

Standard Operating Procedure for PulseNet PFGE of *Vibrio cholera* and *Vibrio parahaemolyticus*

Purpose

To describe the One-Day (24-26 hour) Standardized Laboratory Protocol for Molecular Subtyping of *Vibrio cholera* and *Vibrio parahaemolyticus* by Pulsed-field Gel Electrophoresis (PFGE).

Scope

To provide the PulseNet participants with the same procedure for performing PFGE of *Vibrio cholerae* and *Vibrio parahaemolyticus*, thus ensuring inter-laboratory comparability of the generated results.

Definitions and Terms

1. PFGE: Pulsed-field Gel Electrophoresis
2. DNA: Deoxyribonucleic acid
3. CDC: Centers for Disease Control and Prevention
4. CLRW: Clinical Laboratory Reagent Water

Biosafety Warning

Vibrio cholerae and *Vibrio parahaemolyticus* are human pathogens and can cause serious disease. Always use Biosafety Level 2 practices and extreme caution when transferring and handling strains of these species. Work in a biological safety cabinet when handling large amounts of cells. Disinfect or dispose of all plasticware and glassware that come in contact with the cultures in a safe manner.

Please read all instructions carefully before starting protocol. Treat all plasticware, glassware, pipets, spatulas, etc. that come in contact with the cell suspensions or plugs as contaminated materials and dispose of, or disinfect according to the guidelines of your institution. Disinfect reusable plug molds before they are washed; the disposable plug molds, including the tape and the tab that is used to push the plugs out of the wells, are also contaminated and should be disinfected with 1% Lysol/Amphyll or 90% Ethanol for at least 30 minutes if they will be washed and reused.

Day 0

Grow the culture

Streak an isolated colony from test cultures onto Trypticase Soy Agar with 5% defibrinated sheep blood (TSA-SB) plates (or comparable media) for confluent growth. It is recommended that a storage vial of each culture be created. To do this, stab small screw cap tubes of TSA, HIA, or similar medium with the same inoculating loop used to streak the plate. This will ensure that the same colony can be retested if necessary.

Incubate cultures at 37°C for 14-18 hours.

Day 1

Making plugs

1. Turn on shaker water bath or incubator (54-55°C), stationary water baths (55-60°C) and spectrophotometer (or equivalent instrument such as the Dade Microscan Turbidity meter or bioMérieux Vitek colorimeter).
2. Prepare TE Buffer (10 mM Tris:1 mM EDTA, pH 8.0) as follows:
 - 2.1. 10 ml of 1 M Tris, pH 8.0
 - 2.2. 2 ml of 0.5 M EDTA, pH 8.0
 - 2.3. Dilute to 1000 ml with sterile Ultrapure Clinical Laboratory Reagent Water (CLRW)

The TE Buffer is used to make the plug agarose and also to wash lysed PFGE plugs.

3. Prepare 1% SeaKem Gold agarose in TE Buffer (10 mM Tris:1 mM EDTA, pH 8.0) for PFGE plugs as follows:
 - 3.1. Weigh 0.50 g (or 0.25 g) SeaKem Gold (SKG) agarose into 250 ml screw-cap flask.
 - 3.2. Add 50.0 ml (or 25.0 ml) TE Buffer; swirl gently to disperse agarose.
 - 3.3. Loosen cap, and microwave for 30 seconds; mix gently and repeat for 10 seconds intervals until agarose is completely dissolved.
 - 3.4. Recap flask and return to 55-60°C water bath and equilibrate the agarose in the water bath for 15 minutes or until ready to use.

SAFETY WARNING: USE HEAT-RESISTANT GLOVES WHEN HANDLING HOT FLASKS AFTER MICROWAVING.

SeaKem Gold agarose works well for making PFGE plugs because it provides added strength to the plugs minimizing breakage of plugs during the lysis and washing steps. The time and temperature needed to completely dissolve the agarose is dependent on the specifications of the microwave used, and will have to be determined empirically in each laboratory.

4. Label small tubes (12mm x 75mm Falcon tubes or equivalent) with culture numbers.
5. Prepare Cell Suspension Buffer (100 mM Tris:100 mM EDTA, pH 8.0) as follows:
 - 5.1. 100 ml of 1 M Tris, pH 8.0
 - 5.2. 200 ml of 0.5 M EDTA, pH 8.0
 - 5.3. Dilute to 1000 ml with sterile Ultrapure water (CLRW)
6. Transfer 2 ml of Cell Suspension Buffer (CSB) to small labeled tubes. Use a sterile polyester-fiber or cotton swab that has been moistened with sterile CSB to remove some of the growth from agar plate; suspend cells in CSB by spinning swab gently so cells will be evenly dispersed and formation of aerosols is minimized.

The minimum volume of the cell suspension needed will depend on size of the cuvettes or tubes used to measure the cell concentration and are dependent on the manufacturer's specifications for the spectrophotometer, turbidity meter, or

colorimeter. Keep suspensions on ice if you have more than 6 cultures to process or refrigerate cell suspensions if you cannot adjust their concentration immediately.

7. Adjust concentration of cell suspensions to one of values given below by diluting with sterile CSB or by adding additional cells.
 - 7.1. Spectrophotometer: 610 nm wavelength, absorbance (Optical Density) of 0.9 (range of 0.8-1.0)
 - 7.2. Dade Microscan Turbidity Meter:
 - 7.2.1. 0.35-0.45 (measured in Falcon 2054 tubes)
 - 7.2.2. 0.52-0.64 (measured in Falcon 2057 tubes), *V. cholera*
 - 7.2.3. 0.55-0.65 (measured in Falcon 2057 tubes), *V. parahaemolyticus*
 - 7.3. bioMérieux Vitek colorimeter: 20% transmittance (measured in Falcon 2054 tubes)

The values in step 7 give satisfactory results at CDC; each laboratory may need to establish the optimal concentration needed for satisfactory results.

Casting Plugs

1. Label wells of PFGE plug molds with culture number. When reusable plug molds are used, put strip of tape on lower part of reusable plug mold before labeling wells.

Unused plug agarose can be kept at room temperature and reused 1-2 times. Microwave on low-medium power for 10-15 seconds and mix; repeat for 5-10 seconds intervals until agarose is completely melted. This agarose melts rapidly!

Proteinase K solutions (20 mg/ml) are available commercially. Alternatively, a stock solution of Proteinase K can be prepared from the powder in sterile Ultrapure water (CLRW). For best results, aliquot 300-500 µl into small tubes and store at -20°C until ready to use. Just before use, thaw appropriate number of vials needed for the samples; keep Proteinase K solutions on ice. If the Proteinase K stock solution was prepared from powder, discard any thawed solution at the end of work day. Store commercially prepared Proteinase K solutions according to directions provided by the supplier.

2. Transfer 400 µl adjusted cell suspensions to labeled 1.5-ml microcentrifuge tubes.
3. Add 20 µl of Proteinase K (20 mg/ml stock) to each tube and mix gently with pipet tip. (200 µl are needed for 10 cell suspensions.)
4. Add 400 µl melted 1% SeaKem Gold agarose to 400 µl cell suspension; mix by gently pipetting mixture up and down a few times. Over-pipetting can cause DNA shearing. Maintain temperature of melted agarose by keeping flask in beaker of warm water (55-60°C).
5. Immediately, dispense part of mixture into appropriate well(s) of reusable plug mold. Do not allow bubbles to form. Two plugs of each sample can be made from these amounts of cell suspension and agarose and are useful if repeat testing is required. Allow plugs to solidify at room temperature for 10-15 minutes. They can also be placed in the refrigerator (4°C) for 5 minutes

If disposable plug molds are used for making plugs with 1% SeaKem Gold agarose, use 200 µl cell suspension, 10 µl of Proteinase K (20 mg/ml stock) and 200 µl of agarose; up to 4 plugs can be made from these amounts of cell suspension and agarose.

The generation of cell suspension and the subsequent casting of the plugs should be performed as rapidly as possible in order to minimize premature cell lysis. If large numbers of samples are being prepared, it is recommended that they be processed in batches of around 10 samples at a time. Once the first batch of isolates are in the cell lysis incubation, then start preparing the cells suspensions the next group samples, and so on. All batches can be lysed and washed together, since additional lysis time will not affect the initial batches.

Lysis of Cells in Agarose Plugs

Two plugs (reusable molds) or 3 – 4 plugs (disposable molds) of the same strain can be lysed in the same 50ml tube.

1. Label 50ml polypropylene screw-cap or 50ml Oak Ridge tubes with culture numbers.
2. Prepare Cell Lysis Buffer (50 mM Tris:50 mM EDTA, pH 8.0 + 1% Sarcosyl) as follows:
 - 2.1. 50 ml of 1 M Tris, pH 8.0
 - 2.2. 100 ml of 0.5 M EDTA, pH 8.0
 - 2.3. 100 ml of 10 % Sarcosyl (N-Lauroylsarcosine, Sodium salt)
 - 2.4. Dilute to 1000 ml with sterile Ultrapure water (CLRW)
3. Calculate the total volume of Cell Lysis/Proteinase K Buffer needed as follows:
 - 3.1. 5 ml Cell Lysis Buffer is needed per tube
e. g., 5 ml x 10 tubes = 50 ml
 - 3.2. 25 μ l Proteinase K stock solution (20 mg/ml) is needed per tube of the cell lysis buffer
e. g., 25 μ l x 10 tubes = 250 μ l
4. Prepare the master mix by measuring the correct volume of Cell Lysis Buffer and Proteinase K into appropriate size test tube or flask and mix well.

The final concentration of Proteinase K in the lysis buffer is 0.1 mg/ml, and is different from the concentration that was added to the cell suspension (0.5 mg/ml).

5. Add 5 ml of Proteinase K/Cell Lysis Buffer master mix to each labeled 50 ml tube.
6. Trim excess agarose from top of plugs with scalpel, razor blade or similar instrument. Open reusable plug mold and transfer plugs from mold with a 6-mm wide spatula to appropriately labeled tube. If disposable plug molds are used, remove white tape from bottom of mold and push out plug(s) into appropriately labeled tube. Be sure plugs are under buffer and not on side of tube.

The excess agarose, plug mold, spatula, etc. are contaminated. Discard or disinfect appropriately.

7. Remove tape from reusable mold. Place both sections of the plug mold, spatulas, and scalpel in 90% ethanol, 1% Lysol/Amphyll or other suitable disinfectant. Soak them for 15 minutes before washing them. Discard disposable plug molds or disinfect them in 90% ethanol for 30-60 minutes if they will be washed and reused.
8. Place tubes in rack and incubate in a 54-55°C shaker water bath or incubator for 1.5-2 hours with constant and vigorous agitation (150-175 rpm). If lysing in water bath, be sure water level is above level of lysis buffer in tubes.
9. Pre-heat enough sterile Ultrapure water (CLRW) to 54-55°C so that plugs can be washed two times with 10-15 ml water (200-300 ml for 10 tubes)

Washing of Agarose Plugs After Cell Lysis

Most laboratories will find that their plugs are sufficiently stable to perform the following washing steps at 54-55°C. However, if you notice that your plugs are nicked along the edges or breaking it will be necessary for your laboratory to lower the water bath or incubator to 50°C for the following washing steps.

1. Remove tubes from water bath or incubator, and carefully pour off lysis buffer into an appropriate discard container; plugs can be held in tubes with a screened cap or spatula.

It is important to remove all of the liquid during this and subsequent wash steps by touching edge of tube or screened cap on an absorbent paper towel.

- Add at 10-15 ml sterile Ultrapure water (CLRW) that has been pre-heated to 54-55°C to each tube and shake the tubes in a 54-55°C water bath or incubator for 10-15 minutes.
- Pour off water from the plugs and repeat wash step with pre-heated water (Step 2) one more time.
- Pre-heat enough sterile TE Buffer (10 mM Tris:1 mM EDTA, pH 8.0) in a 54-55°C water bath so that plugs can be washed four times with 10-15 ml TE (400-600 ml for 10 tubes) after beginning last water wash.
- Pour off water, add 10-15 ml pre-heated (54-55°C) sterile TE Buffer, and shake the tubes in 54-55°C water bath or incubator for 10-15 minutes.
- Pour off TE and repeat wash step with pre-heated TE three more times.
- Decant last wash and add 5-10 ml sterile TE. Continue with step 1 in "Restriction Digestion" section or store plugs in TE Buffer at 4°C until needed. Plugs can be transferred to smaller tubes for long term storage.

If restriction digestion is to be done the same day, complete Steps 1-3 of next section (Restriction Digestion) during last TE wash step for optimal use of time.

Restriction Digestion of DNA in Agarose Plugs

A small slice of the plug should be digested with the primary restriction enzyme because less enzyme is required and other slices of the plug can be subjected to restriction analysis with other enzymes, according to the table below. SfiI is recommended as the secondary enzyme for analysis of Vibrio cholerae and Vibrio parahaemolyticus. The use of a secondary enzyme is useful in situations where the PFGE patterns obtained with the primary enzyme from two or more isolates are indistinguishable.

- Label 1.5-ml microcentrifuge tubes with culture numbers; label 3 (10-well gel) or 4 (15-well gel) tubes for Salmonella ser. Braenderup H9812¹ standards.
- Pre-Restriction Incubation Step: Prepare a master mix by diluting the appropriate 10X restriction buffer (Roche Applied Science or equivalent) 1:10 with sterile Ultrapure water (CLRW) according to the following table:

Reagent	µl/Plug Slice	µl/10Plug Slices	µl/15 Plug Slices
CLRW	180 µl	1800 µl	2700 µl
10X Restriction Buffer	20 µl	200 µl	300 µl
Total Volume	200 µl	2000 µl	3000 µl

- Add 200µl diluted restriction buffer (1X) to labeled 1.5-ml microcentrifuge tubes.
- Carefully remove plug from TE with spatula and place in a sterile disposable Petri dish or on large glass slide.
- Cut a 2.0 to 2.5mm wide slice from each test samples and the appropriate number of S. ser. Braenderup H9812 standards with a single edge razor blade (or scalpel, cover slip, etc.) and transfer to tube containing diluted restriction buffer. Be sure plug slice is under buffer. Replace rest of plug into the original tube that contains 5 ml TE buffer and store at 4°C.

PulseNet recommends that the combs with larger teeth (10 mm wide teeth) be used to cast the gels because computer analysis of the gel lanes is more accurate and less tedious than analysis of gel lanes cast with combs with the smaller teeth (5.5 mm). Using combs with smaller teeth is not advised. The number of slices that can be cut from the plugs will depend on the skill and experience of the operator, integrity of the plug, and whether the slices are cut vertically or horizontally (plugs made in disposable molds).

¹ Directions for making and testing PFGE plugs of Salmonella ser. Braenderup H9812 are in PNL05

- 5.1. Incubate sample and control plug slices in a water bath for 5-10 minutes or at room temp for 10-15 minutes.
 - 5.1.1. Incubate samples to be digested with *Sfi*I at 50°C.
 - 5.1.2. Incubate samples to be digested with *Not*I and *Xba*I at 37°C.
- 5.2. After incubation, remove buffer from plug slice using a pipet fitted with 200-250µl tip all the way to bottom of tube and aspirate buffer. Be careful not to damage the plug slice with pipet tip and that plug slice is not discarded with pipet tip.
6. Prepare the restriction enzyme master mix according to the following table. May mix in the same tube that was used for the diluted restriction buffer.

Reagent	µl/Plug Slice	µl/10 Plug Slices	µl/15 Plug Slices
CLRW	177 µl	1777 µl	2655 µl
10X Restriction Buffer	20 µl	200 µl	300 µl
BSA (10mg/ml)	2 µl	20 µl	30 µl
<i>Sfi</i>I or <i>Not</i>I (40U/ µl)	1 µl	10 µl	15 µl
Total Volume	200 µl	2000 µl	3000 µl

Keep vial of restriction enzyme on ice or in insulated storage box (-20°C) at all times.

Enzymes may be purchased in several different stock concentrations. The *Sfi*I stock enzyme should be ordered in concentrated form (40 U/µl). Either form is acceptable for *Not*I restriction. The calculation above is based on a 40U/µl stock concentration and will need to be adjusted for a different stock concentration.

Addition of Bovine Serum Albumin (BSA; highly recommended): Several restriction enzyme vendors specifically recommend the addition of 1X BSA to enzyme restriction mixtures while others do not. PulseNet Central recommends adding BSA to all enzyme restriction mixtures to minimize the incidence of incomplete restriction.

7. Add 200 µl restriction enzyme master mix to each tube. Close tube and mix by tapping gently; be sure plug slices are under enzyme mixture.
8. Incubate sample and control plug slices for 4 hours in the appropriate waterbath.
 - 8.1. Incubate samples digested with *Sfi*I at 50°C.
 - 8.2. Incubate samples digested with *Not*I and *Xba*I at 37°C.
9. If plug slices will be loaded into the wells (Option B, page 9), continue with Steps 1-4 of the next section (Casting an Agarose Gel) approximately 1 hour before restriction digest reaction is finished so the gel can solidify for at least 30 minutes before loading the restricted PFGE plugs.

Casting an Agarose Gel

1. Confirm that water bath is equilibrated to 55-60°C.
2. Make volume of 0.5X Tris-Borate EDTA Buffer (TBE) that is needed for both the gel and electrophoresis running buffer according to one of the following tables.

Reagent	Volume (ml)	Volume(ml)
5X TBE Stock	200	220
CLRW	1800	1980
Total Volume	2000	2200

Reagent	Volume (ml)	Volume (ml)
10X TBE Stock	100	110
CLRW	1900	2090
Total Volume	2000	2200

3. Make 1% SeaKem Gold (SKG) Agarose in 0.5X TBE as follows:
 - 3.1. Weigh appropriate amount of SKG into 500 ml screw-cap flask.
 - 3.1.1. Mix 1.0 g agarose with 100 ml 0.5X TBE for 14cm-wide gel form (10 wells)
 - 3.1.2. Mix 1.5 g agarose with 150 ml 0.5X TBE for 21cm-wide gel form (15 wells)
 - 3.2. Add appropriate amount of 0.5X TBE; swirl gently to disperse agarose.
4. Loosen cap and microwave for 60 seconds; mix gently and repeat for 15 second intervals until agarose is completely dissolved.
5. Return flask to the 55-60°C water bath and equilibrate the agarose in the water bath for 15 minutes or until ready to use.

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Agarose LFTM (Amresco, X174) and Certified Megabase Agarose (Bio-Rad, 161-3108) are the only acceptable alternatives to SeaKem Gold, at this time. The time and temperature needed to completely dissolve the agarose is dependent on the specifications of the microwave used and will have to be determined empirically in each laboratory. Similarly, the optimal running time for each agarose will have to be determined empirically in each laboratory.

6. A small volume (2-5 ml) of melted and cooled (55-60°C) 1% SKG agarose may be wanted to seal wells after plugs are loaded. Prepare as described above. Unused SKG agarose can be kept at room temperature, melted, and reused several times.

Place the gel form on a leveling table and adjust until perfectly leveled. Place the comb holder so the front part (side with small metal screws) and teeth face the bottom of gel frame and the comb teeth touch the gel platform.

7. Remove restricted plug slices from 37°C water bath. Remove enzyme/buffer mixture and add 200 µl 0.5X TBE. Incubate at room temperature for 5 minutes.
8. Remove plug slices from tubes; put comb on bench top and load plug slices on the bottom of the comb teeth as follows:
 - 8.1. Load S. ser. Braenderup H9812 standards on teeth (lanes) 1, 5, 10 (10 well gel) or on teeth 1, 5, 10, 15 (15 well gel).
 - 8.2. Load samples on remaining teeth and note locations.
9. Remove excess buffer with a kimwipe. Allow plug slices to air dry on the comb for 3-5 minutes or seal them to the comb with 1% SKG agarose (55-60°C).
10. Position comb in gel form and confirm that the plug slices are correctly aligned on the bottom of the comb teeth, that the lower edge of the plug slice is flush against the black platform.
11. Carefully pour the agarose (cooled to 55-60°C) into the gel form and remove any bubbles or debris.
12. Put black gel frame in electrophoresis chamber. Add 2 -2.2 L freshly prepared 0.5X TBE. Close cover of unit. (The amount of buffer needed depends on whether residual buffer was left in tubing or if unit was flushed with water after the last gel was run).
13. Turn on the power supply, pump calibrated to a flow rate of 1 liter/minute (setting of about 70) and cooling module (14°C) approximately 30 minutes before gel is to be run.
14. Remove comb after gel solidifies for 30-45 minutes.
15. Fill in wells of gel with melted and cooled (55-60°C) 1% SKG Agarose (optional). Unscrew and remove end gates from gel form; remove excess agarose from sides and bottom of casting platform with a kimwipe. Keep gel on casting platform and carefully place gel inside black gel frame in electrophoresis chamber. Close cover of chamber.

Loading Restricted Plug Slices into the Wells (optional alternate method)

1. Follow steps 1-4 in the section Loading Restricted Plug Slices on the Comb.

Place the gel form on a leveling table and adjust until perfectly leveled before pouring gel. Position the comb holder so that front part (side with small metal screws) and teeth face the bottom of gel and the bottom edge of the comb is 2 mm above the surface of the gel platform.

2. Cool melted SKG agarose in 55-60°C water bath for 15-20 min; carefully pour agarose into gel form (casting stand) fitted with comb. Be sure there are no bubbles.
3. Put black gel frame in electrophoresis chamber. Add 2-2.2 L freshly prepared 0.5X TBE. Close cover of unit. (The amount of buffer depends on whether residual buffer was left in tubing, or if unit was flushed with water after the last gel was run.)
4. Turn on power supply, pump calibrated to a flow rate of 1 liter/minute (setting at about 70), and cooling module (14°C) approximately 30 minutes before gel is to be run.
5. Remove restricted plug slices from 37°C water bath. Remove enzyme/buffer mixture and add 200 µl 0.5X TBE. Incubate at room temperature for 5 minutes.
6. Remove comb after gel solidifies for at least 30 minutes.
7. Remove restricted plug slices from tubes with tapered end of spatula and load into appropriate wells. Gently push plugs to bottom and front of wells with wide end of spatula. Manipulate position with spatula and be sure that are no bubbles.
 - 7.1. Load 5 ser. Braenderup H9812 standards in wells (lanes) 1, 5, 10 (10-well gel) or in wells 1, 5, 10, 15 (15 well gel).
 - 7.2. Load samples in remaining wells.

Loading the plug slices can be tedious; each person has to develop his/her own technique for consistently placing the plug slices in the wells so the lanes will be straight and the bands sharp.

8. Fill in wells of gel with melted 1% SKG Agarose (equilibrated to 55-60°C). Allow to harden for 3-5 minutes. Unscrew and remove end gates from gel form; remove excess agarose from sides and bottom of casting platform with a tissue. Keep gel on casting platform and carefully place gel inside black gel frame in electrophoresis chamber. Close cover of chamber.

Electrophoresis Conditions

Vibrio cholera strains restricted with *SfiI* or *NotI*:

- Select following conditions on CHEF Mapper with a two block program
 - Press the Multi-State button
 - Program with Interrupts?
 - Press 0 for “no” then Enter
 - Block 1 Runtime?
 - Enter 13 hours, then Enter
 - Block 1, State 1:
 - 6.0 Volts, Enter
 - **60° angle**, Enter
 - 2 seconds Initial Switch Time, Enter
 - 10 seconds Final Switch Time, Enter
 - Press 0 for a linear Ramping Factor, Enter
 - Continue with another state (vector)?
 - Press 1 for yes, Enter
 - Block 1, State 2
 - 6.0 Volts, Enter
 - **-60° angle**, Enter
 - 2 seconds Initial Switch Time, Enter
 - 10 seconds Final Switch Time, Enter
 - Press 0 for a linear Ramping Factor, Enter
 - Continue with another state (vector)?
 - Press 0 for No, Enter
 - Continue with another Block?
 - Press 1 for Yes, Enter
 - Block 2 Runtime?
 - 6 hours, Enter
 - Block 2, State 1:
 - 6.0 Volts, Enter
 - **60° angle**, Enter
 - 20 seconds Initial Switch Time, Enter
 - 25 seconds Final Switch Time, Enter
 - Press 0 for a linear Ramping Factor, Enter
 - Continue with another state (vector)?
 - Press 1 for yes, Enter
 - Block 2, State 2
 - 6.0 Volts, Enter
 - **-60° angle**, Enter
 - 20 seconds Initial Switch Time, Enter
 - 25 seconds Final Switch Time, Enter
 - Press 0 for a linear Ramping Factor, Enter
 - Continue with another state (vector)?
 - Press 0 for No, Enter
 - Continue with another Block?

- Press 0 for No, Enter
 - A Program is in memory, please enter another command
 - Press the Start Run button
- Select following conditions on CHEF-DR III
 - Block 1
 - Initial switch time: 2 s
 - Final switch time: 10 s
 - Voltage: 6 V
 - Included Angle: 120°
 - Run time: 13 hours
 - Block 2
 - Initial switch time: 2 s
 - Final switch time: 10 s
 - Voltage: 6 V
 - Included Angle: 120°
 - Run time: 6 hours

Vibrio parahaemolyticus strains restricted with *SfiI* or *NotI*:

- Select following conditions on CHEF Mapper
 - Auto Algorithm
 - 78 kb: low MW
 - 396 kb: high MW
 - Select default values except where noted by pressing "Enter."
 - Change run time to 18-19 hours (See note below)
 - (Default values: Initial switch time = 10 s; Final switch time = 35.03 s)
- Select following conditions on CHEF DR-III
 - Initial switch time: 10 s
 - Final switch time: 35 s
 - Voltage: 6 V
 - Included Angle: 120°
 - Run time: 18-19 hours (See note below)
- Select following conditions on CHEF DR-II.
 - Initial A time: 10s
 - Final A time: 35 s
 - Start Ratio: 1.0 (if applicable)
 - Voltage: 200 V
 - Run time: 19-20 hours (See note below)

The electrophoresis running times recommended above are based on the equipment and reagents used at the CDC. Run times may be different in your laboratory and will have to be optimized for your gels so that the lowest band in the S. ser. Braenderup H9812 standard migrates within 1.0-1.5 cm of the bottom of the gel.

Make note of the initial milliamp (mA) reading on the instrument. The initial mA should be between 110-150 mA. A reading outside of this range may indicate that the 0.5X TBE buffer was prepared improperly and the buffer should be remade.

Staining and Documentation of an Agarose Gel

The following staining procedure describes the use of ethidium bromide to stain PFGE gels. Alternate DNA stains may be used. Please see the “Alternate DNA Stains-Results and Recommendations” posting within the Important PulseNet Documents forum on CDC Team for additional information.

1. When electrophoresis run is over, turn off equipment; remove and stain gel with ethidium bromide. Dilute 40 µl of ethidium bromide stock solution (10 mg/ml) with 400 ml of Ultrapure water (CLRW) (this volume is for a staining box that is approximately 14-cm x 24-cm; a larger container may require a larger amount of staining solution). Stain gel for 20-30 minutes in covered container.

Ethidium bromide is toxic and a mutagen. Stock solutions of 10 mg/ml Ethidium Bromide (EtBr) in water are available from several commercial companies (Amresco X328; Bio-Rad, 161-0433; Sigma, E-1510). The diluted solution can be kept in dark bottle and reused 6-8 times before discarding according to your institution's guidelines for hazardous waste. CDC does not recommend disposing of EtBr down the drain. Aqueous solutions containing EtBr can be filtered through charcoal or degraded using activated carbon destaining or “tea” bags from Amresco (E732-25 Destaining Bags) or other companies, which effectively and safely remove EtBr from solutions and gels. Once the EtBr is removed, the treated aqueous solutions can be discarded down the drain. If you have further questions about EtBr please refer to the Material Safety Data Sheets (MSDS) provided by the vendor or manufacturer.

Currently, the only acceptable alternative stain options are GelRed™ (Biotium, 31010), SYBR® Safe (Invitrogen, S-33102) and SYBR® Gold (Invitrogen, S-11494). Labs are strongly encouraged to follow manufacturer's instructions and test stains in their labs before adopting them for routine use. Importantly, if one of the alternative stains is used, the destaining steps should be omitted.

2. Destain gel in approximately 500 ml reagent grade water for 60-90 minutes; change water every 20 minutes. Capture image a Gel Doc 1000, 2000, EQ, or XR, or equivalent documentation system. If too much background is observed destain for an additional 30-60 minutes.
3. Follow directions given with the imaging equipment to save gel image as a *.1sc or *.scn file; convert this file to *.tif file for analysis with the BioNumerics software program. The gel image should fill the entire window of the imaging equipment (computer) screen (without cutting off wells or lower bands). Ensure that the image is in focus and that there is little to no saturation (over-exposure) in the bands. Additional instructions are provided in PNL07 of the PulseNet QA/QC manual.
4. Drain buffer from electrophoresis chamber and discard. Rinse chamber with 2 L Ultrapure water (CLRW) or, if unit is not going to be used for several days, flush lines with water by letting pump run for 5-10 minutes before draining water from chamber and hoses.
5. If the lowest band in the H9812 standard does not migrate within 1 -1.5 cm of the bottom of the gel, the run time will need to be determined empirically for the conditions in each laboratory.

Please note the following if PFGE results do not have to be available within 24-28 hours:

- *Plugs can be lysed for longer periods of time (3-16 hours).*
- *The washing steps with TE to remove the lysis buffer from the PFGE plugs can be done for longer periods of time (30-45 min) and at lower temperatures (37°C or room temperature). They can be started on Day 1 and finished on Day 2 after overnight refrigeration of the plugs in TE.*

USE OF TRADE NAMES AND COMMERCIAL SOURCES IS FOR IDENTIFICATION PURPOSES ONLY AND DOES NOT IMPLY ENDORSEMENT BY CDC OR THE U.S. DEPARTMENT OF HEALTH AND HUMAN SERVICES.

CLIA Laboratory Procedure Manual Requirements

Efforts have been made to assure that the procedures described in this protocol have been written in accordance with the 1988 Clinical Laboratory Improvement Amendments (CLIA) requirements for a procedure manual (42 CFR 493.1211). However, due to the format required for training, the procedures will require some modifications and additions to customize them for your particular laboratory operation.

Any questions regarding the CLIA requirements for a procedure manual, quality control, quality assurance, etc., should be directed to the agency or accreditation organization responsible for performing your laboratory's CLIA inspection. In addition, some states and accreditation organizations may have more stringent requirements that will need to be addressed.

Formulas of Selected Reagents used in PulseNet Standardized Laboratory Protocol for PFGE

Tris: EDTA Buffer, pH 8.0: (TE, 10 mM Tris: 1 mM EDTA, pH 8.0)

10 ml of 1 M Tris, pH 8.0

2 ml of 0.5 M EDTA, pH 8.0

Dilute to 1000 ml with sterile Ultrapure water (CLRW)

Cell Lysis Buffer: (50 mM Tris: 50 mM EDTA, pH 8.0 + 1% Sarcosine + 0.1 mg/ml Proteinase K)

50 ml of 1 M Tris, pH 8.0

100 ml of 0.5 M EDTA, pH 8.0

100 ml 10% N-Lauroylsarcosine, Sodium salt (Sarcosyl) OR 10 g of N-Lauroylsarcosine, Sodium salt (Sarcosyl)²

Dilute to 1000 ml with Sterile Ultrapure water (CLRW)

Add 25 µl Proteinase K stock solution (20 mg/ml) per 5 ml of cell lysis buffer just before use for a final concentration in the lysis buffer of 0.1 mg/ml Proteinase K.

Restriction Enzyme Master Mix: *Xba*I (50U/plug slice) low concentration stock

Reagent	µl/Plug Slice	µl/Plug Slices	µl/15Plug Slices
CLRW	173 µl	1730 µl	2595 µl
10X H Buffer	20 µl	200 µl	300 µl
BSA (10mg/ml)	2 µl	20 µl	30 µl
<i>Xba</i> I (10U/µl)	5 µl	50 µl	75 µl
Total Volume	200 µl	2000 µl	3000 µl

Keep vials of restriction enzyme on ice or in an insulated storage box (-20°C) at all times.

² *If Sarcosyl powder is added directly to the other components of this reagent, warm the solution to 50- 60°C for 30-60 minutes, or leave at room temperature for about 2 hours to completely dissolve the Sarcosyl*