been published in final form at http://dx.doi.org/10.1002/slct.201600245. This article may be used for noncommercial purposes in accordance With Wiley-VCH Terms and Conditions for self-archiving

Thiacalix[4]arene Derivatives Bearing Imidazole Units: A Ditopic Hard/Soft Receptor for Na⁺ and K⁺/Ag⁺ with an Allosteric Effect and a Reusable Extractant for Dichromate Anions

Jiang-Lin Zhao,^[a] Chong Wu,^[a] Xi Zeng,^[b] Shofiur Rahman,^[c] Paris E. Georghiou,^[c] Mark R.J. Elsegood,^[d] Thomas G. Warwick,^[d] Carl Redshaw,^[e] Simon J. Teat,^[f] and Takehiko Yamato*^[a]

Abstract: Two novel receptors 5,11,17,23-tetra-tert-butyl-25,27bis[(ethoxycarbonyl)methoxy]-26,28-bis-[1-methyl-(imidazole)methoxy]-2,8,14,20-tetra-thiacalix[4]arene (L1) and 5,11,17,23-tetra-tertbutyl-25,27-bis-[(benzyl)methoxy]-26,28-bis-[1-methyl-(imidazole)methoxy]-2,8,14,20-tetrathiacalix[4]arene (L2) possessing imidazole moieties based on thiacalix[4]arene in the 1,3-alternate conformation have been synthesized and characterized. The crystal structures of L1 and L2 have been determined. The binding behaviour towards Li⁺, Na⁺, K⁺ and Ag⁺ ions has been examined by ¹H NMR titration experiments in (CDCl₃/CD₃CN; 10:1, v/v) solution. The exclusive formation of mononuclear complexes of L1 with metal cations is of particular interest revealing a negative allosteric effect in the thiacalix[4]arene family. Liquid-liquid extraction experiments indicate that synthesized L2 can be utilized as an efficient reusable extractant for dichromate anion by controlling the pH of the aqueous solution.

Introduction

Allosteric regulation between subunits within a receptor system is one of nature's elegant strategies for precisely regulating and controlling the diverse functions in biological systems.¹ The use of allosteric regulation has also been utilized by chemists to control molecular function by external stimuli to transduce chemical signals, and to achieve chemical feedback regulation.² Inspired by this, much effort has been devoted in supramolecular chemistry

[a]	Dr. JL. Zhao, Dr. C. Wu, Dr. H. Tomiyasu, Dr. Prof. T. Yamato Department of Applied Chemistry, Faculty of Science and		
	Engineering		
	Saga University		
	Honjo-machi 1, Saga 840-8502 Japan		
	E-mail: yamatot@cc.saga-u.ac.jp		
[b]	Prof. X. Zeng		
	Department Key Laboratory of Macrocyclic and Supramolecular		
	Chemistry of Guizhou Province		
	Guizhou University		
	Guiyang, Guizhou, 550025, China.		
[c]	Dr. S. Rahman, Prof. P. E. Georghiou		
	Department of Chemistry,		
	Memorial University of Newfoundland,		
	St. John's, Newfoundland and Labrador, Canada A1B3X7.		
[d]	Dr. M. R. J. Elsegood, Dr. T. G. Warwick		
	Chemistry Department		
	Loughborough University		
	Loughborough, Leicestershire LR11 3TU, UK		
[e]	Dr. C. Redshaw		
	Department of Chemistry		
	The University of Hull		
	Cottingham Road, Hull, Yorkshire HU6 7RX, UK		
[f]	Dr. S. J. Teat		
-	ALS, Berkeley Lab,		
	1 Cyclotron Road, Berkeley, CA 94720, USA		

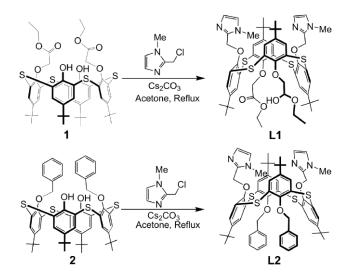
to construct artificial systems to achieve analogous

functions such as molecular recognition and signal amplification.

Di- or poly-topic receptors have been constructed with two or more binding subunits within the same macrocyclic structure.³ It is well known that such systems are suitable candidates for the allosteric regulation of host-guest interactions with metal cations.^{1d,4} Thiacalix[4]arenes are macrocyclic molecules which can be relatively easily functionalized and have pre-organized binding sites. In particular, the 1,3-alternate conformation is capable of possessing binding subunits in the two opposite domains of the molecule, and is thus a potentially ideal structure for allosteric regulation.⁵ Consequently, we have been interested in being able to incorporate two subunits having different binding properties onto a thiacalix[4]arene and to control or maintain a 1,3-alternate conformation. This would therefore permit the construction of a di-topic receptor system which should possess allosteric features.

The development of systems for the fast estimation, removal and separation of silver (soft metal), together with the use of Ag⁺ complexes in photographic materials and their potential use in cancer radioimmunotherapy has attracted the attention of supramolecular chemists towards designing effective Ag⁺-selective receptors.⁶ Recently, we undertook the synthesis and evaluation of thiacalix[4]arene and homotrioxacalix[3]arene ligands as Ag+ ionophores and found that the introduction of imidazole groups at the lower rim of the calixarene skeleton resulted in a high affinity for Aq⁺.⁷ On the other hand, it has been reported that by introducing an ester group at its lower or upper rim, the thiacalix[4]arene derivative can selectively complex alkali metal cations (hard metals), such as Na⁺ and K⁺.⁸ Thus, the introduction of imidazole groups onto the thiacalix[4]arene framework as one binding subunit and the introduction of ester groups on the opposite side as the other binding subunit, was envisioned. Such a system could be used as a ditopic receptor with two binding subunits pre-organised for both hard (Na⁺ and K⁺) and soft (Ag⁺) cations, and with possible switching of complexation preferences.

Herein, the synthesis, X-ray structure, and complexation studies of a novel ditopic receptor L1 which possesses two binding subunits (imidazole and ester moieties) based on a thiacalix[4]arene in the 1,3-*alternate* conformation is reported. In order to further investigate the allosteric effects, the mono-topic receptor L2, with a similar structure, was also produced. Additionally, due to the amphoteric nature of the imidazole group can function not only as an effective cation but also as an anion receptor under appropriate conditions. Also, as a continuation of our search for



Scheme 1. The synthetic routes to receptors L1 and L2.

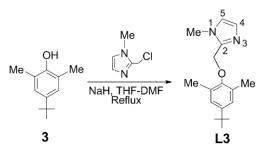
dichromate anion extractants based on thiacalixarene derivatives,^{7,13} the extraction efficiency of the systems described herein towards dichromate anion was conducted by liquid-liquid extraction experiments.

Results and Discussion

Synthesis

The preferential formation of the 1,3-*alternate* conformer of thiacalix[4]arene occurs in the presence of a cesium cation whose size is compatable with that of the thiacalix[4]arene cavity and thus contributes significantly to the cation- π interaction.⁵ Receptor **L1** could therefore be obtained in 55% yield by the stereoselective O-alkylation of **1**¹⁰ with 2-chloromethyl-1-methyl-1*H*-imidazole⁹ in the presence of Cs₂CO₃ in dry acetone (Scheme 1). Similarly, receptor **L2** was obtained in 44% yield by the stereoselective O-alkylation of **2**¹¹ with 2-chloromethyl-1-methyl-1-methyl-1*H*-imidazole in the presence of Cs₂CO₃ in dry acetone (Scheme 1).

The ¹H NMR spectrum of L1 exhibits two singlets for the *tert*butyl protons at up-field positions, *viz* δ 1.10 and 1.26 ppm; two singlets for the aromatic protons at δ 7.24 ppm and 7.46 ppm (Figure S1), all of which is indicative of a *C*₂-symmetric structure for the 1,3-*alternate* conformation. Obviously, receptor L2 is also in the 1,3-*alternate* conformation revealing two singlets for the *tert*-butyl protons (δ 0.82 and 1.15 ppm) and two singlets for the aromatic protons (δ 7.12 and 7.27 ppm, Figure S4). Surprisingly, remarkable shielding effects are experienced by the N-*CH*₃ protons of L1 and L2 (Table 1), compared with the



Scheme 2. The synthetic route of reference compound L3.

Table 1. Partial chemical shifts of L1, L2 and reference compound L3.ª

Compound	Chemical shifts, δ (ppm)		
Compound	-N-CH ₃	H_4	H ₅
L1	2.78	6.65	6.96
L3	3.70	6.82	6.94
L2	2.57	6.68	6.99
$\Delta \delta \mathbf{L1}^{b}$	-0.92	-0.17	+0.02
$\Delta \delta_{L2}{}^{b}$	-1.13	-0.14	+0.05

^a $\Delta\delta$ value is the difference of the chemical shift between L1, L2 and reference compound L3 in CDCl₃ at 27 °C. ^b A plus sign (+) denotes a shift to lower magnetic field, whereas, a negative sign (–) denotes a shift to higher magnetic field.

reference compound L3,⁷ which is prepared by O-alkylation of 4*tert*-butyl-2,6-dimethylphenol 3^{12} with 2-chloromethyl-1-methyl-1*H*-imidazole in the presence of NaH (Scheme 2). It strongly suggests that the heteroaromatic protons of the imidazole groups for both of L1 and L2 are exposed to the ring current shielding effect area operating between two of the thiacalixarene benzene rings.^{7,13}

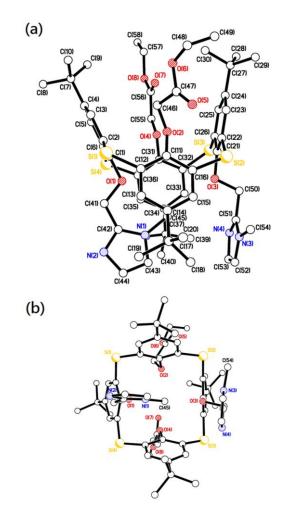


Figure 1. Single-crystal structure of L1 showing (a) the side view (b) the top view. Hydrogen atoms, ethanol and water molecules as solvent of crystallization, and minor disorder components have been omitted for clarity.

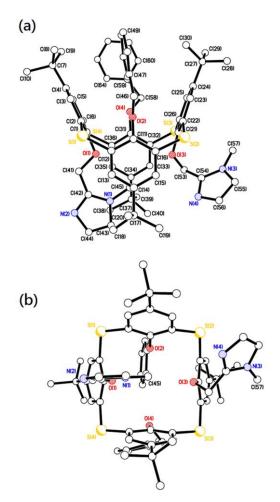


Figure 2. Single-crystal structure of L2 showing (a) the side view (b) the top view. Hydrogen atoms, ethanol and water molecules as solvent of crystallization, and minor disorder components have been omitted for clarity.

Furthermore, X-ray crystallographic analysis further confirmed the molecular structure of L1 and L2 as shown in Figure 1. Both L1 (Fig. 1a) and L2 (Fig. 2a) are in the 1,3alternate conformation. As expected, the imidazole ring containing N(1) and C(45) of L1 and L2 are pointing into the thiacalix[4]arene cavity which is consistent with the significant up-field shift for the N-*CH*₃ group in the ¹H NMR spectra. A slightly distorted square is observed for receptor L1 (Fig. 1b, S1...S2 = 5.636 Å, S2...S3 = 5.562 Å, S3...S4 = 5.546 Å and S4...S1 = 5.579 Å); however, an approximate square is observed for receptor L2 (Fig. 2b, S1...S2 = 5.558 Å). In other words, the cavity size of receptor L1 is a little larger than receptor L2 based on the sum of the S...S distances.

Complexation studies

At the onset of the study, it was anticipated that multi-recognition of both hard metals (Na⁺, K⁺) and soft metals (Ag⁺) by the ditopic receptor **L1** could be detected. This of course, being due to the presence of the two ester moieties⁸ at one face of the thiacalix[4]arene cavity and the two imidazole moieties⁷ at the opposite face. The binding affinities of **L1** and **L2** toward metal cations were therefore evaluated by ¹H NMR titration experiments (CDCl₃:CD₃CN = 10:1, v/v).

¹H NMR titration studies

Both L1 and L2 possess imidazole moieties which would preferentially exhibit higher affinity towards Ag⁺ ion. Hence, we conducted ¹H NMR titration experiments with CF₃SO₃Aq. Titration of a solution of L1 with Ag⁺ ions resulted in dramatically downfield shifts ($\Delta\delta$ = +0.86 ppm) for the imidazole-N-CH₃ protons at δ 3.68 ppm after complexation with 1.0 equiv. Ag⁺ (Figure 3). It was noteworthy that δ 3.68 ppm was almost the same chemical shift for the imidazole-N-CH₃ protons (δ 3.70 ppm, Table 1) of the reference compound L3. It strongly suggested that the imidazole-N-CH₃ group of L1 escaped from the shielding area to the deshielding area upon Ag⁺ capture by receptor L1. Subsequently, the adjacent imidazolyl- proton H_4 was also affected by the change in the position of the N-CH_3 group, and exhibited a downfield shift ($\Delta\delta$ = +0.46 ppm) at δ 7.14 ppm. In contrast, another significantly up-field shift ($\Delta\delta = -0.15$ ppm) for the methylene protons of OCH₂Imid at δ 5.04 ppm was also observed. Thus chemical shift change may be attributed to the OCH₂Imid methylene protons being folded into the shielding area of the thiacalix[4]arene-cavity in order to form an efficient complex with Ag⁺. All of the evidence strongly suggested that Ag⁺ was complexed by the nitrogen atoms of the imidazole moieties via N···Ag⁺ interactions.

Similar phenomena were observed for the titration experiment of L2 with Ag+ (Figure S7). The proton resonances of N-CH₃ and H_4 in the imidazole rings were gradually shifted to downfield upon addition of Ag+ ions which could be attributed to the conformational change of receptor L2 with a concomitant deshielding effect. The methylene protons of OCH2lmid were slightly shifted up-field after being complexed with one equiv. Ag+, which may also be attributed to the OCH₂Imid methylene protons being folded into the shielding field of the thiacalix[4]arene-cavity. Both L1 and L2 exhibited slight chemical shift changes of the aromatic protons which are also attributed to the conformational changes upon complexation. On increasing the titration amount of Ag⁺ to 2.0 equiv., no further significant changes were observed for L1 and L2. According to the corresponding titration curve, the association constants were calculated to be 44 M⁻¹ and 83 M⁻¹, respectively (Figure S16 and S19). All of the observed phenomenon suggested that receptor L1 and L2 possess higher affinity towards Ag⁺ ion via N···Ag⁺ interactions.

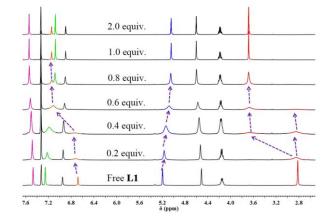


Figure 3. Partial ¹HNMR spectra of L1 (5.0mM) and increasing concentrations of Ag^+ in CDCl₃/CD₃CN(10:1, v/v) at 298K.

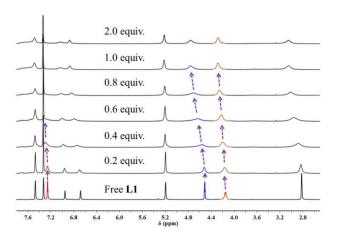


Figure 4. Partial ¹HNMR spectra of L1 (5.0mM) and increasing concentrations of K⁺ in CDCl₃/CD₃CN(10:1, v/v) at 298K.

The affinity towards alkali metal cations was also investigated, given that K⁺ and Na⁺ ions are known to interact with the ester moiety.⁸ ¹H NMR titration experiments with CF₃SO₃K, CF₃SO₃Na and CF₃SO₃Li were carried out. The addition of an equiv. of CF₃SO₃K to L1 caused immediate complexation as demonstrated by the downfield shifts of the ester ethyl group resonances (OCH₂CO, $\Delta\delta$ = +0.25; COOCH₂, $\Delta\delta$ = +0.12 ppm, Figure 4) for the 1:1 complex of L1 with an association constant of 253 M⁻¹ (Figure S17). The slightly downfield shifts of the thiacalixarene benzene protons may be attributed to the decreased shielding effect operating in two of the thiacalixarene benzene rings upon complexation of the ester moieties with K⁺. The signals of the imidazole group's protons were broadened, showing slow ligand exchange on the NMR time scale. A similar phenomenon was observed in the presence of an equiv. of CF₃SO₃Na (OCH₂CO, $\Delta\delta$ = +0.15; COOCH₂, $\Delta\delta$ = +0.07 ppm, Figure S8) which formed a 1:1 complex of L1 with an association constant of 92 M⁻¹ (Figure S18). CF₃SO₃Li did not cause detectable spectral changes for L1 due to the small atom radius which could not form an efficient complex (Figure S9). According to the corresponding association constants for these processes, it is clear that the association constants for alkali metal cations directly follow the radius of the alkali metal cations: $K^+ > Na^+ >$ Li⁺ which reflects the size recognition ability of receptor L1.

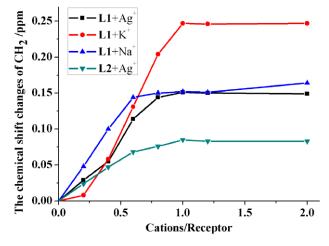


Figure 5. ¹H NMR shift of CH₂ (blue peak) in L1 with Ag⁺, K⁺, Na⁺ and L2 with Ag⁺. [In the presence of 0.2, 0.4, 0.6, 0.8, 1.0, 1.2, 2.0 equiv. of cations in CDCl₃/CD₃CN(10:1, v/v), respectively.]

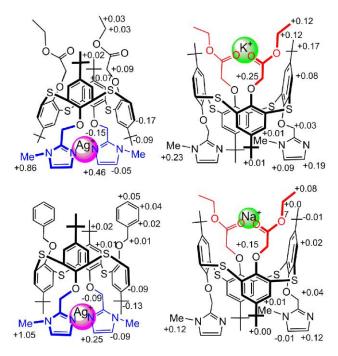


Figure 6. Chemical shift changes of L1 and L2 in the presence of different metal ions; (+) denotes the downfield shift and (-) denotes the up field shift.

Given the fact that receptor **L2** is unable to bind alkali metal cations due to one side having phenyl moieties present, which are inactive for recognition of alkali metal cations, as expected, no significant changes were observed during the ¹H NMR titration experiments with alkali metal cations. The change of the chemical shift of the *CH*₂ groups of receptor **L1** and **L2** plotted against the amount of cations (Ag⁺, K⁺ and Na⁺) is shown in Figure 5. The $\Delta\delta$ value increased in proportion to the amount of cations and became almost constant after the addition of 1.0 equiv. of cations, which indicated the 1:1 stoichiometry of the receptors–cations complex that was formed.

MALDI-TOF-MS data also supported the formation of a stable 1:1 host-guest complex. As shown in Figure S10 ~ S12, the mass spectrum of L1 with cations exhibited peaks at 1187.123, 1119.301 and 1103.285 corresponding to masses of [L1 + Ag]⁺, [L1 + K]⁺ and [L1 + Na]⁺, respectively. And the mass spectrum of L2 with cation exhibited peaks at 1195.107 corresponding to masses of [L2 + Ag]⁺ (Figure S13). In other words, the 1:1 stoichiometry complex between receptors and guest cations were unambiguously confirmed by Mass. Consequently, the proposed binding model and chemical shift changes ($\Delta\delta$) of L1 and L2 with metal ions are summarized in Figure 6.

Allosteric studies

The presence of an allosteric effect with **L1** was studied by ¹H NMR titration. Upon titration with one equiv. of Ag⁺ to the 1:1 M⁺ \subset **L1** (M⁺ = Li⁺, Na⁺ or K⁺) complex solution, the corresponding peaks were dramatically shifted to the same chemical shift positions as were noted for the 1:1 Ag⁺ \subset **L1** complex. This therefore implied that Ag⁺ was bound to the imidazole site and induced the decomplexation of hard metal cations (Li⁺, Na⁺ or K⁺) from the ester site (Figure 7). Furthermore, the 1:1 Ag⁺ \subset **L1** complex was also titrated with an equiv. of M⁺ ions (M⁺ = Li⁺, Na⁺

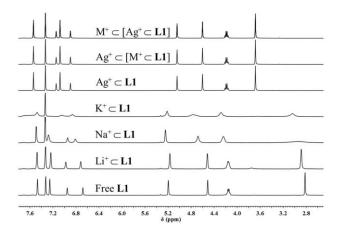


Figure 7. Partial ¹H NMR spectra of receptor L1 (5.0 mM) in CDCl₃/CD₃CN (10:1, v/v) complex with an equiv. of various metal ions; M^+ denoted the alkali metal cations ($M^+ = Li^+$, Na⁺ or K⁺).

or K⁺). None of the spectral patterns of Li⁺ \subset [Ag⁺ \subset L1], Na⁺ \subset [Ag⁺ \subset L1] and K⁺ \subset [Ag⁺ \subset L1] complexes showed any detectable signal changes. These results indicated that the complexation of Ag⁺ completely suppressed the recognition of Li⁺, Na⁺ or K⁺ by the ester moiety. The concept of a negative allosteric effect by receptor L1 is shown in Figure 8.

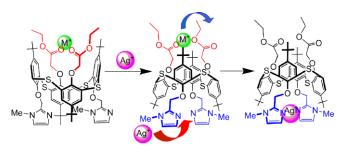


Figure 8. Proposed negative allosteric effect of ditopic recptor L1. M^+ denotes the alkali metal cations (M^+ = Li⁺, Na⁺ or K⁺).

Two-phase solvent extraction

Chromium and its compounds are widely used in our daily life, such as., tanning, plating, leather dyes, and in the photographic industry, all of which produce large quantities of toxic pollutants.¹⁴ However, high concentrations of hexavalent chromium (VI) ion is toxic to the human body, as it can diffuse as $Cr_2O_7^{2-}$ or $HCr_2O_7^{-}$ through cell membranes and oxidize biological molecules.15 Thus, selective treatment of pollutional water containing Cr (VI) prior to discharge is essential. The dichromate $(Cr_2O_4^{2-} \text{ and } HCr_2O_7^{-})$ ions are anions with oxide functionalities at their periphery. These oxide moieties are potential sites for hydrogen bonding to the complexant or host molecule(s). Imidazole group, among such heterocyclic units, the imidazole ring behaves as an excellent hydrogen bond donor moiety in synthetic anion receptor systems, and the acidity of the NH proton of the imidazole can be tuned by changing the electronic properties of the imidazole substituents.¹⁶ Thus, the introduction of a imidazole moiety to thiacalix[4]arene would potentially lead to an effective extractant for dichromate anions.

To investigate further the applicability of the receptors **L1** and **L2** which possess imidazole groups, liquid-liquid extraction experiments were performed to examine the extraction ability of **L1** and **L2** toward dichromate anion from the aqueous phase

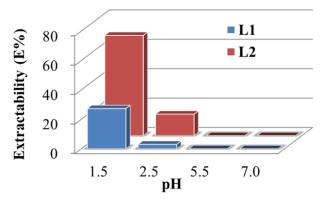


Figure 9. E% values of dichromate anion with ionophores L1 and L2 (2.0 × 10^{-4} M, 2 h at 25 °C) at pH 1.5–7.0 (H₂O/CH₂Cl₂:10/10 (v/v); K₂Cr₂O₇ = 1 × 10^{-4} M).

into the organic phase (dichloromethane) over the range of pH 1.5–7.0. It has been clearly demonstrated that the lower the pH, the higher the extractability is found for both of **L1** and **L2** (Figure 9). This could be attributed to an ion-pair (hydrogen bonded) complex formed in the two-phase extraction system following proton transfer to the nitrogen atoms of the imidazole units in **L1** and **L2** and then complexation of $Cr_2O7^{2-/}HCr_2O7^{-.7}$

Interestingly, according to the reference reports,⁷ the reference compound **L3** showed almost no significant selective binding of dichromate anions even at low pH. It strongly suggests that the thiacalix[4]arene platform plays an important role in confirming cooperative participation of the peripheral imidazole groups. Furthermore, the extraction results also indicated that extractant **L2** was more effective for the extraction of dichromate anions at low pH (pH 1.5) than extractant **L1**. This could be ascribed to the cavity size of extractant **L2** is smaller and more symmetrical than extractant **L1** (Figure 1, X-ray results). In other words, extractant **L2** maybe provided a more ideal mutual distance needed for hydrogen bonding between two imidazole groups to extract dichromate anions.

Additionally, the extractants **L1** and **L2** possess 'protonswitchable' binding sites, namely the imidazole moieties, which can be protonated at low pH.¹⁷ The protonated forms of **L1** and **L2** are more effective at complexing with dichromate anion through hydrogen bonds. In other words, the extraction of dichromate anion by extractants **L1** and **L2** only occurs when the aqueous phase is acidic, especially at lower pH, in order to form efficient hydrogen bonds. Thus, a reusable extractant concept is possible for extractants **L1** and **L2** by controlling the pH of the aqueous solution (Figure 10).

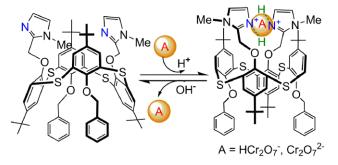


Figure 10. A proposed reusable extractant concept of L2 with Cr₂O₇²⁻.

Conclusions

Two new thiacalix[4]arene receptors L1 and L2 which each possess imidazole moieties in the 1,3-alternate conformation have been synthesized and characterized. The X-ray crystal structures of both L1 and L2 have been determined, confirming their 1,3-alternate conformations. The binding behaviour towards Na⁺, K⁺ and Ag⁺ ions have been examined by ¹H NMR titration experiments in (CDCl₃/CD₃CN; 10:1, v/v) solutions. The ditopic receptor L1 showed affinity not only toward hard alkali metal cations but also toward soft metal Ag⁺ cations, owing to the presence of the two ester moieties at one face of the thiacalix[4]arene cavity and two imidazole moieties at the opposite face. However, the monotopic receptor L2 only exhibited affinity toward Ag⁺ and was unable to bind alkali metal cations due to its lack of the ester groups. The exclusive formation of mononuclear complexes of L1 with metal cations is of particular interest with respect to negative allosteric effects in the thiacalix[4]arene family. These findings further demonstrate that preorganization, suitable conformational changes and affinity have a pronounced effect on the complexation process between the two different arms placed at the two edges of the thiacalix[4]arene platform. Due to its amphoteric nature, the imidazole ring can function as an effective cation and/or anion receptor system. Furthermore, the extraction abilities of L1 and L2 toward dichromate anion were evaluated by a liquid-liquid extraction method. The extraction results indicate that the synthesized L2 can be utilized as a reusable extractant by control of the pH of the aqueous solution.

Electronic Supplementary Information (ESI) available: Details of single-crystal X-ray crystallographic data. ¹H, ¹³C NMR and MS spectra of **L1** and **L2**.

Acknowledgements

This work was performed under the Cooperative Research Program of "Network Joint Research Center for Materials and Devices (In-stitute for Materials Chemistry and Engineering, Kyushu Universi-ty)". We would like to thank the OTEC at Saga University and the International Cooperation Projects of Guizhou Province (No. 20137005) for financial support. The EPSRC is thanked for a travel grant (to CR).

Keywords: Thiacalix[4]arene • Imidazole • Allosteric effect• Dichromate

- a) L. Stryer, *Biochemistry, 4th ed.*, W. H. Freeman, New York, **1995**; b)
 A. Livitzki, *Quantitative Aspects of Allosteric Mechanism*; Springer-Verlag, Berlin, **1978**; c)
 M. Takeuchi, M. Ikeda, A. Sugasaki, S. Shinkai, *Acc. Chem. Res.* **2001**, 34, 865–873; d)
 C. Kremer, A. Lützen. *Chem. Eur. J.* **2013**, 19, 6162-6196.
- [2] a) T. Endoh, N. Sugimoto, *Anal. Chem.* **2015**, 87, 7628–7635; b) A. M. Lifschitz, M. S. Rosen, C. M. McGuirk, and C. A. Mirkin, *J. Am. Chem. Soc.* **2015**, 137, 7252–7261; c) L. Kovbasyuk and R. Kramer, *Chem. Rev.*, **2004**, 104, 3161-3187.
- a) T. Nabeshima, *Coord. Chem. Rev.* **1996**, 148, 151–169; b) B. Linton,
 A.D. Hamilton, *Chem. Rev.* **1997**, 97, 1669–1680; c) T. Nabeshima, S.
 Akine, T. Saiki, *Rev. Heteroatom Chem.* **2000**, 22, 219–239; d) N. Y.
 Edwards and A. L. Possanza, *Supramol. Chem.* **2013**, 25, 446-463.
- [4] a) T. Nabeshima, T. Saiki, S. Akine, *Tetrahedron Lett.* 2004, 45, 4719–4722; b) T. Saiki, J. Iwabuchi, S. Akine, T. Nabeshima, *Tetrahedron Lett.* 2004, 45, 7007–7010; c) T. Nabeshima, Y. Yoshihira, T. Saiki, S. Akine, E. Horn, *J. Am. Chem. Soc.* 2003, 125, 28–29; d) P. D. Beer, J. P. Martin, M. G. B. Drew, *Tetrahedron* 1992, 48, 9917–9928.
- [5] a) H. Tomiyasu, J. L. Zhao, X. L. Ni, X. Zeng, M. R. J. Elsegood, B. Jones, C. Redshaw, S. J. Teat, T. Yamato, *RSC Adv.* 2015, 5, 14747–

14755; b) H. Tomiyasu, C. C. Jin, X. L. Ni, X. Zeng, C. Redshaw and T. Yamato, *Org. Biomol. Chem.* **2014**, 12, 4917–4923; c) J.-L. Zhao, H. Tomiyasu, C. Wu, H. Cong, X. Zeng, S. Rahman, P. E. Georghiou, D. L. Hughes, C. Redshaw, T. Yamato, *Tetrahedron* **2015**, 71, 8521-8527; d) R. Kumar, Y. O. Lee, V. Bhalla, M. Kumar, J. S. Kim, *Chem. Soc. Rev.* **2014**, 43, 4824–4870; e) N. Morohashi, F. Narumi, N. Iki, T. Hattori and S. Miyano, *Chem. Rev.* **2006**, 106, 5291–5316.

- [6] a) S. Singha, D. Kim, H. Seo, S. W. Cho, K. H. Ahn, *Chem. Soc. Rev.* 2015, 44, 4367–4399; b) X. Li, S. L. Gong, W. P. Yang, Y. Li, Y. Y. Chen, X. G. Meng, *J. Incl. Phenom. and Macrocycl. Chem.* 2010, 66, 179–184; c) A. Melaiye, Z. Sun, K. Hindi, A. Milsted, D. Ely, D. H. Reneker, C. A. Tessier, W. J. Youngs, *J. Am. Chem. Soc.* 2005, 127, 2285–2291.
- [7] a) J. L. Zhao, H. Tomiyasu, X. L. Ni, X. Zeng, M. R. J. Elsegood, C. Redshaw, S. Rahman, P. E. Georghiou, T. Yamato, *New J. Chem.* 2014, 38, 6041–6049; b) X. L. Ni, C. C. Jin, X. K. Jiang, M. Takimoto, S. Rahman, X. Zeng, D. L. Hughes, C. Redshaw, T. Yamato, *Org. Biomol. Chem.* 2013, 11, 5435–5442.
- [8] a) X. L. Ni, H. Cong, A. Yoshizawa, S. Rahman, H. Tomiyasu, U. Rayhan, X. Zeng, T. Yamato, *J. Mol. Struct.* 2013, 1046, 110–115; b) E. G. Akgemci, H. Bingol, M. Ersoz, I. Stibor, *Electroanal.*, 2008, 20, 1354–1360; c) T. Yamato, C. P. Casas, M. R. J. Elsegood, S. H. Dale, C. Redshaw, *J. Incl. Phenom. Macrocycl. Chem.* 2006, 55, 31–36; d) N. Iki, F. Narumi, T. Fujimoto, N. Morohashi, S. Miyano, *J. Chem. Soc. Perkin Trans.* 2 1998, 2745–2750.
- [9] C. Liao, X. Zhu, X. G. Sun, S. Dai. Tetrahedron Lett. 2011, 52, 5308– 5310.
- [10] M. Lamouchi, E. Jeanneau, R. Chiriac, D. Ceroni, F. Meganem, A. Brioude, A. W. Coleman, C. Desroches, *Tetrahedron Lett.* **2012**, 53, 2088–2090.
- [11] T. Yamato, C. P. Casas, H. Yamamoto, M. R. J. Elsegood, S. H. Dale, C. Redshaw, J. Incl. Phenom. Macrocycl. Chem. 2005, 54, 261–269.
- [12] U. Svanholm, V. D. Parker, J. Chem. Soc., Perkin Trans. 1 1973, 1, 562–566.
- [13] J. L. Zhao, H. Tomiyasu, X. L. Ni, X. Zeng, M. R. J. Elsegood, C. Redshaw, S. Rahman, P. E. Georghiou, S. J. Teat, T. Yamato, *Org. Biomol. Chem.* 2015, 13, 3476–3483.
- a) C. Raji, T. S. Anirudhan, *Water Res.* **1998**, 32, 3772–3780; b) N.
 Goyal, S. C. Jain, U. C. Banerjee, *Adv. Environ. Res.* **2003**, 7, 311–319.
- [15] P. G. Krishna, J. M. Gladis, U. Rambabu, T. P. Rao and G. R. K. Naidu, *Talanta* **2004**, 63, 541–546.
- [16] P. Molina, A. Tarraga, F. Oton, Org. Biomol. Chem. 2012, 10, 1711– 1724.
- [17] a) M. Tabakci, B. Tabakci, A. D. Beduk, *Tetrahedron* 2012, 68, 4182–4186; b) A. Yilmaz, B. Tabakci, M.Tabakci, *Supramol. Chem.* 2009, 21, 435–441; c) M. Tabakci, S. Memon, M. Yilmaz, *Tetrahedron* 2007, 63, 6861–6865; d) A. Yilmaz, B. Tabakci, E. Akceylan, M. Yilmaz, *Tetrahedron* 2007, 63, 5000–5005.