M, Wu X, Pal SK,
1/187	Kalkum M, Jones JO. Vitamin K epoxide reductase regulation of androgen
1/10/	receptor activity. Oncotarget. 2017;8(8):13818-13831.
1400	
1409	34. Saez E, Olson P, Evans RM. Genetic deficiency in Pparg does not alter
1490	development of experimental prostate cancer Nat Med 2003-9(10)-1265-1266
1491	actorophiene of experimental prostate cancer. Nat Med. 2003, 9(10).1203-1200.
1492	25 Forestan ES Forestan SS Malli MI Chan D. Li C. Lin V. Dudland DS. Foster
1493	55. FOLOULAILTS, FOLOULAILSS, MIAIKI MI, UIEILD, LIUK, KUUIAIIUPS, FOSLEF
1494	CS, Ke 1. The expression of C-FABP and PPAKgamma and their prognostic
1495	significance in prostate cancer. Int J Oncol. 2014;44(1):265-275.
1496	
1497	
1498	
1499	25

1501	
1502	
1503	26 Pag 7 Malli MI Forgatan SS Adamson I Forgatan FS Chan D Foster CS
1504	50. Dau $L$ , Maiki Mi, Fulutionali 55, Audilisuli J, Fulutionali FS, Cheli D, Fusiel CS,
1505	Rudiand PS, Ke Y. A novel cutaneous Fatty Acid-binding protein-related signaling
1506	pathway leading to malignant progression in prostate cancer cells. Genes Cancer.
1507	2013;4(7-8):297-314.
1508	
1509	37. Olokpa E, Bolden A, Stewart LV. The Androgen Receptor Regulates
1510	PPARgamma Expression and Activity in Human Prostate Cancer Cells. J Cell
1511	Physiol. 2016:231(12):2664-2672.
1512	
1513	38. Hauner H. The mode of action of thiazolidinediones. Diabetes Metab Res
1514	Rev 2002:18 Suppl 2:S10-15
1515	Rev. 2002,10 Suppl 2.510 15.
1516	39 Akinyaka TO Stewart IV Troditazona suppresses c-Myc levels in human
1517	57. Akinyeke 10, stewart LV. Hogitazone suppresses c-Myc levels in human
1518	prostate cancer cens via a PPARgamma-independent mechanism. Cancer Biol
1510	Ther. 2011;11(12):1046-1058.
1519	
1520	40. Bolden A, Bernard L, Jones D, Akinyeke T, Stewart LV. The PPAR Gamma
1521	Agonist Troglitazone Regulates Erk 1/2 Phosphorylation via a PPARgamma-
1522	Independent, MEK-Dependent Pathway in Human Prostate Cancer Cells. PPAR
1523	research. 2012;2012:929052.
1524	
1525	41. Flavin R, Peluso S, Nguyen PL, Loda M. Fatty acid synthase as a potential
1526	therapeutic target in cancer. Future Oncol. 2010:6(4):551-562.
1527	
1528	42. Menendez IA, Lupu R, Fatty acid synthase and the lipogenic phenotype in
1529	cancer nathogenesis Nat Rev Cancer 2007.7(10).763-777
1530	cancer pathogenesis. Nat Nev Gancer. 2007,7(10).703 7777.
1531	43 Van de Sande T. De Schrijver F. Heyne W. Verheeven C. Swinnen IV. Pole
1532	45. Vali de Salide 1, De Schrijver E, Heylis W, Verhoeven G, Swinnen JV. Role
1533	of the phosphatidy most of 3 -kinase/PTEN/Akt kinase pathway in the
1534	overexpression of fatty acid synthase in LNCaP prostate cancer cells. Lancer Res.
1535	2002;62(3):642-646.
1536	
1537	44. Migita T, Ruiz S, Fornari A, Fiorentino M, Priolo C, Zadra G, Inazuka F,
1538	Grisanzio C, Palescandolo E, Shin E, Fiore C, Xie W, Kung AL, Febbo PG,
1539	Subramanian A, Mucci L, Ma J, Signoretti S, Stampfer M, Hahn WC, Finn S, Loda M.
1540	Fatty acid synthase: a metabolic enzyme and candidate oncogene in prostate
1541	cancer. I Natl Cancer Inst. 2009:101(7):519-532.
1542	,, ,, ,, ,, ,, ,, ,, ,, ,, ,, ,, ,, ,, ,, ,, ,,
1543	45. Gang X. Yang Y. Zhong I. Jiang K. Pan Y. Karnes RJ. Zhang J. Xu W. Wang G.
1544	Huang H P300 acetyltransferase regulates fatty acid synthase expression linid
1545	motabolism and prostate cancer growth Oncotarget 2016;7(12):15135-15140
1546	inetabolisin and prostate cancer growth. Oncotarget. 2010,7(12).13133-13149.
1547	46 Swinnen IV Rockame T Ioniau & Van Donnel U Oven D Roort I. House W
1548	To. Swinnen Jv, Roskanis I, Joinau S, van Fuppel II, Oyen R, Daeit L, Heylis W,
1549	vernoeven G. Overexpression of fatty acid synthase is an early and common
1550	event in the development of prostate cancer. Int J Cancer. 2002;98(1):19-22.
1551	
1552	47. Van de Sande T, Roskams T, Lerut E, Joniau S, Van Poppel H, Verhoeven G,
1553	Swinnen JV. High-level expression of fatty acid synthase in human prostate
1554	cancer tissues is linked to activation and nuclear localization of Akt/PKB. J
1555	Pathol. 2005;206(2):214-219.
1556	
1557	
1558	
1559	26

1561 1562 1563 48. Hamada S, Horiguchi A, Kuroda K, Ito K, Asano T, Miyai K, Iwaya K. 1564 Increased fatty acid synthase expression in prostate biopsy cores predicts higher 1565 Gleason score in radical prostatectomy specimen. BMC Clin Pathol. 2014;14(1):3. 1566 1567 Dalmau N, Jaumot J, Tauler R, Bedia C. Epithelial-to-mesenchymal 49. 1568 transition involves triacylglycerol accumulation in DU145 prostate cancer cells. 1569 Mol Biosyst. 2015;11(12):3397-3406. 1570 1571 50. Yoshii Y, Furukawa T, Oyama N, Hasegawa Y, Kiyono Y, Nishii R, Waki A, 1572 Tsuji AB, Sogawa C, Wakizaka H, Fukumura T, Yoshii H, Fujibayashi Y, Lewis JS, 1573 Saga T. Fatty acid synthase is a key target in multiple essential tumor functions of 1574 1575 prostate cancer: uptake of radiolabeled acetate as a predictor of the targeted 1576 therapy outcome. PLoS One. 2013;8(5):e64570. 1577 1578 51. Swinnen JV, Esquenet M, Goossens K, Heyns W, Verhoeven G. Androgens 1579 stimulate fatty acid synthase in the human prostate cancer cell line LNCaP. 1580 Cancer Res. 1997;57(6):1086-1090. 1581 1582 52. Graner E, Tang D, Rossi S, Baron A, Migita T, Weinstein LJ, Lechpammer M, 1583 Huesken D, Zimmermann J, Signoretti S, Loda M. The isopeptidase USP2a 1584 regulates the stability of fatty acid synthase in prostate cancer. Cancer Cell. 1585 2004;5(3):253-261. 1586 1587 53. Furuta E, Pai SK, Zhan R, Bandyopadhyay S, Watabe M, Mo YY, Hirota S, 1588 Hosobe S, Tsukada T, Miura K, Kamada S, Saito K, Iiizumi M, Liu W, Ericsson J, 1589 Watabe K. Fatty acid synthase gene is up-regulated by hypoxia via activation of 1590 Akt and sterol regulatory element binding protein-1. Cancer Res. 1591 2008;68(4):1003-1011. 1592 1593 54. Paton CM, Ntambi JM. Biochemical and physiological function of stearoyl-1594 CoA desaturase. Am J Physiol Endocrinol Metab. 2009;297(1):E28-37. 1595 1596 Peck B, Schug ZT, Zhang Q, Dankworth B, Jones DT, Smethurst E, Patel R, 1597 55. 1598 Mason S, Jiang M, Saunders R, Howell M, Mitter R, Spencer-Dene B, Stamp G, 1599 McGarry L, James D, Shanks E, Aboagye EO, Critchlow SE, Leung HY, Harris AL, 1600 Wakelam MJ, Gottlieb E, Schulze A. Inhibition of fatty acid desaturation is 1601 detrimental to cancer cell survival in metabolically compromised environments. 1602 Cancer Metab. 2016;4:6. 1603 1604 56. Fritz V, Benfodda Z, Rodier G, Henriquet C, Iborra F, Avances C, Allory Y, 1605 de la Taille A, Culine S, Blancou H, Cristol JP, Michel F, Sardet C, Fajas L. 1606 Abrogation of de novo lipogenesis by stearoyl-CoA desaturase 1 inhibition 1607 interferes with oncogenic signaling and blocks prostate cancer progression in 1608 mice. Mol Cancer Ther. 2010;9(6):1740-1754. 1609 1610 57. Mason P, Liang B, Li L, Fremgen T, Murphy E, Quinn A, Madden SL, 1611 Biemann HP, Wang B, Cohen A, Komarnitsky S, Jancsics K, Hirth B, Cooper CG, 1612 Lee E, Wilson S, Krumbholz R, Schmid S, Xiang Y, Booker M, Lillie J, Carter K. 1613 1614 SCD1 inhibition causes cancer cell death by depleting mono-unsaturated fatty 1615 acids. PLoS One. 2012;7(3):e33823. 1616 1617 1618 27 1619

1622	
1623	70 Arizona II Kana N. Matauda C. Inaua T. Arai II. Dagraaga in membrana
1624	58. Ariyama H, Kono N, Matsuda S, Inoue T, Arai H. Decrease in memorane
1625	phospholipid unsaturation induces unfolded protein response. J Biol Chem.
1626	2010;285(29):22027-22035.
1627	
1628	59. Rysman E, Brusselmans K, Scheys K, Timmermans L, Derua R, Munck S,
1629	Van Veldhoven PP, Waltregny D, Daniels VW, Machiels J, Vanderhovdonc F,
1630	Smans K. Waelkens E. Verhoeven G. Swinnen IV. De novo lipogenesis protects
1631	cancer cells from free radicals and chemotheraneutics by promoting membrane
1632	linid caturation Cancor Dog 2010.70(20).9117 9126
1633	iipiu saturation. Cancer Res. 2010,70(20).0117-0120.
1624	(0 Vim CL Choi IL Dark CC Chang C Vim E Steanard Co (departuress (CCD)
1034	60. KIIII SJ, CHOI H, PAIK SS, Chang C, KIIII E. Stear Oyr COA desaturase (SCD)
1030	facilitates proliferation of prostate cancer cells through enhancement of
1636	androgen receptor transactivation. Mol Cells. 2011;31(4):371-377.
1637	
1638	61. Hatzivassiliou G, Zhao F, Bauer DE, Andreadis C, Shaw AN, Dhanak D,
1639	Hingorani SR, Tuveson DA, Thompson CB. ATP citrate lyase inhibition can
1640	suppress tumor cell growth. Cancer Cell. 2005;8(4):311-321.
1641	
1642	62. Bauer DE, Hatzivassiliou G, Zhao F, Andreadis C, Thompson CB, ATP
1643	citrate lyase is an important component of cell growth and transformation
1644	On $cogono 2005 \cdot 24(41) \cdot 6314 - 6322$
1645	Oncogene. 2003, 24(41).0314-0322.
1646	62 Doarco NI Vatoc IW Borkhout TA Jackson B. Tow D. Boud H. Camillori D.
1647	Curean and D. Crickille AD. Cheve A. Creat DIL The role of ATD situate luces in the
1648	Sweeney P, Gribble AD, Snaw A, Groot PH. The role of ATP citrate-lyase in the
1649	metabolic regulation of plasma lipids. Hypolipidaemic effects of SB-204990, a
1650	lactone prodrug of the potent ATP citrate-lyase inhibitor SB-201076. Biochem J.
1651	1998;334 ( Pt 1):113-119.
1652	
1653	64. Migita T, Narita T, Nomura K, Miyagi E, Inazuka F, Matsuura M, Ushijima
1654	M, Mashima T, Seimiya H, Satoh Y, Okumura S, Nakagawa K, Ishikawa Y. ATP
1655	citrate lyase: activation and therapeutic implications in non-small cell lung
1656	cancer Cancer Res 2008:68(20):8547-8554
1657	
1658	65 Shah S. Carriveau WI, Li I. Campbell SL, Koninski PK, Lim HW, Daurio N
1659	Trofoly S Won KI Wallace DC Koumonis C Mancuso A Wallon KE Targeting
1660	ACLY consisting a contraction resistant prostate cancer calls to AD antagonism by
1661	ACLT Selisitizes castiation-resistant prostate cancer tens to AK antagonisin by
1662	impinging on an ACLY-AMPK-AR feedback mechanism. Uncotarget.
1663	2016;7(28):43713-43730.
1664	
1665	66. Wilson KM, Giovannucci EL, Mucci LA. Lifestyle and dietary factors in the
1666	prevention of lethal prostate cancer. Asian J Androl. 2012;14(3):365-374.
1667	
1668	67. Zhang X, Zhou G, Sun B, Zhao G, Liu D, Sun J, Liu C, Guo H. Impact of
1669	obesity upon prostate cancer-associated mortality: A meta-analysis of 17 cohort
1670	studies. Oncol Lett. 2015;9(3):1307-1312.
1670	
1672	68. Chavarro JE, Toth TL, Wright DL. Meeker ID. Hauser R. Body mass index in
1673	relation to semen quality, sperm DNA integrity, and serum reproductive
1674	hormone levels among men attending an infertility clinic Fortil Steril
1675	2010.02(7).2222.2221
1676	2010,73(7).2222 <sup>-</sup> 2231.
10/0	
10//	
10/0	20
10/9	28

1681	
1682	
1683	
1684	69. Salji M, Hendry J, Patel A, Ahmad I, Nixon C, Leung HY. Peri-prostatic Fat
1685	Volume Measurement as a Predictive Tool for Castration Resistance in Advanced
1686	Prostate Cancer. Eur Urol Focus. 2017.
1000	
1007	70 Rhindi B. Trottier G. Flharram M. Fernandes KA. Lockwood G. Toi A
1688	Hereasy KM Einelli A Evene A van den Kuvest TH Elechnen NE Meesurement of
1689	Hersey KM, Finein A, Evans A, van der Kwast TH, Flesnner NE. Measurement of
1690	peri-prostatic fat thickness using transrectal ultrasonography (TRUS): a new risk
1691	factor for prostate cancer. BJU Int. 2012;110(7):980-986.
1692	
1693	71. Venkatasubramanian PN, Brendler CB, Plunkett BA, Crawford SE, Fitchev
1694	PS Morgan & Cornwell MI McGuire MS Wyrwicz AM Doll IA Periprostatic
1605	adinaga tiggua from abaga prostate concernationts promotes tumor and
1606	aupose tissue nom obese prostate cancer patients promotes tumor and
1090	endothelial cell proliferation: a functional and MR imaging pilot study. Prostate.
1097	2014;74(3):326-335.
1698	
1699	72. Ribeiro R, Monteiro C, Cunha V, Oliveira MJ, Freitas M, Fraga A, Principe P,
1700	Lobato C. Lobo F. Morais A. Silva V. Sanches-Magalhaes I. Oliveira I. Pina F. Mota-
1701	Dista A Lanas C Madairas D Human navinnastatis adinasa tissua promotos
1702	Pinto A, Lopes C, Medenos R. Human periprostatic adipose tissue promotes
1703	prostate cancer aggressiveness in vitro. J Exp Clin Cancer Res. 2012;31:32.
1704	
1704	73. Laurent V, Guerard A, Mazerolles C, Le Gonidec S, Toulet A, Nieto L, Zaidi
1705	F. Maied B. Garandeau D. Socrier Y. Golzio M. Cadoudal T. Chaoui K. Drav C.
1706	Monsarrat B Schiltz O Wang VV Couderc B Valet P Malavaud B Muller C
1/0/	Devine estation edine estado estado e devining force for exectato concer exectation in
1708	Periprostatic adipocytes act as a driving force for prostate cancer progression in
1709	obesity. Nat Commun. 2016;7:10230.
1710	
1711	74. Weisberg SP, McCann D, Desai M, Rosenbaum M, Leibel RL, Ferrante AW,
1712	Ir. Obesity is associated with macrophage accumulation in adipose tissue. I Clin
1713	Invest 2003.112(12).1796-1808
1714	11/030.2003,112(12).1790 1000.
1715	75 Culiz 7 Droinflommatory autolying interlevelin 6 in prostate
1715	75. Cung Z. Prominanimatory cytokine interfeukin-6 in prostate
1710	carcinogenesis. Am J Clin Exp Urol. 2014;2(3):231-238.
1/1/	
1718	76. Adler HL, McCurdy MA, Kattan MW, Timme TL, Scardino PT, Thompson
1719	TC. Elevated levels of circulating interleukin-6 and transforming growth factor-
1720	heta1 in natients with metastatic prostatic carcinoma [IIro] 1999.161(1).182-
1721	
1722	107.
1723	
1724	//. Wise GJ, Marella VK, Talluri G, Shirazian D. Cytokine variations in patients
1724	with hormone treated prostate cancer. J Urol. 2000;164(3 Pt 1):722-725.
1720	
1/20	78. Yu SH, Zheng Q, Esopi D. Macgregor-Das A. Luo I. Antonarakis ES. Drake
1727	CG Vessella R Morrissev C De Marzo AM Sfanos KS A Paracrine Role for II 6 in
1728	Droctate Cancer Dationte: Lack of Droduction by Drimowy or Metactatic Tymer
1729	Colle Concertainen al D. 2015 2(10) 1155 1100 UNITALY OF MELASTATIC FUITION
1730	Cells. Cancer Immunol Res. 2015;3(10):1175-1184.
1731	
1732	79. Duncan RE, Ahmadian M, Jaworski K, Sarkadi-Nagy E, Sul HS. Regulation
1733	of lipolysis in adipocytes. Annu Rev Nutr. 2007;27:79-101.
1734	
1725	80 Nieman KM Kenny HA Penicka CV Ladanyi A Ruell-Guthrod R 7illhardt
1700	MP Romoro II Carou MS Mills CP Hotamioligil CS Vamada CD Dotor ME Curin
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1/3/	
1738	
1739	29
1740	

1741	
1742	
1743	
1744	K, Lengyel E. Adipocytes promote ovarian cancer metastasis and provide energy
1745	for rapid tumor growth. Nat Med. 2011;17(11):1498-1503.
1746	
1747	81. Diedrich JD, Rajagurubandara E, Herroon MK, Mahapatra G, Huttemann
1748	M, Podgorski I. Bone marrow adipocytes promote the Warburg phenotype in
1749	metastatic prostate tumors via HIF-1alpha activation. Oncotarget.
1750	2016:7(40):64854-64877
1751	
1751	82 Deep C. Agarwal R. Targeting tumor microenvironment with silibining
1752	oz. Deep 0, Agai wai K. Targeting tunior intervention of the sinonim.
1700	promise and potential for a translational cancer chemopreventive strategy. Curr
1754	Cancer Drug Targets. 2013;13(5):486-499.
1755	
1756	83. Nambiar DK, Rajamani P, Singh RP. Silibinin attenuates ionizing radiation-
1757	induced pro-angiogenic response and EMT in prostate cancer cells. Biochem
1758	Biophys Res Commun. 2015;456(1):262-268.
1759	
1760	84. Nambiar DK, Deep G, Singh RP, Agarwal C, Agarwal R. Silibinin inhibits
1761	aberrant lipid metabolism, proliferation and emergence of androgen-
1762	independence in prostate cancer cells via primarily targeting the sterol response
1763	alement binding protoin 1. On sotarget 2014 E (20):10017 10022
1764	element binding protein 1. Oncotarget. 2014;5(20):10017-10055.
1765	
1766	85. Kamisuki S, Mao Q, Abu-Eineiga L, Gu Z, Kugimiya A, Kwon Y, Shinonara T,
1767	Kawazoe Y, Sato S, Asakura K, Choo HY, Sakai J, Wakil SJ, Uesugi M. A small
1768	molecule that blocks fat synthesis by inhibiting the activation of SREBP. Chem
1769	Biol. 2009;16(8):882-892.
1770	
1771	86. Sigingaowa, Sekar S, Gopalakrishnan V, Taghibiglou C. Sterol regulatory
1772	element-binding protein 1 inhibitors decrease pancreatic cancer cell viability
1772	and proliferation Biochem Biophys Res Commun 2017:488(1):136-140
1773	
1775	87 Olofely IM Trantmont of inculin registrance with perovisions proliferator.
1773	orizated resenter samma aganista I Clin Invest 2000.106(4).467.472
1770	activated receptor gamma agomsts. J chin myest. 2000;100(4):407-472.
1///	
1778	88. Wang YL, Frauwirth KA, Rangwala SM, Lazar MA, Thompson CB.
1779	Thiazolidinedione activation of peroxisome proliferator-activated receptor
1780	gamma can enhance mitochondrial potential and promote cell survival. J Biol
1781	Chem. 2002;277(35):31781-31788.
1782	
1783	89. Alfaqih MA, Allott EH, Hamilton RJ, Freeman MR, Freedland SJ. The
1784	current evidence on statin use and prostate cancer prevention: are we there vet?
1785	Nature reviews Urology 2017-14(2)-107-119
1786	nuture reviews of ology. 2017,11(2).107-117.
1787	90 Murtola TI Pennanen P. Suvala H. Blauer M. Vlikomi T. Tammela TI
1788	Fifests of simulation a setulational solution and resignitation on preliferation of
1789	Effects of similastatin, acetyisancync aciu, and fosigntazone on promeration of
1790	normal and cancerous prostate epitnelial cells at therapeutic concentrations.
1791	Prostate. 2009;69(9):1017-1023.
1792	
1793	91. Hoque A, Chen H, Xu XC. Statin induces apoptosis and cell growth arrest in
1794	prostate cancer cells. Cancer Epidemiol Biomarkers Prev. 2008;17(1):88-94.
1795	
1796	
1797	
1798	
1799	.30

- 1802 1803 92. Nielsen SF, Nordestgaard BG, Bojesen SE. Statin use and reduced cancer-1804 related mortality. N Engl J Med. 2012;367(19):1792-1802. 1805 1806 93. Larsen SB, Dehlendorff C, Skriver C, Dalton SO, Jespersen CG, Borre M, 1807 Brasso K, Norgaard M, Johansen C, Sorensen HT, Hallas J, Friis S. Postdiagnosis 1808 Statin Use and Mortality in Danish Patients With Prostate Cancer. J Clin Oncol. 1809 2017: JCO2016718981. 1810 1811 94. Yu O, Eberg M, Benayoun S, Aprikian A, Batist G, Suissa S, Azoulay L. Use 1812 1813 of statins and the risk of death in patients with prostate cancer. J Clin Oncol. 2014;32(1):5-11. 1814 1815 Scosyrev E, Tobis S, Donsky H, Wu G, Joseph J, Rashid H, Messing E. Statin 1816 95. 1817 use and the risk of biochemical recurrence of prostate cancer after definitive 1818 local therapy: a meta-analysis of eight cohort studies. BJU Int. 2013;111(3 Pt 1819 B):E71-77. 1820 1821 96. Park HS, Schoenfeld JD, Mailhot RB, Shive M, Hartman RI, Ogembo R, 1822 Mucci LA. Statins and prostate cancer recurrence following radical 1823 prostatectomy or radiotherapy: a systematic review and meta-analysis. Ann 1824 Oncol. 2013;24(6):1427-1434. 1825 1826 97. Raval AD, Thakker D, Negi H, Vyas A, Salkini MW. Association between 1827 statins and clinical outcomes among men with prostate cancer: a systematic 1828 review and meta-analysis. Prostate Cancer Prostatic Dis. 2016;19(2):222. 1829 1830 98. Murtola TJ, Peltomaa AI, Talala K, Maattanen L, Taari K, Tammela TLJ, 1831 Auvinen A. Statin Use and Prostate Cancer Survival in the Finnish Randomized 1832 Study of Screening for Prostate Cancer. Eur Urol Focus. 2016. 1833 1834 99. Emilsson L, Garcia-Albeniz X, Logan RW, Caniglia EC, Kalager M, Hernan 1835 MA. Examining Bias in Studies of Statin Treatment and Survival in Patients With 1836 1837 Cancer. JAMA Oncol. 2017. 1838 Kuhajda FP. Fatty acid synthase and cancer: new application of an old 1839 100. 1840 pathway. Cancer Res. 2006;66(12):5977-5980. 1841 1842 Pizer ES, Thupari J, Han WF, Pinn ML, Chrest FJ, Frehywot GL, Townsend 101. 1843 CA, Kuhajda FP. Malonyl-coenzyme-A is a potential mediator of cytotoxicity 1844 induced by fatty-acid synthase inhibition in human breast cancer cells and 1845 xenografts. Cancer Res. 2000;60(2):213-218. 1846 1847 102. Kuhajda FP, Pizer ES, Li JN, Mani NS, Frehywot GL, Townsend CA. 1848 Synthesis and antitumor activity of an inhibitor of fatty acid synthase. Proc Natl 1849 Acad Sci U S A. 2000;97(7):3450-3454. 1850 1851 103. Nicot C, Napal L, Relat J, Gonzalez S, Llebaria A, Woldegiorgis G, Marrero 1852 PF, Haro D. C75 activates malonyl-CoA sensitive and insensitive components of 1853 the CPT system. Biochem Biophys Res Commun. 2004;325(3):660-664. 1854 1855 1856 1857 1858 1859
- 1860

1861	
1862	
1863	104 Fritz V Benfodda Z Henriquet C Hure S Cristol IP Michel F Carbonneau
1864	MA Casas E Egias I Metabolic intervention on linid synthesis converging
1865	ma, casas r, rajas E. Metabolic intervention on input synthesis converging
1866	pathways abrogates prostate cancer growth. Uncogene. 2013;32(42):5101-5110.
1867	
1868	105. Yang CS, Wang X, Lu G, Picinich SC. Cancer prevention by tea: animal
1869	studies, molecular mechanisms and human relevance. Nat Rev Cancer.
1870	2009;9(6):429-439.
1871	
1872	106. Schlaepfer IR, Rider L, Rodrigues LU, Gijon MA, Pac CT, Romero L, Cimic A,
1873	Sirintranun SI Clode I M Eckel RH Cramer SD Linid catabolism via CPT1 as a
1974	thereporties target for prostate sanger Mol Cancer Ther 2014.12(10).2261
1074	
1070	23/1.
1876	
1877	107. Kridel SJ, Axelrod F, Rozenkrantz N, Smith JW. Orlistat is a novel inhibitor
1878	of fatty acid synthase with antitumor activity. Cancer Res. 2004;64(6):2070-
1879	2075.
1880	
1881	108. Dean EL Falchook GS. Patel MR. Brenner Al Infante IR. Arkenau HT
1882	Borazanci FH Lonoz IS Dant S Schmid D Erankal AE Janas SE MaCullach M
1883	borazanci En, Lopez JS, Fant S, Schning F, Flanker AE, Jones SF, McCunoch W,
1884	Kemble G, O Farrell M, H. B. Preliminary activity in the first in human study of the
1885	first-in-class fatty acid synthase (FASN) inhibitor, TVB-2640. J Clin Oncol.
1886	2016;34(Supplement):2512.
1887	
1007	109. O'Farrell M, Heuer T, Grimmer K, Crowley R, Waszczuk J, Fridlib M,
1888	Ventura R. Rubio C. Lai I. Ruckley D. McCulloch W. Kemble G. Abstract I.B-214
1889	EASN inhibitor TVD 2640 shows pharmacodynamic affect and avidance of
1890	FASIN IIIIIDILOI TVD-2040 SIOWS pilatiliacouylialiic effect and evidence of
1891	clinical activity in KRAS-mutant NSCLC patients in a phase I study. Cancer Res.
1892	2016;76(14 Supplement).
1893	
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1923	Figure 1. Interaction of lipid notherways in prestate concer
1924	Figure 1: Interaction of lipid pathways in prostate cancer
1925	Some key regulatory proteins PPARG, SREBP and SCAP (red empses) govern
1926	fatty acid metabolism, the activity of these three is observed to fall under the
1927	overall control of Androgens and Androgen Receptor activation (blue ellipse,
1928	solid black arrows). Downstream of this hub of control (solid black arrows) are
1929	the effector of lipid synthesis, ACC, ACLY, FASN and SCD1 (red ellipses with
1930	dashed black arrows showing progression through the pathway) un-regulation
1931	of these effectors is also implicated in Prostate Cancer (PC) progression Several
1932	therapoutic agents Warfarin Estactatin Orlistat (groop rootangles red lines) are
1933	line apeutic agents, wai fai in, ratostatin, Offistat (green rectangles reu nies) are
1934	known to block key processes in lipid metabolism and have a negative effect on
1935	PC progression. In addition to those proteins resident within the prostate
1036	tumour cells themselves, within the microenvironment including nearby peri-
1037	prostatic fat, there are various other factors (yellow ellipses) that promote
1029	prostate cancer growth and progression. Matrix metallo-proteases, MMP2 and
1930	MMP9 promote metastasis whilst CCL7 and CCR3 have been linked to generation
1939	of an adjooking gradient giving directionality to tumour cell migration
1940	of an adipolatic gradient grang an eetionanty to tamour een migration.
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# Table 1: Key lipid regulating genes

Gene	Function
SREBP1	Transcription factor, binding sterol regulatory element- 1 (SRE1) sites. Activity is governed by AR. It governs lipid homeostasis and metabolism as well as sterol biosynthesis.
PPARG	Transcription factor of the nuclear hormone receptor family, binding PPAR response elements (PPRE). It governs the activity of genes involved in lipid metabolism and adipocyte differentiation.
FASN	Enzyme responsible for the generation of long chain saturated fatty acids from acetyl-CoA and malonyl-CoA.
SCD1	Enzyme downstream of FASN responsible for the rate limiting step of converting of saturated fatty acids to unsaturated fatty acids, by insertion of a double bond at the $\Delta 9$ position.
ACLY	Enzyme upstream of FASN responsible for the conversion of citrate to acetyl-CoA.
ACC	Enzyme that bridges the gap between ACLY and FASN, responsible for conversion of acetyl Co-A to malonyl- CoA.

