PROTEIN-CODING VARIANTS IMPLICATE NOVEL GENES RELATED TO LIPID HOMEOSTASIS CONTRIBUTING TO BODY FAT DISTRIBUTION

Anne E Justice[¥,1,2], Tugce Karaderi[¥,3,4], Heather M Highland[¥,1,5], Kristin L Young[¥,1], Mariaelisa Graff[¥,1], Yingchang Lu[¥,6,7,8], Valérie Turcot[9], Paul L Auer[10], Rebecca S Fine[11,12,13], Xiuqing Guo[14], Claudia Schurmann[7,8], Adelheid Lempradl[15], Eirini Marouli[16], Anubha Mahajan[3], Thomas W Winkler[17], Adam E Locke[18,19], Carolina Medina-Gomez[20,21], Tõnu Esko[11,13,22], Sailaja Vedantam[11,12,13], Ayush Giri[23], Ken Sin Lo[9], Tamuno Alfred[7], Poorva Mudgal[24], Maggie CY Ng[24,25], Nancy L Heard-Costa[26,27], Mary F Feitosa[28], Alisa K Manning[11,29,30] , Sara M Willems[31], Suthesh Sivapalaratnam[30,32,33], Goncalo Abecasis[18], Dewan S Alam[34], Matthew Allison[35], Philippe Amouyel[36,37,38], Zorayr Arzumanyan[14], Beverley Balkau[39], Lisa Bastarache[40], Sven Bergmann[41,42], Lawrence F Bielak[43], Matthias Blüher[44,45], Michael Boehnke[18], Heiner Boeing[46], Eric Boerwinkle[47,48], Carsten A Böger[49], Jette Bork-Jensen[50], Erwin P Bottinger[7], Donald W Bowden[24,25,51], Ivan Brandslund[52,53], Linda Broer[21], Amber A Burt[54], Adam S Butterworth[55,56], Mark J Caulfield[16,57], Giancarlo Cesana[58], John C Chambers[59,60,61,62,63], Daniel I Chasman[11,64,65,66], Yii-Der Ida Chen[14], Rajiv Chowdhury[55], Cramer Christensen[67], Audrey Y Chu[27,65], Francis S Collins[68], James P Cook[69], Amanda J Cox[24,25,70], David S Crosslin[71], John Danesh[55,56,72,73], Paul IW de Bakker[74,75], Simon de Denus[9,76], Renée de Mutsert[77], George Dedoussis[78], Ellen W Demerath[79], Joe G Dennis[80], Josh C Denny[40], Emanuele Di Angelantonio[55,56], Marcus Dörr[81,82], Fotios Drenos[83,84], Marie-Pierre Dubé[9,85], Alison M Dunning[86], Douglas F Easton[80,86], Paul Elliott[87], Evangelos Evangelou[61,88], Aliki-Eleni Farmaki[78], Shuang Feng[18], Ele Ferrannini[89,90], Jean Ferrieres[91], Jose C Florez[11,29,30], Myriam Fornage[92], Caroline S Fox[27], Paul W Franks[93,94,95], Nele Friedrich[96], Wei Gan[3], Ilaria Gandin[97], Paolo Gasparini[98,99], Vilmantas Giedraitis[100], Giorgia Girotto[98,99], Mathias Gorski[17,49], Harald Grallert[101,102,103], Niels Grarup[50], Megan L Grove[47], Stefan Gustafsson[104], Jeff Haessler[105], Torben Hansen[50], Andrew T Hattersley[106], Caroline Hayward[107], Iris M Heid[17,108], Oddgeir L Holmen[109], G Kees Hovingh[110], Joanna MM Howson[55], Yao Hu[111], Yi-Jen Hung[112,113],

Kristian Hveem[109,114], M Arfan Ikram[20,115,116], Erik Ingelsson[104,117], Anne U Jackson[18], Gail P Jarvik[54,118], Yucheng Jia [14], Torben Jørgensen[119,120,121], Pekka Jousilahti[122], Johanne M Justesen[50], Bratati Kahali[123,124,125,126], Maria Karaleftheri[127], Sharon LR Kardia[43], Fredrik Karpe[128,129], Frank Kee[130], Hidetoshi Kitajima[3], Pirjo Komulainen[131,132,133], Jaspal S Kooner[60,62,63,134], Peter Kovacs[44], Bernhard K Krämer[135], Kari Kuulasmaa[122], Johanna Kuusisto[136], Markku Laakso[136], Timo A Lakka[131,132,133], David Lamparter[41,42], Leslie A Lange[137], Claudia Langenberg[31], Eric B Larson[54,138,139], Nanette R Lee[140,141], Wen-Jane Lee[142,143], Terho Lehtimäki[144,145], Cora E Lewis[146], Huaixing Li[111], Jin Li[147], Ruifang Li-Gao[77], Li-An Lin[92], Xu Lin[111], Lars Lind[148], Jaana Lindström[122], Allan Linneberg[121,149,150], Ching-Ti Liu[151], Dajiang J Liu[152], Jian'an Luan[31], Leo-Pekka Lyytikäinen[144,145], Stuart MacGregor[153], Reedik Mägi[22], Satu Männistö[122], Gaëlle Marenne[72], Jonathan Marten[107], Nicholas GD Masca[154,155], Mark I McCarthy[3,128,129], Karina Meidtner[101,156], Evelin Mihailov[22], Leena Moilanen[157], Marie Moitry[158,159], Dennis O Mook-Kanamori[77,160], Anna Morgan[98], Andrew P Morris[3,69], Martina Müller-Nurasyid[108,161,162], Patricia B Munroe[16,57], Narisu Narisu[68], Christopher P Nelson[154,155], Matt Neville[128,129], Ioanna Ntalla[16], Jeffrey R O'Connel[163], Katharine R Owen[128,129], Oluf Pedersen[50], Gina M Peloso[151], Craig E Pennell[164], Markus Perola[122,165,166], James A Perry[163], John RB Perry[31], Tune H Pers[50,167], Ailith Pirie[86], Ozren Polasek[168,169], Olli T Raitakari[170,171], Asif Rasheed[172], Chelsea K Raulerson[137], Rainer Rauramaa[131,132,133], Dermot F Reilly[173], Alex P Reiner[105,174], Paul M Ridker[65,66,175], Manuel A Rivas[11,176], Neil R Robertson[3,128], Antonietta Robino[177], Igor Rudan[169], Katherine S Ruth[178], Danish Saleheen[172,179], Veikko Salomaa[122], Nilesh J Samani[154,155], Pamela J Schreiner[180], Matthias B Schulze[101,156], Robert A Scott[31], Marcelo P Segura-Lepe[61], Xueling Sim[18,181], Andrew J Slater[182,183], Kerrin S Small[184], Blair H Smith[185,186], Jennifer A Smith[43], Lorraine Southam[3,72], Timothy D Spector[184], Elizabeth K Speliotes[123,124,125], Kari Stefansson[187,188], Valgerdur Steinthorsdottir[187], Kathleen E Stirrups[16,33], Konstantin Strauch[108,189], Heather M Stringham[18], Michael Stumvoll[44,45], Liang Sun[190,191], Praveen Surendran[55], Karin MA Swart[192], Jean-Claude Tardif[9,85], Kent D Taylor[14], Alexander Teumer[193], Deborah J Thompson[80], Gudmar Thorleifsson[187], Unnur Thorsteinsdottir[187,188], Betina H

Thuesen[121], Anke Tönjes[194], Mina Torres[195], Emmanouil Tsafantakis[196], Jaakko Tuomilehto[122,197,198,199], André G Uitterlinden[20,21], Matti Uusitupa[200], Cornelia M van Duijn[20], Mauno Vanhala[201,202], Rohit Varma[195], Sita H Vermeulen[203], Henrik Vestergaard[50,204], Veronique Vitart[107], Thomas F Vogt[205], Dragana Vuckovic[99], Lynne E Wagenknecht[206], Mark Walker[207], Lars Wallentin[208], Feijie Wang[111], Carol A Wang[164], Shuai Wang[151], Nicholas J Wareham[31], Helen R Warren[16,57], Dawn M Waterworth[209], Jennifer Wessel[210], Harvey D White[211], Cristen J Willer[123,124,212], James G Wilson[213], Andrew R Wood[178], Ying Wu[137], Hanieh Yaghootkar[178], Jie Yao[14], Laura M Yerges-Armstrong[163,214], Robin Young[55,215], Eleftheria Zeggini[72], Xiaowei Zhan[216], Weihua Zhang[60,61], Jing Hua Zhao[31], Wei Zhao[179], He Zheng[111], Wei Zhou[123,124], M Carola Zillikens[20,21], CHD Exome+ Consortium, EPIC-CVD Consortium, ExomeBP Consortium, Global Lipids Genetic Consortium, GoT2D Genes Consortium, InterAct, ReproGen Consortium, T2D-Genes Consortium, The MAGIC Investigators, Fernando Rivadeneira[20,21], Ingrid B Borecki[28], John A Pospisilik[15], Panos Deloukas[16,217], Timothy M Frayling[178], Guillaume Lettre[9,85], Karen L Mohlke[137], Jerome I Rotter[14], Zoltán Kutalik[42,218], Joel N Hirschhorn[11,13,219], L Adrienne Cupples[Ł,27,151], Ruth JF Loos[Ł,7,8,220], *Kari E North[*,Ł,221], *Cecilia M Lindgren [*,Ł,3,222]

¥ These authors contributed equally to this work.

Ł These authors jointly supervised this work.

***CORRESPONDING AUTHORS**

Prof. Kari North

Department of Epidemiology

University of North Carolina at Chapel Hill

137 East Franklin Street

Suite 306

Chapel Hill, NC 27514

Prof. Cecilia M Lindgren

The Big Data Institute, Li Ka Shing Centre for Health Information and Discovery

University of Oxford
Roosevelt Drive
Oxford
OX3 7BN
United Kingdom
celi@well.ox.ac.uk

AFFILIATIONS

1. Department of Epidemiology, University of North Carolina, Chapel Hill, NC, 27514, USA
2. Biomedical and Translational Informatics, Geisinger Health, Danville, PA 17822
3. Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford, OX3 7BN, UK
4. Department of Biological Sciences, Faculty of Arts and Sciences, Eastern Mediterranean University, Famagusta, Cyprus
5. Human Genetics Center, The University of Texas School of Public Health, The University of Texas MD Anderson Cancer Center UTHealth Graduate School of Biomedical Sciences, The University of Texas Health Science Center at Houston, Houston, TX, 77030, USA
6. Division of Epidemiology, Department of Medicine, Vanderbilt-Ingram Cancer Center, Vanderbilt Epidemiology Center, Vanderbilt University School of Medicine, Nashville, TN, 37203, USA
7. The Charles Bronfman Institute for Personalized Medicine, Icahn School of Medicine at Mount Sinai, New York, NY, 10029, USA
8. The Genetics of Obesity and Related Metabolic Traits Program, Icahn School of Medicine at Mount Sinai, New York, NY, 10069, USA
9. Montreal Heart Institute, Universite de Montreal, Montreal, Quebec, H1T 1C8, Canada
10. Zilber School of Public Health, University of Wisconsin-Milwaukee, Milwaukee, WI, 53201, USA
11. Broad Institute of MIT and Harvard, Cambridge, MA, 02142, USA
12. Department of Genetics, Harvard Medical School, Boston, MA, 02115, USA

13. Division of Endocrinology and Center for Basic and Translational Obesity Research, Boston Children's Hospital, Boston, MA, 02115, USA
14. Institute for Translational Genomics and Population Sciences, LABioMed at Harbor-UCLA Medical Center, Torrance, CA, 90502, USA
15. Max Planck Institute of Immunobiology and Epigenetics, Freiburg, 79108, Germany
16. William Harvey Research Institute, Barts and The London School of Medicine and Dentistry, Queen Mary University of London, London, EC1M 6BQ, UK
17. Department of Genetic Epidemiology, University of Regensburg, Regensburg, D-93051, Germany
18. Department of Biostatistics and Center for Statistical Genetics, University of Michigan, Ann Arbor, MI, 48109, USA
19. McDonnell Genome Institute, Washington University School of Medicine, Saint Louis, MO, 63108, USA
20. Department of Epidemiology, Erasmus Medical Center, Rotterdam, 3015 GE, The Netherlands
21. Department of Internal Medicine, Erasmus Medical Center, Rotterdam, 3015 GE, The Netherlands
22. Estonian Genome Center, University of Tartu, Tartu, 51010, Estonia
23. Division of Epidemiology, Department of Medicine, Institute for Medicine and Public Health, Vanderbilt Genetics Institute, Vanderbilt University, Nashville, TN, 37203, USA
24. Center for Diabetes Research, Wake Forest School of Medicine, Winston-Salem, NC, 27157, USA
25. Center for Genomics and Personalized Medicine Research, Wake Forest School of Medicine, Winston-Salem, NC, 27157, USA
26. Department of Neurology, Boston University School of Medicine, Boston, MA, 02118, USA
27. NHLBI Framingham Heart Study, Framingham, MA, 01702, USA

28. Division of Statistical Genomics, Department of Genetics, Washington University School of Medicine, St. Louis, MO, 63108, USA
29. Department of Medicine, Harvard University Medical School, Boston, MA, 02115, USA
30. Massachusetts General Hospital, Boston, MA, 02114, USA
31. MRC Epidemiology Unit, University of Cambridge School of Clinical Medicine, Institute of Metabolic Science, Cambridge, CB2 0QQ, UK
32. Department of Vascular Medicine, AMC, Amsterdam, 1105 AZ, The Netherlands
33. Department of Haematology, University of Cambridge, Cambridge, CB2 0PT, UK
34. School of Kinesiology and Health Science, Faculty of Health, York University, Toronto
35. Department of Family Medicine & Public Health, University of California, San Diego, La Jolla, CA, 92093, USA
36. INSERM U1167, Lille, F-59019, France
37. Institut Pasteur de Lille, U1167, Lille, F-59019, France
38. Universite de Lille, U1167 - RID-AGE - Risk factors and molecular determinants of aging-related diseases, Lille, F-59019, France
39. INSERM U1018, Centre de recherche en Épidemiologie et Sante des Populations (CESP), Villejuif, France
40. Department of Biomedical Informatics, Vanderbilt University, Nashville, TN, 37203, USA
41. Department of Computational Biology, University of Lausanne, Lausanne, 1011, Switzerland
42. Swiss Institute of Bioinformatics, Lausanne, 1015, Switzerland
43. Department of Epidemiology, School of Public Health, University of Michigan, Ann Arbor, MI, 48109, USA
44. IFB Adiposity Diseases, University of Leipzig, Leipzig, 04103, Germany
45. University of Leipzig, Department of Medicine, Leipzig, 04103, Germany

46. Department of Epidemiology, German Institute of Human Nutrition Potsdam-Rehbruecke (DIfE), Nuthetal, 14558, Germany
47. School of Public Health, Human Genetics Center, The University of Texas Health Science Center at Houston, Houston, TX, 77030, USA
48. Human Genome Sequencing Center, Baylor College of Medicine, Houston, TX, 77030 USA
49. Department of Nephrology, University Hospital Regensburg, Regensburg, 93042, Germany
50. The Novo Nordisk Foundation Center for Basic Metabolic Research, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, 2100, Denmark
51. Department of Biochemistry, Wake Forest School of Medicine, Winston-Salem, NC 27157, USA
52. Department of Clinical Biochemistry, Lillebaelt Hospital, Vejle, 7100, Denmark
53. Institute of Regional Health Research, University of Southern Denmark, Odense, 5000, Denmark
54. Department of Medicine, University of Washington, Seattle, WA, 98195, USA
55. MRC/BHF Cardiovascular Epidemiology Unit, Department of Public Health and Primary Care, University of Cambridge, Cambridge, CB1 8RN, UK
56. NIHR Blood and Transplant Research Unit in Donor Health and Genomics, Department of Public Health and Primary Care, University of Cambridge, Cambridge CB1 8RN, UK
57. NIHR Barts Cardiovascular Research Unit, Barts and The London School of Medicine & Dentistry, Queen Mary University of London, London, EC1M 6BQ, UK
58. Research Centre on Public Health, University of Milano-Bicocca, Monza, 20900, Italy
59. Lee Kong Chian School of Medicine, Nanyang Technological University, Singapore 308232, Singapore
60. Department of Cardiology, London North West Healthcare NHS Trust, Ealing Hospital, Middlesex, UB1 3HW, UK

61. Department of Epidemiology and Biostatistics, School of Public Health, Imperial College London, London, W2 1PG, UK
62. Imperial College Healthcare NHS Trust, London, W12 0HS, UK
63. MRC-PHE Centre for Environment and Health, Imperial College London, London, W2 1PG, UK
64. Division of Genetics, Brigham and Women's Hospital and Harvard Medical School, Boston, MA, 02115, USA
65. Division of Preventive Medicine, Brigham and Women's and Harvard Medical School, Boston, MA, 02215, USA
66. Harvard Medical School, Boston, MA, 02115, USA
67. Medical department, Lillebaelt Hospital, Vejle, 7100, Denmark
68. Medical Genomics and Metabolic Genetics Branch, National Human Genome Research Institute, National Institutes of Health, Bethesda, MD, 20892, USA
69. Department of Biostatistics, University of Liverpool, Liverpool, L69 3GL, UK
70. Menzies Health Institute Queensland, Griffith University, Southport, QLD, Australia
71. Department of Biomedical Infomatics and Medical Education, University of Washington, Seattle, WA, 98195, USA
72. Wellcome Trust Sanger Institute, Hinxton, CB10 1SA, UK
73. British Heart Foundation Cambridge Centre of Excellence, Department of Medicine, University of Cambridge, Cambridge, CB2 0QQ, UK
74. Julius Center for Health Sciences and Primary Care, University Medical Center Utrecht, Utrecht, The Netherlands
75. Department of Genetics, Center for Molecular Medicine, University Medical Center Utrecht, Utrecht, 3584 CX, The Netherlands
76. Faculty of Pharmacy, Universite de Montreal, Montreal, Quebec, H3T 1J4, Canada

77. Department of Clinical Epidemiology, Leiden University Medical Center, Leiden, 2300RC, The Netherlands
78. Department of Nutrition and Dietetics, School of Health Science and Education, Harokopio University, Athens, 17671, Greece
79. Division of Epidemiology & Community Health, School of Public Health, University of Minnesota, Minneapolis, MN, 55454, USA
80. Centre for Cancer Genetic Epidemiology, Department of Public Health and Primary Care, University of Cambridge, Cambridge, CB1 8RN, UK
81. Department of Internal Medicine B, University Medicine Greifswald, Greifswald, 17475, Germany
82. DZHK (German Centre for Cardiovascular Research), partner site Greifswald, Greifswald, 17475, Germany
83. Institute of Cardiovascular Science, University College London, London, WC1E 6JF, UK
84. MRC Integrative Epidemiology Unit, School of Social and Community Medicine, University of Bristol, Bristol, BS8 2BN, UK
85. Department of Medicine, Faculty of Medicine, Universite de Montreal, Montreal, Quebec, H3T 1J4, Canada
86. Centre for Cancer Genetic Epidemiology, Department of Oncology, University of Cambridge, Cambridge, CB1 8RN, UK
87. Department of Epidemiology and Biostatistics, MRC-PHE Centre for Environment and Health, School of Public Health, Imperial College London, London, W2 1PG, UK
88. Department of Hygiene and Epidemiology, University of Ioannina Medical School, Ioannina, 45110, Greece
89. CNR Institute of Clinical Physiology, Pisa, Italy

90. Department of Clinical & Experimental Medicine, University of Pisa, Italy
91. Toulouse University School of Medicine, Toulouse, TSA 50032 31059, France
92. Institute of Molecular Medicine, The University of Texas Health Science Center at Houston, Houston, TX, 77030, USA
93. Department of Clinical Sciences, Genetic and Molecular Epidemiology Unit, Lund University, Malmo, SE-20502, Sweden
94. Department of Nutrition, Harvard School of Public Health, Boston, MA, 02115, USA
95. Department of Public Health and Clinical Medicine, Unit of Medicine, Umeå University, Umeå, 901 87, Sweden
96. Institute of Clinical Chemistry and Laboratory Medicine, University Medicine Greifswald, Greifswald, 17475, Germany
97. Ilaria Gandin, Research Unit, AREA Science Park, Trieste, 34149, Italy
98. Department of Medical Sciences, University of Trieste, Trieste, 34137, Italy
99. Institute for Maternal and Child Health - IRCCS "Burlo Garofolo", Trieste, Italy
100. Geriatrics, Department of Public Health, Uppsala University, Uppsala, 751 85, Sweden
101. German Center for Diabetes Research, München-Neuherberg, 85764, Germany

102. Institute of Epidemiology II, Helmholtz Zentrum München - German Research Center for Environmental Health, Neuherberg, 85764, Germany
103. Research Unit of Molecular Epidemiology, Helmholtz Zentrum München - German Research Center for Environmental Health, Neuherberg, 85764, Germany
104. Department of Medical Sciences, Molecular Epidemiology and Science for Life Laboratory, Uppsala University, Uppsala, 751 41, Sweden

105. Division of Public Health Sciences, Fred Hutchinson Cancer Research Center, Seattle WA, 98109, USA
106. University of Exeter Medical School, University of Exeter, Exeter, EX2 5DW, UK
107. MRC Human Genetics Unit, Institute of Genetics and Molecular Medicine, University of Edinburgh, Edinburgh, EH4 2XU, UK
108. Institute of Genetic Epidemiology, Helmholtz Zentrum München - German Research Center for Environmental Health, Neuherberg, 85764, Germany
109. K.G. Jebsen Center for Genetic Epidemiology, Department of Public Health, NTNU, Norwegian University of Science and Technology, Trondheim, 7600, Norway
110. AMC, Department of Vascular Medicine, Amsterdam, 1105 AZ, The Netherlands
111. Key Laboratory of Nutrition and Metabolism, Institute for Nutritional Sciences, Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences, University of the Chinese Academy of Sciences, Shanghai, People's Republic of China, Shanghai, 200031, China
112. Division of Endocrinology and Metabolism, Department of Internal Medicine, Tri-Service General Hospital, Taipei, Taiwan 114, Taiwan
113. School of Medicine, National Defense Medical Center, Taipei, Taiwan 114, Taiwan
114. HUNT Research center, Department of Public Health, Norwegian University of Science and Technology, Levanger, 7600, Norway
115. Department of Neurology, Erasmus Medical Center, Rotterdam, 3015 GE, The Netherlands
116. Department of Radiology, Erasmus Medical Center, Rotterdam, 3015 GE, The Netherlands
117. Department of Medicine, Division of Cardiovascular Medicine, Stanford University School of Medicine, Stanford, CA, 943 05, USA
118. Department of Genome Sciences, University of Washington, Seattle, WA, 98195, USA
119. Faculty of medicine, Aalborg University, Aalborg, DK-9000, Denmark

120. Department of Public Health, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, 2200, Denmark
121. Research Center for Prevention and Health, Capital Region of Denmark, Glostrup, DK-2600, Denmark
122. National Institute for Health and Welfare, Helsinki, FI-00271, Finland
123. Department of Computational Medicine and Bioinformatics, University of Michigan, Ann Arbor, MI, 48109, USA
124. Department of Internal Medicine, University of Michigan, Ann Arbor, MI, 48109, USA
125. Division of Gastroenterology, University of Michigan, Ann Arbor, MI, 48109, USA
126. Centre for Brain Research, Indian Institute of Science, Bangalore 560012, India
127. Echinos Medical Centre, Echinos, Greece
128. Oxford Centre for Diabetes, Endocrinology and Metabolism, Radcliffe Department of Medicine, University of Oxford, Oxford, OX3 7LE, UK
129. Oxford NIHR Biomedical Research Centre, Oxford University Hospitals Trust, Oxford, OX3 7LE, UK
130. UKCRC Centre of Excellence for Public Health Research, Queens University Belfast, Belfast, UK, BT12 6BJ, UK
131. Foundation for Research in Health Exercise and Nutrition, Kuopio Research Institute of Exercise Medicine, Kuopio, 70100, Finland
132. Institute of Biomedicine, School of Medicine, University of Eastern Finland, Kuopio Campus, 70210, Finland
133. Department of Clinical Physiology and Nuclear Medicine, Kuopio University Hospital, Kuopio, Finland

134. National Heart and Lung Institute, Imperial College London, Hammersmith Hospital Campus, London, W12 0NN, UK
135. University Medical Centre Mannheim, 5th Medical Department, University of Heidelberg, Mannheim, 68167, Germany
136. Institute of Clinical Medicine, Internal Medicine, University of Eastern Finland and Kuopio University Hospital, Kuopio, 70210, Finland
137. Department of Genetics, University of North Carolina, Chapel Hill, NC, 27599, USA
138. Kaiser Permanente Washington Health Research Institute Seattle WA 98101
139. Department of Health Services, University of Washington, Seattle WA 98101
140. Department of Anthropology, Sociology, and History, University of San Carlos, Cebu City, 6000, Philippines
141. USC-Office of Population Studies Foundation, Inc., University of San Carlos, Cebu City, 6000, Philippines
142. Department of Medical Research, Taichung Veterans General Hospital, Taichung, Taiwan 407, Taiwan
143. Department of Social Work, Tunghai University, Taichung, Taiwan
144. Department of Clinical Chemistry, Fimlab Laboratories, Tampere, 33521, Finland
145. Department of Clinical Chemistry, Finnish Cardiovascular Research Center - Tampere, Faculty of Medicine and Life Sciences, University of Tampere, Tampere 33014, Finland
146. Division of Preventive Medicine University of Alabama at Birmingham, Birmingham, AL 35205, USA
147. Department of Medicine, Division of Cardiovascular Medicine, Stanford University School of Medicine, Palo Alto, CA, 94304, USA
148. Uppsala University, Uppsala, 75185, Sweden

149. Department of Clinical Experimental Research, Rigshospitalet, Copenhagen, DK-2200, Denmark
150. Department of Clinical Medicine, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, 2200, Denmark
151. Department of Biostatistics, Boston University School of Public Health, Boston, MA, 02118, USA
152. Department of Public Health Sciences, Institute for Personalized Medicine, the Pennsylvania State University College of Medicine, Hershey, PA, 17033, USA
153. QIMR Berghofer Medical Research Institute, Brisbane, Queensland, 4006, Australia
154. Department of Cardiovascular Sciences, Univeristy of Leicester, Glenfield Hospital, Leicester, LE3 9QP, UK
155. NIHR Leicester Cardiovascular Biomedical Research Unit, Glenfield Hospital, Leicester, LE3 9QP, UK
156. Department of Molecular Epidemiology, German Institute of Human Nutrition Potsdam-Rehbruecke (DIfE), Nuthetal, 14558, Germany
157. Department of Medicine, Kuopio University Hospital, Kuopio, 70210, Finland
158. Department of Epidemiology and Public Health, University of Strasbourg, Strasbourg, F-67085, France
159. Department of Public Health, University Hospital of Strasbourg, Strasbourg, F-67081, France
160. Department of Public Health and Primary Care, Leiden University Medical Center, Leiden, 2300RC, The Netherlands
161. Department of Medicine I, University Hospital Grosshadern, Ludwig-Maximilians-Universitat, Munich, 81377, Germany
162. DZHK (German Centre for Cardiovascular Research), partner site Munich Heart Alliance, Munich, 80802, Germany

163. Program for Personalized and Genomic Medicine, Department of Medicine, University of Maryland School of Medicine, Baltimore, MD, 21201, US
164. School of Women's and Infants' Health, The University of Western Australia, Perth, Western Australia, 6009, Australia
165. University of Helsinki, Institute for Molecular Medicine (FIMM) and Diabetes and Obesity Research Program, Helsinki, FI00014, Finland
166. University of Tartu, Estonian Genome Center, Tartu, Estonia, Tartu, 51010, Estonia
167. Department of Epidemiology Research, Statens Serum Institut, Copenhagen, 2200, Denmark
168. School of Medicine, University of Split, Split, 21000, Croatia
169. Centre for Global Health Research, Usher Institute of Population Health Sciences and Informatics, University of Edinburgh, Edinburgh, EH8 9AG, UK
170. Department of Clinical Physiology and Nuclear Medicine, Turku University Hospital, Turku, 20521, Finland
171. Research Centre of Applied and Preventive Cardiovascular Medicine, University of Turku, Turku, 20520, Finland
172. Centre for Non-Communicable Diseases, Karachi, Pakistan
173. Merck, Sharp & Dohme, Genetics and Pharmacogenomics, Boston, MA, 02115, USA
174. Department of Epidemiology, University of Washington, Seattle, WA, 98195, USA
175. Division of Cardiovascular Medicine, Brigham and Women's Hospital and Harvard Medical School, Boston, MA, 02115, USA
176. Nuffield Department of Clinical Medicine, Oxford, OX37 BN, UK
177. Institute for Maternal and Child Health - IRCCS "Burlo Garofolo", Trieste, 34137, Italy
178. Genetics of Complex Traits, University of Exeter Medical School, University of Exeter, Exeter, EX2 5DW, UK

179. Department of Biostatistics and Epidemiology, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA, 19104, USA
180. Division of Epidemiology & Community Health University of Minnesota, Minneapolis, MN, 55454, USA
181. Saw Swee Hock School of Public Health, National University Health System, National University of Singapore, Singapore 117549, Singapore
182. Genetics, Target Sciences, GlaxoSmithKline, Research Triangle Park, NC, 27709, US
183. OmicSoft a QIAGEN Company, Cary, NC, 27513, US
184. Department of Twin Research and Genetic Epidemiology, King's College London, London, SE1 7EH, UK
185. Division of Population Health Sciences, Ninewells Hospital and Medical School, University of Dundee, Dundee, UK
186. Generation Scotland, Centre for Genomic and Experimental Medicine, University of Edinburgh, Edinburgh, EH4 2XU, UK
187. deCODE Genetics/Amgen inc., Reykjavik, 101, Iceland
188. Faculty of Medicine, University of Iceland, Reykjavik, 101, Iceland
189. Chair of Genetic Epidemiology, IBE, Faculty of Medicine, LMU Munich, 81377, Germany
190. Biodemography of Aging Research Unit, Social Science Research Institute, Duke University, Durham, NC, 27708, USA
191. Department of Public Health, University of Helsinki, Helsinki, FI-00014, Finland
192. VU University Medical Center, Department of Epidemiology and Biostatistics, Amsterdam, 1007 MB, The Netherlands
193. Institute for Community Medicine, University Medicine Greifswald, Greifswald, 17475, Germany

194. Center for Pediatric Research, Department for Women's and Child Health, University of Leipzig, Leipzig, 04103, Germany
195. USC Roski Eye Institute, Department of Ophthalmology, Keck School of Medicine of the University of Southern California, Los Angeles, CA, 90033, USA
196. Anogia Medical Centre, Anogia, Greece
197. Centre for Vascular Prevention, Danube-University Krems, Krems, 3500, Austria
198. Dasman Diabetes Institute, Dasman, 15462, Kuwait
199. Diabetes Research Group, King Abdulaziz University, Jeddah, 21589, Saudi Arabia
200. Department of Public Health and Clinical Nutrition, University of Eastern Finland, Kuopio, 70210, Finland
201. Central Finland Central Hospital, Jyvaskyla, 40620, Finland
202. University of Eastern Finland, Kuopio, 70210, Finland
203. Radboud Institute for Health Sciences, Radboud University Medical Center, Nijmegen, 6500 HB, The Netherlands
204. Steno Diabetes Center Copenhagen, Gentofte, 2800, Denmark
205. Merck, Sharp & Dohme, Cardiometabolic Disease, Kenilworth, NJ, 07033, USA
206. Division of Public Health Sciences, Wake Forest School of Medicine, Winston-Salem, NC, 27157, USA
207. Institute of Cellular Medicine, The Medical School, Newcastle University, Newcastle, NE2 4HH, UK
208. Department of Medical Sciences, Cardiology, Uppsala Clinical Research Center, Uppsala University, Uppsala, 752 37, Sweden
209. Genetics, Target Sciences, GlaxoSmithKline, King of Prussia, PA, US

210. Departments of Epidemiology & Medicine, Diabetes Translational Research Center, Fairbanks School of Public Health & School of Medicine, Indiana University, Indiana, IN, 46202, USA
211. Green Lane Cardiovascular Service, Auckland City Hospital and University of Auckland, Auckland, New Zealand
212. Department of Human Genetics, University of Michigan, Ann Arbor, MI, 48109, USA
213. Department of Physiology and Biophysics, University of Mississippi Medical Center, Jackson, MS, 39216, USA
214. GlaxoSmithKline, King of Prussia, PA, 19406, USA
215. University of Glasgow, Glasgow, G12 8QQ, UK
216. Department of Clinical Sciences, Quantitative Biomedical Research Center, Center for the Genetics of Host Defense, University of Texas Southwestern Medical Center, Dallas, TX, 75390, USA
217. Princess Al-Jawhara Al-Brahim Centre of Excellence in Research of Hereditary Disorders (PACER-HD), King Abdulaziz University, Jeddah, 21589, Saudi Arabia
218. Institute of Social and Preventive Medicine, Lausanne University Hospital, Lausanne, 1010, Switzerland
219. Departments of Pediatrics and Genetics, Harvard Medical School, Boston, MA, 02115, USA
220. The Mindich Child Health and Development Institute, Icahn School of Medicine at Mount Sinai, New York, NY, 10069, USA
221. Department of Epidemiology and Carolina Center of Genome Sciences, Chapel Hill, NC, 27514, USA
222. Li Ka Shing Centre for Health Information and Discovery, The Big Data Institute, University of Oxford, Oxford, OX3 7BN, UK

ABSTRACT

Body fat distribution is a heritable risk factor for a range of adverse health consequences, including hyperlipidemia and type 2 diabetes. To identify protein-coding variants associated with body fat distribution, assessed by waist-to-hip ratio adjusted for body mass index, we analyzed 246,328 predicted coding and splice site variants available on exome arrays in up to 344,369 individuals from five major ancestries for discovery and 132,177 independent European-ancestry individuals for validation. We identified 15 common (minor allele frequency, MAF ≥ 5%) and 9 low frequency or rare (MAF < 5%) coding variants that have not been reported previously. Pathway/gene set enrichment analyses of all associated variants highlight lipid particle, adiponectin level, abnormal white adipose tissue physiology, and bone development and morphology as processes affecting fat distribution and body shape. Furthermore, the cross-trait associations and the analyses of variant and gene function highlight a strong connection to lipids, cardiovascular traits, and type 2 diabetes. In functional follow-up analyses, specifically in *Drosophila* RNAi-knockdown crosses, we observed a significant increase in the total body triglyceride levels for two genes (*DNAH10* and *PLXND1*). By examining variants often poorly tagged or entirely missed by genome-wide association studies, we implicate novel genes in fat distribution, stressing the importance of interrogating low-frequency and protein-coding variants.

Body fat distribution, as assessed by waist-to-hip ratio (WHR), is a heritable trait and a well-established risk factor for adverse metabolic outcomes[1-6]. A high WHR often indicates a large presence of intra-abdominal fat whereas a low WHR is correlated with a greater accumulation of gluteofemoral fat. Lower values of WHR have been consistently associated with lower risk of cardiometabolic diseases like type 2 diabetes (T2D)[7,8], or differences in bone structure and gluteal muscle mass[9]. These epidemiological associations are consistent with the results of our previously reported genome-wide association study (GWAS) of 49 loci associated with WHR (after adjusting for body mass index, WHRadjBMI)[10]. Notably, a genetic predisposition to higher WHRadjBMI is associated with increased risk of T2D and coronary heart disease (CHD), and this association appears to be causal[9].

More recently, large-scale genetic studies have identified ~125 common loci for central obesity, primarily non-coding variants of relatively modest effect, for different measures of body fat distribution[10-16]. Large scale interrogation of both common (minor allele frequency [MAF]≥5%) and low frequency or rare (MAF<5%) coding and splice site variation may lead to additional insights into the genetic and biological etiology of central obesity by narrowing in on causal genes contributing to trait variance. Thus, we set out to identify protein-coding and splice site variants associated with WHRadjBMI using exome array data and to explore their contribution to variation in WHRadjBMI through multiple follow-up analyses.

RESULTS

Protein-coding and splice site variation associated with body fat distribution

We conducted a 2-stage fixed-effects meta-analysis testing both additive and recessive models in order to detect protein-coding genetic variants that influence WHRadjBMI (**Online Methods, Figure 1**). Our stage 1 meta-analysis included up to 246,328 variants (218,195 with MAF<5%) from up to 344,369 individuals from 74 studies of European (N=288,492), South Asian (N=29,315), African

(N=15,687), East Asian (N=6,800) and Hispanic/Latino (N=4,075) descent, genotyped with an ExomeChip array (**Supplementary Tables 1-3).** For stage 2, we assessed 70 suggestively significant ($P<2x10^{-6}$) variants from stage 1 (**Online Methods, Supplementary Data 1-3**) in two independent cohorts from the United Kingdom [UK Biobank (UKBB), N=119,572] and Iceland (deCODE, N=12,605) (**Online Methods, Supplementary Data 1-3**) for a total stage 1+2 sample size of 476,546 (88% European). Variants were considered statistically significant in the total meta-analyzed sample (stage 1+2) when they achieved a significance threshold of $P<2x10^{-7}$ after Bonferroni correction for multiple testing (0.05/246,328 variants tested), and considered novel if they were greater than one megabase (Mb) from a previously-identified WHRadjBMI lead SNP[10-16].

In stages 1 and 2 combined all ancestry meta-analyses, we identified 48 coding variants (16 novel) across 43 genes, 47 identified assuming an additive model, and one more variant under a recessive model (**Table 1**). Due to the possible heterogeneity introduced by combining multiple ancestries[17], we also performed a European-only meta-analysis. Here, four additional coding variants were significant (three novel) assuming an additive model. Of these 52 significant variants (48 from the all ancestry and 4 from the European-only analyses), eleven were of low frequency, including seven novel variants in *RAPGEF3*, *FGFR2, R3HDML, HIST1H1T, PCNXL3, ACVR1C,* and *DARS2* (**Table 1, Supplementary Figures 1-7)**. These low frequency variants tended to display larger effect estimates than any of the previously reported common variants (**Figure 2**)[10]. In general, variants with MAF<1% had effect sizes approximately three times greater than those of common variants (MAF>5%). There are likely additional rare variants with smaller effects sizes that we are underpowered to detect. However, in the absence of common variants with similarly large effects, our results point to the importance of investigating rare and low frequency variants to identify variants with large effects (**Figure 2**).

Given the established differences in the genetic underpinnings between sexes for WHRadjBMI[10,11], we also performed sex-stratified analyses and report variants that were array-wide

497 significant ($P<2x10^{-7}$) in at least one sex stratum and exhibit significant sex-specific effects
498 ($P_{sexhet}<7.14x10^{-4}$, see **Online Methods**). We found four additional novel variants that were not identified
499 in the sex-combined meta-analyses (in *UGGT2* and *MMP14* for men only; and *DSTYK* and *ANGPTL4* for
500 women only) (**Table 2, Supplementary Figures 7-11**). Variants in *UGGT2* and *ANGPTL4* were of low
501 frequency (MAF_{men}=0.6% and MAF_{women}=1.9%, respectively). Additionally, 14 variants from the sex-
502 combined meta-analyses displayed stronger effects in women, including the novel, low frequency
503 variant in *ACVR1C* (rs55920843, MAF=1.1%). Overall, 19 of the 56 variants (32%) identified across all
504 meta-analyses (48 from all ancestry, 4 from European-only and 4 from sex-stratified analyses) showed
505 significant sex-specific effects on WHRadjBMI (**Figure 1**): 16 variants with significantly stronger effects in
506 women, and three in men (**Figure 1**).

507 In summary, we identified 56 array-wide significant coding variants ($P<2.0x10^{-7}$); 43 common (14
508 novel) and 13 low frequency or rare variants (9 novel). For all 55 significant variants from the additive
509 model only (47 from all ancestry, 4 from European-only, and 4 from sex-specific analyses), we examined
510 potential collider bias, i.e. potential bias in effect estimates caused by adjusting for a correlated and
511 heritable covariate like BMI, for the relevant sex stratum and ancestry (**Online Methods, Supplementary**
512 **Table 7, Supplementary Note 1**). Overall, 38 of the 55 variants were robust against collider bias[18,19]
513 across all primary and secondary meta-analyses ($P<2x10^{-7}$ following correction), and an additional three
514 variants were robust against collider bias in the women-only analysis but not in the sex-combined. The
515 effect estimates of the remaining 14 variants (**Supplementary Table 7, Supplementary Note 1**) did not
516 remain array-wide significant following correction. Thus, these 14 variants warrant further functional
517 investigations to quantify their impact on WHR, as a true association may still exist, although the effect
518 may be slightly overestimated in the current analysis.

519 Using stage 1 meta-analysis results, we then aggregated low frequency variants across genes
520 and tested their joint effect with both SKAT and burden tests[20] (**Supplementary Table 8**, **Online**

Methods). We identified five genes that reached array-wide significance ($P<2.5x10^{-6}$, 0.05/16,222 genes tested), *RAPGEF3*, *ACVR1C*, *ANGPTL4*, *DNAI1*, and *NOP2*. However, while all genes analyzed included more than one variant, none remained significant after conditioning on the single variant with the most significant p-value, suggesting these associations are driven by a single variant.

Conditional analyses

We next implemented conditional analyses to determine (1) the number of independent association signals the 56 array-wide significant coding (23 novel) variants represent, and (2) whether the 33 variants near known GWAS association signals (<+/- 1Mb) represented independent novel association signals. To determine if these variants were independent association signals, we used approximate joint conditional analyses to test for independence in stage 1 (**Online Methods**; **Supplementary Table 4**)[20]. Only the *RSPO3-KIAA0408* locus contains two independent variants 291 Kb apart, rs1892172 in *RSPO3* (MAF=46.1%, $P_{conditional}=4.37x10^{-23}$ in the combined sexes, and $P_{conditional}=2.4x10^{-20}$ in women) and rs139745911 in *KIAA0408* (MAF=0.9%, $P_{conditional}=3.68x10^{-11}$ in the combined sexes, and $P_{conditional}=1.46x10^{-11}$ in women; **Figure 3**).

Further, 33 of our significant variants are within one Mb of previously identified GWAS tag SNPs for WHRadjBMI. We again used approximate joint conditional analysis to test for independence in the stage 1 meta-analysis dataset and obtained further complementary evidence from the UKBB dataset where necessary (**Online Methods**). We identified one coding variant representing a novel independent signal in a known locus [*RREB1*; stage1 meta-analysis, rs1334576, EAF = 0.44, $P_{conditional}= 3.06x10^{-7}$, (**Supplementary Table 5, Figure 3 [B]**); UKBB analysis, rs1334576, *RREB1*, $P_{conditional}= 1.24x10^{-8}$, (**Supplementary Table 6**) in the sex-combined analysis.

In summary, we identified a total of 56 WHRadjBMI-associated coding variants in 41 independent association signals. Of these 41 independent association signals, 24 are new or independent of known GWAS-identified tag SNPs (either >1MB +/- or array-wide significant following conditional analyses) (**Figure 1**). The remaining non-GWAS-independent variants may assist in narrowing in on the causal variant or gene underlying these established association signals.

Gene set and pathway enrichment analysis

To determine if the significant coding variants highlight novel biological pathways and/or provide additional support for previously identified biological pathways, we applied two complementary pathway analysis methods using the EC-DEPICT (ExomeChip Data-driven Expression Prioritized Integration for Complex Traits) pathway analysis tool,[21,22] and PASCAL[23] (**Online Methods**). We examined 361 variants with suggestive significance ($P<5x10^{-4}$)[10,17] from the combined ancestries and combined sexes analysis, as well as variants that exhibited significant sex-specific effects ($P_{sexhet}<5x10^{-4}$).

The sex-combined analyses identified 49 significantly enriched gene sets (FDR<0.05) that grouped into 25 meta-gene sets (**Supplementary Note 2, Supplementary Data 4-5**). We noted a cluster of meta-gene sets with direct relevance to metabolic aspects of obesity ("enhanced lipolysis," "abnormal glucose homeostasis," "increased circulating insulin level," and "decreased susceptibility to diet-induced obesity"). While these pathway groups had previously been identified in the GWAS DEPICT analysis (**Figure 4**), many of the individual gene sets within these meta-gene sets were not significant in the previous GWAS analysis, such as "insulin resistance," "abnormal white adipose tissue physiology," and "abnormal fat cell morphology" **(Supplementary Data 4**, **Figure 4**, **Supplementary Figure 12a**), but represent similar biological underpinnings implied by the shared meta-gene sets. These analyses highlight novel genes that fall outside known GWAS loci, based on their strong contribution to the significantly enriched gene sets related to adipocyte and insulin biology (**Figure 4**).

To focus on novel findings, we conducted pathway analyses after excluding variants from previous WHRadjBMI analyses[10] (**Supplemental Note 2**). Seventy-five loci/genes were included in the DEPICT analysis, and we identified 26 significantly enriched gene sets (13 meta-gene sets). Here, all but one gene set, "lipid particle size", were related to skeletal biology. This result likely reflects an effect on the pelvic skeleton (hip circumference), shared signaling pathways between bone and fat (such as TGF-beta) and shared developmental origin[24] (**Supplementary Data 5**, **Supplementary Figure 12b**).

We used PASCAL (**Online Methods**) to further distinguish between enrichment based on *coding-only* variant associations (this study) and *regulatory-only* variant associations (up to 20 kb upstream of the gene from a previous GIANT study[10]). For completeness, we also compared the coding pathways to those that could be identified in the total previous GWAS effort (using both *coding and regulatory* variants) by PASCAL. The analysis revealed 109 significantly enriched coding pathways (FDR<0.05; **Supplementary Table 9**). A total of 111 gene sets were identified only in the coding+regulatory analysis that included ExomeChip data. Thus, while we observed high concordance in the -log10 (p-values) between ExomeChip and GWAS gene set enrichment (Pearson's r (coding *vs* regulatory only) = 0.38, $P<10^{-300}$; Pearson's r (coding *vs* coding+regulatory) = 0.51, $P<10^{-300}$), there are gene sets that seem to be enriched *specifically* for variants in coding regions (e.g., decreased susceptibility to diet-induced obesity, abnormal skeletal morphology) or unique to variants in regulatory regions (e.g. transcriptional regulation of white adipocytes) (**Supplementary Figure 13**).

Cross-trait associations

To assess the relevance of our identified variants with cardiometabolic (lipids, diabetes-related, blood pressure), anthropometric (height and BMI), and reproductive traits (age at menopause and menarche), we conducted association lookups from existing ExomeChip studies of 15 traits (**Supplementary Data 6**, **Supplementary Figure 14**). We found that variants in *STAB1* and *PLCB3* display the greatest number of significant cross-trait associations, each with seven different traits ($P<9.8x10^{-4}$,

0.05/51 variants tested). Of note, these two genes cluster together with *RSPO3*, *DNAH10*, *MNS1*, *COBLL1*, *CCDC92*, and *ITIH3*. The WHR-increasing allele in this cluster of variants exhibit a pattern of increased cardiometabolic risk (e.g. increased fasting insulin [FI], two-hour glucose [TwoHGlu], and triglycerides [TG]; and decreased high-density lipoprotein cholesterol [HDL]), but also decreased BMI (**Supplementary Data 6**, **Supplementary Figure 14**). Among the traits we examined, height (19 variants), HDL (18 variants), and BMI (16 variants) have the greatest number of significant associations with WHRadjBMI-associated ExomeChip variants. Many of our novel variants exhibit significant associations with lipid-related traits, including variants in *DAGLB* (HDL), *MGA* (HDL, TG), *RASIP1* (low-density lipoprotein cholesterol [LDL], TG, total cholesterol [TC]), and *IZUMO1* (LDL, TG, TC). Further, significant cross-trait associations are consistent with expected direction of effect for several traits, i.e. the WHR-increasing allele is associated with higher values of TG, DBP, fasting insulin, TC , LDL and T2D when compared to the WHR across all significant variants ($P<9.8x10^{-4}$). The WHR-increasing allele decreases HDL for 89% of significantly associated variants (**Supplementary Data 6**, **Supplementary Figure 14)**.

Given the established correlation between total body fat percentage and WHR (*R*= 0.052 to 0.483)[25-27], we examined the association of our top exome variants with both total body fat percentage and truncal fat percentage available in a sub-sample of up to 118,160 participants of UKBB (**Supplementary Tables 10-11**). Seven of the common novel variants were significantly associated (*P*<0.001, 0.05/48 variants examined) with both total body and truncal fat percentage in the sexes-combined analysis (*COBLL1*, *UHRF1BP1*, *WSCD2*, *CCDC92*, *IFI30*, *MPV17L2*, *IZUMO1*). Only one of our tag SNPs, rs7607980 in *COBLL1*, is nearby a known total body fat percentage GWAS locus (rs6738627; R^2=0.1989, distance=6751 bp, with our tag SNP)[28]. Two additional variants, rs62266958 in *EFCAB12* and rs224331 in *GDF5*, were significantly associated with truncal fat in the women-only analysis. Of the nine SNPs associated with at least one of these two traits, all variants displayed much greater magnitude of effect on truncal fat compared to total body fat (**Supplementary Figure 15**).

Previous studies have demonstrated the importance of examining common and rare variants within genes with mutations known to cause monogenic diseases[29,30]. We assessed enrichment of our WHRadjBMI within genes that cause monogenic forms of lipodystrophy) and/or insulin resistance (**Supplementary Data 7**). No significant enrichment was observed (**Supplementary Figure 16**). For lipodystrophy, the lack of significant findings may be due in part to the small number of implicated genes and the relatively small number of variants in monogenic disease causing genes, reflecting their intolerance of variation.

Genetic architecture of WHRadjBMI coding variants

We used summary statistics from our stage 1 results to estimate the phenotypic variance explained by ExomeChip coding variants. We calculated the variance explained by subsets of SNPs across various significance thresholds ($P< 2x10^{-7}$ to 0.2) and conservatively estimated using only independent tag SNPs (**Supplementary Table 12**, **Online Methods**, and **Supplementary Figure 17)**. The 22 independent significant coding SNPs in stage 1 account for 0.28% of phenotypic variance in WHRadjBMI. For independent variants that reached suggestive significance in stage 1 ($P<2x10^{-6}$), 33 SNPs explain 0.38% of the variation; however, the 1,786 independent SNPs with a liberal threshold of P<0.02 explain 13 times more variation (5.12%). While these large effect estimates may be subject to winner's curse, for array-wide significant variants, we detected a consistent relationship between effect magnitude and MAF in our stage 2 analyses in UK Biobank and deCODE (**Supplementary Data 1-3**). Notably, the Exomechip coding variants explained less of the phenotypic variance than in our previous GIANT investigation, wherein 49 significant SNPs explained 1.4% of the variance in WHRadjBMI. When considering all coding variants on the ExomeChip in men and women together, 46 SNPs with a $P<2x10^{-6}$ and 5,917 SNPs with a P<0.02 explain 0.51% and 13.75% of the variance in WHRadjBMI, respectively. As expected given the design of the ExomeChip, the majority of the variance explained is attributable to rare and low frequency coding variants (independent SNPs with MAF<1% and MAF<5% explain 5.18%

and 5.58%, respectively). However, for rare and low frequency variants, those that passed significance in stage 1 explain only 0.10% of the variance in WHRadjBMI. As in **Figure 2**, these results also indicate that there are additional coding variants associated with WHRadjBMI that remain to be discovered, particularly rare and low frequency variants with larger effects than common variants. Due to observed differences in association strength between women and men, we estimated variance explained for the same set of SNPs in women and men separately. As observed in previous studies[10], there was significantly ($P_{RsqDiff}$<0.002=0.05/21, Bonferroni-corrected threshold) more variance explained in women compared to men at each significance threshold considered (differences ranged from 0.24% to 0.91%).

To better understand the potential clinical impact of WHRadjBMI associated variants, we conducted penetrance analysis using the UKBB population (both sexes combined, and men- and women-only). We compared the number of carriers and non-carriers of the minor allele for each of our significant variants in centrally obese and non-obese individuals to determine if there is a significant accumulation of the minor allele in either the centrally obese or non-obese groups (**Online Methods**). Three rare and low frequency variants (MAF ≤ 1%) with larger effect sizes (effect size > 0.90) were included in the penetrance analysis using World Health Organization (WHO- obese women WHR>0.85 and obese men WHR>0.90) WHR cut-offs for central obesity. Of these, one SNV (rs55920843-*ACVR1C*; $P_{sex\text{-}combined}=9.25\times10^{-5}$; $P_{women}=4.85\times10^{-5}$) showed a statistically significant difference in the number of carriers and non-carriers of the minor allele when the two strata were compared (sex-combined obese carriers=2.2%; non-obese carriers=2.6%; women obese carriers=2.1%; non-obese women carriers=2.6% (**Supplementary Table 13, Supplementary Figure 18**). These differences were significant in women, but not in men ($P_{men}<5.5\times10^{-3}$ after Bonferroni correction for 9 tests) and agree with our overall meta-analysis results, where the minor allele (G) was significantly associated with higher WHRadjBMI in women only (**Tables 1 and 2**).

Evidence for functional role of significant variants

Drosophila Knockdown

Considering the genetic evidence of adipose and insulin biology in determining body fat distribution[10], and the lipid signature of the variants described here, we examined whole-body triglycerides levels in adult *Drosophila*, a model organism in which the fat body is an organ functionally analogous to mammalian liver and adipose tissue and triglycerides are the major source of fat storage[31]. Of the 51 genes harboring our 56 significantly associated variants, we identified 27 with *Drosophila* orthologues for functional follow-up analyses. In order to prioritize genes for follow-up, we selected genes with large changes in triglyceride storage levels (> 20% increase or > 40% decrease, as chance alone is unlikely to cause changes of this magnitude, although some decrease is expected) after considering each corresponding orthologue in an existing large-scale screen for adipose with ≤2 replicates per knockdown strain.[31] Two orthologues, for *PLXND1* and *DNAH10*, from two separate loci met these criteria. For these two genes, we conducted additional knockdown experiments with ≥5 replicates using tissue-specific drivers (fat body [cg-Gal4] and neuronal [elav-Gal4] specific RNAi-knockdowns) (**Supplementary Table 14**). A significant (P<0.025, 0.05/2 orthologues) increase in the total body triglyceride levels was observed in *DNAH10* orthologue knockdown strains for both the fat body and neuronal drivers. However, only the neuronal driver knockdown for *PLXND1* produced a significant change in triglyceride storage. *DNAH10* and *PLXND1* both lie within previous GWAS identified regions (**Box 1**). Adjacent genes have been highlighted as likely candidates for the *DNAH10* association region, including *CCDC92* and *ZNF664* based on eQTL evidence. However, our fly knockdown results support *DNAH10* as the causal genes underlying this association. Of note, rs11057353 in *DNAH10* showed suggestive significance after conditioning on the known GWAS variants in nearby *CCDC92* (sex-combined $P_{conditional}=7.56\times10^{-7}$; women-only rs11057353 $P_{conditional}= 5.86\times10^{-7}$, **Supplementary Table 6**; thus providing some evidence of multiple causal variants/genes underlying this association signal. Further

analyses are needed to determine whether the implicated coding variants from the current analysis are the putatively functional variants.

eQTL Lookups

To gain a better understanding of the potential functionality of novel and low frequency variants, we examined the *cis*-association of the identified variants with expression level of nearby genes in subcutaneous adipose tissue, visceral omental adipose tissue, skeletal muscle and pancreas from GTEx[32], and assessed whether the exome and eQTL associations implicated the same signal (**Online Methods**, **Supplementary Data 8-9, Supplementary Table 15**). The lead exome variant was associated with expression level of the coding gene itself for *DAGLB, MLXIPL, CCDC92, MAPKBP1, LRRC36* and *UQCC1*. However, at three of these loci (*MLXIPL, MAPKBP1,* and *LRRC36)*, the lead exome variant is also associated with expression level of additional nearby genes, and at three additional loci, the lead exome variant is only associated with expression level of nearby genes (*HEMK1* at *C3orf18; NT5DC2, SMIM4* and *TMEM110* at *STAB1/ITIH3;* and *C6orf106* at *UHRF1BP1*). Although detected with a missense variant, these loci are also consistent with a regulatory mechanism of effect as they are significantly associated with expression levels of genes, and the association signal may well be due to LD with nearby regulatory variants.

Some of the coding genes implicated by eQTL analyses are known to be involved in adipocyte differentiation or insulin sensitivity: e. g. for *MLXIPL*, the encoded carbohydrate responsive element binding protein is a transcription factor, regulating glucose-mediated induction of *de novo* lipogenesis in adipose tissue, and expression of its *beta*-isoform in adipose tissue is positively correlated with adipose insulin sensitivity[33,34]. For *CCDC92,* the reduced adipocyte lipid accumulation upon knockdown confirmed the involvement of its encoded protein in adipose differentiation[35].

Biological Curation

To gain further insight into the possible functional role of the identified variants, we conducted thorough searches of the literature and publicly available bioinformatics databases (**Supplementary Data 9-10**, **Box 1**, **Online Method**). Many of our novel low frequency variants are in genes that are intolerant of nonsynonymous mutations (e.g. *ACVR1C*, *DARS2*, *FGFR2*; ExAC Constraint Scores >0.5). Like previously identified GWAS variants, several of our novel coding variants lie within genes that are involved in glucose homeostasis (e.g. *ACVR1C*, *UGGT2*, *ANGPTL4*), angiogenesis (*RASIP1*), adipogenesis (*RAPGEF3*), and lipid biology (*ANGPTL4*, *DAGLB*) (**Supplementary Data 9**, **Box 1**).

DISCUSSION

Our two-staged approach to analysis of coding variants from ExomeChip data in up to 476,546 individuals identified a total of 56 array-wide significant variants in 41 independent association signals, including 24 newly identified (23 novel and one independent of known GWAS signals) that influence WHRadjBMI. Nine of these variants were low frequency or rare, indicating an important role for low frequency variants in the polygenic architecture of fat distribution and providing further insights into its underlying etiology. While, due to their rarity, these coding variants only explain a small proportion of the trait variance at a population level, they may, given their predicted role, be more functionally tractable than non-coding variants and have a critical impact at the individual and clinical level. For instance, the association between a low frequency variant (rs11209026; R381Q; MAF<5% in ExAC) located in the *IL23R* gene encoding a subunit of the interleukin 23 (IL23) receptor and multiple inflammatory diseases (such as psoriasis[36], rheumatoid arthritis[37], ankylosing spondylitis[38], and inflammatory bowel diseases[39]) led to the development of new therapies, targeting IL23 and IL12 in the same pathway (reviewed in [40-42]). Although a large proportion of variance at the population level still

needs to be accounted for in these inflammatory diseases, the contribution of this association to understanding disease mechanisms and the development of new therapies has been tremendous (reviewed in [41,42]). Thus, we are encouraged that our associated low frequency coding variants displayed large effect sizes; all but one of the nine novel low frequency variants had an effect size larger than the 49 SNPs reported in Shungin *et al.* 2015, and some of these effect sizes were up to 7-fold larger than those previously reported for GWAS. This finding mirrors results for other cardiometabolic traits[43], and suggests variants of possible clinical significance with even larger effect and lower frequency variants will likely be detected through larger additional genome-wide scans of many more individuals.

We continue to observe sexual dimorphism in the genetic architecture of WHRadjBMI[11]. Overall, we identified 19 coding variants that display significant sex differences, of which 16 (84%) display larger effects in women compared to men. Of the variants outside of GWAS loci, we reported three (two with MAF<5%) that show a significantly stronger effect in women and two (one with MAF<5%) that show a stronger effect in men. Additionally, genetic variants continue to explain a higher proportion of the phenotypic variation in body fat distribution in women compared to men[10,11]. Of the novel female (*DSTYK* and *ANGPTL4*) and male (*UGGT2* and *MMP14*) specific signals, only *ANGPTL4* implicated fat distribution related biology associated with both lipid biology and cardiovascular traits (**Box 1**). Sexual dimorphism in fat distribution is apparent from childhood and throughout adult life[44-46], and at sexually dimorphic loci, hormones with different levels in men and women may interact with genomic and epigenomic factors to regulate gene activity, though this remains to be experimentally documented. Dissecting the underlying molecular mechanisms of the sexual dimorphism in body fat distribution, and also how it is correlated with – and causing – important comorbidities like T2D and cardiovascular diseases will be crucial for improved understanding of disease risk and pathogenesis.

Overall, we observe fewer significant associations between WHRadjBMI and coding variants on the ExomeChip than Turcot *et al.* (*In press*) examining the association of low frequency and rare coding

754 variants with BMI. In line with these observations, we identify fewer pathways and cross-trait
755 associations. One reason for fewer WHRadjBMI implicated variants and pathways may be smaller
756 sample size ($N_{WHRadjBMI}$ = 476,546, N_{BMI} = 718,639), and thus, lower statistical power. Power, however, is
757 likely not the only contributing factor. For example, Turcot *et al.* (*In Press*) have comparative sample
758 sizes between BMI and that of Marouli *et al*.[22] studying height (N_{height} = 711,428). However, greater than
759 seven times the number of coding variants are identified for height than for BMI, indicating that perhaps
760 a number of other factors, including trait architecture, heritability (possibly overestimated in some
761 phenotypes), and phenotype precision, likely all contribute to our study's capacity to identify low
762 frequency and rare variants with large effects. Further, it is possible that the comparative lack of
763 significant findings for WHRadjBMI and BMI compared to height may be a result of higher selective
764 pressure against genetic predisposition to cardiometabolic phenotypes, such as BMI and WHR. As
765 evolutionary theory predicts that harmful alleles will be low frequency[47], we may need larger sample
766 sizes to detect rare variants that have so far escaped selective pressures. Lastly, the ExomeChip is
767 limited by the variants that are present on the chip, which was largely dictated by sequencing studies in
768 European-ancestry populations and a MAF detection criteria of ~0.012%. It is likely that though an
769 increased sample size, use of chips designed to detect variation across a range of continental ancestries,
770 and/or alternative study designs, future studies will detect additional variation from the entire allele
771 frequency spectrum that contributes to fat distribution phenotypes.

772 The collected genetic and epidemiologic evidence has now demonstrated that fat distribution
773 (as measured by increased WHRadjBMI) is correlated with increased risk of T2D and CVD, and that this
774 association is likely causal with potential mediation through blood pressure, triglyceride-rich
775 lipoproteins, glucose, and insulin[9]. This observation yields an immediate follow-up question: Which
776 mechanisms regulate depot-specific fat accumulation and are risks for disease, driven by increased
777 visceral or decreased subcutaneous adipose tissue mass (or both)? Pathway analysis identified several

778 novel pathways and gene sets related to metabolism and adipose regulation, bone growth and
779 development. Similarly, expression/eQTL results support the function and relevance of adipogenesis,
780 adipocyte biology, and insulin signaling, supporting our previous findings for WHRadjBMI[10]. We also
781 provide evidence suggesting known biological functions and pathways contributing to body fat
782 distribution (e.g., diet-induced obesity, angiogenesis, bone growth and morphology, and enhanced
783 lipolysis).

784 A seminal finding from this study is the importance of lipid metabolism for body fat distribution.
785 In fact, pathway analyses that highlight enhanced lipolysis, cross-trait associations with circulating lipid
786 levels, existing biological evidence from the literature, and knockdown experiments in *Drosophila*
787 examining triglyceride storage point to novel candidate genes (*ANGPTL4*, *ACVR1C*, *DAGLB*, *MGA*, *RASIP1*,
788 and *IZUMO1*) and new candidates in known regions (*DNAH10*[10] and *MLXIPL*[14]) related to lipid biology
789 and its role in fat storage. Newly implicated genes of interest include *ACVR1C*, *MLXIPL*, and *ANGPTL4*, all
790 of which are involved in lipid homeostasis; all are excellent candidate genes for central adiposity.
791 Carriers of inactivating mutations in *ANGPTL4* (*Angiopoietin Like 4*), for example, display low triglyceride
792 levels and low risk of coronary artery disease[48]. *ACVR1C* encodes the activin receptor-like kinase 7
793 protein (ALK7), a receptor for the transcription factor TGFB-1, well known for its central role in growth
794 and development in general[49-53], and adipocyte development in particular[53]. *ACVR1C* exhibits the highest
795 expression in adipose tissue, but is also highly expressed in the brain[54-56]. In mice, decreased activity of
796 *ACVR1C* upregulates PPARγ and C/EBPα pathways and increases lipolysis in adipocytes, thus decreasing
797 weight and diabetes in mice[54,57,58]. Such activity is suggestive of a role for ALK7 in adipose tissue
798 signaling and therefore for therapeutic targets for human obesity. *MLXIPL*, also important for lipid
799 metabolism and postnatal cellular growth, is a transcription factor which activates triglyceride synthesis
800 genes in a glucose-dependent manner[59,60]. The lead exome variant in this gene is highly conserved, most
801 likely damaging, and is associated with reduced *MLXIPL* expression in adipose tissue. Furthermore, in a

recent longitudinal, *in vitro* transcriptome analysis of adipogenesis in human adipose-derived stromal cells, gene expression of *MLXIPL* was up-regulated during the maturation of adipocytes, suggesting a critical role in the regulation of adipocyte size and accumulation[61].

Taken together, our 24 novel variants for WHRadjBMI offer new biology, highlighting the importance of lipid metabolism in the genetic underpinnings of body fat distribution. We continue to demonstrate the critical role of adipocyte biology and insulin resistance for central obesity and offer support for potentially causal genes underlying previously identified fat distribution GWAS loci. Notably, our findings offer potential new therapeutic targets for intervention in the risks associated with abdominal fat accumulation, and represents a major advance in our understanding of the underlying biology and genetic architecture of central adiposity.

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HDW; (Health) AL; (HELIC MANOLIS) EZ, GD; (HELIC Pomak) EZ, GD; (HUNT-MI) KH, CJW; (Inter99) TH, TJ; (IRASFS) LEW, EKS; (Jackson Heart Study (JHS)) JGW; (KORA S4) KS, IMH; (Leipzig-Adults) MB, PK; (LOLIPOP-Exome) JCC, JSK; (LOLIPOP-OmniEE) JCC, JSK; (MESA) JIR, XG; (METSIM) JK, ML; (MONICA-Brianza) GC; (Montreal Heart Institute Biobank (MHIBB)) MPD, GL, SdD, JCT; (MORGAM Central Laboratory) MP; (MORGAM Data Centre) KK; (OBB) FK; (PCOS) APM, CML; (PIVUS) CML, LL; (PRIME - Belfast) FK; (PRIME - Lille) PA; (PRIME - Strasbourg) MM; (PRIME - Toulouse) JF; (PROMIS) DS; (QC) MAR; (RISC) BB, EF, MW; (Rotterdam Study I) AGU, MAI; (SEARCH) AMD; (SHIP/SHIP-Trend) MD; (SIBS) DFE; (SOLID TIMI-52) DMW; (SORBS) APM, MS, AT; (The Mount Sinai BioMe Biobank) EPB, RJFL; (The NEO Study) DOMK; (The NHAPC study, The GBTDS study) XL; (The Western Australian Pregnancy Cohort (Raine) Study) CEP, SM; (TwinsUK) TDS; (ULSAM) APM; (Vejle Biobank) IB, CC, OP; (WGHS) DIC, PMR; (Women's Health Initiative) PLA; (WTCCC-UKT2D) MIM, KRO; (YFS) TL, OTRa; Genotyping of contributing studies: (1958 Birth Cohort) KES; (Airwave) EE, MPSL; (AMC PAS) SS; (Amish) LMYA, JAP; (ARIC) EWD, MG; (BBMRI-NL) SHV, LB, CMvD, PIWdB; (BRAVE) EDA; (Cambridge Cancer Studies) JGD; (CARDIA) MF; (CHD Exome + Consortium) ASB, JMMH, DFR, JD, RY(Clear/eMERGE (Seattle)) GPJ; (CROATIA_Korcula) VV; (DIACORE) CAB, MG; (DPS) AUJ, JL; (DRSEXTRA) PK; (EGCUT) TE; (EPIC-Potsdam) MBS, KM; (EpiHealth) EI, PWF; (Family Heart Study) KDT; (Fenland, EPIC) RAS; (Fenland, EPIC, InterAct) NJW, CL; (FUSION) NN; (FVG) IG, AM; (Generation Scotland) CH; (Genetic Epidemiology Network of Arteriopathy (GENOA)) SLRK, JAS; (GRAPHIC) NJS; (GSK-STABILITY) DMW; (Health) JBJ; (HELIC MANOLIS) LS; (HELIC Pomak) LS; (Inter99) TH, NG; (KORA) MMN; (KORA S4) KS, HG; (Leipzig-Adults) AM; (LOLIPOP-Exome) JCC, JSK; (LOLIPOP-OmniEE) JCC, JSK; (MESA) JIR, YDIC, KDT; (METSIM) JK, ML; (Montreal Heart Institute Biobank (MHIBB)) MPD; (OBB) FK; (PCOS) APM; (PIVUS) CML; (Rotterdam Study I) AGU, CMG, FR; (SDC) JMJ, HV; (SEARCH) AlMD; (SOLID TIMI-52) DMW; (SORBS) APM; (The Mount Sinai BioMe Biobank) EPB, RJFL, YL, CS; (The NEO Study) RLG; (The NHAPC study, The GBTDS study) XL, HL, YH; (The Western Australian Pregnancy Cohort (Raine) Study) CEP, SM; (TUDR) ZA; (TwinsUK) APM; (ULSAM) APM; (WGHS)

DIC, AYC; (Women's Health Initiative) APR; (WTCCC-UKT2D) MIM; (YFS) TL, LPL; Phenotyping of contributing studies: (Airwave) EE; (AMC PAS) SS; (Amish) LM YA; (ARIC) EWD; (ARIC, Add Health) KEN; (BBMRI-NL) SHV; (BRAVE) EDA; (BRIGHT) MJC; (CARL) AR, GG; (Cebu Longitudinal Health and Nutrition Survey) NRL; (CHES) RV, MT; (Clear/eMERGE (Seattle)) GPJ, AAB; (CROATIA_Korcula) OP, IR; (DIACORE) CAB, BKK; (DPS) AUJ, JL; (EFSOCH) ATH; (EGCUT) EM; (EPIC-Potsdam) HB; (EpiHealth) EI; (EXTEND) ATH; (Family Heart Study) MFF; (Fenland, EPIC, InterAct) NJW; (FIN-D2D 2007) LM, MV; (FINRISK) SM; (FINRISK 2007 (T2D)) PJ, HS; (Framingham Heart Study) CSF; (Generation Scotland) CH, BHS; (Genetic Epidemiology Network of Arteriopathy (GENOA)) SLRK, JAS; (GRAPHIC) NJS; (GSK-STABILITY) LW, HDW; (Health) AL, BHT; (HELIC MANOLIS) LS, AEF, ET; (HELIC Pomak) LS, AEF, MK; (HUNT-MI) KH, OH; (Inter99) TJ, NG; (IRASFS) LEW, BK; (KORA) MMN; (LASA (BBMRI-NL)) KMAS; (Leipzig-Adults) MB, PK; (LOLIPOP-Exome) JCC, JSK; (LOLIPOP-OmniEE) JCC, JSK; (MESA) MA; (Montreal Heart Institute Biobank (MHIBB)) GL, KSL, VT; (MORGAM Data Centre) KK; (OBB) FK, MN; (PCOS) CML; (PIVUS) LL; (PRIME - Belfast) FK; (PRIME - Lille) PA; (PRIME - Strasbourg) MM; (PRIME - Toulouse) JF; (RISC) BB, EF; (Rotterdam Study I) MAI, CMGFR, MCZ; (SHIP/SHIP-Trend) NF; (SORBS) MS, AT; (The Mount Sinai BioMe Biobank) EPB, YL, CS; (The NEO Study) RdM; (The NHAPC study, The GBTDS study) XL, HL, LS, FW; (The Western Australian Pregnancy Cohort (Raine) Study) CEP; (TUDR) YJH, WJL; (TwinsUK) TDS, KSS; (ULSAM) VG; (WGHS) DIC, PMR; (Women's Health Initiative) APR; (WTCCC-UKT2D) MIM, KRO; (YFS) TL, OTR; Data analysis of contributing studies: (1958 Birth Cohort) KES, IN; (Airwave) EE, MPSL; (AMC PAS) SS; (Amish) JRO'C, LMYA, JAP; (ARIC, Add Health) KEN, KLY, MG; (BBMRI-NL) LB; (BRAVE) RC, DSA; (BRIGHT) HRW; (Cambridge Cancer Studies) JGD, AP, DJT; (CARDIA) MF, LAL; (CARL) AR, DV; (Cebu Longitudinal Health and Nutrition Survey) YW; (CHD Exome + Consortium) ASB, JMMH, DFR, RY, PS; (CHES) YJ; (CROATIA_Korcula) VV; (deCODE) VS, GT; (DHS) AJC, PM, MCYN; (DIACORE) CAB, MG; (EFSOCH) HY; (EGCUT) TE, RM; (eMERGE (Seattle)) DSC; (ENDO) TK; (EPIC) JHZ; (EPIC-Potsdam) KM; (EpiHealth) SG; (EXTEND) HY; (Family Heart Study) MFF; (Fenland) JaL; (Fenland, EPIC) RAS; (Fenland, InterAct) SMW;

(Finrisk Extremes and QC) SV; (Framingham Heart Study) CTL, NLHC; (FVG) IG; (Generation Scotland) CH, JM; (Genetic Epidemiology Network of Arteriopathy (GENOA)) LFB; (GIANT-Analyst) AEJ; (GRAPHIC) NJS, NGDM, CPN; (GSK-STABILITY) DMW, AS; (Health) JBJ; (HELIC MANOLIS) LS; (HELIC Pomak) LS; (HUNT-MI) WZ; (Inter99) NG; (IRASFS) BK; (Jackson Heart Study (JHS)) LAL, JL; (KORA S4) TWW; (LASA (BBMRI-NL)) KMAS; (Leipzig-Adults) AM; (LOLIPOP-Exome) JCC, JSK, WZ; (LOLIPOP-OmniEE) JCC, JSK, WZ; (MESA) JIR, XG, JY; (METSIM) XS; (Montreal Heart Institute Biobank (MHIBB)) JCT, GL, KSL, VT; (OBB) AM; (PCOS) APM, TK; (PIVUS) NR; (PROMIS) AR, WZ; (QC GoT2D/T2D-GENES (FUSION, METSIM, etc)) AEL; (RISC) HY; (Rotterdam Study I) CMG, FR; (SHIP/SHIP-Trend) AT; (SOLID TIMI-52) DMW, AS; (SORBS) APM; (The Mount Sinai BioMe Biobank) YL, CS; (The NEO Study) RLG; (The NHAPC study, The GBTDS study) XL, HL, YH; (The Western Australian Pregnancy Cohort (Raine) Study) CAW; (UK Biobank) ARW; (ULSAM) APM, AM; (WGHS) DIC, AYC; (Women's Health Initiative) PLA, JH; (WTCCC-UKT2D) WG; (YFS) LPL.

METHODS

Studies

Stage 1 consisted of 74 studies (12 case/control studies, 59 population-based studies, and five family studies) comprising 344,369 adult individuals of the following ancestries: 1) European descent (N= 288,492), 2) African (N= 15,687), 3) South Asian (N= 29,315), 4) East Asian (N=6,800), and 5) Hispanic (N=4,075). Stage 1 meta-analyses were carried out in each ancestry separately and in the all ancestries group, for both sex-combined and sex-specific analyses. Follow-up analyses were undertaken in 132,177 individuals of European ancestry from the deCODE anthropometric study and UK Biobank (**Supplementary Tables 1-3**). Conditional analyses were performed in the all ancestries and European descent groups.

Phenotypes

For each study, WHR (waist circumference divided by hip circumference) was corrected for age, BMI, and the genomic principal components (derived from GWAS data, the variants with MAF >1% on the ExomeChip, and ancestry informative markers available on the ExomeChip), as well as any additional study-specific covariates (e.g. recruiting center), in a linear regression model. For studies with non-related individuals, residuals were calculated separately by sex, whereas for family-based studies sex was included as a covariate in models with both men and women. Additionally, residuals for case/control studies were calculated separately. Finally, residuals were inverse normal transformed and used as the outcome in association analyses. Phenotype descriptives by study are shown in Supplementary Table 3.

Genotypes and QC

The majority of studies followed a standardized protocol and performed genotype calling using the algorithms indicated in Supplementary Table 2, which typically included zCall[3]. For 10 studies participating in the Cohorts for Heart and Aging Research in Genomic Epidemiology (CHARGE) Consortium, the raw intensity data for the samples from seven genotyping centers were assembled into a single project for joint calling[4]. Study-specific quality control (QC) measures of the genotyped variants were implemented before association analysis (**Supplementary Tables 1-2**).

Study-level statistical analyses

Individual cohorts were analyzed for each ancestry separately, in sex-combined and sex-specific groups, with either RAREMETALWORKER (http://genome.sph.umich.edu/wiki/RAREMETALWORKER) or RVTESTs (http://zhanxw.github.io/rvtests/), to associate inverse normal transformed WHRadjBMI with genotype accounting for cryptic relatedness (kinship matrix) in a linear mixed model. These software programs are designed to perform score-statistic based rare-variant association analysis, can accommodate both unrelated and related individuals, and provide single-variant results and variance-

covariance matrices. The covariance matrix captures linkage disequilibrium (LD) relationships between markers within 1 Mb, which is used for gene-level meta-analyses and conditional analyses[62,63]. Single-variant analyses were performed for both additive and recessive models.

Centralized quality-control

Individual cohorts identified ancestry population outliers based on 1000 Genome Project phase 1 ancestry reference populations. A centralized quality-control procedure implemented in EasyQC[64] was applied to individual cohort association summary statistics to identify cohort-specific problems: (1) assessment of possible errors in phenotype residual transformation; (2) comparison of allele frequency alignment against 1000 Genomes Project phase 1 reference data to pinpoint any potential strand issues, and (3) examination of quantile-quantile (QQ) plots per study to identify any inflation arising from population stratification, cryptic relatedness and genotype biases.

Meta-analyses

Meta-analyses were carried out in parallel by two different analysts at two sites using RAREMETAL[62]. During the meta-analyses, we excluded variants if they had call rate <95%, Hardy-Weinberg equilibrium P-value $<1x10^{-7}$, or large allele frequency deviations from reference populations (>0.6 for all ancestries analyses and >0.3 for ancestry-specific population analyses). We also excluded from downstream analyses markers not present on the Illumina ExomeChip array 1.0, variants on the Y-chromosome or the mitochondrial genome, indels, multiallelic variants, and problematic variants based on the Blat-based sequence alignment analyses. Significance for single-variant analyses was defined at an array-wide level ($P<2x10^{-7}$). For all suggestive significant variants from Stage 1, we tested for significant sex differences. We calculated Psexhet for each SNP, testing for difference between women-specific and men-specific beta estimates and standard errors using EasyStrata[11,65] Each SNP that reached Psexhet<0.05/# of variants tested was considered significant.

For the gene-based analyses, we applied two sets of criteria to select variants with a MAF<5% within each ancestry based on coding variant annotation from five prediction algorithms (PolyPhen2, HumDiv and HumVar, LRT, MutationTaster and SIFT)[65,66]. Our broad gene-based tests included nonsense, stop-loss, splice site, and missense variants annotated as damaging by at least one algorithm mentioned above. Our strict gene-based tests included only nonsense, stop-loss, splice site, and missense variants annotated as damaging by all five algorithms. These analyses were performed using the sequence kernel association test (SKAT) and variable threshold (VT) methods. Statistical significance for gene-based tests was set at a Bonferroni-corrected threshold of $P<2.5x10^{-6}$. All gene-based tests were performed in RAREMETAL[62].

Genomic inflation

We observed a marked genomic inflation of the test statistics even after controlling for population stratification (linear mixed model) arising mainly from common markers; λGC in the primary meta-analysis (combined ancestries and combined sexes) was 1.08 and 1.43 for all and only common markers, respectively (**Supplementary Figures 4 and 7** and **Supplementary Table 16**). Such inflation is expected for a highly polygenic trait like WHRadjBMI, for studies using a non-random set of variants across the genome, and is consistent with our very large sample size[64,67,68].

Conditional analyses

The RAREMETAL R-package[62] was used to identify independent WHRadjBMI association signals across all ancestries and European meta-analysis results. RAREMETAL performs conditional analyses by using covariance matrices to distinguish true signals from the shadows of adjacent significant variants in LD. First, we identified the lead variants ($P<2x10^{-7}$) based on a 1Mb window centered on the most significantly associated variant. We then conditioned on the lead variants in RAREMETAL and kept new

lead signals at $P<2x10^{-7}$ for conditioning in a second round of analysis. The process was repeated until no additional signal emerged below the pre-specified P-value threshold ($P<2x10^{-7}$).

To test if the associations detected were independent of the previously published WHRadjBMI variants [10,14,16], we performed conditional analyses in the stage 1 discovery set if the GWAS variant or its proxy ($r^2 \geq 0.8$) was present on the ExomeChip using RAREMETAL[62]. All variants identified in our meta-analysis and the previously published variants were also present in the UK Biobank dataset[69]. This dataset was used as a replacement dataset if a good proxy was not present on the ExomeChip as well as a replication dataset for the variants present on the ExomeChip. All conditional analyses in the UK Biobank dataset were performed using SNPTEST[70-72]. The conditional analyses were carried out reciprocally, conditioning on the exome chip variant and then the previously published variant. An association was considered independent of the previously published association if there was a statistically significant association detected prior to the conditional analysis ($P<2x10^{-7}$) with both the exome chip variant and the previously published variant, and the observed association with both or either of the variants disappeared upon conditional analysis ($P>0.05$). A conditional p-value between $9x10^{-6}$ and 0.05 was considered inconclusive. However, a conditional p-value < $9x10^{-6}$ was also considered suggestive.

Stage 2 meta-analyses

In our Stage 2, we sought to validate a total of 70 variants from Stage 1 that met $P<2x10^{-6}$ in two independent studies, the UK Biobank (Release 1[69]) and Iceland (deCODE), comprising 119,572 and 12,605 individuals, respectively (Supplementary Tables 1-3). The same QC and analytical methodology were used for these studies. Genotyping, study descriptions and phenotype descriptives are provided in Supplementary Tables 11, 12 and 13. For the combined analysis of Stage 1 plus 2, we used the inverse-variance weighted fixed effects meta-analysis method. Significant associations were defined as those

nominally significant (P<0.05) in the Stage 2 study and for the combined meta-analysis (Stage 1 plus Stage 2) significance was set at $P<2x10^{-7}$.

Pathway enrichment analyses: DEPICT

We adapted DEPICT, a gene set enrichment analysis method for GWAS data, for use with the ExomeChip ('EC-DEPICT'); this method is also described in a companion manuscript[22]. DEPICT's primary innovation is the use of "reconstituted" gene sets, where many different types of gene sets (e.g. canonical pathways, protein-protein interaction networks, and mouse phenotypes) were extended through the use of large-scale microarray data (see Pers et al.[21] for details). EC-DEPICT computes p-values based on Swedish ExomeChip data (Malmö Diet and Cancer (MDC), All New Diabetics in Scania (ANDIS), and Scania Diabetes Registry (SDR) cohorts, N=11,899) and, unlike DEPICT, takes as input only the genes directly containing the significant (coding) variants rather than all genes within a specified amount of linkage disequilibrium (see **Supplementary Note 2**).

Two analyses were performed for WHRadjBMI ExomeChip: one with all variants $p<5x10^{-4}$ (49 significant gene sets in 25 meta-gene sets, FDR <0.05) and one with all variants > 1 Mb from known GWAS loci [10] (26 significant gene sets in 13 meta-gene sets, FDR <0.05). Affinity propagation clustering[73] was used to group highly correlated gene sets into "meta-gene sets"; for each meta-gene set, the member gene set with the best p-value was used as representative for purposes of visualization (see Supplementary Note). DEPICT for ExomeChip was written using the Python programming language, and the code can be found at https://github.com/RebeccaFine/obesity-ec-depict.

Pathway enrichment analyses: PASCAL

We also applied the PASCAL pathway analysis tool[23] to exome-wide association summary statistics from Stage 1 for all coding variants. The method derives gene-based scores (both SUM and MAX statistics) and subsequently tests for over-representation of high gene scores in predefined

biological pathways. We used standard pathway libraries from KEGG, REACTOME and BIOCARTA, and also added dichotomized (Z-score>3) reconstituted gene sets from DEPICT[21]. To accurately estimate SNP-by-SNP correlations even for rare variants, we used the UK10K data (TwinsUK[74] and ALSPAC[75] studies , N=3781). In order to separate the contribution of regulatory variants from the coding variants, we also applied PASCAL to association summary statistics of only regulatory variants (20 kb upstream) and regulatory+coding variants from the Shungin et al[10] study. In this way, we could comment on what is gained by analyzing coding variants available on ExomeChip arrays. We performed both MAX and SUM estimations for pathway enrichment. MAX is more sensitive to genesets driven primarily by a single signal, while SUM is better when there are multiple variant associations in the same gene.

Monogenic obesity enrichment analyses

We compiled two lists consisting of 31 genes with strong evidence that disruption causes monogenic forms of insulin resistance or diabetes; and 8 genes with evidence that disruption causes monogenic forms of lipodystrophy. To test for enrichment of association, we conducted simulations by matching each gene with others based on gene length and number of variants tested, to create a matched set of genes. We generated 1,000 matched gene sets from our data, and assessed how often the number of variants exceeding set significance thresholds was greater than in our monogenic obesity gene set.

Variance explained

We estimated the phenotypic variance explained by the association signals in Stage 1 all ancestries analyses for men, women, and combined sexes[76]. For each associated region, we pruned subsets of SNPs within 500 kb, as this threshold was comparable with previous studies, of the SNPs with the lowest P-value and used varying P value thresholds (ranging from $2x10^{-7}$ to 0.02) from the combined sexes results. Additionally, we examined all variants and independent variants across a range of MAF

thresholds. The variance explained by each subset of SNPs in each strata was estimated by summing the variance explained by the individual top coding variants. For the comparison of variance explained between men and women, we tested for the significance of the differences assuming that the weighted sum of chi-squared distributed variables tend to a Gaussian distribution ensured by Lyapunov's central limit theorem.[76,77]

Cross-trait lookups

To carefully explore the relationship between WHRadjBMI and related cardiometabolic, anthropometric, and reproductive traits, association results for the 51 WHRadjBMI coding SNPs were requested from existing or on-going meta-analyses from 7 consortia, including ExomeChip data from GIANT (BMI, height), Global Lipids Genetics Consortium Results (GLGC) (total cholesterol, triglycerides, HDL-cholesterol, LDL-cholesterol), International Consortium for Blood Pressure (IBPC)[78] (systolic and diastolic blood pressure), Meta-Analyses of Glucose and Insulin-related traits Consortium (MAGIC) (glycemic traits), and DIAbetes Genetics Replication And Meta-analysis (DIAGRAM) consortium (type 2 diabetes). For coronary artery disease, we accessed 1000 Genomes Project-imputed GWAS data released by CARDIoGRAMplusC4D[79] and for the ReproGen consortium (age at menarche and menopause) we used a combination of ExomeChip and 1000 Genomes Project-Imputed GWAS data. Heatmaps were generated in R v3.3.2 using gplots (https://CRAN.R-project.org/package=gplots). We used Euclidean distance based on p-value and direction of effect and complete linkage clustering for the dendrograms.

Body-fat percentage associations

We performed body fat percent and truncal fat percent look-up of 48 of the 56 identified variants (tables 1 and 2) that were available in the UK Biobank, Release 1[69], data (notably some of the rare variants in table 1 and 2 were not available) to further characterize their effects on WHRadjBMI.

Genome-wide association analyses for body fat percent and truncal fat percent were carried out in the UK Biobank. Prior to analysis, phenotype data were filtered to exclude pregnant or possibly pregnant women, individuals with body mass index < 15, and without genetically confirmed European ancestry, resulting in a sample size of 120,286. Estimated measures of body fat percent and truncal fat percent were obtained using the Tanita BC418MA body composition analyzer (Tanita, Tokyo, Japan). Individuals were not required to fast and did not follow any specific instructions prior to the bioimpedance measurements. SNPTEST was used to perform the analyses based on residuals adjusted for age, 15 principle components, assessment center and the genotyping chip[70].

Collider bias

In order to evaluate SNPs for possible collider bias, we used results from an ongoing association analysis from GIANT on BMI to first identify SNPs with effects in the opposite direction between WHRadjBMI and BMI and with P<0.05 for BMI. For each SNP that met these criteria, WHRadjBMI associations were adjusted for the correlation between the two traits to obtain new effect estimates using the following equations:

$$\beta_{\text{corrected}} = \beta_{WHRadjBMI} + \beta_{BMI} \times \rho$$

,and

$$SE_{corrected} = \sqrt{(SE^2_{WHRadjBMI} + \rho^2 \times SE^2_{BMI}}$$

where ρ is the phenotypic correlation between WHR and BMI (0.49).

Drosophila RNAi knockdown experiments

For each gene in which coding variants were associated with WHRadjBMI in the final combined meta-analysis ($P < 2\times10^{-7}$), its corresponding Drosophila orthologues were identified in the Ensembl

ortholog database (www.ensembl.org), when available. Drosophila triglyceride content values were mined from a publicly available genome-wide fat screen data set [31] to identify potential genes for follow-up knockdowns. Estimated values represent fractional changes in triglyceride content in adult male flies. Data are from male progeny resulting from crosses of male UAS-RNAi flies from the Vienna Drosophila Resource Center (VDRC) and Hsp70-GAL4; Tub-GAL8ts virgin females. Two-to-five-day-old males were sorted into groups of 20 and subjected to two one-hour wet heatshocks four days apart. On the seventh day, flies were picked in groups of eight, manually crushed and sonicated, and the lysates heat-inactivated for 10 min in a thermocycler at 95 °C. Centrifuge-cleared supernatants were then used for triglyceride (GPO Trinder, Sigma) and protein (Pierce) determination. Triglyceride values from these adult-induced ubiquitous RNAi knockdown individuals were normalized to those obtained in parallel from non-heatshocked progeny from the very same crosses. The screen comprised one to three biological replicates. We followed up each gene with a >0.2 increase or >0.4 decrease in triglyceride content.

Orthologues for two genes were brought forward for follow-up, *DNAH10* and *PLXND1*. For both genes, we generated adipose tissue (cg-Gal4) and neuronal (elav-Gal4) specific RNAi-knockdown crosses, leveraging upstream activation sequence (UAS)-inducible short-hairpin knockdown lines, available through the VDRC (Vienna *Drosophila* Resource Center). We crossed male UAS-RNAi flies and elav-GAL4 or CG-GAL4 virgin female flies. All fly experiments were carried out at 25°C. Five-to-seven-day-old males were sorted into groups of 20, weighed and homogenated in PBS with 0.05% Tween with Lysing Matrix D in a beadshaker. The homogenate was heat-inactivated for 10 min in a thermocycler at 70°C. 10µl of the homogenate was subsequently used in a triglyceride assay (Sigma, Serum Triglyceride Determination Kit) which was carried out in duplicate according to protocol, with one alteration: the samples were cleared of residual particulate debris by centrifugation before absorbance reading. Resulting triglyceride

values were normalized to fly weight and larval/population density. We used the non-parametric Kruskall-Wallis test to compare wild type with knockdown lines.

Expression quantitative trait loci (eQTLs) analysis

We queried the significant variant (Exome coding SNPs)-gene pairs associated with eGenes across five metabolically relevant tissues (skeletal muscle, subcutaneous adipose, visceral adipose, liver and pancreas) with at least 70 samples in the GTEx database[32]. For each tissue, variants were selected based on the following thresholds: the minor allele was observed in at least 10 samples, and the minor allele frequency was ≥ 0.01. eGenes, genes with a significant eQTL, are defined on a false discovery rate (FDR)[80] threshold of ≤0.05 of beta distribution-adjusted empirical p-value from FastQTL. Nominal p-values were generated for each variant-gene pair by testing the alternative hypothesis that the slope of a linear regression model between genotype and expression deviates from 0. To identify the list of all significant variant-gene pairs associated with eGenes, a genome-wide empirical p-value threshold[64], pt, was defined as the empirical p-value of the gene closest to the 0.05 FDR threshold. pt was then used to calculate a nominal p-value threshold for each gene based on the beta distribution model (from FastQTL) of the minimum p-value distribution f(pmin) obtained from the permutations for the gene. For each gene, variants with a nominal p-value below the gene-level threshold were considered significant and included in the final list of variant-gene pairs[64]. For each eGene, we also listed the most significantly associated variants (eSNP). Only these exome SNPs with $r^2 > 0.8$ with eSNPs were considered for the biological interpretation (Supplementary eQTL GTEx).

We also performed cis-eQTL analysis in 770 METSIM subcutaneous adipose tissue samples as described in Civelek, et al.[81] A false discovery rate (FDR) was calculated using all p-values from the cis-eQTL detection in the q-value package in R. Variants associated with nearby genes at an FDR less than 1% were considered to be significant (equivalent p-value $< 2.46 \times 10^{-4}$).

For loci with more than one microarray probeset of the same gene associated with the exome variant, we selected the probeset that provided the strongest LD r2 between the exome variant and the eSNP. In reciprocal conditional analysis, we conditioned on the lead exome variant by including it as a covariate in the cis-eQTL detection and reporting the p-value of the eSNP and vice versa. We considered the signals to be coincident if both the lead exome variant and the eSNP were no longer significant after conditioning on the other and the variants were in high pairwise LD ($r2 > 0.80$).

For loci that also harbored reported GWAS variants, we performed reciprocal conditional analysis between the GWAS lead variant and the lead eSNP. For loci with more than one reported GWAS variant, the GWAS lead variant with the strongest LD r2 with the lead eSNP was reported.

Penetrance analysis

Phenotype and genotype data from the UK Biobank (UKBB) were used for the penetrance analysis. Three of 16 rare and low frequency variants (MAF ≤ 1%) detected in the final Stage 1 plus 2 meta-analysis were available in the UKBB and had relatively larger effect sizes (>0.90). The phenotype data for these three variants were stratified with respect to waist-to-hip ratio (WHR) using the World Health Organization (WHO) guidelines. These guidelines consider women and men with WHR greater than 0.85 and 0.90 as obese, respectively. Genotype and allele counts were obtained for the available variants and these were used to calculate the number of carriers of the minor allele. The number of carriers for women, men and all combined was then compared between two strata (obese vs. non-obese) using a $\chi 2$ test. The significance threshold was determined by using a Bonferroni correction for the number of tests performed ($0.05/9=5.5x10^{-3}$)).

DATA AVAILABILITY

Summary statistics of all analyses are available at https://www.broadinstitute.org/collaboration/giant/.

1317 **BOXES**

Box 1. Genes of biological interest harboring WHR-associated variants

PLXND1- (3:129284818, rs2625973, known locus) The major allele of a common non-synonymous variant in Plexin D1 (L1412V, MAF=26.7%) is associated with increased WHRadjBMI (β (SE)= 0.0156 (0.0024), P-value=9.16E-11). *PLXND1* is a semaphorin class 3 and 4 receptor gene, and therefore, is involved in cell to cell signaling and regulation of growth in development for a number of different cell and tissue types, including those in the cardiovascular system, skeleton, kidneys, and the central nervous system[82-86]. Mutations in this gene are associated with Moebius syndrome[87-90], and persistent truncus arteriosus[84,91]. *PLXND1* is involved in angiogenesis as part of the SEMA and VEGF signalling pathways[92-95]. *PLXND1* was implicated in the development of T2D through its interaction with SEMA3E in mice. *SEMA3E* and *PLXND1* are upregulated in adipose tissue in response to diet-induced obesity, creating a cascade of adipose inflammation, insulin resistance, and diabetes mellitus[86]. *PLXND1* is highly expressed in adipose (both subcutaneous and visceral) (GTeX). *PLXND1* is highly intolerant of mutations and therefore highly conserved (**Supplementary Table 12**). Last, our lead variant is predicted as damaging or possibly damaging for all algorithms examined (SIFT, Polyphen2/HDIV, Polyphen2/HVAR, LRT, MutationTaster).

ACVR1C– (2:158412701, rs55920843, novel locus) The major allele of a low frequency non-synonymous variant in activin A receptor type 1C (rs55920843, N150H, MAF=1.1%) is associated with increased WHRadjBMI (β (SE)= 0.0652 (0.0105), P-value= 4.81E-10). *ACVR1C*, also called Activin receptor-like kinase 7 (*ALK7*), is a type I receptor for TGFB (Transforming Growth Factor, Beta-1), and is integral for the activation of SMAD transcription factors; therefore, *ACVR1C* plays an important role in cellular growth and differentiation[49-53], including adipocytes[53]. Mouse Acvr1c decreases secretion of insulin and

is involved in lipid storage[96]. Mouse Acvr1c decreases secretion of insulin and is involved in lipid storage[54,57,58]. *ACVR1C* exhibits the highest expression in adipose tissue, but is also highly expressed in the brain[54-56]. Expression is associated with body fat, carbohydrate metabolism and lipids in both obese and lean individuals[55]. *ACVR1C* is moderately tolerant of variants (EXaC Constraint Scores: synonymous= -0.86, nonsynonymous = 1.25, LoF = 0.04). Last, our lead variant is predicted as damaging for two of five algorithms examined (LRT and MutationTaster).

FGFR2– (10:123279643, rs138315382, novel locus) The minor allele of a rare synonymous variant in Fibroblast Growth Factor Receptor 2 (rs138315382, MAF=0.09%) is associated with increased WHRadjBMI (β (SE) = 0.258 (0.049), P-value= 1.38E-07). The extracellular portion of the FGFR2 protein binds with fibroblast growth factors, influencing mitogenesis and differentiation. Mutations in this gene have been associated with many rare monogenic disorders, including skeletal deformities, craniosynostosis, eye abnormalities, and LADD syndrome, as well as several cancers including breast, lung, and gastric cancer. Methylation of *FGFR2* is associated with high birth weight percentile[97]. *FGFR2* is tolerant of synonymous mutations, but highly intolerant of missense and loss-of-function mutations (ExAC Constraint scores: synonymous=-0.9, missense=2.74, LoF=1.0) (**Supplementary Table 13**). Last, this variant is not predicted to be damaging based on any of the 5 algorithms tested.

ANGPTL4 – (19:8429323, rs116843064, novel locus) The major allele of a nonsynonymous low frequency variant in Angiopoietin Like 4 (rs116843064, E40K, EAF=98.1%) is associated with increased WHRadjBMI (β (SE) = 0.064 (0.011) P-value= 1.20E-09). *ANGPTL4* encodes a glycosylated, secreted protein containing a C-terminal fibrinogen domain. The encoded protein is induced by peroxisome proliferation activators and functions as a serum hormone that regulates glucose homeostasis, triglyceride metabolism[98,99], and insulin sensitivity[100]. Angptl4-deficient mice have hypotriglyceridemia and increased lipoprotein lipase

(LPL) activity, while transgenic mice overexpressing Angplt4 in the liver have higher plasma triglyceride levels and decreased LPL activity[101]. The major allele of rs116843064 has been previously associated with increased risk of coronary heart disease and increased TG[48]. *ANGPTL4* is moderately tolerant of mutations (ExAC constraint scores synonymous=1.18, missense=0.21, LoF=0.0). Last, our lead variant is predicted damaging for four of five algorithms (SIFT, Polyphen 2/HDIV, Polyphen2/HVAR, and MutationTaster).

RREB1- (6:7211818, rs1334576, novel association signal) The major allele of a common non-synonymous variant in the Ras responsive element binding protein 1 (rs1334576, G195R, EAF=56%) is associated with increased WHRadjBMI (β (SE)=0.017 (0.002), P-value=$3.9x10^{-15}$). This variant is independent of the previously reported GWAS signal in the *RREB1* region (rs1294410; 6:6738752[10]). The protein encoded by this gene is a zinc finger transcription factor that binds to RAS-responsive elements (RREs) of gene promoters. It has been shown that the calcitonin gene promoter contains an RRE and that the encoded protein binds there and increases expression of calcitonin, which may be involved in Ras/Raf-mediated cell differentiation[102-104]. The ras responsive transcription factor *RREB1* is a candidate gene for type 2 diabetes associated end-stage kidney disease[103]. This variant is highly intolerant to loss of function (ExAC constraint score LoF = 1).

DAGLB- (7:6449496, rs2303361, novel locus) The minor allele of a common non-synonymous variant (rs2303361,Q664R, MAF=22%) in *DAGLB* (Diacylglycerol lipase beta) is associated with increased WHRadjBMI (β (SE)= 0.0136 (0.0025), P-value=6.24x10-8). *DAGLB* is a diacylglycerol (DAG) lipase that catalyzes the hydrolysis of DAG to 2-arachidonoyl-glycerol, the most abundant endocannabinoid in tissues. In the brain, DAGL activity is required for axonal growth during development and for retrograde synaptic signaling at mature synapses (2-AG)[105]. The *DAGLB* rs702485 (7:6449272, r^2= 0.306 and D'=1

with rs2303361) variant has been previously associated with high-density lipoprotein cholesterol (HDL) previously. Pathway analysis indicate a role in the triglyceride lipase activity pathway [106]. *DAGLB* is tolerant of synonymous mutations, but intolerant of missense and loss of function mutations (ExAC Constraint scores: synonymous=-0.76, missense=1.07, LoF=0.94). Last, this variant is not predicted to be damaging by any of the algorithms tested.

MLXIPL (7:73012042, rs35332062 and 7:73020337, rs3812316, known locus) The major alleles of two common non-synonymous variants (A358V, MAF=12%; Q241H, MAF=12%) in *MLXIPL* (MLX interacting protein like) are associated with increased WHRadjBMI (β (SE)= 0.02 (0.0033), P-value=1.78×10^{-9}; β (SE)= 0.0213 (0.0034), P-value=1.98×10^{-10}). These variants are in strong linkage disequilibrium (r^2=1.00, D'=1.00, 1000 Genomes CEU). This gene encodes a basic helix-loop-helix leucine zipper transcription factor of the Myc/Max/Mad superfamily. This protein forms a heterodimeric complex and binds and activates carbohydrate response element (ChoRE) motifs in the promoters of triglyceride synthesis genes in a glucose-dependent manner[59,60]. This gene is possibly involved in the growth hormone signaling pathway and lipid metabolism. The WHRadjBMI-associated variant in this gene has been associated with the levels, risk of non-alcoholic fatty liver disease and coronary artery disease . This gene possibly involved in the growth hormone signaling pathway and lipid metabolism. The WHRadjBMI-associated variant rs3812316 in this gene has been associated with the levels, risk of non-alcoholic fatty liver disease and coronary artery disease[59,107,108]. Furthermore, Williams-Beuren syndrome (an autosomal dominant disorder characterized by short stature, abnormal weight gain, various cardiovascular defects, and mental retardation) is caused by a deletion of about 26 genes from the long arm of chromosome 7 including *MLXIPL*. *MLXIPL* is generally intolerant to variation, and therefore conserved (ExAC Constraint scores: synonymous = 0.48, missense=1.16, LoF=0.68). Last, both variants reported here are predicted as possible or probably damaging by one of the algorithms tested

(PolyPhen).

RAPGEF3 (12:48143315, rs145878042, novel locus) The major allele of a low frequency non-synonymous variant in Rap Guanine-Nucleotide-Exchange Factor (GEF) 3 (rs145878042, L300P, MAF=1.1%) is associated with increased WHRadjBMI (β (SE)=0.085 (0.010), P-value = $7.15E^{-17}$). *RAPGEF3* is an intracellular cAMP sensor, also known as Epac (the Exchange Protein directly Activated by Cyclic AMP). Among its many known functions, RAPGEF3 regulates the ATP sensitivity of the KATP channel involved in insulin secretion[109], may be important in regulating adipocyte differentiation[110-112], and plays an important role in regulating adiposity and energy balance[113], and plays an important role in regulating adiposity and energy balance[113]. *RAPGEF3* is tolerant of mutations (ExAC Constraint Scores: synonymous = -0.47, nonsynonymous = 0.32, LoF = 0). Last, our lead variant is predicted as damaging or possibly damaging for all five algorithms examined herein (SIFT, Polyphen2/HDIV, Polyphen2/HVAR, LRT, MutationTaster).

TBX15 (1:119427467, rs61730011, known locus) The major allele of a low frequency non-synonymous variant in T-box 15 (rs61730011, M460R, MAF=4.3%) is associated with increased WHRadjBMI (β(SE)=0.041(0.005)). T-box 15 (*TBX15*) is a developmental transcription factor expressed in adipose tissue, but with higher expression in visceral adipose tissue than in subcutaneous adipose tissue, and strongly downregulated in overweight and obese individuals[114]. *TBX15* negatively controls depot-specific adipocyte differentiation and function[115] and regulates glycolytic myofiber identity and muscle metabolism[116]. *TBX15* is moderately intolerant of mutations and therefore conserved (ExAC Constraint Scores: synonymous = 0.42, nonsynonymous = 0.65, LoF = 0.88). Last, our lead variant is predicted as damaging or possibly damaging for four algorithms (Polyphen2/HDIV, Polyphen2/HVAR, LRT, MutationTaster).

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FIGURES

Figure 1. Summary of meta-analysis study design and workflow. Abbreviations:

EUR- European, AFR- African, SAS- South Asian, EAS- East Asian, and HIS- Hispanic/Latino ancestry.

Figure 2. Minor allele frequency compared to estimated effect. This scatter plot displays the relationship between minor allele frequency (MAF) and the estimated effect (β) for each significant coding variant in our meta-analyses. All novel WHRadjBMI variants are highlighted in orange, and variants identified only in models that assume recessive inheritance are denoted by diamonds and only in sex-specific analyses by triangles. Eighty percent power was calculated based on the total sample size in the Stage 1+2 meta-analysis and $P=2x10^{-7}$. Estimated effects are shown in original units (cm/cm) calculated by using effect sizes in standard deviation (SD) units times SD of WHR in the ARIC study (sexes combined=0.067, men=0.052, women=0.080).

Figure 3. Regional association plots for known loci with novel coding signals. Point color reflects r^2 calculated from the ARIC dataset. In a) there are two independent variants in *RSPO3* and *KIAA0408,* as shown by conditional analysis. In b) we have a variant in *RREB1* that is independent of the GWAS variant rs1294421.

Figure 4. Heat maps showing DEPICT gene set enrichment results. For any given square, the color indicates how strongly the corresponding gene (shown on the x-axis) is predicted to belong to the reconstituted gene set (y-axis). This value is based on the gene's z-score for gene set inclusion in DEPICT's reconstituted gene sets, where red indicates a higher and blue a lower z-score. To visually reduce redundancy and increase clarity, we chose one representative "meta-gene set" for each group of highly correlated gene sets based on affinity propagation clustering (Online Methods, Supplementary Information). Heatmap intensity and DEPICT P-values (see P-values in Supplementary Data 4-5) correspond to the most significantly enriched gene set within the meta-gene set. Annotations for the

genes indicate (1) the minor allele frequency of the significant ExomeChip (EC) variant (shades of blue; if multiple variants, the lowest-frequency variant was kept), (2) whether the variant's P-value reached array-wide significance (<2x10-7) or suggestive significance (<5x10-4) (shades of purple), (3) whether the variant was novel, overlapping "relaxed" GWAS signals from Shungin et al.[10] (GWAS P<5x10-4), or overlapping "stringent" GWAS signals (GWAS P<5x10-8) (shades of pink), and (4) whether the gene was included in the gene set enrichment analysis or excluded by filters (shades of brown/orange) (Online Methods and Supplementary Information). Annotations for the gene sets indicate if the meta-gene set was found significant (shades of green; FDR <0.01, <0.05, or not significant) in the DEPICT analysis of GWAS results from Shungin et al.

TABLES

Table 1. Association results for Combined Sexes. Association results based on an additive or recessive model for coding variants that met array-wide significance (P<2x10-07) in the sex-combined meta-analyses.

Locus (+/- 1Mb of given variant)	Chr:Position (GRCh37)[b]	rsID	Effect Allele	Other Allele	Gene[c]	Amino Acid Change[c]	If locus is known, nearby (< 1 MB) published variant(s) [d]	N	Effect Allele Frequency	Effect size (SD/allele)[e]	SE	P-value	P-value for Sex-heterogeneity[f]
Variants in Novel Loci													
All Ancestry Additive model Sex-combined analyses													
1	2:158412701	rs55920843	T	G	*ACVR1C*	N150H	-	455,526	0.989	0.065	0.011	***4.8E-10***	**1.7E-07**
2	3:50597092	rs1034405	G	A	*C3orf18*	A162V	-	455,424	0.135	0.016	0.003	***1.9E-07***	8.8E-01
3	4:120528327	rs3733526	G	A	*PDE5A*	A41V	-	461,521	0.187	0.015	0.003	***2.6E-08***	5.2E-03
4	6:26108117	rs146860658	T	C	*HIST1H1T*	A69T	-	217,995	0.001	0.229	0.042	***4.3E-08***	6.3E-01
5	7:6449496	rs2303361	C	T	*DAGLB*	Q664R	-	475,748	0.221	0.014	0.003	***6.2E-08***	3.4E-03
6	10:123279643	rs138315382	T	C	*FGFR2*	synonymous	-	236,962	0.001	0.258	0.049	***1.4E-07***	1.1E-01
7	11:65403651	rs7114037	C	A	*PCNXL3*	H1822Q	-	448,861	0.954	0.029	0.005	***1.8E-08***	4.4E-01
8	12:48143315	rs145878042	A	G	*RAPGEF3*	L300P	-	470,513	0.990	0.085	0.010	***7.2E-17***	7.3E-03
9	12:108618630	rs3764002	C	T	*WSCD2*	T266I	-	474,637	0.737	0.014	0.002	***9.8E-10***	5.5E-01
10	15:42032383	rs17677991	G	C	*MGA*	P1523A	-	469,874	0.345	0.015	0.002	***3.5E-11***	9.1E-01
	16:4432029	rs3810818	A	C	*VASN*	E384A	-	424,163	0.231	0.016	0.003	***2.0E-09***	3.3E-01
11	16:4445327	rs3747579	C	T	*CORO7*	R193Q	-	453,078	0.299	0.018	0.002	***2.2E-13***	4.3E-02
	16:4484396	rs1139653	A	T	*DNAJA3*	N75Y	-	434,331	0.284	0.015	0.002	***4.3E-10***	1.4E-01

12	19:49232226	rs2287922	A	G	*RASIP1*	R601C	-	430,272	0.494	0.014	0.002	***1.6E-09***	3.7E-02
	19:49244220	rs2307019	G	A	*IZUMO1*	A333V	-	476,147	0.558	0.012	0.002	***4.7E-08***	3.9E-02
13	20:42965811	rs144098855	T	C	*R3HDML*	P5L	-	428,768	0.001	0.172	0.032	***9.7E-08***	1.0E+00
European Ancestry Additive model Sex-combined analyses													
14	1:173802608	rs35515638	G	A	*DARS2*	K196R	-	352,646	0.001	0.201	0.038	***1.4E-07***	6.0E-02
15	14:58838668	rs1051860	A	G	*ARID4A*	synonymous	-	367,079	0.411	0.013	0.002	***2.2E-08***	1.3E-01
16	15:42115747	rs3959569	C	G	*MAPKBP1*	R1240H	-	253,703	0.349	0.017	0.003	***2.0E-08***	6.3E-01

Variants in Previously Identified Loci

All Ancestry Additive model Sex-combined analyses													
1	1:119427467	rs61730011	A	C	*TBX15*	M566R	rs2645294, rs12731372, rs12143789, rs1106529	441,461	0.957	0.041	0.005	***2.2E-14***	6.7E-01
	1:119469188	rs10494217	T	G		H156N		472,259	0.174	0.018	0.003	***1.4E-10***	6.0E-01
2	1:154987704	rs141845046	C	T	*ZBTB7B*	P190S	rs905938	476,440	0.976	0.037	0.007	***3.8E-08***	**7.9E-07**
3	2:165551201	rs7607980	T	C	*COBLL1*	N941D	rs1128249, rs10195252, rs12692737, rs12692738, rs17185198	389,883	0.879	0.026	0.004	***1.6E-13***	**3.0E-30**
4	2:188343497	rs7586970	T	C	*TFPI*	N221S	rs1569135	452,638	0.697	0.016	0.002	***3.0E-12***	6.3E-01
5	3:52558008	rs13303	T	C	*STAB1*	M113T	rs2276824	470,111	0.445	0.019	0.002	***5.5E-18***	6.7E-02
	3:52833805	rs3617	C	A	*ITIH3*	Q315K		452,150	0.541	0.015	0.002	***1.6E-12***	4.0E-01
6	3:129137188	rs62266958	C	T	*EFCAB12*	R197H	rs10804591	476,382	0.936	0.036	0.004	***8.3E-17***	**9.3E-05**
	3:129284818	rs2625973	A	C	*PLXND1*	L1412V		476,338	0.733	0.016	0.002	***9.2E-11***	**1.6E-05**
7	4:89625427	rs1804080	G	C	*HERC3*	E946Q	rs9991328	446,080	0.838	0.021	0.003	***1.5E-12***	**4.1E-06**
	4:89668859	rs7657817	C	T	*FAM13A*	V443I		476,383	0.815	0.016	0.003	***5.0E-09***	**9.6E-05**
8	5:176516631	rs1966265	A	G	*FGFR4*	V10I	rs6556301	455,246	0.236	0.023	0.003	***1.7E-19***	2.1E-01
9	6:7211818	**rs1334576**[g]	G	A	*RREB1*	G195R	rs1294410	451,044	0.565	0.017	0.002	***3.9E-15***	1.5E-01
10	6:34827085	rs9469913	A	T	*UHRF1BP1*	Q984H	rs1776897	309,684	0.847	0.021	0.004	***1.2E-08***	2.7E-01

11	6:127476516	rs1892172	A	G	*RSPO3*	synonymous	rs11961815, rs72959041, rs1936805	476,358	0.543	0.031	0.002	***2.6E-47***	**7.7E-09**
	6:127767954	**rs139745911**[g]	A	G	*KIAA0408*	P504S		391,469	0.010	0.103	0.012	***6.8E-19***	**2.0E-04**
12	7:73012042	rs35332062	G	A	*MLXIPL*	A358V	rs6976930	451,158	0.880	0.020	0.003	***1.8E-09***	1.5E-01
	7:73020337	rs3812316	C	G		Q241H		454,738	0.881	0.021	0.003	***2.0E-10***	5.8E-02
13	10:95931087	rs17417407	T	G	*PLCE1*	R240L	rs10786152	476,475	0.173	0.018	0.003	***2.5E-11***	5.9E-01
14	11:64031241	rs35169799	T	C	*PLCB3*	S778L	rs11231693	476,457	0.061	0.034	0.004	***9.1E-15***	**1.3E-04**
15	12:123444507	rs58843120	G	T	*ABDB9*	F92L	rs4765219, rs863750	466,498	0.987	0.053	0.009	***1.3E-08***	3.5E-01
	12:124265687	rs11057353	T	C	*DNAH10*	S228P		476,360	0.373	0.018	0.002	***2.1E-16***	**2.7E-08**
	12:124330311	rs34934281	C	T		T1785M		476,395	0.889	0.025	0.003	***2.9E-14***	**3.1E-08**
	12:124427306	rs11057401	T	A	*CCDC92*	S53C		467,649	0.695	0.029	0.002	***7.3E-37***	**5.5E-11**
16	15:56756285	rs1715919	G	T	*MNS1*	Q55P	rs8030605	476,274	0.096	0.023	0.004	***8.8E-11***	2.7E-02
17	16:67397580	rs9922085	G	C	*LRRC36*	R101P	rs6499129	469,474	0.938	0.034	0.005	***3.8E-13***	5.9E-01
	16:67409180	rs8052655	G	A		G388S		474,035	0.939	0.034	0.005	***5.5E-13***	4.0E-01
18	19:18285944	rs11554159	A	G	*IFI30*	R76Q	rs12608504	476,389	0.257	0.015	0.002	***3.5E-10***	3.1E-03
	19:18304700	rs874628	G	A	*MPV17L2*	M72V		476,388	0.271	0.015	0.002	***1.2E-10***	2.5E-03
19	20:33971914	rs4911494	T	C	*UQCC1*	R51Q	rs224333	451,064	0.602	0.018	0.002	***2.5E-16***	1.5E-03
	20:34022387	rs224331	A	C	*GDF5*	S276A		345,805	0.644	0.017	0.003	***1.8E-11***	3.2E-03
All Ancestry Recessive model Sex-combined analyses													
20	17:17425631	rs897453	C	T	*PEMT*	V58L	rs4646404	476,546	0.569	0.025	0.004	***4.1E-11***	8.2E-01
European Ancestry Additive model Sex-combined analyses													
6	3:129293256	rs2255703	T	C	*PLXND1*	M870V	rs10804591	420,520	0.620	0.014	0.002	***3.1E-09***	**1.6E-04**

Abbreviations: GRCh37=human genome assembly build 37;rsID=based on dbSNP; VEP=Ensembl Variant Effect Predictor toolset; GTEx=Genotype-Tissue Expression project;SD=standard deviation; SE=standard error;N=sample size

a Coding variants refer to variants located in the exons and splicing junction regions.

b Variant positions are reported according to Human assembly build 37 and their alleles are coded based on the positive strand.

c The gene the variant falls in and amino acid change from the most abundant coding transcript is shown (protein annotation is based on VEP toolset and transcript abundance from GTEx database).

d Previously published variants within +/-1Mb are from Shungin et al.[10], except for rs6976930 and rs10786152 from Graff et al.[14] and rs6499129 from Ng. et al [16].

e Effect size is based on standard deviation (SD) per effect allele

f P-value for sex heterogeneity, testing for difference between women-specific and men-specific beta estimates and standard errors, was calculated using EasyStrata: Winkler, T.W. et al. EasyStrata: evaluation and visualization of stratified genome-wide association meta-analysis data. Bioinformatics 2015: 31, 259-61.PMID: 25260699. Bolded P-values met significance threshold after bonferonni correction (P-value<7.14E-04; i.e. 0.05/70 variants).

g **rs1334576 in *RREB1*** is a new signal in a known locus that is independent from the known signal, rs1294410; **rs139745911 in *KIAA0408*** is a new signal in a known locus that is independent from all known signals rs11961815, rs72959041, rs1936805, in a known locus (see Supplementary 8A/B).

Table 2. Association results for Sex-stratified analyses. Association results based on an additive or recessive model for coding variants that met array-wide significance (P<2x10-07) in the sex-specific meta-analyses and reach bonferonni corrected P-value for sex hetergeneity (P_{sexhet}<7.14E-04).

Locus (+/- 1Mb of a given variant)	Chr:Position (GRCh37)[c]	rsID	Effect Allele	Other Allele	Gene[d]	Amino Acid Change[d]	Identified in sex-combined analyses[e]	If locus is known, nearby (< 1 MB) published variant(s) [f]	P_{sexhet}	Men					Women				
										N	EAF	Effect[h] (SD/ allele)	SE	P	N	EAF	Effect[h] (SD/ allele)	SE	P
Variants in Novel Loci																			
All Ancestry Additive model Men only analyses																			
1	13:96665697	rs148108950	A	G	*UGGT2*	P175L	No	-	**1.5E-06**	203,009	0.006	0.130	0.024	***6.1E-08***	221,390	0.004	-0.044	0.027	1.1E-01
2	14:23312594	rs1042704	A	G	*MMP14*	D273N	No	-	**2.6E-04**	226,646	0.202	0.021	0.004	***2.6E-08***	250,018	0.197	0.002	0.004	6.1E-01
All Ancestry Additive model Women only analyses																			
3	1:205130413	rs3851294	G	A	*DSTYK*	C641R	No	-	**9.8E-08**	225,803	0.914	-0.005	0.005	3.4E-01	249,471	0.912	0.034	0.005	***4.5E-11***
4	2:158412701	rs55920843	T	G	*ACVR1C*	N150H	Yes	-	**1.7E-07**	210,071	0.989	0.006	0.015	7.2E-01	245,808	0.989	0.113	0.014	***1.7E-15***
5	19:8429323	rs116843064	G	A	*ANGPTL4*	E40K	No	-	**1.3E-07**	203,098	0.981	-0.017	0.011	1.4E-01	243,351	0.981	0.064	0.011	***1.2E-09***
Variants in Previously Identified Loci																			
All Ancestry Additive model Women only analyses																			
1	1:154987704	rs141845046	C	T	*ZBTB7B*	P190S	Yes	rs905938	**7.9E-07**	226,709	0.975	0.004	0.010	6.9E-01	250,084	0.977	0.070	0.010	***2.3E-13***
2	2:165551201	rs7607980	T	C	*COBLL1*	N941D	Yes	rs1128249, rs10195252, rs12692737, rs12692738, rs17185198	**3.0E-30**	173,600	0.880	-0.018	0.005	5.8E-04	216,636	0.878	0.062	0.005	***6.7E-39***
3	3:129137188	rs62266958	C	T	*EFCAB12*	R197H	Yes	rs10804591	**9.3E-05**	226,690	0.937	0.018	0.006	3.1E-03	250,045	0.936	0.051	0.006	***8.1E-18***
	3:129284818	rs2625973	A	C	*PLXND1*	L1412V	Yes		**1.6E-05**	226,650	0.736	0.005	0.003	1.9E-01	250,023	0.730	0.025	0.003	***8.2E-14***
	3:129293256	rs2255703	T	C		M870V	Yes		**5.0E-04**	226,681	0.609	0.003	0.003	3.1E-01	250,069	0.602	0.018	0.003	***1.9E-09***
4	4:89625427	rs1804080	G	C	*HERC3*	E946Q	Yes	rs9991328	**4.1E-06**	222,556	0.839	0.008	0.004	6.6E-02	223,877	0.837	0.034	0.004	***2.1E-16***
	4:89668859	rs7657817	C	T	*FAM13A*	V443I	Yes		**9.6E-05**	226,680	0.816	0.006	0.004	1.5E-01	242,970	0.815	0.026	0.004	***5.9E-12***

5	6:127476516	rs1892172	A	G	*RSPO3*	synonymous	Yes	rs11961815, rs72959041, rs1936805	**7.7E-09**	226,677	0.541	0.018	0.003	***5.6E-10***	250,034	0.545	0.042	0.003	***3.4E-48***
	6:127767954	**rs139745911**[i]	A	G	*KIAA0408*	P504S	Yes		**2.0E-04**	188,079	0.010	0.057	0.017	6.8E-04	205,203	0.010	0.143	0.016	***5.9E-19***
6	11:64031241	rs35169799	T	C	*PLCB3*	S778L	Yes	rs11231693	**1.3E-04**	226,713	0.061	0.016	0.006	9.6E-03	250,097	0.061	0.049	0.006	***6.7E-16***
7	12:124265687	rs11057353	T	C	*DNAH10*	S228P	Yes	rs4765219, rs863750	**2.7E-08**	226,659	0.370	0.005	0.003	8.3E-02	250,054	0.376	0.029	0.003	***3.1E-22***
	12:124330311	rs34934281	C	T		T1785M	Yes		**3.1E-08**	226,682	0.891	0.006	0.005	1.9E-01	250,066	0.887	0.043	0.005	***1.4E-20***
	12:124427306	rs11057401	T	A	*CCDC92*	S53C	Yes		**5.5E-11**	223,324	0.701	0.013	0.003	4.3E-05	244,678	0.689	0.043	0.003	***1.0E-41***

Abbreviations: GRCh37=human genome assembly build 37;rsID=based on dbSNP; VEP=Ensembl Variant Effect Predictor toolset; GTEx=Genotype-Tissue Expression project;SD=standard deviation; SE=standard error;N=sample size

a Coding variants refer to variants located in the exons and splicing junction regions.

b Bonferonni corrected Pvalue for the number of SNPs tested for sex-heterogeneity is <7.14E-04 i.e. 0.05/70 variants.

c Variant positions are reported according to Human assembly build 37 and their alleles are coded based on the positive strand.

d The gene the variant falls in and amino acid change from the most abundant coding transcript is shown (protein annotation is based on VEP toolset and transcript abundance from GTEx database).

e Variant was also identified as array-wide significant in the sex-combined analyses.

f Previously published variants within +/-1Mb are from Shungin D et al. New genetic loci link adipose and insulin biology to body fat distribution. Nature 2015; 518, 187–196 doi:10.1038/nature14132 (PMID 25673412).

g P-value for sex heterogeneity, testing for difference between women-specific and men-specific beta estimates and standard errors, was calculated using EasyStrata: Winkler, T.W. et al. EasyStrata: evaluation and visualization of stratified genome-wide as Bioinformatics 2015: 31, 259-61. PMID: 25260699.

h Effect size is based on standard deviation (SD) per effect allele

i **rs139745911 in KIAA0408** is a new signal in a known locus that is independent from all known signals rs11961815, rs72959041, rs1936805, in a known locus (see Supplementary 8A/B).

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